#### **REVIEW**



# **The modulation of necroptosis and its therapeutic potentials**

**Chun Kim[1](http://orcid.org/0000-0001-9497-708X)**

Accepted: 3 March 2021 / Published online: 12 March 2021 © The Korean Society of Toxicogenomics and Toxicoproteomics 2021

#### **Abstract**

**Purpose of review** Necroptosis is a form of cell death regulated by specifc cellular protein machinery. Although the cell death is tightly controlled like apoptosis, another type of programed cell death, the biological features of necroptosis rather resemble necrosis that is defned as an uncontrolled accidental cell death. The pathway executing necroptosis relies on a protein kinase, RIPK3, and its downstream efector molecule, MLKL. Upon necroptosis initiating signals, both RIPK3 and MLKL undergo extensive post-translation modifcations to construct a death complex called necrosome, fnally leading to lysis of cell membrane. Preclinical mouse models demonstrated the physiological importance of necroptosis in the progress of various infammation-associated diseases. The objective of this brief review is to introduce a new emerging concept in cell death biology and to provide a frst entry into the research feld of necroptosis.

**Recent fndings** The uncovering of necroptosis pathway brought a fundamental change in the basic concept that necrotic cell death is passive and unregulated. Currently, multiple small molecules that can target necrotic cell death are under development and some of them are under clinical trials to evaluate their therapeutic potentials. Better understanding of the molecular mechanism leveraging necroptosis will provide an unprecedented opportunity to pathological necrosis-driven human diseases.

**Keywords** Cell death · Necroptosis · RIPK1 · RIPK3 · MLKL · Infammation

# **Introduction**

Cell death is a fnal event to cease all biological functions in unicellular organisms, but the same cellular event is a dynamic process which controls homeostasis and immunity in multicellular organisms. In humans, as many as a hundred billion cells die each day and are replaced by other cells (Nagata [2018](#page-4-0)). Therefore, the equilibrium between cell death and proliferation is crucial for the maintenance of tissue homeostasis. For example, the continuous cell deathinduced shedding of intestinal epithelial cells during normal tissue homeostasis is a crucial process to maintain intestinal barrier integrity (Vereecke et al. [2011\)](#page-4-1). Cell death also plays important functions in infammation and immunity. It can restrict viral replication (Danthi [2016](#page-3-0)), but at the same time, excessive cell death can cause tissue damage and infammation (Wallach et al. [2014\)](#page-4-2). Indeed, cell death has been acknowledged as an eminent pathological feature of various

 $\boxtimes$  Chun Kim chunkim@hanyang.ac.kr infammatory diseases (Kist and Vucic [2021](#page-4-3)). Consequently, the cell death process during immune responses is required to be tightly coordinated to ensure benefcial immunological efects without resulting in detrimental tissue damage.

The role of cell death in tissue homeostasis, immunity and infammation have been extensively discussed in the context of two diferent types of cell death: apoptosis and necrosis. Apoptosis is a programed cell death whereby a cell commits a suicide by utilizing its own cellular signaling cascades in response to specifc stimuli (Nagata [2018](#page-4-0)). Apoptotic cell dies by a self-destruction program coded by caspase-dependent signaling, and the cellular dead body is then cleared by the neighboring tissue-resident phagocytes. Since the process does not permit leakage of intracellular components, apoptosis is believed as a noninfammatory form of cell death.

In contrast to apoptosis, necrosis is an accidental cell death occurring in a fatally damaged cell by an external fatal insult (Galluzzi [2007](#page-4-4)). The necrotic cell death was originally regarded as a simple failure of cell survival caused by toxic interference of essential cellular functions. A necrotic cell undergoes swelling, followed by uncontrolled rupture of the cell membrane, resulting in a massive release of cellular

 $1$  Department of Molecular and Life Science, Hanyang University, ERICA Campus, Ansan, Republic of Korea

contents to surrounding tissue environment. The unleashed cellular contents function as danger-associated molecular patterns (DAMPs) and often create infammation in tissues (Venereau et al. [2015\)](#page-4-5).

Early studies considered apoptosis as the only form of regulated cell death and necrosis was seen as an unregulated and, therefore, uncontrollable accidental cell death. The traditional concept of necrosis, which was considered as a passive cell death, has been changed since the discovery of a specifc form of regulated cell death called necroptosis. As in necrosis, necroptosis executes a highly infammatory cell death, accompanying by release of DAMPs (Newton and Manning [2016](#page-4-6)). While the phenotype and the consequence of cell death similar to necrosis, collective biochemical and genetic evidence revealed that necroptosis occur as a programed cell death regulated by Receptor Interacting Protein Kinase 3 (RIPK3) and its substrate, Mixed Lineage Kinase Like (MLKL). During necroptosis, modifed MLKL triggers plasma membrane permeabilization and therefore, promotes the release of proinfammatory substances from the dead cell (Grootjans et al. [2017\)](#page-4-7).

Accumulating evidence suggests that necroptosis is a physiologically relevant cell death and it is critical for tissue homeostasis, immunity, and infammation. In this brief review, I will discuss about our current understanding of necroptosis and its implication in tissue homeostasis and infammation.

#### **Overview of necroptosis**

Apoptosis, which is a form of caspase-dependent programed cell death, is characterized by several distinct biological features such as shrinkage of the cell, forming membrane blebs, and nuclear fragmentation (Nagata [2018\)](#page-4-0). This cell death can be triggered by a wide variety of stimuli and conditions such as genotoxic stress, deprivation of growth factors, and occupation of death receptors by their specifc ligands. Apoptosis is believed to be a non-infammatory cell death since apoptotic cells generate "eat me" signals on their cell surface assisting rapid phagocytosis by surrounding tissueresident immune cells. Because of this silent nature of cell death, apoptosis is utilized during normal development to balance cell populations in tissues (Voss and Strasser [2020](#page-4-8)).

The term, necroptosis was frst introduced when a group of researchers recognized a cell death accompanied by necrotic morphological features was caspase-independent and inhibited by a small molecule called, necrostatin-1(Nec-1) (Degterev [2005](#page-3-1)). In contrast to apoptotic cell death, which is governed by caspase-mediated series of molecular events, necroptosis deploys apical protein kinases for its execution and does not require caspases. Although it was initially thought that RIPK1 is a mandatory driver of necroptosis, currently necroptosis is defned as RIPK3 and MLKL-dependent cell death (Galluzzi [2018](#page-4-9)). The main reason why RIPK1 was thought to be required for necroptosis is that Nec‐1, a RIPK1 inhibitor, potently inhibits TNFdependent necroptosis. In fact, many studies still utilize Nec-1 to define a cell death as necroptosis. However, it is now very clear that RIPK1 can function both as an inhibitor and a promoter of necroptosis depending on the cellular contexts (Voss and Strasser [2020](#page-4-8)) and therefore the absence of RIPK1 often results in necroptosis-driven infammation in vivo due to lack of the kinase-independent survival function of RIPK1 (Ito [2016;](#page-4-10) Lin [2016;](#page-4-11) Newton [2016a\)](#page-4-12).

The physiological role of RIPK1 in cell survival and death has been well-demonstrated by mouse genetic studies. RIPK1 defcient mice display postnatal lethality (Kelliher [1998\)](#page-4-13). It was initially thought that the lethal phenotype is delivered by excessive apoptosis in the animals. But later, it was found that the defciency of Fas-associated protein with death domain (FADD), which would inactivate apoptosis, did not rescue the lethal phenotype of RIPK1 KO mice (Zhang [2011](#page-4-14)). Furthermore, loss of RIPK3 also did not prevent the lethality resulted from RIPK1 defciency in the animals. The postnatal death of RIPK1 KO mice was eventually rescued by the combined ablation of Caspase-8 and RIPK3, indicating that RIPK1 can repress both caspase‐8‐dependent apoptosis and RIPK3‐dependent necroptosis (Dillon [2014](#page-3-2); Rickard [2014](#page-4-15)). Interestingly, animals with a kinase-inactive form of RIPK1 are viable, unlike RIPK1 KO mice (Berger [2014](#page-3-3); Polykratis [2014](#page-4-16)). The result demonstrates that a kinase-independent scafolding function of RIPK1 is responsible for the pro-survival efect of RIPK1. Taken together, RIPK1 can either promotes apoptosis and necroptosis via its kinase activity or suppress cell death by its kinase-independent function. In this regard, it is important to keep it in mind that inhibition of a cell death by RIPK1 inhibitors does not necessarily indicate the mode of cell death is necroptosis.

### **Activation of necroptosis**

Necroptosis can be triggered by diferent initiating signals such as death receptors, some toll-like receptors, and unknown ligands sensed by protein, Z-DNA binding protein 1(ZBP1) (Grootjans et al. [2017\)](#page-4-7). All these necroptotic signals employ proteins contain RIP Homotypic Interaction Motif (RHIM) and the RHIM provides a critical protein–protein interaction interface that regulates the formation of death-inducing protein complexes (Rebsamen [2009\)](#page-4-17). There are only four proteins that contain RHIM in the mammalian system: RIPK1, RIPK3, ZBP1 and TIR-domain-containing adapter-inducing interferon-β (TRIF).

The molecular mechanism of necroptosis is most wellestablished in TNFR1 signaling pathway(Fig. [1](#page-2-0)) (Wajant and



<span id="page-2-0"></span>**Fig. 1** TNFR1 signaling pathway. Upon ligation of TNF, TNFR1 forms a large membrane-associated complex that is responsible for infammation and cell survival-promoting cellular signaling pathways. During the process, if the survival-promoting signal is attenuated, death-inducing cytoplasmic complex can potentiate caspase-8 dependent apoptosis. RIPK3-dependent necroptosis is believed as an alternative backup cell death program, which occurs when apoptosis is compromised. The RIPK3-dependent modifcation of MLKL induces the translocation of MLKL to cell membrane resulting in lysis of a cell.

Siegmund [2019](#page-4-18)). TNFR1 is a member of the death receptor family. Upon the ligation of TNF, the death domain (DD) in TNFR1 enables to recruit DD-containing proteins including TNFR1-associated death domain protein (TRADD) and RIPK1 (Haas [2009\)](#page-4-19). The ubiquitin ligases, cIAP1/2, which preexist as cytoplasmic complexes with TNF receptorassociated factor 2 (TRAF2) (Zheng et al. [2010](#page-4-20)), are then recruited to the TFNR1 membrane complex, followed by LUBAC, an E3 ubiquitin ligase complex composed of proteins, SHARPIN, HOIP, and HOIL-1 (Spit et al. 2019). The ubiquitin chains assembled by the coordination of cIAP1/2 and LUBAC allow the recruitment of ubiquitin binding proteins such as TAK1-binding protein 2/3 (TAB2/3) and NF-κB essential modulator (NEMO), resulting in the activation of NF-kB signaling pathway. Once TNF-mediated proinfammatory and cell survival signals are established, later the TNFR1 membrane complex transforms into a cell deathinducting cytoplasmic complex associated with FADD, caspase-8 and RIPK3 (Grootjans et al. [2017](#page-4-7)). The cytoplasmic death complex can induce caspase-8-dependent apoptosis and normally, RIPK3 is inactivated in the complex by its cleavage by caspase-8 (Feng [2007](#page-3-4)). Nevertheless, under caspase-inactive conditions, RIPK3-dependent necroptotic pathway begins by the autophosphorylation of RIPK3, which enables interaction with its efector, MLKL. The RIPK3 dependent phosphorylation of MLKL allows oligomerization of MLKL, leading to lysis cell membranes (Sun [2012](#page-4-21); Petrie et al. [2019](#page-4-22)).

## **Pathological implications of necroptosis**

Necrotic cell death is found in various pathological conditions (Nieminen [2003\)](#page-4-23). Despite its presence in a broad range of clinical states, necrosis in human disease was not perceived as a potential therapeutic target until very recently since it was thought that necrotic cell death is an uncontrollable process. However, the discovery of necroptosis, which is regulated by cellular machinery, provided a new opportunity to target pathological necrosis.

Recent studies performed in preclinical animal models have revealed the potential beneft of targeting necroptosis in various human diseases, including sepsis, tissue injuries, chronic infammatory disease, and neurodegenerations (Spit et al. [2019;](#page-4-24) Khoury et al. [2020](#page-4-25); Molnar [2019\)](#page-4-26). Notably, some studies utilized only specifc RIPK1 inhibitors or RIPK1-defcient mice in testing necroptosis as a potential therapeutic target. Importantly, as discussed before, RIPK1 can activate both apoptosis and necroptosis, and can potentiate NF-kB-dependent infammation. Therefore, RIPK1 dependent phenotypes do not necessarily correlate with necroptosis-driven pathological outcomes. In this section, I will briefy discuss only studies that confrmed the pathological contribution of necroptosis in the absence of activity of RIPK3 or MLKL.

The inflammatory nature of necroptosis proposes its potential involvement in the pathologies of acute infammation. A mouse model of TNF-induced systemic infammatory response syndrome (SIRS) showed that loss of RIPK3 in animals were protected against lethal SIRS (Duprez [2011](#page-3-5)). In the same study, ablation of RIPK3 also prevented cecal ligation and puncture-induced sepsis.

In addition to acute infammation models, necroptosis has been implicated in chronic infammation-associated disease models. A model of nonalcoholic fatty liver disease (NAFLD) demonstrated that when RIPK3-deficient mice were fed with methionine- and choline-deficient (MCD), MCD diet-induced liver injury, steatosis and fbrosis were attenuated (Afonso [2015\)](#page-3-6). Moreover, the high levels of RIPK3 expression in patient livers were correlated with poor prognosis of alcoholic cirrhosis (Zhang [2018](#page-4-27)). In another chronic infammation-associated animal model, RIPK3-defciency reduced atherosclerotic lesions, which were promoted by loss of LDL receptor (Lin [2013](#page-4-28)).

Necroptosis also appears to contribute to multiple acute tissue injury models. A study using a model of kidney I/R reported protection of RIPK3-defcient mice but interestingly, the protection was minimal in MLKL-defcient mice (Newton [2016b\)](#page-4-29). Another acute kidney model induced by toxic folic acid demonstrated that RIPK3 or MLKL did not contribute to the early stage of renal injury, but later when the TWEAK signaling-dependent necroptosis occurred, ablation of RIPK3 or MLKL reduced the late injury of kidney in the model (Martin-Sanchez [2018\)](#page-4-30). In addition to kidney models, RIPK3-defcient mice were protected from long-term adverse post-infarct remodeling following I/R injury-induced myocardial infarction (Luedde [2014](#page-4-31)). Furthermore, the contribution of RIPK3 to lung injury was reported in a murine model of acute lung injury (ALI). The study demonstrated that both necroptosis-dependent and -independent functions of RIPK3 can contribute to the phenotype of an LPS-induced ALI model. Importantly, RIPK3 inhibitor ameliorated lung injury and reduced infammation in this model (Chen [2018](#page-3-7)).

Beside acute tissue injury models, necroptosis has been reported in neurodegenerative diseases. RIPK3 defciency markedly improves neurological and systemic disease in a mouse model of Gaucher's disease (GD). When GD was induced by daily injection of a GlcCerase inhibitor, RIPK3 defcient mice showed considerably improved survival and motor coordination. In a mouse amyotrophic lateral sclerosis (ALS) model, the RIPK1, RIPK3 and MLKL–dependent necroptosis was observed in the central nerve system. In this optineurin-defcient ALS mice, the axonal pathology was rescued by RIPK3-deficiecy (Ito [2016](#page-4-10)).

Collectively, the studies discussed above synchronously propose crucial roles of necroptosis in the development of pathophysiology. It is worth to note that some studies suggested there are MLKL-independent, therefore necroptosis-independent roles of RIPK3 (Chen [2018;](#page-3-7) Alvarez-Diaz [2016](#page-3-8)). In this regard, when studying the role of necroptosis in vivo, it is important to confrm if the identifed RIPK3 dependent phenotypes are also MLKL-dependent, therefore, ensuring *bona fde* necroptosis-driven pathology.

## **Conclusion**

Investigating the details of the mechanism regulating necroptosis is critical to understand physiological roles of necroptosis in the pathogenesis of human diseases. The key signaling molecules in necroptotic pathway are largely distinct from those involved in other pathways including apoptosis and infammation. So far, therapeutic potentials of targeting necroptosis have been reported mainly in mouse models. It remains to be demonstrated if blocking necroptosis is a viable strategy to treat human diseases.

The development of specifc inhibitors against RIPK3 was initially perused to achieve specifc therapeutic benefts in the necroptosis-associated pathophysiology of the disease. But it turned out that the inhibition of RIPK3 either by specifc inhibitors or mutations leading to inactivation of its kinase activity not only block necroptosis but promotes spontaneous apoptosis (Mandal [2014\)](#page-4-32). Hence, it seems a specifc inhibitor against MLKL will be a better strategy to target necroptosis in the clinic.

**Acknowledgements** This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korean government (MSIT) (No. 2020R1F1A1074601).

#### **Declarations**

**Conflict of Interest** The author declares that there is no confict of interest.

**Human and animal rights** This review article contains published animal studies, which were performed under institutional and national guidelines.

## **References**

- <span id="page-3-6"></span>Afonso MB et al (2015) Necroptosis is a key pathogenic event in human and experimental murine models of non-alcoholic steatohepatitis. Clin Sci (Lond) 129:721–739. [https://doi.org/10.](https://doi.org/10.1042/CS20140732) [1042/CS20140732](https://doi.org/10.1042/CS20140732)
- <span id="page-3-8"></span>Alvarez-Diaz S et al (2016) The pseudokinase MLKL and the kinase RIPK3 have distinct roles in autoimmune disease caused by loss of death-receptor-induced apoptosis. Immunity 45:513–526. <https://doi.org/10.1016/j.immuni.2016.07.016>
- <span id="page-3-3"></span>Berger SB et al (2014) Cutting Edge: RIP1 kinase activity is dispensable for normal development but is a key regulator of infammation in SHARPIN-defcient mice. J Immunol 192:5476–5480. <https://doi.org/10.4049/jimmunol.1400499>
- <span id="page-3-7"></span>Chen J et al (2018) RIP3 dependent NLRP3 infammasome activation is implicated in acute lung injury in mice. J Transl Med 16:233. <https://doi.org/10.1186/s12967-018-1606-4>
- Choi ME, Price DR, Ryter SW, Choi AMK (2019) Necroptosis: a crucial pathogenic mediator of human disease. JCI Insight. <https://doi.org/10.1172/jci.insight.128834>
- <span id="page-3-0"></span>Danthi P (2016) Viruses and the diversity of cell death. Annu Rev Virol 3:533–553. [https://doi.org/10.1146/annurev-virol](https://doi.org/10.1146/annurev-virology-110615-042435) [ogy-110615-042435](https://doi.org/10.1146/annurev-virology-110615-042435)
- <span id="page-3-1"></span>Degterev A et al (2005) Chemical inhibitor of nonapoptotic cell death with therapeutic potential for ischemic brain injury. Nat Chem Biol 1:112–119. <https://doi.org/10.1038/nchembio711>
- <span id="page-3-2"></span>Dillon CP et al (2014) RIPK1 blocks early postnatal lethality mediated by caspase-8 and RIPK3. Cell 157:1189–1202. [https://doi.](https://doi.org/10.1016/j.cell.2014.04.018) [org/10.1016/j.cell.2014.04.018](https://doi.org/10.1016/j.cell.2014.04.018)
- <span id="page-3-5"></span>Duprez L et al (2011) RIP kinase-dependent necrosis drives lethal systemic infammatory response syndrome. Immunity 35:908– 918. <https://doi.org/10.1016/j.immuni.2011.09.020>
- <span id="page-3-4"></span>Feng S et al (2007) Cleavage of RIP3 inactivates its caspase-independent apoptosis pathway by removal of kinase domain. Cell Signal 19:2056–2067. [https://doi.org/10.1016/j.cellsig.2007.](https://doi.org/10.1016/j.cellsig.2007.05.016) [05.016](https://doi.org/10.1016/j.cellsig.2007.05.016)
- <span id="page-4-4"></span>Galluzzi L et al (2007) Cell death modalities: classifcation and pathophysiological implications. Cell Death Difer 14:1237–1243. <https://doi.org/10.1038/sj.cdd.4402148>
- <span id="page-4-9"></span>Galluzzi L et al (2018) Molecular mechanisms of cell death: recommendations of the Nomenclature Committee on Cell Death 2018. Cell Death Difer 25:486–541. [https://doi.org/10.1038/](https://doi.org/10.1038/s41418-017-0012-4) [s41418-017-0012-4](https://doi.org/10.1038/s41418-017-0012-4)
- <span id="page-4-7"></span>Grootjans S, Vanden Berghe T, Vandenabeele P (2017) Initiation and execution mechanisms of necroptosis: an overview. Cell Death Difer 24:1184–1195.<https://doi.org/10.1038/cdd.2017.65>
- <span id="page-4-19"></span>Haas TL et al (2009) Recruitment of the linear ubiquitin chain assembly complex stabilizes the TNF-R1 signaling complex and is required for TNF-mediated gene induction. Mol Cell 36:831–844. <https://doi.org/10.1016/j.molcel.2009.10.013>
- <span id="page-4-10"></span>Ito Y et al (2016) RIPK1 mediates axonal degeneration by promoting infammation and necroptosis in ALS. Science 353:603–608. <https://doi.org/10.1126/science.aaf6803>
- <span id="page-4-13"></span>Kelliher MA et al (1998) The death domain kinase RIP mediates the TNF-induced NF-kappaB signal. Immunity 8:297–303. [https://](https://doi.org/10.1016/s1074-7613(00)80535-x) [doi.org/10.1016/s1074-7613\(00\)80535-x](https://doi.org/10.1016/s1074-7613(00)80535-x)
- <span id="page-4-25"></span>Khoury MK, Gupta K, Franco SR, Liu B (2020) Necroptosis in the pathophysiology of disease. Am J Pathol 190:272–285. [https://](https://doi.org/10.1016/j.ajpath.2019.10.012) [doi.org/10.1016/j.ajpath.2019.10.012](https://doi.org/10.1016/j.ajpath.2019.10.012)
- <span id="page-4-3"></span>Kist M, Vucic D (2021) Vucic D (2021)) Cell death pathways: intricate connections and disease implications. EMBO J. [https://doi.org/10.](https://doi.org/10.15252/embj.2020106700) [15252/embj.2020106700](https://doi.org/10.15252/embj.2020106700)
- <span id="page-4-28"></span>Lin J et al (2013) A role of RIP3-mediated macrophage necrosis in atherosclerosis development. Cell Rep 3:200–210. [https://doi.org/](https://doi.org/10.1016/j.celrep.2012.12.012) [10.1016/j.celrep.2012.12.012](https://doi.org/10.1016/j.celrep.2012.12.012)
- <span id="page-4-11"></span>Lin J et al (2016) RIPK1 counteracts ZBP1-mediated necroptosis to inhibit infammation. Nature 540:124–128. [https://doi.org/10.](https://doi.org/10.1038/nature20558) [1038/nature20558](https://doi.org/10.1038/nature20558)
- <span id="page-4-31"></span>Luedde M et al (2014) RIP3, a kinase promoting necroptotic cell death, mediates adverse remodelling after myocardial infarction. Cardiovasc Res 103:206–216. <https://doi.org/10.1093/cvr/cvu146>
- <span id="page-4-32"></span>Mandal P et al (2014) RIP3 induces apoptosis independent of pronecrotic kinase activity. Mol Cell 56:481–495. [https://doi.org/10.](https://doi.org/10.1016/j.molcel.2014.10.021) [1016/j.molcel.2014.10.021](https://doi.org/10.1016/j.molcel.2014.10.021)
- <span id="page-4-30"></span>Martin-Sanchez D et al (2018) TWEAK and RIPK1 mediate a second wave of cell death during AKI. Proc Natl Acad Sci U S A 115:4182–4187.<https://doi.org/10.1073/pnas.1716578115>
- <span id="page-4-26"></span>Molnar T et al (2019) Current translational potential and underlying molecular mechanisms of necroptosis. Cell Death Dis 10:860. <https://doi.org/10.1038/s41419-019-2094-z>
- <span id="page-4-0"></span>Nagata S (2018) Apoptosis and clearance of apoptotic cells. Annu Rev Immunol 36:489–517. [https://doi.org/10.1146/annurev-immun](https://doi.org/10.1146/annurev-immunol-042617-053010) [ol-042617-053010](https://doi.org/10.1146/annurev-immunol-042617-053010)
- Newton K (2020) Multitasking kinase RIPK1 regulates cell death and infammation. Cold Spring Harb Perspect Biol. [https://doi.org/10.](https://doi.org/10.1101/cshperspect.a036368) [1101/cshperspect.a036368](https://doi.org/10.1101/cshperspect.a036368)
- <span id="page-4-6"></span>Newton K, Manning G (2016) Necroptosis and infammation. Annu Rev Biochem 85:743–763. [https://doi.org/10.1146/annurev-bioch](https://doi.org/10.1146/annurev-biochem-060815-014830) [em-060815-014830](https://doi.org/10.1146/annurev-biochem-060815-014830)
- <span id="page-4-12"></span>Newton K et al (2016a) RIPK1 inhibits ZBP1-driven necroptosis during development. Nature 540:129–133. [https://doi.org/10.1038/](https://doi.org/10.1038/nature20559) [nature20559](https://doi.org/10.1038/nature20559)
- <span id="page-4-29"></span>Newton K et al (2016b) RIPK3 defciency or catalytically inactive RIPK1 provides greater beneft than MLKL defciency in mouse models of inflammation and tissue injury. Cell Death Differ 23:1565–1576.<https://doi.org/10.1038/cdd.2016.46>
- <span id="page-4-23"></span>Nieminen AL (2003) Apoptosis and necrosis in health and disease: role of mitochondria. Int Rev Cytol 224:29–55. [https://doi.org/](https://doi.org/10.1016/s0074-7696(05)24002-0) [10.1016/s0074-7696\(05\)24002-0](https://doi.org/10.1016/s0074-7696(05)24002-0)
- <span id="page-4-22"></span>Petrie EJ, Czabotar PE, Murphy JM (2019) The structural basis of necroptotic cell death signaling. Trends Biochem Sci 44:53–63. <https://doi.org/10.1016/j.tibs.2018.11.002>
- <span id="page-4-16"></span>Polykratis A et al (2014) Cutting edge: RIPK1 Kinase inactive mice are viable and protected from TNF-induced necroptosis in vivo. J Immunol 193:1539–1543. [https://doi.org/10.4049/jimmunol.](https://doi.org/10.4049/jimmunol.1400590) [1400590](https://doi.org/10.4049/jimmunol.1400590)
- <span id="page-4-17"></span>Rebsamen M et al (2009) DAI/ZBP1 recruits RIP1 and RIP3 through RIP homotypic interaction motifs to activate NF-kappaB. EMBO Rep 10:916–922.<https://doi.org/10.1038/embor.2009.109>
- <span id="page-4-15"></span>Rickard JA et al (2014) RIPK1 regulates RIPK3-MLKL-driven systemic inflammation and emergency hematopoiesis. Cell 157:1175–1188.<https://doi.org/10.1016/j.cell.2014.04.019>
- <span id="page-4-24"></span>Spit M, Rieser E, Walczak H (2019) Linear ubiquitination at a glance. J Cell Sci.<https://doi.org/10.1242/jcs.208512>
- <span id="page-4-21"></span>Sun L et al (2012) Mixed lineage kinase domain-like protein mediates necrosis signaling downstream of RIP3 kinase. Cell 148:213–227. <https://doi.org/10.1016/j.cell.2011.11.031>
- <span id="page-4-5"></span>Venereau E, Ceriotti C, Bianchi ME (2015) DAMPs from cell death to new life. Front Immunol 6:422. [https://doi.org/10.3389/fmmu.](https://doi.org/10.3389/fimmu.2015.00422) [2015.00422](https://doi.org/10.3389/fimmu.2015.00422)
- <span id="page-4-1"></span>Vereecke L, Beyaert R, van Loo G (2011) Enterocyte death and intestinal barrier maintenance in homeostasis and disease. Trends Mol Med 17:584–593. <https://doi.org/10.1016/j.molmed.2011.05.011>
- <span id="page-4-8"></span>Voss AK, Strasser A (2020) The essentials of developmental apoptosis. F1000Res.<https://doi.org/10.12688/f1000research.21571.1>
- <span id="page-4-18"></span>Wajant H, Siegmund D (2019) TNFR1 and TNFR2 in the control of the life and death balance of macrophages. Front Cell Dev Biol 7:91. <https://doi.org/10.3389/fcell.2019.00091>
- <span id="page-4-2"></span>Wallach D, Kang TB, Kovalenko A (2014) Concepts of tissue injury and cell death in infammation: a historical perspective. Nat Rev Immunol 14:51–59.<https://doi.org/10.1038/nri3561>
- <span id="page-4-14"></span>Zhang H et al (2011) Functional complementation between FADD and RIP1 in embryos and lymphocytes. Nature 471:373–376. [https://](https://doi.org/10.1038/nature09878) [doi.org/10.1038/nature09878](https://doi.org/10.1038/nature09878)
- <span id="page-4-27"></span>Zhang Z et al (2018) RIPK3-mediated necroptosis and neutrophil infltration are associated with poor prognosis in patients with alcoholic cirrhosis. J Immunol Res 2018:1509851. [https://doi.org/](https://doi.org/10.1155/2018/1509851) [10.1155/2018/1509851](https://doi.org/10.1155/2018/1509851)
- <span id="page-4-20"></span>Zheng C, Kabaleeswaran V, Wang Y, Cheng G, Wu H (2010) Crystal structures of the TRAF2: cIAP2 and the TRAF1: TRAF2: cIAP2 complexes: affinity, specificity, and regulation. Mol Cell 38:101-113.<https://doi.org/10.1016/j.molcel.2010.03.009>

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.