

# Bambusicolous fungi

Dong Qin Dai<sup>1,2,3,4,5</sup> · Rungtiwa Phookamsak<sup>1,3,4,5</sup> · Nalin N. Wijayawardene<sup>4,5</sup> ·  
Wen Jing Li<sup>1,3,4,5</sup> · D. Jayarama Bhat<sup>6,7</sup> · Jian Chu Xu<sup>1,3</sup> · Joanne E. Taylor<sup>8</sup> ·  
Kevin D. Hyde<sup>1,3,4,5</sup> · E. Chukeatirote<sup>4,5</sup>

Received: 22 March 2016 / Accepted: 11 April 2016 / Published online: 21 May 2016  
© School of Science 2016

**Abstract** Forty-three species of microfungi from bamboo are treated, including one new family, *Occultibambusaceae*, three new genera, *Neoanthostomella*, *Occultibambusa* and *Seriascoma*, 27 new species, one renamed species and 15 re-described or re-illustrated species, and four designated reference specimens are treated in this paper, the majority of which are saprobic on dead culms. To determine species identification, separate phylogenetical analyses for each group are carried out, based on molecular data from this study and sequences downloaded from GenBank. Morphologically similar species and phylogenetically close taxa are compared and discussed. In addition a list of bambusicolous fungi published since Hyde and colleagues in 2002 is provided.

✉ Kevin D. Hyde  
kdhyde3@gmail.com

<sup>1</sup> Key Laboratory for Plant Diversity and Biogeography of East Asia, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, People's Republic of China

<sup>2</sup> Center for Yunnan Plateau Biological Resources Protection and Utilization, College of Biological Resource and Food Engineering, Qujing Normal University, Qujing, Yunnan 655011, People's Republic of China

<sup>3</sup> World Agroforestry Centre, East and Central Asia, Heilongtan, Kunming 650201, People's Republic of China

<sup>4</sup> School of Science, Mae Fah Luang University, Chiang Rai 57100, Thailand

<sup>5</sup> Centre of Excellence in Fungal Research, Mae Fah Luang University, Chiang Rai 57100, Thailand

<sup>6</sup> Department of Botany, Goa University, Goa, India

<sup>7</sup> No. 128/1-J, Azad Housing Society, Curca, P.O., Goa Velha 403108, India

<sup>8</sup> Royal Botanic Garden Edinburgh, 20A Inverleith Row, Edinburgh EH3 5LR, UK

**Keywords** *Dothideomycetes* · *Sordariomycetes* · *Botryosphaerales* · *Chaetosphaerales* · *Hypocreales* · *Myriangiales* · *Pleosporales* · *Sordariales* · *Togniniales* · *Tubeufiales* · *Valsariales* · *Xylariales* · *Apiosporaceae* · *Bambusicolaceae* · *Botryosphaeriaceae* · *Diatrypaceae* · *Lasiosphaeriaceae* · *Myriangiaceae* · *Nectriaceae* · *Roussoellaceae* · *Stachybotryaceae* · *Togniniaceae* · *Tubeufiaceae* · *Valsariaceae* · *Xylariaceae*

## Table of Contents

Ascomycota

*Pezizomycotina*

**Class *Dothideomycetes***

***Botryosphaerales***

***Botryosphaeriaceae***

**1. *Neodeightonia microspora*** D.Q. Dai & K.D.

Hyde, *sp. nov.* ..... 6

**2. *Neodeightonia subglobosa***

C. Booth ..... 6

***Myriangiales***

***Myriangiaceae***

**3. *Mendogia bambusina*** Racib.

..... 13

**4. *Mendogia macrostroma*** D.Q. Dai & K.D. Hyde,

*sp. nov.* ..... 13

**Pleosporales****Bambusicolaceae**

5. *Bambusicola didymospora* Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 18
6. *Bambusicola pustulata* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 20
7. *Bambusicola thailandica* Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 22
8. *Bambusicola triseptatispora* Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 22
9. *Occultibambusaceae* D.Q. Dai & K.D. Hyde, *fam. nov.* ..... 25
10. *Occultibambusa* D.Q. Dai & K.D. Hyde, *gen. nov.* ..... 26
11. *Occultibambusa bambusae* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 26
12. *Occultibambusa fusispora* Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 28
13. *Occultibambusa pustula* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 30
14. *Seriascoma* Phookamsak, D.Q. Dai & K.D. Hyde, *gen. nov.* ..... 30
15. *Seriascoma didymospora* Phookamsak, D.Q. Dai, S.C.Karunarathana & K.D. Hyde, *sp. nov.*..... 32

**Roussoellaceae**

16. *Roussoella mukdahanensis* Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 32
17. *Roussoella neopustulans* D.Q. Dai, J.K. Liu & K.D. Hyde ..... 35
18. *Roussoella pseudohysterioides* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 37
19. *Roussoella tuberculata* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 19
20. *Roussoella siamensis* Phookamsak, J.K. Liu & K.D. Hyde ..... 40

**Tubeufiales****Tubeufiaceae**

21. *Tubeufia javanica* Penz. & Sacc. .... 41
22. *Tubeufia longiseta* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 42

**Valsariales****Valsariaceae**

23. *Bambusaria bambusae* (J.N. Kapoor & H.S. Gill) Jaklitsch et al. .... 44

**Class Sordariomycetes****Chaetosphaeriales genera, incertae sedis**

24. *Leptospora bambusae* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 44

**Hypocreales****Stachybotryaceae**

25. *Myrothecium thailandicum* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 48
26. *Myrothecium chiangmaiense* D. Q. Dai & K.D. Hyde, *sp. nov.* ..... 49
27. *Myrothecium cylindrosporium* D.Q. Dai & K.D. Hyde, *nom. nov.* ..... 51
28. *Myrothecium uttaraditense* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 52

**Nectriaceae**

29. *Nectria pseudotrichia* Berk. & M.A. Curtis ..... 54

**Sordariales****Lasiosphaeriaceae**

30. *Cercophora thailandica* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 57

**Togniniales****Togniniaceae**

31. *Phaeoacremonium sphinctrophorum* L. Mostert, Summerb. & Crous ..... 59

**Xylariales****Apiosporaceae**

32. *Arthrinium longistromum* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 62

33. *Arthrinium rasikravindrae* Shiv M. Singh et al. .... 32
34. *Arthrinium thailandicum* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 66
35. *Arthrinium yunnanum* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 69
- Diatrypaceae**
36. *Eutypa linearis* Rehm ..... 69
37. *Peroneutypa scoparia* (Schwein.) Carmarán & A.I. Romero ..... 73
- Xylariaceae**
38. *Anthostomella pseudobambusicola* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 75
39. *Astrocystis mirabilis* Berk. & Broome ..... 75
40. *Daldinia bambusicola* Y.M. Ju et al. .... 40
41. *Hypoxylon pseudefendleri* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 82
42. *Hypoxylon neosublenormandii* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 82
43. *Neoanthostomella* D.Q. Dai & K.D. Hyde, *gen. nov.* ..... 84
44. *Neoanthostomella pseudostromatica* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 84
45. *Vamsapriya bambusicola* D.Q. Dai et al. .... 86
46. *Xylaria bambusicola* Y.M. Ju & J.D. Rogers ..... 88
- Sordariomycetes genera, incertae sedis**
47. *Pleurophragmium bambusinum* D.Q. Dai & K.D. Hyde, *sp. nov.* ..... 92

## Introduction

Bamboo, members of the grass family *Poaceae* and subfamily *Bambusoideae*, comprise more than 115 genera with approximately 1,450 species (Gratani et al. 2008; Kelchner and Group 2013; Bamboo Botanicals 2016). Bamboo species are found growing in diverse climates, from cold mountain areas to hot tropical regions (Kelchner and Group 2013). They are distributed in almost all countries of Asia, Australia, Africa and the Americas (Bystrakova et al. 2003; Shimokawa et al. 2009; Song et al. 2011). Europe however, has no native bamboo species (INBAR 2016). In Thailand, bamboos are widely distributed in several forest types, especially mixed deciduous

forest and in regenerated forests (Kirschner et al. 2009). Bamboo has a long history of use in Asian furniture and construction of roof thatches (Lin 2004). In China and India, bamboo has been used to hold up simple suspension bridges, either by making cables of split bamboo or twisting whole culms of sufficiently pliable bamboo together (Peters 1987). Bamboo shoots are also edible (Dransfield and Widjaja 1995) and are used in numerous Asian culinary dishes and broths, and are available in supermarkets in various sliced forms, both as fresh and canned versions (Idris and Mohamad 2002). Bamboo is used in Chinese traditional medicine for treating infections and healing (Janssen 1991).

Studies on the taxonomy and phylogeny of bamboo fungi can be of economic significance (Zhang and Zhang 2000; Jiang et al. 2002; Dharmananda 2004; Zhou et al. 2005; Singhal et al. 2013). Some fungi are pathogens causing spots or malformations on bamboo leaves and culms (Xu et al. 2006, 2007). Examples include *Linearistroma lineare* (Rehm) Höhn., *Shiraria bambusicola* P. Henn. and *Calonectria* spp., which can affect the growth and development of bamboo, and cause serious economic losses (Tewari 1993; Mohanan 2002; Li et al. 2009). However some fungi can be beneficial to humans. *Shiraia bambusicola* is of medical importance because of its metabolite, hypocrellin (Hudson et al. 1994), which has promising applications in photodynamic therapy (PDT) for anticancer treatment (Liu et al. 2012b).

A review of the literature on bamboo-associated fungi reveals that nearly 1100 species, which belong to 228 genera, have so far been described or recorded worldwide (Hyde et al. 2002a, b). There have been several large studies on bambusicolous fungi. Hino and Katumoto (1960) published ‘Icones Fungorum Bambusicolum Japonicorum’ and listed 258 species of fungi from bamboo in Japan. Petrini et al. (1989) investigated the fungi on bamboo in France and listed 63 species, including two new species. Eriksson and Yue (1998) provided a checklist of the ascomycetes on bamboo that included 587 names. Zhang and Wang (1999) recorded 213 species (including 104 species, belonging 60 genera of ascomycetes), described from bamboo in China.

There have been few thorough studies on the taxonomy and phylogeny of fungi on bamboo (Hyde et al. 2002a). Generally bambusicolous fungi have been poorly documented (Rehm 1913, 1914; Sydow and Sydow 1913, 1914b; Eriksson and Yue 1998; Zhou and Hyde 2001, 2002). In Japan, ca. 300 species are known from bamboo (Tanaka and Harada 2004) and from 2003 to 2015, Tanaka described 48 new species, which belong to 19 genera, including seven new genera from bamboo in Japan (Tanaka and Harada 2003a, b, 2004, 2005a, b; Shirouzu and Harada 2004; Hatakeyama et al. 2005, 2008; Sato et al. 2008; Tanaka et al. 2005, 2009, 2015). Since 2011 until now, 60 new species, belonging to 29 genera, including 16 new genera were published on bamboo in Thailand (Liu et al. 2011, 2012a, 2014, 2015; Dai et al. 2012, 2014a, b, c,

2015, 2016; Phookamsak et al. 2014, 2015; Adamčík et al. 2015; Ariyawansa et al. 2015; Senanayake et al. 2015).

As pointed by Hyde et al. (2002b), the hitherto knowledge on bamboo fungi is incomplete and largely remained at cataloguing stage. Numerous bamboo fungi lack molecular data and defined studies. In this paper, we provide detailed taxonomic and phylogenetic information on bambusicolous fungi from Thailand, including one taxon from China.

## Materials and methods

### Collection and isolation

Dead bamboo culms were collected from China and Thailand. The samples were placed in Ziplock plastic bags and brought to the laboratory. Fresh materials were examined by using stereo and compound microscopes. The specimens were incubated in a moist chamber for 3–7 days at room temperature, if they did not sporulate. Microscopic observations were made in distilled water. Staining of the ascus apical ring was tested in Melzer's reagent. India ink was used to observe the mucilaginous sheath and appendages of spores. Micro-morphological characters were examined by differential interference contrast (DIC), using a Nikon ECLIPSE 80i or Ni compound microscope with a Cannon 550D or a 600D digital camera. Fruiting bodies were observed by stereomicroscopy using a Zeiss Stereo Discovery V8 and photographed by AxioCam ERc 5 s. Measurements were made using Tarosoft (R) Image Frame Work software.

The fungi were isolated based on the way of single spore isolation. Ascomata or conidiomata were cut horizontally, the hymenium containing ascospores, conidia mass or conidiophores bearing conidia (hyphomycetes) were transferred to a drop of sterile water on a flamed concave microscope slide. The hymenium contents or conidia mass were broken up and separated by using the sterile needles until the single spores suspended in the sterile water. The spore suspension was siphoned by a sterile pipette and dropped on the surface of a Petri dish with 2 % Difco potato-dextrose agar (PDA) media. To obtain germinating spores, the plates were kept within 24–48 h at room temperature, under 12 h light/12 h dark. The germinating spores were individually transferred to three fresh plates. Cultures were growing on PDA or MEA (33.6 g/L sterile distilled water, Difco malt extract) in 15–45 days, at 25–32 °C, under 12 h light/12 h dark for recording growth rates and culture characters. Colours rated according to Komerup and Wanscher (1978). Pieces of growing colonies were cut and transferred on WA (15 g/l sterile distilled water) placed nearby the sterile bamboo pieces to sporulate.

The growing colonies were transferred to 1.5 ml microcentrifuge tubes with PDA to store at 4 °C and suspended in 2 ml screw cap microcentrifuge tubes with 15 % glycerol for storage at –20 °C. Type materials are

deposited at the herbarium of Mae Fah Luang University, Chiang Rai, Thailand (MFLU) and Herbarium of Kunming Institute of Botany, Chinese Academy of Sciences (KUN). Living cultures are deposited at Mae Fah Luang University Culture Collection (MFLUCC), CBS-KNAW Fungal Biodiversity Centre, Utrecht, The Netherlands (CBS), Mycothèque de l'Université catholique de Louvain (MUCL), Kunming Culture Collection (KUMCC) and in the International Collection of Microorganisms from Plants (ICMP), Landcare Research, New Zealand. Facesoffungi (Jayasiri et al. 2015) and Index Fungorum (2016) numbers were provided for new taxa.

### DNA extraction, PCR amplification and sequencing

Fungal isolates were grown on PDA for 15–30 day at 27 °C and genomic DNA was extracted from fresh mycelia, following the protocols of Biospin Fungus Genomic DNA Extraction Kit (BioFlux®). If cultures were unavailable, fungal fruiting bodies were used to directly extract DNA using an OMEGA E.Z.N.A.® Forensic DNA Kit following the manufacturer's instructions.

ITS5 and ITS4, NS1 and NS4 (White et al. 1990), and LROR and LR5 (Vilgalys and Hester 1990) primers were used for the amplification of internal transcribed spacers (ITS), small subunit rDNA (SSU) and large subunit rDNA (LSU) respectively. Translation elongation factor 1- $\alpha$  gene region (TEF 1-alpha) and RNA polymerase II second largest subunit (RPB2) genes were amplified by using EF1-983 F and EF1-2218R (Rehner 2001), fRPB2-5f and fRPB2-7cr primers (Liu et al. 1999) respectively.

Polymerase chain reaction (PCR) amplification was carried out as follows: the final volume of the PCR reaction was 25  $\mu$ l, which contained 1  $\mu$ l of DNA template, 1  $\mu$ l of each forward and reverse primers, 12.5  $\mu$ l of 2 $\times$  Power Taq PCR Master Mix (mixture of *EasyTaq*<sup>TM</sup> DNA Polymerase, dNTPs, and optimized buffer, Beijing Bio Teke Corporation (Bio Teke), PR China) and 9.5  $\mu$ l Distilled-Deionized-Water. The PCR thermal cycle program of ITS, SSU, LSU and TEF 1-alpha genes amplifications were provided as: initially 94 °C for 3 min, followed by 35 cycles of denaturation at 94 °C for 30 s, annealing at 55 °C for 50 s, elongation at 72 °C for 1 min, and final extension at 72 °C for 10 min. The PCR thermal cycle program for the RPB2 gene was provided as initially 95 °C for 5 min, followed by 35 cycles of denaturation at 95 °C for 1 min, annealing at 52 °C for 2 min, elongation at 72 °C for 90 s, and final extension at 72 °C for 10 min. The quality of PCR products were checked by 1 % Biowest agarose gel electrophoresis. Amplified PCR fragments were sequenced at Shanghai Majorbio Bio-Pharm Technology Co., Ltd. and BGI Tech Solutions Co., Ltd. (BGI-Tech), P.R. China. Generated new sequences of ITS, LSU, SSU, RPB2 and TEF1 regions are deposited in GenBank.

## Phylogenetic analyses

Based on blast searches in GenBank, using LSU or ITS sequence data, separate phylogenetic analyses were carried out to determine the placements of each fungal group. Additional sequences were downloaded from GenBank, based on blast search and recent publications.

Single gene sequence alignments were generated with MAFFT v. 7.215 (Kato and Standley 2013; <http://mafft.cbrc.jp/alignment/server/index.html>) and edited manually when necessary in MEGA6 version 6.0 (Tamura et al. 2013) or BioEdit v. 7.0 (Hall 2004).

Maximum-likelihood (ML) analyses including 1000 bootstrap replicates were run using RAxMLGUI v.1.0. (Stamatakis 2006; Silvestro and Michalak 2011). Alignments in PHYLIP format were exchanged and loaded from the website (<http://sing.ei.uvigo.es/ALTER/>). The online tool Findmodel (<http://www.hiv.lanl.gov/content/sequence/findmodel/findmodel.html>) was used to determine the best nucleotide substitution model for each partition data.

Maximum-parsimony (MP) analyses were carried out in PAUP v. 4.0b10 (Swofford 2002) with 1000 replications. Maxtrees were set to 1000, branches of zero length were collapsed, and all multiple equally most parsimonious trees were saved. The robustness of the most parsimonious trees was evaluated from 1 000 bootstrap replications (Phillips et al. 2013).

Bayesian analyses were performed by using MrBayes v. 3.0b4 (Ronquist and Huelsenbeck 2003). The model of evolution was performed by using MrModeltest v. 2.2 (Nylander 2004). Posterior probabilities (PP) (Rannala and Yang 1996; Zhaxybayeva and Gogarten 2002) were determined by Markov Chain Monte Carlo sampling (MCMC) in MrBayes v. 3.0b4 (Huelsenbeck and Ronquist 2001). Six simultaneous Markov chains were run for 1,000,000 generations and trees were sampled every 100th generation. The burn-in was set to 0.25, and the run was automatically stopped when the average standard deviation of split frequencies reached below 0.01 (Maharachchikumbura et al. 2015).

Trees were visualized with TreeView (Page 1996) or FigTree v. 1.4.0 (<http://tree.bio.ed.ac.uk/software/figtree/>), and additionally layouts were done with Adobe Illustrator CS v. 5. Maximum-likelihood bootstrap values (MLBP) and Maximum-parsimony bootstrap values (MPBP) equal or greater than 50 % are given for each tree. Bayesian posterior probabilities (BYPP) > 0.90 are indicated as thickened lines.

## Results

### Phylogeny

Due to the diversity of taxa found, we made the individual trees at the family or order levels. Phylogenetic trees for each data set

are provided below. Phylogenetically similar taxa are discussed. All new sequence data are deposited in GenBank and GenBank accession numbers are provided in Tables 1.

### Taxonomy

***Dothideomycetes*** O.E. Erikss. & Winka, Myconet 1(1): 5 (1997)

*Dothideomycetes* is one of the largest classes within Ascomycota with an estimated 19,000 species (Kirk et al. 2008; Schoch et al. 2009). Hyde et al. (2013) re-examined 105 family types of *Dothideomycetes* and provided multi-gene analyses for 64 families. Wijayawardene et al. (2014) listed all genera accepted in *Dothideomycetes* (belonging to 23 orders and 110 families), including pleomorphic and nonpleomorphic genera. Rossman et al. (2015) provided recommended names for pleomorphic genera in *Dothideomycetes*.

***Botryosphaeriales*** C.L. Schoch, et al., Mycologia 98(6): 1050 (2007) [2006]

The order *Botryosphaeriales* was introduced by Schoch et al. (2006) with a single family *Botryosphaeriaceae*. Liu et al. (2012a) provided a phylogenetic analysis of *Botryosphaeriales* and included the families *Botryosphaeriaceae* and *Phyllostictaceae*. Hyde et al. (2013) stated that the order *Botryosphaeriales* comprises three families: *Botryosphaeriaceae*, *Planistromellaceae* and *Phyllostictaceae*. Subsequently, Slippers et al. (2013) introduced the families *Aplosporellaceae*, *Melanopsaceae* and *Saccharataceae*. Wijayawardene et al. (2014) recognized six families, viz. *Aplosporellaceae*, *Botryosphaeriaceae*, *Melanopsaceae*, *Phyllostictaceae*, *Planistromellaceae* and *Saccharataceae*, in this order.

***Botryosphaeriaceae*** Theiss. & Syd. [as ‘*Botryosphaeriaceae*’], Anns mycol. 16(1/2): 16 (1918)

The family *Botryosphaeriaceae* was introduced by Theissen and Sydow (1918). This family has been well circumscribed by several authors (Crous et al. 2006b; Phillips et al. 2008, 2013; Liu et al. 2012a; Slippers et al. 2013; Hyde et al. 2014; Linaldeddu et al. 2015). Liu et al. (2012a) accepted 29 genera in this family, based on morphology. Phillips et al. (2013) considered only the genera known from culture and based the number of accepted genera on morphology and phylogeny. Thus 17 genera were recognized by Phillips et al. (2013) using morphological features and combined phylogenetic analysis of SSU, ITS, LSU, EF1- $\alpha$  and  $\beta$ -tubulin sequence data, and further keys to the genera and species were provided. Crous et al. (2015) introduced *Eutiarospora*, *Marasasiomyces*, *Mucoharknessia* and *Sakireeta*, which Wijayawardene et al. (2016) accepted in their phylogenetic analyses.

*Neodeightonia* C. Booth, in Punithalingam, Mycol. Pap. 119: 17 (1970) [1969]

The genus *Neodeightonia* was introduced by Booth (Punithalingam 1969), and placed in the family *Botryosphaeriaceae*. This genus comprises four species, viz. *N. microspora* (new species in this study), *N. palmicola*, *N. phoenicum* and *N. subglobosa* based on morphological and phylogenetic analyses (Phillips et al. 2008, 2013; Liu et al. 2012).

*Neodeightonia microspora* D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum number*: IF552019; *Facesoffungi number*: FoF 01965; Figs. 1 and 2

*Etymology*: In reference to the small ascospores.

*Holotype*: MFLU 15–1201

*Saprobic* on dead bamboo culms. **Sexual morph**: *Ascstromata* 100–150  $\mu\text{m}$  high, 95–150  $\mu\text{m}$  diam., dark brown to black, immersed under epidermis to erumpent, solitary, visible as minute black dots or papillae on host tissue, uni-loculate, locules globose to subglobose or fused, coriaceous, vertically orientated to the host surface, with a central ostiole. *Peridium* 15–20  $\mu\text{m}$  wide, comprising several layers of cells, with relatively thick brown to black walls of *textura angularis*, broader at the base. *Hamathecium* comprising only asci. *Asci* 70–110  $\times$  14–20  $\mu\text{m}$  ( $\bar{x}$  = 90.8  $\times$  18.5  $\mu\text{m}$ ,  $n$  = 20), 8-spored, bitunicate, fissitunicate, clavate to cylindro-clavate, with a 25–45  $\mu\text{m}$  long pedicel, apically rounded, with a well-developed ocular chamber. *Ascospores* 10–12  $\times$  4.5–6  $\mu\text{m}$  ( $\bar{x}$  = 10.5  $\times$  5.4  $\mu\text{m}$ ,  $n$  = 30), 2–3-seriate, hyaline, aseptate, obovoid, usually wider at the apex, thick-walled, surrounded by a distinctive, structured, mucilaginous sheath. **Asexual morph**: Undetermined.

*Culture characters*: Ascospores germinating on PDA within 24 h and germ tubes produced from apex. Colonies growing fast on PDA, reaching 5 cm in 1 week at 28 °C, under 12 h light/12 h dark, effuse, velvety to hairy, circular, initially white, becoming dark brown to black after 1 week.

*Material examined*: THAILAND, Payae, Bah Huay Kawkhand, Amphet, on dead culms of bamboo, 31 October 2011, Dong-Qin Dai DDQ00228 (MFLU 15–1201, **holotype**); *ibid.* (KUN HKAS88710, **isotype**), living culture, MFLUCC 11–0483, KUMCC; Chiang Rai, Doi Pui, on dead culms of bamboo, 1 September 2011, Dong-Qin Dai DDQ00111 (MFLU 15–1314, **paratype**), living cultures, MFLUCC 11–0504.

*Notes*: *Neodeightonia microspora* is characterized by clavate to cylindro-clavate asci with long pedicels and small obovoid ascospores, with a thick mucilaginous sheath. The new taxon differs from *N. subglobosa* and *N. palmicola* in its smaller obovoid ascospores (10–12  $\times$  4.5–6  $\mu\text{m}$ ) as compared to *N. subglobosa* (ellipsoidal to fusiform, 21–26  $\times$  7.5–9.5  $\mu\text{m}$ ) and *N. palmicola* (ellipsoidal to fusiform, 23–31.5  $\times$  8.5–12.5  $\mu\text{m}$ ). It also differs in having asci with a long pedicel. In the phylogenetic analyses (Fig. 1) *N. microspora* separates from other *Neodeightonia* species with high

bootstrap support (94/96 % MPBP/MLBP, BYPP value greater than 0.90).

*Neodeightonia subglobosa* C. Booth, in Punithalingam, Mycol. Pap. 119: 19 (1970) [1969]

*Facesoffungi number*: FoF 01966; Figs. 3 and 4

*Synonymy*: See index Fungorum

*Saprobic* on dead bamboo culms. **Sexual morphs**: *Ascstromata* 160–220  $\mu\text{m}$  high, 250–370  $\mu\text{m}$  diam., immersed under epidermis to erumpent, gregarious, visible as minute black dots or papilla on host tissue, dark brown to black, uniloculate, globose to subglobose, coriaceous, with a centrally located ostiole, papillate. *Peridium* 17–40  $\mu\text{m}$  wide, comprising several layers, with outer layers composed of relatively thick brown to black-walled cells of *textura angularis*, inner layers composed of light brown to hyaline cells of *textura angularis*, poorly developed at the base. *Hamathecium* of hyaline, septate, up to 2–4  $\mu\text{m}$  wide pseudoparaphyses. *Asci* 110–150  $\times$  16–20  $\mu\text{m}$  ( $\bar{x}$  = 124.8  $\times$  19.5  $\mu\text{m}$ ,  $n$  = 20), 8-spored, bitunicate, fissitunicate, cylindro-clavate, pedicellate, apically rounded with well-developed ocular chamber. *Ascospores* 17–21  $\times$  8–10.5  $\mu\text{m}$  ( $\bar{x}$  = 20.5  $\times$  8.4  $\mu\text{m}$ ,  $n$  = 20), 2–3-seriate, hyaline, aseptate, obovoid, usually wider at the apex, thick-walled, surrounded by a distinctive structured mucilaginous sheath. **Asexual morph**: Fertile on pine needles on WA after 1 month. *Conidiomata* 150–200  $\mu\text{m}$  diam., pycnidial, immersed to superficial, solitary to gregarious, globose to subglobose, black, coriaceous. *Conidiogenous cells* 5–12.5  $\times$  2–3  $\mu\text{m}$  ( $\bar{x}$  = 9.8  $\times$  2.6  $\mu\text{m}$ ,  $n$  = 20), holoblastic, cylindrical, hyaline, straight to curved, smooth-walled. *Conidia* 11–13.5  $\times$  8–10.5  $\mu\text{m}$  ( $\bar{x}$  = 12.6  $\times$  9.3  $\mu\text{m}$ ,  $n$  = 20), globose to subglobose, hyaline, aseptate, smooth-walled, guttulate.

*Culture characters*: Ascospores germinating on PDA within 24 h and germ tubes produced from apex. Colonies growing fast on PDA, reaching 5 cm in 1 week at 28 °C, under 12 h light/12 h dark, effuse, velvety to hairy, circular, white in first week, dark brown to black after 1 week from below and above.

*Material examined*: THAILAND, Chiang Rai, Mae Fah Luang University, on dead culms of bamboo, 1 August 2011, Dong-Qin Dai DDQ00087 (MFLU 15–1189, **reference specimen designated here**); *ibid.* (KUN, HKAS88699, duplicates of reference specimen), living culture, MFLUCC 11–0482, MFLUCC 11–0607; asexual morph, 19 June 2011, Dong-Qin Dai DDQ00041, living cultures, MFLUCC 11–0388.

*Notes*: *Neodeightonia subglobosa* was originally collected from *Bambusa arundinacea* Willd. (synonym of *Bambusa bambos* (L.) Voss) in Africa, and introduced by Punithalingam (1969). Phillips et al. (2008) isolated this species, which causes keratomycosis of the eye, from a human host in an unknown location, illustrated only the asexual morph, and provided a sequence based on the culture CBS

**Table 1** Newly sequence data with GenBank accession numbers in this study are provided

Organism	Strain	Collection_date	Country	Host	Specimen_voucher	Type	GenBank accession numbers				
							LSU	SSU	ITS	RPB2	RRR
<i>Neodeigthonia microspora</i>	MFLUCC 11-0483	31-Oct-2011	Thailand	bamboo	MFLU 15-1201	holotype	KU863099	KU872104	KU940110	/	/
<i>Neodeigthonia microspora</i>	MFLUCC 11-0504	1-Sep-2011	Thailand	bamboo	MFLU 15-1314		KU863100	KU872105	KU940111	/	/
<i>Neodeigthonia subglobosa</i>	MFLUCC 11-0482	1-Aug-2011	Thailand	bamboo	MFLU 15-1189		KU863101	KU872106	KU940112	/	KU940184
<i>Neodeigthonia subglobosa</i>	MFLUCC 11-0607	1-Aug-2011	Thailand	bamboo	MFLU 15-1189		KU863102	KU872107	KU940113	/	KU940185
<i>Neodeigthonia subglobosa</i>	MFLUCC 11-0388	19-Jun-2011	Thailand	bamboo	/		KU863103	KU872108	KU940114	/	KU940186
<i>Mendogtia macrostroma</i>	/	5-Oct-2012	Thailand	bamboo	MFLU 13-0642	holotype	KU863104	KU872109	KU940115	KU940162	KU940187
<i>Bambusicola didymospora</i>	MFLUCC 10-0557	10-Mar-2010	Thailand	bamboo	MFLU 11-0149	holotype	KU863105	KU872110	KU940116	KU940163	KU940188
<i>Bambusicola didymospora</i>	MFLUCC 15-0189	5-Dec-2014	Thailand	bamboo	MFLU 16-0882		KU863106	KU872111	KU940117	KU940164	KU940189
<i>Bambusicola pustulata</i>	MFLUCC 15-0190	5-Dec-2014	Thailand	bamboo	MFLU 15-1216	holotype	KU863107	KU872112	KU940118	KU940165	KU940190
<i>Bambusicola thailandica</i>	MFLUCC 11-0147	13-Aug-2010	Thailand	bamboo	MFLU 11-0183	holotype	KU863108	KU872113	KU940119	KU940166	KU940191
<i>Bambusicola triseptatispora</i>	MFLUCC 11-0166	13-Apr-2011	Thailand	bamboo	MFLU 11-0202	holotype	KU863109	/	KU940120	KU940167	/
<i>Bambusicola splendida</i>	MFLUCC 11-0611	1-Aug-2011	Thailand	bamboo	/		KU863110	KU872114	KU940121	KU940168	/
<i>Bambusicola massarina</i>	MFLUCC 11-0135	16-Jun-2010	Thailand	bamboo	MFLU 11-0171		KU863111	KU872115	KU940122	KU940169	KU940192
<i>Occlitibambusa bambusae</i>	MFLUCC 13-0855	9-Jul-2013	Thailand	bamboo	MFLU 15-1212	holotype	KU863112	KU872116	KU940123	KU940170	KU940193
<i>Occlitibambusa bambusae</i>	MFLUCC 11-0394	14-May-2011	Thailand	bamboo	MFLU 16-0880		KU863113	KU872117	KU940124	KU940171	KU940194
<i>Occlitibambusa fusispora</i>	MFLUCC 11-0127	29-Jun-2010	Thailand	bamboo	MFLU 11-0163	holotype	KU863114	/	KU940125	KU940172	KU940195
<i>Occlitibambusa pustula</i>	MFLUCC 11-0502	19-Jul-2011	Thailand	bamboo	MFLU 15-1185	holotype	KU863115	KU872118	KU940126	/	/
<i>Seriascooma didymospora</i>	MFLUCC 11-0179	25-Oct-2010	Thailand	bamboo	MFLU 11-0215	holotype	KU863116	KU872119	KU940127	KU940173	KU940196
<i>Seriascooma didymospora</i>	MFLUCC 11-0194	13-Jan-2011	Thailand	bamboo	MFLU 11-0230		KU863117	KU872120	KU940128	KU940174	KU940197
<i>Rousoella mukdahanensis</i>	MFLUCC 11-0201	13-Apr-2011	Thailand	bamboo	MFLU 11-0237	holotype	KU863118	KU872121	KU940129	/	/
<i>Rousoella neopustulans</i>	MFLUCC 12-0003	13-Apr-2011	Thailand	bamboo	MFLU 11-0241		KU863119	KU872122	KU940130	/	/
		9-Jul-2013	Thailand	bamboo	MFLU 15-1209	holotype	KU863120	KU872123	KU940131	/	KU940198

Table 1 (continued)

Organism	Strain	Collection_date	Country	Host	Specimen_voucher	Type	GenBank accession numbers				
							LSU	SSU	ITS	RPB2	RRR
<i>Roussoella pseudohysterioides</i>	MFLUCC 13-0852	9-Jul-2013	Thailand	bamboo	MFLU 15-1211	holotype	KU863121	KU872124	KU940132	/	KU940199
<i>Roussoella tuberculata</i>	MFLUCC 11-0854	13-Aug-2010	Thailand	bamboo	MFLU 11-0185	holotype	/	KU872125	/	/	/
<i>Roussoella siamensis</i>	MFLUCC 15-0149	6-Oct-2014	Thailand	bamboo	MFLU 15-1215	holotype	/	/	KU940133	/	/
<i>Tubeufia longiseta</i>	MFLUCC 12-0846	31-Jul-2012	Thailand	bamboo	MFLU 15-1203	holotype	KU863122	/	KU940134	/	/
<i>Leptosporella bambusae</i>	MFLUCC 11-0395	14-May-2011	Thailand	bamboo	MFLU 15-1180	holotype	KU863123	KU872126	KU940135	/	KU940200
<i>Myrothecium thailandicum</i>	MFLUCC 11-0506	26-May-2011	Thailand	bamboo	MFLU 15-1181	holotype	KU863124	KU872127	KU940136	KU940175	KU940201
<i>Myrothecium changmatense</i>	MFLUCC 11-0392	14-May-2011	Thailand	bamboo	MFLU 14-0823	holotype	/	KU872128	/	/	/
<i>Myrothecium cylindrosporium</i>	MFLUCC 11-0632	28-Oct-2011	Thailand	bamboo	MFLU 15-1197	holotype	KU863125	KU872129	KU940137	/	/
<i>Myrothecium uttaradiense</i>	MFLUCC 11-0391	12-May-2011	Thailand	bamboo	MFLU 15-1179	holotype	KU863126	KU872130	KU940138	/	/
<i>Nectria pseudotrichia</i>	MFLUCC 12-0845	31-Jul-2012	Thailand	bamboo	MFLU 15-1202	holotype	KU863127	KU872131	KU940139	KU940176	/
<i>Cercophora thailandica</i>	MFLUCC 11-0629	6-Oct-2011	Thailand	bamboo	MFLU 15-1196	holotype	KU863128	KU872132	KU940140	/	KU940202
<i>Phaeoacremonium sphinctrothorum</i>	MFLUCC 11-0481	19-Jul-2011	Thailand	bamboo	MFLU 15-1184	holotype	KU863129	/	KU940141	/	/
<i>Arthrinium longistromum</i>	MFLUCC 11-0479	17-Jul-2011	Thailand	bamboo	/		KU863130	/	KU940142	/	/
<i>Arthrinium longistromum</i>	MFLUCC 15-0203	4-Oct-2014	Thailand	bamboo	MFLU 15-1227	holotype	KU863131	KU872133	KU940143	/	/
<i>Arthrinium rasilkravindrae</i>	MFLUCC 11-0616	1-Oct-2011	Thailand	bamboo	/		KU863132	KU872134	KU940144	/	/
<i>Arthrinium rasilkravindrae</i>	MFLUCC 15-0202	4-Oct-2014	Thailand	bamboo	MFLU 15-1226	holotype	KU863133	/	KU940145	/	/
<i>Arthrinium thailandicum</i>	MFLUCC 15-0199	4-Oct-2014	Thailand	bamboo	MFLU 15-1224	holotype	KU863134	/	KU940146	/	/
<i>Arthrinium thailandicum</i>	MFLUCC 15-0002	7-Jul-2014	China	bamboo	MFLU 15-0382	holotype	KU863135	KU872135	KU940147	KU940177	/
<i>Arthrinium yunnanum</i>	DDQ00281	7-Jul-2014	China	bamboo	/		KU863136	KU872136	KU940148	KU940178	/
<i>Eutypa linearis</i>	MFLUCC 15-0198	6-Dec-2014	Thailand	bamboo	MFLU 15-1223	holotype	KU863137	KU872137	KU940149	/	KU940203
<i>Eutypa linearis</i>		19-Jul-2011	Thailand	bamboo	MFLU 15-1186	holotype	KU863138	KU872138	KU940150	/	KU940204





448.91. Phillips et al. (2013) however, indicated CBS 448.91 as ex-type without any justification. Liu et al. (2012a) illustrated the sexual morph of this species based on a new collection. In this study, new isolates and specimens were obtained collected on bamboo in Thailand with both sexual and asexual morphs. Thus a reference specimen is designated here, and the species is re-described and illustrated species, due to no available strain linked to Punithalingam (1969). Punithalingam (1969) stated that *N. subglobosa* has brown and 1-septate ascospores. However, these characters were not observed in this study. Phylogenetic analyses show three new strains in the same branch, with another two strains of *N. subglobosa* (MFLUCC 11–0163 and CBS 448.91), clustering with high bootstrap support (98/94 % MPBP/MLBP) (Fig. 1).

#### *Myriangiales* Starbäck 1899

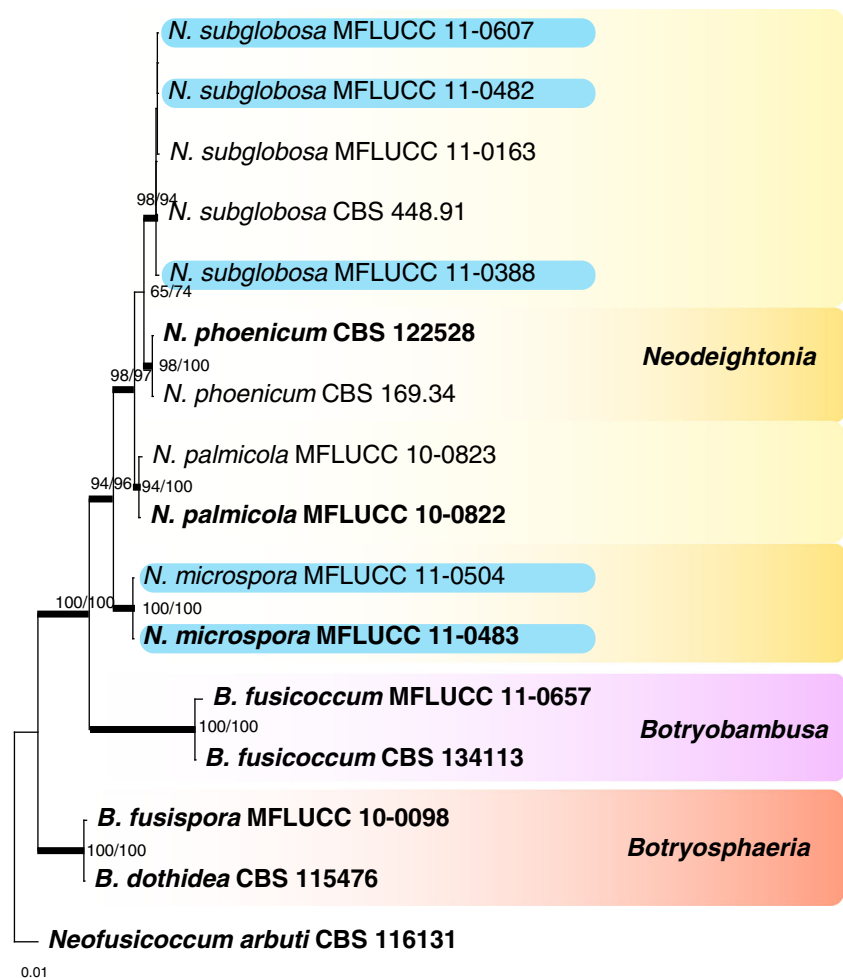
The order *Myriangiales* was introduced by Starbäck (1899) based on a genus producing crustose, loculate ascostromata, with muriform ascospores (Hyde et al. 2013). Three families, *Cookellaceae*, *Elsinoaceae* and *Myriangiaceae*, were included in *Myriangiales* by Kirk et al. (2008). Based on phylogenetic analyses, Lumbsch and Huhndorf (2010), Hyde et al.

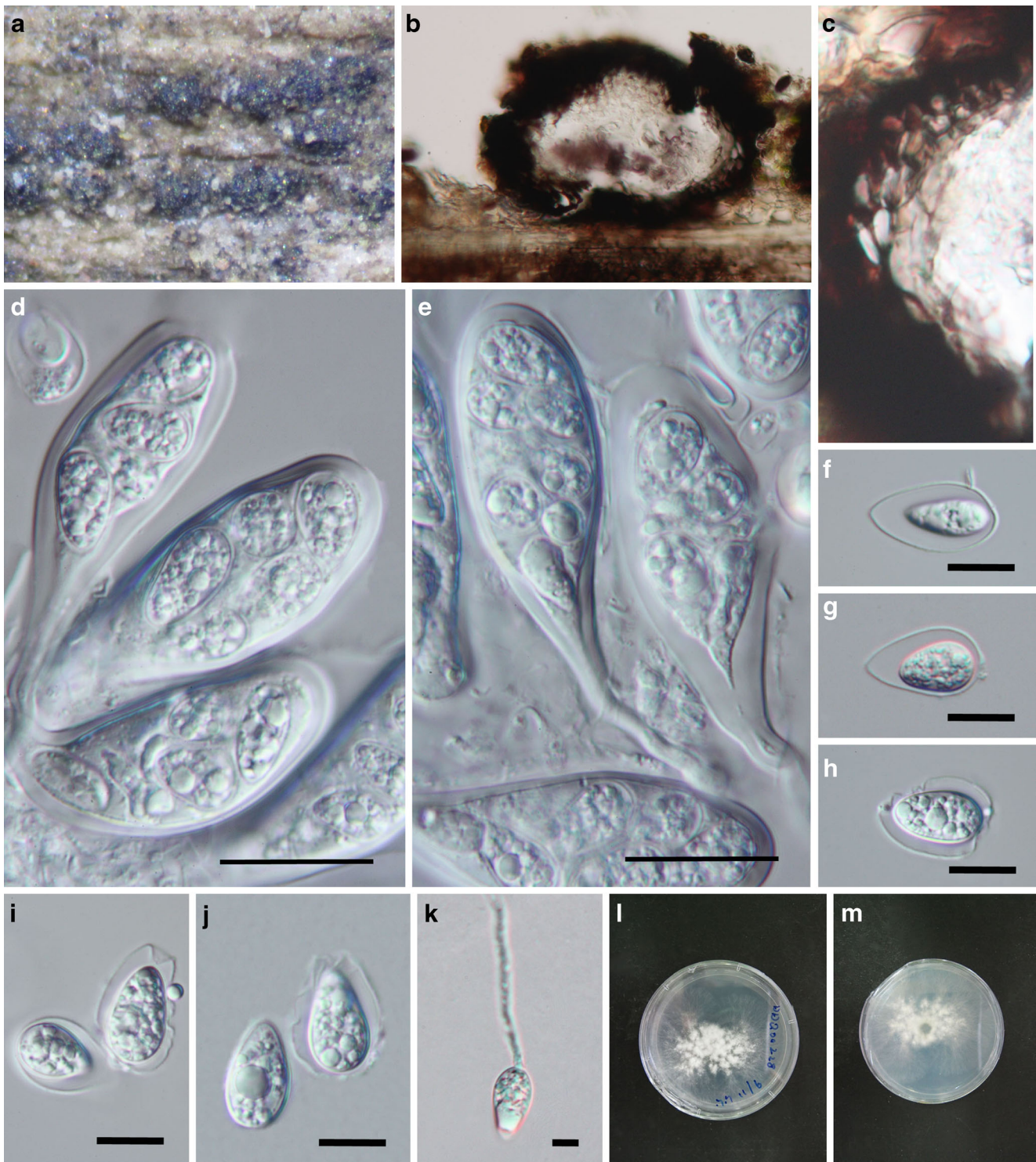
(2013), Wijayawardene et al. (2014) accepted only *Elsinoaceae* and *Myriangiaceae* in this order.

*Myriangiaceae* Nyl., Mém. Soc. Sci. nat. Cherbourg 2: 9 (1854)

The family *Myriangiaceae* was introduced by Nylander (1854). Von Arx and Müller (1975) placed 16 genera in the family, but Lumbsch and Huhndorf (2010) included only four genera (*Anhelliella*, *Diplothea*, *Eurytheca*, *Myriangium*). Dissanayake et al. (2014) re-examined the generic type specimens and accepted ten genera, providing descriptions and discussion on the generic types of *Anhelliella*, *Ascostratum*, *Butleria*, *Dictyocyclus*, *Diplothea*, *Eurytheca*, *Hemimyriangium*, *Micularia*, *Myriangium* and *Zukaliopsis*. However, only *Anhelliella* and *Myriangium* have molecular data (Pinho et al. 2012; Hyde et al. 2013; Wijayawardene et al. 2014). The family *Myriangiaceae* is characterized by having only one ascus in each locule, and superficial, erumpent, dark ascostromata. The asci are clavate to subglobose or globose and ascospores smooth to verruculose and muriform, occasionally with only transverse septa (Miller 1938; Hyde et al. 2013; Dissanayake et al. 2014). The formations of the asci of

**Fig. 1** Maximum likelihood phylogenetic tree (lnL = -2178.664062) generated by RAxML (GTR+G model), based on combined SSU, ITS and LSU sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes and branches with Bayesian posterior probabilities greater than 0.90 are in bold. The original isolate numbers codes are noted after the species names. The tree is rooted to *Neofusicoccum arbuti* (CBS 116131). Ex-type strains are in bold. Newly generated sequences are highlighted with a blue background



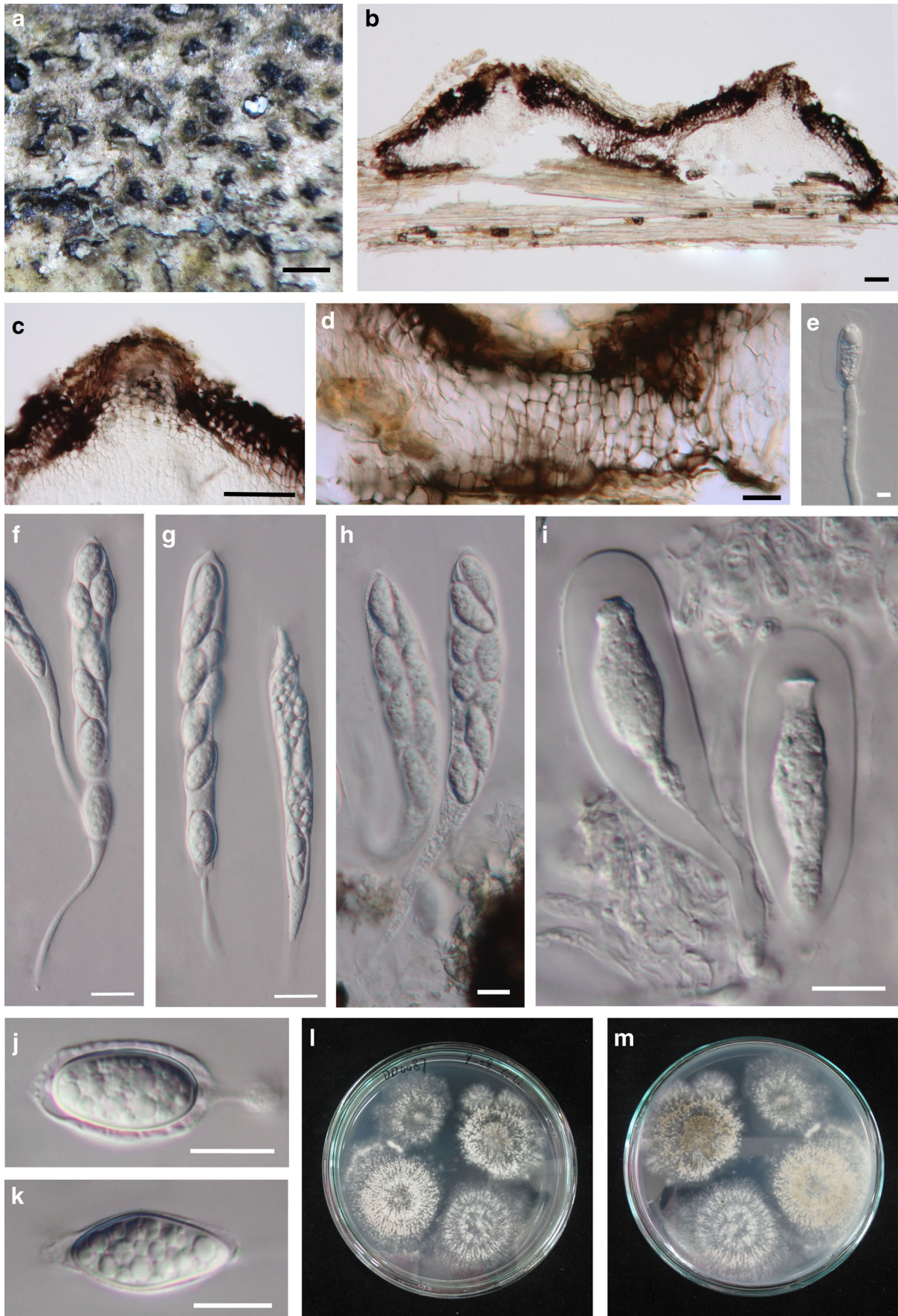


**Fig. 2** *Neodeightonia microspora* (MFLU 15–1201, **holotype**). **a** Appearance of ascomata on bamboo host. **b** Vertical section of ascoma. **c** Peridium. **d, e** Asci with eight ascospores. **f–j** Ascospores surrounded

by mucilaginous sheath. **k** Germinating ascospore. **l, m** Cultures on PDA. Scale bars: **d, e** = 20  $\mu$ m, **f–j** = 10  $\mu$ m

this family are two types. Type I: The asci are scattered irregularly over the whole ascostromata. Type II: The asci are localized in definite regions, usually forming at the base of the ascostromata, which are differentiated into sterile and fertile

regions (Miller 1940; Dissanayake et al. 2014). Species of *Myriangiaceae* are usually epiphytic on bark, leaves and branches of various plants (Hyde et al. 2013; Dissanayake et al. 2014).



◀ **Fig. 3** *Neodeightonia subglobosa* (MFLU 15–1189, reference specimen). **a** Appearance of ascomata on bamboo. **b** Vertical section of ascoma. **c** Ostiole of ascoma. **d** Peridium. **e** Germinating ascospores. **f–g** Asci. **i** Immature asci. **j, k** Ascospores. **l, m** Culture on PDA. Scale bars: a = 200  $\mu\text{m}$ , b, c = 50  $\mu\text{m}$ , d–k = 10  $\mu\text{m}$

*Mendogia* Racib., Parasit. Alg. Pilze Java's (Jakarta) 3: 31 (1900)

*Epiphytic* on living bamboo culms or palms. **Sexual morph:** *Ascostromata* small to large, solitary to gregarious, superficial, round to irregular, black, carbonaceous, flattened, with central area slightly raised, and peripheral wall of stromata cracking, pseudoparenchymatous tissue and asci growing under stroma, revealing the dark brown inner issue, with irregular edge, rugose to rugulose at the surface, loculate, easily removable from the host, with asci separated by paraphysoid-like filaments or pseudoparenchymatous cells. *Wall of ascostromata* consisting of two layers, outer layer composed of black, thin carbonaceous tissue, easily breaking; inner layer composed of large, hyaline to pale-brown to dark brown pseudoparenchymatous cells of *textura angularis*. *Hamathecium* with asci or paraphysoid-like filaments. *Asci* 8-spored, thick-walled, bitunicate, fissionate, subglobose to clavate, or cylindrical, with rounded pedicel, with a distinct ocular chamber. *Ascospores* irregularly arranged, ellipsoidal, muriform, with rounded ends, hyaline, smooth-walled. **Asexual morph:** Undetermined.

*Type species: Mendogia bambusina* Racib., Parasit. Alg. Pilze Java's (Jakarta) 3: 31 (1900)

*Notes:* *Mendogia* was introduced by Raciborski (1900) based on a single species on a bamboo host, collected in Java, Indonesia. Two more species *Mendogia manaosensis* (Henn.) Theiss. & Syd. and *Mendogia philippinensis* (Syd. & P. Syd.) Arx & E. Müll. observed on palms were named under this genus (Hennings 1904; von Arx and Müller 1975). *Mendogia* is characterized by small to large, black, rather flattened ascostromata, and subglobose to clavate asci, producing eight muriform ascospores. This genus was earlier placed in family *Schizothyriaceae* by von Arx and Müller (1975). Without molecular data, Hyde et al. (2013) and Wijayawardene et al. (2014) followed this placement.

In this study, we re-examined the type species *Mendogia bambusina* Racib. and introduce a new species, *Mendogia macrostroma*, from a bamboo host. Multi-gene (SSU, LSU and TEF1) phylogenetic analyses indicate that *Mendogia* belongs to order *Myriangiales* (97/97 % MPBP/MLBP) in the family *Myriangiaceae* (94/97 % MPBP/MLBP) (Fig. 5). This genus can phylogenetically be separated from similar studied genera (*Anhellia* and *Myriangium*) with high bootstrap support (99/100 % MPBP/MLBP) (Fig. 5).

*Mendogia bambusina* Racib., Parasit. Alg. Pilze Java's (Jakarta) 3: 31 (1900)

*Facesoffungi* number: FoF 01967; Fig. 6

*Epiphytic* on living bamboo culms. **Sexual morph:** *Ascostromata* 0.98–1.85 mm diam., scattered, solitary to gregarious, superficial on host surface, black, flattened, circular to round, hemispherical or shield-shape, rugulose, sunken near the edge, easily removed from the host, loculate, glabrous, walls rough, carbonaceous at the outer surface, with centre sterile region and fertile region located near the margin. *Fertile region* locule-like, 100–210  $\mu\text{m}$  high, 260–420  $\mu\text{m}$  diam. in vertical section, forming as circular ring near the edge of ascostroma, immersed in ascostroma, raised, subglobose to hemispherical, with asci separated by paraphysoid-like filaments, discharging asci and ascospores via cracking of the outer walls, with slit-like opening. *Peridium* 15–60  $\mu\text{m}$  wide, thick-walled, of unequal thickness, thick at the rim, slightly thin at the base, composed of two layers, of dark brown to black cells, outer layers comprising thick, black, melanized cells, carbonaceous, arranged in a *textura angularis*, inner layers comprising thick, brown to dark brown, pseudoparenchymatous cells, arranged in *textura angularis*. *Hamathecium* composed of dense, 1.5–4  $\mu\text{m}$  wide, paraphysoid-like filaments, septate, constricted at the septum, anastomosing among the asci. *Asci* arranged in type II, (55–)60–75(–80)(–90)  $\times$  (17–)18–22(–25)  $\mu\text{m}$  ( $\bar{x}$  = 71.6  $\times$  20.5  $\mu\text{m}$ ,  $n$  = 25), 8-spored, bitunicate, fissionate, cylindrical-clavate to clavate, short pedicellate, apically rounded with well-developed ocular chamber, thickened at the apex. *Ascospores* (13.5–)17–20(–25)  $\times$  (5–)6–8  $\mu\text{m}$  ( $\bar{x}$  = 19.4  $\times$  6.7  $\mu\text{m}$ ,  $n$  = 30), overlapping, uni- to tri-seriate, hyaline, ellipsoidal to clavate, muriform, with 3–5 transverse septa and 1–2 longitudinal septa, constricted at the septum, smooth-walled. **Asexual morph:** Undetermined.

*Material examined:* INDONESIA, Jawa Barat (West) District, Java, Buitenzorg, on bamboo, 1907, G. v. Höhnel (S, F48345).

*Notes:* *Mendogia bambusina* differs from other *Mendogia* species in having asci arranged in a fertile region lying as concentric rings near the edge of ascostroma, with muriform ascospores, having 4–5 transverse septa and 1–2 longitudinal septa.

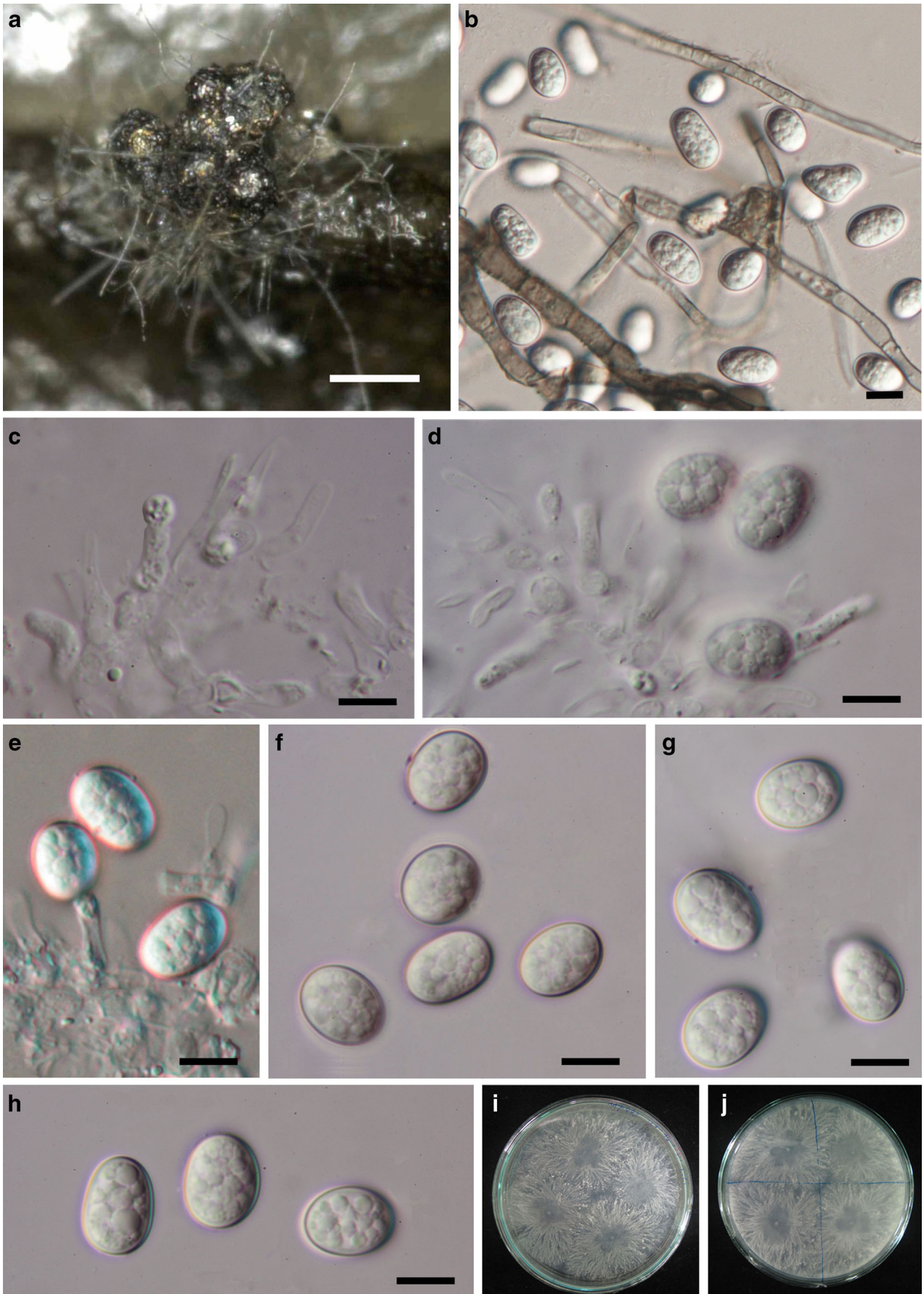
*Mendogia macrostroma* D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum* number: IF552020; *Facesoffungi* number: FoF 01968; Fig. 7

*Etymology:* In reference to its large stromata.

*Holotype:* MFLU 13–0642

*Epiphytic* on living bamboo culms. **Sexual morph:** *Ascostromata* 10–20  $\times$  5–15 mm, 130–200  $\mu\text{m}$  high, solitary, or gregarious up to 25  $\times$  20 mm, superficial, round, elliptical to irregular, black, coriaceous, flattened, with central area slightly raised, and peripheral wall of stromata cracking, revealing the dark brown inner issue, pseudoparenchymatous tissue and asci growing under stroma, with irregular edge, rugose to rugulose at surface, multi-loculate, easily removable from the host. *Surface cells of ascostromata* 20–30  $\mu\text{m}$  thick,

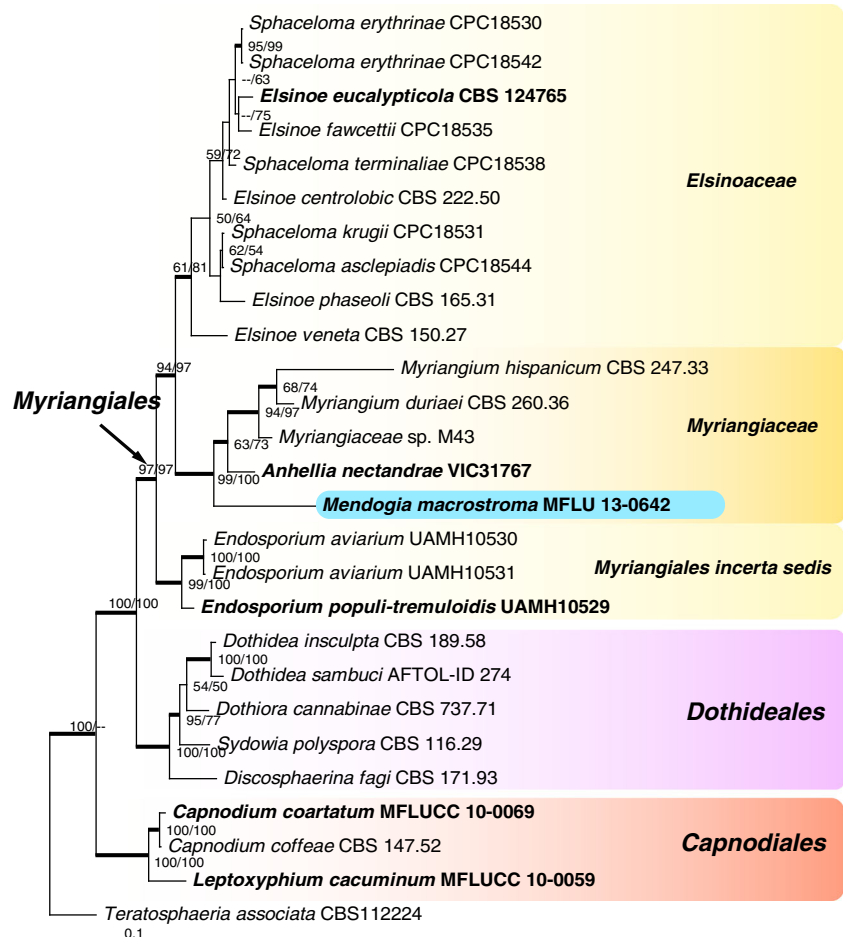


**Fig. 4** *Neodeightonia subglobosa* (MFLUCC 11–0388). **a** Appearance of conidiomata on pine needles. **b, f–h** Conidia. **c–e** Conidiogenous cells. **i, j** Cultures on PDA. Scale bars: a = 200  $\mu\text{m}$ , b–h = 10  $\mu\text{m}$

composed of black tissue, easily breaking. *Cells between asci* 100–150  $\mu\text{m}$  thick composed of large, hyaline, pseudoparenchymatous cells of *textura angularis*. *Hamathecium* with only asci; *pseudoparaphyses* not observed. *Asci* arranged in type II, 72–85  $\times$  28–33.5  $\mu\text{m}$  ( $\bar{x}$  = 76.6  $\times$  31.2  $\mu\text{m}$ ,  $n$  = 20), 8-spored, bitunicate, fissitunicate, subglobose to clavate, with rounded pedicel, with a distinct ocular chamber. *Ascospores* 20–27  $\times$  9–11  $\mu\text{m}$  ( $\bar{x}$  = 24.2  $\times$  10.1  $\mu\text{m}$ ,  $n$  = 20), irregularly arranged, hyaline, ellipsoidal, with rounded ends, muriform, with 4–6 transverse septa and 2–4 longitudinal septa, constricted at the septa, smooth-walled, narrow at basal end, occasionally narrow at both ends. **Asexual morph:** Undetermined.

**Material examined:** THAILAND, Chiang Rai, Khun Korn Waterfall, on living culms of bamboo, 5 October 2012, Dong-Qin Dai DDQ00255 (MFLU 13–0642, **holotype**); *Ibid.* (KUN HKAS88716, **isotype**); *Ibid.*, August 2014, R. Phookamsak RP0134 (KUN HKAS83874).

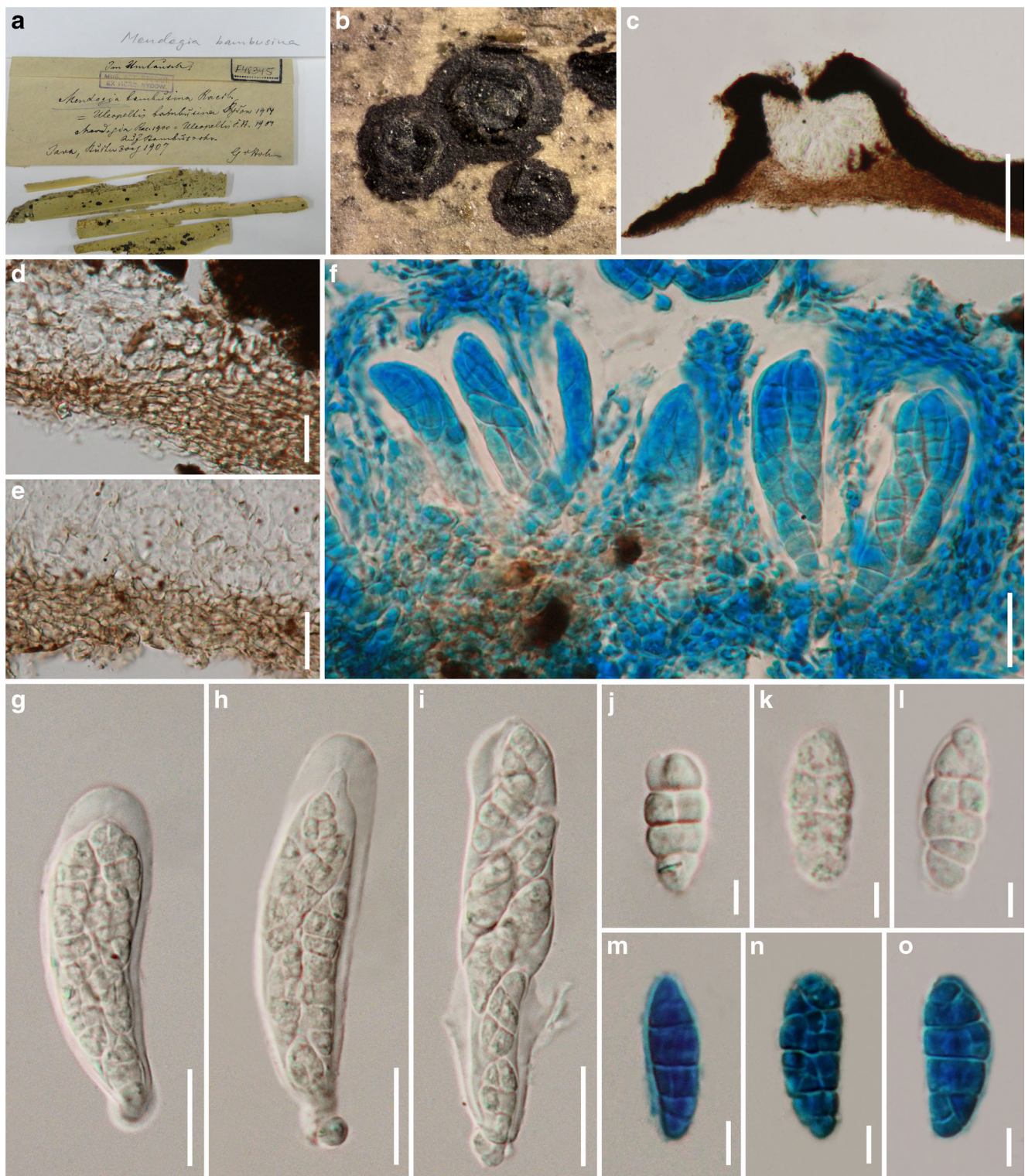
**Fig. 5** Maximum likelihood phylogenetic tree (lnL = –2981.957640) generated by RAxML (GTR+G model) based on combined SSU, LSU and TEF1 sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes and branches with Bayesian posterior probabilities greater than 0.90 are in bold. The original isolate or specimen numbers are noted after the species names. The tree is rooted to *Teratosphaeria associata* (CBS 112224). Ex-type strains are in bold. Newly generated sequence is highlighted with a blue background



**Notes:** *Mendogia macrostroma* differs morphologically from other species of *Mendogia* by its large ascostromata (10–20  $\times$  8–15 mm vs. less than 4 mm diam.) and wider asci (28–33.5  $\mu\text{m}$  vs. less than 25  $\mu\text{m}$ ) (Hennings 1904; Sydow and Sydow 1914a, 1917). The new taxon can be compared with *Myriangium haraeaeum* F.L. Tai & C.T. Wei (synonym: *Myriangium bambusae* Hara 1913) in having dark, flattened ascostromata and muriform ascospores (Tai and Wei 1933; Eriksson and Yue 1998). However, the new species differs in having larger ascostromata (10–20  $\times$  8–15 mm vs. 1–3 mm in diam.). Moreover, *Mendogia macrostroma* has subglobose to clavate asci, whereas *Myriangium haraeaeum* has cylindrical asci (Tai and Wei 1933).

#### **Pleosporales** Luttr. ex M.E. Barr 1987

*Pleosporales*, the largest order of *Dothideomycetes*, was introduced by Nitschke (1869), and later validated by Barr (1987). Luttrell (1975) included eight families in this order, Kirk et al. (2008) listed 23 families, Lumbsch and Huhndorf (2010) accepted 28 families and Zhang et al. (2012) included 25 families in the phylogenetic analysis. Forty-one families were placed in *Pleosporales* by Hyde et al. (2013), based on the



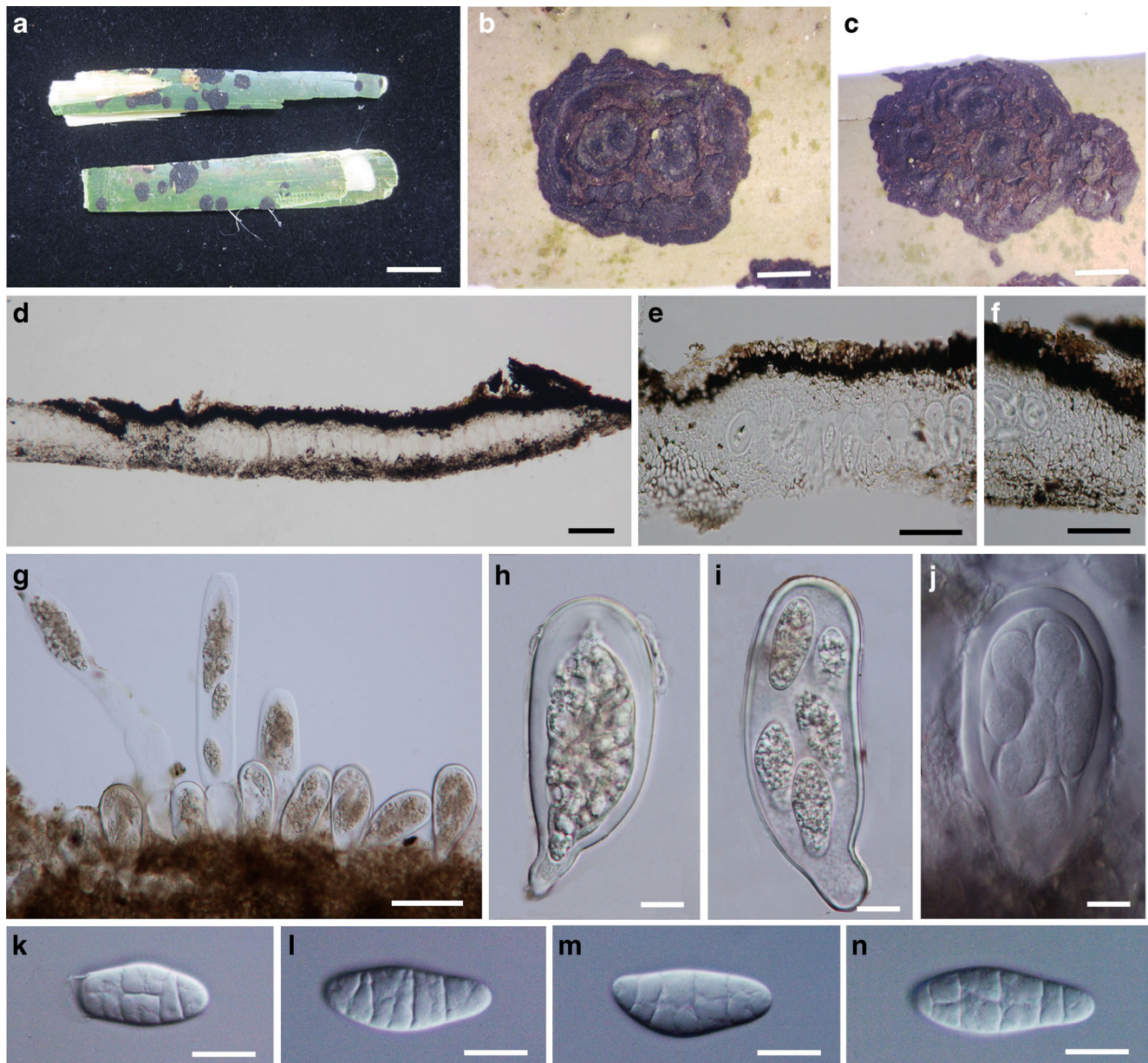
**Fig. 6** *Mendogia bambusina* (S F48345). **a** Herbarium materials. **b** Black ascostromata on bamboo host. **c** Section of ascostroma. **d, e** Peridium. **f** Asci separated by paraphysoid-like filaments. **g–i** Asci. **j–o**

Ascospores (**m–o**: Ascospores stained in cotton blue). Scale bars: **c** = 100  $\mu$ m, **d–i** = 20  $\mu$ m, **j–o** = 5  $\mu$ m

multi-gene analyses and morphological study of family types. However, Wijayawardene et al. (2014) accepted 39 families. Two new families (*Parabambusicolaceae*

Kaz. Tanaka & K. Hiray. and *Sulcatisporaceae* Kaz. Tanaka & K. Hiray.) were introduced by Tanaka et al. (2015) in the suborder *Massarineae* of the *Pleosporales*.





**Fig. 7** *Mendogia macrostroma* (MFLU 13–0642, **holotype**). **a** Type material. **b, c** Black ascostromata on host. **d** Vertical section of ascostroma. **e, f** Surface cells of ascostromata and

pseudoparenchymatous cells between asci. **g–j** Asci (**g**: Showing fissitunicate dehiscence). **k–n** Ascospores. Scale bars: **a** = 2 mm, **b, c** = 1 mm, **d–g** = 100  $\mu\text{m}$ , **h–j** = 20  $\mu\text{m}$

Ariyawansa et al. (2015) introduced three more new families *Ascocylindricaceae* Abdel-Wahab et al., *Caryosporaceae* H. Zhang et al. and *Wicklowiaceae* Ariyawansa & K.D. Hyde in *Pleosporales*.

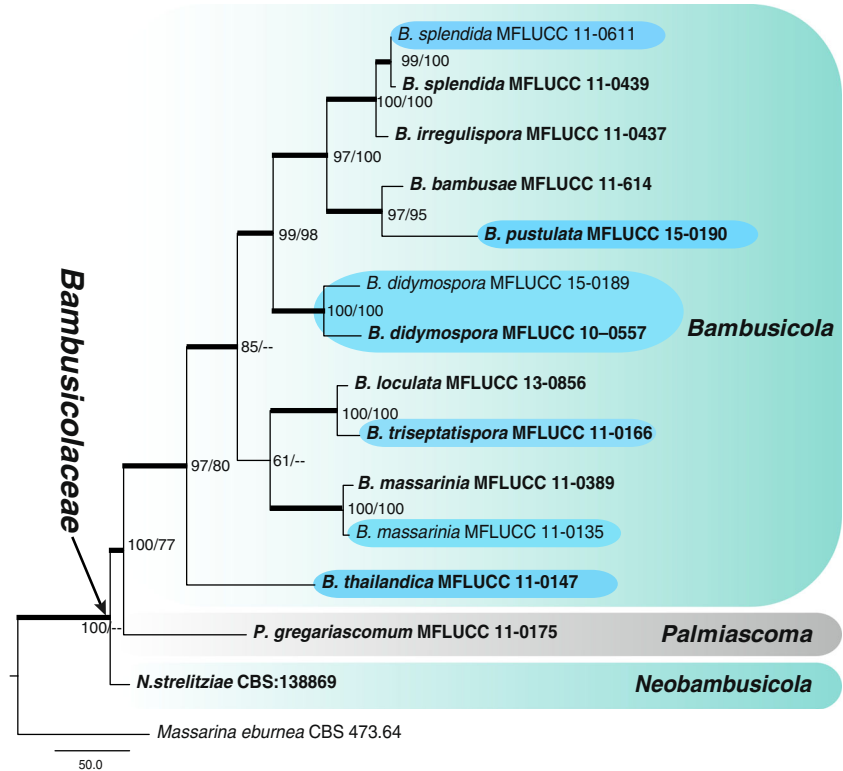
***Bambusicolaceae*** D.Q. Dai & K.D. Hyde, Fungal Diversity 63: 49 (2013)

The family *Bambusicolaceae* was introduced by Hyde et al. (2013), with the type genus *Bambusicola*, to accommodate taxa with small, immersed ascromata, with cellular to trabeculate pseudoparaphyses, cylindrical, bitunicate asci and fusiform ascospores, and usually inhabiting bamboo. Crous et al. (2014b) isolated a related asexual taxon from

leaves of *Strelitzia nicolai* (*Strelitziaceae*) and introduced a new genus *Neobambusicola*. Liu et al. (2015) described *Palmiascoma* from a dead palm frond, thus widening the family concept (Dai et al. 2015). The species of *Bambusicolaceae* usually produce asexual morph on host substrate or on culture, with holoblastic, annelidic or phialidic conidiogenous cells (Dai et al. 2012; Crous et al. 2014b; Liu et al. 2015). In this paper, we provide a multi-gene (SSU, LSU, TEF1 and RPB2 sequence data) phylogram for all genera of *Bambusicolaceae* (Fig. 8).

***Bambusicola*** D.Q. Dai & K.D. Hyde, Cryptog. Mycol. 33(3): 367 (2012)

**Fig. 8** Maximum likelihood phylogenetic tree generated by RAxML (GTR+G model) based on analysis of combined SSU, LSU, TEF1 and RPB2 sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes, and branches with Bayesian posterior probabilities greater than 0.90 are in bold. The original isolate numbers are noted after the species names. The tree is rooted to *Massarina eburnea* (CBS 473.64). Ex-type strains are in *bold*. Newly generated sequences are highlighted with a blue background



*Bambusicola* was introduced by Dai et al. (2012) and typified by *B. massarinia* D.Q. Dai & K.D. Hyde. This genus is characterized by ascostromata forming dark to light raised areas on the host surface with an asexual morph typified by holoblastic, annelidic conidiogenous cells and cylindrical conidia. *Bambusicola* now contains nine species including four new species described in this paper (Dai et al. 2012, 2015).

Key to species of *Bambusicolaceae*

- 1. Fruiting bodies ascostromata.....2
- 1. Fruiting bodies conidiomata only.....10
- 2. Ascospores hyaline.....3
- 2. Ascospores dark brown.....*Palmiascoma gregariascomum*
- 3. Ascostromata containing a single locule.....5
- 4. Ascostromata containing more than one locule.....***Bambusicola loculata***
- 5. Ascospores 3-septate.....6
- 5. Ascospores 1-septate.....7
- 6. Ascostromata less than 160 μm high × 450 μm diam.....***B. thailandica***
- 6. Ascostromata more than 160 μm high × 450 μm diam.....***B. triseptatispora***
- 7. Ascostromata forming dark, rounded, raised spots, on host surface, with a black halo around the ostiolar opening.....8
- 7. Ascostromata forming light coloured, raised spots, on the host surface, with split ostiolar opening.....9
- 8. Asexual morph with cylindrical conidia.....***B. massarinia***

- 8. Asexual morph with oblong to ellipsoidal conidia.....***B. didymospora***
- 9. Ascospores larger than 15 μm × 3 μm.....*B. bambusae*
- 9. Ascospores smaller than 15 μm × 3 μm.....*B. pustulata*
- 10. Conidia fusoid-ellipsoid.....*Neobambusicola strelitziae*
- 10. Conidia cylindrical.....11
- 11. Conidiomata acerose, conidia 15–18 × 1.5–3 μm.....***B. irregulispora***
- 11. Conidiomata subglobose, conidia 20–30 × 3.5–5 μm.....***B. splendida***

***Bambusicola didymospora*** Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.*

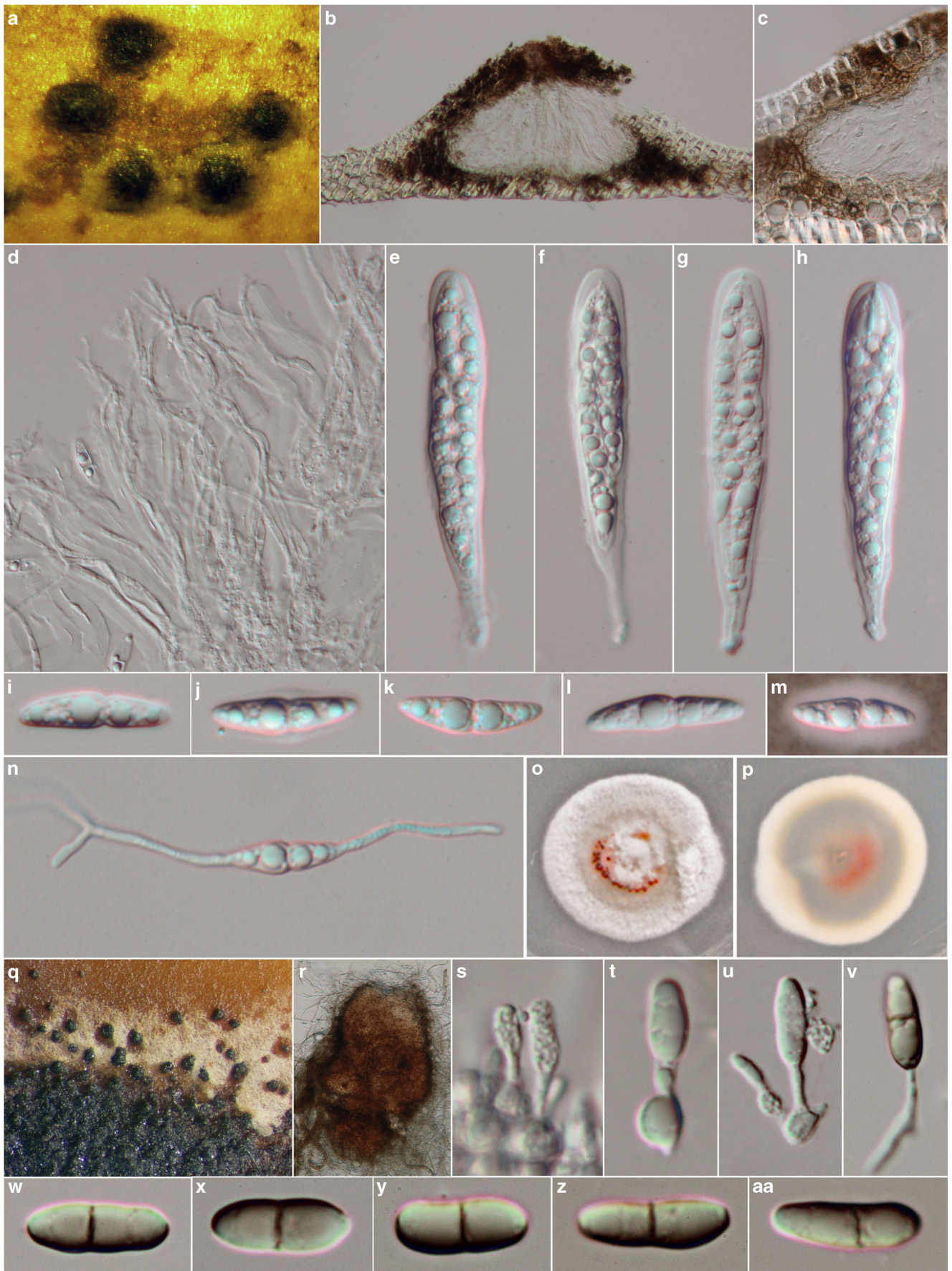
*Index Fungorum*: IF552021; *Facesoffungi* number: FoF 01969; Fig. 9

*Etymology*: The epithet “didymospora” refers to the didymosporous ascospores and conidia.

*Holotype*: MFLU 11–0149

*Saprobic* on dead bamboo culms, as raised, black, dome-shaped structures, on host surface. **Sexual morph**:

**Fig. 9** *Bambusicola didymospora* (MFLU 11–0149, holotype). **a** Appearance of ascostromata on host surface. **b** Vertical section through ascoma. **c** Peridium. **d** Pseudoparaphyses. **e–h** Asci. **i–l** Ascospores. **m** Ascospore stained in Indian ink. **n** Ascospore germination on WA. **o, p** Culture characteristics (upper and lower sides). **q, r** Conidiomata produced on culture colony. **s–v** Conidiogenous cells producing conidia. **w–aa** Conidia. Scale bars: b = 100 μm, c, r = 50 μm, d, e–h, n = 20 μm, i–m = 10 μm, s–aa = 5 μm



*Ascostromata* 90–155 µm high, 310–400 µm diam., solitary, scattered to gregarious, immersed in host cortex, with red pigment in the ostiole, with flattened base, convex or dome-shaped, uni-loculate, rarely bi-loculate, with centrally located ostiole. *Peridium* 16–33 µm wide, thin- to thick-walled, of unequal thickness, poorly developed at the base, composed of several layers of dark brown to black, pseudoparenchymatous cells, intermixed with host cells, arranged in a *textura angularis*, coriaceous. *Hamathecium* of dense, 0.5–1.5 µm wide, filamentous, indistinct, septate, cellular pseudoparaphyses, not constricted at the septum, slightly rough-walled, anastomosing at the apex, embedded in a hyaline gelatinous matrix. *Asci* 55–85 × 9–13 µm ( $\bar{x}$  = 71 × 11.2 µm,  $n$  = 20), 8-spored, bitunicate, fissitunicate, cylindrical-clavate to clavate, short pedicellate, apically rounded, with an ocular chamber. *Ascospores* 17–23 × 4.5–6 µm ( $\bar{x}$  = 20.7 × 5.6 µm,  $n$  = 25), overlapping bi-seriate, hyaline, fusiform, with rounded to acute ends, 1-septate, constricted at the septum, upper cell larger than lower cell, smooth-walled, guttulate, surrounded by a mucilaginous sheath. **Asexual morph:** Coelomycetous, produced in culture after 4 months. *Conidiomata* 90–210 µm high, 95–180 µm diam., pycnidial, solitary, semi-immersed to embedded in agar, visible as raised, black dots, ovoid to obpyriform, or irregular in shape, uni-loculate, ostiolate, covered by vegetative hyphae. *Wall of conidiomata* thin-walled, composed of a few layers of orange-brown to dark brown, pseudoparenchymatous cells, arranged in *textura angularis*. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* (2–)5–8 × 1–3 µm ( $\bar{x}$  = 5.5 × 2 µm,  $n$  = 30), enteroblastic, phialidic, determinate, discrete, cylindrical to ampulliform, hyaline, aseptate, apically attenuated, with distinct periclinal thickening, smooth-walled. *Conidia* (9.5–)10–13(–15) × 4–5 µm ( $\bar{x}$  = 12.5 × 4.5 µm,  $n$  = 30), pale brown, oblong to ellipsoidal, rounded apex, with truncate base, 1-septate, slightly constricted at the septum, smooth-walled.

**Culture characters:** Colonies on MEA 26–30 mm diam. after 2 weeks at 25–30 °C, under 12 h light/12 h dark, circular, dense, flat, slightly raised, dull with entire edge, fluffy to floccose, with regular edge; white at the margin, becoming greyish at the center with red droplets; reverse white to cream at the margin, greenish-grey at middle, pale yellowish to reddish, slightly radiating with pale yellowish concentric ring; non-pigmented, forming black pycnidia at the margin.

**Material examined:** THAILAND, Chiang Rai, Muang District, Huai Mae Sai Waterfall, on dead culms of bamboo, 10 March 2010, R. Phookamsak, RP0028 (MFLU 11–0149, **holotype**), ex-type living culture, MFLUCC 10–0557, KUMCC; Phang-Nga Province, Amphoe Mueang Phangnga, Tambon Tham Nam Phut, forest, 8°26'24"N 98°32'15"E, on dead culms of bamboo, 5 December 2014, Kevin D. Hyde DDQ00288 (MFLU 16–0882, **paratype**), living cultures, MFLUCC 15–0189, MUCL 55886.

**Notes:** *Bambusicola didymospora* is similar to *B. massarinia* D.Q. Dai & K.D. Hyde, *B. bambusae* D.Q. Dai & K.D. Hyde, *B. loculata* D.Q. Dai & K.D. Hyde and *B. pustulata* D.Q. Dai & K.D. Hyde in having didymosporous, fusiform ascospores with rounded ends. However, *B. didymospora* differs from *B. massarinia* in its asexual morph. *Bambusicola didymospora* forms oblong to ellipsoidal, brown to reddish brown conidia, with phialidic conidiogenous cells, while *B. massarinia* forms cylindrical, pale brown conidia with many guttules; phialidic conidiogenous cells were not observed (Dai et al. 2012). *Bambusicola didymospora* differs from *B. loculata* in having scattered ascostromata, while *B. loculata* forms multi-loculate ascostromata (Dai et al. 2015). *Bambusicola didymospora* differs from *B. bambusae* and *B. pustulata* in having immersed ascostromata strongly raised from host surface, black, dome-shaped, however, in *B. bambusae* and *B. pustulata* darken only at the slightly raised cracks (Dai et al. 2012). Multi-gene phylogenetic analyses show that *B. didymospora* forms a robust clade close to *B. bambusae* and *B. pustulata*, but is distinct with high bootstrap support (99/98 % MPBP/MLBP, BYPP value greater than 0.90).

***Bambusicola pustulata*** D.Q. Dai & K.D. Hyde, **sp. nov.**

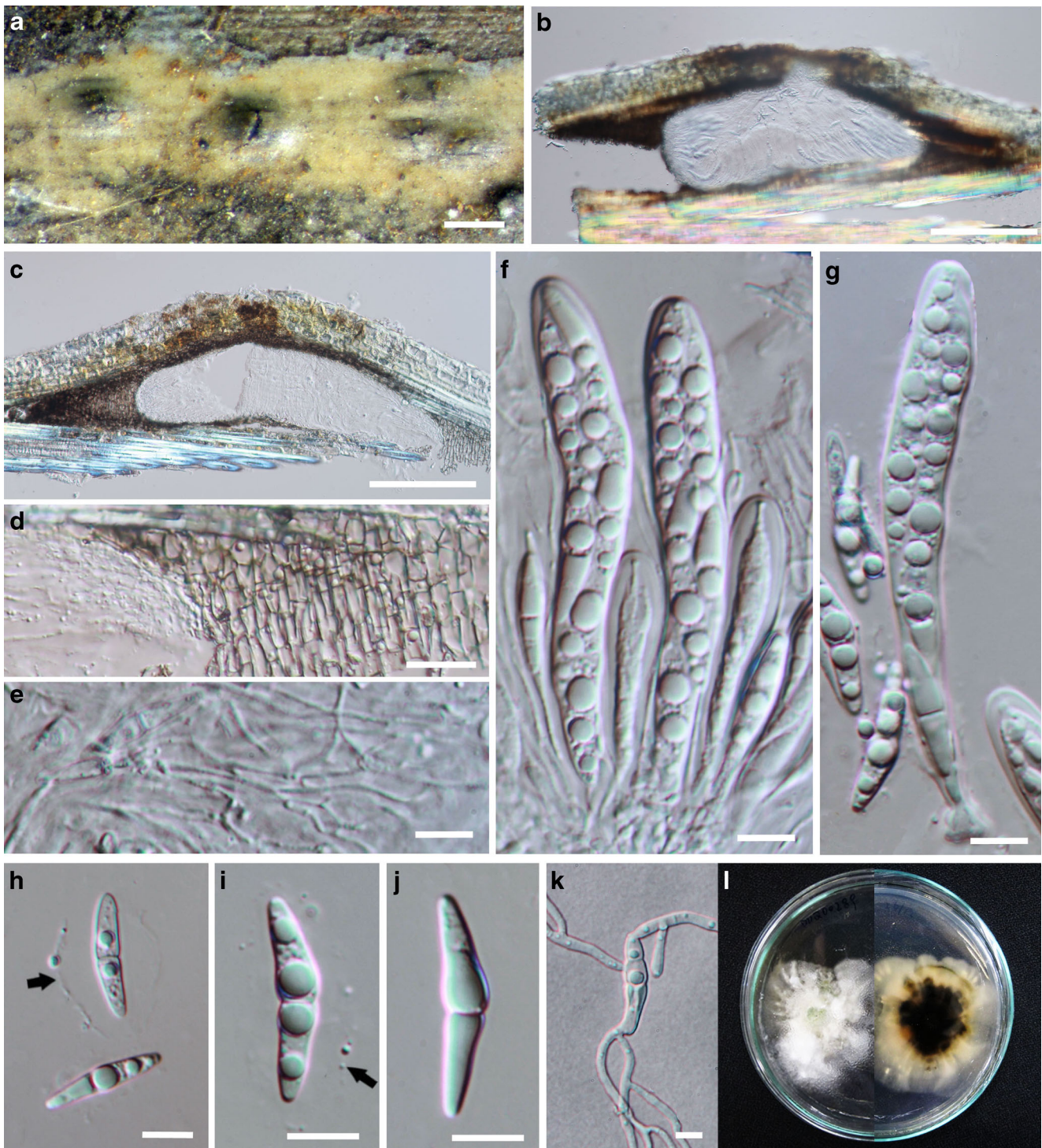
**Index Fungorum numbers:** IF552022; **Facesoffungi numbers:** FoF 01970; **Fig. 10**

**Etymology:** In reference to the ascomata rising as pustules.

**Holotype:** MFLU 15–1216

**Saprobic** on dead culm of bamboo, forming dark pustule-like spots on the host surface with ascostromata breaking through raised cracks at the centre. **Sexual morph:** *Ascostromata* 100–125 µm high, 250–300 µm diam., solitary, scattered or in groups of 2–5, immersed under the host tissue, uni-loculate, subglobose, light brown, coriaceous, ostiolate at the centre, black at the apex of ostiolar opening, 40–45 µm diam. and 30–35 µm high. *Peridium* comprising several layers of host and fungal tissues, laterally 30–45 µm thick, composed of brown to hyaline cells of *textura angularis*, intermingled with host cells, basal part poorly developed; wedge of palisade-like cells at the periphery: 80–105 µm thick, composed of large, thick-walled, dark brown, 4.5–8 × 1–3 µm, cells. *Hamathecium* of dense, long, 0.5–1 µm broad, septate, branched, anastomosing, trabeculate pseudoparaphyses, occurring between and above the asci. *Asci* 40–60 × 5.5–7 µm ( $\bar{x}$  = 52.6 × 6.3 µm,  $n$  = 10), 8-spored, bitunicate, fissitunicate, cylindrical, with a shallow apical chamber and a short furcate pedicel. *Ascospores* 11.5–17.5 × 2.5–3.8 µm ( $\bar{x}$  = 14.2 × 2.9 µm,  $n$  = 20), 2–3-seriate, hyaline, fusiform, 1-septate, occasionally with large upper cell, with narrowly rounded ends, smooth-walled, surrounded by a thick gelatinous sheath 2.5–4.5 µm wide; each cell with 1–2 guttules. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced at both ends. *Colonies* slow



**Fig. 10** *Bambusicola pustulata* (MFLU 15–1216, **holotype**). **a** Appearance of ascostromata on bamboo host. **b, c** Vertical section of ascostromata. **d** Peridium. **e** Pseudoparaphyses. **f, g** Asci. **h–j**

Ascospores (**h, i** Ascospores surrounded by gelatinous sheath). **k** Germinating ascospore. **l** Cultures on PDA. Scale bars: **a** = 500  $\mu\text{m}$ , **b, c** = 100  $\mu\text{m}$ , **d** = 10  $\mu\text{m}$ , **e–k** = 5  $\mu\text{m}$

growing, attaining 35 mm diam. after 40 days at room temperature (25–28 °C), under 12 h light/12 h dark, circular, white on the periphery and yellowish green in center from above, dark yellow to black from below.

**Material examined:** THAILAND, Phang-Nga Province, Amphoe Mueang Phang-nga, Tambon Tham Nam Phut,

forest, 8°26'24"N 98°32'15"E, on dead culms of bamboo, 5 December 2014, Kevin D. Hyde DDQ00289 (MFLU 15–1216, **holotype**); *Ibid.* (KUN HKAS88723, **isotype**), ex-type living cultures, MFLUCC 15–0190, MUCL 55887.

**Notes:** *Bambusicola pustulata* is similar to *B. bambusae* in having small, immersed ascostromata, cylindrical asci and

slightly broad fusiform, 1-septate, hyaline ascospores (Dai et al. 2012). However, this new taxon differs by having dark ascostromata and smaller ascospores ( $14.2 \times 2.9 \mu\text{m}$  vs.  $17.6 \times 3.4 \mu\text{m}$ ), and greener culture (Dai et al. 2012). The branch length of the phylogenetic tree (Fig. 8) also indicates they are different species.

***Bambusicola thailandica*** Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum*: IF552023; *Facesoffungi* number: FoF 01971; Fig. 11

*Etymology*: The epithet “thailandica” refers to the country where the fungus was collected.

*Holotype*: MFLU 11–0183

*Saprobic* on dead bamboo culms. **Sexual morph**: *Ascostromata* 90–155  $\mu\text{m}$  high, 310–400  $\mu\text{m}$  diam., solitary, scattered to gregarious, immersed in host cortex to semi-immersed, erumpent through host tissue at maturity, visible as raised, dark spots on host surface, with flattened base, convex or dome-shaped, wedge-shaped at sides, uniloculate, glabrous, with centrally located ostiole, with a clypeus and flattened tope. *Peridium* 14–28  $\mu\text{m}$  wide, thin-to thick-walled, of unequal thickness, poorly developed at the base, composed of several layers of dark brown to black, pseudoparenchymatous cells intermixed with host cells, arranged in a *textura angularis* to *textura epidermoidea*, coriaceous. *Hamathecium* of dense, 1–2  $\mu\text{m}$  wide, filamentous, indistinctly septate, smooth-walled, cellular pseudoparaphyses, anastomosing at the apex, embedded in a hyaline, gelatinous matrix. *Asci* (60–)70–90(–97)  $\times$  10–12(–14)  $\mu\text{m}$  ( $\bar{x} = 75.5 \times 12 \mu\text{m}$ ,  $n=20$ ), 8-spored, bitunicate, fissitunicate, broadly cylindrical to cylindri-clavate, with a short pedicel, apically rounded with well-developed ocular chamber. *Ascospores* (16–)18–22(–24)  $\times$  (3.5–)4–6  $\mu\text{m}$  ( $\bar{x} = 20.8 \times 5 \mu\text{m}$ ,  $n=30$ ), overlapping, uni- to bi-seriate, hyaline, fusiform, with acute ends, 3-septate, slightly constricted at the median septum, smooth-walled. **Asexual morph**: Undetermined.

*Culture characters*: Colonies on MEA fast growing, 65–70 mm diam. after 4 weeks at 25–30 °C, under 12 h light/12 h dark, circular, medium dense, flat, slightly raised, smooth, with entire edge, fluffy to floccose, white to cream, reverse white to cream at the margin, reddish grey to greenish grey at middle, pale grey at the center, slightly radiating, not producing pigments.

*Material examined*: THAILAND, Chiang Rai Province, Muang District, Mae Fah Luang University campus grounds, on dead culms of bamboo, 13 August 2010, R. Phookamsak, RP0063 (MFLU 11–0183, **holotype**), ex-type living cultures, MFLUCC 11–0147, KUMCC.

*Notes*: *Bambusicola thailandica* is morphologically similar to *B. triseptatispora* in having phragmospores, however, differs from latter in having a thinner and lighter peridium. Moreover, ascostromata in *B. thailandica* have a flattened top,

**Fig. 11** *Bambusicola thailandica* (MFLU 11–0183, **holotype**). **a** Appearance of ascostromata on host surface. **b** Vertical section through ascostroma. **c, d** Section through peridium. **e** Pseudoparaphyses. **f–h** *Asci*. **i–m** *Ascospores*. **n, o** Culture characteristics (upper and lower sides). Scale bars: b = 100  $\mu\text{m}$ , c–h = 20  $\mu\text{m}$ , i–m = 5  $\mu\text{m}$

whereas, these in *B. triseptatispora* are pointed. *Bambusicola thailandica* forms a distinct clade in multi-gene analyses, at the base of *Bambusicola*, whereas *B. triseptatispora* clusters with *B. loculata* (Fig. 8).

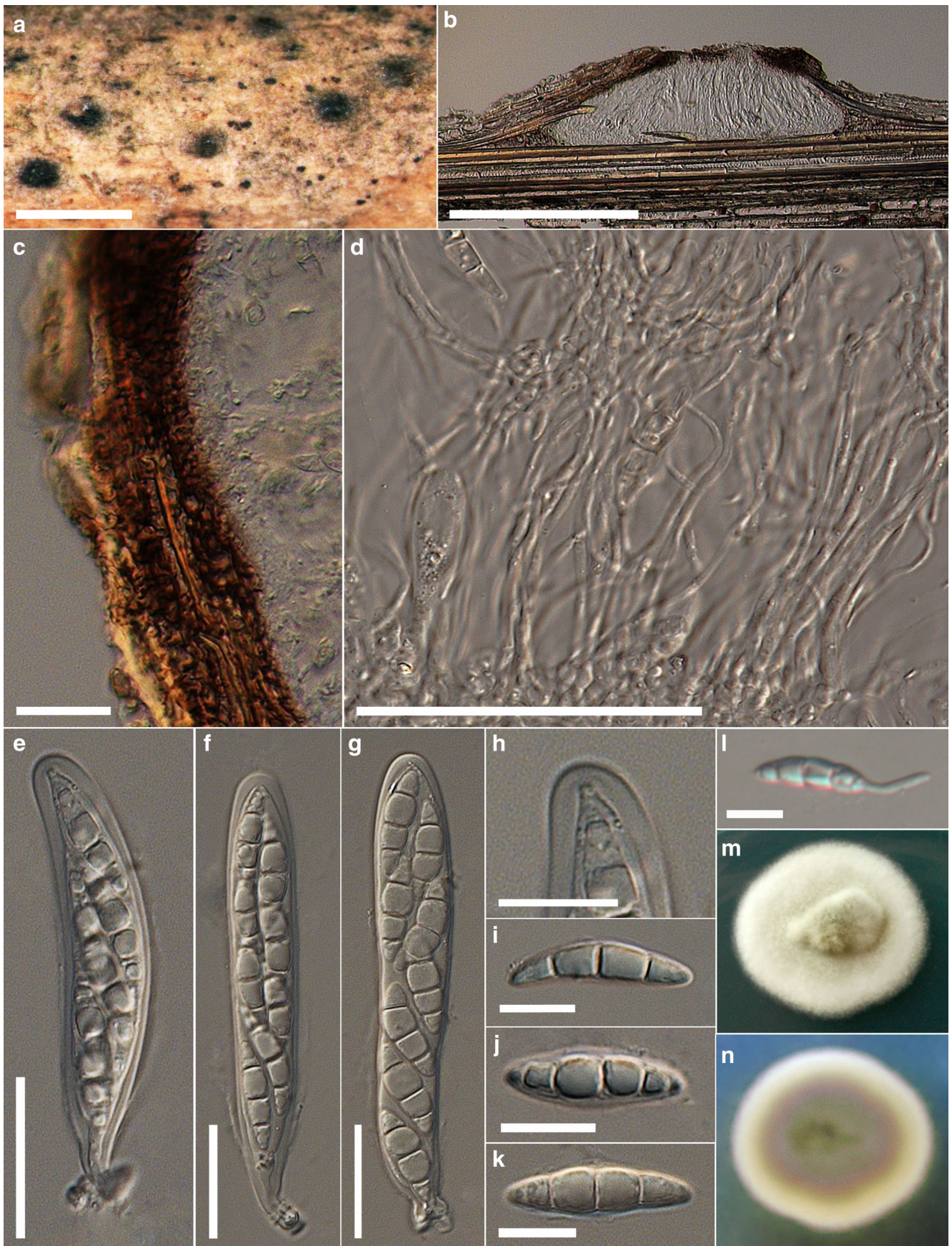
***Bambusicola triseptatispora*** Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.*

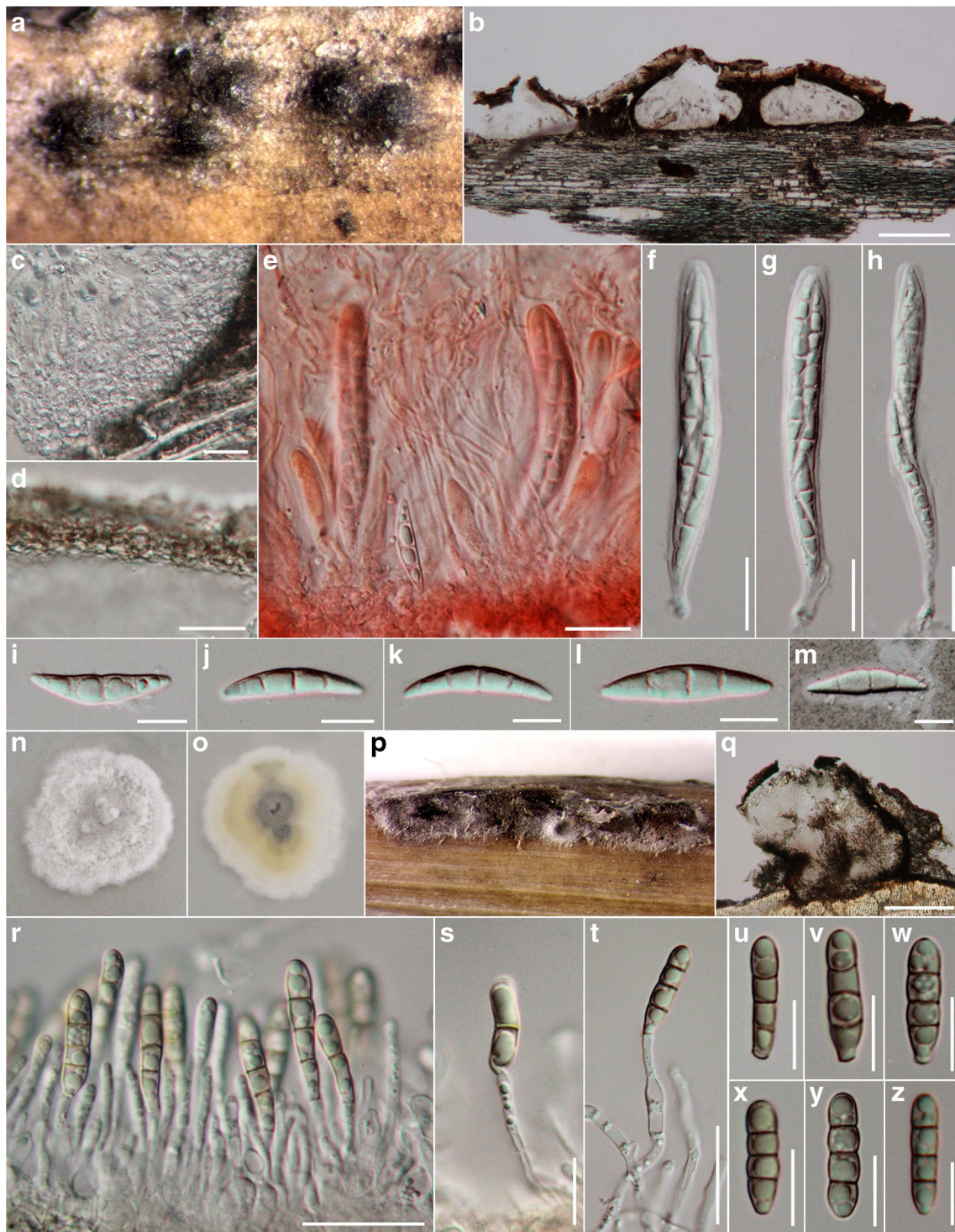
*Index Fungorum*: IF552024; *Facesoffungi* number: FoF 01972; Fig. 12

*Etymology*: The epithet “triseptatispora” refers to the 3-septate ascospores and conidia.

*Holotype*: MFLU 11–0202

*Saprobic* on rotten dead bamboo culms. **Sexual morph**: *Ascostromata* 180–310  $\mu\text{m}$  high, 470–730  $\mu\text{m}$  diam., scattered to clustered, solitary to gregarious, immersed, sub-epidermal, visible as raised, black structures on the host surface, with flattened base, convex or dome-shaped to ampulliform, uni-loculate; sometimes forming pseudostroma, bi- to tri-loculate, glabrous, centrally ostiolate ascostromata. *Peridium* 10–40  $\mu\text{m}$  wide, thin- to thick-walled, of unequal thickness, slightly thick at the sides, poorly developed at the base, composed of several layers of brown to dark brown, pseudoparenchymatous cells interdispersed with host cells, with inner layers comprising several layers of hyaline *textura angularis* cells, outer layers comprising several layers of flattened, brown to dark brown *textura angularis* cells, coriaceous. *Hamathecium* dense, filamentous, 1–2  $\mu\text{m}$  wide, distinctly septate, smooth-walled, cellular pseudoparaphyses, anastomosing at the apex, embedded in a hyaline gelatinous matrix. *Asci* (78–)80–100(–110)  $\times$  10–12(–14)  $\mu\text{m}$  ( $\bar{x} = 95 \times 11.1 \mu\text{m}$ ,  $n=30$ ), 8-spored, bitunicate, fissitunicate, cylindrical to cylindri-clavate, short pedicellate, apically rounded, with well-developed ocular chamber. *Ascospores* (25–)26–30(–31)  $\times$  4–6  $\mu\text{m}$  ( $\bar{x} = 27.7 \times 5.5 \mu\text{m}$ ,  $n=30$ ), overlapping, uni- to bi-seriate, hyaline to pale brown, fusiform, with acute ends, 3-septate, rarely constricted at septa, smooth-walled, surrounded by an irregular mucilaginous sheath. **Asexual morph**: Coelomycetous, produced on bamboo pieces on WA after 2 months. *Conidiomata* 350–480  $\mu\text{m}$  high, 420–700  $\mu\text{m}$  diam., pycnidial, solitary to gregarious, semi-immersed at the base, becoming superficial, visible as raised, black dots on colonies, eustromatic or sometimes pseudostromatic, irregularly shaped, uni- to multi-loculate, covered by vegetative hyphae, with indistinct ostioles. *Wall of Conidiomata* 17–60  $\mu\text{m}$  wide, thin- to thick-walled, of unequal thickness, slightly thick at sides, composed of several layers, of dark





**Fig. 12** *Bambusicola triseptatispora* (MFLU 11–0202, holotype). **a** Appearance of ascostromata on host surface. **b** Vertical section through ascostromata. **c**, **d** Section through peridium. **e** Asci with pseudoparaphyses stained with congo red. **f–h** Asci. **i–l** Ascospores. **m** Ascospore stained with Indian ink. **n**, **o** Cultures characters (upper and

lower sides). **p** Conidiomata produced on bamboo pieces on WA. **q** Section through pycnidia. **r–t** Conidiogenous cells attached with conidia. **u–z** Conidia. Scale bars: b, q = 200  $\mu\text{m}$ , c–h, r, t = 20  $\mu\text{m}$ , i–m, s, u–z = 10  $\mu\text{m}$

brown to black, *textura angularis* to *textura prismatica* pseudoparenchymatous cells mixed with host cells. *Conidiophores* reduced to conidiogenous cells.

*Conidiogenous cells* (9–)(10–)12–28(–32)  $\times$  1.5–3  $\mu\text{m}$  ( $\bar{x}$  = 18.8  $\times$  2.2  $\mu\text{m}$ ,  $n=30$ ), holoblastic, determinate, discrete, cylindrical to irregularly-shaped, hyaline, with distinct



periclinal thickening, smooth. *Conidia* (15–)18–20(–22) × 3–5 μm ( $\bar{x}$  = 19.3 × 4.1 μm,  $n$  = 30), cylindrical to cylindric-clavate, narrow towards ends, apex rounded, with truncate base, initially 1-septate and hyaline, becoming 3-septate and brown at maturity, constricted at the septa, smooth, with guttules.

**Culture characters:** Colonies on PDA fast growing, 50–60 mm diam. after 4 weeks at 25–30 °C, under 12 h light/12 h dark, irregular, medium dense to dense, flat, slightly raised, rough with lobate edge, fluffy to floccose, or velvety; white to cream at the margin, white-grey at the centre; reverse white to cream at the margin, pale yellowish to yellowish at middle, yellowish grey to dark grey at the center, slightly radiating outwards; forming grey to white tufts, with pycnidia and yellowish droplets surrounding the colonies.

**Material examined:** THAILAND, Mukdahan Province, Nongsung District, Wang Hai Village, on dead culms of bamboo, 13 April 2011, R. Phookamsak, RP0082 (MFLU11–0202, **holotype**), ex-type living cultures, MFLUCC 11–0166, KUMCC.

**Notes:** *Bambusicola triseptatispora* is similar to *B. thailandica* in having phragmosporous, fusiform, 3-septate ascospores. However, *B. triseptatispora* differs from *B. thailandica* by the colour of its ascospores, ascostromata and asexual morph. *Bambusicola triseptatispora* has phragmosporous, fusiform, pale brown, 3-septate, smooth ascospores with uni- to multi-loculate pseudoascostromata and phragmosporous, cylindric-clavate, brown, 3-septate conidia, whereas *B. thailandica* forms phragmosporous, fusiform, hyaline, ascospores slightly constricted at median septum, with scattered, uni-loculate ascostromata. The asexual morph of *B. triseptatispora* is similar to *B. splendida*. However, *B. triseptatispora* has smaller conidia and longer conidiogenous cells. Based on multi-gene phylogenetic analyses (Fig. 8), *B. triseptatispora* forms a robust clade with *B. loculata*, which is similar, but has multi-loculate ascostromata and 1-septate ascospores (Dai et al. 2015).

***Occultibambusaceae*** D.Q. Dai & K.D. Hyde, **fam. nov.**

**Index Fungorum number:** IF552012; **Facesoffungi number:** FoF 01973

**Saprobic** on dead bamboo culms or teak branches, forming dark, ascostromata on raised areas, with ostiolate opening. **Sexual morph:** *Ascostromata* solitary, scattered or gregarious, subglobose, uni- or multi-loculate, greyish to dark brown, coriaceous, with a central, papillate, rounded ostiole, internally lined with periphyses. *Peridium* comprising host and fungal tissues or, only fungal tissue brown and thick-walled to hyaline and thin-walled cells of *textura angularis*. *Hamathecium* composed of long, septate, cellular pseudoparaphyses, above the asci. *Asci* 8-spored, bitunicate, broadly cylindrical to clavate, with a short furcate pedicel, with a shallow ocular chamber. *Ascospores* 2–3-seriate, slightly broad-fusiform, 1–3-septate, hyaline, pale brown to dark brown, surrounded by a

gelatinous sheath, with guttulate cells. **Asexual morph:** Coelomycetous, produced on bamboo pieces on WA. *Conidiomata* eustromatic, immersed to partly immersed, solitary to gregarious, globose to subglobose, conical in section, dark ostiolate, with a short neck. *Conidiomata wall* with several layers, composed of dark to hyaline cells of *textura angularis*. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* enteroblastic, phialidic, determinate, discrete, ampulliform to cylindrical, smooth-walled. *Conidia* obovoid, cylindrical to oblong, 0–3-septate, hyaline to pale brown, rounded at the apex, smooth-walled, guttulate.

**Notes:** *Occultibambusaceae* is introduced herein to accommodate two new genera *Occultibambusa* and *Seriascoma*, a coelomycetous genus *Versicolorisporium* (Hatakeyama et al. 2008) and *Neoccultibambusa* introduced in Doilom et al. (2016).

Species of the family *Occultibambusaceae* usually occur on monocotyledons, such as bamboo, but have also been found on hardwood trees such as teak (*Tectona grandis* L.f., *Lamiaceae*). This family is characterized by immersed, solitary to gregarious ascostromata with black ostioles, broadly cylindrical to clavate, bitunicate asci, cellular pseudoparaphyses and broad-fusiform, hyaline to dark brown ascospores with 1–3 septa. The asexual morphs are morphologically diverse. For example, the conidia in the genus *Occultibambusa* are oblong and pale brown, whereas *Seriascoma* has cylindrical and hyaline conidia. Obovoid conidia with a darker middle cell are observed in *Versicolorisporium* (Hatakeyama et al. 2008). However, they are all formed in eustromatic conidiomata and have phialidic conidiogenous cells with enteroblastic conidiogenesis.

Members of *Occultibambusaceae* are similar to species of *Bambusicola*, *Lophiostoma*, and *Massarina* in having fusiform ascospores and clavate asci (Zhang et al. 2009; Dai et al. 2012, 2015). However, *Bambusicola* has hyaline ascospores and asexual morphs with usually annellidic conidiogenous cells (Dai et al. 2012). Ascospores of the species of *Occultibambusaceae* are mostly brown, no annellidic conidiogenous cells observed. *Lophiostoma* has ascostromata containing a compressed/crest-like ostiole (Zhang et al. 2009; Thambugala et al. 2015), while those in *Occultibambusaceae* are papillate. *Lophiostoma* and *Massarina* differ by cylindrical or oblong asci (Zhang et al. 2009; Thambugala et al. 2015).

The family *Occultibambusaceae* is phylogenetically close to the family *Biatrisporaceae* (Fig. 13). However, the species of *Biatrisporaceae* are manglicolous and the dark brown ascospores usually have hyaline, rounded, swollen ends which release mucilage (Hyde et al. 2013). Members of *Occultibambusaceae* are separated from *Biatrisporaceae* by high bootstrap support (100 % MLBP, BYPP greater than 0.90).

*Type: Occultibambusa* D.Q. Dai & K.D. Hyde

*Occultibambusa* D.Q. Dai & K.D. Hyde, *gen. nov.*

*Index Fungorum number:* IF552013; *Facesoffungi number:* FoF 01974

*Etymology:* Occulti means hidden in Latin, and refers to members of this genus hidden on bamboo substrates.

*Saprobic* on dead bamboo culms, forming dark, round, black spots on host surface, with ascostromata on raised areas, with an ostiolate opening. **Sexual morph:** *Ascostromata* solitary, scattered, immersed under host tissue, subglobose, conical in section, uni-loculate, black at ostiolar region, coriaceous, with a central, papillate, rounded ostiole, internally lined with periphyses. *Peridium* comprising host and fungal tissues, composed of brown and thick-walled to hyaline and thin-walled cells of *textura angularis* intermingled with host cells. *Hamathecium* composed of long, septate, cellular pseudoparaphyses above the asci. *Asci* 8-spored, bitunicate, broadly cylindrical to clavate, with a short furcate pedicel, and a shallow ocular chamber. *Ascospores* 2–3-seriate, broad-fusiform, 1-septate, usually with larger upper cell, narrowly acute at both ends, dark brown, straight to curved, surrounded by a gelatinous sheath, guttulate. **Asexual morph:** Coelomycetous, produced on bamboo pieces on WA. *Conidiomata* eustromatic, immersed to partly-immersed, solitary to gregarious, globose to subglobose, black, with a black papillae or short neck. *Wall of conidiomata* comprising several layers, composed of dark to hyaline cells of *textura angularis*. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* enteroblastic, phialidic, determinate, ampulliform to cylindrical, smooth-walled. *Conidia* oblong, aseptate, pale brown to hyaline, rounded at the apex, smooth-walled, guttulate.

*Type species: Occultibambusa bambusae* D.Q. Dai & K.D. Hyde

*Notes:* The genus *Occultibambusa* is characterized by immersed, dark coloured, ascostromata with periphysate ostioles, broad cylindrical to clavate asci, and dark brown, fusiform, 1-septate ascospores. The asexual morphs of *Occultibambusa* produce black necks at the center of the conidiomata. This character is not observed in other genera of the family *Occultibambusaceae*. Characters such as pale brown and oblong conidia are similar with *Bambusicola* species (Dai et al. 2012), however, annellidic conidiogenous cells are not observed in *Occultibambusa*.

*Occultibambusa bambusae* D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum number:* IF552016; *Facesoffungi number:* FoF 01975; Fig. 14

*Etymology:* Refers to the bamboo host.

*Holotype:* MFLU 15–1212

*Saprobic* on dead bamboo culms, forming dark, round spots on host surface, with ascostromata on raised areas, but

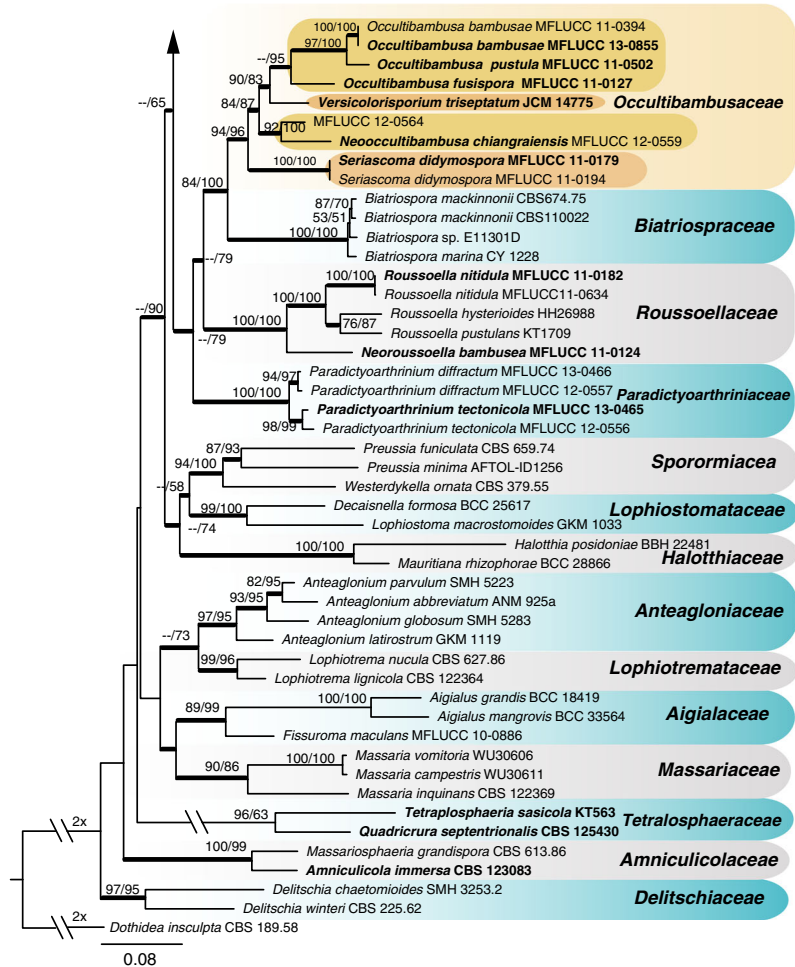
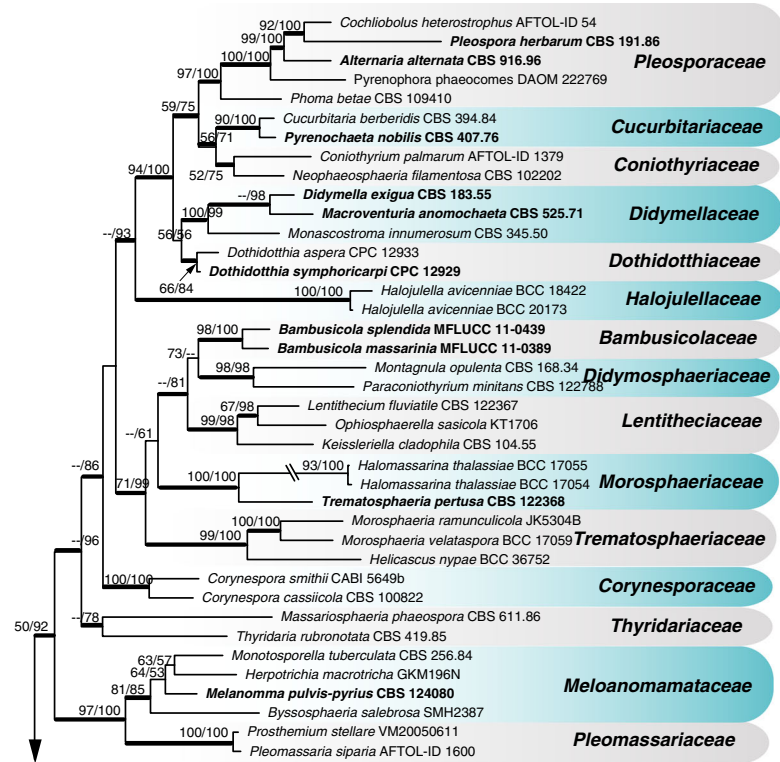
still immersed under host tissue. **Sexual morph:** *Ascostromata* 150–200 µm high, 400–550 µm diam., solitary, scattered, immersed, subglobose, slightly conical in section, uni-loculate, black at the ostiolar regions, coriaceous, with a central, rounded, papillate ostiole, lined internally with periphyses. *Peridium* comprising host and fungal tissues, 30–45 µm thick, composed of brown to hyaline, thick- to thin-walled cells of *textura angularis*, intermingled with host cells, with basal part 15–30 µm thick; wedge of palisade-like cells at the periphery: 50–100 µm thick, composed of large, brown cells. *Hamathecium* composed of long, 2–3.5 µm wide, septate, cellular pseudoparaphyses branching above the asci. *Asci* (50–)60–80(–90) × (9.5–)11.5–14.5(–15) µm ( $\bar{x}$  = 70.8 × 13.1 µm,  $n$  = 20), 8-spored, bitunicate, broadly cylindrical, with a short furcate pedicel, with a shallow ocular apical chamber. *Ascospores* (22–)23.5–27.5 × 4.5–7 µm ( $\bar{x}$  = 25.7 × 6.3 µm,  $n$  = 20), 2–3-seriate, slightly broad and fusiform, 1-septate, occasionally with larger upper cell, narrowly acute at both ends, dark brown, straight to curved, surrounded by a gelatinous sheath, each cell with 2–3 guttules. **Asexual morph:** Undetermined.

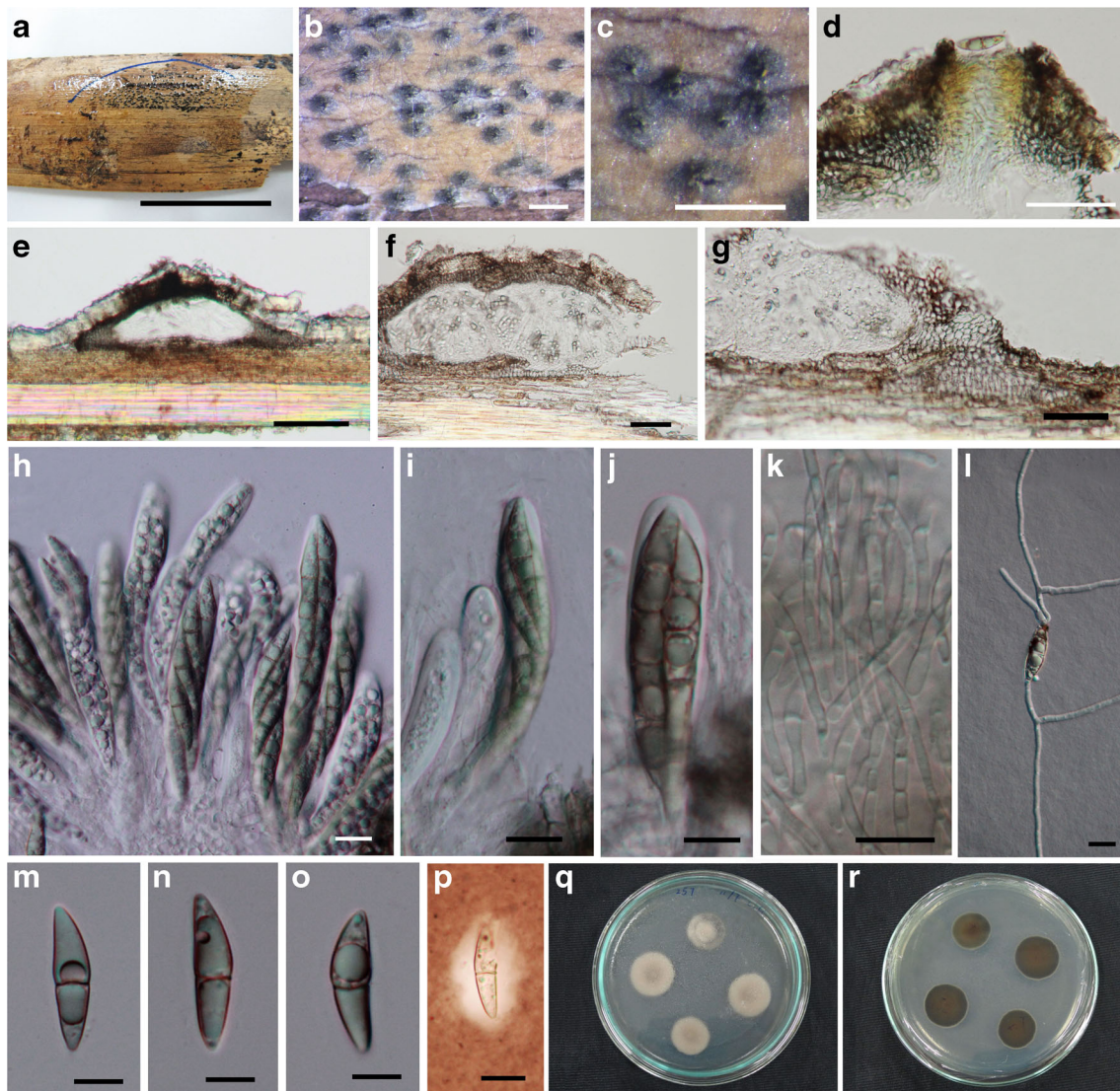
*Cultural characters:* Ascospores germinating on PDA within 24 h with germ tubes produced from both cells. Colonies growing slowly on PDA, reaching 1.5–2 mm in 18 days at 28 °C, under 12 h light/12 h dark, circular, floccose, with even margin, dark brown above and below. Mycelium immersed and superficial in the media, composed of branched, septate, smooth-walled, brown hyphae.

*Material examined:* THAILAND, Chiang Rai Province, Mae Fah Luang University, on dead culms of bamboo, 9 July 2013, Dong-Qin Dai, DDQ00262 (MFLU 15–1212, **holotype**); *Ibid.* (KUN HKAS88749, **isotype**), ex-type living cultures, MFLUCC 13–0855, CBS 139960, MUCL 55882; Chiang Mai, Mae Sae Village, on dead culms of bamboo, 14 May 2011, Dong-Qin Dai, DDQ00025 (MFLU 16–0880), living culture, MFLUCC 11–0394.

*Notes:* *Occultibambusa bambusae* is similar to *O. fusispora* in having clavate asci and fusiform, dark brown ascospores. However, *O. bambusae* differs in its larger ascostromata (150–200 µm high, 400–550 µm diam. vs. 135–185 µm high, 240–275 µm diam.). The branch length of phylogenetic tree shows they are different species (Fig. 13). *Occultibambusa bambusae* can produce ascostromata on bamboo pieces on WA after 3 months, however, asexual morph are not observed from culture.

**Fig. 13** Maximum likelihood phylogenetic tree generated by RAxML (GTR+G model) based on combined SSU, LSU, TEF1 and RPB2 sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes, and branches with Bayesian posterior probabilities greater than 0.90 are given in bold. The original isolate numbers are noted after the species names. The tree is rooted to *Dothidea insculpta* (CBS 189.58). Ex-type strains are in bold. The new family is highlighted with a yellow background





**Fig. 14** *Occultibambusa bambusae* (MFLU 15–1212, holotype). **a–c** Ascostromata developing on bamboo culm. **d** Ostiole with periphyses. **e, f** Vertical section of ascostroma (**e** Ascostroma with wedge of palisade-like cells at the periphery). **g** Wall of ascostroma. **h–j** Asci containing

ascospores. **k** Pseudoparaphyses. **l** Germinating ascospore. **m–o** Dark brown ascospores with a single septum. **p** Ascospore with gelatinous sheath. **q, r** Cultures on MEA. Scale bars: **a** = 5 cm, **b, c** = 500  $\mu\text{m}$ , **e** = 100  $\mu\text{m}$ , **d, f, g** = 50  $\mu\text{m}$ , **h–p** = 10  $\mu\text{m}$

*Occultibambusa fusispora* Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.*

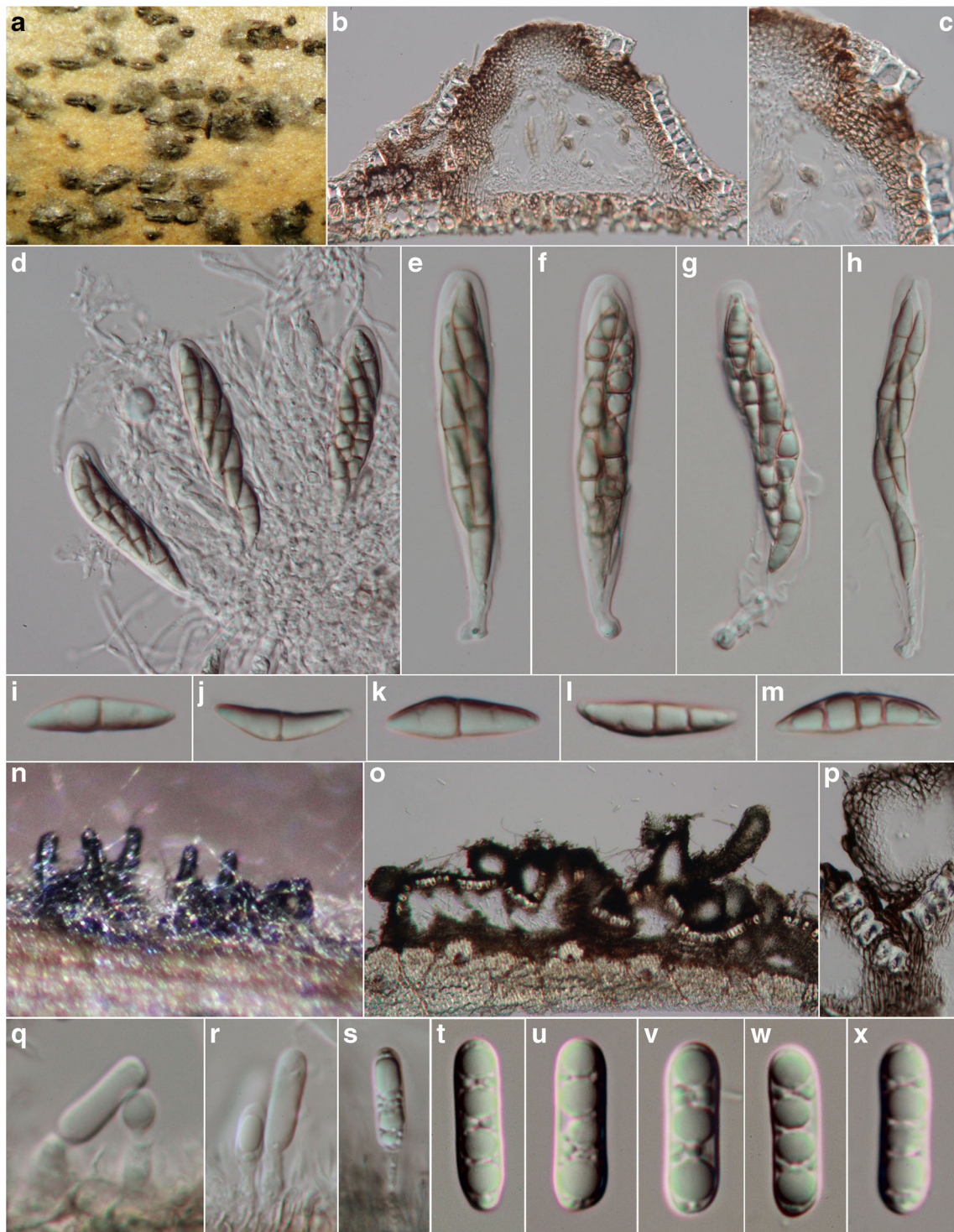
*Index Fungorum*: IF552017; *Facesoffungi* number: FoF 01976; Fig. 15

*Etymology*: The epithet “fusispora” refers to the fusiform ascospores.

*Holotype*: MFLU 11–0163

*Saprobic* on dead bamboo culm. **Sexual morph**: *Ascomata* 135–185  $\mu\text{m}$  high, 240–275  $\mu\text{m}$  diam., scattered, gregarious, subepidermal, semi-immersed to erumpent through host surface, raised, brown to dark brown, with black papillate dots, conical with wedged sides, uni-loculate, ostiole central with minute papilla. *Peridium* 7–60  $\mu\text{m}$  wide, pseudostromatic, with thin- to thick-walled cells of unequal thickness, slightly thick

at the sides and towards apex, poorly developed at the base, composed of several layers of small, brown to dark brown, *textura angularis* to *textura prismatica* pseudoparenchymatous cells. *Hamathecium* composed of dense, broad, 1.8–2.7  $\mu\text{m}$  wide, filamentous, distinctly septate, cellular pseudoparaphyses, not constricted at the septa, anastomosing at the apex, embedded in a gelatinous matrix. *Asci* (60–)65–90(–110)  $\times$  (11–)12–14(–15)(–16)  $\mu\text{m}$  ( $\bar{x}$  = 75.6  $\times$  13.2  $\mu\text{m}$ ,  $n$  = 30), 8-spored, bitunicate, fission-tunicate, clavate to cylindrical-clavate, short pedicellate, with furcate to obtuse ends, apically rounded with well-developed, ocular chamber. *Ascospores* (20–)22–25(–26)  $\times$  5–6(–6.5)  $\mu\text{m}$  ( $\bar{x}$  = 23.9  $\times$  5.7  $\mu\text{m}$ ,  $n$  = 30), overlapping, bi-seriate, fusiform, with acute ends, light brown, mostly 1-septate, rarely 2–



**Fig. 15** *Occultibambusa fusispora* (MFLU 11–0163, holotype). **a** Appearance of ascostromata on host surface. **b** Vertical section through ascostroma. **c** Section through peridium. **d** Asci with pseudoparaphyses. **e–h** Asci. **i–m** Ascospores. **n** Pycnostroma produced on bamboo pieces

on WA. **o** Section through pycnostroma. **p** Pycnidial wall. **q–s** Conidiogenous cells. **t–x** Conidia. Scale bars: **o** = 200  $\mu\text{m}$ , **b** = 100  $\mu\text{m}$ , **c** = 50  $\mu\text{m}$ , **d–h**, **p** = 20  $\mu\text{m}$ , **i–m**, **q–s** = 10  $\mu\text{m}$ , **t–x** = 5  $\mu\text{m}$

3-septate, not constricted at the septum, slightly curved, smooth-walled. **Asexual morph:** Coeloemycetous, produced on bamboo pieces on WA after 12 weeks. *Conidiomata* 160–250  $\mu\text{m}$  high (including beaks),

500–800  $\mu\text{m}$  diam., eustromatic, subepidermal, solitary to gregarious, clustered, visible as raised, black, elongate spikes, covered by sparse hyphae on bamboo pieces, elongate irregular in shape, multi-loculate,

glabrous, ostiolate with long beaked papilla. *Locules* 140–250  $\mu\text{m}$  high (including neck), 110–230  $\mu\text{m}$  diam., gregarious, clustered, immersed in cortex of host to erumpent through host surface by papilla, conical to ampulliform, or irregular-shaped. *Neck* 50–195  $\mu\text{m}$  high 50–100  $\mu\text{m}$  diam., in each locule, ostiole central, slightly oblique, with short to long beaks. *Wall of conidiomata* 10–45  $\mu\text{m}$  wide, thin- to thick-walled, of unequal thickness, thick at the sides and base of pycnidia, composed of several layers of dark brown to black, *textura angularis* to *textura prismatica* pseudoparenchymatous cells, with host cells and fungal tissue. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* (6–)7–12(–15)  $\times$  2–4  $\mu\text{m}$  ( $\bar{x}$  = 10.2  $\times$  2.7  $\mu\text{m}$ ,  $n$  = 30), enteroblastic, phialidic, determinate, discrete, cylindrical to ampulliform or lageniform, hyaline, aseptate, apically attenuated, with indistinct periclinal thickening, smooth-walled. *Conidia* (14–)15–17(–19)  $\times$  4–5(–5.5)  $\mu\text{m}$  ( $\bar{x}$  = 16.5  $\times$  5.2  $\mu\text{m}$ ,  $n$  = 30), oblong to cylindrical, with rounded ends, hyaline, aseptate, smooth-walled, with large guttules.

**Culture characters:** Colonies on MEA fast growing, 55–60 mm diam. after 4 weeks at 25–30 °C, under 12 h light/12 h dark, colonies irregular, dense, flat, slightly raised, dull to rough, with undulate edge, velvety, radiating, wrinkled and folded at the margin, green to brown greenish at the margin, dull green to white-grey at the centre from above, brown greenish at the margin, dark greenish to black at the centre from below, no pigments on PDA.

**Material examined:** THAILAND, Chiang Rai Province, Mae Fah Luang District, Doi Tung, on dead branch of bamboo, 29 June 2010, R. Phookamsak, RP0043 (MFLU 11–0163, **holotype**), ex-type living cultures, MFLUCC 11–0127, KUMCC.

**Notes:** Morphologically, *Occultabambusa fusispora* is similar to *O. bambusae* in having clavate asci and fusiform, brown ascospores. However, they differ in the size of asci and ascospores, ascospore septation, sheath surrounding the ascospores and asexual morph. *Occultabambusa fusispora* has longer asci (60–110  $\mu\text{m}$  vs. 50–90  $\mu\text{m}$ ) and smaller ascospores (22–25  $\times$  5–6  $\mu\text{m}$  vs. 23.5–27.5  $\times$  4.5–7  $\mu\text{m}$ ). In addition, *O. fusispora* has 1–3-septate ascospores, lacks a mucilaginous sheath. *Occultabambusa bambusae* has 1-septate ascospores with a mucilaginous sheath. Based on multi-gene phylogenetic analyses (Fig. 13), *O. fusispora* forms a robust clade which is distinct from *O. bambusae* (95 % MLBP, BYPP greater than 0.90). Therefore, we introduce a second species in the genus *Occultabambusa*.

***Occultabambusa pustula*** D.Q. Dai & K.D. Hyde, *sp. nov.*

**Index Fungorum number:** IF552018; ***Facesoffungi* number:** FoF 01977; Fig. 16

**Etymology:** Refers to its pustule-like ascomata.

**Holotype:** MFLU 15–1212

**Saprobic** on decaying bamboo culms, forming black, rounded, pustule-like spots on host surface, with immersed ascostromata on raised areas. **Sexual morphs:** *Ascostromata* 150–200  $\mu\text{m}$  high, 200–300  $\mu\text{m}$  diam., immersed, subepidermal, slightly conical, black, with a centrally located ostiole. *Peridium* 30–80  $\mu\text{m}$  thick, composed of brown and thick-walled to hyaline and thin-walled *textura angularis*, upper peridium combined with epidermal cells. *Hamathecium* composed of dense, long, up to 2  $\mu\text{m}$  wide, cellular pseudoparaphyses above the asci. *Asci* 80–105  $\times$  8–12  $\mu\text{m}$  ( $\bar{x}$  = 90.4  $\times$  11.4  $\mu\text{m}$ ,  $n$  = 20), 8-spored, bitunicate, cylindrical, with a short furcate pedicel, with a shallow ocular chamber. *Ascospores* 22–25  $\times$  5–5.5  $\mu\text{m}$  ( $\bar{x}$  = 21.6  $\times$  5.3  $\mu\text{m}$ ,  $n$  = 20), 2–3-seriate, slightly broad-fusiform, 1-septate, hyaline to pale brown, guttulate, surrounded by a gelatinous sheath. **Asexual morphs:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from both ends. Colonies growing slowly on PDA, reaching 40 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, circular, edge regular, flocculent, dark brown from above and below. Mycelium superficial to immersed in media, composed of branched, septate, smooth hyphae.

**Material examined:** THAILAND, Chiang Rai, Mae Fah Luang University, on dead culm of bamboo, 19 July 2011, Dong-Qin Dai DDQ00081 (MFLU 15–1185, **holotype**); *Ibid.* (KUN HKAS88695, **isotype**), ex-type living culture, MFLUCC 11–0502.

**Notes:** *Occultabambusa pustula* is similar to *O. bambusae* in having broad fusiform, 1-septate ascospores, but the latter has darker brown ascospores. Further distinctions between *O. pustula* and *O. bambusae* is that the former has raised and darker ascomata and there are also colony differences. Branch length in the phylogenetic tree (Fig. 13) shows that they are different species with high bootstrap support value (97/100 % MPBP/MLBP, BYPP greater than 0.90).

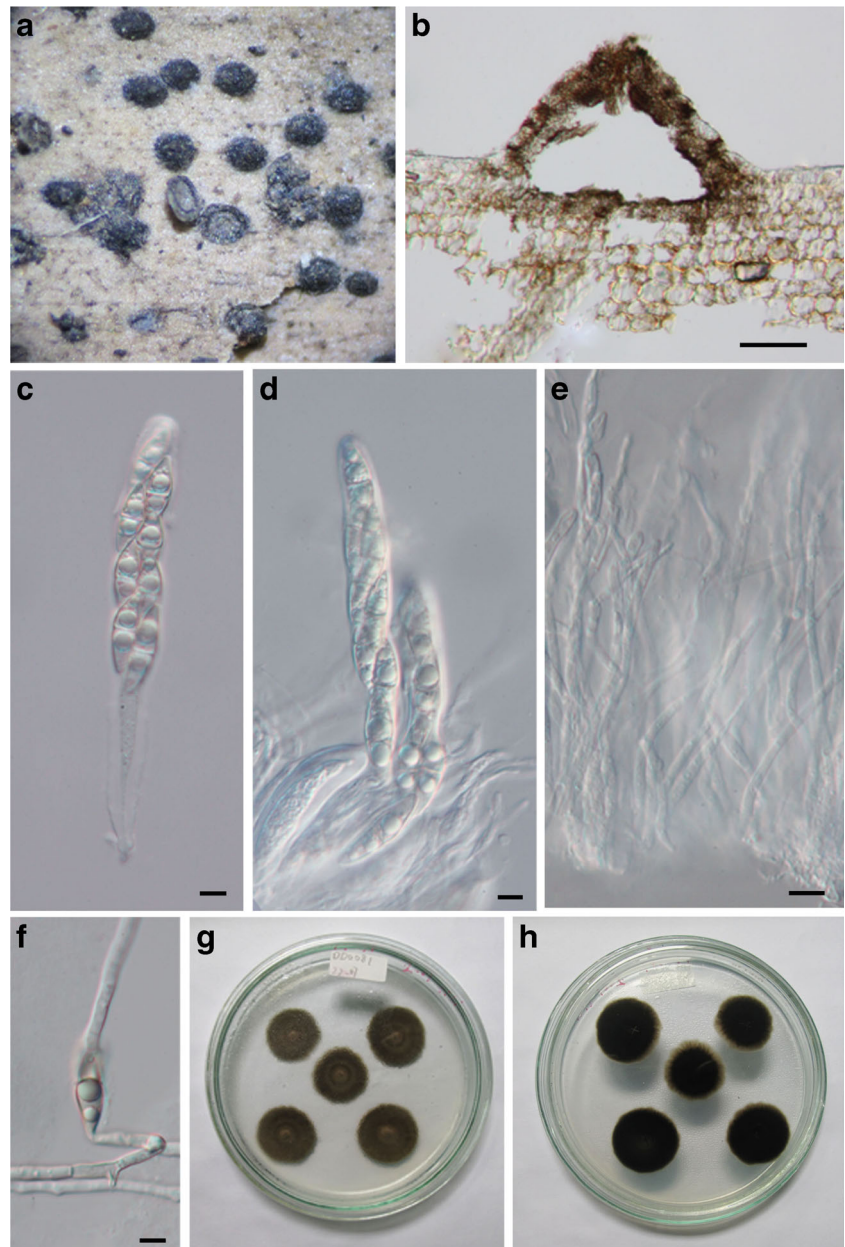
***Seriascoma*** Phookamsak, D.Q. Dai & K.D. Hyde, *gen. nov.*

**Index Fungorum number:** IF552014; ***Facesoffungi* number:** FoF 01978

**Etymology:** The generic epithet “*Seriascoma*” refers to the elongate rows of ascomata which form on the host surface.

**Saprobic** on bamboo culms. **Sexual morph:** *Ascostromata* gregarious, clustered, immersed beneath clypeus, coriaceous, raised, erumpent in linear rows, with slit-like opening, multiloculate, ostiolate. *Locules* arranged in rows, immersed, clustered, gregarious, subglobose to ampulliform, or quadrilateral, glabrous, central ostiole with minute papilla. *Peridium* thick-walled, composed of several layers of small, dark brown pseudoparenchymatous cells of *textura angularis*. *Hamathecium* composed of broad, filamentous, distinctly, septate, cellular, pseudoparaphyses. *Asci* 8-spored, bitunicate, fissitunicate,

**Fig. 16** *Occultibambusa pustula* (MFLU 15–1185, holotype). **a** Appearance of ascostromata on bamboo. **b** Vertical section of ascostromata. **c, d** Asci. **e** Pseudoparaphyses. **f** Germinating ascospores. **h, i** Culture on PDA. Scale bars: b = 50  $\mu\text{m}$ , c–f = 5  $\mu\text{m}$



clavate, long pedicellate, apically rounded, with well-developed, ocular chamber. *Ascospores* overlapping, uni- to tri-septate, clavate to fusiform, with rounded ends, hyaline, 1-septate, slightly constricted at the septum, straight to curved, smooth-walled, with guttules. **Asexual morph:** Coelomycetous, *Conidiomata* eustromatic, solitary to gregarious, immersed in cortex layers, raised, black, elongate on bamboo pieces, conical, uni-loculate, glabrous, covered by vegetative hyphae. *Wall of conidiomata* comprising host and fungi tissue, with several layers composed of dark brown to black, *textura angularis* pseudoparenchymatous cells. *Conidiophores* arising from the basal cavity, reduced to conidiogenous cells. *Conidiogenous cells* holoblastic, phialidic, determinate,

discrete, cylindrical to ampulliform or lageniform, hyaline, aseptate, apically attenuated, with distinct periclinal thickening, smooth-walled. *Conidia* oblong, with rounded to obtuse ends, hyaline, aseptate, smooth-walled.

*Type species: Seriascoma didymospora* Phookamsak, D.Q. Dai & K.D. Hyde

*Notes:* *Seriascoma* is introduced to accommodate Dothideomycete species forming elongated, multiloculate, coriaceous ascostromata, embedded beneath a clypeus, erumpent and splitting the host epidemis parallel with the venation to form a slit like opening. The ascospores are didymosporous, clavate to fusiform and hyaline. The genus is commonly found on bamboo. *Seriascoma* morphologically differs from other genera in the family *Occultabambusaceae*

in the position of its ascostromata under a clypeus. The asexual morph of *Seriascoma* is also different from *Occultabambusa* and *Versicolorisporium*. *Seriascoma* has hyaline, oblong, aseptate conidia, similar to *Occultabambusa*, but the conidia of *Seriascoma* ( $4.9 \times 1.9 \mu\text{m}$ ) are smaller than *Occultabambusa* ( $16.5 \times 5.2 \mu\text{m}$ ). *Versicolorisporium* forms brown, clavate and septate conidia. Based on multi-gene analyses (Fig. 13), *Seriascoma* forms a robust clade, close to *Neooccultabambusa* (Doilom et al. 2016). Ascostroma of *Seriascoma* are similar to those of *Apiospora* which are commonly found on bamboo. However, *Apiospora* has unitunicate asci and belongs to *Sordariomycetes* (Senanayake et al. 2015). Therefore, we propose to introduce the new genus *Seriascoma*.

***Seriascoma didymospora*** Phookamsak, D.Q. Dai, S.C.Karunarathana & K.D. Hyde, *sp. nov.*

*Index Fungorum number*: IF552015; *Facesoffungi number*: FoF 01979; Fig. 17

*Etymology*: The epithet “*didymospora*” refers to the ascospores having two cells.

*Holotype*: MFLU 11–0215

*Saprobic* on bamboo culms. **Sexual morph**: *Ascostromata* 150–320  $\mu\text{m}$  high, 1000–1900  $\mu\text{m}$  diam., gregarious, clustered, immersed beneath clypeus, coriaceous, raised, erumpent in linear rows, with slit-like opening, multi-loculate, ostiolate. *Locules* 120–230  $\mu\text{m}$  high, 180–300  $\mu\text{m}$  diam., arranged in rows, clustered, gregarious, subglobose to ampulliform, glabrous, central ostiole with minute papilla. *Peridium* 15–85  $\mu\text{m}$  wide, thin- to thick-walled, sometimes unequal in thickness, thicker at the sides towards apex, thinner at the base, composed of several layers of small, dark brown pseudoparenchymatous cells of *textura angularis*. *Hamathecium* composed of 2–2.5  $\mu\text{m}$  wide, broad, filamentous, distinctly septate cellular pseudoparaphyses. *Asci* (56–)60–75(–80)  $\times$  8–11(–13)  $\mu\text{m}$  ( $\bar{x}$  =  $68 \times 10 \mu\text{m}$ ,  $n=30$ ), 8-spored, bitunicate, fissitunicate, clavate, long pedicellate, with furcate to obtuse ends, apically rounded, with well-developed ocular chamber. *Ascospores* 11–12(–14.5)  $\times$  3–4  $\mu\text{m}$  ( $\bar{x}$  =  $11.9 \times 3.9 \mu\text{m}$ ,  $n=30$ ), overlapping, uni- to triseriate, didymosporous, clavate to fusiform, with rounded ends, hyaline, slightly constricted at the septum, straight to curved, with upper cell shorter and wider than lower cell, smooth-walled, with large and small guttules. **Asexual morph**: Coelomycetous, produced on bamboo pieces on WA after 4 months. *Conidiomata* 110–170  $\mu\text{m}$  high, 250–470  $\mu\text{m}$  diam., eustromatic, solitary to gregarious, semi-immersed in cortex layers, to erumpent, raised, black, elongate on bamboo pieces, conical, uni-loculate, glabrous, covered by vegetative hyphae. *Wall of conidiomata* 8–80  $\mu\text{m}$  thick, thin- to thick-walled, of unequal thickness, thick at the sides, comprised of host and fungi tissue, with several layers of dark brown to black, pseudoparenchymatous cells of *textura angularis*. *Conidiophores* arising from the base of the cavity,

reduced to conidiogenous cells. *Conidiogenous cells* 4–7(–8)  $\times$  1.5–3  $\mu\text{m}$  ( $\bar{x}$  =  $5.7 \times 2.4 \mu\text{m}$ ,  $n=30$ ), enteroblastic, phialidic, determinate, discrete, cylindrical to ampulliform or lageniform, hyaline, aseptate, apically attenuated, with distinct periclinal thickening, smooth-walled. *Conidia* 4–5.5  $\times$  1.5–2  $\mu\text{m}$  ( $\bar{x}$  =  $4.9 \times 1.9 \mu\text{m}$ ,  $n=30$ ), oblong, with rounded to obtuse ends, hyaline, aseptate, smooth-walled.

*Culture characters*: Colonies on PDA fast growing, 64–73 mm diam. after 4 weeks at 25–30 °C, under 12 h light/12 h dark, colonies circular, dense, flat, slightly raised, smooth with entire edge, velvety to floccose, slightly radiating, aspect smooth, slightly raised at the centre, dull green to dark green from above, blackish green from below.

*Material examined*: THAILAND, Chiang Rai Province, Mae Jun District, Huai Kang Pla waterfall, on dead clumps of bamboo, 25 October 2010, R. Phookamsak, RP0095 (MFLU 11–0215, **holotype**), ex-type living cultures, MFLUCC 11–0179, KUMCC; Muang District, Khun Korn waterfall, on dead culms of bamboo, 13 January 2011, S.C.Karunarathana, RP0110 (MFLU 11–0230, paratype), living cultures, MFLUCC 11–0194, KUMCC.

***Rousoellaceae*** J.K. Liu, et al. *Phytotaxa* 181(1): 7 (2014)

The family *Rousoellaceae*, introduced by Liu et al. (2014), is characterized by immersed, gregarious, clypeate ascostromata, cylindrical, bitunicate asci and 2-celled, brown, ornamented ascospores. Wijayawardene et al. (2014) included five genera, *Appendispora*, *Cytoplea*, *Neorousoella*, *Rousoella* and *Rousoellopsis*, in this family. Members of *Rousoellaceae* mostly occur on monocotyledons, such as bamboo and palms (Hyde et al. 1996; Hyde 1997; Liu et al. 2014). Crous et al. (2014b), however, described a new species, *Rousoella acaciae*, from *Acacia*, a dicotyledonous plant. The asexual morphs of this family are linked to *Cytoplea*, *Melanconiopsis* and *Neomelanconium* (Liu et al. 2014). Ariyawansa et al. (2015) reported a microconidial state occurring in the same conidiomata of *Rousoella* species.

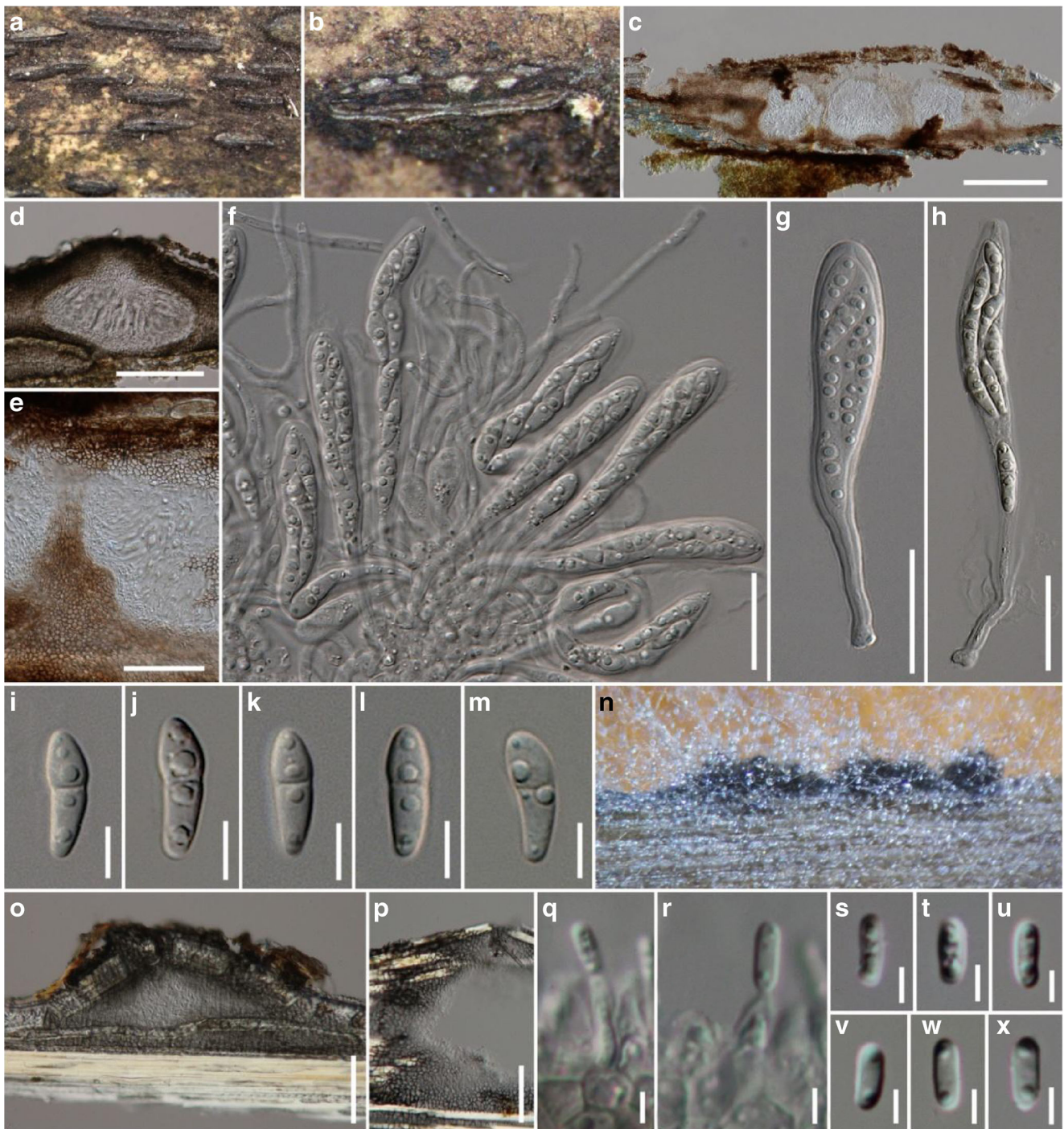
***Rousoella*** Sacc., in Saccardo & Paoletti, *Atti Inst. Veneto Sci. lett.*, ed Arti, Sér. 6 6: 410 (1888)

*Rousoella* was introduced by Saccardo and Paoletti (1888), with the type species *R. nitidula* Sacc. & Paol., which was collected from bamboo in Malaysia. This genus was previously placed in order *Xylariales* (Müller and von Arx 1962), as from the asci and the paraphyses, it was very difficult to distinguish if they were unitunicate or bitunicate. However, Hyde et al. (1996) re-examined the type material and demonstrated that *Rousoella* has bitunicate asci, lacking a blue staining ring. Hyde (1997) reviewed *Rousoella* with a modified key provided. The asexual morph of *Rousoella* links to *Cytoplea* (Liu et al. 2014).

***Rousoella mukdahanensis*** Phookamsak, D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum*: IF552025; *Facesoffungi number*: FoF 01980; Fig. 18





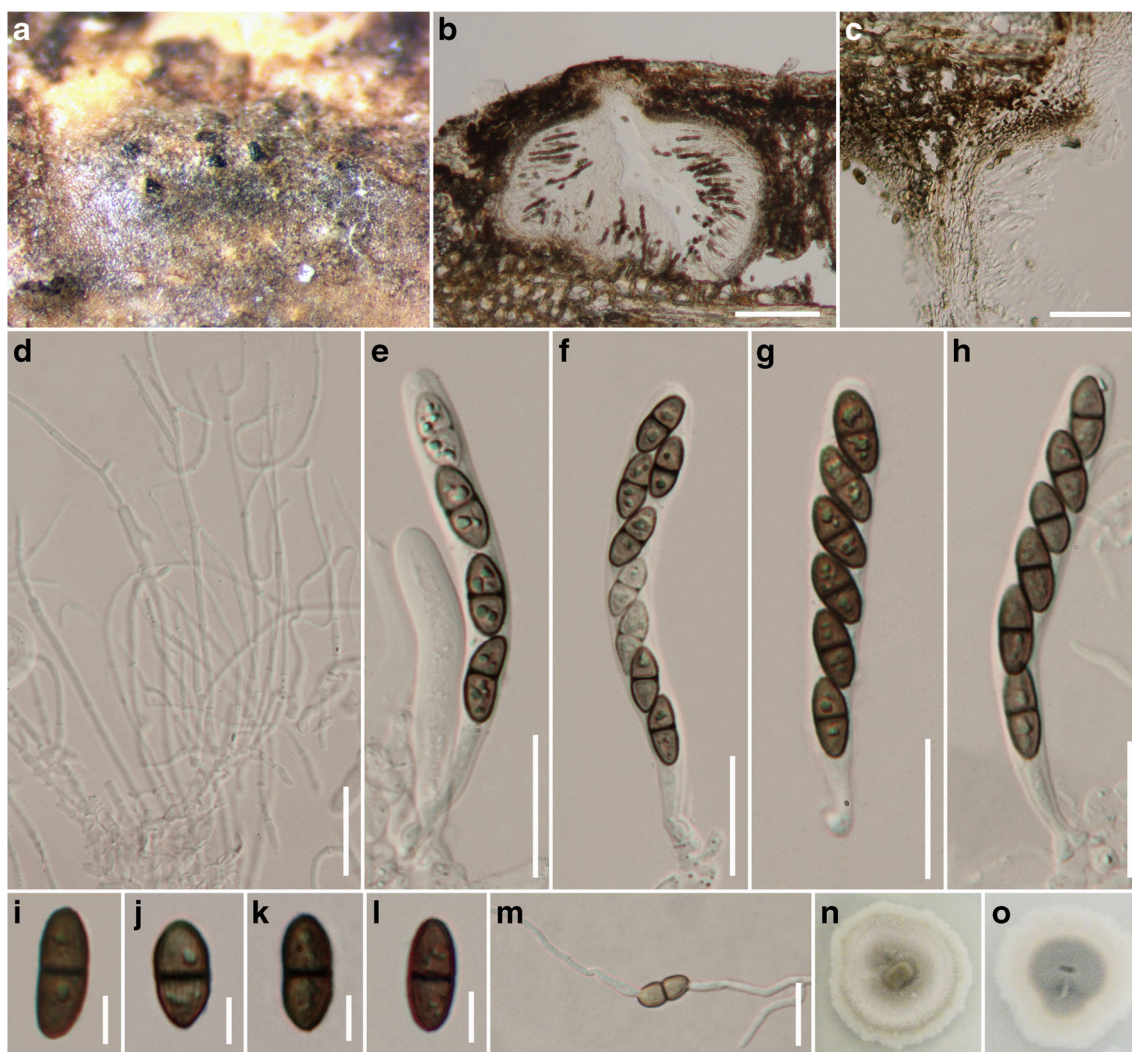
**Fig. 17** *Seriascoma didymospora* (MFLU 11–0215, holotype). **a** Appearance of ascostromata on host surface (MFLU 11–0230). **b** Horizontal section through ascostroma (MFLU 11–0230). **c** Vertical section through ascostroma (MFLU 11–0230). **d** Section through locule (MFLU 11–0215). **e** Section through peridium (MFLU 11–0230). **f** Asci with pseudoparaphyses (MFLU 11–0230). **g–h** Asci (MFLU 11–0230).

**i–m** Ascospores (MFLU 11–0230). Asexual morph from MFLU 11–0215; **n** Rows of conidiomata produced on bamboo pieces on WA. **o** Section through conidiomata. **p** Conidiomata wall. **q–r** Conidiogenous cells. **s–x** Conidia. Scale bars: c = 200  $\mu\text{m}$ , d, o = 100  $\mu\text{m}$ , e, p = 50  $\mu\text{m}$ , f–h = 20  $\mu\text{m}$ , i–m = 5  $\mu\text{m}$ , q–x = 2  $\mu\text{m}$

*Etymology*: The epithet “mukdahanensis” refers to the Mukdahan Province, one of the Northeastern provinces in Thailand, where the fungus was found.

*Holotype*: MFLU 11–0237

*Saprobic* on dead bamboo culms, forming raised, dark brown areas with black dots on host surface. **Sexual morph**: *Ascostromata* pseudostromatic, 280–350  $\mu\text{m}$  high, 520–880  $\mu\text{m}$  diam., solitary to gregarious, immersed, uni-



**Fig. 18** *Roussoella mukdahanensis* (MFLU 11–0237, holotype). **a** Appearance of scostromata on host surface. **b** Vertical section through ascostroma. **c** Section through peridium. **d** Pseudoparaphyses. **e–h** Asci.

**i–l** Ascospores. **m** Germinating ascospore. **n–o** Culture characters (n = upper side, o = lower side). Scale bars: b = 100  $\mu\text{m}$ , c = 50  $\mu\text{m}$ , d–h, m = 20  $\mu\text{m}$ , i–l = 5  $\mu\text{m}$

multi-loculate. *Locules* 250–350  $\mu\text{m}$  high, 300–420  $\mu\text{m}$  diam., scattered to clustered, forming groups immersed in ascostroma to erumpent through host surface by papilla, globose to subglobose, glabrous, ostioles central, with minute papilla, periphyses. *Peridium* 15–40  $\mu\text{m}$  wide, thin-walled, of equal thickness, comprising several layers of flattened, brown to dark brown, *textura angularis* to *textura prismatica* pseudoparenchymatous cells, *textura epidermoidea* beside papilla. *Hamathecium* composed of dense, 1.5–2.5  $\mu\text{m}$  wide, cellular pseudoparaphyses, distinctly septate, not constricted at the septa, branched, anastomosing at the apex, embedded in a gelatinous matrix. *Asci* (65–)70–80(–89)  $\times$  (7–)8–10  $\mu\text{m}$  ( $\bar{x}$  = 77.9  $\times$  8.8  $\mu\text{m}$ ,  $n$  = 30), 4–6–8-spored, bitunicate, cylindrical to cylindrical-clavate, short to long pedicellate with furcate or knob-like pedicel, apically rounded with indistinct ocular chamber. *Ascospores* (10.5–)11–13(–14)  $\times$  (4–)5–6  $\mu\text{m}$  ( $\bar{x}$  = 12.4  $\times$  5.4  $\mu\text{m}$ ,  $n$  = 30),

overlapping, uni-seriate, ellipsoidal to fusiform, with rounded to acute ends, brown to dark brown, 1-septate, not constricted at the septum, rough-walled, striate or longitudinally ribbed. **Asexual morph**: Undetermined.

*Culture characters*: Colonies on MEA slow growing, 38–40 mm diam. after 4 weeks at 25–30  $^{\circ}\text{C}$ , under 12 h light/12 h dark, colonies irregular, dense, slightly raised to umbonate, dull with undulate edge, floccose to velvety, slightly radiating, the margin separated by yellowish brown concentric ring, white to cream at the margin, white to grey outwards towards the centre, grey at the centre from above, white to cream at the margin, grey to dark grey at the centre, separated from margin by orangish brown concentric ring from below, not producing pigments.

*Material examined*: THAILAND, Mukdahan Province, Nongsung District, Wang Hai village, on dead culms of bamboo, 13 April 2011, R. Phookamsak, RP0117 (MFLU 11–

0237, **holotype**); *Ibid.* (BBH **isotype**), ex-type living cultures, MFLUCC 11–0201, KUMCC.

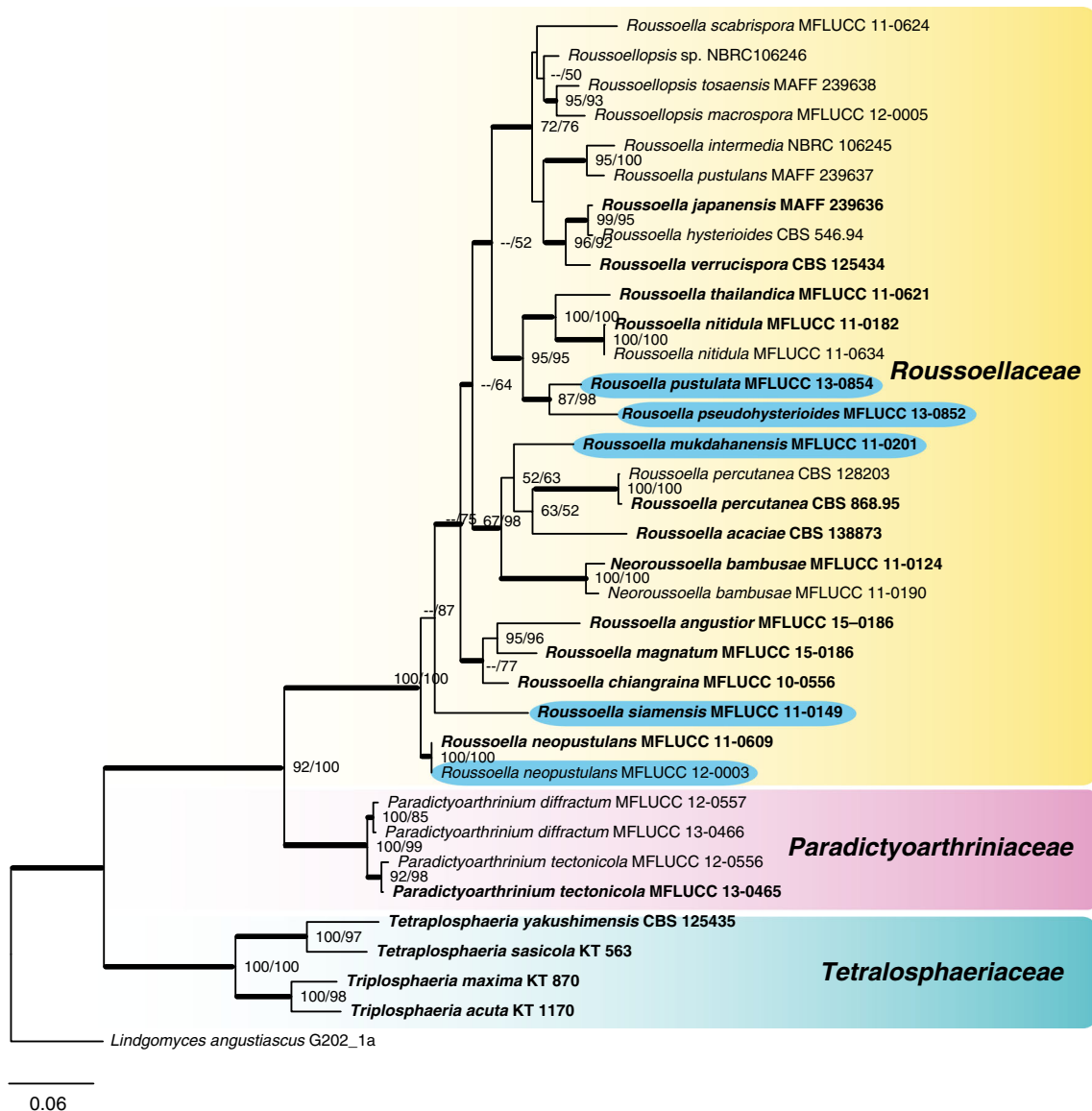
*Notes:* *Rousoella mukdahanensis* is similar to *R. donacicola*, *R. minutella*, *R. munkii*, *R. neopustulans* and *R. pustulans* in the size of asci and ascospores, however, they can be differentiated by their ascostroma size and morphology on the host surface (Saccardo 1891; Aptroot 1995; Ju et al. 1996; Hyde 1997; Liu et al. 2014). *Rousoella mukdahanensis* is most similar to *R. neopustulans* and *R. pustulans* in sharing the size range of asci and ascospores and papillate ostioles in *R. neopustulans* (Liu et al. 2014). Based on the multi-gene phylogenetic analyses (Fig. 19), *Rousoella mukdahanensis* distinctly separates from the morphologically similar species.

Liu et al. (2014) mentioned that *Rousoella* species were barely distinguishable by morphological characters, however, they can be separated by phylogenetic investigation.

*Rousoella neopustulans* D.Q. Dai, J.K. Liu & K.D. Hyde, in Liu et al. Phytotaxa 181(1): 1–33 (2014)

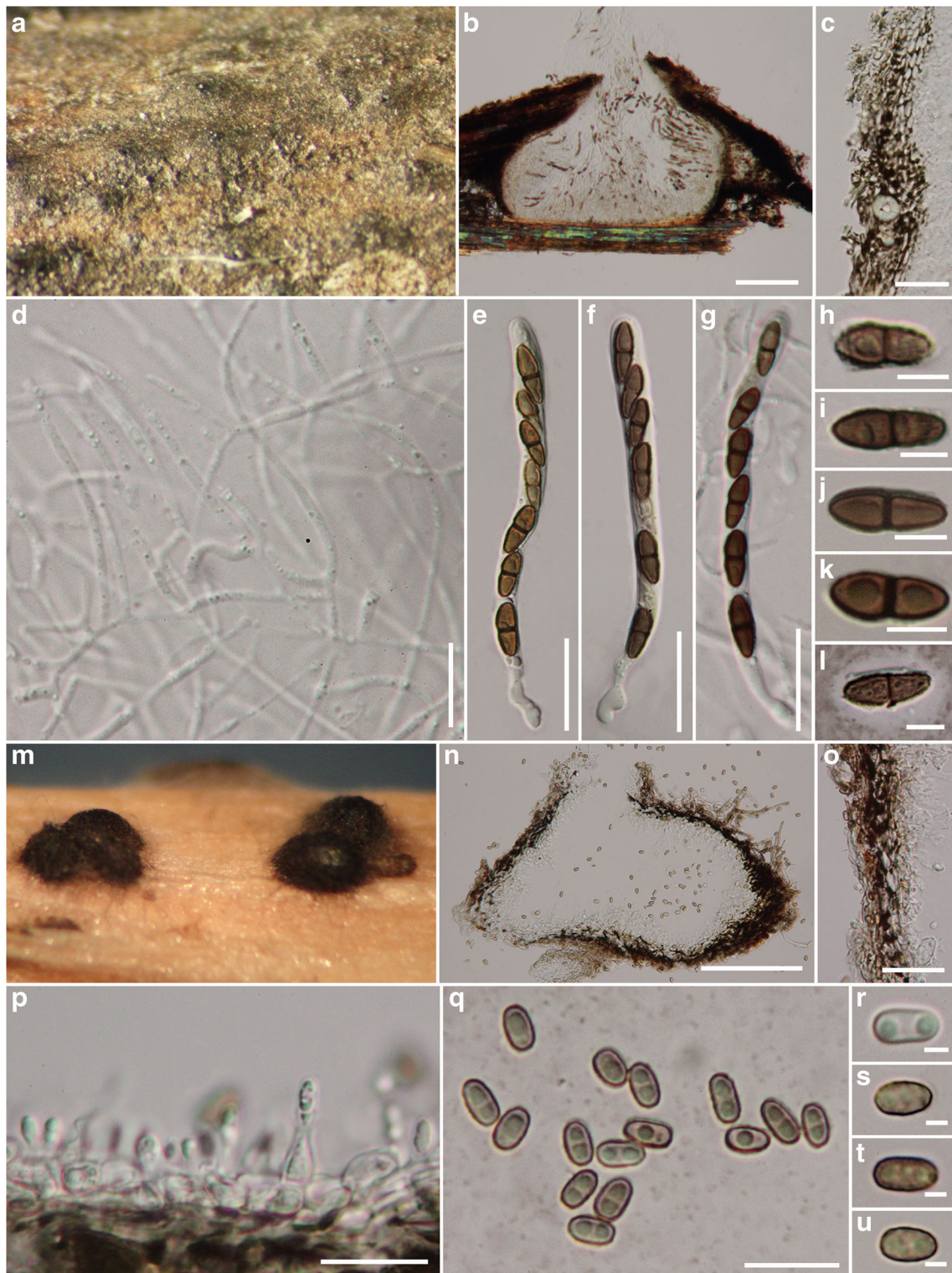
*Facesoffungi* number: FoF 01981; Fig. 20

*Saprobic* on dead culms of bamboo, forming raised, brown to dark brown area, with black dots on host surface. **Sexual morph:** *Ascostromata* 210–280 µm high, 300–400 µm diam., gregarious, immersed to erumpent through host surface by papilla, globose to subglobose, uni-loculate, glabrous, with a central ostiole, with minute papilla. *Peridium* 9.5–25.5 µm wide, thin-walled, of unequal thickness, slightly thinner at



**Fig. 19** Maximum likelihood phylogenetic tree generated by RAxML (GTR+G model) based on combined SSU, LSU and TEF1 sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes, and branches with Bayesian posterior probabilities

greater than 0.90 are given in bold. The original isolate numbers are noted after the species names. The tree is rooted to *Lindgomyces angustiascus* (G202\_1a). Ex-type strains are in bold and newly generated sequences are highlighted with a blue background



**Fig. 20** *Roussoella neopustulans* (MFLU 11–0241). **a** Appearance of ascostromata on host surface. **b** Vertical section through ascostroma. **c** Section through peridium. **d** Pseudoparaphyses. **e–g** Asci. **h–k** Ascospores. **l** Ascospore stained with Indian ink. **m** Conidiomata on

bamboo pieces on WA. **n** Vertical section through conidioma. **o** Section through pycnidial wall. **p** Conidiogenous cells. **q** Conidia stained with Indian ink. **r–u** Conidia. Scale bars: **b, n** = 100  $\mu\text{m}$ , **c, e, f, g, o** = 20  $\mu\text{m}$ , **d, p, q** = 10  $\mu\text{m}$ , **h–l** = 5  $\mu\text{m}$ , **r–u** = 2  $\mu\text{m}$

the base, composed of several layers of flattened, brown to dark brown, *textura prismatica* pseudoparenchymatous cells,

rarely *textura angularis*. *Hamathecium* composed of dense, 2–2.5  $\mu\text{m}$  wide, cellular pseudoparaphyses, distinctly septate,

constricted at the septum, anastomosing at the apex, embedded in a gelatinous matrix. *Asci* (63–)75–90(–96) × 5–6(–7)  $\mu\text{m}$  ( $\bar{x}$  = 80.1 × 5.9  $\mu\text{m}$ ,  $n$  = 30), 8-spored, bitunicate, cylindrical, short pedicellate, with furcate or knob-like pedicel, apically rounded with indistinct ocular chamber. *Ascospores* (8–)9–12 × 3–4(–5)  $\mu\text{m}$  ( $\bar{x}$  = 10.3 × 3.8  $\mu\text{m}$ ,  $n$  = 30), overlapping, uni-seriate, ellipsoidal to fusiform, with rounded to acute ends, brown to dark brown, 1-septate, slightly constricted at the septum, rough-walled, with striations or longitudinally ribbed, surrounded by mucilaginous sheath. **Asexual morph:** produced on bamboo pieces on WA after 12 weeks. *Conidiomata* 140–300  $\mu\text{m}$  high, 170–320  $\mu\text{m}$  diam., pycnidial, solitary to gregarious, clustered, superficial, visible as blackdots on bamboo pieces, variable in shape, globose to subglobose, lenticular, dome-shaped to irregular-shaped, uni- to multi-loculate, covered by hyphae, ostiole central, with pore-like opening. *Wall of conidiomata* 12–25.5  $\mu\text{m}$  wide, thin-walled, of equal thickness, composed of two types cell layers, inner part comprising several layers of flattened, dark brown to black, *textura angularis* pseudoparenchymatous cells, barely distinguishable from conidiogenous cells, outer part comprising a few layers of *textura epidermoidea* to *textura intricata*. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* (2.5–)3–6 × (2–)3–5  $\mu\text{m}$  ( $\bar{x}$  = 4.8 × 3.9  $\mu\text{m}$ ,  $n$  = 30), enteroblastic, annellidic and phialidic, determinate, discrete, unbranched, ampulliform to lageniform, hyaline, aseptate, smooth-walled. *Conidia* 4–5.5(–6) × 2–3  $\mu\text{m}$  ( $\bar{x}$  = 5.2 × 3  $\mu\text{m}$ ,  $n$  = 30), oblong to ellipsoidal, with rounded to obtuse ends, initially hyaline, becoming brown, aseptate, verrucose.

**Culture characters:** Colonies on PDA fast growing, 30–40 mm diam. after 2 weeks at 25–30 °C, colonies irregular, sparse to medium dense, slightly raised to umbonate, surface rough, with entire edge, woolly at the margin, floccose to velvety in the centre, wrinkled at the centre, colonies white at the margin, grey to greenish grey in the middle, with white-grey at the centre from above, white to cream at the margin, brown to dark brown at the centre from below, not producing pigments.

**Material examined:** THAILAND, Mukdahan Province, Nongsung District, Wang Hai Village, on dead culms of bamboo, 13 April 2011, R. Phookamsak, RP0121 (MFLU 11–0241), living cultures, MFLUCC 12–0003, KUMCC.

**Notes:** *Rousoella neopustulans* was introduced by Liu et al. (2014), with a description of only the sexual morph. This species was recollected in the same country, and the new collection sporulated on bamboo pieces on water agar. Thus this species is redescribed and illustrated here with both sexual and asexual morphs.

This species is morphologically similar to *R. pustulans*. However, *R. neopustulans* has smaller ascospores but larger asci (Liu et al. 2014). Additionally, *R. neopustulans* ascospores have linear striations along their entire length, while those of *R. pustulans* have

shorter and irregular striations. Based on multi-gene phylogenetic analyses (Fig. 19), *R. neopustulans* forms a distinct basal clade in the family *Rousoellaceae*. *Rousoella pustulans* has no known asexual morph.

***Rousoella pseudohysterioides*** D.Q. Dai & K.D. Hyde, *sp. nov.*

**Index Fungorum number:** IF552026; **Facesoffungi number:** FoF 01982; Fig. 21

**Etymology:** Refers to the similarity with *Rousoella hysterioides*.

**Holotype:** MFLU 15–1209

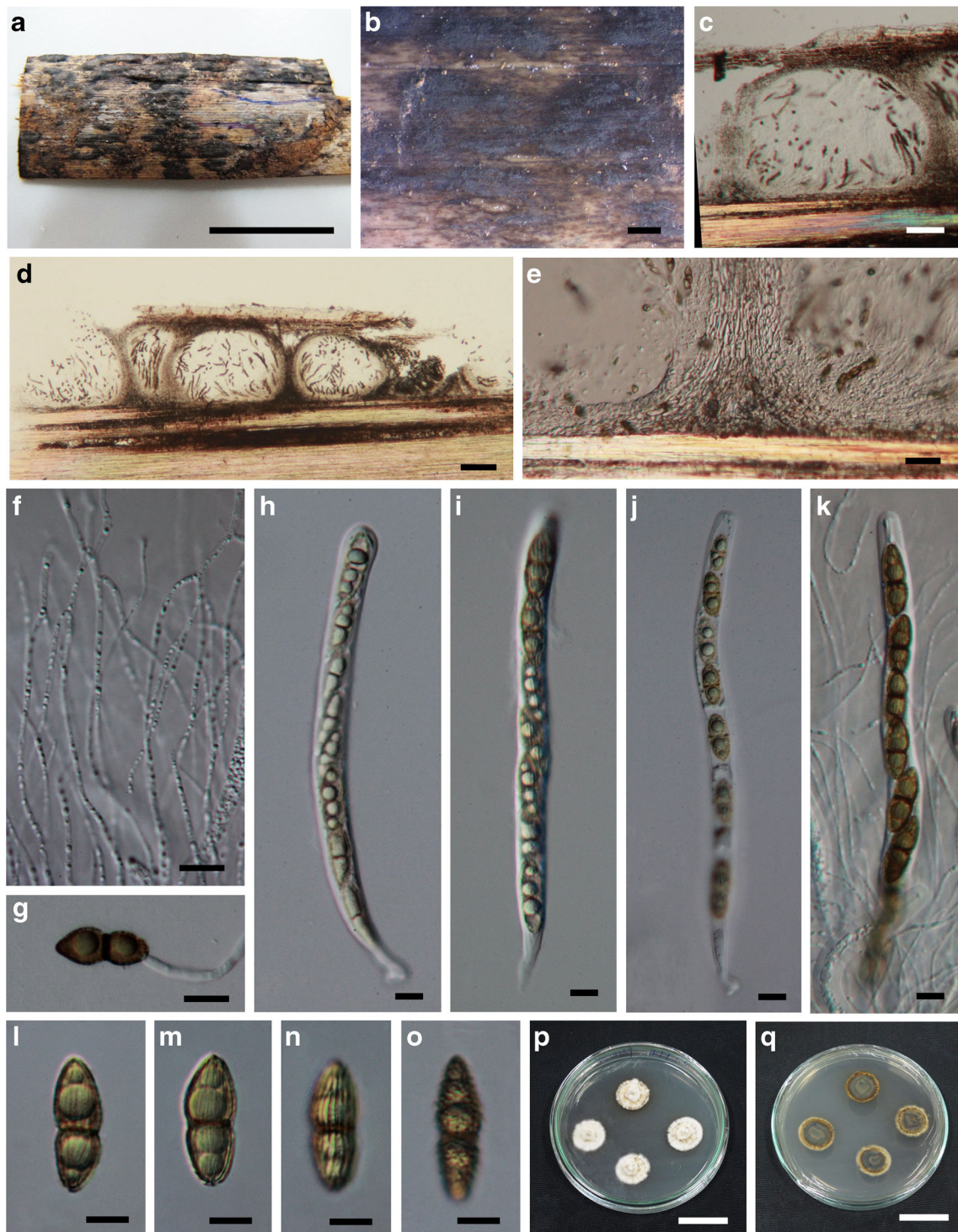
**Saprobic** on decaying bamboo culms. **Sexual morph:** *Ascstromata* forming under black area, up to 3–5 mm long and 1–2 mm wide, raised at maturity, ellipsoidal to irregular, black, coriaceous. *Locules* in vertical section 200–250  $\mu\text{m}$  high, 170–350  $\mu\text{m}$  diam., solitary to gregarious, subglobose to ellipsoidal, dark brown, with ostiolate opening. *Peridium* 15–25  $\mu\text{m}$  wide, composed of dark brown to hyaline cells of compressed *textura angularis*, with upper wall 15–45  $\mu\text{m}$  wide, darker, comprising host and fungal tissues. *Hamathecium* comprising dense, 2–3.5  $\mu\text{m}$  wide, cellular pseudoparaphyses, indistinctly septate, anastomosing and branching at the apex, embedded in a gelatinous matrix. *Asci* 100–270 × 7–15  $\mu\text{m}$  ( $\bar{x}$  = 148.2 × 9.7  $\mu\text{m}$ ,  $n$  = 20), 8-spored, bitunicate, cylindrical, with a short furcate pedicel, with an apical ocular chamber. *Ascospores* 12.5–20 × 4–6  $\mu\text{m}$  ( $\bar{x}$  = 16.1 × 5.2  $\mu\text{m}$ ,  $n$  = 20), uniseriate, fusiform-ellipsoidal, 1-septate, constricted at the septum, narrow at both ends, with longitudinally striate and verrucose wall ornamentation. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from lower end or both ends. Colonies circular, slow growing, 25 mm diam. in 30 days at 28–32 °C, under 12 h light/12 h dark, cottony, yellowish brown at the centre, light coloured and floccose at the periphery from above, yellow to dark brown to orange from below.

**Material examined:** THAILAND, Chiang Rai Province, Mae Fah Luang University, on dead culm of bamboo, 9 July 2013, Dong-Qin Dai DDQ00259 (MFLU 15–1209); *Ibid.* (KUN HKAS88717, **isotype**), ex-type living cultures, MFLUCC 13–0852, CBS 139992.

**Note:** *Rousoella pseudohysterioides* is similar to *R. hysterioides* in having black stromatic ascomata, cylindrical asci and fusiform-ellipsoidal ascospores with longitudinal striations (Hyde et al. 1996). *Rousoella pseudohysterioides*, however, differs by its smaller ascospores (12.5–20 × 4–6  $\mu\text{m}$  vs. 18–34 × 6–8  $\mu\text{m}$ ) (Hyde et al. 1996). Moreover, *R. pseudohysterioides* has verrucose ascospores, a character not observed in *R. hysterioides*. In the phylogenetic tree (Fig. 19) *R. pseudohysterioides* is close to *R. pustulata*, while the branch length indicates that they are different species.

***Rousoella tuberculata*** D.Q. Dai & K.D. Hyde, *sp. nov.*



**Fig. 21** *Roussoella pseudohysterioides* (MFLU 15–1209, holotype). **a**, **b** Ascostromata developing on bamboo culm. **c**, **d** Vertical sections of ascostromata. **e** Peridium. **f** Branched pseudoparaphyses. **g** Germinating

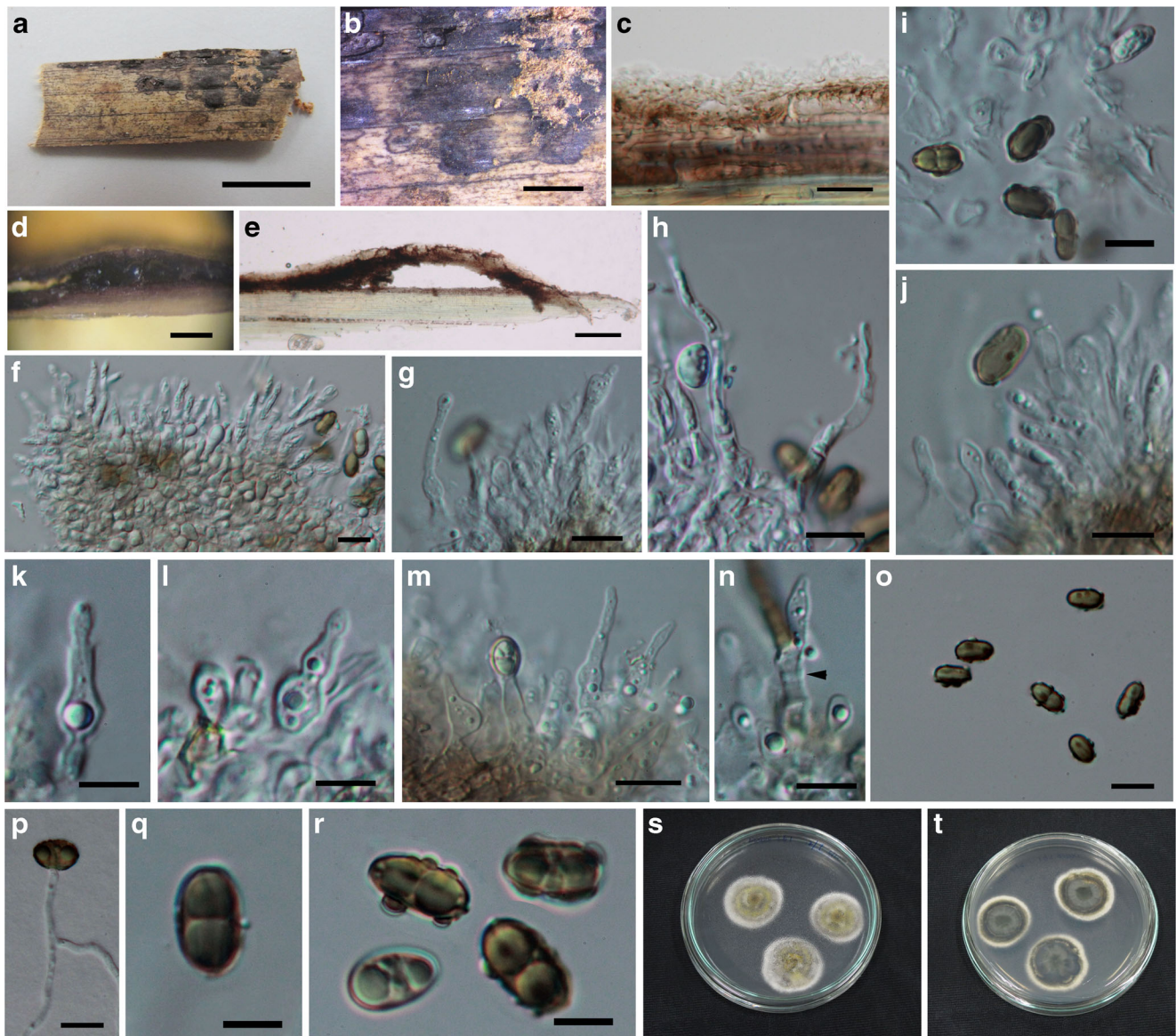
ascospore. **h–k** Asci containing eight ascospores. **l–o** Dark brown ascospores. **p**, **q** Cultures on MEA. Scale bars: **a** = 2 cm, **b** = 1 mm, **c** = 50  $\mu\text{m}$ , **d** = 100  $\mu\text{m}$ , **e** = 20  $\mu\text{m}$ , **f–k** = 10  $\mu\text{m}$ , **l–o** = 5  $\mu\text{m}$ , **p**, **q** = 25 mm

*Index Fungorum* number: IF552027; *Facesoffungi* number: FoF 01983; Fig. 22

*Etymology*: Refers to the conidia covered by tubercles.

*Holotype*: MFLU 15–1211

*Saprobic* on decaying bamboo culms. **Sexual morph**: Undetermined. **Asexual morph**: *Stromata* forming under a blackened area, up to 1–2 mm long and 2–3 mm wide, and becoming raised at maturity, ellipsoidal to irregular.



**Fig. 22** *Roussoella tuberculata* (MFLU 15–1211, holotype). **a, b** Conidiomata on bamboo culm. **c** Basal wall of conidioma. **d, e** Vertical sections of conidiomata. **f, g, i–n** Conidiogenous cells developing conidia (**n**: showing annellidic conidiogenous cells). **h** Paraphyses. **p**

Germinating conidia. **o, q, r** Dark brown conidia (**o, r**: Mature conidia covered by small tubercles). **s, t** Cultures on MEA. Scale bars: **a** = 2 cm, **b** = 2 mm, **b, d, e** = 200  $\mu\text{m}$ , **c** = 20  $\mu\text{m}$ , **f–j, m, o** = 10  $\mu\text{m}$ , **k, l, n, p–r** = 5  $\mu\text{m}$

*Conidiomata* 150–250  $\mu\text{m}$  high, 500–800  $\mu\text{m}$  diam., eustromatic, immersed in the stromata, globose to subglobose, dark brown, with ostiolate opening. *Conidiomatal wall* comprising two layers of cells of *textura angularis*, with dark to brown outer thin layer 5  $\mu\text{m}$  thick, with 10–17  $\mu\text{m}$  thick, hyaline, conidiogenous inner layer. *Paraphyses* 30–40  $\mu\text{m}$  long  $\times$  2–3.5  $\mu\text{m}$  wide, septate, wide at the base, curved. *Conidiophores* 5–12.5  $\times$  2.5–4  $\mu\text{m}$  ( $\bar{x}$  = 7.4  $\times$  3.6  $\mu\text{m}$ ,  $n$  = 20) short, hyaline, wide at base, smooth, formed from the inner cells of the conidiomata wall. *Conidiogenous cells* 2.5–12  $\times$  1.5–3.5  $\mu\text{m}$  ( $\bar{x}$  = 8.1  $\times$  2.5  $\mu\text{m}$ ,  $n$  = 20) obclavate, enteroblastic, phialidic, becoming annellidic, with 1–3 barely

discernible annellations, indeterminate, discrete, cylindrical to ampulliform, hyaline, smooth-walled. *Conidia* 8.5–10  $\times$  4.5–5.5  $\mu\text{m}$  ( $\bar{x}$  = 9.5  $\times$  5.2  $\mu\text{m}$ ,  $n$  = 20), ellipsoidal to oblong, occasionally obovoid, aseptate, straight, rounded at both ends, brown to dark brown, smooth-walled, with wall sparsely covered by small, roughened tubercles, inside usually containing 1–2 large guttules.

*Culture characters*: Conidia germinating on MEA within 24 h and germ tubes produced from sides. Colonies slow growing, 30 mm diam. after 15 days at 28–32  $^{\circ}\text{C}$ , under 12 h light/12 h dark, circular, with even margin, floccose at the center, drift white at margin, and light greenish brown at the centre.

**Material examined:** THAILAND, Chiang Rai Province, Mae Fah Luang University, on dead culms of bamboo, 9 July 2013, Dong-Qin Dai, DDQ00261 (MFLU 15–1211, **holotype**); *Ibid.* (KUN HKAS88719, **isotype**), ex-type living cultures, MFLUCC 13–0854, ICMP 20529.

**Notes:** *Roussoella tuberculata* is characterized by immersed, eustromatic conidiomata, enteroblastic, phialidic, annellidic conidiogenous cells and ellipsoidal conidia covered by small tubercles. *Roussoella tuberculata* is similar to *R. hysterioides*, but differs in having annellidic conidiogenous cells (Hyde et al. 1996). Conidia of *Roussoella* species usually have verrucose wall ornamentation (Hyde et al. 1996; Liu et al. 2014), while *R. tuberculata* has conidia with small, roughened tubercles.

***Roussoella siamensis*** Phookamsak, J.K. Liu & K.D. Hyde., in Liu et al. *Phytotaxa* 181(1): 1–33 (2014)

**Facesoffungi number:** FoF 01984; Figs. 23

**Saprobic** on dead bamboo culms. **Sexual morph:** *Ascstromata* 120–150  $\mu\text{m}$  high, 620–750  $\mu\text{m}$  diam., gregarious, immersed within cortex, visible as raised, brown to dark brown areas on the host surface, shield-shaped to continuously low convex, uni- to multi-loculate, coriaceous. *Locules* 70–120  $\mu\text{m}$  high, 130–350  $\mu\text{m}$  diam., scattered to clustered, gregarious, immersed in ascstromata, lenticular to subglobose, with a flattened base, glabrous, ostiole central, with a minute ostiole embedded beneath a clypeus, becoming pore-like opening on host surface at maturity. *Peridium* 7–15  $\mu\text{m}$  wide, thin-walled of unequal thickness, thinner at the base, composed of several layers of flattened, brown to dark brown, compressed *textura angularis* pseudoparenchymatous cells. *Hamathecium* composed of dense, 1–2  $\mu\text{m}$  wide, cellular pseudoparaphyses, indistinctly septate, not constricted at the septum, anastomosing and branching at the apex, embedded in a gelatinous matrix. *Asci* (67–)70–90(–94)  $\times$  6–8  $\mu\text{m}$  ( $\bar{x}$  = 77.8  $\times$  7.3  $\mu\text{m}$ ,  $n=25$ ), 8-spored, bitunicate, cylindrical, short to long pedicellate with furcate or foot-like pedicel, apically rounded, with an indistinct ocular chamber. *Ascospores* (8.5–)10–12(–14)  $\times$  4–5  $\mu\text{m}$  ( $\bar{x}$  = 10.8  $\times$  4.6  $\mu\text{m}$ ,  $n=30$ ), overlapping, uni-seriate, ellipsoidal to fusiform, with rounded ends, brown to dark brown, 1-septate, constricted at the septum, rough-walled, longitudinally ribbed. **Asexual morph:** Coelomycetous, produced on bamboo pieces on WA after 12 weeks. *Conidiomata* 360–940  $\mu\text{m}$  high, 560–1300  $\mu\text{m}$  diam., solitary to gregarious, clustered, semi-immersed to superficial, visible as black irregular pycnostroma, covered by hyphae on bamboo pieces, globose to subglobose, uni-loculate, becoming glabrous at maturity, indistinctly ostiolate. *Conidiomata wall* 20–45  $\mu\text{m}$  wide, thin-walled, of equal thickness, composed of four strata, an inner layer comprising conidiogenous cells arising from a thin layer of hyaline

cells, a stratum of 3–5 layers of dark brown to black, pseudoparenchymatous cells, arranged in a *textura angularis*, a median stratum comprising several layers of flattened, hyaline cells, arranged in *textura angularis* to *textura prismatica*, and an outer stratum comprising several layers of dark brown to black stromatic cells, arranged in *textura intricata*. *Conidiophores* arising from the basal cavity around conidiomata, unbranched, 0–1 septate, or reduced to conidiogenous cells. *Conidiogenous cells* 3–6(–10)  $\times$  2.5–5  $\mu\text{m}$  ( $\bar{x}$  = 5.3  $\times$  3.7  $\mu\text{m}$ ,  $n=30$ ), enteroblastic, phialidic or broadly annellidic, determinate, discrete, unbranched, ampulliform to lageniform, or irregular in shape, hyaline, aseptate, smooth-walled, with distinct periclinal thickening. *Conidia* 4–5  $\times$  3–4  $\mu\text{m}$  ( $\bar{x}$  = 4.8  $\times$  3.4  $\mu\text{m}$ ,  $n=50$ ), globose to subglobose, initially hyaline, becoming brown, aseptate, rough-walled, verrucose.

**Culture characters:** Colonies on PDA fast growing, 55–60 mm diam. after 4 weeks at 25–30 °C, colonies irregular, medium dense, slightly raised to umbonate, dull with undulate edge, floccose to woolly, slightly radiating, pale yellowish to brownish orange at the margin, white to cream medianly, pale yellowish at the centre from above, pale yellowish at the margin, brown to golden brown medianly, dark brown to black at the centre from below, produced yellowish brown pigment in agar.

**Material examined:** THAILAND, Chiang Rai Province, Muang District, Mae Fah Luang University campus grounds, on dead culms of *Bambusa* sp., 13 August 2010, R. Phookamsak, RP0065 (MFLU 11–0185, **holotype**), ex-type living cultures, MFLUCC 11–0149, KUMCC.

**Notes:** *Roussoella siamensis* was introduced by Liu et al. (2014), and lacked the asexual morph description. In this study, the ex-type culture produced a coelomycetous asexual morph on bamboo pieces on water agar. Thus, we redescribe and illustrate this species based on morphological characters of both sexual and asexual morphs.

The species was described as forming multi-loculate ascromata with cylindrical, bitunicate asci and brown, striate, didymosporous ascospores (Liu et al. 2014). Based on the morphological characters, *Roussoella siamensis* is similar to *R. chiangraina* and *R. neopustulans* (Liu et al. 2014). However, *R. siamensis* differs from *R. chiangraina* and *R. neopustulans* by forming multi-loculate ascstromata, and having larger asci, ascospores and conidia than *R. chiangraina* and *R. neopustulans*. Based on multi-gene phylogenetic analyses, *R. siamensis* forms a single basal clade (Fig. 19).

***Tubeufiales*** Boonmee & K.D. Hyde, *Fungal Diversity* 68 (1): 245 (2014)

The order *Tubeufiales* was introduced by Boonmee et al. (2014), and accommodates a single family *Tubeufiaceae* (Boonmee et al. 2011). Boonmee et al. (2014) examined the sexual and asexual morphs, and provided a key to the 19 accepted genera (including five asexual genera).





**Fig. 23** *Rousoella siamensis* (MFLU 11–0185, holotype). **a** Appearance of ascostromata on host surface. **b** Vertical section through ascostroma. **c** Section through peridium between locules. **d** Pseudoparaphyses. **e–h** Asci. **i–l** Ascospores. **m** Germinating ascospore.

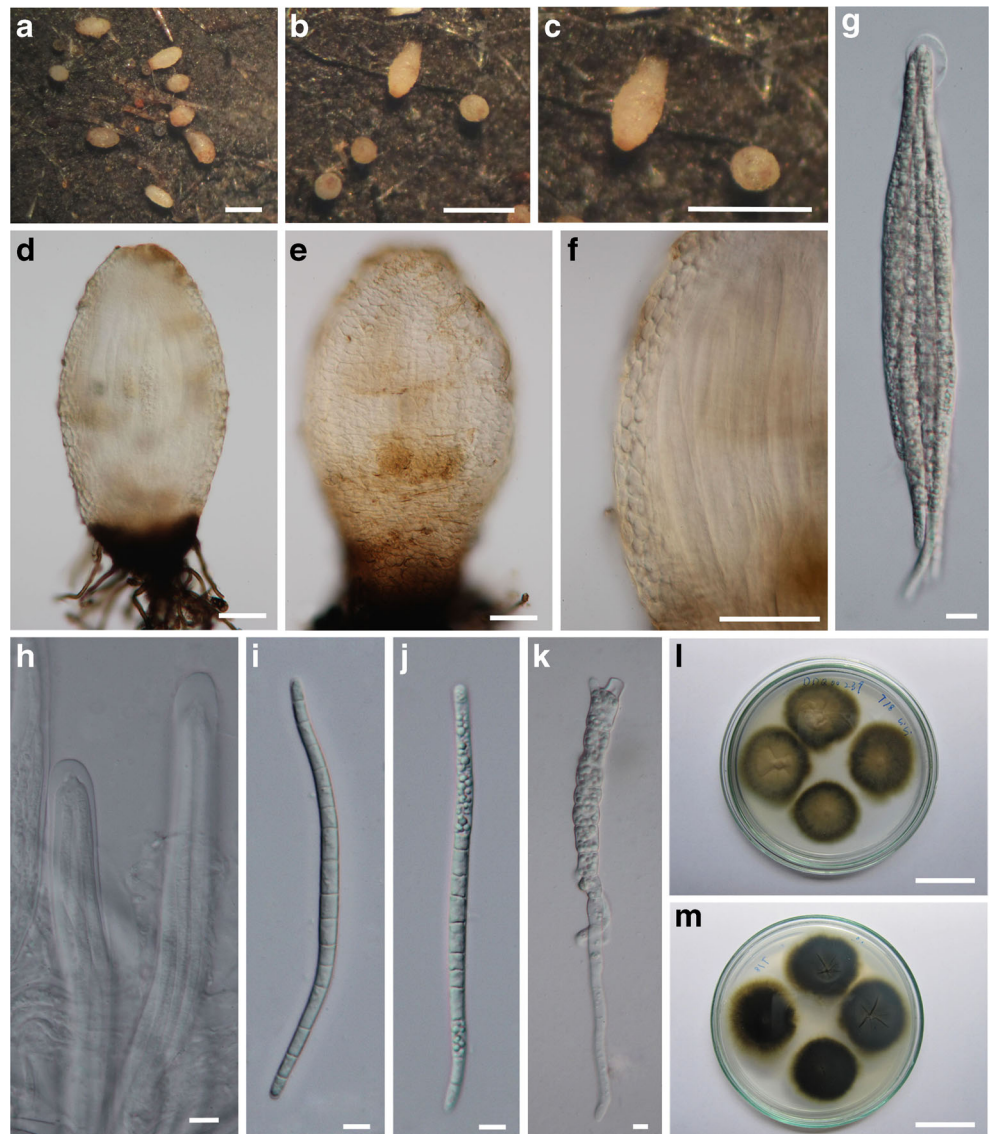
**n** Conidiomata on bamboo pieces on WA. **o** Vertical section through conidioma. **p** Section through pycnidial wall. **q–s** Conidiogenous cells. **t–w** Conidia. Scale bars: **b**, **o** = 100  $\mu\text{m}$ , **c–h**, **p** = 20  $\mu\text{m}$ , **m** = 10  $\mu\text{m}$ , **i–l**, **q–s**, **t** = 5  $\mu\text{m}$ , **u–w** = 2  $\mu\text{m}$

Subsequently, Wijayawardene et al. (2014) recognized 21 genera in *Tubeufiaceae*.

*Tubeufia javanica* Penz. & Sacc., Malpighia 11(11–12): 517 (1898) [1897], Fig. 24

*Material examined*: THAILAND, Chiang Rai, Muang, Khun Korn Waterfall, on dead culm-sheaths of bamboo, 31 July 2012, Dong-Qin Dai DDQ00239 (MFLU 13–0371, **epitype**); *Ibid.* (PDD 104450, iso-epitype).

**Fig. 24** *Tubeufia javanica* (MFLU 13–0371, **epitype**). **a–c** Oblong to clavate ascomata on host surface. **d** Ascoma forming on dark brown hyphae. **e, f** Peridium of ascomata **g** Broken ascus. **h** Cylindrical asci with ocular chambers. **i, j** Hyaline to pale brown, filiform, multi-septate ascospores. **k** Germinating ascospore. **l, m** Dark brown colonies on PDA media. Scale bears: **a–c** = 400  $\mu\text{m}$ , **d–f** = 50  $\mu\text{m}$ , **g, h** = 10  $\mu\text{m}$ , **i–k** = 5  $\mu\text{m}$ , **l, m** = 25 mm



**Notes:** *Tubeufia javanica*, the family type of *Tubeufiaceae*, was described by Penzig and Saccardo (1897) based on a specimen collected from bamboo. It was recollected, epitypified and redescribed in Boonmee et al. (2014). In this paper, we reillustrate this important species to show the typical morphology of *Tubeufiaceae*.

***Tubeufia longiseta*** D.Q. Dai & K.D. Hyde, *sp. nov.*

**Index Fungorum umbers:** IF552029; **Facesoffungi number:** FoF 01985; **Fig. 25**

**Etymology:** In reference to its ascomata with long setae.

**Holotype:** MFLU 15–1215

**Saprobic** on bamboo culm. **Sexual morph:** *Ascomata* 250–370  $\mu\text{m}$  high, 180–250  $\mu\text{m}$  diam., superficial, solitary, gregarious or in loose groups, subglobose, obovoid, black, with a central ostiole. *Setae* 100–350  $\times$  3.5–5.5  $\mu\text{m}$ , dense, covering the whole ascoma, black, unbranched, septate, dark brown, thick-walled. *Peridium* 25–40  $\mu\text{m}$  wide, composed of

7–11 layers of cells of *textura angularis*; outer stratum composed of large, dark brown, thick-walled, 11–21  $\times$  6.5–14.5  $\mu\text{m}$  cells; inner stratum composed of small, hyaline, thin-walled cells. *Hamathecium* comprising 2–2.5  $\mu\text{m}$  wide, septate, branched, hyaline, cellular pseudoparaphyses embedded in a gelatinous matrix. *Asci* 90–140  $\times$  14.5–18  $\mu\text{m}$  ( $\bar{x}$  = 118.1  $\times$  16.8  $\mu\text{m}$ ,  $n=20$ ), 8-spored, bitunicate, fissitunicate, cylindrical, short pedicellate, apically rounded, with an ocular chamber. *Ascospores* 38.5–55.5  $\times$  4.5–6  $\mu\text{m}$  ( $\bar{x}$  = 47.3  $\times$  5.6  $\mu\text{m}$ ,  $n=20$ ), long subfusiform, elongate, pointed at the ends, slightly curved, 10–13-septate, not constricted at the septa, hyaline, smooth-walled. **Asexual morph:** Undetermined.

**Cultural characters:** Ascospores germinating on PDA within 24 h. Colonies growing slowly on PDA, reaching 2 mm in 30 d at 28  $^{\circ}\text{C}$ , under 12 h light/12 h dark, effuse, velvety to hairy, edge fimbriate, olive to olive brown, dark



**Fig. 25** *Tubeufia longiseta* (MFLU 15–1215, holotype). **a** Herbarium specimen. **b–d** Black ascomata with setae on host surface. **e, f** Vertical sections of ascomata. **g** Peridium. **h** Dark brown setae. **i–k** Asci. **l, m**

Sterile cells from PDA. **n, o** Cultures on PDA. **p–t** Ascospores. **u** Germinating ascospore. Scale bars: **a** = 5 mm, **b–d** = 1 mm, **e–g** = 50  $\mu\text{m}$ , **h–u** = 10  $\mu\text{m}$

brown on PDA media. Mycelium superficial and partially immersed, branched, septate, pale brown to olivaceous brown, smooth, sterile at 60 days.

**Material examined:** THAILAND, Phang-Nga, Doi Nang Hong, Tham Thong Lang, Thap Put District, 8°32'11"N 98°33'35"E, on dead culms of bamboo, 6 October 2014, Kevin D. Hyde DDQ00287 (MFLU 15–1215, **holotype**); *Ibid.* (KUN HKAS88722, **isotype**), ex-type living cultures, MFLUCC 15–0188, KUMCC.

**Notes:** *Tubeufia longiseta* is similar to *T. acacia* in having superficial, subglobose, black ascomata, covered with black setae, cylindrical asci and long subfusiform, elongate ascospores. However, *T. longiseta* has smaller ascomata (250–370 µm high, 180–250 µm diam. vs. 300–375 µm high, 225–255 µm diam.), ascospores with more transverse septa (10–13 vs. 5–7 septa), and is found on different substrates (bamboo vs. acacia). Black ascomata with long, black setae can be observed in several genera of *Tubeufiaceae*, such as *Acanthohelicospora*, *Acanthostigma*, *Chlamydotubeufia*, *Helicangiospora*, *Neocanthostigma* and *Thaxteriellopsis* (Boonmee et al. 2014). However, phylogenetic analyses confirm that this new taxon belongs to *Tubeufia sensu stricto* and is closely related to *T. javanica* (Fig. 26). The latter species differs in having white to yellowish, oblong to clavate ascomata and broad, filiform ascospores (Boonmee et al. 2014).

**Valsariales** Jaklitsch, K.D. Hyde & Voglmayr, Fungal Diversity 73:167 (2015)

The order *Valsariales* was introduced by Jaklitsch et al. (2015) to accommodate the single family *Valsariaceae*. Jaklitsch et al. (2015) revised *Valsaria* and introduced *Valsariaceae* based on multi-gene phylogeny analyses (SSU, LSU, ITS, RPB2 and tef1 sequences). Jaklitsch et al. (2015) re-examined and re-collected type species and provided full descriptions and illustrations. Two new combined genera were introduced by Jaklitsch et al. (2015). Thus, *Valsariaceae* includes three genera, *Bambusaria*, *Myrmaecium* and *Valsaria*.

***Bambusaria bambusae*** (J.N. Kapoor & H.S. Gill) Jaklitsch, D.Q. Dai, K.D. Hyde & Voglmayr, Fungal Diversity 73: 196 (2015), Figs. 27 and 28

**Material examined:** THAILAND, Chiang Rai, Mae Fah Luang University, on dead culms of bamboo, 1 August 2011, Dong-Qin Dai, DDQ0097 (MFLU 15–1192); Doi Pui, 1 September 2011, Dong-Qin Dai, DDQ00109 (MFLU 15–1194).

**Notes:** The monotypic genus *Bambusaria* was introduced by Jaklitsch et al. (2015) and is typified by *B. bambusae* (J.N. Kapoor & H.S. Gill) Jaklitsch, D.Q. Dai, K.D. Hyde (basonym: *Valsaria bambusae* J.N. Kapoor & H.S. Gill). *Bambusaria bambusae* was originally isolated from bamboo in India by Kapoor and Gill (1961). The same species was recollected on *Thyrsostachys siamensis* (*Bambuseae*) and published in Jaklitsch et al. (2015). In the present study, this

species is re-illustrated based on new collections (sexual and asexual morph specimens).

***Sordariomycetes*** O.E. Erikss. & Winka, Myconet 1(1): 10 (1997)

*Sordariomycetes* is one of the largest classes of Ascomycota consisting of 15 orders, 64 families, 1119 genera, and 10564 species (Kirk et al. 2008). Maharachchikumbura et al. (2015) provided an updated outline of *Sordariomycetes* and a backbone tree incorporating asexual and sexual genera in the class. Six subclasses, 28 orders, 90 families and 1344 genera were included by Maharachchikumbura et al. (2015).

***Chaetosphaeriales*** Huhndorf, A.N. Mill. & F.A. Fernández, Mycologia 96(2): 378 (2004)

The order *Chaetosphaeriales* was introduced by Huhndorf et al. (2004). Lumbsch and Huhndorf (2010) included a single family *Chaetosphaeriaceae* with ten genera in this order. Based on combined LSU, SSU, TEF and RPB2 genes analyses, Maharachchikumbura et al. (2015) moved the family *Helminthosphaeriaceae* with seven genera into *Chaetosphaeriales* and recognized 35 genera within *Chaetosphaeriaceae*, and placed six genera, as *Chaetosphaeriales*, genera *incertae sedis*.

***Chaetosphaeriales*, genera *incertae sedis***

***Leptosporella*** Penz. & Sacc., Malpighia 11(9–10): 406 (1897)

*Leptosporella* was introduced by Penzig and Saccardo (1897), and typified by *L. gregaria* Penz. & Sacc. Lumbsch and Huhndorf (2010) placed *Leptosporella* in Sordariomycetidae, genera *incertae sedis*. Huhndorf and Miller (2011) re-examined the holotype of *L. gregaria* and collected fresh specimens. Based on LSU sequence data, Huhndorf and Miller (2011) moved *Leptosporella* into *Chaetosphaeriales*. However, the family placement of *Leptosporella* is still not yet determined (Maharachchikumbura et al. 2015).

***Leptosporella bambusae*** D.Q. Dai & K.D. Hyde, *sp. nov.*

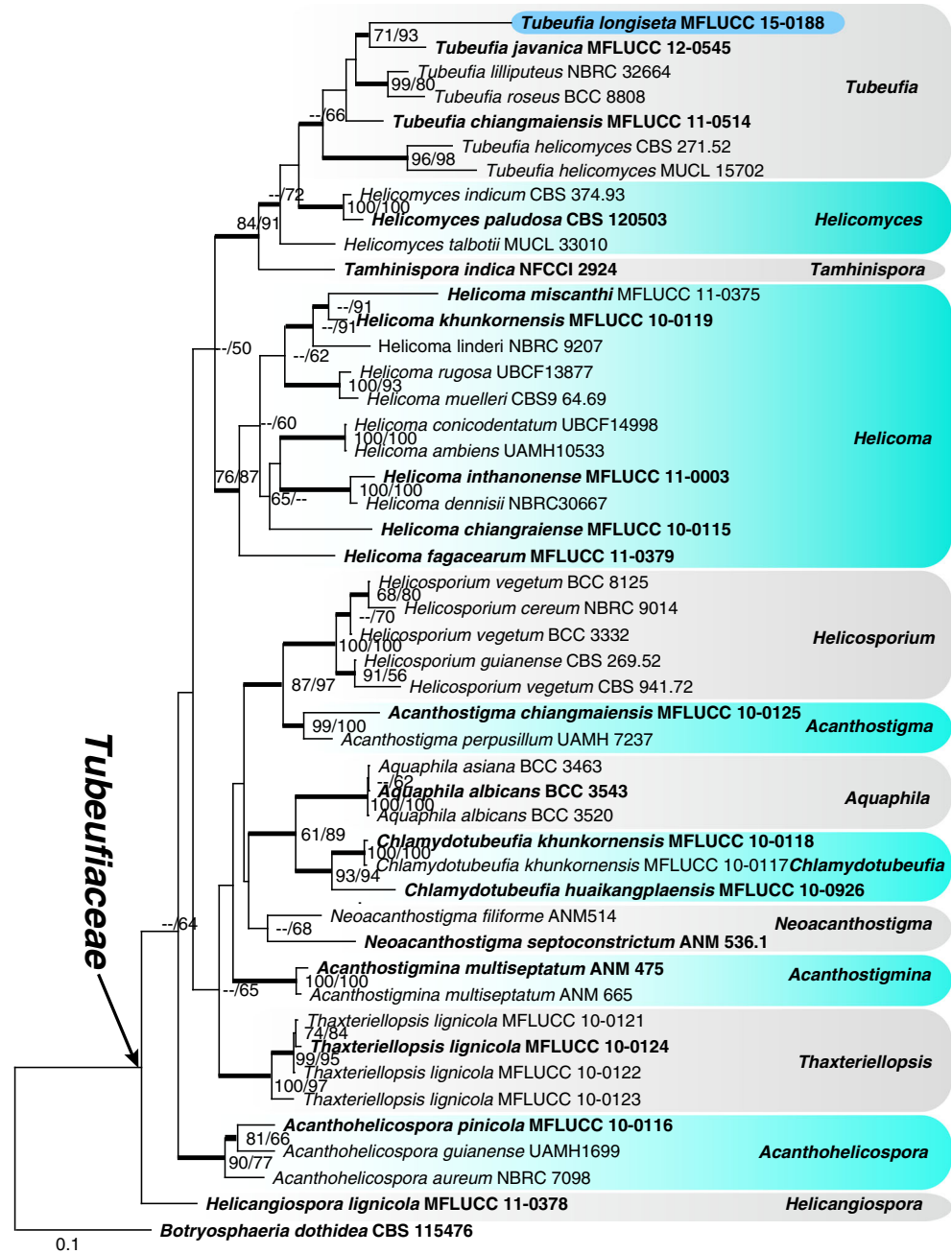
***Index Fungorum numbers:*** IF552030; ***Facesoffungi numbers:*** FoF 01986; Figs. 29 and 30

***Etymology:*** Refers to the bamboo host.

***Holotype:*** MFLU 15–1203

***Saprobic*** on dead bamboo culms. **Sexual morph:** *Ascomata* forming under a black pseudostromata, immersed within cortex, becoming raised and conical when mature, 300–450 µm high, 700–1000 µm long and 500–850 µm wide, coriaceous, with black and round papilla; in vertical section 200–250 µm high, 500–850 µm diam., solitary, subglobose, dark brown, with flattened base, with centrally located ostiole, periphyses not observed. *Peridium* 15–25 µm wide, composed of dark brown to hyaline cells of *textura angularis*, with poorly developed lower and upper walls. *Hamathecium* of dense, 2–3.5 µm wide, long, filamentous, unbranched, indistinctly septate, paraphyses. *Asci*

**Fig. 26** Maximum likelihood phylogenetic tree (lnL = -1912.497367) generated by RAxML (GTR+G model) based on combined LSU and ITS sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes, and branches with Bayesian posterior probabilities greater than 0.90 are given in bold. The original isolate numbers are noted after the species names. The tree is rooted to *Botryosphaeria dothidea* (CBS 115476). Ex-type strains are in bold and newly generated sequences are highlighted with a blue background



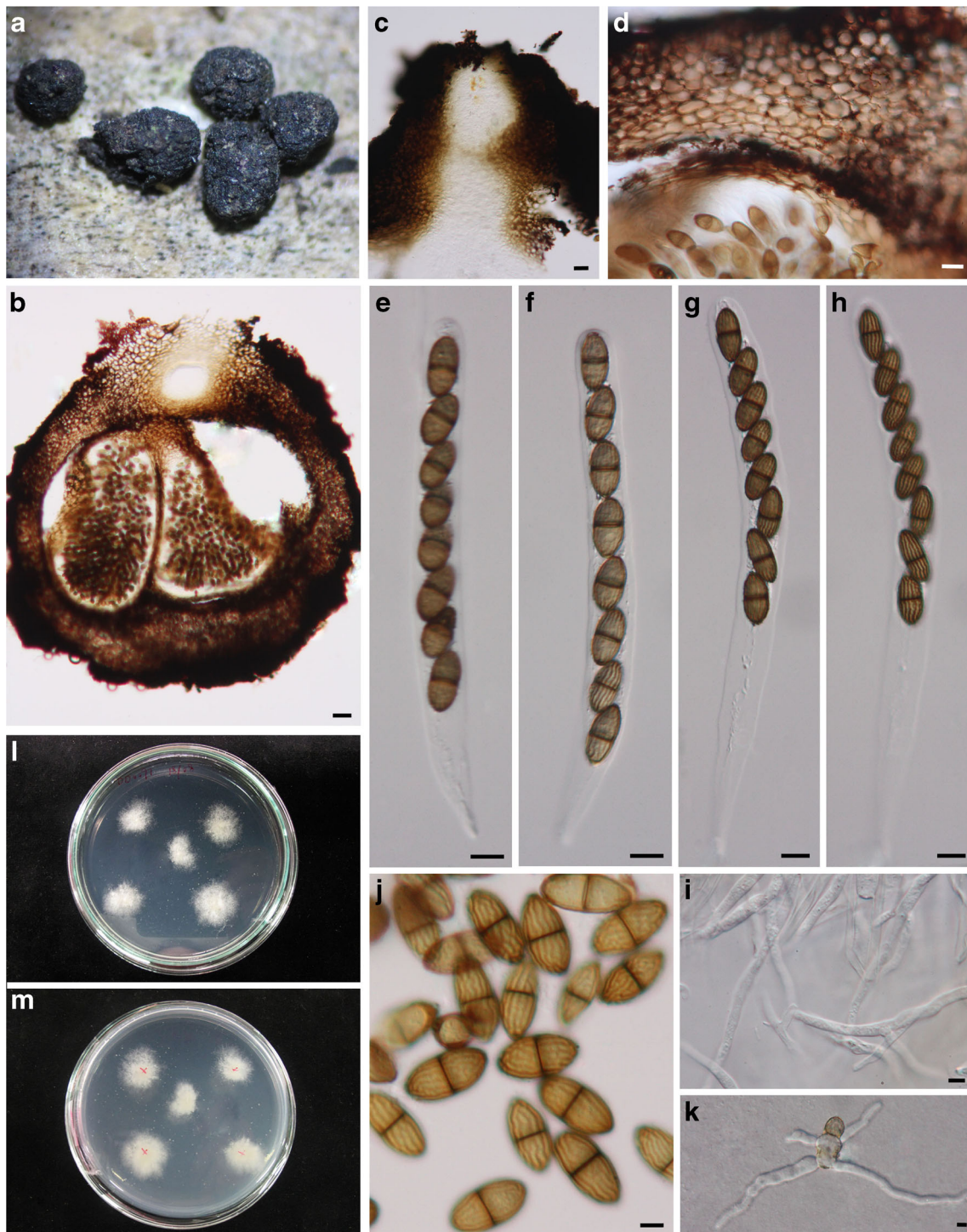
100–195.5 × 9–13.5 μm ( $\bar{x}$  = 168.6 × 10.8 μm,  $n$  = 20), 2–8-spored, unitunicate, cylindrical, with a short furcate pedicel, with a J-, apical ring. *Ascospores* 130–175 × 2–3 μm ( $\bar{x}$  = 160.3 × 2.4 μm,  $n$  = 20), fasciculate, hyaline, long fusiform, 2–6-septate, narrow and acute at both ends, curved, with abundant small and round guttules, smooth-walled. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 48 h and germ tubes produced from both ends. **Colonies** growing slowly on PDA, reaching 20 mm in 45 d at 25–32 °C, under 12 h light/12 h dark, circular, with even margin, floccose, white and high convex in

centre, brown at margin. Mycelium immersed and superficial in the media, composed of branched, septate, smooth-walled, brown hyphae.

**Material examined:** THAILAND, Chiang Rai, Khun Korn Waterfall, on dead culms of bamboo, 31 July 2012, Dong-Qin Dai DDQ00243 (MFLU 15–1203, **holotype**); *Ibid.* (KUN HKAS88712, **isotype**), ex-type living cultures, MFLUCC 12–0846, MUCL 55879.

**Notes:** Currently 13 records of *Leptospora* are listed in Index Fungorum (2016) of which 9 are considered to be species of *Leptospora*. Members of *Leptospora* usually occur on wood or branches (Rehm 1901; von



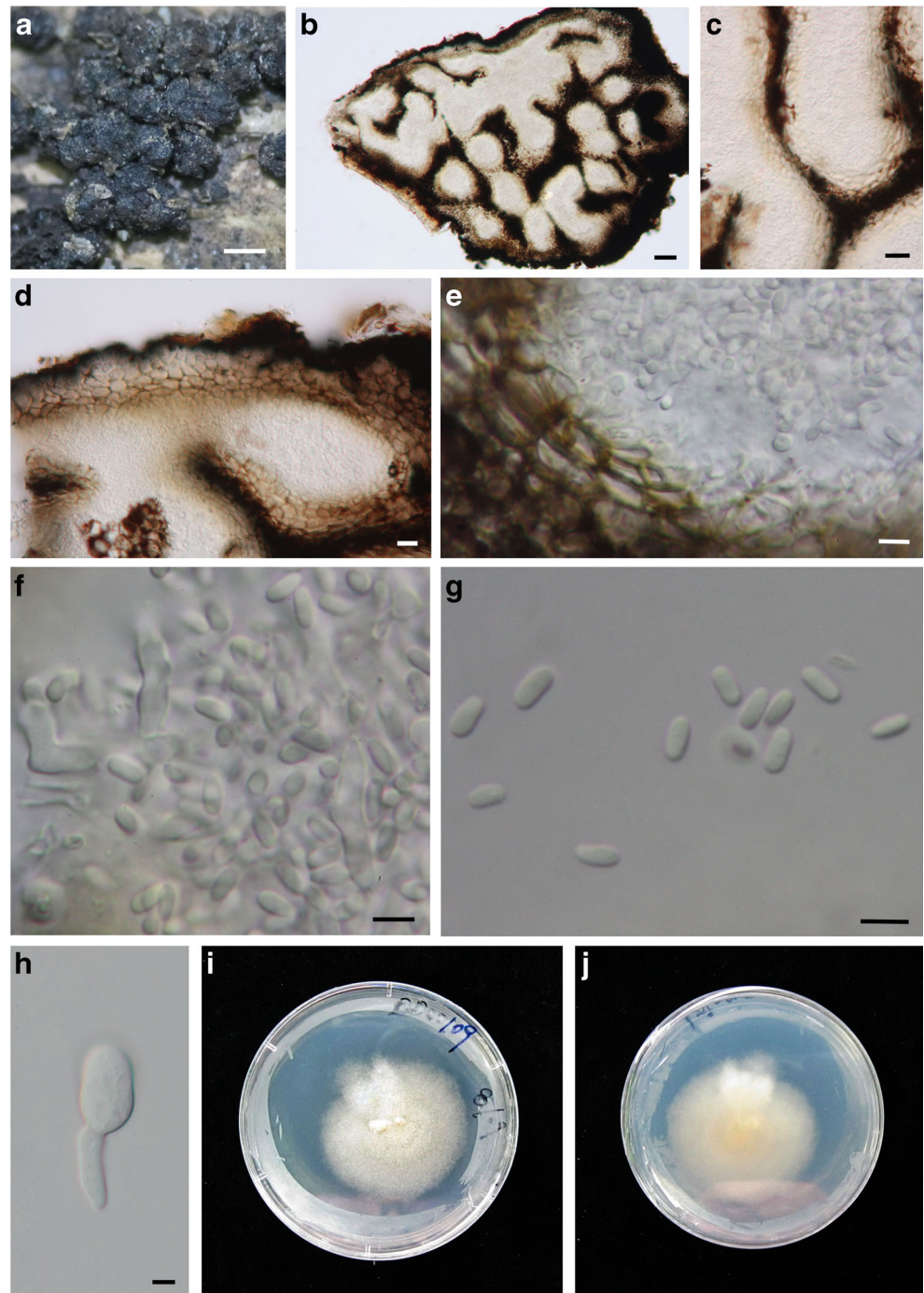
**Fig. 27** *Bambusaria bambusae* (MFLU 15–1192). **a** ascostromata on bamboo. **b** Vertical section of ascostroma. **c** Ostiole of locule. **d** Peridium of ascostroma and locule. **e–h** Asci. **i** Pseudoparaphyses. **j** Ascospores

with striations. **k** Germinating ascospore. **l, m** Culture on PDA. Scale bars: **b, c** = 50  $\mu\text{m}$ , **d–h** = 10  $\mu\text{m}$ , **i–k** = 5  $\mu\text{m}$

Höhnelt 1909; Spegazzini 1912; Sydow 1938; Chardón 1939; Hansford 1957; Huhndorf and Miller 2011) and occasionally on ferns and *Rosa* sp. (Edward et al. 1972; Index Fungorum 2016). However, our new taxon, *Leptospora bambusae*, was collected from bamboo.

*Leptospora bambusae* can be compared with the generic type, *L. gregaria*, but it has narrower ascospores (150–175  $\times$  2–3  $\mu\text{m}$  vs. 107–137  $\times$  2.8–4  $\mu\text{m}$ ) and possesses septa (2–6-septate vs. 0-septate) (Huhndorf and Miller 2011). In the phylogenetic analyses (Fig. 29), the

**Fig. 28** *Bambusaria bambusae* (MFLU 15–1194). **a** Conidiomata on bamboo. **b** Vertical section of conidioma. **c** Conidiolocules. **d** Wall of conidiolocules. **e, f** Conidiogenous cells and conidia. **g** Conidia **h** Germinating conidium. **i, j** Cultures on PDA. Scale bars: a = 1 mm, b = 50  $\mu$ m, c–e = 10  $\mu$ m, f–h = 5  $\mu$ m



new taxon clusters in *Leptospora* with strong support (100/99 % MPBP/MLBP).

**Hypocreales** Lindau, in Engler & Prantl, Nat. Pflanzenfam., Teil. I (Leipzig) 1(1): 343 (1897)

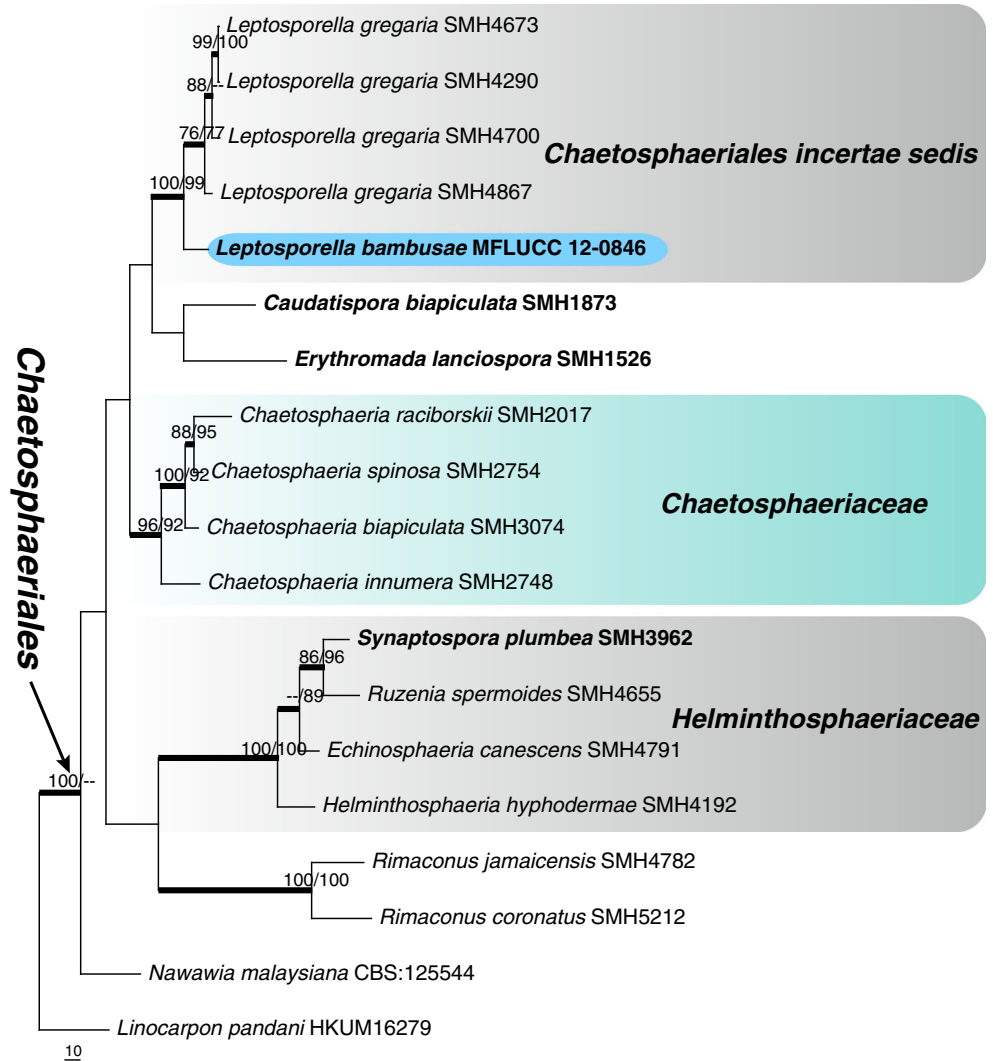
*Hypocreales* are highly diverse in the tropics and subtropics (Pöldmaa 2011; Maharachchikumbura et al. 2015). Kirk et al. (2008) indicated that *Hypocreales* comprised seven families, 237 genera and 2647 species. Maharachchikumbura et al. (2015) included *Bionectriaceae*, *Clavicipitaceae*, *Cordycipitaceae*, *Hypocreaceae*, *Nectriaceae*, *Niessliaceae*,

*Ophiocordycipitaceae*, *Stachybotryaceae* and *Tilachliaceae* in the order, based on phylogenetic analyses.

**Stachybotryaceae** L. Lombard & Crous [as '*Stachybotriaceae*'], in Crous et al., Persoonia, Mol. Phyl. Evol. Fungi 32: 283 (2014)

*Stachybotryaceae*, typified by *Stachybotrys* Corda, was introduced by Crous et al. (2014a). This family was established to accommodate the genera *Myrothecium*, *Peethambara* and *Stachybotrys* (Crous et al. 2014a), three genera earlier placed in *incertae sedis* of the order *Hypocreales* (Lumbsch and

**Fig. 29** Maximum likelihood phylogenetic tree (lnL = -3059.137345) generated by RAxML (GTR+G model) based on LSU sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes, and branches with Bayesian posterior probabilities greater than 0.90 are given in bold. The original isolate numbers are noted after the species names. The tree is rooted to *Linocarpon pandani* (HKUM16279). Ex-type strains are in **bold** and a newly generated sequence is highlighted with a blue background



Huhndorf 2010; Crous et al. 2014a). A further four genera, *Albosynnema*, *Parasarcopodium*, *Sarcopodium* and *Scopinella* were transferred to *Stachybotryaceae* by Maharachchikumbura et al. (2015).

***Myrothecium*** Tode, Fung. mecklenb. sel. (Lüneburg) 1: 25 (1790)

*Myrothecium* was introduced by Tode (1790). The genus was placed in *Hypocreales* order *incertae sedis* by Castlebury et al. (2004), Lumbsch and Huhndorf (2010), Summerbell et al. (2011) and Wijayawardene et al. (2012). Later, it was transferred to *Stachybotryaceae* by Crous et al. (2014). Maharachchikumbura et al. (2015) verified this placement by phylogenetic analyses. *Myrothecium* species are usually found in decaying habitats, soil and rotting leaves (Pidoplichko and Kirilenko 1969; Tulloch 1972; Saccardo 1886; Udagawa and Awao 1984; Matsushima 1989; Jiang et al. 2014; Wu et al. 2015) occasionally on dung (Faurel and Schotter 1965) and algae (DiCosmo et al. 1980). Liu et al. (2015) introduced a *Myrothecium* species occurring on

bamboo. Chen et al. (2016) revised *Myrothecium* in detail, with phylogenetic analyses of ITS, ATP6, EF1- $\alpha$ , LSU, RPB1 and SSU sequence data. In this paper three new species were isolated from bamboo and are described and illustrated.

***Myrothecium thailandicum*** D.Q. Dai & K.D. Hyde, *sp. nov.*

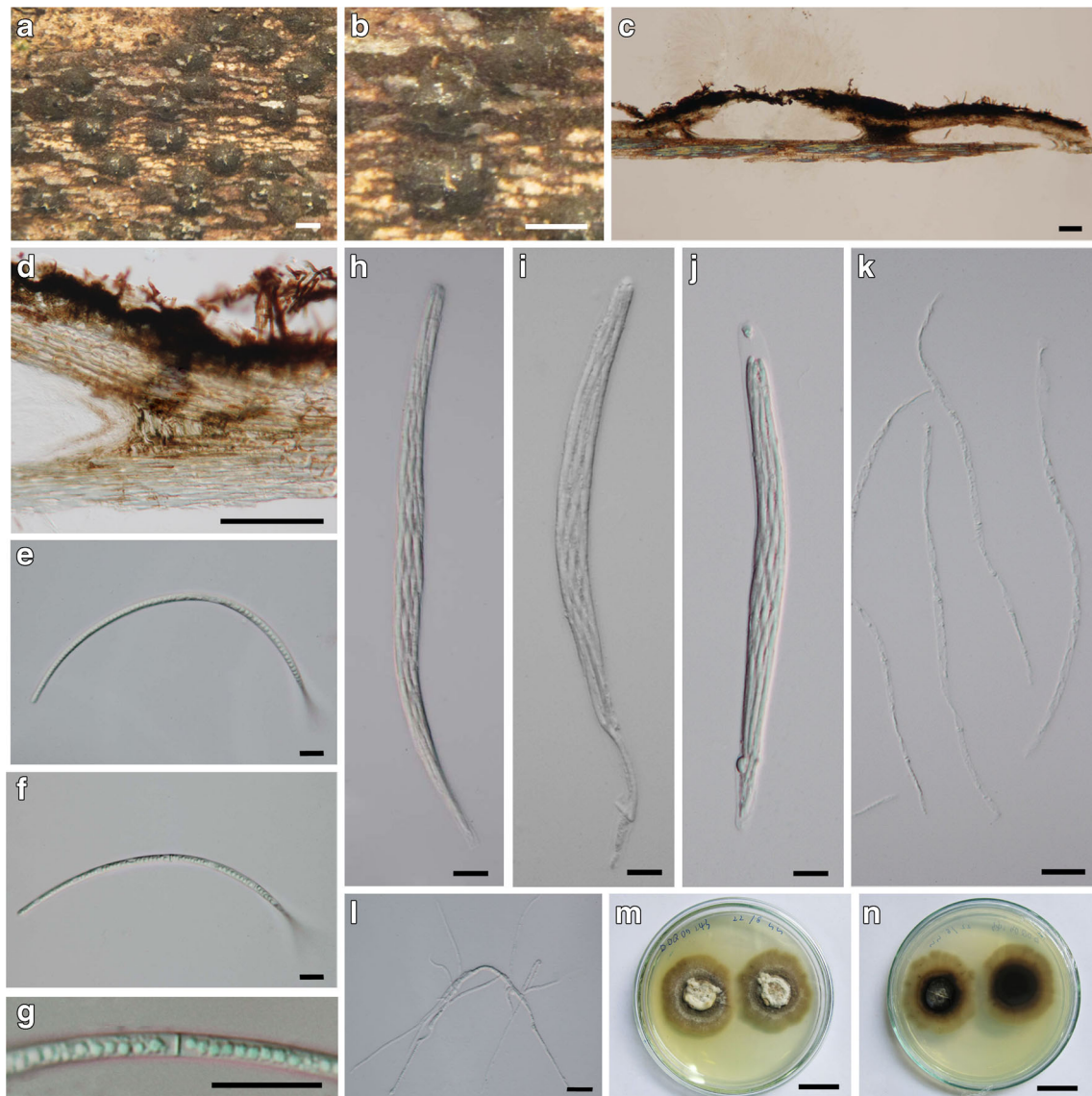
*Index Fungorum* number: IF552031; *Facesoffungi* numbers: IF 01987; Figs. 31 and 32

*Entomology*: Refers to the country where this species first collected.

*Holotype*: MFLU 15-1180

*Saprobic* on decaying bamboo culms. **Sexual morph**: Undetermined. **Asexual morph**: *Mycelium* partly immersed on the substrate, partly superficial, composed of septate, branched, hyaline, smooth hyphae. *Synnemata* 600–1000  $\mu\text{m}$  long, apical part 45–60  $\mu\text{m}$  wide, basal part 20–30  $\mu\text{m}$  wide, superficial, gregarious, white, apically crowned by slimy, glistening, dark green conidial head. *Conidiophores* macronematous, cylindrical, pale brown,





**Fig. 30** *Leptosorella bambusae* (MFLU 15–1203, **holotype**). **a, b** Appearance of ascostromata on bamboo host. **c** Vertical section of ascostromata. **d** Peridium at the side. **e–g** Hyaline ascospores (**g**:

showing septum of ascospore). **h–j** Asci (**j**: showing ascap apical ring). **k** Paraphyses. **l** Geminating spore. **m, n** Cultures on PDA. Scale bars: **a, b** = 1 mm, **c, d** = 100  $\mu\text{m}$ , **e–l** = 10  $\mu\text{m}$ , **m, n** = 25 mm

septate, unbranched, straight, smooth, single conidiophores  $400\text{--}650 \times 4\text{--}5 \mu\text{m}$ . *Conidiogenous cells*  $15\text{--}20 \times 1.5\text{--}2 \mu\text{m}$  ( $\bar{x} = 17.3 \times 1.7 \mu\text{m}$ ,  $n=20$ ), phialidic, integrated or discrete, cylindrical, pale brown to hyaline, smooth-walled, straight. *Conidia*  $6.5\text{--}10 \times 2.5\text{--}4.5 \mu\text{m}$  ( $\bar{x} = 8.5 \times 3.4 \mu\text{m}$ ,  $n=20$ ), cylindrical, narrow at the apex, truncate at the base, straight, aseptate, hyaline, smooth-walled, with guttules.

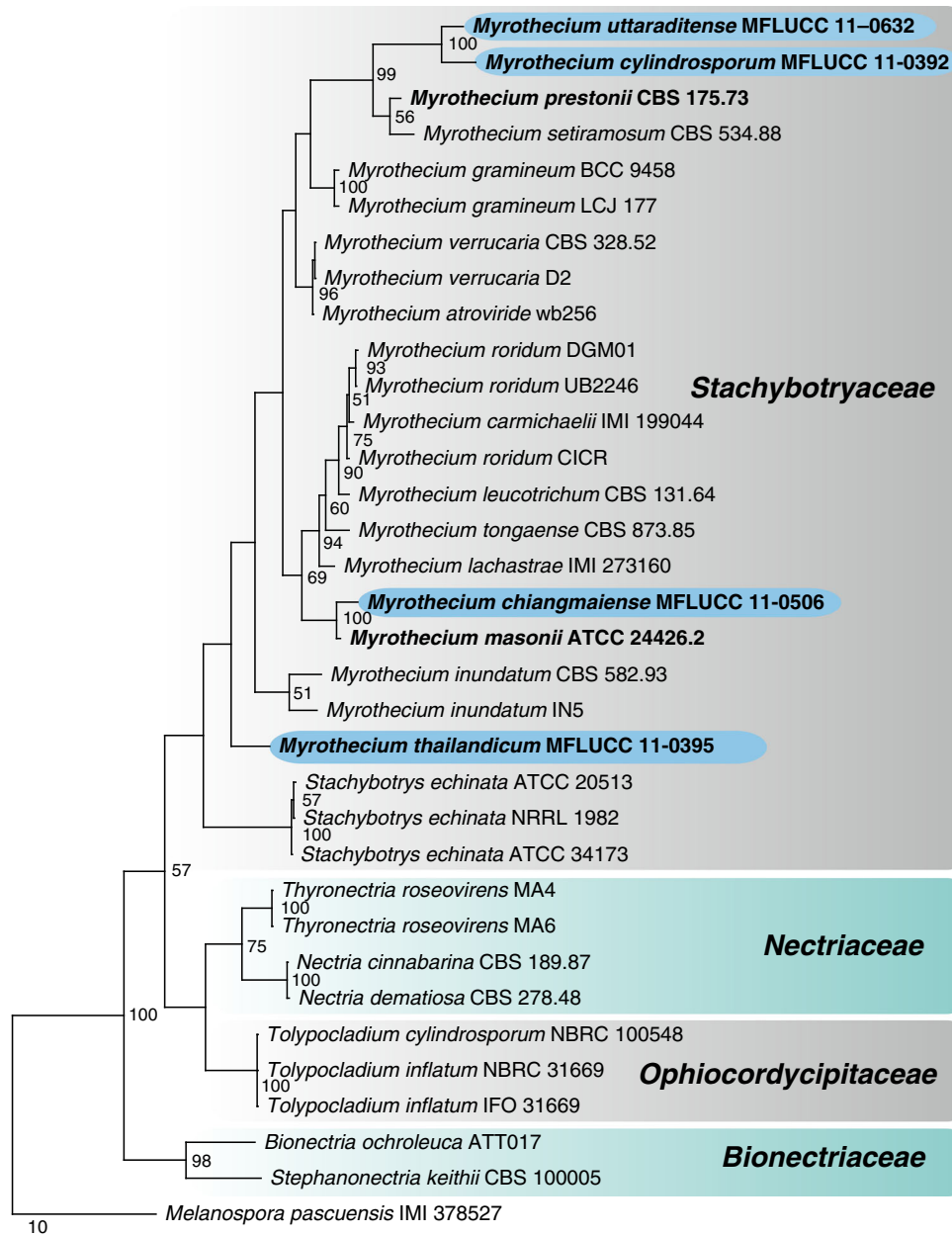
**Culture characters:** Conidia germinating on PDA within 24 h and germ tubes developing from both apices. Colonies growing slowly on PDA, reaching 4 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, circular, with regular edge, hyaline from above and below. Mycelium superficial to immersed in media, with branched, septate, smooth hyphae.

**Material examined:** THAILAND, Chiang Mai, Mae Sae Village, on dead culms of bamboo, 14 May 2011, Dong-Qin Dai, DDQ00029 (MFLU 15–1180, **holotype**); *Ibid.* (KUN HKAS88691, **isotype**), living cultures, MFLUCC 11–0395, KUMCC.

**Notes:** *Myrothecium thailandicum* can be phylogenetically distinguished from other species (Fig. 31). This species differs from *M. chiangmaiense* in having shorter conidiophores forming together, wider conidia ( $2.5\text{--}4.5 \mu\text{m}$  vs.  $1.5\text{--}2.5 \mu\text{m}$ ), and different culture (culture with a circle in middle and with regular edge vs. one without circle and with irregular edge).

*Myrothecium chiangmaiense* D. Q. Dai & K.D. Hyde, *sp. nov.*

**Fig. 31** One of six equally most parsimonious trees obtained from the ITS sequence data. MP values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes. The original isolate codes are noted after the species names. The tree is rooted to *Melanospora pascuensis* (IMI 378527). Ex-type strains are in **bold** and newly generated sequences are highlighted with a **blue** background



*Index Fungorum number*: IF552032, *Facesoffungi number*: FoF 01988; Fig. 33

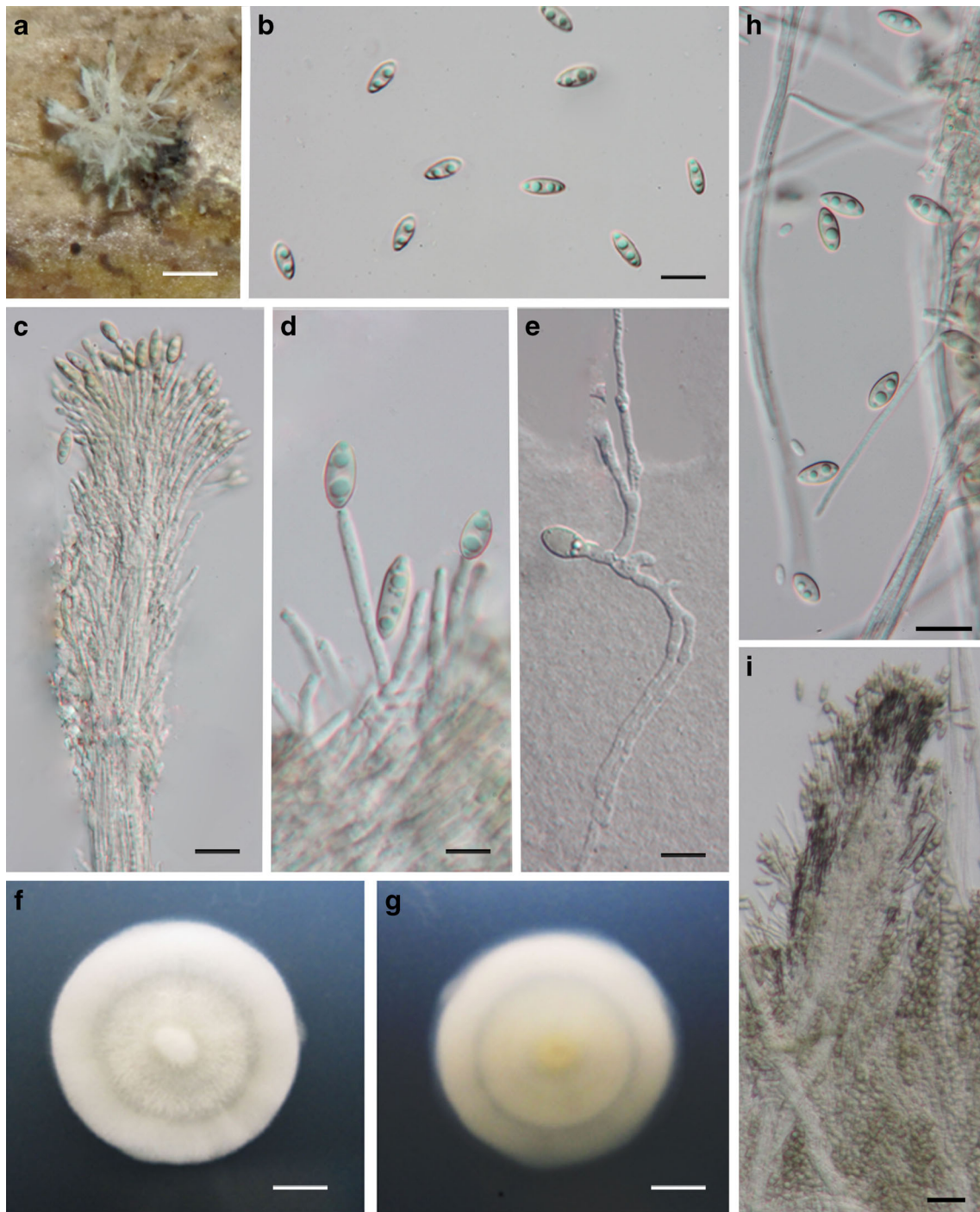
*Entomology*: Refers to the province where this species first collected.

*Holotype*: MFLU 15-1181

*Saprobic* on decaying bamboo culms. *Mycelium* immersed in substrate, composed of septate, branched, hyphae. **Sexual morph**: Undertermined. **Asexual morph**: *Synnemata* 1000–2000 × 500–700 μm ( $\bar{x}$  = 1450 × 640 μm,  $n=20$ ), superficial, solitary, creamish white, with conidiophores arranged in a palisade layer, sometimes proliferating 2–3 times vertically, apically crowned by a slimy, glistening, greenish brown conidial head. *Conidiophores* macronematous, synnematus, white, branched, septate,

straight, with white and flocculent hyphae at the base. *Conidiogenous cells* 10–30 × 4–5 μm ( $\bar{x}$  = 25.3 × 4.7 μm,  $n=20$ ), phialidic, cylindrical, hyaline, smooth, straight. *Conidia* 4–7 × 1.5–2.5 μm ( $\bar{x}$  = 5.6 × 1.9 μm,  $n=20$ ) μm, cylindrical, long ellipsoid, narrow at basal end, straight to slightly curved, aseptate, hyaline, smooth-walled, guttulate.

*Culture characters*: Ascospores germinating on PDA within 24 h and germ tubes produced from both ends. Colonies growing fast on PDA, reaching 30 cm in 2 weeks at 28 °C, under 12 h light/12 h dark, circular, irregular edge, flocculent, snow white from above and below. *Mycelium* superficial to immersed in media, branched, septate, smooth.



**Fig. 32** *Myrothecium thailandicum* (MFLU 15–1180, holotype). **a** Synnemata gregariously on bamboo. **b, h** Conidia. **c, i** Conidiophores. **d** Conidiogenous cells. **e** Germinating conidium. **f, g** Cultures on PDA. Scale bars: **a** = 10 mm, **b–e** = 10  $\mu$ m, **f, g** = 30 mm

*Material examined*: THAILAND, Chiang Mai, Mae Sae Village, on dead culms of bamboo, 14 May 2011, Dong-Qin Dai DDQ00031 (MFLU 15–1181, holotype); *Ibid.* (KUN HKAS88692, isotype), living cultures, MFLUCC 11–0506, CBS 139986.

*Notes*: *Myrothecium chiangmaiense* is close to *M. masonii* M.C. Tulloch based on the phylogenetic analysis (Fig. 31). However, *M. masonii* was isolated from leaves of *Glyceria*

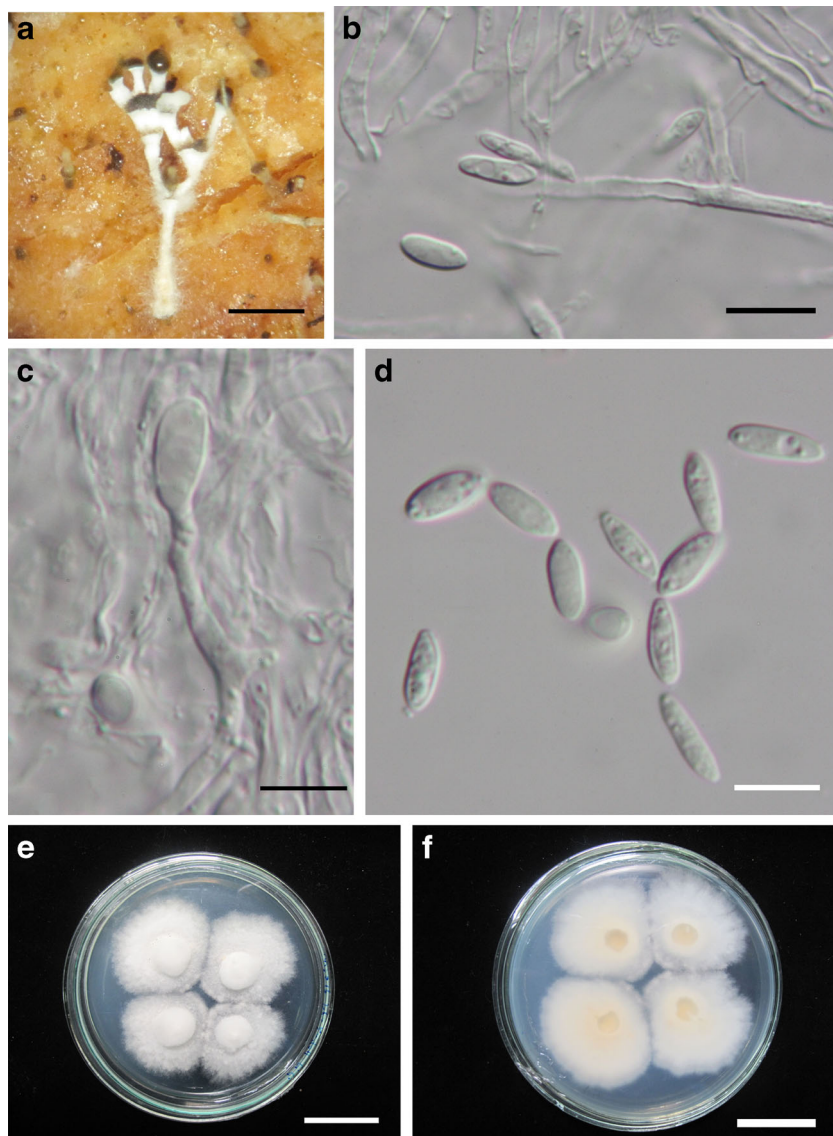
R.Br. in UK (Tulloch 1972), but our species is from culms of bamboo, in Thailand.

*Myrothecium cylindrosporum* D.Q. Dai & K.D.Hyde, *nom. nov.*

*Index Fungorum Number*: IF552044

*Synonym*: *Myrothecium macrosporum* D.Q. Dai & K.D.Hyde, in Liu et al., Fungal Diversity: 10.1007/s13225-015-0324-y, [33] (2015)

**Fig. 33** *Myrothecium chiangmaiense* (MFLU 15–1181, **holotype**) **a** Conidiophores on surface of host. **b, d** Hyaline conidia. **c** Conidiogenous cell. **e, f** Cultures on PDA. Scale bars: **a** = 500  $\mu\text{m}$ , **b–d** = 5  $\mu\text{m}$ , **e, f** = 25 mm



**Notes:** Jiang et al. (2014) described *Myrothecium macrosporum* H.F. Wang & T.Y. Zhang from forest soil. This taxon has pale greenish brown, to dark greenish brown, obovoid to pyriform conidia. Due to Index Fungorum (2016) releasing *Myrothecium macrosporum* H.F. Wang & T.Y. Zhang late, Liu et al. (2015), also introduced *Myrothecium macrosporum* D.Q. Dai & K.D. Hyde to accommodate another *Myrothecium* species which has large, hyaline, cylindrical conidia. As, the name *Myrothecium macrosporum* D.Q. Dai & K.D. Hyde is invalid, we herein provide a new name *Myrothecium cylindrosporum* **nom. nov.** to accommodate this taxon.

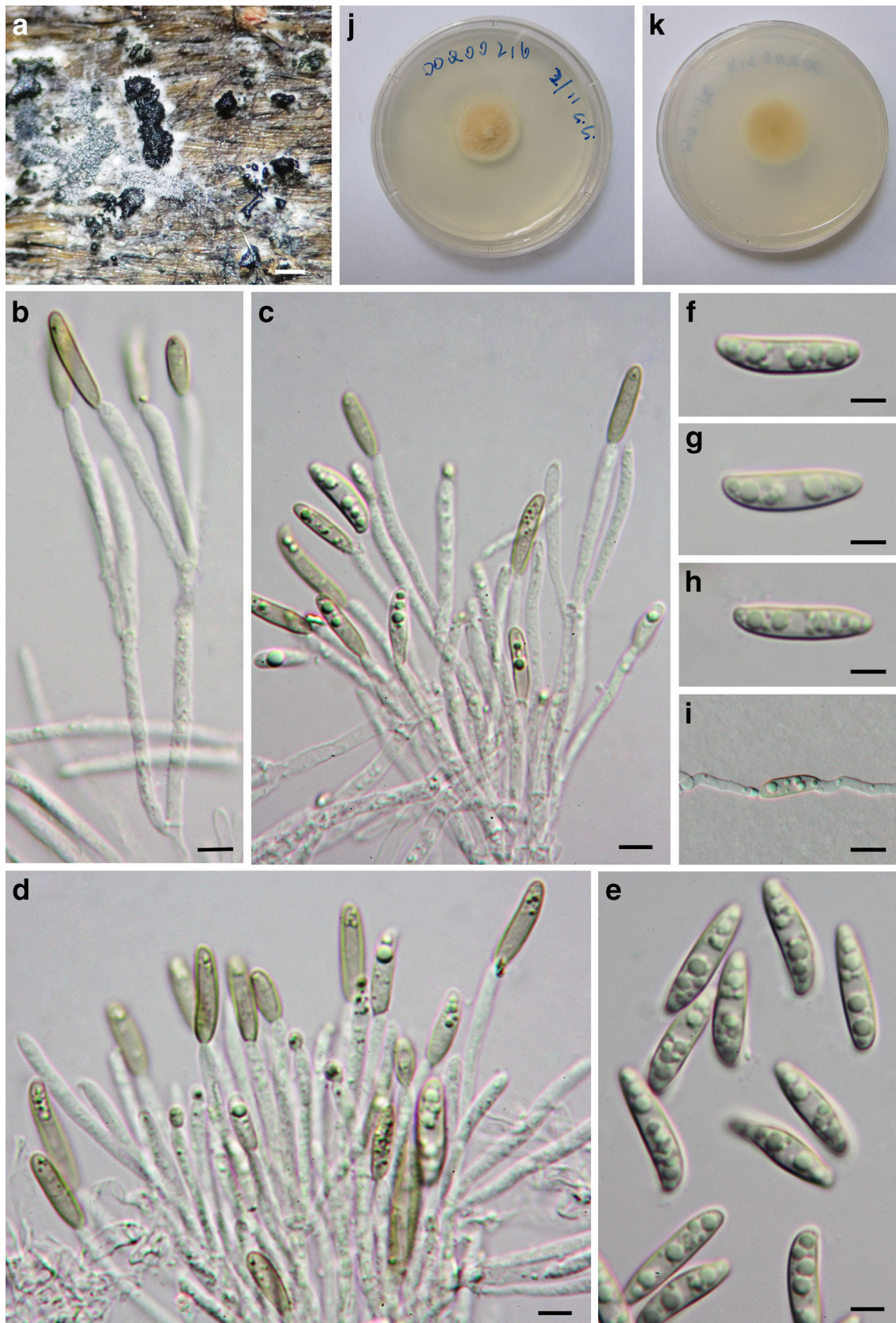
*Myrothecium uttaraditense* D.Q. Dai & K.D. Hyde, **sp. nov.**

**Index Fungorum number:** IF552033; **Facesoffungi number:** FoF 01989; Fig. 34

**Entomology:** Refers to the province where this species first collected.

**Holotype:** MFLU 15–1197

**Saprobic** on decaying bamboo culms. **Sexual morph:** Undetermined. **Asexual morph:** *Mycelium* immersed in the substrate, composed of septate, branched hyphae. *Sporodochia* comprising white mycelium, round to irregular, at first dark green, becoming black with a white margin, without setae. *Conidiophores* macronematous, cylindrical, brown to dark brown, septate, branching irregularly penicillate, straight, smooth, single conidiophores  $60\text{--}100 \times 2\text{--}3 \mu\text{m}$  ( $\bar{x} = 75.3 \times 2.6 \mu\text{m}$ ,  $n=20$ ). *Conidiogenous cells*  $12\text{--}25 \times 1.5\text{--}2.5 \mu\text{m}$  ( $\bar{x} = 20.6 \times 2.1 \mu\text{m}$ ,  $n=20$ ), phialidic, integrated or discrete, cylindrical, pale brown, smooth, straight. *Conidia*  $10\text{--}15 \times 2\text{--}3.5 \mu\text{m}$  ( $\bar{x} = 12.6 \times 3.1 \mu\text{m}$ ,  $n=20$ )  $\mu\text{m}$ ,



**Fig. 34** *Myrothecium uttaraditense* (MFLU 15–1197, holotype). **a** Sporodochium on bamboo. **b–d** Conidiophores and conidia. **f–h** Conidia. **i** Geminating conidia. **j, k** Cultures on PDA. Scale bars: **a** = 500  $\mu\text{m}$ , **b–i** = 5  $\mu\text{m}$

cylindrical to long ellipsoid, narrow and truncate at basal end, aseptate, pale brown, straight to slightly curve, smooth-walled, guttulate.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from both apices. Colonies growing slowly on PDA, reaching 5 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, circular, orange from above and below. Mycelium superficial to immersed in media, branched, septate, smooth.

**Material examined:** THAILAND, Uttaradit Province, on dead culms of bamboo, 28 October 2011, Dong-Qin Dai DDQ00216 (MFLU 15–1197, **holotype**); *Ibid.* (KUN HKAS88748, **isotype**), living cultures, MFLUCC 11–0632, KUMCC.

**Notes:** *Myrothecium uttaraditense* is morphologically similar to *M. inundatum* Tode and *M. roridum* Tode in its dark sporodochia surrounded by white mycelium. However, *M. uttaraditense* phylogenetically separates from the above species (Fig. 31). In addition, *M. inundatum*, was isolated from *Agaricus*, and *M. roridum* from leaves, stems or seeds of Angiosperms and Gymnosperms (Saccardo 1886; Worapong et al. 2009), whereas, our new species was isolated from bamboo culms. In the phylogenetic tree (Fig. 31), *M. uttaraditense* is closest to *M. cylindrosporum*, but differs in having pigmented and shorter conidia (pale brown and 10–15 µm long vs. hyaline and 25–35 µm long).

**Nectriaceae** Tul. & C. Tul. [as ‘*Nectriei*’], *Select. fung. carpol.* (Paris) 3: 3 (1865)

**Nectriaceae**, typified by *Nectria* (Fr.) Fr., was introduced by Tulasne and Tulasne (1865). This family is characterized by light to bright-coloured (usually white, yellow, orange-red or purple), soft-textured, superficial, uniloculate perithecia, unitunicate asci, and phialidic asexual morphs (Hirooka et al. 2012; Lombard et al. 2015). Chaverri et al. (2011) placed seven genera in the family. Lumbsch and Huhndorf (2010) included 26 genera and of these, Hirooka et al. (2012) accepted 16 genera. Lombard et al. (2015) mentioned that **Nectriaceae** contains around 55 genera based on sexual and asexual morphs and resolved the identity of 47 genera based on DNA sequences of 10 loci (*act1*, *act*, *cmdA*, *his3*, *ITS*, *LSU*, *rpb1*, *rpb2*, *tefl* and *tub2*). Maharachchikumbura et al. (2015) listed 67 genera within **Nectriaceae**. This family presently includes approximately 900 species (Lombard et al. 2015). Notable asexual morph genera of **Nectriaceae** are *Acremonium*, *Clonostachys*, *Cylindrocarpon*, *Cylindrocladium*, *Fusarium*, *Stilbella* and *Tubercularia* (Seifert et al. 2011).

**Nectria** (Fr.) Fr., *Summa veg. Scand.*, *Section Post.* (Stockholm): 387 (1849)

The genus *Nectria* is characterized by its brightly coloured perithecia with 82 estimated species (Kirk et al. 2008). However, more than 1000 records are listed in *Index Fungorum* (2016). Hirooka et al. (2012) revised the genus

*Nectria* with 29 species based on morphology and phylogenetic analyses and provided full descriptions, illustrations and keys to sexual and asexual species. The asexual morphs of *Nectria* are linked to *Acremonium*-like, *Cylindrocarpon*, *Gyrostroma*, *Stilbella*, *Tubercularia*, and *Zythiostroma* species (Kirk et al. 2008; Wijayawardene et al. 2012; Hirooka et al. 2012).

**Nectria pseudotrichia** Berk. & M.A. Curtis, *J. Acad. Nat. Sci. Philadelphia* 2, 2: 289. 1853.

**Facesoffungi number:** FoF 01990; Figs. 35 and 36

**Saprobic** on bamboo culms. **Sexual morph:** Not observed.

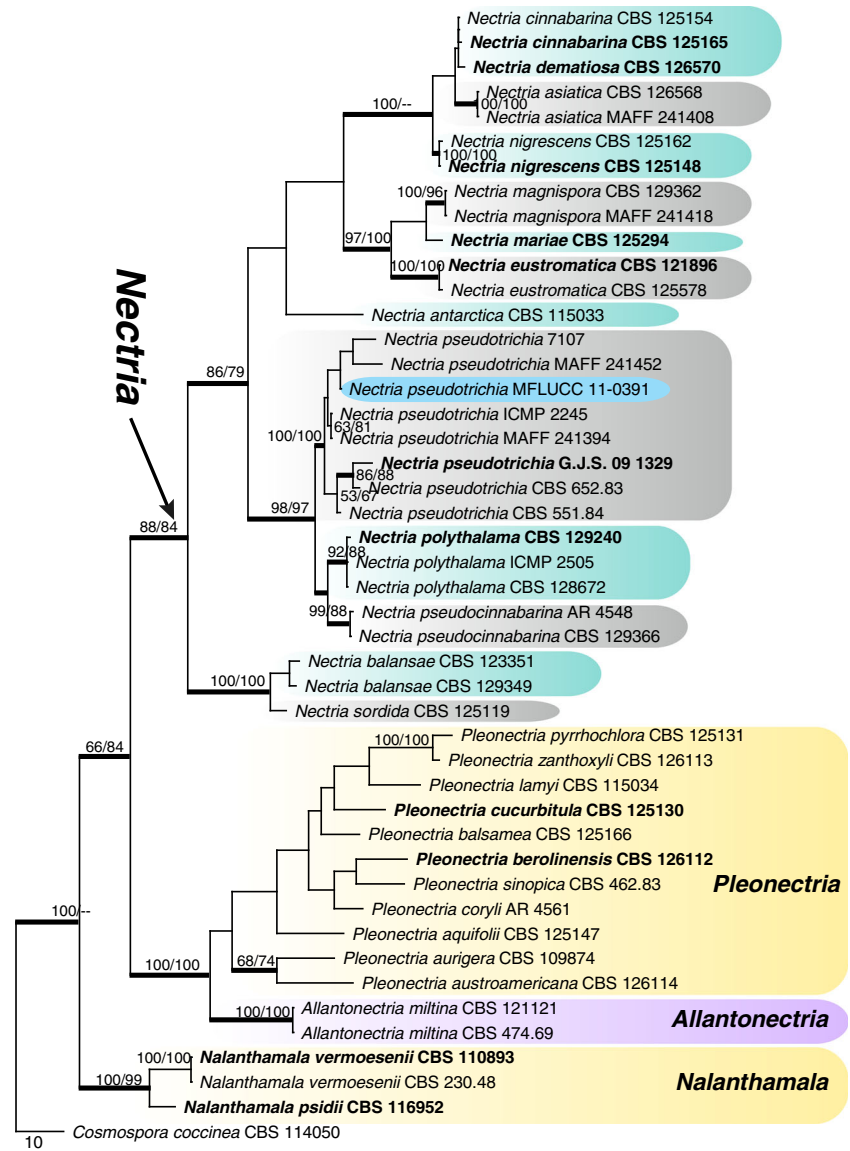
**Asexual morph:** Colonies effuse. Mycelium partly immersed on the substrate, composed of septate, branched, brown, hyphae. Conidiophores macronematous, synnematos, orange, septate, branched. Synnemata solitary or gregarious, erect, rigid, orange, branched in top half, hyaline at the tip, velvety, smooth, composed of compactly arranged conidiophores, 1.5–3.5 mm long, 200–270 µm wide at the base, 110–150 µm wide in the middle, 20–50 µm wide at the tip, fertile only apically, usually flexuous to curved in upper parts. Conidiogenous cells 12–23 × 1.5–3 µm ( $\bar{x}$  = 15.1 × 1.9 µm, *n* = 20), enteroblastic, phialidic, integrated or discrete, cylindrical, hyaline, smooth. Conidial mass 75–120 µm diam., globose, light orange in colour. Conidia 3.5–5 × 2–3 µm ( $\bar{x}$  = 4.6 × 2.7 µm, *n* = 20) µm, cylindrical, straight, aseptate, hyaline, smooth-walled.

**Culture characters:** Conidia germinating on PDA within 24 h. Colonies fast growing on PDA, reaching 90 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, flat, circular, hyaline with reddish orange pigmentation from above and below. Mycelium immersed in media, later on the surface, with septate, branched, smooth, hyaline hyphae.

**Material examined:** THAILAND, Chiang Rai, Doi Fung, on dead culms of bamboo, 12 May 2011, Dong-Qin Dai DDQ0005 (MFLU 15–1179); *Ibid.* (KUN HKAS88690), living cultures, MFLUCC 11–0391, KUMCC.

**Notes:** *Nectria pseudotrichia* was introduced by Berkeley and Curtis (1853) based on a sexual morph on bark. This species is widely spread especially in tropical areas and usually occurs on dead wood (Hirooka et al. 2012). Hirooka et al. (2012) linked *N. pseudotrichia* to *Tubercularia lateritia* (Berk.) Seifert. An epitype was established by Hirooka et al. (2012) with a detailed description and illustration. Our new collection (MFLUCC 11–0391) grouped with seven *N. pseudotrichia* strains including ex-epitype (CBS 129240) with high bootstrap support (100/100 % MPBP/MLBP) (Fig. 35). However, our collection has longer, branched synnemata (1.5–3.5 mm vs. 1–2.7 mm compared to its epitype) and a smaller conidia mass (75–120 µm vs. 120–350 µm in diam.) (Hirooka et al. 2012). Zhao et al. (2011) mentioned that, sometimes, two different *Nectria* species may share the same sequence data of the internal transcribed spacers of ribosomal RNA (*ITS*) and 28S rDNA (*LSU*), while differing in

**Fig. 35** One of eight equally most parsimonious trees resulting from 1000 bootstrap replicates obtained from combined ITS, LSU sequence data. Maximum likelihood phylogenetic tree (lnL = -3031.102133) was generated by RAxML (GTR+G model). MP/ML values (>50 %) are given at the nodes, and branches with Bayesian posterior probabilities greater than 0.90 are given in bold. The original isolate numbers are noted after the species names. The tree is rooted to *Cosmospora coccinea* (CBS 114050). Ex-type strains are in bold and newly generated sequence is highlighted with a blue background



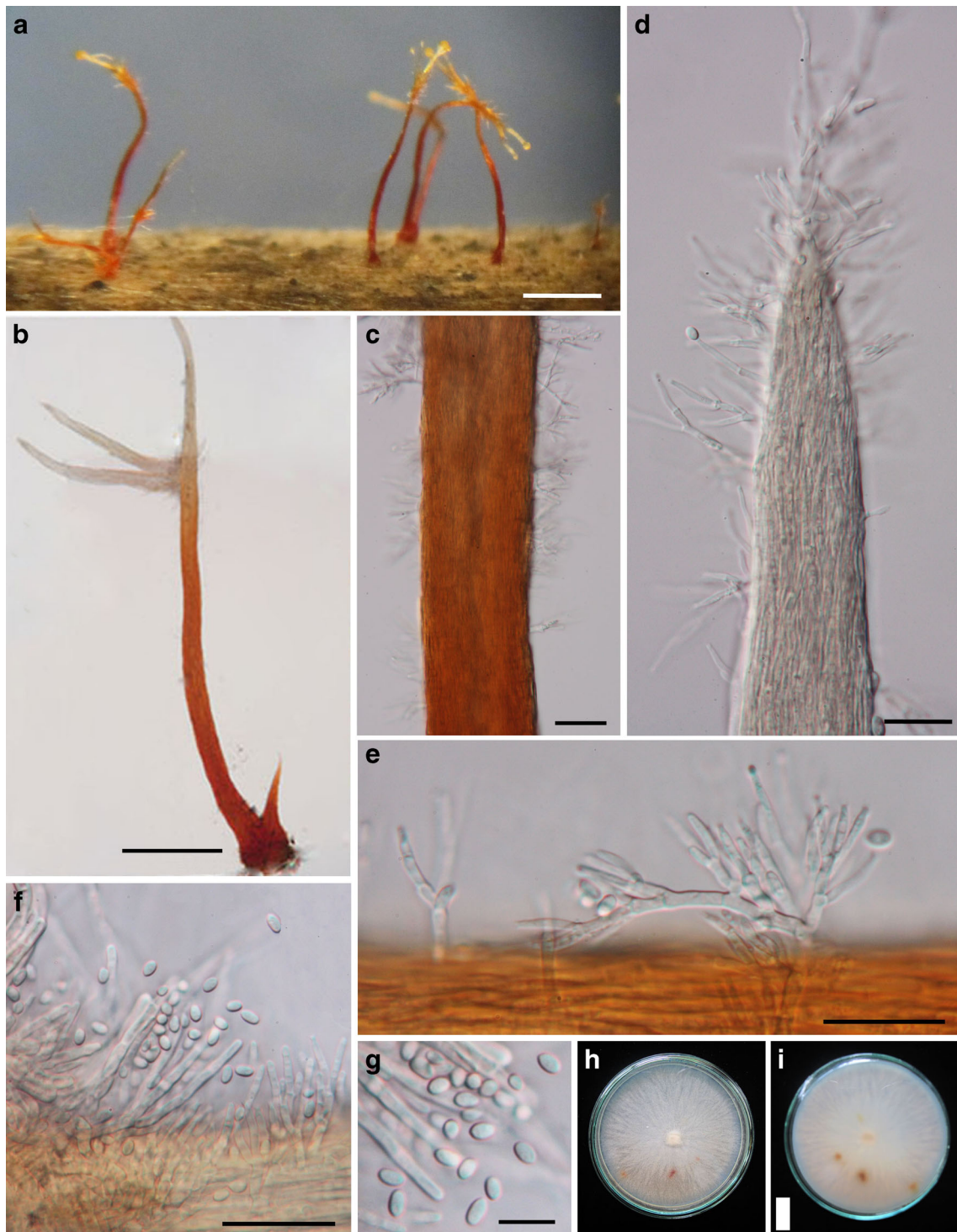
morphology. In the absence of an appropriate DNA barcode for *Nectria*, we provisionally report our collection as *N. pseudotrachia*.

***Sordariales*** Chadeff. ex D. Hawksw. & O.E. Erikss., Syst. Ascom. 5(1): 182 (1986)

The order *Sordariales* was introduced by Hawksworth and Eriksson (1986). Huhndorf et al. (2004) mentioned that 14 families are included in this order. However, Kirk et al. (2008) included only five families with 97 genera. Lumbsch and Huhndorf (2010) and Maharachchikumbura et al. (2015) accepted three families, *Chaetomiaceae*, *Lasiosphaeriaceae* and *Sordariaceae* and placed around 20 genera in *Sordariales* genera *incerta sedis*. The species accommodated in the order *Sordariales* usually have erumpent to superficial, perithecial ascomata with large-celled ascomal walls, unitunicate asci and one to two celled ascospores with appendages or sheaths (Huhndorf et al. 2004; Zhang et al. 2006).

***Lasiosphaeriaceae*** Nannf., Nova Acta R. Soc. Scient. upsal., Ser. 4 8(no. 2): 50 (1932)

*Lasiosphaeriaceae*, introduced by Nannfeldt (1932), is typified by *Lasiosphaeria* Ces. & De Not. Kirk et al. (2008) included 36 genera in this family and of these, Lumbsch and Huhndorf (2010) accepted 31 genera, whereas Maharachchikumbura et al. (2015) listed 35 genera. The family *Lasiosphaeriaceae* is the largest and most morphologically diverse familial taxon in the order *Sordariales* (Huhndorf et al. 2004). It is also highly polyphyletic (Miller and Huhndorf 2004; Huhndorf et al. 2004; Cai et al. 2006; Krays et al. 2015), containing several paraphyletic genera (Miller and Huhndorf 2005; Krays et al. 2015). Krays et al. (2015) demonstrated that the monophyletic family *Sordariaceae* phylogenetically nested among *Lasiosphaeriaceae* with taxa traditionally placed in the family. The phylogenetic tree given in this paper shows a



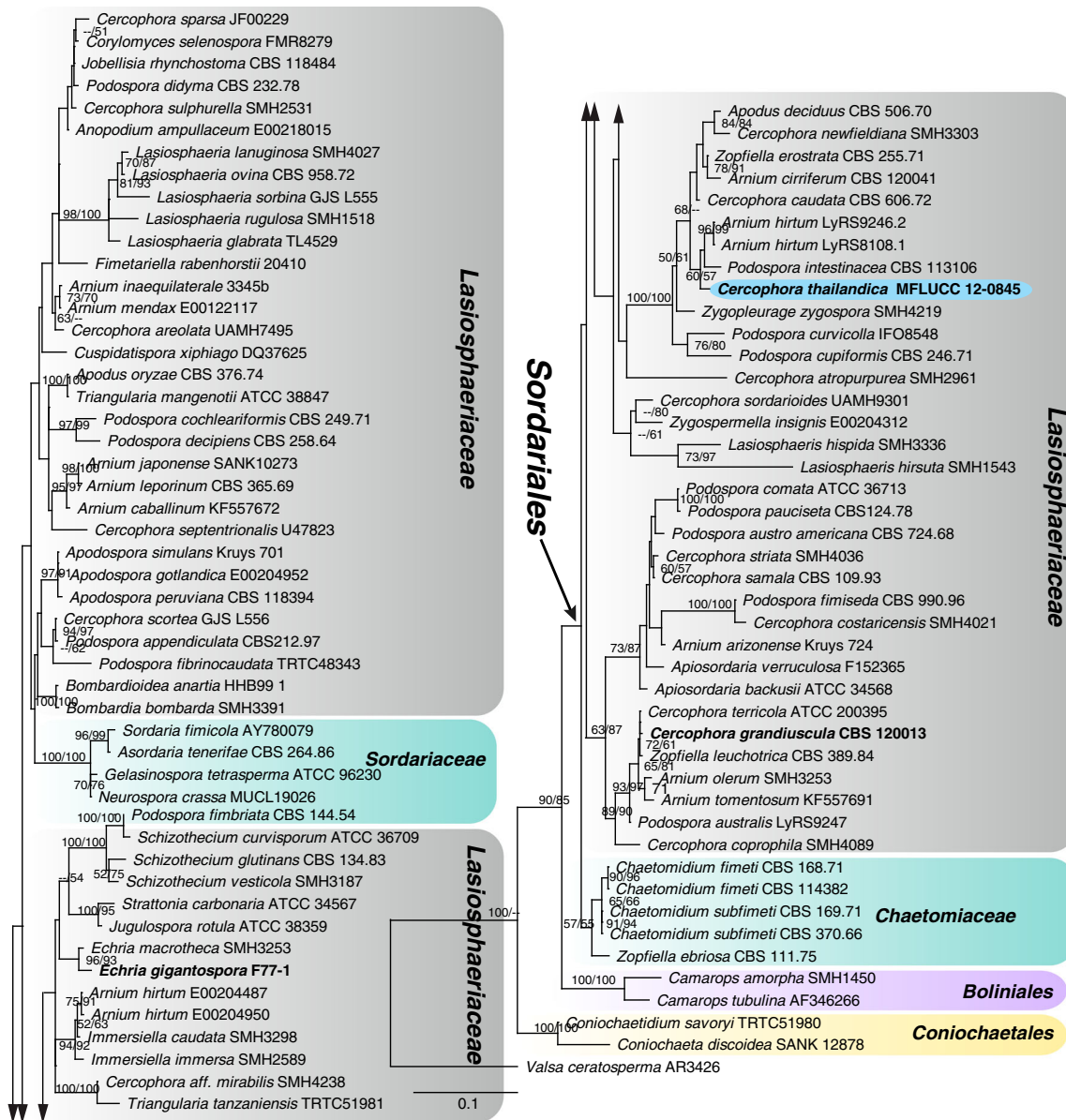
**Fig. 36** *Nectria pseudotrichia* (MFLU 15–1179). **a, b** Synnemata. **c** Middle part of synnema. **d** Apex of synnema with conidiogenous cells. **e** Conidiogenous cells. **f, g** Conidia. **h–i** Cultures on PDA. Scale bars: **a** = 1 mm, **b** = 500  $\mu\text{m}$ , **c** = 50  $\mu\text{m}$ , **d–f** = 25  $\mu\text{m}$ , **g** = 10  $\mu\text{m}$

similar result (Fig. 37). The asexual morph of *Lasiosphaeriaceae* can be linked to *Angulimaya*, *Bagadiella*, *Cladorrhinum*, *Mammaria* and *Phialophora*-like taxa (Wijayawardene et al. 2012).

*Cercophora* Fuckel, Jb. nassau. Ver. Naturk. 23–24: 244 (1870) [1869–70]

The genus *Cercophora* was introduced by Fuckel (1870) to accommodate taxa having membranous to coriaceous, dark ascomata and hyaline, cylindrical ascospores which develop a swollen, pigmented head (Miller and Huhndorf 2001). *Cercophora* can be compared with genera such as *Arnium*, *Lasiosphaeria*, *Podospora* and





**Fig. 37** Maximum likelihood phylogenetic tree generated by RAxML (GTR+G model) based on LSU sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes. The

*Zopfiella*. However, species of *Lasiosphaeria* do not form ascospores with a swollen, pigmented head (Seaver 1912; Taylor et al. 2001; Miller and Huhndorf 2001). Members of *Arnium* and *Podospora* generally grow on dung and produce ascospores with gelatinous appendages (Krug and Cain 1972; Lorenzo and Havrylenko 2001; Melo et al. 2015). *Zopfiella* has septate ascospores with a hyaline pedicel which often collapses (Cai et al. 2006). Phylogenetic analyses (Miller and Huhndorf 2005; Cai et al. 2006; Krays et al. 2015) showed that *Arnium*, *Cercophora*, *Podospora* and *Zopfiella* are polyphyletic. Thus, presently members in these groups are distinguished following the conventional taxonomy of Lundqvist (1972).

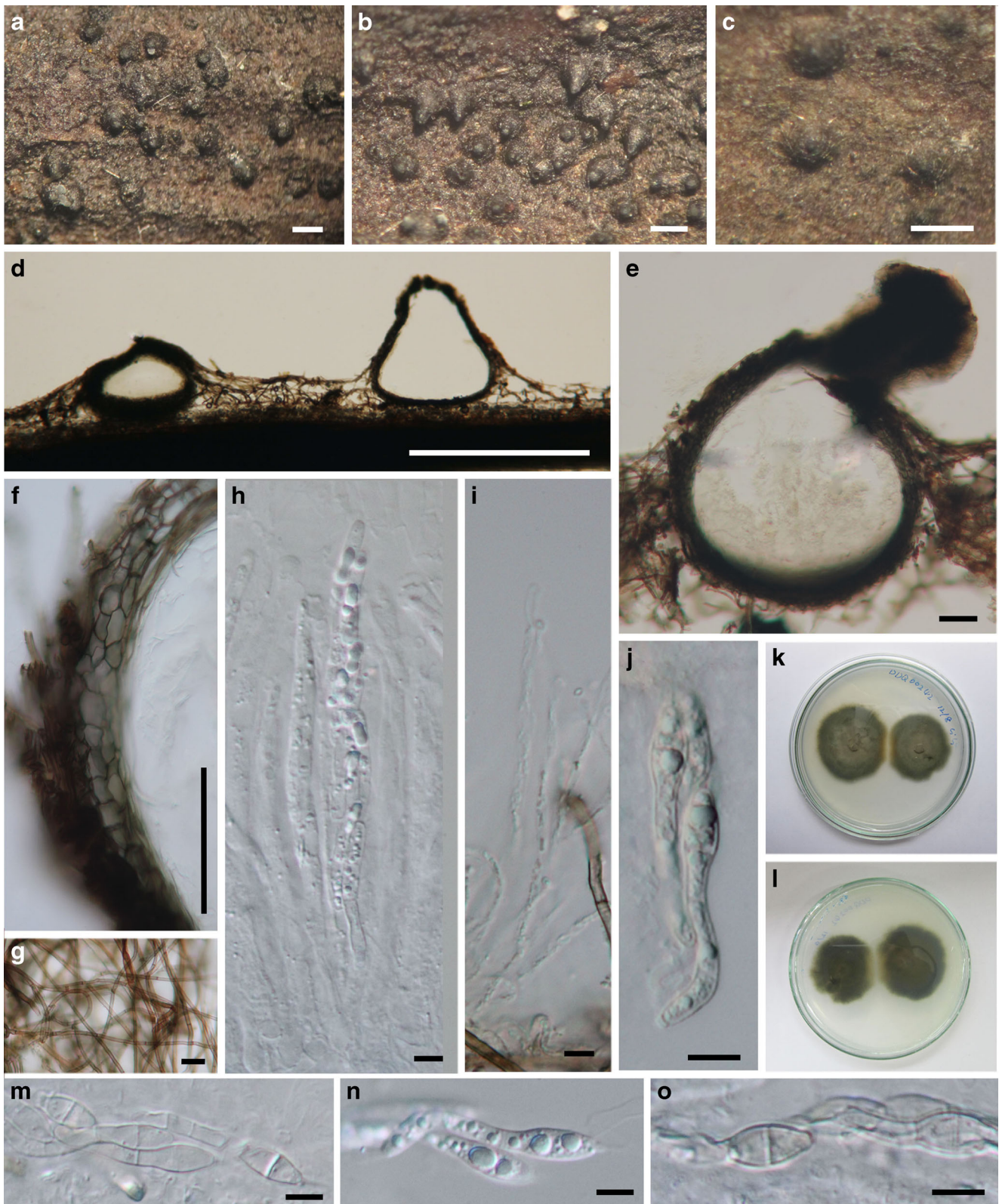
original isolate numbers are noted after the species names. The tree is rooted to *Valsa ceratosperma* (AR3426). Ex-type strains are in **bold** and newly generated sequences are highlighted with a blue background

***Cercophora thailandica*** D.Q. Dai & K.D. Hyde, *sp. nov.*  
Index Fungorum number: IF552034; Facesoffungi number: FoF 01991; Fig. 38

**Entomology:** In reference to the country where this species first collected.

**Holotype:** MFLU 15–1202

**Saprobic** on rotting bamboo culms. *Hyphae* 1.5–3 µm wide, superficial, dark brown, septate, branching, thick-walled, surrounding ascomata in a false subiculum. **Sexual morph:** *Ascomata* 350–480 µm high, 300–350 µm diam., perithecial, solitary, superficial, with base immersed in the hyphae, subglobose to pyriform, black, membranous, centrally located ostiole with a short neck, without periphyses, easily removed, with a rough surface and lackingsurface



**Fig. 38** *Cercophora thailandica* (MFLU 15–1202, holotype). **a–c** Appearance of ascomata on bamboo host. **d, e** Vertical sections of ascomata. **f** Peridium. **g** Brown mycelium. **h** Asci. **i** Paraphyses. **j, m–o** Ascospores. **k, l** Colonies on PDA. Scale bars: **a–d** = 500  $\mu\text{m}$ , **e, f** = 50  $\mu\text{m}$ , **g–j, m–o** = 10  $\mu\text{m}$

hairs, 150–180 × 100–120 µm. *Peridium* 10–25 µm thick, comprising two layers; outer layer composed of dark brown, 7–20 × 5–9 µm cells of *textura angularis*; inner layer thin, composed of elongated, spindly, light brown to hyaline cells. *Hamathecium* of septate, unbranched, 1.5–4.5 µm wide, filiform-ventricose paraphyses. *Asci* 90–110 × 10–15 µm ( $\bar{x}$  = 101.7 × 10.8 µm,  $n=20$ ), 8-spored, unitunicate, cylindrical, apex blunt, with a short pedicel. *Ascospores* 30–35 × 3–3.5 µm, 2-seriate, cylindrical, hyaline, curved, rounded at both ends; upper part becoming swollen into narrowly fusiform to ellipsoid, 9.5–15 × 4–6.5 µm ( $\bar{x}$  = 10.9 × 5.3 µm,  $n=20$ ) 0–1-septate, hyaline to pale brown head, with conical appendage at the apex, 3–5 µm long, truncate at the base; lower part elongated 18–23.5 × 3–3.5 µm ( $\bar{x}$  = 20.5 × 3.2 µm,  $n=20$ ) 3-septate, curved, slightly sigmoid, with 5–7 µm long and 2 µm wide appendage at the base. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 48 h and germ tubes produced from both ends. Colonies circular, with even margin, dark brown, floccose, fast growing, 40 mm diam. in 45 days at 25–32 °C, under 12 h light/12 h dark, flat, olive to olive brown from above and below. Mycelium immersed and partially superficial, branched, septate, pale brown to olivaceous brown, smooth, asexual spores not formed within 60 days.

**Material examined:** THAILAND, Chiang Rai, Khun Korn Waterfall, on rotting culm of bamboo, 31 July 2012, Dong-Qin Dai DDQ00242 (MFLU 15–1202, **holotype**); *Ibid.* (KUN HKAS88711, **isotype**), living cultures, MFLUCC 12–0845, KUMCC.

**Notes:** *Cercophora* species usually grow on dung or occasionally rotten wood (Miller and Huhndorf 2001). *Cercophora thailandica* is similar to *C. atropurpurea* in ascospores having a swollen head. However, *C. thailandica* has shorter ascomata with a short neck (350–480 µm vs. 620–790 µm in height) (Miller and Huhndorf 2001). This new species is phylogenetically close to *Arnium hirtum* (LyRS9246.2 and LyRS8108.1) and *Podospora intestinacea* (CBS 113106). *Arnium hirtum* has ascomata with hairs, and large dark brown ascospores (more than 45 × 20 µm), with long appendages (more than 35 × 7 µm) (Lundqvist 1972). *Podospora intestinacea* has ascospores with typical dark brown apical cell and a hyaline basal pedicel (Doveri 2008). *Arnium*, *Cercophora* and *Podospora* are interesting because of their coprophilous habitat. They generally appear on herbivore dung, however, our new species was collected on a rotten bamboo culm.

**Togniniales** Senan., Maharachch. & K.D. Hyde, Fungal Diversity 72: 220 (2015)

*Togniniales* was introduced by Maharachchikumbura et al. (2015) and contains a single family, *Togniniaceae*. This family was previously placed in the *Calosphaeriales* (Mostert et al.

2003). Based on SSU and LSU gene analysis, Mostert et al. (2006) transferred *Togniniaceae* to *Diaporthales*. Maharachchikumbura et al. (2015) indicated that *Togniniaceae* formed a distinct clade from *Diaporthales* in the subclass *Diaportheomycetidae* based on phylogenetic analysis of combined LSU, SSU, TEF and RPB2 sequence data. Lumbsch and Huhndorf (2010) included a single genus *Togninia* in the family *Togniniaceae*. Maharachchikumbura et al. (2015) listed three genera, including a sexual morph genus *Phaeoacremonium*. Gramaje et al. (2015) presented a case for retaining the genus *Phaeoacremonium* over that of *Togninia*.

**Phaeoacremonium sphinctrophorum** L. Mostert et al., Stud. Mycol. 54: 85 (2006)

**Facesoffungi numbers:** FoF 01992; Figs. 39 and 40

**Saprobic** on dead bamboo culms. **Sexual morph:** *Ascomata* perithecial, 100–150 µm high, 170–250 µm diam., dark brown to black, immersed under epidermis to erumpent, gregarious, uniloculate, globose to subglobose, coriaceous; with a central neck 20–40 µm long × 10–15 µm wide, black, straight to curved. *Peridium* comprising several strata, outer stratum 8–10 µm thick, composed of small, dark brown to brown cells of *textura angularis*; inner stratum composed of larger, pale brown to hyaline cells of *textura angularis*. *Paraphyses* 15–25 µm long × 2–3.5 µm wide, hyaline, septate. *Asci* form small fascicles arising from short ascogenous hyphae, 15–20 × 3.5–4 µm ( $\bar{x}$  = 18.8 × 3.7 µm,  $n=20$ ), 8-spored, unitunicate, clavate to cylindro-clavate, pedicellate, apically rounded to truncate, J-; ascogenous hyphae hyaline, branched. *Ascospores* 3–4 × 1–1.5 µm ( $\bar{x}$  = 3.5 × 1.4 µm,  $n=30$ ), 2–3-seriate, allantoid, hyaline, rounded at both ends, smooth-walled. **Asexual morph:** Fertile on PDA after 2 months. *Conidiophores* micronematous, septate, hyaline to brown, smooth, verrucose wall not observed. *Conidiogenous cells* enteroblastic, monophialidic, terminal or lateral, hyaline, smooth; phialides of three types: type I 2–7 × 1.5–2 µm cylindrical, raised from the hypha; type II 7–12 × 1.5–3 µm elongate-ampulliform; type III 17–21 × 1.5–2.5 µm subcylindrical. *Conidia* 3–4.5 × 1.5–2 µm ( $\bar{x}$  = 3.8 × 1.7 µm,  $n=20$ ), oblong, ellipsoidal or obovoid, hyaline, smooth-walled.

**Culture Characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from both ends. Colonies growing fast on PDA, reaching 50 mm in 1 week at 28 °C, under 12 h light/12 h dark, circular, irregular edge, white from above and pale red from reverse view. Mycelium superficial to immersed in media, branched, septate, smooth.

**Material examined:** THAILAND, Chiang Mai, Mae Taeng, Mushroom Research Foundation, on dead culms of bamboo, 6 October 2011, Dong-Qin Dai DDQ00210 (MFLU 15–1196); *Ibid.* (KUN HKAS88705), living cultures, MFLUCC 11–0629, MUCL 55877.

**Notes:** *Phaeoacremonium* species are commonly isolated from stems and branches of diseased woody hosts,

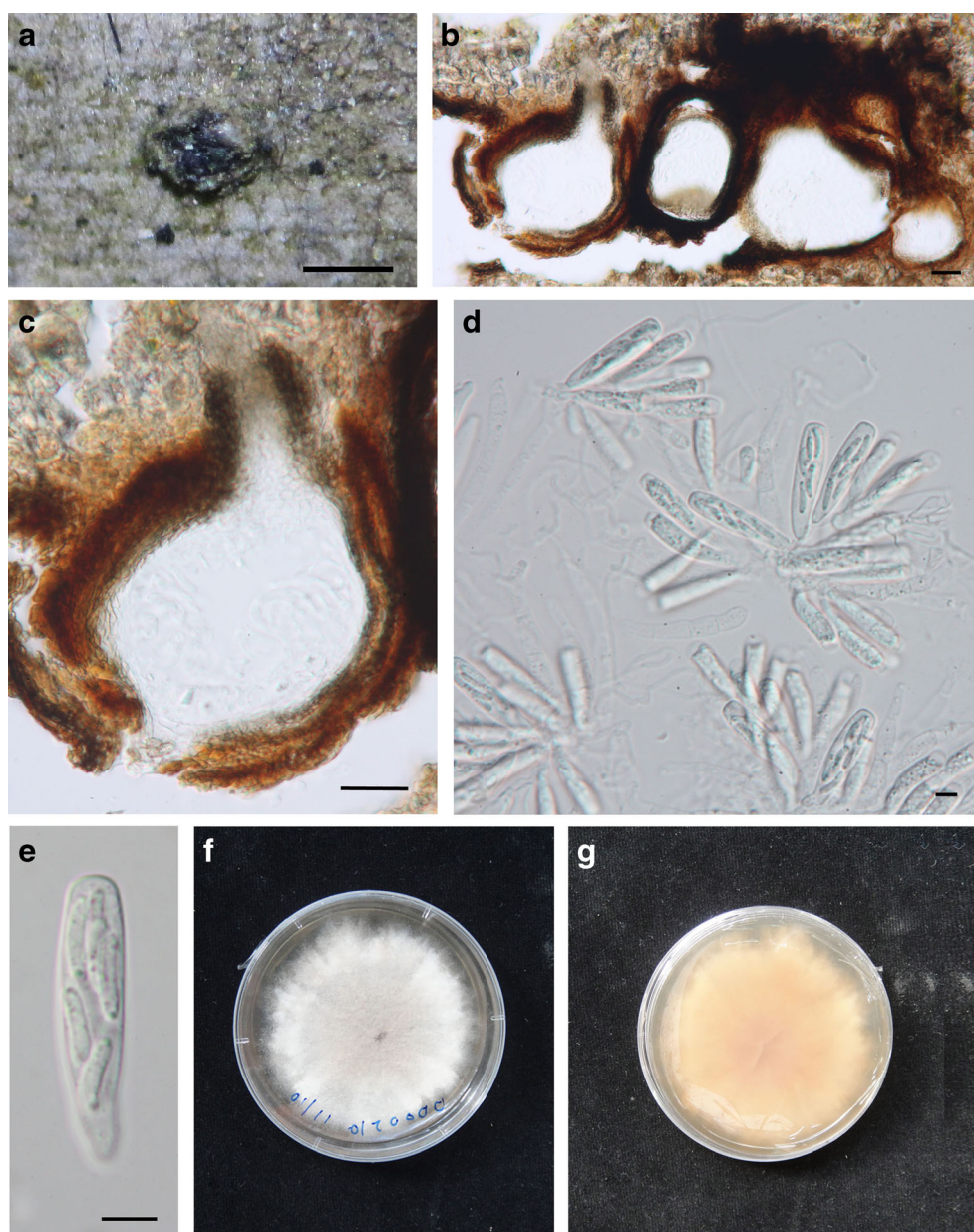
humans with phaeohyphomycosis, and occasionally from soil (Crous et al. 1996; Mostert et al. 2003, 2006; Essakhi et al. 2008; Graham et al. 2009; Gramaje et al. 2015). Gramaje et al. (2015) monographed *Phaeocremonium* and expanded the genus, which presently contains 46 species and its sexual morph, *Togninia*, containing 26 epithets. *Phaeocremonium sphinctrophorum* was described by Mostert et al. (2006) based on asexual morph characters of a collection inhabiting a human in Canada. In our study, we collected the sexual morph on a bamboo stub in Thailand, and observed the asexual morph on PDA, which has same morphology with the type. Our collection forms a clade with two strains of *P. sphinctrophorum*, including the type

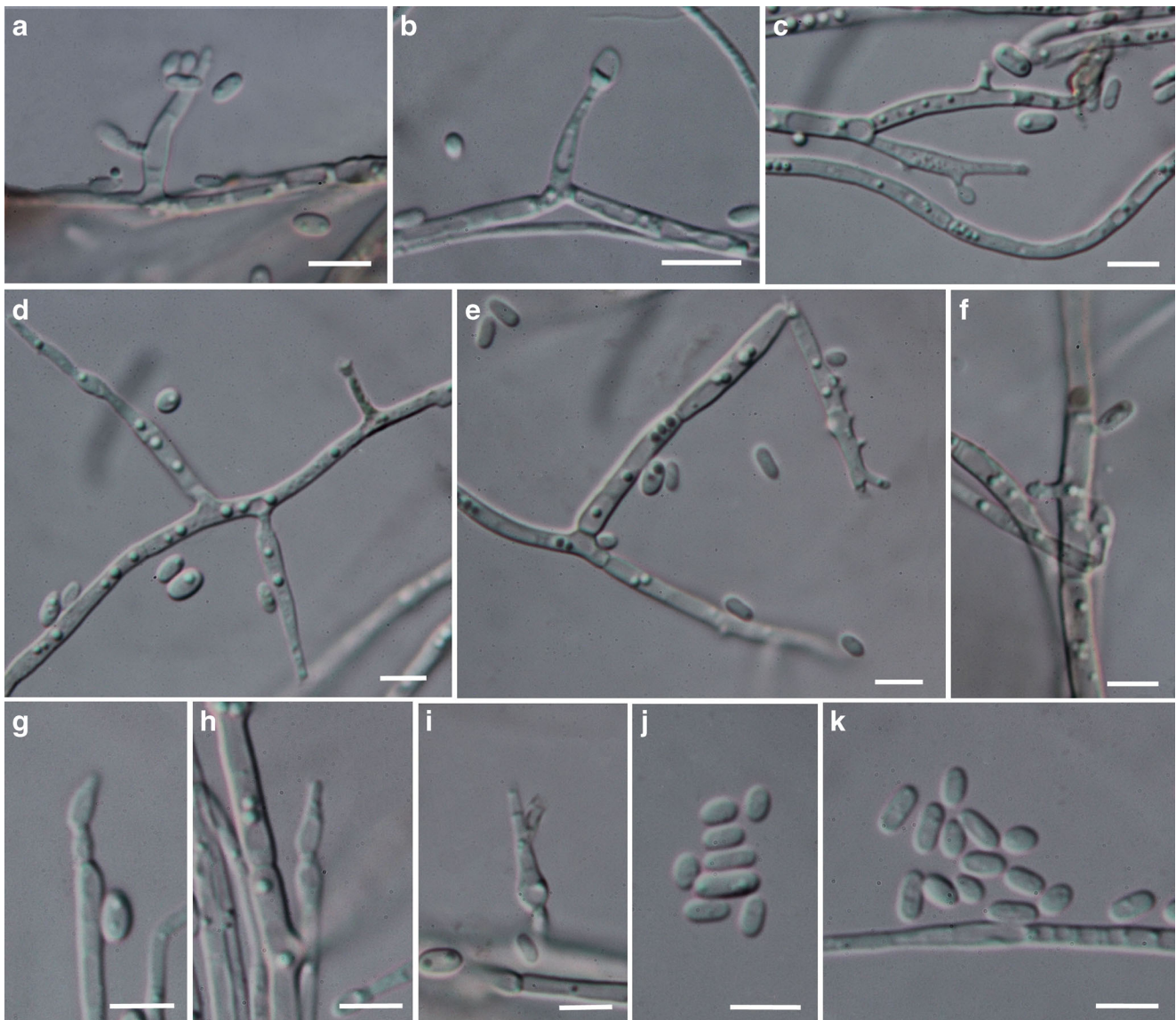
strain (CBS 337.90) in the phylogeny tree (Fig. 41) with high bootstrap support (99/91 % MPBP/MLBP).

*Xylariales* Nannf., Nova Acta R. Soc. Scient. upsal., Ser. 4 8(no. 2): 66 (1932)

The order *Xylariales* is a large taxon of unitunicate, perithecial ascomycetes within the class *Sordariomycetes* (Smith et al. 2003; Kirk et al. 2008; Jaklitsch and Voglmayr 2012; Daranagama et al. 2015; Maharachchikumbura et al. 2015; Senanayake et al. 2015). Smith et al. (2003) stated that this order contains seven families, over 92 genera and 795 species. Kirk et al. (2008) included nine families, 209 genera and 2487 species. Lumbsch and Huhndorf (2010) accepted six families and 154 genera. Based on phylogenetic

**Fig. 39** *Phaeocremonium sphinctrophorum* (MFLU 15–1196). **a** Appearance of ascomata on bamboo. **b, c** Vertical sections of ascomata. **d, e** Asci containing ascospores. **f, g** Cultures on PDA. Scale bars: a = 1 mm, b, c = 20  $\mu$ m, d–e = 5  $\mu$ m





**Fig. 40** Asexual morph of *Phaeoacremonium sphinctrophorum* from PDA (MFLUCC 11–0629). **a–i** Conidiogenous cells (**a, c, f** type I phialides, **g–i**: type II phialides, **b, d, e**: type III phialides). **j, k** Conidia. Scale bars: **a–k** = 5  $\mu$ m

analysis, eleven families are circumscribed in the *Xylariales* by Maharachchikumbura et al. (2015). Senanayake et al. (2015) transferred *Amphisphaeriaceae* to *Amphisphaeriales*, and introduced two new families *Lopadostomaceae* and *Pseudomassariaceae*, based on morphological and phylogenetic evidence. Thus, this order is expanded to contain 12 families.

**Apiosporaceae** K.D. Hyde et al., Sydowia 50(1): 23 (1998)

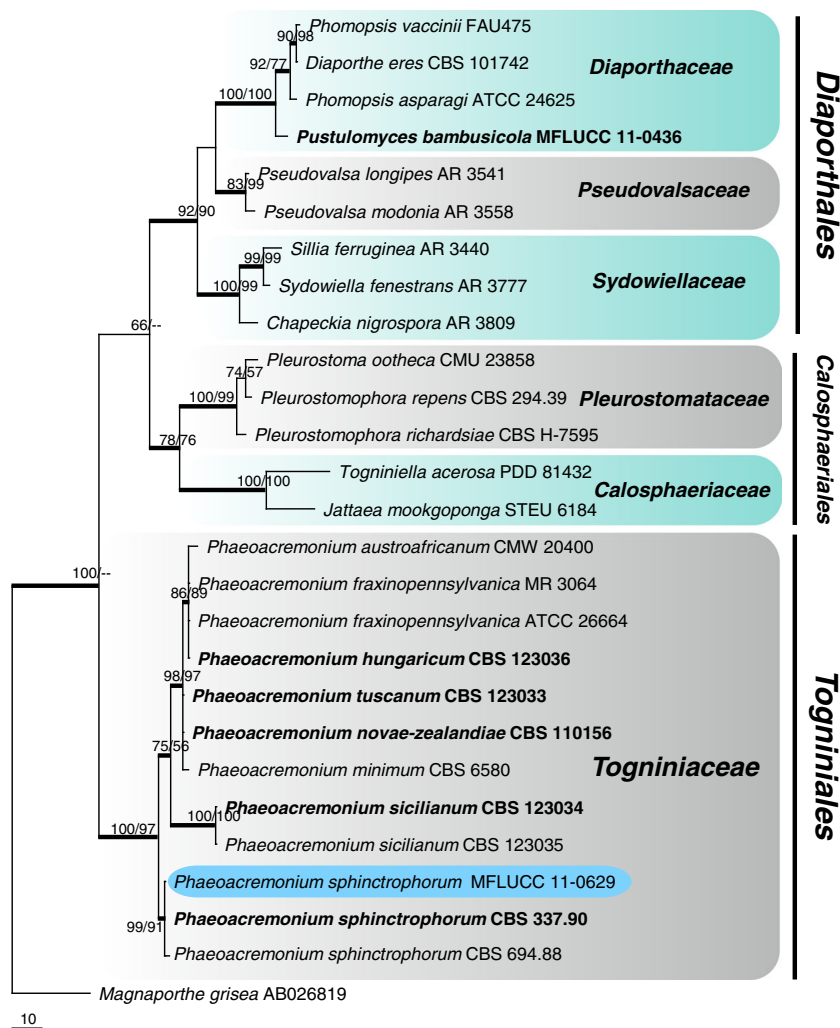
*Apiosporaceae* was introduced by Hyde et al. (1998), and typified by *Apiospora* Sacc. The family placement of *Apiospora* was undetermined during 1875 to 1995 (Müller and von Arx 1962; Barr 1976, 1990; Barr and Cannon 1994; Hawksworth et al. 1995). Subsequently Hyde et al. (1998) re-examined the type (*Apiospora montagnei* Sacc.) and introduced the family *Apiosporaceae* to accommodate *Apiospora*

(Sexual morph of *Arthrinium*) and *Appendicospora*. The family placement of *Apiospora* has been clarified as *Apiosporaceae* has been recognized in order *Xylariales* by several authors (Huhndorf et al. 2004; Zhang et al. 2006; Jaklitsch and Voglmayr 2012; Dai et al. 2014c; Sharma et al. 2014; Senanayake et al. 2015). Crous and Groenewald (2013) synonymised *Apiospora* under *Arthrinium* Kunze. Thus, this family contains six genera, *Appendicospora*, *Arthrinium*, *Dictyoarthrinium*, *Endocalyx*, *Scyphospora* and *Spegazzinia* (Maharachchikumbura et al. 2015).

**Arthrinium** Kunze, in Kunze & Schmidt, Mykol. Hefte 1: 9 (1817) : Fr., Syst. Mycol. 1: xlv (1821)

*Arthrinium*, introduced by Schmidt and Kunze (1817), is widespread in diverse ecological niches (Crous and Groenewald 2013). The genus is morphologically and phylogenetically delimited by Crous and Groenewald

**Fig. 41** Maximum likelihood phylogenetic tree (lnL = -2056.989940) generated by RAxML (GTR+G model) based on LSU sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes. The original isolate numbers are noted after the species names. The tree is rooted to *Magnaporthe grisea* (M82). Ex-type strains are in **bold** and newly generated sequences are highlighted with a blue background



(2013) and Senanayake et al. (2015). The distinguishing characters of *Arthrinium* are the basauxic conidiogenesis in the asexual morph, with apiospores in the sexual morph (Samuels et al. 1981; Bahl 2006; Senanayake et al. 2015).

*Arthrinium longistromum* D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum number*: IF552035; *Facesoffungi number*: FoF 01993; Figs. 42 and 43

*Entomology*: Refers to its long stromata.

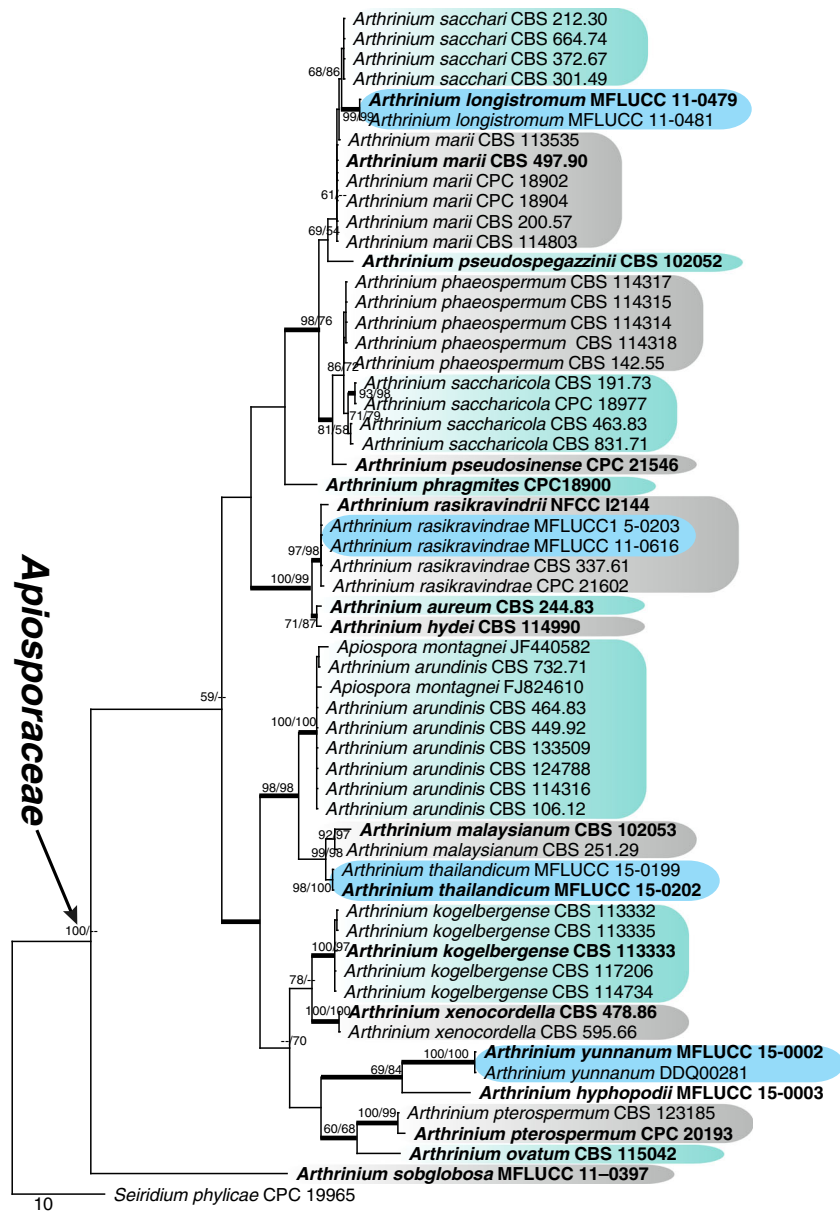
*Holotype*: MFLU 15-1184

*Saprobic* on decaying bamboo culms, forming black, linear, raised areas on the host surface, with ascostromata breaking through raised cracks at the black centre. **Sexual morph**: *Ascostromata* 5–30 mm long, 0.25–0.5 mm wide, 0.25–0.3 mm high, gregarious, raised, irregularly and sinuously filiform, black, with long axis broken at the apex, with black dot-like ostioles of ascostromata. *Ascomata* perithecial, 140–280 µm diam., 145–265 µm high, 10–30 gregarious in one ascostroma, subglobose, sometimes with flattened base, slight

brown to reddish brown, membranous, with a centrally located, periphysate, 50–80 × 60–75 µm wide, distinctive ostiole. *Peridium* laterally 10–15 µm thick, composed of brown and small light brown to reddish brown to hyaline cells of compressed *textura angularis* to *textura intricata*; upper wall near ostiole 25–35 µm thick, black to reddish black. *Cells between perithecia* 50–100 µm thick, comprising host and fungal tissues, composed of small, brown cells of *textura angularis*. *Hamathecium* composed of few, 4.5–7 µm broad, septate, unbranched, paraphyses. *Asci* 70–95 × 15.5–20 µm ( $\bar{x}$  = 79.7.6 × 18.4 µm,  $n$  = 20), 8-spored, unitunicate, broad cylindrical to clavate, with rounded apex and a short pedicel, J-. *Ascospores* 20–30 × 4.5–6.5 µm ( $\bar{x}$  = 23.1 × 5.7 µm,  $n$  = 20), 2–3-seriate, 1-septate, elliptical to broad fusiform, with a large, upper cell and small lower cell, with narrowly rounded ends, hyaline, smooth-walled, with many guttules, without a sheath. **Asexual morph**: Undermined.

*Culture characters*: Ascospores germinating on PDA within 24 h and germ tubes developing from the upper cell. Colonies slowly growing on PDA, reaching 60 mm in 4 weeks

**Fig. 42** One of four equally most parsimonious trees resulting from 1000 bootstrap replicates obtained from the ITS sequence data. Maximum likelihood phylogenetic tree (lnL = -2217.066944) was generated by RAxML (GTR+G model). MP/ML values (>50 %) are given at the nodes, and branches with Bayesian posterior probabilities greater than 0.90 are given in *bold*. The original isolate numbers are noted after the species names. The tree is rooted to *Seiridium phylicae* (CPC 19965). Ex-type strains are in *bold* and newly generated sequences are highlighted with a blue background



at 26 °C, under 12 h light/12 h dark, cottony, circular, with irregular edge, white from above, brown to dark brown in centre from below. Mycelium superficial to immersed in media, with branched, septate, smooth hyphae.

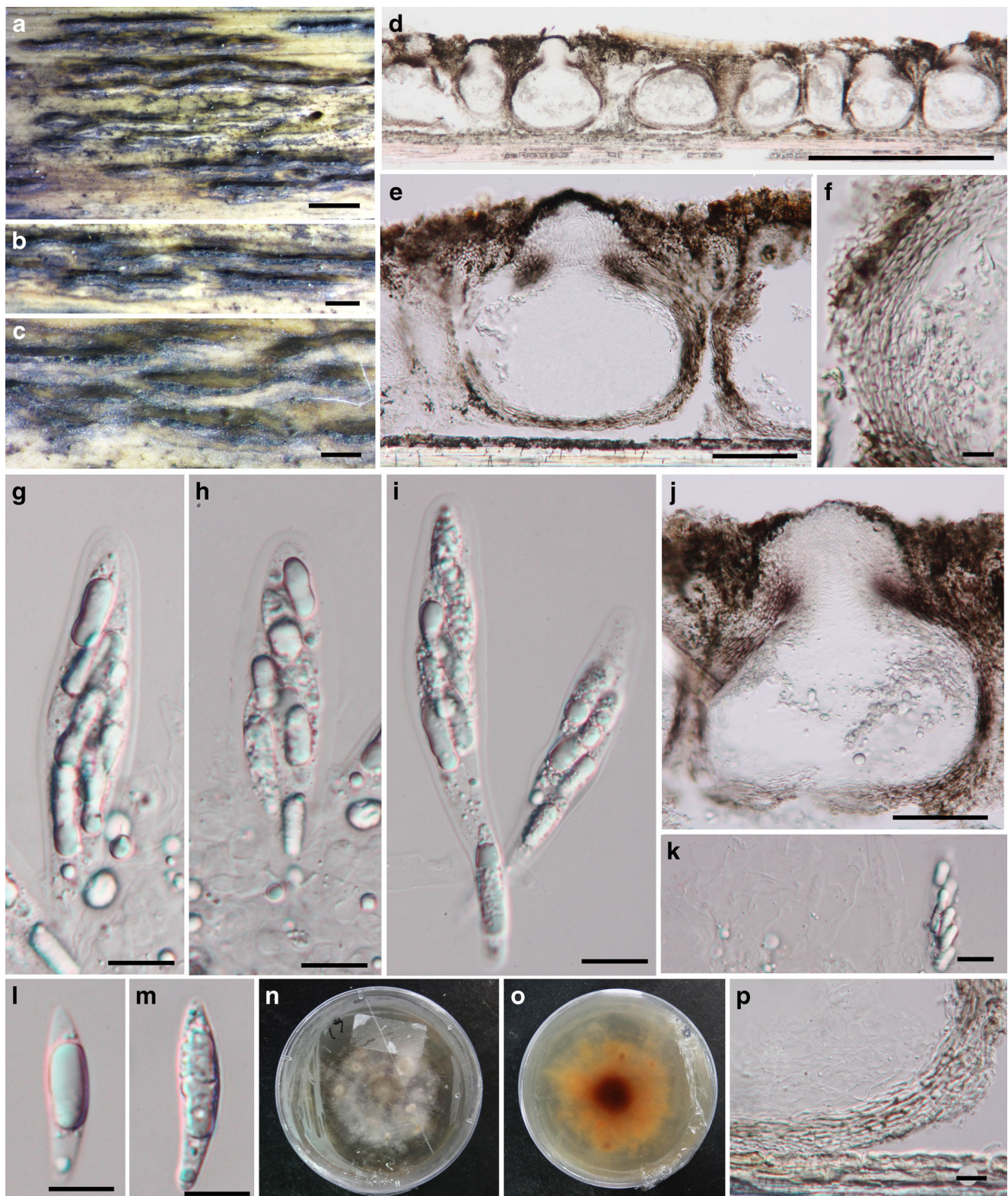
**Material examined:** THAILAND, Chiang Rai, Mae Fah Luang University, on dead culms of bamboo, 19 July 2011, Dong-Qin Dai DDQ00079 (MFLU 15-1184, **holotype**); *Ibid.* (KUN HKAS88694, **isotype**), living cultures, MFLUCC 11-0481, KUMCC; *Ibid.*, 17 July 2011, Dong-Qin Dai DDQ00074, living culture, MFLUCC 11-0479.

**Notes:** *Arthrinium longistromum* is characterized by the linear ascostromata (5–30 mm long) containing more than ten perithecia. *Arthrinium longistromum* is phylogenetically close to *A. sacchari* (Speg.) M.B. Ellis and *A. marii*

Larrondo & Calvo (Fig. 42). However, the branch length indicates that they are different species (Fig. 42).

*Arthrinium rasikravindrae* Shiv M. Singh et al., [as ‘*rasikravindrii*’], in Singh et al., Mycotaxon 122: 452 (2012) *Facesoffungi* number: FoF 01994; Figs. 44 and 45

**Saprobic** on dead bamboo culms, forming black, fusiform spots, with ascostromata breaking through raised cracks at the black centre. **Sexual morph:** *Ascostromata* 0.8–1.7 mm long, 0.4–0.6 mm wide, 0.15–0.2 mm high, solitary, immersed, fusiform, black, with a long broken axis at the top. *Ascomata* perithecial, 100–150 µm high, 200–300 µm diam., 3–5 perithecia immersed within the ascostromata, subglobose with a flattened base, dark brown, membranous, with centrally located, periphysate, 45–60 × 40–50 µm wide distinctive ostiole.



**Fig. 43** *Arthrinium longistromum* (MFLU 15–1184, holotype). **a–c** Dark, long ascostromata on bamboo host. **d** Vertical section of ascostroma. **e, j** Vertical sections of perithecia (**j**: showing periphysate

ostiole). **f, p** Peridium. **g–i** Asci. **k** Paraphyses. **l, m** Ascospores. **n, o** Cultures on PDA. Scale bars: **a** = 1 mm, **b–d** = 500  $\mu\text{m}$ , **e** = 100  $\mu\text{m}$ , **f** = 10  $\mu\text{m}$ , **j** = 50  $\mu\text{m}$ , **g–m, p** = 10  $\mu\text{m}$

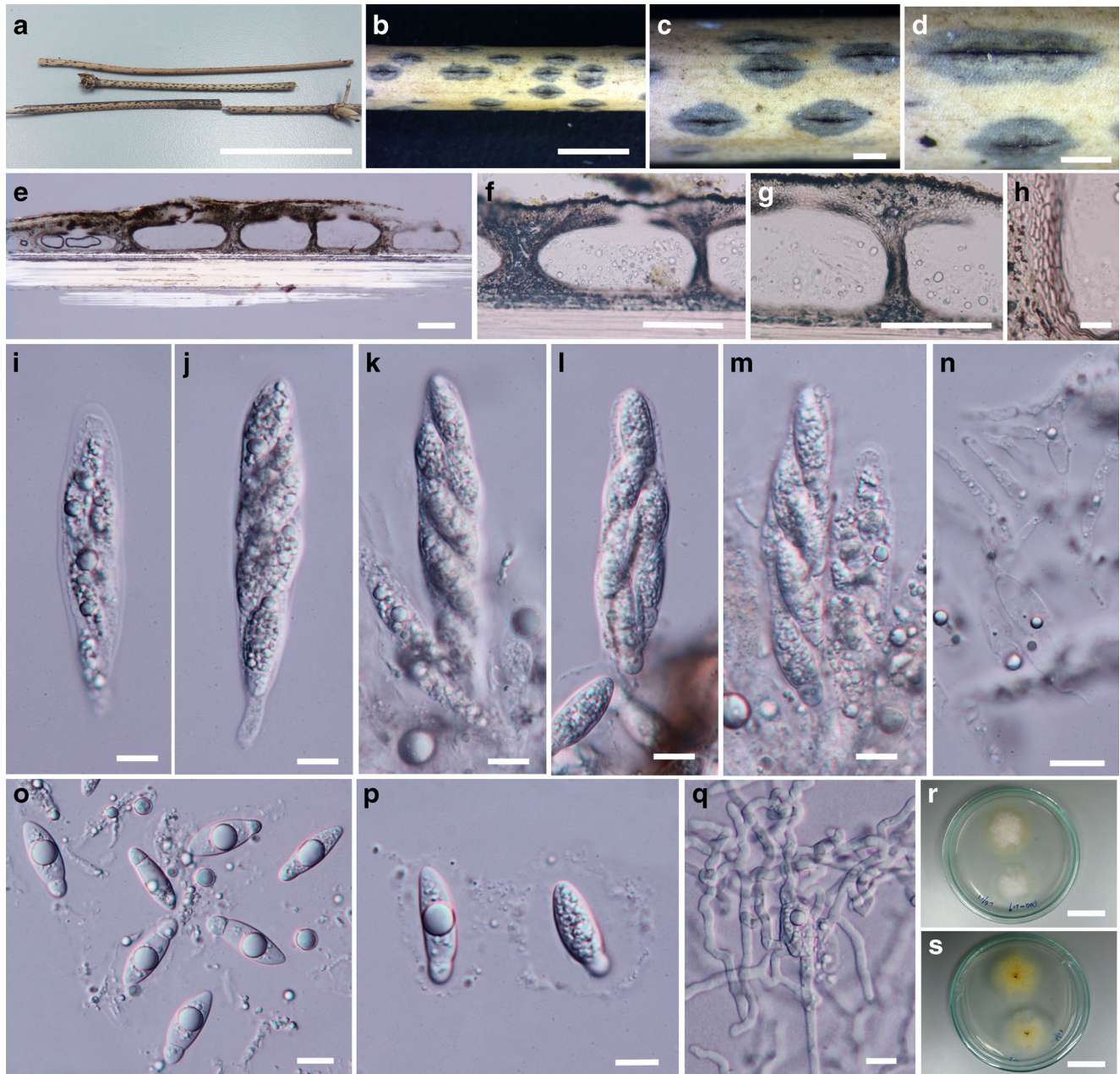
*Peridium* composed of dark brown to brown to hyaline cells of compressed *textura angularis* to *textura intricata*, laterally

5–10  $\mu\text{m}$  thick; upper wall around ostiole 10–15  $\mu\text{m}$  thick, with black pigment. *Cells between perithecia* 50–250  $\mu\text{m}$



thick, comprising host and fungal tissues, composed of small, brown cells of *textura angularis*. *Hamathecium* composed of dense, long, 1.5–3  $\mu\text{m}$  broad, septate, unbranched, paraphyses. *Asci* 70–90  $\times$  15–17.5  $\mu\text{m}$  ( $\bar{x}$  = 82.1  $\times$  16.6  $\mu\text{m}$ ,  $n=20$ ), 8-spored, unitunicate, broadly cylindrical to long clavate, with a short pedicel. *Ascospores* 21.5–24.5  $\times$  7–9.5  $\mu\text{m}$  ( $\bar{x}$  = 23.1  $\times$  8.3  $\mu\text{m}$ ,  $n=20$ ), 2–3-seriate, 1-septate, ellipsoidal to broad fusiform, with a large, occasionally curved, upper cell and a small lower cell, with narrowly rounded ends, hyaline, smooth-walled,

with many guttules and one large guttule in the centre, surrounded by 3.5–9  $\mu\text{m}$  wide gelatinous sheath attached at both ends. **Asexual morph:** Fertile on PDA, after 1 month. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* basauxic, cylindrical, discrete, smooth-walled. *Conidia* 7–10  $\times$  4.5–8.5  $\mu\text{m}$  ( $\bar{x}$  = 8.1  $\times$  6.8  $\mu\text{m}$ ,  $n=20$ ), lenticular, globose to subglobose, occasionally elongated to ellipsoidal, dark brown, smooth-walled, with a longitudinal, hyaline, thin, germ-slit.



**Fig. 44** *Arthrinium rasikravindrii* (MFLU 15–1227). **a** Bamboo specimens. **b–d** Black ascostromata on bamboo host. **e** Vertical section of ascostroma. **f, g** Vertical sections of perithecia. **h** Peridium. **i** Immature

ascus. **j–m** Mature asci. **n** Paraphyses. **o–p** Ascospores. **q** Germinating ascospore. **r, s** Cultures on PDA. Scale bars: **a** = 5 mm, **b** = 1 mm, **c, d** = 500  $\mu\text{m}$ , **e–g** = 100  $\mu\text{m}$ , **h–q** = 10  $\mu\text{m}$

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes developing from the upper cell. Colonies fast growing on PDA, reaching 90 mm in 2 weeks at 26 °C, under 12 h light/12 h dark, cottony, circular, with irregular edge, white from above, brown to dark brown in centre from below. Mycelium superficial to immersed in media, with branched, septate, smooth hyphae.

**Material examined:** THAILAND, Krabi, Wat Tham Sua Temple, 8°07'34"N 98°55'28"E, limestone outcrop, on dead culms of bamboo, 4 October 2014, Kevin D. Hyde DDQ00309 (MFLU 15–1227); *Ibid.* (KUN HKAS88734), living cultures, MFLUCC 15–0203, MUCL 55894; Chiang Rai, Mae Fah Luang University, 1 October 2011, Dong-Qin Dai DDQ00102, living culture, MFLUCC 11–0616.

**Notes:** *Arthrinium rasikravindrae* was introduced by Singh et al. (2012) based on the asexual morph characters and phylogeny analyses. The type was originally isolated from soil in Norway (Singh et al. 2012). Singh et al. (2012) also reported that this species occurred on wood from marine coast and *Pinus thunbergii* Parl., as well as stubble of *Triticum aestivum* L. However, only the asexual morph was observed by Singh et al. (2012). In this paper, we describe and illustrate both sexual and asexual morphs (Figs. 44 and 45). The conidia of our collection (MFLUCC 15–0203) are smaller than the type ( $7\text{--}10 \times 4.5\text{--}8.5 \mu\text{m}$  vs.  $10\text{--}15 \times 6\text{--}10.5 \mu\text{m}$ ) (Singh et al. 2012). However, in the phylogenetic tree (Fig. 42), our two isolates form same branch with ex-type strain (NFCCI 2144),

as well as two authentic strains (CBS 337.61 and CPC 21602) with high bootstrap support (97/98 % MPBP/MLBP).

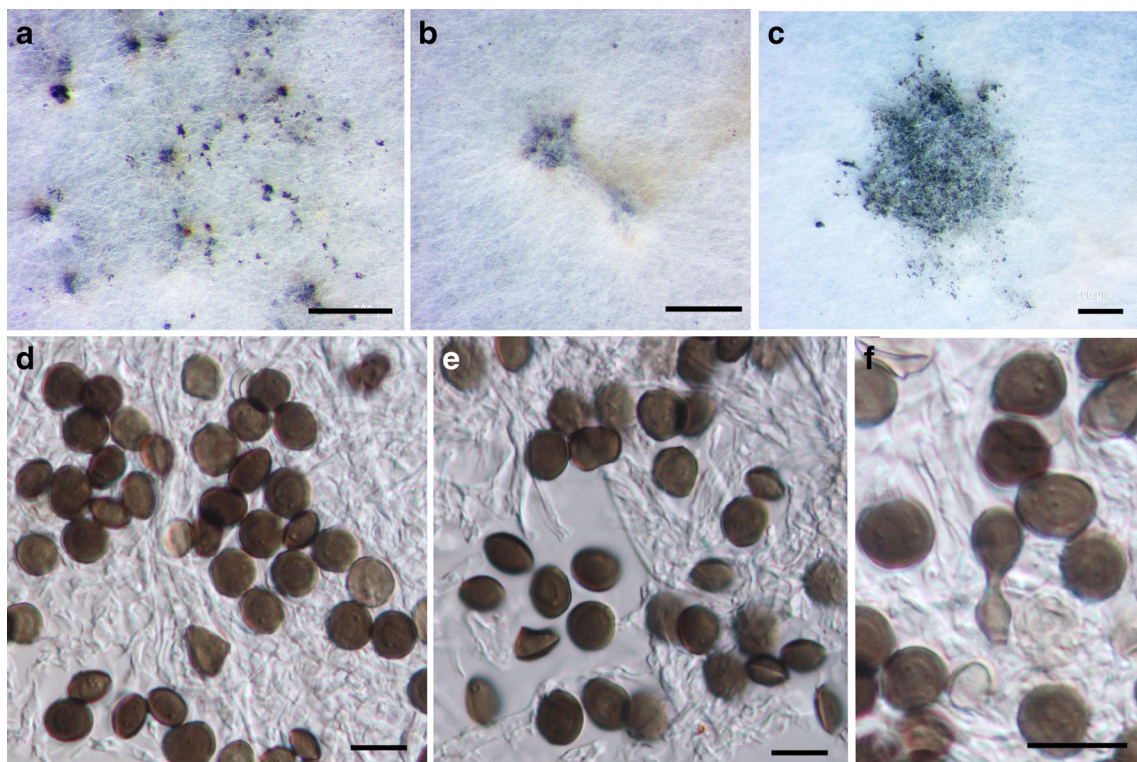
***Arthrinium thailandicum*** D.Q. Dai & K.D. Hyde, *sp. nov.*

***Index Fungorum* number:** IF552036; ***Facesoffungi* number:** FoF 01995; Figs. 46 and 47

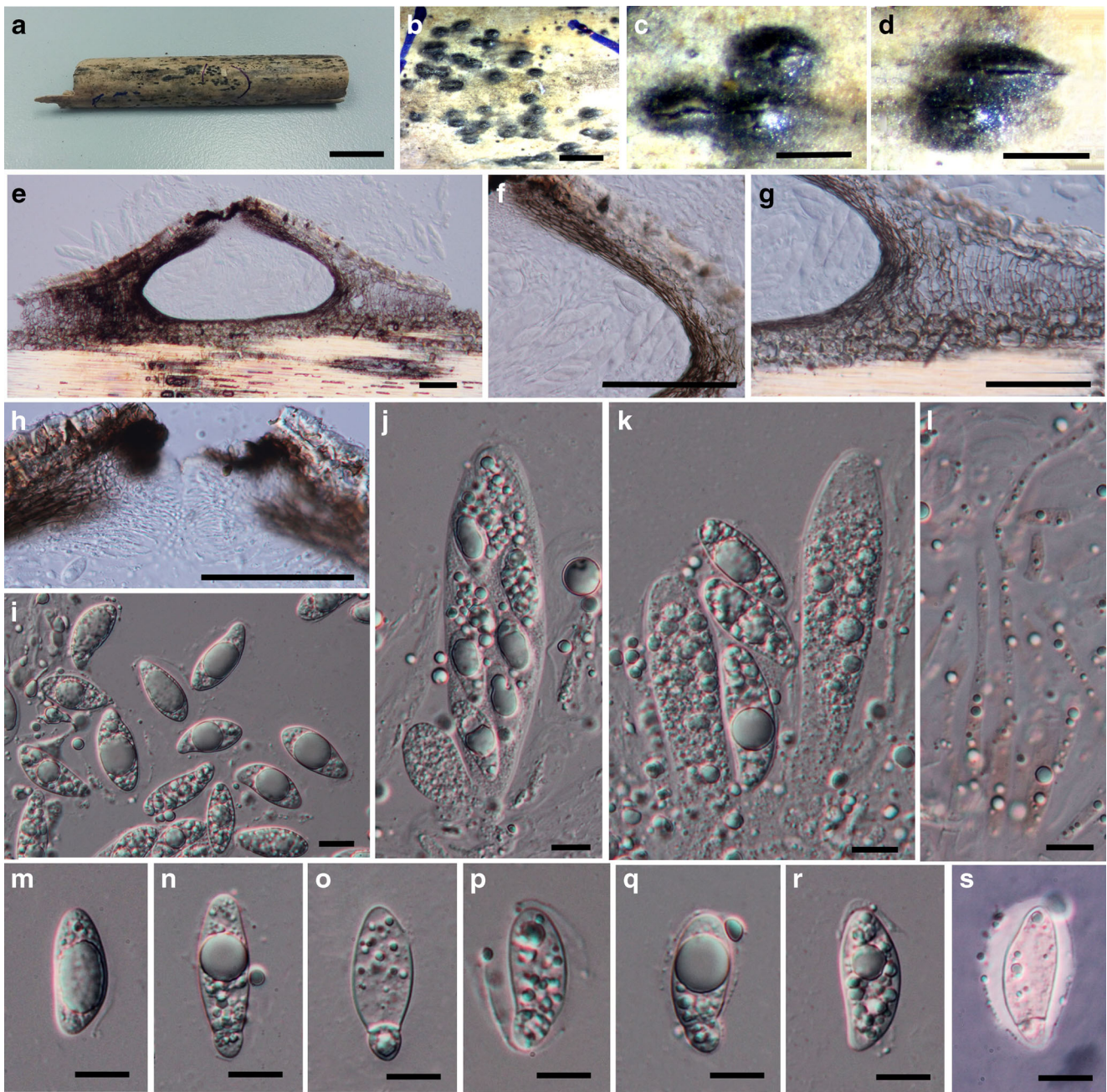
***Entomology:*** In reference to the country where this species was collected.

***Holotype:*** MFLU 15–1226

***Saprobic*** on dead bamboo culms, forming black, lenticular spots with ascostromata breaking through raised cracks at the black centre. **Sexual morph:** *Ascostromata* 450–990  $\mu\text{m}$  long, 300–550  $\mu\text{m}$  wide, 150–200  $\mu\text{m}$  high, solitary to gregarious, immersed, fusiform to ellipsoid, black, with long axis broken at the top. *Ascomata* perithecal, 145–160  $\mu\text{m}$  high, 250–280  $\mu\text{m}$  diam., immersed within the ascostromata, subglobose, dark brown, membranous, with a single perithecium, with a distinctive centrally located, periphysate, 50–75  $\mu\text{m}$  wide, ostiole, covered by a black clypeus. *Peridium* thin to thick, comprising several strata, composed of dark brown to brown to hyaline cells of compressed *textura angularis*, thinner at the base, laterally 5–10  $\mu\text{m}$  thick, with upper wall around ostiole, laterally 15–25  $\mu\text{m}$  thick; surrounded by 150–250  $\mu\text{m}$  thick stromatic tissue composed of large, brown cells of *textura prismatica* to *textura angularis*. *Hamathecium* composed of dense, long, 2–5  $\mu\text{m}$  broad, septate, unbranched, paraphyses. *Asci* 80–100  $\times$  16–20  $\mu\text{m}$  ( $\bar{x} = 94.4 \times 19.8 \mu\text{m}$ ,  $n = 20$ ), 8-spored, unitunicate,



**Fig. 45** Asexual morph of *Arthrinium rasikravindrae* (MFLUCC 15–0203). **a, b** Colony on PDA producing conidia mass. **c** Conidia mass. **d–f** Conidia. Scale bars: a–c = 500  $\mu\text{m}$ , d–f = 10  $\mu\text{m}$

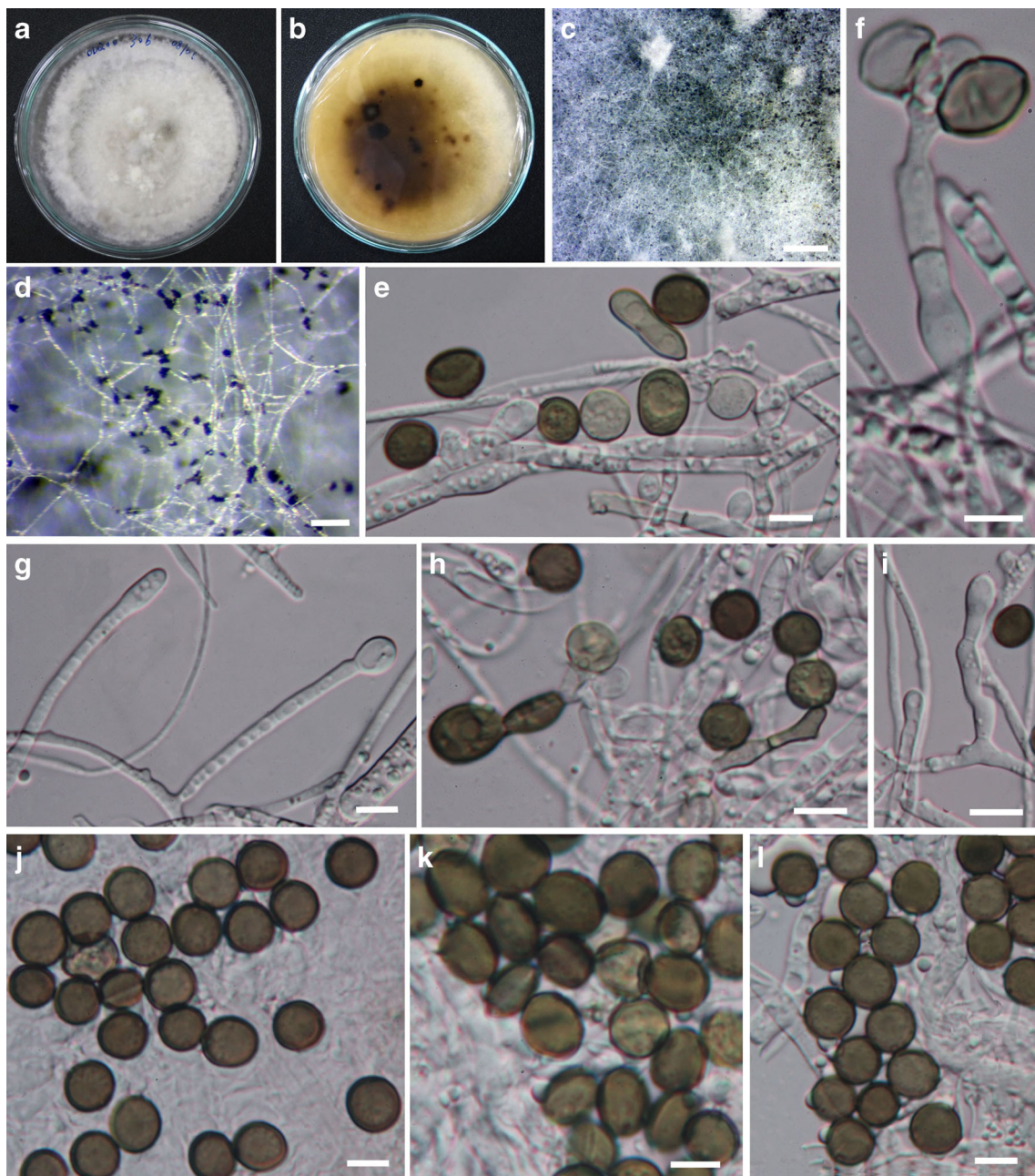


**Fig. 46** *Arthrinium thailanicum* (MFLU 15–1226, holotype). **a** Type specimen, a bamboo culm. **b–d** Appearance of ascostromata on host. **e** Vertical section of ascostroma. **f** Peridium of ascoma. **g** Peridium of ascoma surrounded by stromatic tissue. **h** Ostiole opening with

periphyses. **j, k** Asci. **l** Paraphyses in congo red. **i, m** Ascospores. **s** Ascospore with sheath in indian ink. Scale bars: a=2 mm, b=1 mm, c, d=500  $\mu\text{m}$ , e–h=50  $\mu\text{m}$ , i–s=10  $\mu\text{m}$

broadly cylindrical to long clavate, with a short pedicel. *Ascospores* 22–30  $\times$  8–12.5  $\mu\text{m}$  ( $\bar{x}$  = 25.3  $\times$  10.3  $\mu\text{m}$ ,  $n=20$ ), 2–3-seriate, 1-septate, elliptical, with a large, curved, upper cell and small lower cell, with narrowly rounded ends, hyaline, smooth-walled, with many guttules, surrounded by 3–5  $\mu\text{m}$  wide gelatinous sheath attached, at both ends. **Asexual morph:** Fertile on PDA, after 1 month. *Conidiophore mother cells* 5.5–11  $\times$  3–4.5  $\mu\text{m}$  ( $\bar{x}$  = 7.6  $\times$  3.8  $\mu\text{m}$ ,  $n=20$ ), ampulliform to cylindrical,

smooth-walled. *Conidiophores* reduced to conidiogenous cells. *Conidiogenous cells* 11.5–39  $\times$  2–3.5  $\mu\text{m}$  ( $\bar{x}$  = 26.7  $\times$  2.6  $\mu\text{m}$ ,  $n=20$ ), basauxic, polyblastic, sympodial, cylindrical, discrete, sometimes branched, smooth-walled. *Conidia* 5–9  $\times$  5–8  $\mu\text{m}$  ( $\bar{x}$  = 7.1  $\times$  6.2  $\mu\text{m}$ ,  $n=20$ ), lenticular, globose to subglobose, occasionally elongated to ellipsoidal, dark brown, smooth-walled, with a truncate basal scar, occasionally with a longitudinal, hyaline, thin, germ-slit.



**Fig. 47** Asexual morph of *Arthrinium thailandicum* (MFLUCC 15–0202, ex-type). **a, b** Colony on PDA. **c, d** Mycelium with conidia. **e, f, i, j** Conidiogenous cells. **h** Elongated conidia. **k** Conidia from above view. **l** Conidia from side view. Scale bars: **c, d** = 1 mm, **e–l** = 5  $\mu$ m

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes developing from the upper cell. Colonies fast growing on PDA, reaching 90 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, cottony, circular, with irregular edge, white from above, brown to dark brown in the centre from below. Mycelium superficial to immersed in media, with branched, septate, smooth hyphae.

**Material examined:** THAILAND, Krabi, Krabi Noi, Wat Tham Sua Temple, 8°07'34"N 98°55'28"E, limestone outcrop, on dead culms of bamboo, 4 October

2014, Kevin D. Hyde DDQ00308 (MFLU 15–1226, **holotype**); *Ibid.* (KUN HKAS88733, **isotype**), ex-living cultures, MFLUCC 15–0202, KUMCC; *Ibid.*, DDQ00304 (MFLU 15–1224); *Ibid.* (KUN HKAS88731), living culture, MFLUCC 15–0199, KUMCC.

**Notes:** *Arthrinium thailandicum* is phylogenetically close to *A. malaysianum* Crous (Fig. 42) introduced by Crous and Groenewald (2013) with description of only the asexual morph. *Arthrinium malaysianum* has short, unbranched

conidiogenous cells ( $4\text{--}7 \times 3\text{--}5 \mu\text{m}$ ), whereas *A. thailandicum* has longer, branched conidiogenous cells ( $11.5\text{--}39 \times 2\text{--}3.5 \mu\text{m}$ ). Their conidia overlap in size but the average is larger in *A. thailandicum* (Crous and Groenewald 2013).

***Arthrinium yunnanum*** D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum number*: IF552037; *Facesoffungi number*: FoF 01996; Figs. 48 and 49

*Entomology*: Refers to the province where the species was first collected.

*Holotype*: MFLU 15–0382

*Saprobic* on decaying bamboo culms, forming black, lenticular spots on the host surface, with ascostromata breaking through raised cracks at the black centre. **Sexual morph**: *Ascostromata* 0.7–1.5 mm long, 0.35–0.55 mm wide, 0.2–0.3 mm high, solitary, or occasionally two gregarious, immersed, subepidermal, fusiform to ellipsoid, black, with a long slit opening at the top. *Ascomata* perithecial, 150–300  $\mu\text{m}$  diam., 155–200  $\mu\text{m}$  high in section, 2–3 immersed under ascostromata, subglobose, light brown to reddish brown, membranous, distinctive ostiole at the centre, with a black clypeus. *Peridium* laterally 15–25  $\mu\text{m}$  thick, composed of brown and small light brown to reddish brown to hyaline, elongated cells of *textura angularis*; surrounded by 100–300  $\mu\text{m}$  stromatic tissues, composed of small cells of *textura angularis*. *Hamathecium* composed of dense, 2–7  $\mu\text{m}$  broad, septate, unbranched, paraphyses. *Asci* 85–100  $\times$  30–35  $\mu\text{m}$  ( $\bar{x}$  = 92.3  $\times$  33.1  $\mu\text{m}$ ,  $n=20$ ), 8-spored, unitunicate, broad cylindrical to subglobose, with narrow apex. *Ascospores* 28–37  $\times$  9–11  $\mu\text{m}$  ( $\bar{x}$  = 32.7  $\times$  10.2  $\mu\text{m}$ ,  $n=20$ ), 2–3-seriate, 1-septate, elliptical, with a large, curved, upper cell and small lower cell, with narrowly rounded ends, hyaline, smooth-walled, with many guttules, surrounded by a 5–20  $\mu\text{m}$  wide, gelatinous sheath. **Asexual morph**: Coelomycetous, fertile on PDA, after 1 month. *Conidiomata* 300–350  $\mu\text{m}$  diam., 350–400  $\mu\text{m}$  high, stromatal, sporodochial, solitary to gregarious, superficial on media, surrounded by hyaline colonies, globose to subglobose, coriaceous. *Conidiophore mother cells* 6.5–9  $\times$  2.5–4  $\mu\text{m}$ , cylindrical, smooth-walled, arising from the stroma. *Conidiogenous cells* 16.5–50  $\times$  2–4  $\mu\text{m}$  ( $\bar{x}$  = 37.8  $\times$  2.9  $\mu\text{m}$ ,  $n=20$ ), basauxic, cylindrical, discrete, smooth-walled. *Conidia* 17.5–26.5  $\times$  15.5–25  $\mu\text{m}$  ( $\bar{x}$  = 22.3  $\times$  9.2  $\mu\text{m}$ ,  $n=20$ ), lenticular, globose to obovoid, dark brown, smooth-walled, with a truncate basal scar.

*Culture characters*: Ascospores germinating on PDA within 24 h and germ tubes developing from the upper part. Colonies fast growing on PDA, reaching 90 mm in 2 weeks at 26 °C, under 12 h light/12 h dark, cottony, circular, with irregular edge, white from above, brown to dark brown in centre from below. Mycelium superficial to immersed in media, with branched, septate, smooth hyphae.

*Material examined*: CHINA, Yunnan, Kunming, Kunming Institute of Botany, Chinese Academy of Science, on dead culms of *Phyllostachys nigra*, 7 July 2014, Dong-Qin Dai DDQ00279 (MFLU 15–0382, **holotype**); *Ibid.* (KUN HKAS83867 **isotype**), ex-type living cultures, MFLUCC 15–0002, CBS 139958.

*Notes*: *Arthrinium yunnanum* is phylogenetically close to *A. hyphopodii* D.Q. Dai & K.D. Hyde (Fig. 42). However, *A. yunnanum* differs from the latter in having longer conidiogenous cells (16.5–50  $\times$  2–4  $\mu\text{m}$  vs. 4–6  $\times$  2–3.5  $\mu\text{m}$ ) and larger conidia (17.5–26.5  $\times$  15.5–25  $\mu\text{m}$  vs. 5–10  $\times$  4–8  $\mu\text{m}$ ) (Senanayake et al. 2015). *Arthrinium yunnanum* is morphological similar to *A. setosa* Samuels and *A. bambusae* (Turconi) Sivan. However, *A. setosa* has smaller asci (65–85  $\times$  20–28  $\mu\text{m}$  vs. 85–100  $\times$  30–35  $\mu\text{m}$ ) whereas *A. bambusae* has smaller conidia (15–19  $\times$  15–17  $\mu\text{m}$  vs. 17.5–26.5  $\times$  15.5–25  $\mu\text{m}$ ) (Samuels et al. 1981; Sivanesan 1983). Moreover, the blast results using the LSU sequence in NCBI's GenBank indicates that *A. yunnanum* is a distinct species.

***Diatrypaceae*** Nitschke [as '*Diatrypeae*'], Verh. naturh. Ver. preuss. Rheinl. 26: 73 (1869)

Kirk et al. (2008) included 13 genera in this family. However, Lumbsch and Huhndorf (2010) accepted ten genera, while Maharachchikumbura et al. (2015) list 15 genera in *Diatrypaceae*. The phylogenetic analyses (Acero et al. 2004; Trouillas et al. 2011, 2015) showed some common genera are polyphyletic, such as *Diatrype*, *Diatrypella*, *Cryptosphaeria*, *Eutypa* and *Eutypella*. This family is characterized by perithecial ascomata, immersed in a well developed eu- or pseudostroma, often long-necked with unitunicate, long stalked asci, often with a truncate apex, and allantoid, hyaline to pale brown ascospores (Carmarán et al. 2006; Vasilyeva and Stephenson 2004, 2006). Members of this family are common worldwide, typically occurring on a broad range of dead or declining woody angiosperms (Acero et al. 2004).

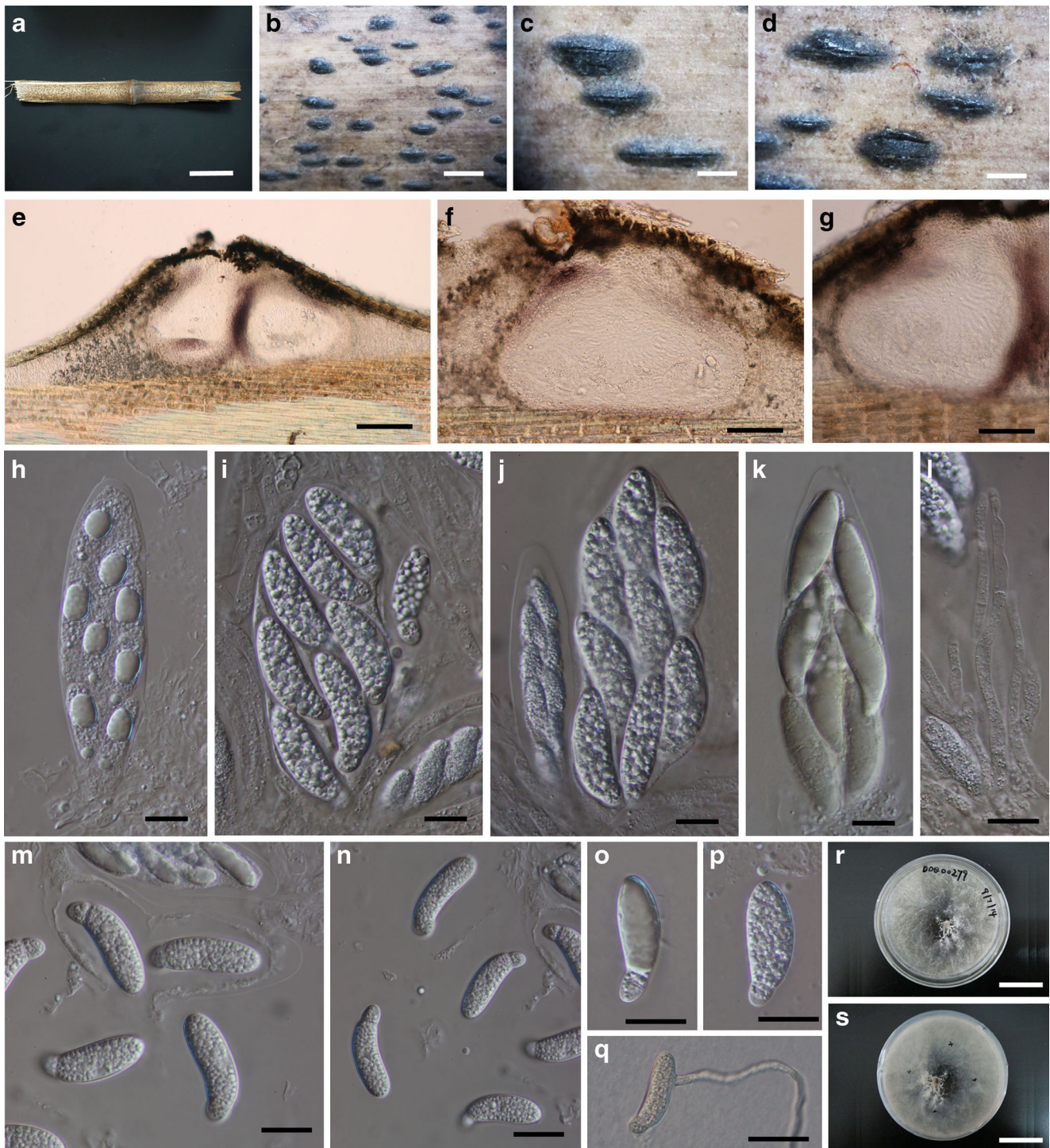
***Eutypa*** Tul. & C. Tul., Select. fung. carpol. (Paris) 2: 52 (1863)

The genus *Eutypa* was established by Tulasne and Tulasne (1863), and typified by *Eutypa lata* (Pers.) Tul. & C. Tul.. Kirk et al. (2008) accepted 32 species in this genus, however, more than 200 epithets are listed in Index Fungorum (2016). *Eutypa* species have a wide host range, and can cause various diseases on plants (DeScenzo et al. 1999; Rolshausen et al. 2006). Asexual morphs of this genus are linked to *Libertella* and *Cytosporina* (Kirk et al. 2008).

***Eutypa linearis*** Rehm, Anns mycol. 5(6): 523 (1907)

*Facesoffungi number*: FoF 01997; Figs. 50 and 51

*Saprobic* on dead bamboo culms, forming black parallel to naviculate ascostromata on the host, breaking through the host tissue as a slit when mature, exposing black ostioles. **Sexual morph**: *Ascostromata* 1.5–6 mm long, 300–650  $\mu\text{m}$  wide,

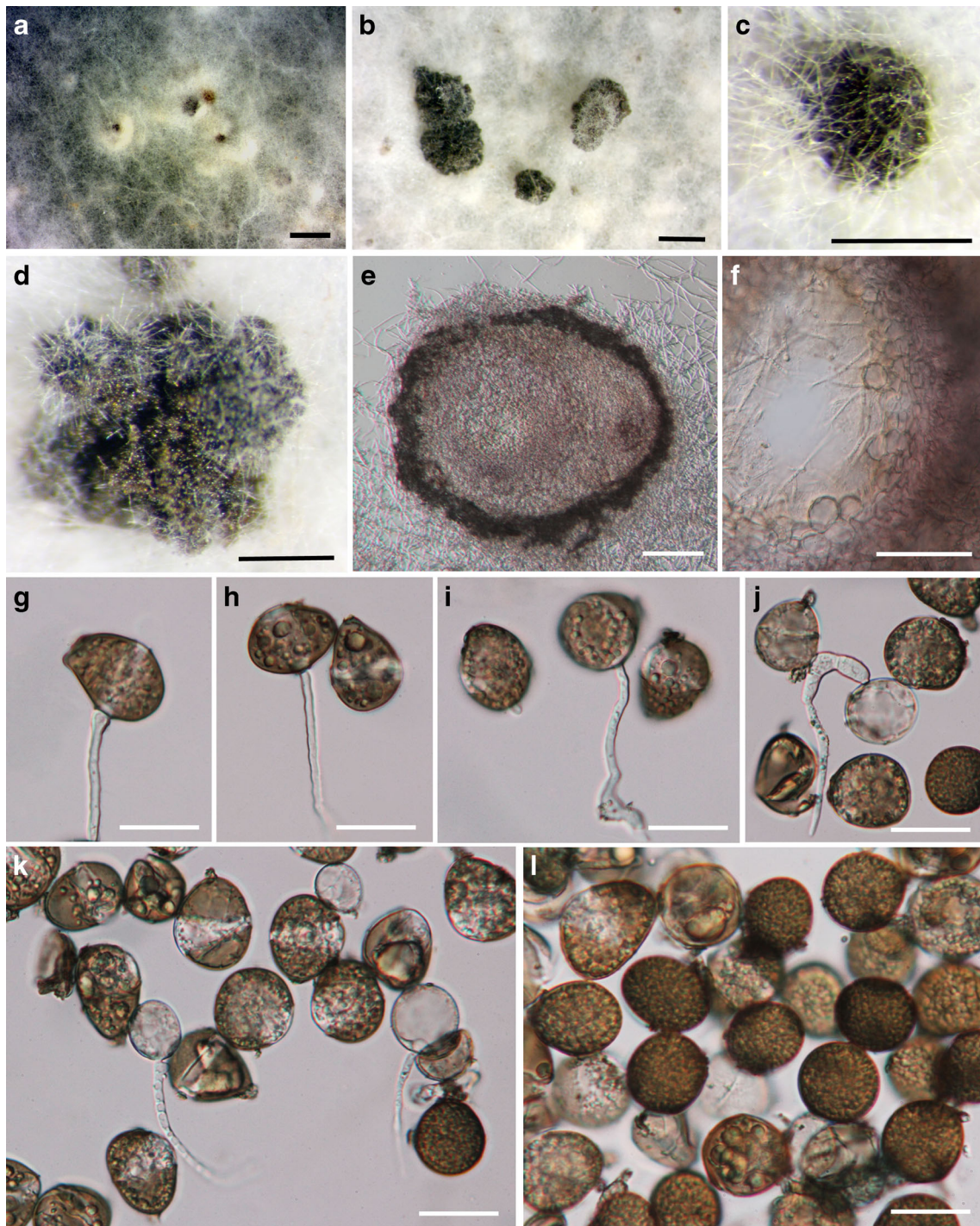


**Fig. 48** *Arthrinium yunnanum* (MFLU 1515–0382, **holotype**). **a** Bamboo specimen. **b–d** Black ascostromata on bamboo host. **e** Vertical section of ascostroma. **f, g** Vertical sections of ascoma. **h** Immature ascus. **i–j** Mature asci. **k** Ascus stained by Meltzer's. **l** Paraphyses. **m–p**

Ascospores. **q** Germinating ascospore. **r, s** Cultures on PDA. Scale bars: **a** = 5 cm, **b–d** = 500  $\mu$ m, **e** = 200  $\mu$ m, **f** = 150  $\mu$ m, **g** = 100  $\mu$ m, **h–l** = 10  $\mu$ m, **m–q** = 5  $\mu$ m, **r, s** = 25 cm

and 300–500  $\mu$ m high, immersed under the host, long fusiform, irregular ellipsoidal. Upper cells of stromata near the perithecial ostiole are black, thick-walled. Stromatic tissue is yellow, compact. *Ascomata* perithecial, 100–150  $\mu$ m diam., 250–400  $\mu$ m high, immersed in stromata, obpyriform, with a

central, ostiolate, opening, 95–105  $\times$  45–65  $\mu$ m, erumpent over stromata, with periphyses. *Peridium* 6–10  $\mu$ m thick, composed of brown to hyaline, elongate, cells of *textura angularis*, surrounded by yellow stromatic tissue. *Hamathecium* with ca. 1  $\mu$ m wide, unbranched, paraphyses.



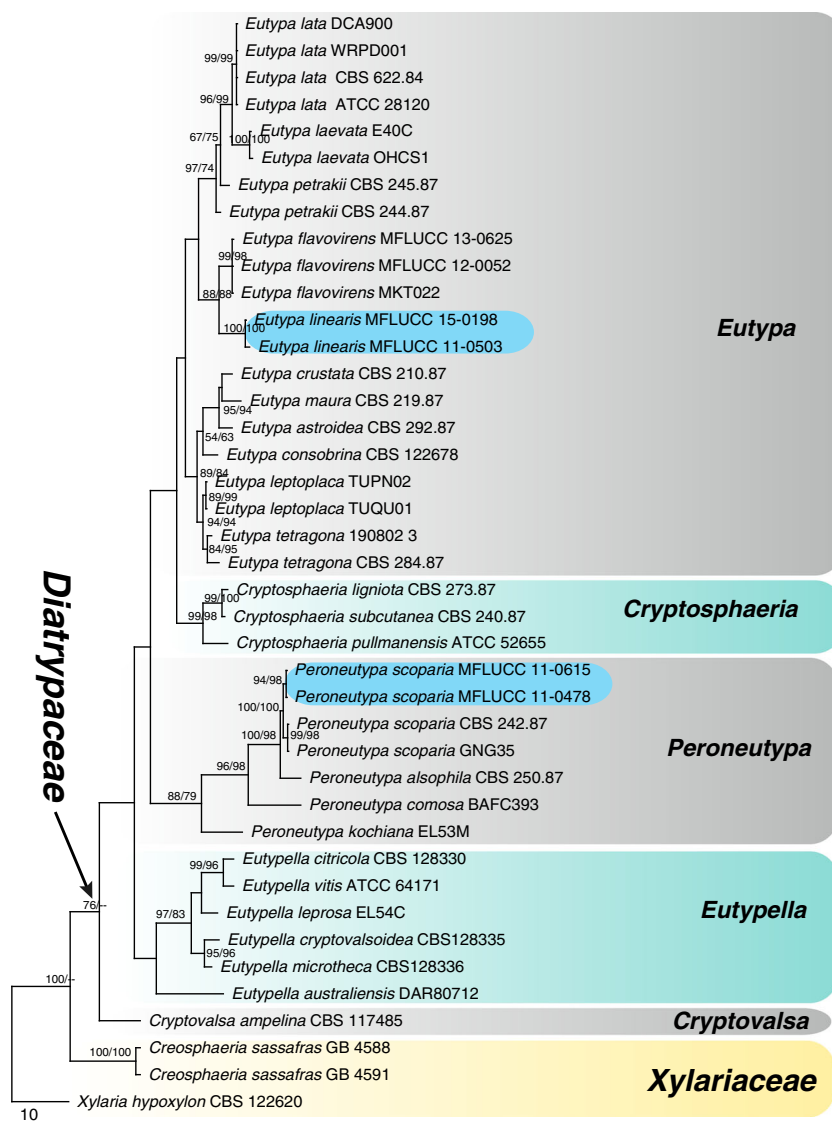
**Fig. 49** Asexual morph of *Arthrinium yunnanum* (MFLUCC 15–0002, ex-type). **a** Colony on PDA producing conidiomata. **b–d** Conidia mass. **e** Vertical section of stromata. **f** stromata wall. **g–j** Conidiogenous

cells. **k, l** Conidia. Scale bars: **a, b** = 1 mm, **c, d** = 500  $\mu\text{m}$ , **e** = 100  $\mu\text{m}$ , **f** = 50  $\mu\text{m}$ , **g–l** = 20  $\mu\text{m}$

*Asci* 50–90  $\times$  5–7.5  $\mu\text{m}$  ( $\bar{x}$  = 70.1  $\times$  6.2  $\mu\text{m}$ ,  $n$  = 20), 8-spored, unitunicate, cylindrical, with a long and thin peridial, apex truncate, with a small, J-, apical ring. *Ascospores* 5–7  $\times$  1–2  $\mu\text{m}$  ( $\bar{x}$  = 6.1  $\times$  1.4  $\mu\text{m}$ ,  $n$  = 20), 2–3-seriate, allantoid, aseptate, rounded at both ends, hyaline, smooth-walled. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from both ends. Colonies growing fast on PDA, reaching 9 mm in 2 weeks at 28  $^{\circ}\text{C}$ , under 12 h light/12 h dark, circular, edge irregular, hyaline from above and below. Mycelium superficial to immersed in media, branched, septate, smooth.

**Fig. 50** One of three equally most parsimonious trees resulting from 1000 bootstrap replicates obtained from ITS sequence data. Maximum likelihood phylogenetic tree ( $\ln L = -2688.878680$ ) was generated by RAxML (GTR+G model). MP/ML values ( $>50\%$ ) are given at the nodes, and branches. The original isolate numbers are noted after the species names. The tree is rooted to *Xylaria hypoxylon* (CBS122620). Newly generated sequences are highlighted with a blue background



**Material examined:** THAILAND, Phang Nga, Thap Put District, Tham Thong Lang, Doi Nang Hong, on dead culms of bamboo, 6 December 2014, Kevin D. Hyde DDQ00303 (MFLU 15–1223, **reference specimen designated here**); *Ibid.* (KUN HKAS88730, **duplicate of reference specimen**), living cultures, MFLUCC 15–0198, MUCL 55892; Chiang Rai, Mae Fah Luang University, on dead culms of bamboo, 19 July 2011, Dong-Qin Dai DDQ00082 (MFLU 15–1186); *Ibid.* (KUN HKAS88696), living culture, MFLUCC 11–0503.

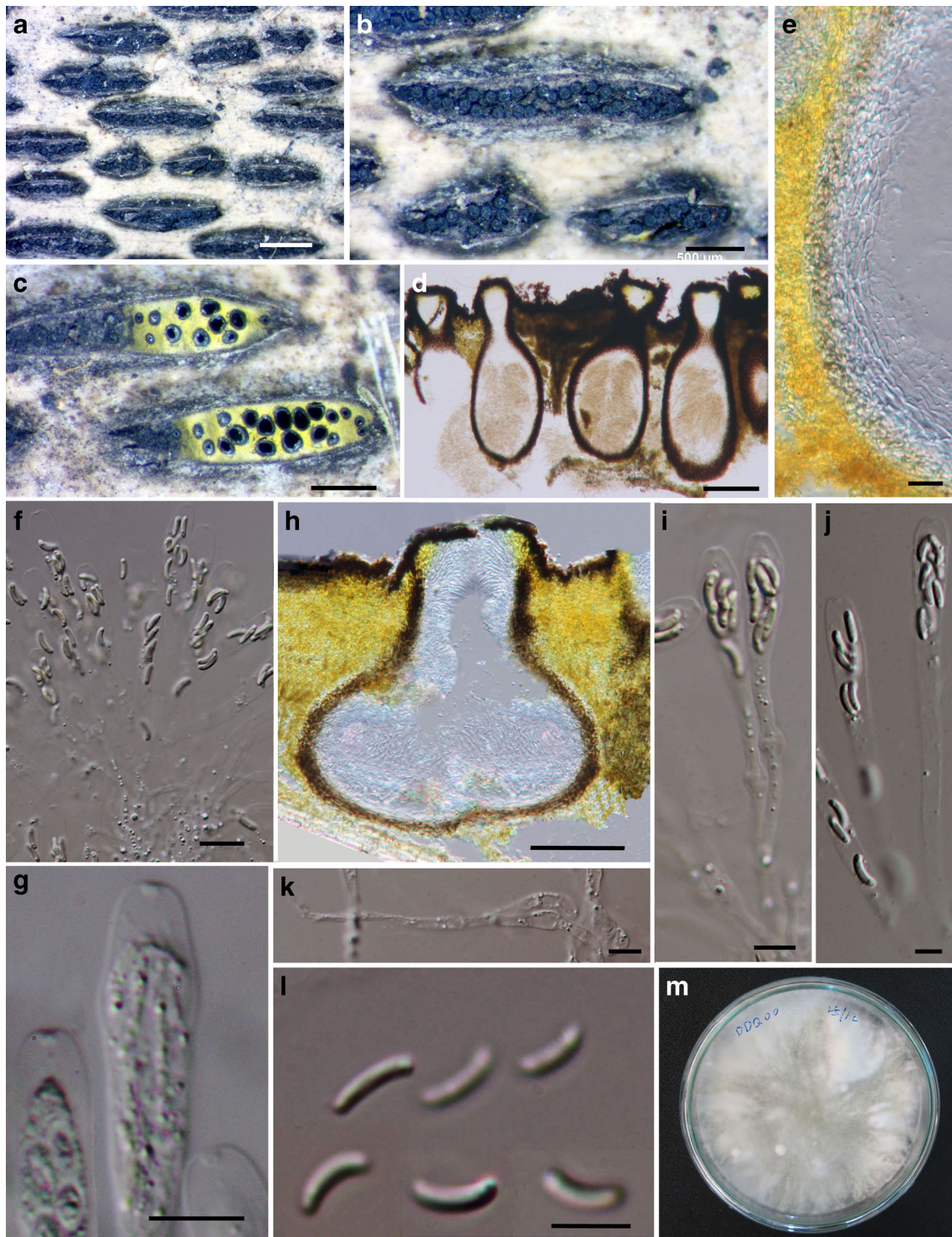
**Notes:** *Eutypa linearis* was described by Rehm (1907), based on a specimen collected in Brazil. This species occurred on culms of *Arundinariae* (bamboo) (Rehm 1907). Eriksson and Yue (1998) provided a line drawing of stromata of *Eutypa linearis*. Our new collections have the same size of ascospores with that in Rehm (1907), but larger stromata (1.5–6 mm vs. 0.5–3 mm in length). The phylogenetic tree (Fig. 50) shows *Eutypa linearis* is

closest to *E. flavovirens*. However, *E. flavovirens* has smaller (less than 1.5 mm in length) round stromata, and occurs on twigs (Senanayake et al. 2015). *Eutypa linearis* is a common species occurring on bamboo (Eriksson and Yue 1998), however, no molecular data is available in GenBank, and thus reference specimen is designated here.

***Peroneutypa*** Berl., Icon. fung. (Abellini) 3(3–4): 80 (1902)

*Peroneutypa* is typified by *Peroneutypa bellula* (Desm.) Berl. and was introduced by Berlese (1902). Berlese (1968) amended this genus to accommodate species with small, clavate asci and perithecia with long necks. It is listed as a synonym of *Eutypella* (Nitschke) Sacc in Index Fungorum (2016). However, Carmarán et al. (2006) accepted *Peroneutypa* with seven species, and there are approximately 50 epithets listed in Index Fungorum (2016).





**Fig. 51** *Eutypa linearis* (MFLU 15–1223, reference specimen). **a, b** Appearance of ascostromata on bamboo host. **c** Horizontal slice through ascostromata. **d, h** Vertical sections of ascostromata in stroma. **e**

Peridium of ascoma. **g** Apical ring of asci. **f, i, j** Asci. **k** Paraphyses. **l** Ascospores. **m** Culture on PDA. Scale bars: **a** = 1 mm, **b, c** = 500  $\mu$ m, **d** = 50  $\mu$ m, **e, f** = 10  $\mu$ m, **h** = 100  $\mu$ m, **g, k–l** = 5  $\mu$ m

*Peroneutypa scoparia* (Schwein.) Carmarán & A.I. Romero, in Carmarán et al., Fungal Diversity Res. Ser. 23: 84 (2006)

*Facesoffungi* number: FoF 01998; Fig. 52

Synonym: See Index Fungorum.

*Saprobic* on dead bamboo culms, necks breaking through host epidermis, after maturity. **Sexual morph:** *Ascomata* perithecial, 350–450  $\mu$ m diam., 150–170  $\mu$ m high, solitary to 2–3

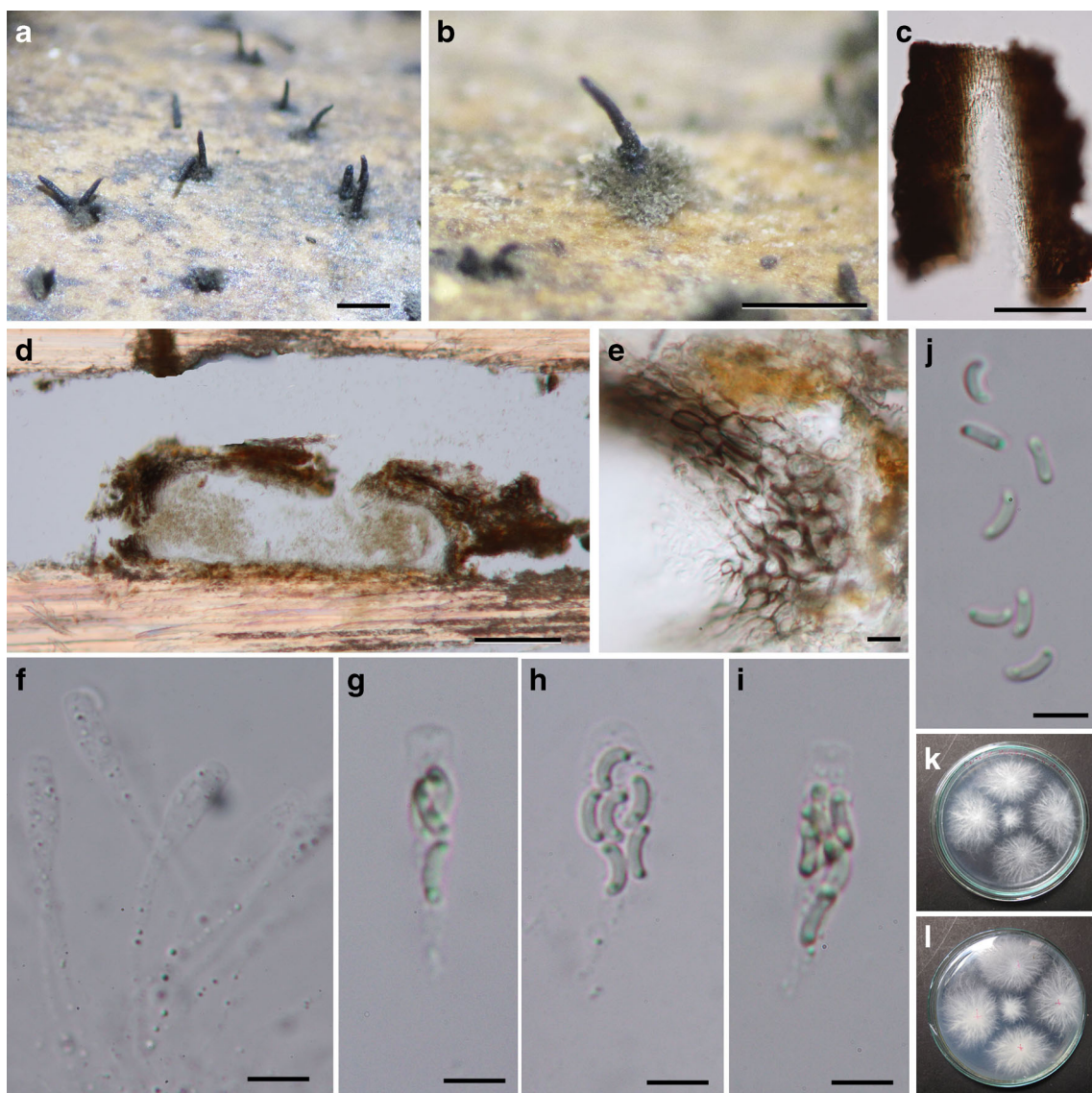
gregarious, immersed in the cortex, globose to subglobose, dark brown to black, coriaceous. *Necks of ascomata* 1.5–3 mm long, 100–150  $\mu\text{m}$  wide, black, straight to curved, with periphyses. *Peridium* 15–25  $\mu\text{m}$  thick, comprising hyaline to dark brown cells of *textura angularis*. *Hamathecium* comprising only asci, paraphyses not observed. *Asci* 21–40.5  $\times$  3.5–5.5  $\mu\text{m}$  ( $\bar{x}$  = 32.6.3  $\times$  4.9  $\mu\text{m}$ ,  $n$  = 20), 8–6-spored, unitunicate, clavate to cylindro-clavate, apically rounded to truncate, with a 15–20  $\mu\text{m}$  long pedicel, J-. *Ascospores* 3.5–4.5  $\times$  1–1.5  $\mu\text{m}$  ( $\bar{x}$  = 4.1  $\times$  1.4  $\mu\text{m}$ ,  $n$  = 30), 2–3-seriate to irregular arranged, allantoid, hyaline, rounded at both ends, smooth-walled. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from both ends. Colonies growing fast on PDA, reaching 45 mm in 1 week at 28  $^{\circ}\text{C}$ ,

under 12 h light/12 h dark, circular, edge irregular, white from above and below. Mycelium superficial to immersed in media, branched, septate, smooth.

**Material examined:** THAILAND, Chiang Rai, Mae Fah Luang University, on dead culms of bamboo, 16 July 2011, Dong-Qin Dai DDQ00074 (MFLU 15–1183, **reference specimen designated here**); *Ibid.* (KUN HKAS88693, **duplicate of reference specimen**), living culture, MFLUCC 11–0478; Jiew Santonkok, on dead culms of bamboo, 11 August 2011, Dong-Qin Dai DDQ00101 (MFLU 16–0881), living culture, MFLUCC 11–0615.

**Notes:** *Peroneutypa scoparia* was combined by Carmaran et al. (2006) based on *Sphaeria scoparia* Schwein. Pongcharoen et al. (2006) isolated this species from leaves of *Garcinia dulcis* (Roxb.) Kurz, collected in Songkhla



**Fig. 52** *Peroneutypa scoparia* (MFLU 15–1183, **reference specimen**). **a, b** Perithecia immersed in host tissue, appearing as long, black necks. **c** Section of perithecial neck. **d** Vertical section of perithecium. **e** Peridium.

**f** Immature asci. **g–i** Asci. **j** Ascospores. **k, l** Culture on PDA. Scale bars: **a, b** = 3 mm, **c** = 50  $\mu\text{m}$ , **d** = 100  $\mu\text{m}$ , **e** = 10  $\mu\text{m}$ , **f–j** = 5  $\mu\text{m}$

Province, Thailand. We recollected same species on bamboo in northern Thailand and designated it as a reference specimen.

**Xylariaceae** Tul. & C. Tul. [as ‘Xylariei’], *Select. fung. carpol.* (Paris) 2: 3 (1863)

*Xylariaceae* is one of the best-known pyrenomycete families in *Ascomycetes* and is distributed throughout the world (Rogers and Ju 1998, 2012; Ju and Rogers 1999; Peláez et al. 2008; Rogers et al. 2008; Chareprasert et al. 2012). Members of *Xylariaceae* are mainly wood inhabitants and prevalent in tropical and subtropical regions (Chareprasert et al. 2012). The xylariaceous fungi generally have well-developed stromata (Ju and Rogers 1999). Ju and Rogers (1996) accepted 38 genera and Whalley et al. (1996) recognized 40. Lumbsch and Huhndorf (2007) listed 76 genera. Subsequently Maharachchikumbura et al. (2015) included 86 genera in this family. The phylogenetic studies of *Xylariaceae* based on ribosomal DNA or other gene sequences have been carried out by many authors (Peláez et al. 2008; Tang et al. 2009; Hsieh et al. 2010; Jaklitsch and Voglmayr 2012; Stadler et al. 2013; Daranagama et al. 2015).

***Anthostomella*** Sacc., *Atti Soc. Veneto-Trent. Sci. Nat., Padova, Sér. 4 4*: 84 (1875)

*Anthostomella*, introduced by Saccardo (1875), is a species-rich genus, with more than 300 putative species (Francis 1975; Lu and Hyde 2000). Hyde (1996) described 14 species (of which 10 were new) on palms. Lu and Hyde (2000) provided a monograph of *Anthostomella*, with keys and illustrations of 86 species. Daranagama et al. (2015) revised the genus *Anthostomella* based on analysis of combined ITS, RPB2,  $\beta$ -tubulin and LSU and introduced three new species. *Anthostomella* is presently regarded as a polyphyletic genus (Stadler et al. 2013; Daranagama et al. 2015). However, very few species of the genus have been sequenced. *Anthostomella* has virgariella-like and nodulisporium-like asexual morphs (Daranagama et al. 2015).

***Anthostomella pseudobambusicola*** D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum number*: IF552038; *Facesoffungi number*: FoF 01999; Fig. 53

*Entomology*: Refers to the morphologically being similar to *Anthostomella bambusicola*.

*Holotype*: MFLU 16–0255

*Saprobic* on dead bamboo culms, visible as conical, dark, raised, 0.9–1.3 mm diam., 0.5 mm high, solitary to gregarious pseudostromata with a central blackened ostiole. **Sexual morph**: *Ascomata* 550–650  $\mu\text{m}$  diam., 400–450  $\mu\text{m}$  high, immersed, globose to subglobose, dark brown, coriaceous, centrally ostiolate, with a black, 60–100  $\mu\text{m}$  thick, pseudoclypeus. *Peridium* laterally 10–15  $\mu\text{m}$  thick, comprising several layers of compressed, brown to hyaline cells of *textura angularis*. *Hamathecium* of dense, long, 2–2.5  $\mu\text{m}$  wide, septate, paraphyses, intermixed with asci. *Asci* 115–

155  $\times$  5–7  $\mu\text{m}$  ( $\bar{x}$  = 136.1  $\times$  6.2  $\mu\text{m}$ ,  $n=20$ ), 4–8-spored, unitunicate, cylindrical, with a 50–70  $\mu\text{m}$  long furcate pedicel, with a 0.5–1 high  $\times$  1–1.5 diam. J+, wedge-shaped, subapical ring. *Ascospores* 8.5–11.5  $\times$  4–5  $\mu\text{m}$  ( $\bar{x}$  = 10.1  $\times$  4.3  $\mu\text{m}$ ,  $n=20$ ), uniseriate, ellipsoid, slightly pointed at the ends, aseptate, dark brown, guttulate, smooth-walled, with a straight germ-slit, extending over the full length longitudinally.

**Asexual morph**: Undetermined.

*Culture characters*: Ascospores germinating on PDA within 48 h with germ tubes produced from the central part of the spore. Colonies growing slowly on PDA, reaching 4 mm in 4 weeks at 28 °C, under 12 h light/12 h dark, effuse, velvety to hairy, white from above, pale yellowish brown from the below. Mycelium superficial and immersed, branched, septate, smooth, hyaline, irregular.

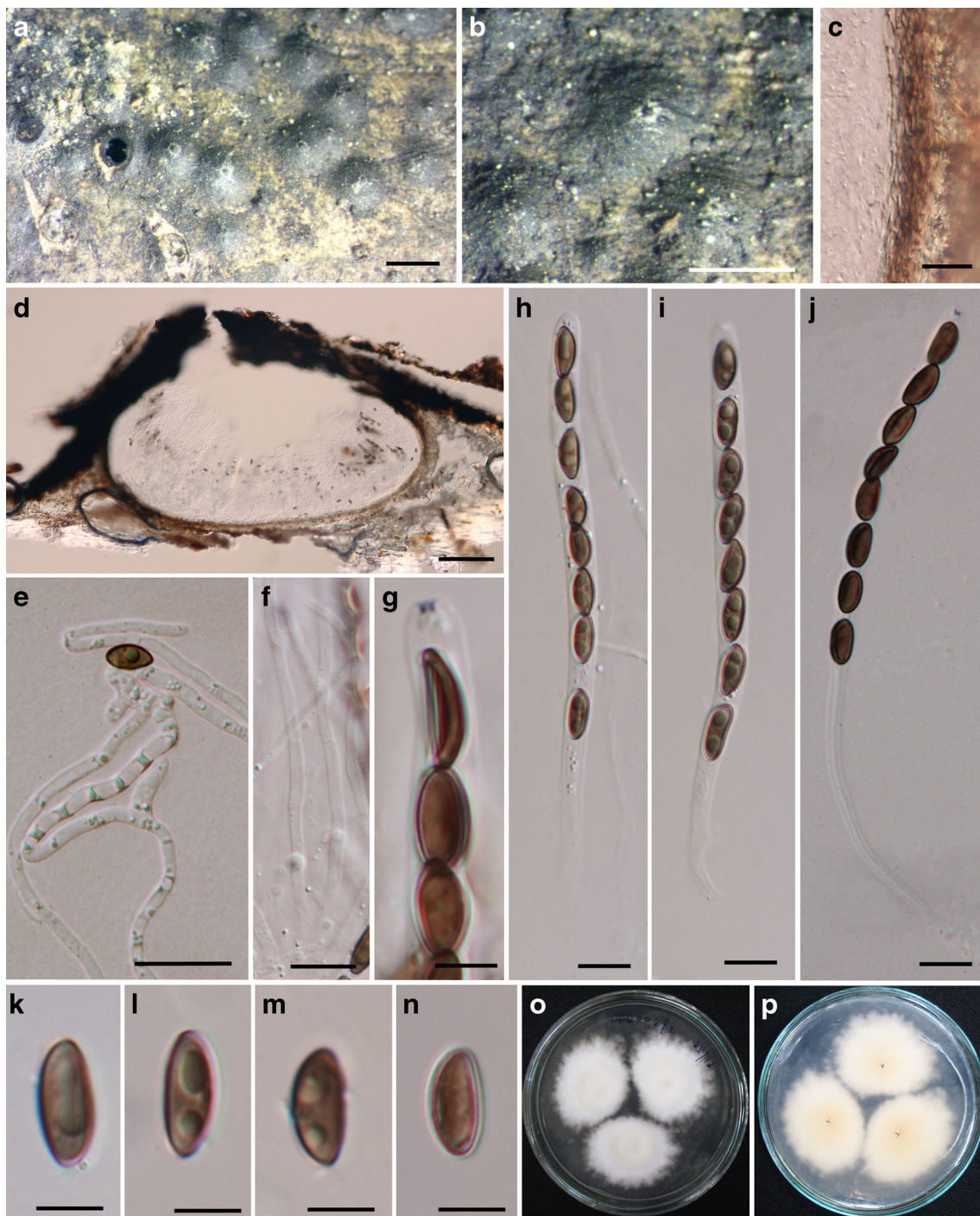
*Material examined*: THAILAND, Doi Nang Hong, Tham Thong Lang, Thap Put District, Phang-Nga, on dead culms of bamboo, 6 December 2014, Kevin D. Hyde DDQ00292 (MFLU 16–0255, **holotype**); *Ibid.* (KUN HKAS92503, **isotype**), living cultures, MFLUCC 15–0192, KUMCC.

*Notes*: *Anthostomella pseudobambusicola* resembles *A. bambusicola* Hohn. in having immersed ascomata, raising the host tissue, visible as conical structures, with a central blackened ostiole, and black clypeus (Lu and Hyde 2000). However, *Anthostomella pseudobambusicola* has smaller asci (115–155  $\times$  5–7  $\mu\text{m}$  vs. 155–175  $\times$  11.5–14  $\mu\text{m}$ ) and ascospores (8.5–11.5  $\times$  4–5  $\mu\text{m}$  vs. 16.5–24  $\times$  7.5–10  $\mu\text{m}$ ) (Lu and Hyde 2000). Lu and Hyde (2000) re-examined the holotype of *A. bambusicola* and noted its short-pedicellate asci lacking a visible apical apparatus. *Anthostomella pseudobambusicola* has long-pedicellate asci (50–70  $\mu\text{m}$  long), with a J+, wedge-shaped, subapical ring. In the phylogenetic tree (Fig. 54), the new taxon groups with *Anthostomella eucalyptorum* Crous & M.J. Wingf. and *Anthostomella proteae* S.J. Lee & Crous. However, the morphological characters are not similar to these species (Lee and Crous 2003; Crous et al. 2006a).

***Astrocystis*** Berk. & Broome, *J. Linn. Soc., Bot.* 14(no. 74): 123 (1873) [1875]

The genus *Astrocystis*, typified by *A. mirabilis* Berk. & Broome (Berkeley and Broome 1874), is usually found on monocotyledons, and is a typical member of *Xylariaceae* (Smith and Hyde 2001). This genus is characterized by uni-, to multi-peritheciate pseudoascostromata which develop beneath the host epidermis and become erumpent through the host tissue to become stellate and appear superficial (Ju and Rogers 1990; Læssøe and Spooner 1993; Smith and Hyde 2001). To date, 21 species are listed in *Index Fungorum* (2016). The asexual morph of *Astrocystis* is allied to the genus *Acanthodochium* (Smith and Hyde 2001).

***Astrocystis mirabilis*** Berk. & Broome, *J. Linn. Soc., Bot.* 14(no. 74): 123 (1873) [1875]



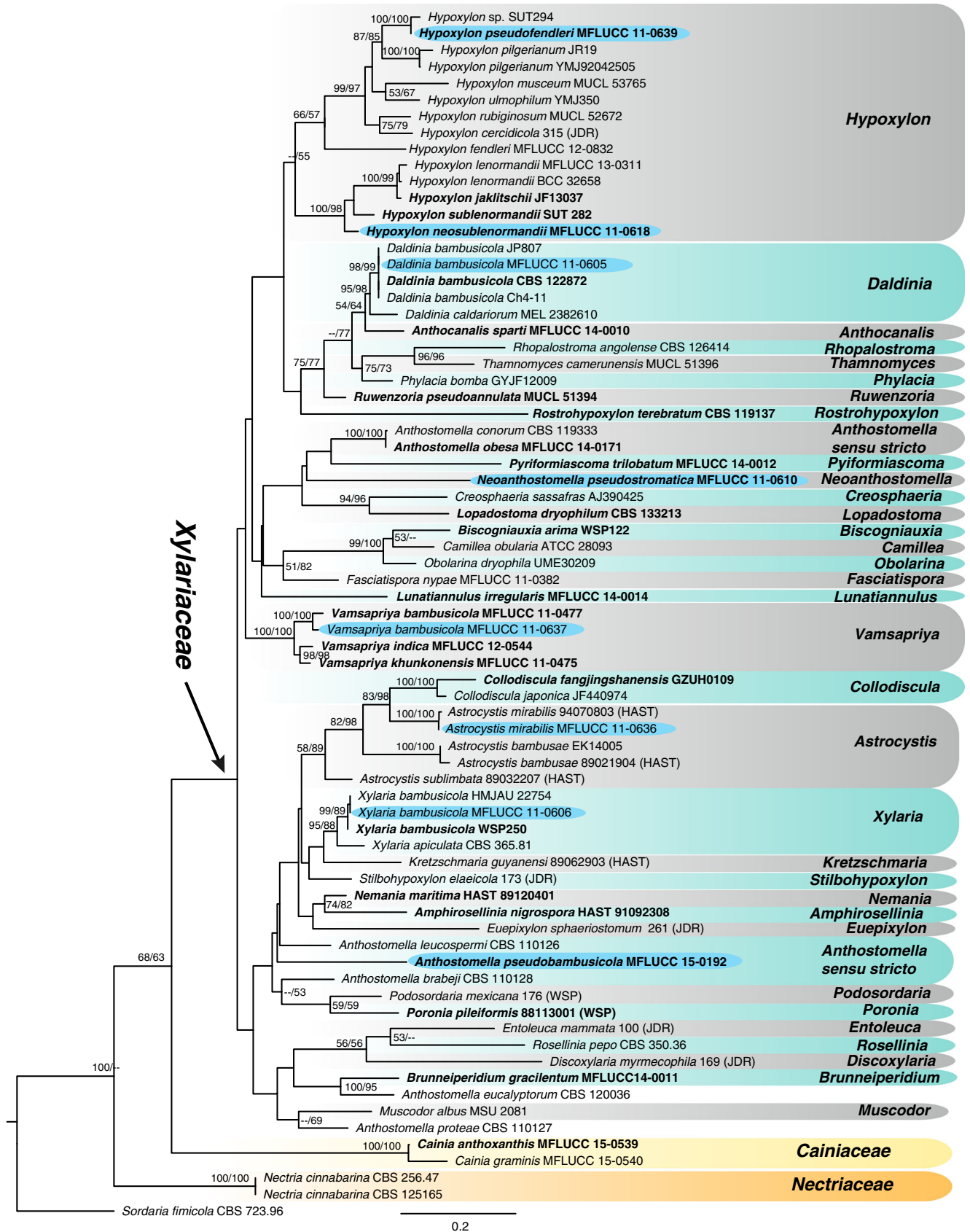
**Fig. 53** *Anthostomella pseudobambusicola* (MFLU 16–0255, holotype). **a, b** Appearance of pseudostromata on bamboo host. **c** Peridium of ascoma. **d** Vertical section of pseudostroma. **e** Germinating

ascospore. **f** Paraphyses. **g** Apical ring of asci stained by Melzer's reagent. **h–j** Asci. **k–n** Ascospores. **o, p** Culture on PDA. Scale bars: **a, b** = 1 mm, **c** = 10  $\mu\text{m}$ , **d** = 100  $\mu\text{m}$ , **e** = 50  $\mu\text{m}$ , **f, h–j** = 10  $\mu\text{m}$ , **g, k–n** = 5  $\mu\text{m}$

*Facesoffungi* number: FoF 02000; Fig. 55

*Saprobic* on dead bamboo culms, forming black, erumpent, star-like flanged pseudostromata, visible by the ruptured host tissue. **Sexual morph:** *Ascomata* perithecial, 350–600  $\mu\text{m}$  diam., 300–400  $\mu\text{m}$  high, solitary to gregarious, immersed becoming erumpent raised and superficial,

**Fig. 54** Maximum likelihood phylogenetic tree (lnL = -3555.676955) generated by RAxML (GTR + G model) based on analysis of ITS sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes, and branches. The original isolate numbers are noted after the species names. The tree is rooted to *Sordaria fimicola* (CBS 723.96). Ex-type strains are in **bold**. Newly generated sequences are highlighted with a blue background



subglobose to hemispherical, black, carbonaceous, with flattened top, surrounding by star-like flanged pseudostromata tissue, each pseudostroma containing a single perithecium with a centrally papillate ostiole, 35  $\mu\text{m}$  diam, lacking periphyses. *Peridium* 30.5–85  $\mu\text{m}$  thick, outer stratum comprising black, fragile, carbonaceous tissue; inner stratum composed of brown to hyaline cells of *textura angularis*. *Hamathecium* comprising dense, long, 2–5.5  $\mu\text{m}$  wide, septate, paraphyses intermixed with asci. *Asci* 95–120  $\times$  8.5–12  $\mu\text{m}$  ( $\bar{x}$  = 107.9  $\times$  11.2  $\mu\text{m}$ ,  $n$  = 20), 8-spored, unitunicate, cylindrical, with a short pedicel, with a 3–4.5 high  $\times$  1.5–2.5 diam. J+, apical ring. *Ascospores* 10–12  $\times$  4.5–6  $\mu\text{m}$  ( $\bar{x}$  = 11.8  $\times$  5.4  $\mu\text{m}$ ,  $n$  = 20), uniseriate, ellipsoid, slightly pointed at the ends, aseptate, brown to dark brown, guttulate, smooth-walled, with germ-slit slightly less than full length, with a 1.5–2.5  $\mu\text{m}$  thick gelatinous sheath. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from both ends of the spore. Colonies growing fast on PDA, reaching 90 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, cottony circular, edge irregular, white from above, brown in centre from below. Mycelium superficial to immersed in media, with septate, branched, smooth hyphae.

**Material examined:** THAILAND, Uttaradit Province, on dead culms of bamboo, 28 October 2011, Dong-Qin Dai DDQ00220 (MFLU 15–1199, **references specimen designated here**); *Ibid.* (KUN HKAS88708, **duplicate of reference specimen**), living culture, MFLUCC 11–0636.

**Notes:** *Astrocystis mirabilis*, the type of *Astrocystis* was originally described from bamboo by Berkeley and Broome (1874). Ju and Rogers (1990) re-examined type materials of two similar species, *A. mirabilis* and *A. bambusae*, with descriptions, line drawings of the ascostromata and ascospores, but assigned *A. mirabilis* to *Rosellinia*. Smith and Hyde (2001) restored this species in *Astrocystis*. Bahl et al. (2005) included *A. mirabilis* (AY862572) in a phylogenetic analyses based on ITS sequence data, and indicated its close relationship with *A. bambusae* (AY862573), however, AY862572 is designated as *Rosellinia mirabilis* in GenBank, and although Bahl et al. (2005) discussed the morphological differences and recommended separating *Astrocystis* and *Rosellinia*, no morphological study was undertaken. In this study we collected a specimen which has same morphology as *A. mirabilis* and when compared with the holotype of *A. mirabilis*, re-examined by Ju and Rogers (1990), this new collection (MFLU 15–1199) is similar. It has the same length, but smaller (4.5–6  $\mu\text{m}$  vs. 6–7.5  $\mu\text{m}$ ) width ascospores, and has shorter asci (95–120  $\mu\text{m}$  vs. 110–190  $\mu\text{m}$  long). This new collection is morphologically and phylogenetically (Fig. 53) identified as *A. mirabilis*, and is

designated as the reference specimen. The blast result shows that ITS gene AY862572 is same with that of MFLU 15–1199. Thus its name should be changed to *A. mirabilis* in GenBank.

*Astrocystis mirabilis* is morphologically similar to *A. bambusae* (Ju and Rogers 1990; Læssøe and Spooner 1993). The main distinctions between them are: the apex of the perithecia of *A. mirabilis* is flattened, however that of *A. bambusae* is usually rounded (Ju and Rogers 1990); and the ascospores of *A. mirabilis* are broad relative to their length and with the length usually less than 12  $\mu\text{m}$ ; however those of *A. bambusae* are narrow, with the length occasionally up to 16  $\mu\text{m}$  (Ju and Rogers 1990).

***Daldinia*** Ces. & De Not., Comm. Soc. crittog. Ital. 1(4): 197 (1863)

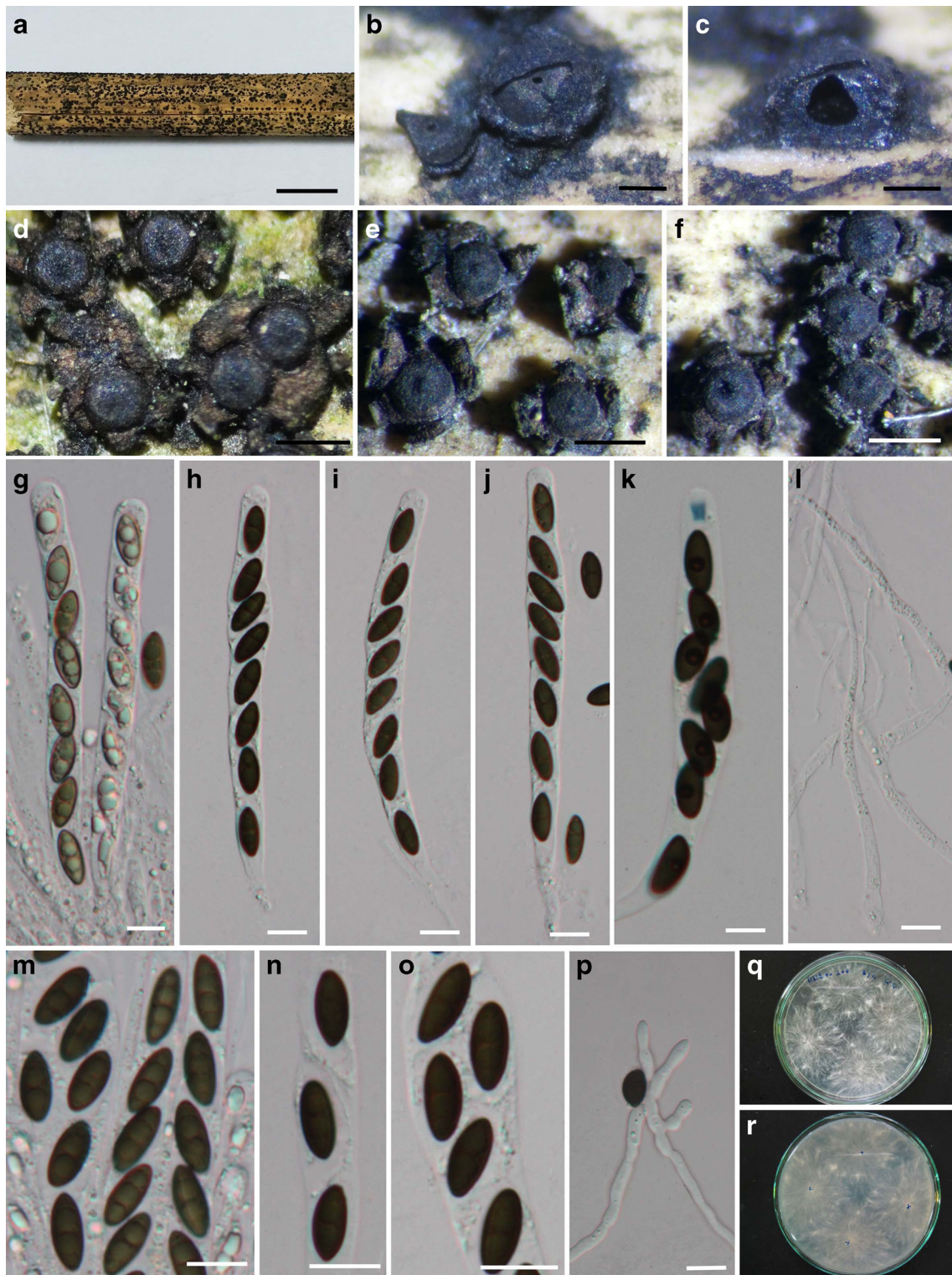
*Daldinia* was introduced by Cesati and De Notaris (1863) to honor Agostino Daldini (1817–1895), and comprises 19 species including the type species *D. concentrica* (Bolton) Ces. & De Not. (Ju et al. 1997; Stadler et al. 2014). *Daldinia* species usually grow on decaying wood (Ju et al. 1997, 1999) and are occasionally found as endophytes (Pažoutová et al. 2013). To date, 98 records of *Daldinia* species are listed in Index Fungorum (2016). Petrini and Müller (1986) established five species found on monocotyledons. The asexual morph of *Daldinia* can be linked to *Nodulisporium* Preuss (Wijayawardene et al. 2012; Stadler et al. 2014).

***Daldinia bambusicola*** Y.M. Ju, et al., Mycotaxon 61: 253. 1997

***Facesoffungi* number:** FoF 02001; Fig. 56

**Saprobic** on decaying bamboo culms, forming large, globose, fruiting bodies. *Stromata* 12–20 mm diam. solitary to gregarious, superficial, globose, sessile, smooth, surface dark vinaceous to black, with inconspicuous perithecial outlines; tissue between perithecia black brown to light brown, and below the perithecial layer composed of alternating zones; darker zones are dark vinaceous brown, pithy to woody, 0.1–0.2 mm thick; lighter zones are white to greyish brown, pithy to woody, persistent, 0.5–2 mm thick, and above the perithecial layer composed of black to dark brown tissue. *Perithecia* 250–450  $\mu\text{m}$  diam., 630–715  $\mu\text{m}$  high, immersed, arranged in a row under the outermost wall of stromata, obovoid to tubular, lower part round and dark brown. *Ostioles* slightly papillate. *Hamathecium* composed of long, 3.5–5.5  $\mu\text{m}$  broad, hyaline, septate, hyphae-like paraphyses. *Asci* 120–160  $\times$  5–7.5  $\mu\text{m}$  ( $\bar{x}$  = 146.1  $\times$  6.6  $\mu\text{m}$ ,  $n$  = 20), 6–8-spored, unitunicate, cylindrical, with thin, 70–100  $\mu\text{m}$  long pedicel and a round, 1–1.5 high  $\times$  2.5–3 diam. J+, apical ring, truncate at the apex. *Ascospores* 7–8  $\times$  3.5–4.5  $\mu\text{m}$  ( $\bar{x}$  = 7.3  $\times$  3.8  $\mu\text{m}$ ,  $n$  = 20), uniseriate, aseptate, curved, elliptical, hyaline, becoming dark brown, with narrowly rounded ends, smooth-walled.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from both ends



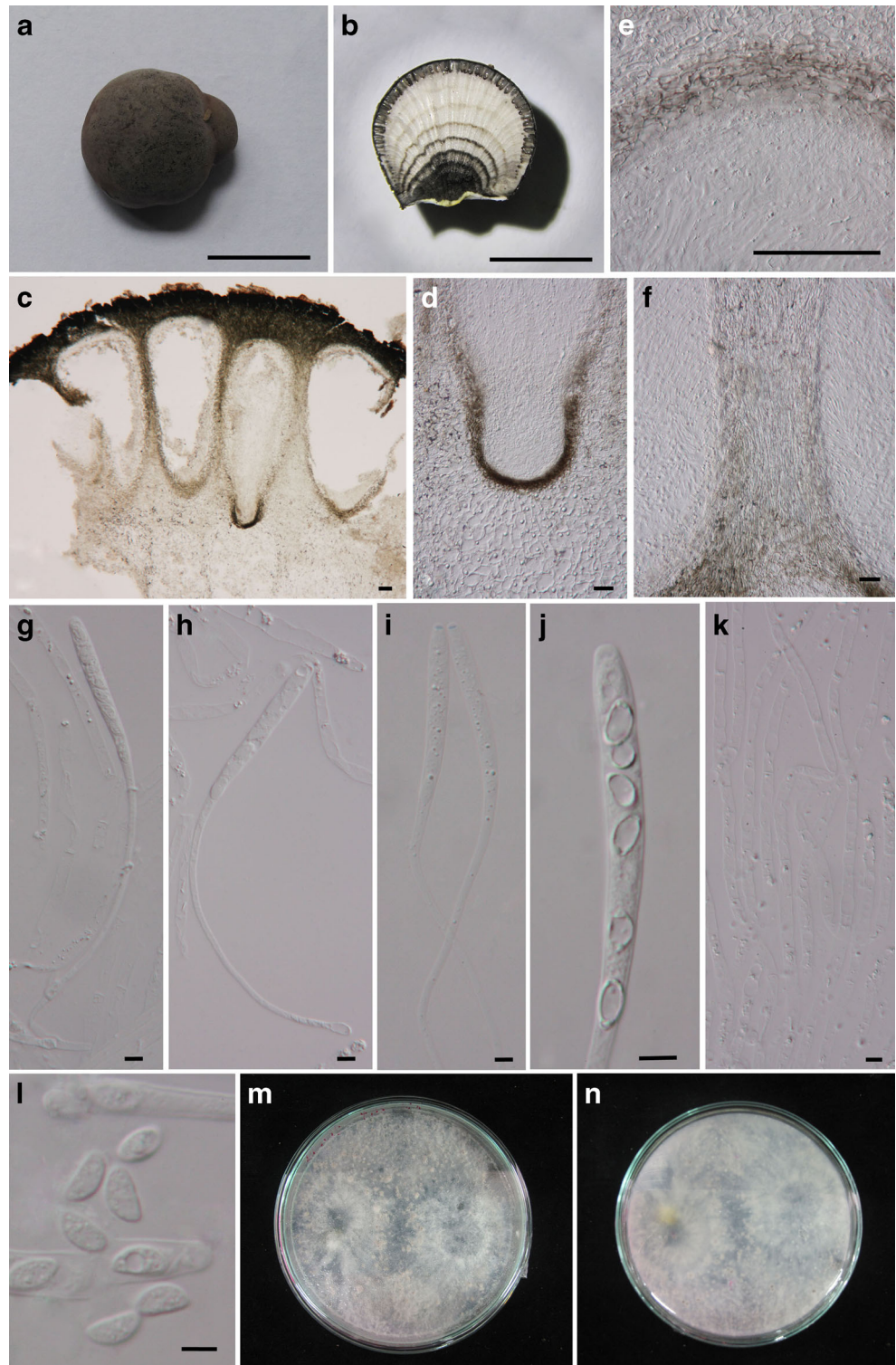
**Fig. 55** *Astrocystis mirabilis* (MFLU 15–1199, reference specimen). **a** Bamboo culm with fungal fruiting bodies. **b–f** Black perithecia. **j–k** Asci. **l** Paraphyses. **m–o** Ascospores (**n, o**: showing sheath of ascospores). **p**

Germinating ascospore. **q, r** Cultures on PDA. Scale bars: **a** = 5 mm, **b, c** = 200  $\mu\text{m}$ , **d, f** = 500  $\mu\text{m}$ , **g–p** = 10  $\mu\text{m}$

of spore. Colonies growing fast on PDA, reaching 90 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, cottony circular, edge irregular, white from above,

brown in centre from below. Mycelium superficial to immersed in media, with septate, branched, smooth hyphae.

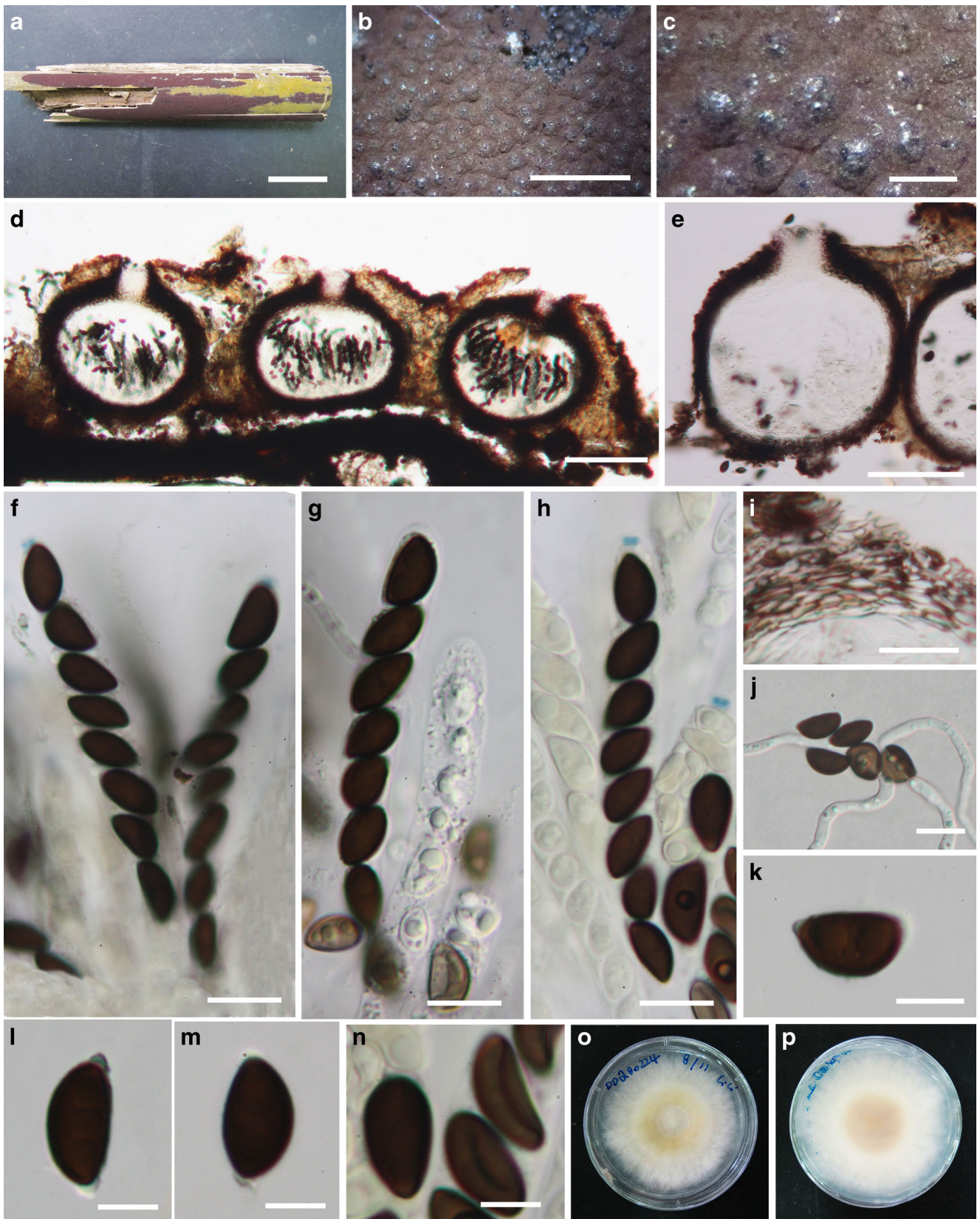
**Fig. 56** *Daldinia bambusicola* (MFLU 15–1187). **a, b** Stromata. **c** Vertical section of ascostroma. **d** Lower part of perithecium. **e** The tissue between perithecia. **f–j** Asci with long pedicel. **k** Paraphyses. **l** Ascospores. **m, n** Culture on PDA. Scale bars: a, b = 10 mm, c–f = 50  $\mu\text{m}$ , g–l = 5  $\mu\text{m}$



*Material examined:* THAILAND, Chiang Rai, Mae Fah Luang University, on dead culms of bamboo, 1 August 2011, Dong-Qin Dai DDQ00085 (MFLU 15–1187); *ibid.* (KUN HKAS88697), living cultures, MFLUCC 11–0605, CBS 139987.

**Fig. 57** *Hypoxylon pseudedefleri* (MFLU 15–1200, holotype). **a** Appearance of stromata on bamboo host. **b, c** Purplish-brown stromata. **d** Vertical section of stroma. **e** Vertical section of perithecia. **f–h** Asci containing eight ascospores, with J+, apical rings. **i** Wall of perithecium. **j** Germinating ascospores. **k–n** Dark brown ascospores. **o, p** Cultures on PDA. Scale bars: a = 3 mm, b = 1 mm, c = 200  $\mu\text{m}$ , d = 100  $\mu\text{m}$ , e = 50  $\mu\text{m}$ , f–j = 10  $\mu\text{m}$ , k–n = 5  $\mu\text{m}$





Notes: *Daldinia bambusicola* is specifically associated with bamboo and commonly distributed in tropical regions

with a likely record in the USA (Ju et al. 1997; Stadler et al. 2014). This species was originally collected in Chiang Mai

Province, Thailand by Ju et al. (1997). We recollected the same species in Chiang Rai Province, and it is illustrated in Fig. 56. Phylogenetic analyses (Fig. 53) show that the new collection groups well with ex-type strain (CBS 122872) of *D. bambusicola* with high bootstrap support value (100/100 % MPBP/MLBP). Ju et al. (1997) mentioned *Daldinia bambusicola* has darker, zonate internal tissue in the stromata. However, Stadler et al. (2014) re-examined the holotype (WSP 69652), and did not observe this character, probably due to the dry and broken material. The fresh collection (MFLU 15–1187) circumscribed by phylogenetic analyses (Fig. 53) has exactly the same features as described by Ju et al. (1997).

**Hypoxylon** Bull., Hist. Champ. Fr. (Paris) 1: 168 (1791)

*Hypoxylon* is presently one of the most species diverse genera in *Xylariaceae*; the species are saprobes or facultative parasites, and they are frequently encountered as endophytes (Ju and Rogers 1996; Suwannasai et al 2005; Cruz and Cortez 2015; Sir et al. 2015). *Hypoxylon* is one of the typical genera of *Xylariaceae* in having well-developed stromata, within which multiple perithecia are usually embedded, an ascus apical ring usually bluing in iodine, and dark brown, one-celled ascospores with a germ slit (Ju and Rogers 1996; Hsieh et al. 2005). Kuhnert et al. (2014, 2015) reported several chemical compounds extracted from *Hypoxylon* species. Asexual morphs of this genus are *Nodulisporium*-like and *Triplicaria* (Ju and Rogers 1996; Wijayawardene et al. 2012; Kuhnert et al. 2015; Senanayake et al. 2015).

***Hypoxylon pseudefendleri*** D.Q. Dai & K.D. Hyde, *sp. nov.*

*Index Fungorum number*: IF552039; *Facesoffungi number*: FoF 02002; Fig. 57

*Entomology*: Refers to the morphologically being similar to *Hypoxylon fendleri*.

*Holotype*: MFLU 15–1200

*Saprobic* on dead bamboo culms, forming large purplish-brown stromata with raised necks of perithecia, appearing as black spots on stromata. **Sexual morph**: *Stromata* gregarious, 5–30 cm long, 3–10 cm broad, 0.3–0.5 mm thick, usually covering the whole surfaces of host culm, superficial, effuse-pulvinate, flattened, purplish-brown. *Topical layer of stromata* comprising darker tissue. *Cells between perithecia* are stromatic tissue, yellowish brown, large and lighter than perithecial wall. *Perithecia* 500–850  $\mu\text{m}$  diam., 350–500  $\mu\text{m}$  high, immersed in stromata, globose to subglobose, brown to dark brown, coriaceous, with short raised necks. *Necks* 50–55  $\times$  35–45  $\mu\text{m}$  long, forming at the center of perithecia, appearing as darker spots on stromata surface when mature, without periphyses, slightly higher than the stromata surface. *Peridium* laterally 20  $\mu\text{m}$  thick, composed of dark brown and thick-walled, to hyaline and thinner walled, cells of *textura angularis*. *Hamathecium* of dense, long, 2.5–4.5  $\mu\text{m}$  wide, septate, paraphyses. *Asci* 70–90.5  $\times$  6.5–10.5  $\mu\text{m}$  ( $\bar{x}$  =

80.6  $\times$  8.6  $\mu\text{m}$ ,  $n=20$ ), 8-spored, unitunicate, cylindrical, occasionally with a short pedicel, with a 1–1.5 high  $\times$  1.5–2.5 diam. J+, apical ring. *Ascospores* 9–11.5  $\times$  4.5–6.5  $\mu\text{m}$  ( $\bar{x}$  = 10.2  $\times$  5.7  $\mu\text{m}$ ,  $n=20$ ), ellipsoid, slightly pointed at the ends, aseptate, dark brown, guttulate, smooth-walled. **Asexual morph**: Undetermined.

*Culture characters*: Ascospores germinating on PDA within 24 h and germ tubes developing from both ends of spore. Colonies fast growing on PDA, reaching 80 mm in 3 weeks at 28 °C, 12 h light/12 h dark, cottony, circular, diffuse from centre, with irregular edge, white with pale yellow in centre from above and below. Mycelium superficial to immersed in media, with branched, septate, smooth hyphae.

*Material examined*: THAILAND, Phitsanulok Province, on dead culms of bamboo, 29 October 2011, Dong-Qin Dai DDQ00224 (MFLU 15–1200, **holotype**); *ibid.* (KUN HKAS88709, **isotype**), living cultures, MFLUCC 11–0639, KUMCC.

*Notes*: *Hypoxylon pseudefendleri* is morphologically similar with *H. fendleri* in having large, purplish-brown stromata, perithecia with a central raised neck, appearing as black spots on stromata (Ju and Rogers 1996; Suwannasai et al. 2013). However, *H. pseudefendleri* has shorter (9–11.5  $\mu\text{m}$  vs. 12–13  $\mu\text{m}$  long) and wider ascospores (4.5–6.5  $\mu\text{m}$  vs. 4  $\mu\text{m}$  wide) (Cooke 1883). Moreover, the phylogenetic analyses (Fig. 53) shows they are different species. Phylogenetically, *Hypoxylon pseudefendleri* is close to *H. pilgerianum* (Fig. 53), whereas, *H. pseudefendleri* has larger stromata 5–30 cm long  $\times$  3–10 cm broad, rather than 0.2–6 cm long  $\times$  0.2–3 cm broad (Ju and Rogers 1996).

***Hypoxylon neosublenormandii*** D.Q. Dai & K.D. Hyde, *sp. nov.*

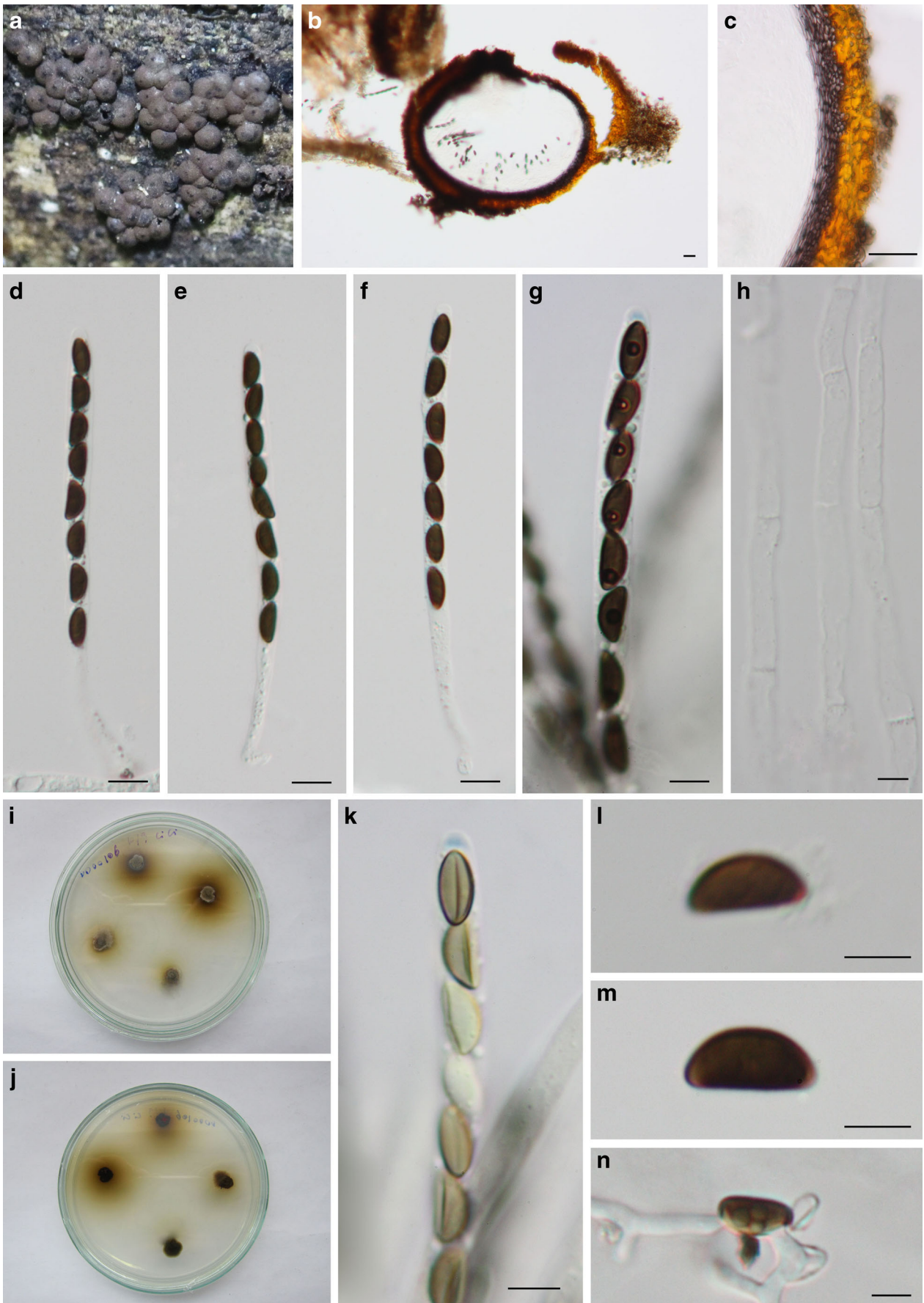
*Index Fungorum number*: IF552040; *Facesoffungi number*: FoF 02003; Fig. 58

*Entomology*: Refers to the morphologically being similar to *Hypoxylon sublenormandii*.

*Holotype*: MFLU 15–1193

*Saprobic* on dead bamboo culms. **Sexual morph**: *Stromata* 300–380  $\mu\text{m}$  diam., 250–300  $\mu\text{m}$  high, gregarious, superficial, globose to subglobose, purplish-brown, carbonaceous to coriaceous, surface smooth, containing a single perithecium. *Ostiole* papillate, forming at the center of stromata, with a black opening, without periphyses. *Peridium* composed of two strata, outer stratum 15–28  $\mu\text{m}$  thick, composed of orange coloured tissue; inner stratum 10–15  $\mu\text{m}$  thick, composed of dark brown and thick-walled cells of *textura angularis*. *Hamathecium* with long, 2.5–4  $\mu\text{m}$  wide, septate, paraphyses,

**Fig. 58** *Hypoxylon neosublenormandii* (MFLU 15–1193, **holotype**). **a** Appearance of stromata on bamboo. **b** Vertical section of stroma. **c** Peridium. **d, g** Asci. **h** Paraphyses. **i, j** Cultures on PDA. **k** Asci with a J+, apical ring. **l, m** Ascospores. **n** Germinating ascospores. Scale bars: b, c = 20  $\mu\text{m}$ , d–h = 10  $\mu\text{m}$ , i–l = 5  $\mu\text{m}$



intermixed with the asci. *Asci* 98–120 × 5–7 μm ( $\bar{x}$  = 109.4 × 6.1 μm,  $n$  = 20), 8-spored, unitunicate, cylindrical, with a short furcate pedicel, with a 1–1.5 high × 1.5–2.5 diam. J+, apical ring. *Ascospores* 8.5–10 × 4–5 μm ( $\bar{x}$  = 9.3 × 4.6 μm,  $n$  = 20), ellipsoid, slightly pointed at the ends, aseptate, dark brown, guttulate, smooth-walled, with a longitudinal germ-slit on one side. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 48 h and germ tubes produced from both ends. Colonies growing slowly on PDA, reaching 30 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, circular, irregular edge, with little mycelium, dark brown from above and below. Mycelium superficial to immersed in media, branched, septate, smooth.

**Material examined:** THAILAND, Chiang Rai, Doi Pui, on dead culms of bamboo, 1 August 2011, Dong-Qin Dai DDQ00106 (MFLU 15–1193, **holotype**); *Ibid.* (HKU HKAS88702, **isotype**), living cultures, MFLUCC 11–0618, KUMCC.

**Notes:** *Hypoxyton neosublenormandii* resembles *H. sublenormandii* Suwann. et al., a species originally observed on a bamboo substrate, in Kanchanaburi Province of Thailand by Suwannasai et al. (2005). They are morphologically similar in having superficial, globose to subglobose, purplish-brown, carbonaceous ascomata. However, *H. neosublenormandii* has wider asci (5–7 μm wide vs. 3.8–5 μm). In addition, *H. neosublenormandii* forms a separate branch from *H. sublenormandii* (SUT 282, ex-type) with high bootstrap support (100/98 % MPBP/MLBP). *Hypoxyton sublenormandii* can be morphologically compared with *H. jaklitschii* and *H. lenormandii* (Kuhnert et al. 2015). However the phylogenetic tree (Fig. 54) indicates that they are different species.

***Neoanthostomella*** D.Q. Dai & K.D. Hyde, **gen. nov.**

**Indexfungorum number:** IF552041; **Facesoffungi number:** FoF 02004

**Entomology:** Refers to the morphological similarities with *Anthostomella*.

**Saprobic** on dead bamboo culms, forming large, blackened, circular to elliptical, raised areas, visible as pustules on the host surface. **Sexual morph:** *Pseudostromata* solitary to gregarious, circular to elliptical, raised as blacked areas with host tissue. *Ascomata* 2–5 growing together in a single pseudostroma, immersed, globose to subglobose, dark brown, coriaceous, with a central, periphysate, ostiolate neck. *Peridium* comprising several layers of compressed, brown to hyaline cells of *textura angularis*. *Hamathecium* of dense, long, septate, paraphyses intermixed with asci. *Asci* 8-spored, unitunicate, cylindrical, with a short furcate pedicel, without an apical ring. *Ascospores* uni- to bi-seriate, sometimes overlapping, ellipsoid, slightly pointed at the ends, aseptate, dark brown, guttulate, smooth-walled, with mucilaginous sheath, with a straight germ slit extending over the full-length. **Asexual morph:** Undetermined.

**Type species:** *Neoanthostomella pseudostromatica* D.Q. Dai & K.D. Hyde

**Notes:** *Neoanthostomella* is characterized by large, blackened, raised pseudostromata on the host tissue, unitunicate, and cylindrical asci containing eight dark brown ascospores, surrounded by a thick mucilaginous sheath. This genus is similar to *Anthostomella* in having cylindrical asci containing eight dark brown ascospores. However, the feature of several ascomata clustering in a large pseudostroma distinguishes it from *Anthostomella* (Lu and Hyde 2000). From the phylogenetic tree in Fig. 54, *Neoanthostomella* forms a single branch within the family *Xylariaceae*, and can therefore also be phylogenetically separated from *Anthostomella*.

***Neoanthostomella pseudostromatica*** D.Q. Dai & K.D. Hyde, **sp. nov.**

**Index Fungorum number:** IF552042; **Facesoffungi number:** FoF 02005; Fig. 59

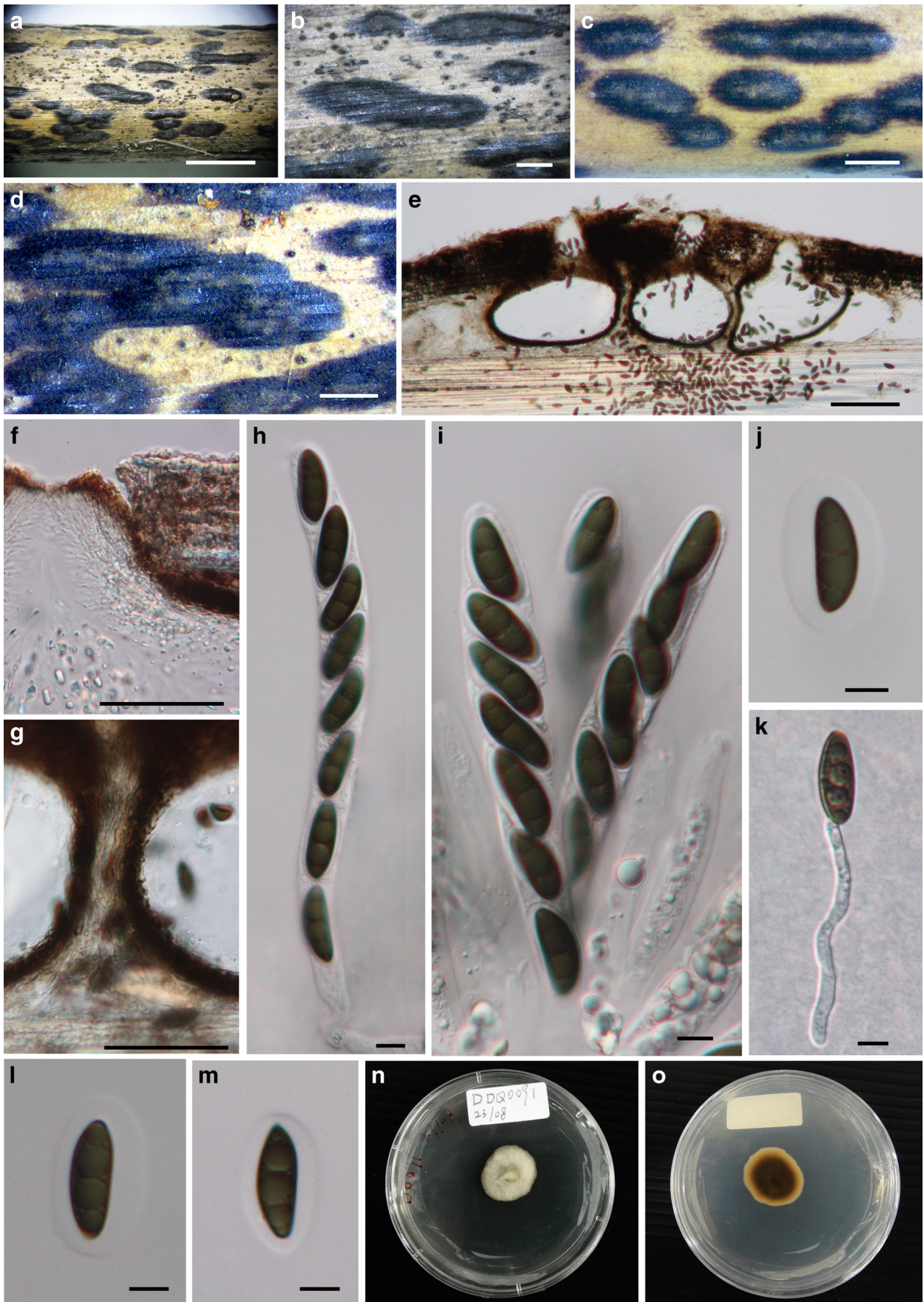
**Entomology:** Refers to the distinguishing feature of the pseudostroma.

**Holotype:** MFLU 15–1190

**Saprobic** on dead bamboo culms, forming large, blackened, circular to elliptical, raised areas, visible as pustules on the host surface. **Sexual morph:** *Pseudostromata* 1.5–4.8 mm long, 0.65–1.5 mm wide, 0.25–0.35 mm high, solitary to gregarious, circular to elliptical, black areas raising the host tissue. *Ascomata* 150–300 μm diam., 160–280 μm high, 2–5 growing together in a single pseudostroma, immersed, globose to subglobose, dark brown, coriaceous, with a central periphysate ostiolate neck, 35–45 μm diam., 55–90 μm long. *Peridium* laterally 5–10 μm thick, comprising several layers of compressed, brown to hyaline cells of *textura angularis*. *Hamathecium* of dense, long, 2.5–4 μm wide, septate, paraphyses intermixed with asci. *Asci* 75–110 × 8.5–13.5 μm ( $\bar{x}$  = 85.1 × 10.2 μm,  $n$  = 20), 8-spored, unitunicate, cylindrical, with a short furcate pedicel, lacking an apical ring. *Ascospores* 11.5–15 × 4–5.5 μm ( $\bar{x}$  = 13.5 × 5.1 μm,  $n$  = 20), 1–2-seriate to overlapping uniseriate, ellipsoid, slightly pointed at the ends, aseptate, dark brown, guttulate, smooth-walled, with a 1.5–3.5 μm thick mucilaginous sheath, with a straight germ slit, extending the full-length. **Asexual morph:** Undetermined.

**Culture characters:** Ascospores germinating on PDA within 48 h with germ tubes produced from lower end. Colonies growing slowly on PDA, reaching 5 mm in 4 weeks at 28 °C, effuse, velvety to hairy, white from above, dark brown from the below. Mycelium superficial and immersed, branched, septate, smooth, hyaline, irregular.

**Fig. 59** *Neoanthostomella pseudostromatica* (MFLU 15–1190, **holotype**). **a–d** Appearance of pseudostromata on bamboo host. **e** Vertical section of pseudostroma. **f** Ostiole. **g** Peridium. **h, i** Asci. **k** Germinating ascospore. **j, l, m** Ascospores. **n, o** Culture on PDA. Scale bars: a = 5 mm, b–d = 1 mm, e = 100 μm, f–g = 50 μm, h–m = 5 μm



**Material examined:** THAILAND, Chiang Rai, Mae Fah Luang University, on dead culms of bamboo, 1 August 2011, Dong-Qin Dai DDQ00091 (MFLU 15–1190, **holotype**); *Ibid.* (KUN HKAS88700, **isotype**), living cultures, MFLUCC 11–0610, KUMCC.

**Notes:** *Neoanthostomella pseudostromatica* is similar to *Anthostomella tumulosa* (Roberge ex Desm.) Sacc. in its globose ascomata with a central, periphysate, ostiolar canal (Lu and Hyde 2000). However, *N. pseudostromatica* can be distinguished as the asci lack an apical ring (*A. tumulosa* has a J+, subapical ring) and smaller ascospores (11.5–15 × 4–5.5 μm vs. 17.5–22.5 × 6.5–9.5 μm) (Lu and Hyde 2000). According to Eriksson and Yue (1998), and Lu and Hyde (2000), several *Anthostomella* species occurred on bamboo, such as *Anthostomella bambusicola*, *A. eructans*, *A. longa*, *A. nitidissima*, *A. palmicola*, *A. puiggarii*, *A. punctulata*, *A. rehmi*, *A. tomicoides*. These species have solitary, papillate ascomata, *Neoanthostomella pseudostromatica* however, differs in having 2–5 ascomata aggregating in a black pseudostroma, and each ascoma with a central, periphysate, ostiolar neck.

**Vamsapriya** Gawas & Bhat, Mycotaxon 94: 150 (2006)

*Vamsapriya*, typified by *V. indica* Gawas & Bhat, was originally described by Gawas and Bhat (2005), based on a collection from a bamboo host. Later Pratibha and Bhat (2008) described a second species *V. mahabaleshwariensis* Pratibha & Bhat, collected on an unidentified dead twig. Dai et al. (2014c) recollected and epityped the genus type, *V. indica*, and introduced two more species based on the phylogenetic analyses. However, molecular data of *V. mahabaleshwariensis* is still unavailable in GenBank. *Vamsapriya* was established in family *Xylariaceae* by Dai et al. (2014c) and Liu et al. (2015) based on multi-gene analyses. Initially *Vamsapriya* species were only found as asexual morphs by Gawas and Bhat (2005), Pratibha and Bhat (2008) and Dai et al. (2014c). In this study we observed the sexual morph of *V. bambusicola* in Fig. 60.

An earlier reference was also found with a detailed description of a fungus with similar morphology to that of *Vamsapriya* Gawas & Bhat, but named as *Tretophragmia* Subram. & Natarajan (Subramanian and Natarajan 1972). The genus, typified by *T. longispora*, is characterized by conspicuous, long, compact, apically flared, synnematous, branched, dark brown conidiphores, monotretic conidiogenous cells and solitary, long, transversely septate, flexuous, smooth, dark brown conidia. The fungus was isolated from dead bamboo twigs, in Coonoor, Tamil Nadu, India.

It is possible that *Tretophragmia* and *Vamsapriya* could be congeneric. These two genera are distinguished by a single character. That is, in *Tretophragmia* the conidia are solitary, whereas they are catenate in *Vamsapriya*. However, the type of *Tretophragmia* has not been sequenced and until that is done, the fungi are maintained as two distinct genera.

Based on a blast search of LSU sequence data in GenBank, *Vamsapriya* species are close to *Fasciatispora nypae* K.D. Hyde (type species of *Fasciatispora*). However, *Vamsapriya* is distinct from *Fasciatispora* in having hyaline apiospores, whereas *Fasciatispora* has dark ascospores typically with a pallid band (Alias et al. 1994; Hyde 1991, 1995; Hidayat et al. 2007). Furthermore, in the phylogenetic tree (Fig. 54) based on ITS sequence data, *Vamsapriya* species form a clade in *Xylariaceae*, and separate from *Fasciatispora nypae* (MFLUCC 11–0382, reference strain designated by Liu et al. (2015)).

**Vamsapriya bambusicola** D.Q. Dai, D.J. Bhat & K.D. Hyde, Cryptog. Mycol. 35(4): 353 (2014)

Facesoffungi number: FoF 02006; Figs. 60 and 61

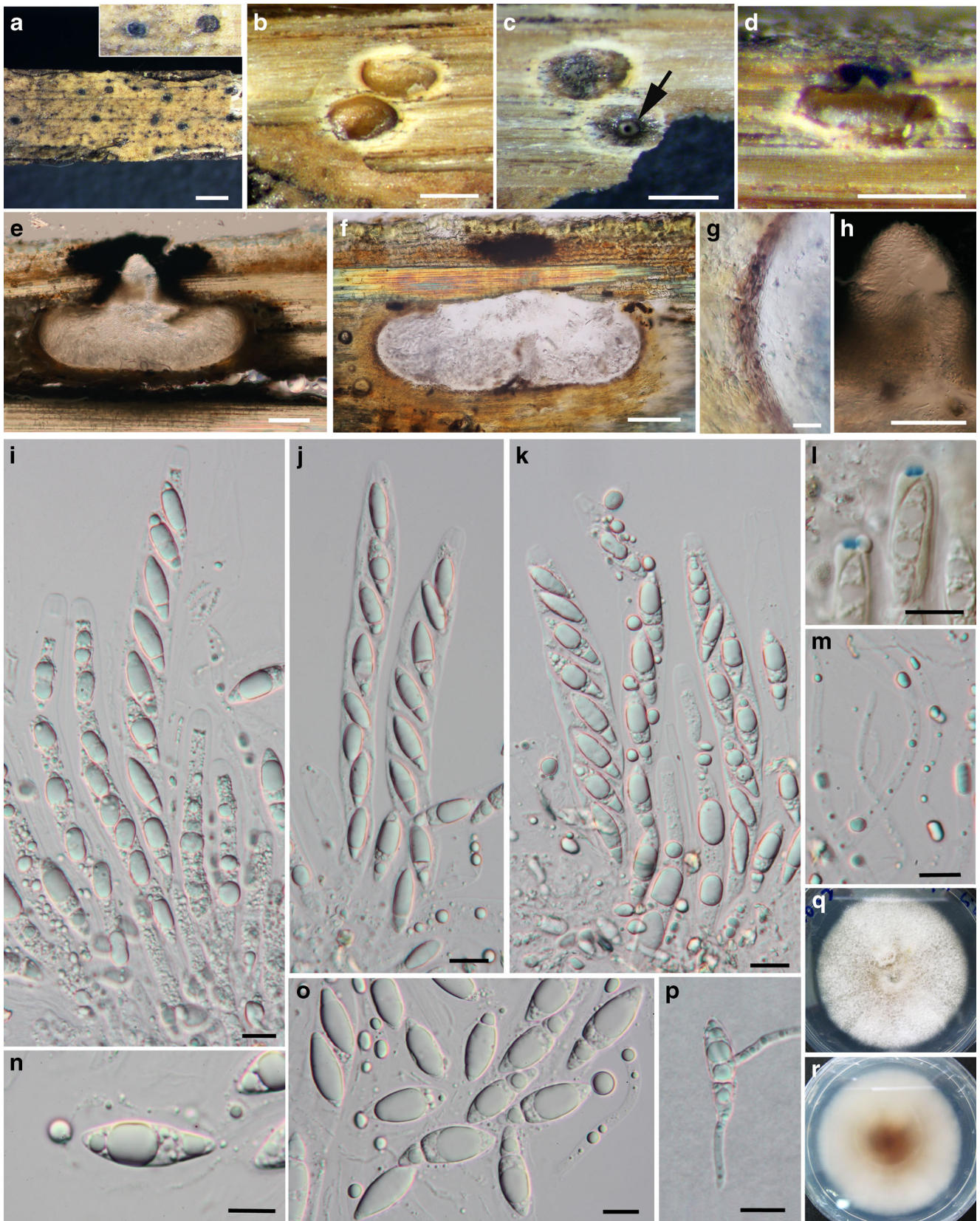
**Saprobic** on dead bamboo culms, immersed within the host cortex, black pigmented, clypeus-like tissue around ostiolar opening, visible as black, circular, 120–320 μm diam., spots on the host surface. **Sexual morph:** *Ascomata* 450–600 × 380–500 μm, 400–500 μm high, solitary, scattered, immersed within the host cortex, subglobose, light brown, papillate, with black clypeus-like tissue surrounding the papillate ostiolar opening. *Ostioles* 60–80 μm diam., 100–120 μm high, raised from center of ascomata, with periphyses. *Peridium* laterally 10–15 μm thick, composed of thin-walled, light brown to hyaline cells of *textura angularis*. *Hamathecium* comprising long, 1.5–3 μm wide, hyaline, septate paraphyses. *Asci* 115–140 × 10.5–18.5 μm ( $\bar{x}$  = 124.8 × 13.8 μm,  $n$  = 20), 8-spored, unitunicate, cylindrical, short pedicellate, with a 1.5–2.5 high × 2.5–3.5 diam. J+, apical ring and round to truncate apex. *Ascospores* 20.5–26.5 × 6.5–7.5(–7.9) μm ( $\bar{x}$  = 23.4 × 7.4 μm,  $n$  = 20), overlapping uniseriate to 1–2-seriate, apiosporous, fusiform to broad fusiform, 1-septate, hyaline, pointed at both ends, usually with a large guttule in upper cell, and a small round guttule in lower cell, surrounded by a small inconspicuous mucilaginous sheath. **Asexual morph:** Refer to Dai et al. (2014c).

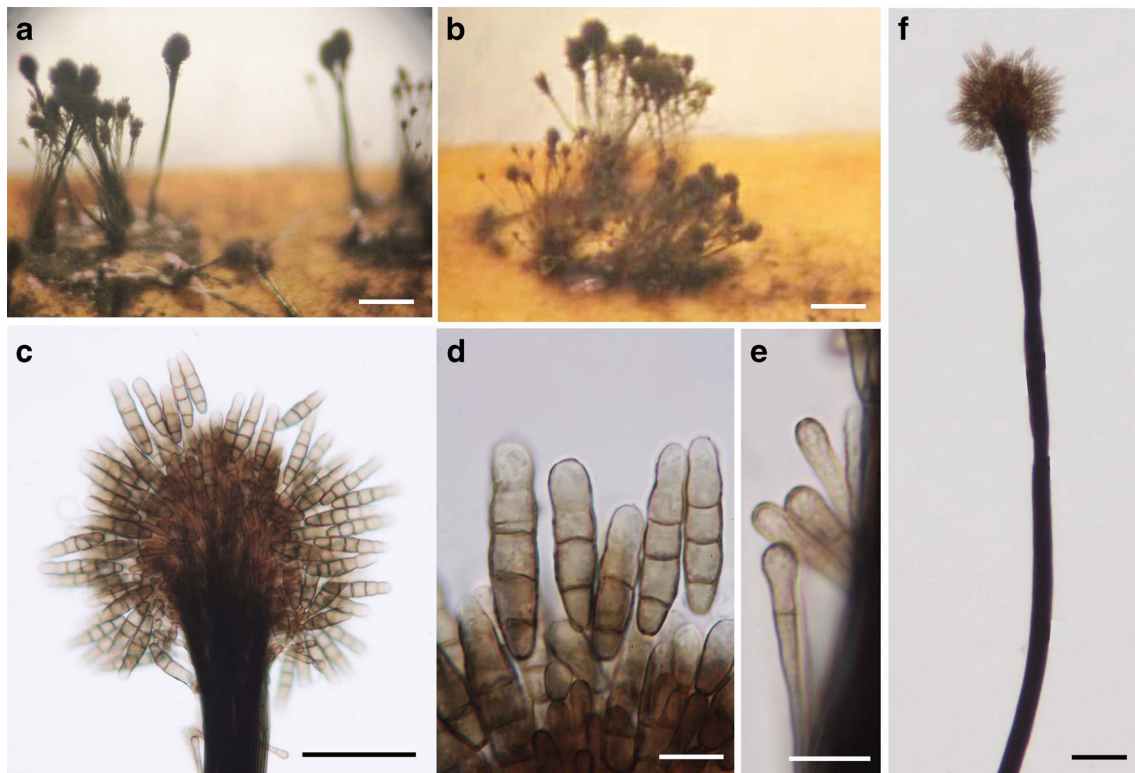
**Cultural characters:** Ascospores germinating on PDA within 36 h and germ tubes produced from both cells. Colonies growing fast on PDA, reaching 5 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, cottony and light-coloured at the centre, circular, from above; light brown from the below. Mycelium immersed in the media, composed of branched, septate, smooth, hyaline, hyphae.

**Fig. 60** *Vamsapriya bambusicola* (MFLU 15–1315). **a** Species visible as blacked ostiolar opening on host surface. **b, c** Horizontal section of ascomata (**c**: Showing ostiolar opening). **d** Vertical section of ascoma showing black clypeus around papillate ostiole. **e, f** Vertical section of ascomata. **j** Peridium. **h** Ostiole with periphyses. **i–k** Asci. **l** Ascospores stained blue in Melzer's reagent. **m** Paraphyses. **n, o** Ascospores (**n** showing ascospore surrounding by sheath). **p** Germinating ascospore. **p, r** Cultures on PDA. Scale bars: a = 1 mm, b–d = 500 μm, e, f = 100 μm, h = 50 μm, i–p = 10 μm

*Material examined:* THAILAND, Uttaradit Province, on dead culms of bamboo, 28 October 2011, Dong-Qin Dai,

DDQ00221 (MFLU 15–1315); *Ibid.* (KUN, HKAS88972), living cultures, MFLUCC 11–0637, KUMCC.





**Fig. 61** *Vamsapriya bambusicola* (MFLU 13–0368, holotype). **a, b** Synnemata on bamboo host. **c** Apical part of synnema. **d** Conidia. **e** Conidiogenous cells. **f** Synnema. Scale bars: a, b = 500  $\mu\text{m}$ , c, f = 100  $\mu\text{m}$ , d, e = 10  $\mu\text{m}$

*Notes:* *Vamsapriya bambusicola* was introduced by Dai et al. (2014c) to accommodate a hyphomycetous fungus with dark synnemata, monotretic and enteroblastic, conidiogenous cells, and cylindrical, brown, septate conidia. The newly recognized sexual morph strain (MFLUCC 11–0637) is linked to type strain (MFLUCC 11–0477) by ITS phylogenetic analyses with high bootstrap support (100/100 % MPBP/MLBP) (Fig. 53).

*Vamsapriya bambusicola* is similar to members of *Apioclypea* K.D. Hyde by having immersed ascomata with a rounded, black clypeus and hyaline apiospores (Hyde et al. 1998). *Vamsapriya bambusicola* however, has longer ascospores (can reach to 26.5  $\mu\text{m}$  long), compared to *Apioclypea* species which are shorter than 25  $\mu\text{m}$  (Hyde 1994; Hyde et al. 1998; Taylor and Hyde 2003). So far *Apioclypea* species were only observed on palms and are placed in family *Clypeosphaeriaceae* (Hyde et al. 1998; Taylor and Hyde 2003). Smith et al. (2003) indicated *Apioclypea* is polyphyletic, and submitted a LSU gene of *Apioclypea* sp. to GenBank. However, the blast search using LSU gene of *Vamsapriya bambusicola* in GenBank shows they are not related. Further collections of these genera are needed to establish their natural placement in Xylariomycetidae (Maharachchikumbura et al. 2015; Senanayake et al. 2015).

*Xylaria* Hill ex Schrank, Baier. Fl. (München) 1: 200 (1789)

*Xylaria* are commonly seen on dead wood, but can also be found in soil or on various other substrates, such as decaying fallen leaves, petioles, herbaceous stems, dung, grasses, seeds or fruits and wood (Ju and Tzean 1985; Rogers 1986; Rogers and Samuels 1986). They are also dominant endophytes in the tropics (Rodrigues and Petrini (1997). Kirk et al. (2008) reported 300 species in *Xylaria*, however, currently more than 700 are listed in Index Fungorum (2016). Ju et al. (2012) synonymized *Penzigia* under *Xylaria* and provided a key for the major penzigoid species of *Xylaria*. Asexual morphs are *Moelleroclavus*, *Geniculosporium*-like, *Xylocoremium* (Kirk et al. 2008; Wijayawardene et al. 2012).

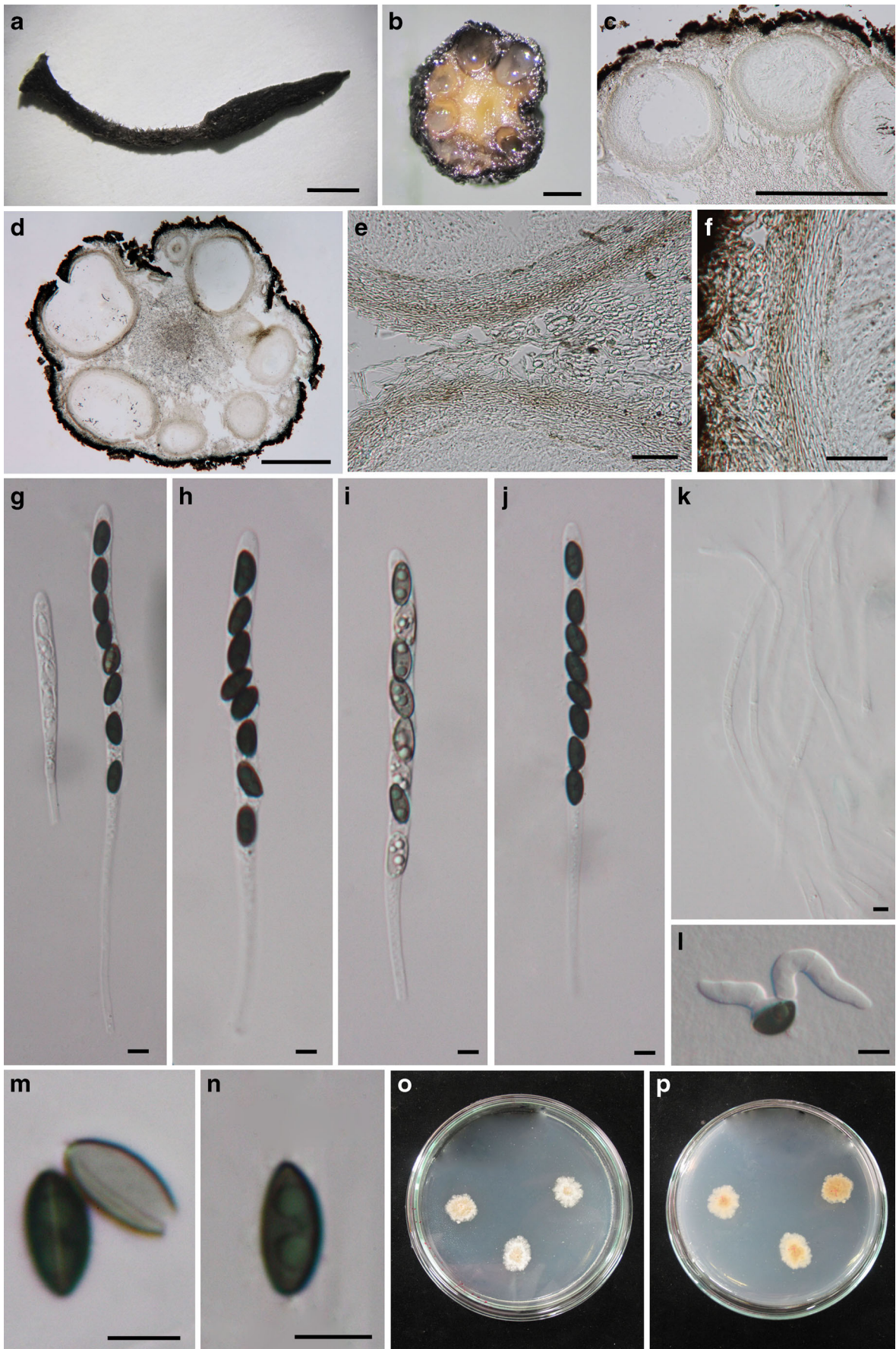
*Xylaria bambusicola* Y.M. Ju & J.D. Rogers, Mycotaxon 73: 400 (1999)

*Facesoffungi* numbers: FoF 02007; Fig. 62

*Saprobic* on dead bamboo culms, forming large, long, *Xylaria*-like stromata on bamboo host. *Stromata* solitary to densely gregarious, superficial, clavate to cylindrical, black, carbonaceous to coriaceous, unbranched to

**Fig. 62** *Xylaria bambusicola* (MFLU 15–1188). **a** Stromata. **b–d** Horizontal section of stroma. **e, f** Peridium. **g–j** Asci. **k** Paraphyses. **l, m** Ascospores **n** Germinating ascospore. **o, p** Culture on PDA. Scale bars: a = 5 mm, b–d = 500  $\mu\text{m}$ , e, f = 50  $\mu\text{m}$ , k–m = 5  $\mu\text{m}$





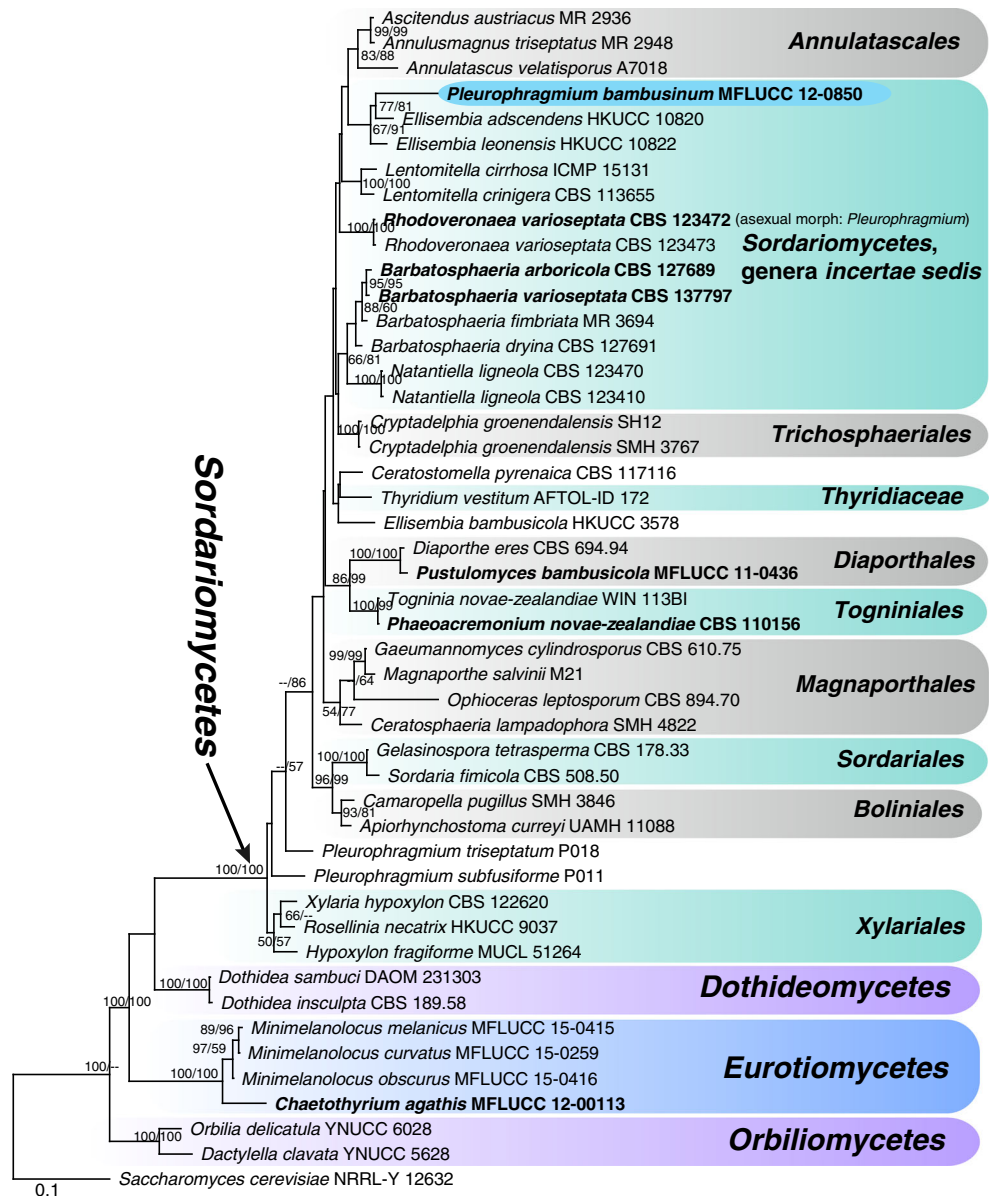
branched, 2–6 cm long in total; well defined, slender, black stipes, covered by short black hairs, 2–3 mm diam.; top fertile part 1–3 cm long, 5–6 mm diam.. *Outer layer of stromata* 25–55 µm thick composed of black tissue. *Cells between perithecia* light brown to hyaline, of *textura intricata* to *oblita*. *Perithecia* 300–650 µm diam., immersed, arranged under outside layer of stromata, globose, membranous, light brown. *Peridium* laterally 20–45 µm thick, composed of small, light brown to hyaline cells of *textura oblita*. *Hamathecium* composed of long, hyaline, septate, 1.5–2 µm broad, paraphyses. *Asci* 95–120 × 4.5–5.5 µm ( $\bar{x}$  = 99.7 × 4.8 µm,  $n=20$ ), 8-spored, unitunicate, cylindrical, with a long, thin pedicel and a 1–1.5 high × 2–3 diam. J+, round apical ring. *Ascospores* 6–8 × 3–3.5 µm ( $\bar{x}$  =

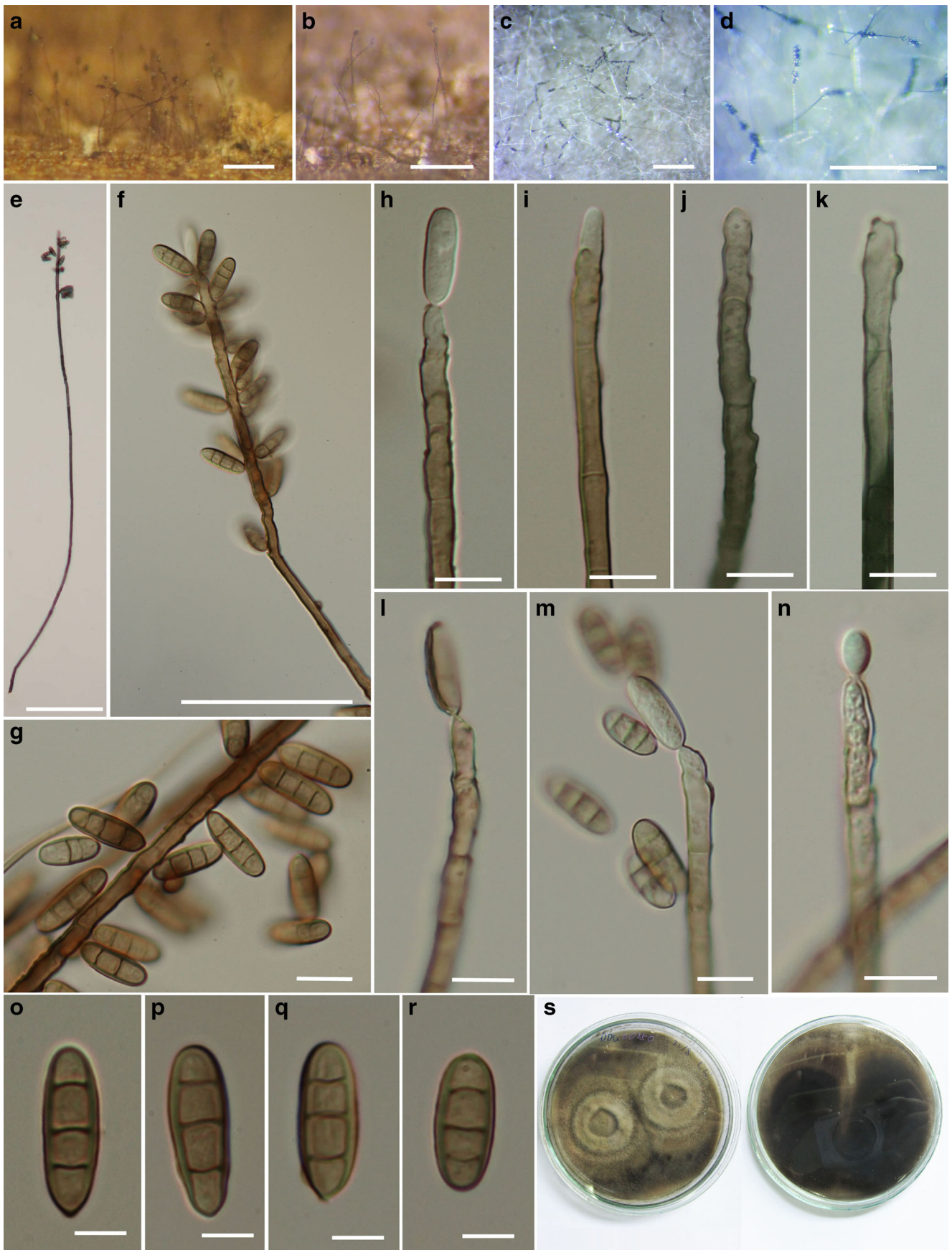
**Fig. 64** *Pleurophragmium bambusinum* (MFLU 15–1207, holotype: a, b, e, j, k observed from host substrate; c, d, f, h, i, g, l–r observed from PDA culture). **a, b** Conidiomata on bamboo host. **c, d** Sporulation on PDA. **e** Conidiophore. **f** Upper part of conidiophore. **g–n** Polyblastic conidiogenous cells (**g, h, l–n** Conidiogenous cells producing conidia). **o–r** brown conidia with 3 septa. **s** Colonies on PDA. Scale bars: a, b = 1 mm, c, d = 500 µm, e = 100 µm, f = 50 µm, h–n = 10 µm, q–r = 5 µm

6.8 × 3.4 µm,  $n=20$ ), uniseriate, aseptate, elliptical, light brown to brown, with narrowly rounded ends, smooth-walled, with asymmetric, full length, germ-slit.

*Culture characters:* Ascospores germinating on PDA within 24 h and germ tubes produced from both ends of spore. Colonies growing fast on PDA, reaching 90 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, cottony circular, irregular edge, white from above,

**Fig. 63** Maximum likelihood phylogenetic tree (lnL = -2178.664062) generated by RAxML (GTR + G model) based on LSU sequence data. MP/ML values (>50 %) resulting from 1000 bootstrap replicates are given at the nodes. The original isolate numbers codes are noted after the species names. The tree is rooted to *Saccharomyces cerevisiae* (CBS 116131). Ex-type strains are in bold. Newly generated sequences are highlighted with a blue background





brown in centre from below. Mycelium superficial to immersed in/on media, with branched, septate, smooth hyphae.

**Material examined:** THAILAND, Chiang Rai, Mae Fah Luang University, on dead culms of bamboo, 1 August 2011, Dong-Qin Dai DDQ00086 (MFLU 15–1188); *ibid.* (KUN HKAS88698), living cultures, MFLUCC 11–0606, CBS 139988.

**Notes:** *Xylaria bambusicola* was originally described by Ju and Rogers (1999) based on a collection from bamboo, in Taiwan. Our new strain (MFLUCC 11–0606) groups within the type strain of *Xylaria bambusicola* (WSP205) with high bootstrap support (100 %/99 % MPBP/MLBP) (Fig. 54). The new collection has similar morphology with the type, however, is shorter in asci (95–120  $\mu\text{m}$  vs. 120–150  $\mu\text{m}$  in length) and ascospores (6–8  $\mu\text{m}$  vs. 9.5–11  $\mu\text{m}$  long) (Ju and Rogers 1999). *Xylaria bambusicola* can be compared with *X. apiculata* and *X. arbuscula* in having black *Xylaria*-like stromata. However, *Xylaria apiculata* has larger ascospores (20–22  $\times$  7  $\mu\text{m}$  vs. 6–8  $\times$  3–3.5  $\mu\text{m}$ ) (Cooke 1879). *Xylaria bambusicola* differs from *X. arbuscula* in its smaller ascospores (6–8  $\times$  3–3.5  $\mu\text{m}$  vs. 14–16  $\times$  5–7  $\mu\text{m}$ ) (Saccardo 1878) and in having a full length germ-slit (Ju and Rogers 1999).

**Sordariomycetes, genera incertae sedis**

***Pleurophragmium*** Costantin, Mucéd. Simpl. (Paris): 100 (1888)

*Pleurophragmium* was established by Costantin (1888) with *P. bicolor* Costantin as type species, and was placed in Ascomyceta, genera *incertae sedis*. The genus is characterized by single unbranched, brown to dark brown conidiophores, with polyblastic, integrated, terminal, sympodial, denticulate conidiogenous cells, producing ellipsoid to fusiform, or clavate, brown, 0–7-septate conidia, with pointed bases (De Hoog 1985; Abarca et al. 2007; Ma et al. 2014). The feature of dark conidia is used to distinguish *Pleurophragmium* from *Dactylaria* (De Hoog 1985; Abarca et al. 2007). D'Souza and Bhat (2012) compared 23 species of *Pleurophragmium* based on their substrates and locations and provided the key for species. Réblová (2009) reported a sexual species *Rhodoveronaea varioseptata* Arzanlou, et al. with a *Pleurophragmium* asexual morph, and provided a phylogenetic analysis based on LSU sequence data. Our phylogenetic tree in Fig. 63, shows *Pleurophragmium* clusters in *Sordariomycetes*, genera *incertae sedis*. More than 20 species were introduced in this genus (Abarca et al. 2007), and its species may be polyphyletic. However, very little molecular data is available in GenBank, and fresh collections are requested to clarify the order and family level placement of the genus, as well as species relationships.

***Pleurophragmium bambusinum*** D.Q. Dai & K.D. Hyde, *sp. nov.*

**Indexfungorum number:** IF552043; **Facesoffungi number:** FoF 02008; **Fig.** 64

**Etymology:** Refers to the host bamboo.

**Holotype:** MFLU 15–1207

**Saprobic** on decaying bamboo culms. **Mycelium** immersed in the substrate, composed of septate, branched, hyphae. **Sexual morph:** Undetermined. **Asexual morph:** **Conidiophores** 800–1900  $\times$  3–4.5  $\mu\text{m}$  ( $\bar{x}$  = 968.3  $\times$  3.9  $\mu\text{m}$ ,  $n=20$ ) macronematous, cylindrical, brown to dark brown, septate, unbranched, straight to slightly flexuous. **Conidiogenous cells** 12–25  $\times$  3–4.5  $\mu\text{m}$  ( $\bar{x}$  = 17.3  $\times$  4.1  $\mu\text{m}$ ,  $n=20$ ), polyblastic, integrated, terminal, sympodial, cylindrical, pale brown, smooth, straight, denticulate. **Conidia** 13–21  $\times$  5–7  $\mu\text{m}$  ( $\bar{x}$  = 17.6  $\times$  6.2  $\mu\text{m}$ ,  $n=20$ )  $\mu\text{m}$ , ellipsoid, slightly narrow at towards base, straight to slightly curved, 3-septate, brown, smooth-walled, thick-walled, without guttules.

**Culture characters:** Ascospores germinating on PDA within 24 h and germ tubes produced from basal end. Colonies growing slowly on PDA, reaching 5 mm in 2 weeks at 28 °C, under 12 h light/12 h dark, circular, dark brown from above and below. Mycelium superficial to immersed in media, branched, septate, smooth. Fertile after 2 months in room temperature on PDA.

**Material examined:** THAILAND, Chiang Rai, Hnong Kao Hong, Dong Mada, Mea Laos, on dead culms of bamboo, 3 August 2012, D. Jayarama Bhat DDQ00250 (MFLU 15–1207, **holotype**); *Ibid.* (KUN HKAS83941, **isotype**), living cultures, MFLUCC 12–0850, KUMCC.

**Notes:** *Pleurophragmium bambusicola* is characterized by polyblastic, sympodial, denticulate conidiogenous cells and 3-septate, brown, thick-walled conidia. D'Souza and Bhat (2012) described *P. indicum* M.A. D'Souza & Bhat and *P. varioseptatum* Matsush. from bamboo. However, *P. indicum* has larger conidia (20–30  $\times$  4.5–11  $\mu\text{m}$  vs. 13–21  $\times$  5–7  $\mu\text{m}$ ) with varied coloration (middle cells dark brown and end cells pale brown vs. identical color). *Pleurophragmium bambusicola* differs from *P. varioseptatum* in having smooth-walled, ellipsoid conidia (D'Souza and Bhat 2012).

## Discussion

This study treats 43 fungal species representing 24 genera, including 29 new species, three new genera and one new family. They are arranged into two classes. Twenty-one fungal species belonging to eight genera (*Bambusaria*, *Bambusicola*, *Mendogia*, *Neodeightonia*, *Occultibambusa*, *Roussoella*, *Seriascoma* and *Tubeufia*), of seven families (*Bambusicolaceae*, *Botryosphaeriaceae*, *Myriangiaceae*, *Occultibambusaceae*, *Roussoellaceae*, *Tubeufiaceae* and

**Table 2** List of bambusicolous fungi published since Hyde et al. (2002b)

Taxa	Bamboo host	Substrate	Country	Order/Family	References
<i>Amphibambusa bambusicola</i>	Bamboo	On dead culms	Thailand	<i>Amphisphaeriales/Amphisphaeriaceae</i>	Liu et al. (2015)
<i>Anthostomella pseudobambusicola</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Xylariaceae</i>	This study
<i>Arthrinium hyphopodii</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Apiosporaceae</i>	Senanayake et al. (2015)
<i>A. longistromum</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Apiosporaceae</i>	This study
<i>A. rasikravindrae</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Apiosporaceae</i>	This study
<i>A. subglobosa</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Apiosporaceae</i>	Senanayake et al. (2015)
<i>A. thailandicum</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Apiosporaceae</i>	This study
<i>A. yunnanum</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Apiosporaceae</i>	This study
<i>Astrocystis mirabilis</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Xylariaceae</i>	This study
<i>Astrosphaeriella africana</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Liu et al. (2011)
<i>A. bambusae</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>A. exorrhiza</i>	<i>Thysanolaena maxima</i>	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>A. fusispora</i>	<i>Phyllostachis bambusoides</i>	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>A. neofusispora</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>A. neostellata</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>A. stellata</i>	<i>Phyllostachis bambusoides</i>	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Liu et al. (2011); Phookamsak et al. (2015)
<i>A. thailandica</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>A. thysanolaenae</i>	<i>Thysanolaena maxima</i>	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>A. tornata</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>Bambusaria bambusae</i>	<i>Thyrsostachys stamensis</i>	On dead culms	Thailand	<i>Pleosporales/Astrosphaeriellaceae</i>	Phookamsak et al. (2015)
<i>Bambusicola bambusae</i>	Bamboo	On dead culms	Thailand	<i>Valsariales/Valsariaceae</i>	Jaklitsch et al. (2015); This study
<i>B. didymospora</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	Dai et al. (2012)
<i>B. irregularispora</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	This study
<i>B. loculata</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	Dai et al. (2012)
<i>B. massarina</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	Dai et al. (2015)
<i>B. pustulata</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	Dai et al. (2012)
<i>B. splendida</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	This study
<i>B. thailandica</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	Dai et al. (2012)
<i>B. triseptatispora</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	This study
<i>Bambusistroma didymosporum</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	This study
<i>Botryobambusa fusisporum</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Bambusicolaceae</i>	Adamčík et al. (2015)
				<i>Botryosphaeriales/Botryosphaeriaceae</i>	Liu et al. (2012a)

Table 2 (continued)

Taxa	Bamboo host	Substrate	Country	Order/Family	References
<i>Brunneoclavivora bambusae</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Halotheiaceae</i>	Ariyawansa et al. (2015)
<i>Cercophora thailandica</i>	Bamboo	On dead culms	Thailand	<i>Sordariales/Lasiothraeriaceae</i>	This study
<i>Collodiscula fangjingshanensis</i>	Bamboo	On dead culms	China	<i>Xylariales/Xylariaceae</i>	Liu et al. (2015)
<i>C. leigongshanensis</i>	Bamboo	On dead culms	China	<i>Xylariales/Xylariaceae</i>	Liu et al. (2015)
<i>Daldinia bambusicola</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Xylariaceae</i>	This study
<i>Dicystosporium pseudomusae</i>	Bamboo	On dead culms	Japan	<i>Pleosporales/Dicystosporiaceae</i>	Tanaka et al. (2015)
<i>Dinemasporium bambusicola</i>	<i>Pleuroblastus chino</i>	On dead culms	Japan	<i>Xylariomycetidae/ Incertae sedis</i>	Hashimoto et al. (2015)
<i>D. cruciferum</i>	Bamboo	On dead culms	Japan	<i>Xylariomycetidae/ Incertae sedis</i>	Hashimoto et al. (2015)
<i>D. japonicum</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Xylariomycetidae/ Incertae sedis</i>	Hashimoto et al. (2015)
<i>D. longicapillatum</i>	Bamboo	On dead culms	Japan	<i>Xylariomycetidae/ Incertae sedis</i>	Hashimoto et al. (2015)
<i>D. parastrigosum</i>	Bamboo	On dead culms	Japan	<i>Xylariomycetidae/ Incertae sedis</i>	Hashimoto et al. (2015)
<i>D. rishiriense</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Xylariomycetidae/ Incertae sedis</i>	Hashimoto et al. (2015)
<i>D. sasae</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Xylariomycetidae/ Incertae sedis</i>	Hashimoto et al. (2015)
<i>D. strigosum</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Xylariomycetidae/ Incertae sedis</i>	Hashimoto et al. (2015)
<i>Dothiorella thailandica</i>	Bamboo	On dead culms	Thailand	<i>Botryosphaeriales/Botryosphaeriaceae</i>	Liu et al. (2012a); Phillips et al. (2013)
<i>Embryonispora bambusicola</i>	Bamboo	On dead culms	China	<i>Ascomycota/ Incertae sedis</i>	Zhao et al. (2014)
<i>Eriosporella bambusicola</i>	Bamboo	On dead culms	Thailand	<i>Capnodiales/Incertae sedis</i>	Dai et al. (2014b)
<i>Eunypha linearis</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Diatrypaceae</i>	This study
<i>Fissuroma aggregata</i> ≡ <i>Astrophaerella aggregata</i>	<i>Phyllostachys bambusoides</i>	On dead culms	Japan, Thailand	<i>Pleosporales/Aigialaceae</i>	Tanaka and Harada (2005a); Liu et al. (2011); Phookamsak et al. (2015)
<i>F. bambusae</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Aigialaceae</i>	Phookamsak et al. (2015)
<i>F. neoaggregata</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Aigialaceae</i>	Phookamsak et al. (2015)
<i>F. thailandicum</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Aigialaceae</i>	Phookamsak et al. (2015)
<i>Flammeascooma bambusae</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Antegloniaceae</i>	Liu et al. (2015)
<i>Goosiomycetes bambusicola</i>	<i>Bambusa arundinacea</i>	On dead culms	China	<i>Ascomycota/ Incertae sedis</i>	Dubey and Neelima (2014)
<i>Gibberella bambusae</i>	Bamboo	On dead culms	China	<i>Hypocreales/Nectriaceae</i>	Zhang and Zhuang (2003)
<i>Gregarithecium curvisporum</i>	<i>Sasa</i> sp.	On dead culms	Japan	<i>Pleosporales/Dicystosporiaceae</i>	Tanaka et al. (2015)
<i>Helminthosporium bambusicola</i>	<i>Bambusa</i> sp.	On dead culms	China	<i>Pleosporales/Massariniaceae</i>	Zhang et al. (2010)
<i>Hypoxylon laminosum</i>	Bamboo	On dead culms	French	<i>Xylariales/Xylariaceae</i>	Kuhnert et al. (2014)
<i>H. neosublenormandii</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Xylariaceae</i>	This study
<i>H. pseudofendleri</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Xylariaceae</i>	This study
<i>H. sublenormandii</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Xylariaceae</i>	Suwannasat et al. (2005)

Table 2 (continued)

Taxa	Bamboo host	Substrate	Country	Order/Family	References
<i>Kamalamyces indicus</i>	<i>Dendrocalamus strictus</i>	On dead culms	Thailand	<i>Tubeufiales/Tubeufiaceae</i>	Boonmee et al. (2011)
<i>Katumotoa bambusicola</i>	<i>Sasa kurilensis</i> ;	On dead culms	Japan	<i>Pleosporales/Lentitheciaceae</i>	Tanaka and Harada (2005a); Tanaka et al. (2015)
<i>Leptosporella bambusae</i>	Bamboo	On dead culms	Thailand	<i>Chaetosphaeriales/Incertae sedis</i>	This study
<i>Linocarpon bambusicola</i>	<i>Bambusa</i> sp.	On dead culms	Philippines	<i>Xylariales/Incertae sedis</i>	Cai et al. (2004)
<i>Lophiostoma arundinis</i>	<i>Sasa palmata</i>	On dead culms	Japan	<i>Pleosporales/Lophiostomataceae</i>	Tanaka and Harada (2003a)
<i>Massarina arundinariae</i>	<i>Phyllostachys bambusoides</i>	On dead culms	Japan	<i>Pleosporales/Massarinaeae</i>	Tanaka and Harada (2003b)
<i>M. bambusina</i>	<i>Sasa kurilensis</i> ; <i>Phyllostachys bambusoides</i>	On dead culms	Japan	<i>Pleosporales/Massarinaeae</i>	Tanaka and Harada (2003b)
<i>M. ryukyensis</i>	<i>Pleioblastus linearis</i>	On dead culms	Japan	<i>Pleosporales/Massarinaeae</i>	Tanaka and Harada (2003b)
<i>Mendogia bambusina</i>	Bamboo	On living culms	Thailand	<i>Myrangiiales/Myrangiaceae</i>	This study
<i>Myrothecium thailandicum</i>	Bamboo	On dead culms	Thailand	<i>Hypoerales/Stachybotryaceae</i>	This study
<i>M. chiangmaiense</i>	Bamboo	On dead culms	Thailand	<i>Hypoerales/Stachybotryaceae</i>	This study
<i>M. cylindrosporum</i>	Bamboo	On dead culms	Thailand	<i>Hypoerales/Stachybotryaceae</i>	Liu et al. (2015); This study
<i>M. uttaraditense</i>	Bamboo	On dead culms	Thailand	<i>Hypoerales/Stachybotryaceae</i>	This study
<i>Nectria pseudotrictia</i>	Bamboo	On dead culms	Thailand	<i>Hypoerales/Nectriaceae</i>	This study
<i>Neoanthostomella pseudostromatica</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Xylariaceae</i>	This study
<i>Neodeightonia microspora</i>	Bamboo	On dead culms	Thailand	<i>Botryosphaeriales/Botryosphaeriaceae</i>	This study
<i>N. subglobosa</i>	<i>Bambusa arundinacea</i>	On dead culms	Thailand	<i>Botryosphaeriales/Botryosphaeriaceae</i>	Liu et al. (2012a); This study
<i>Neogaeanomyces bambusicola</i>	Bamboo	On dead culms	Thailand	<i>Magnaporthales/Magnaporthaceae</i>	Liu et al. (2015)
<i>Neokalmusia brevispora</i>	<i>Sasa senanensis</i> ; <i>Sasa kurilensis</i>	On dead culms	Japan	<i>Pleosporales/Didymosphaeriaceae</i>	Tanaka and Harada (2004); Ariyawansa et al. (2014)
<i>Neokalmusia didymospora</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Didymosphaeriaceae</i>	Dai et al. (2016)
<i>Neokalmusia scabrisspora</i>	<i>Phyllostachys bambusoides</i>	On dead culms	Japan	<i>Pleosporales/Didymosphaeriaceae</i>	Tanaka et al. (2005)
<i>Neophiophlaerella sasicola</i>	<i>Sasa senanensis</i> ; <i>Sasa kurilensis</i>	On dead culms	Japan	<i>Pleosporales/Lentitheciaceae</i>	Tanaka et al. (2015)
<i>Neourussoella bambusae</i>	<i>Bambusa</i> sp.	On dead culms	Thailand	<i>Pleosporales/Roussioellaceae</i>	Liu et al. (2014)
<i>Occlitibambusa bambusae</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Occlitibambusaceae</i>	This study
<i>O. fisispora</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Occlitibambusaceae</i>	This study
<i>O. pustula</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Occlitibambusaceae</i>	This study
<i>Oxydothis bambusicola</i>	<i>Bambusa</i> sp.	On dead culms	China	<i>Xylariales/Incertae sedis</i>	Shenoy et al. (2005)
<i>Parabambusicola bambusina</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Pleosporales/Parabambusicolaceae</i>	Tanaka et al. (2015)
<i>Passalora bambusae</i>	<i>Bambusa spinosa</i>	On culms	India	<i>Capnodiales/Mycosphaerellaceae</i>	Kamal (2010)
<i>Peroneutypa scoparia</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Diatrypaceae</i>	This study
<i>Phaeoacromonium sphinctrophorum</i>	Bamboo	On dead culms	Thailand	<i>Togniniales/Togniniaceae</i>	This study
<i>Phaeoisaria pseudoclematidis</i>	Bamboo	On dead culms	Thailand	<i>Xylariales/Diatrypaceae</i>	Liu et al. (2015)

Table 2 (continued)

Taxa	Bamboo host	Substrate	Country	Order/Family	References
<i>Phaeosphaeria bambusae</i>	<i>Pleiotblastus simoni</i>	On dead culms	Japan	<i>Pleosporales/Phaeosphaeriaceae</i>	Tanaka and Harada (2004)
<i>P. oryzae</i>	<i>Bambusa multiplex</i>	On dead culms	Japan	<i>Pleosporales/Phaeosphaeriaceae</i>	Tanaka and Harada (2004)
<i>Phaeosphaeria</i> sp.	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Pleosporales/Phaeosphaeriaceae</i>	Tanaka and Harada (2004)
<i>Phialosporostilbe gregariclava</i>	<i>Sasa nipponica</i>	On dead culms	Japan	Ascomycota incertae sedis	Shirouzu and Harada (2004)
<i>Pleurophragmium bambusinum</i>	Bamboo	On dead culms	Thailand	<i>Sordariomycetes</i> genera, <i>incertae sedis</i>	This study
<i>Polyphosphaeria fusca</i>	<i>Pleiotblastus chino</i> ; <i>Phyllostachys bambusoides</i> ; <i>Chimonobambusa marmorata</i> ; <i>Sasa kurilensis</i>	On dead culms	Japan	<i>Pleosporales/Tetraplospiraeraceae</i>	Tanaka et al. (2009)
<i>Pseudoastrospiraeriella africana</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Pseudoastrospiraeriellaceae</i>	Phookamsak et al. (2015)
<i>P. bambusae</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Pseudoastrospiraeriellaceae</i>	Phookamsak et al. (2015)
<i>P. longicolla</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Pseudoastrospiraeriellaceae</i>	Phookamsak et al. (2015)
<i>P. papillata</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Pseudoastrospiraeriellaceae</i>	Phookamsak et al. (2015)
<i>P. thailandensis</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Pseudoastrospiraeriellaceae</i>	Phookamsak et al. (2015)
<i>Pseudotachnella yakushimensis</i>	<i>Pleiotblastus</i> sp.	On dead culms	Japan	Ascomycota incertae sedis	Sato et al. (2008)
<i>Pseudotetraploa curviappendiculata</i>	<i>Sasa kurilensis</i> ; <i>Sasa senanensis</i>	On dead culms	Japan	<i>Pleosporales/Tetraplospiraeraceae</i>	Hatakeyama et al. (2005); Tanaka et al. (2009)
<i>P. javanica</i>	<i>Sasa</i> sp.; <i>Phyllostachys bambusoides</i>	On dead culms	Japan	<i>Pleosporales/Tetraplospiraeraceae</i>	Hatakeyama et al. (2005); Tanaka et al. 2009
<i>P. longissima</i>	<i>Pleiotblastus chino</i>	On dead culms	Japan	<i>Pleosporales/Tetraplospiraeraceae</i>	Hatakeyama et al. (2005); Tanaka et al. 2009
<i>Psiloglonium sasicola</i>	Bamboo	On dead culms	Thailand	<i>Hysteriales/Hysteriaceae</i>	Liu et al. (2015)
<i>Pteridiospora chiangraiensis</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Astrospiraeriellaceae</i>	Phookamsak et al. (2015)
<i>Pteridiospora javanica</i>	Bamboo	On dead culms	Thailand, Indonesia	<i>Pleosporales/Astrospiraeriellaceae</i>	Phookamsak et al. (2014)
<i>Pustulomyces bambusicola</i>	Bamboo	On dead culms	Thailand	<i>Diaporiales/Diaportheaceae</i>	Dai et al. (2014a)
<i>Quadriclura bicornis</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Pleosporales/Tetraplospiraeraceae</i>	Tanaka et al. (2009)
<i>Q. meridionalis</i>	Bamboo	On dead culms	Japan	<i>Pleosporales/Tetraplospiraeraceae</i>	Tanaka et al. (2009)
<i>Q. septentrionalis</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Pleosporales/Tetraplospiraeraceae</i>	Tanaka et al. (2009)
<i>Rehmidothis bambusae</i>	<i>Bambusa vulgaris</i>	On dead culms	India	<i>Phyllachorales/Phyllachoraceae</i>	Verma et al. (2008)
<i>Roussoella angustior</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Roussoellaceae</i>	Ariyawansa et al. (2015)
<i>R. chiangraina</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Roussoellaceae</i>	Liu et al. (2014)
<i>R. intermedia</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	<i>Pleosporales/Roussoellaceae</i>	Liu et al. (2014)
<i>R. japonensis</i>	<i>Sasa veitchii</i> var. <i>veitchii</i>	On dead culms	Japan	<i>Pleosporales/Roussoellaceae</i>	Liu et al. (2014)
<i>R. magnatum</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Roussoellaceae</i>	Ariyawansa et al. (2015)
<i>R. mukdahanensis</i>	Bamboo	On dead culms	Thailand	<i>Pleosporales/Roussoellaceae</i>	This study



Table 2 (continued)

Taxa	Bamboo host	Substrate	Country	Order/Family	References
<i>R. neopustulans</i>	Bamboo	On dead culms	Thailand	Pleosporales/Rousoellaceae	Liu et al. (2014); This study
<i>R. nitidula</i>	Bamboo	On dead culms	Malaysia/Thailand	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>R. pseudohysterioides</i>	Bamboo	On dead culms	Thailand	Pleosporales/Rousoellaceae	This study
<i>R. pustulans</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>R. pustulata</i>	Bamboo	On dead culms	Thailand	Pleosporales/Rousoellaceae	This study
<i>R. scabrisspora</i>	Bamboo	On dead culms	Thailand	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>R. siamensis</i>	<i>Bambusa</i> sp.	On dead culms	Thailand	Pleosporales/Rousoellaceae	Liu et al. (2014); This study
<i>R. thailandica</i>	Bamboo	On dead culms	Thailand	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>R. verrucispora</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>Rousoellopsis japonica</i>	<i>Phyllostachys bambusoides</i>	On dead culms	Japan	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>R. macrospora</i>	Bamboo	On dead culms	Thailand	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>Rousoellopsis</i> sp.	<i>Sasa kurilensis</i>	On dead culms	Japan	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>R. tosaensis</i>	Bamboo	On dead culms	Japan	Pleosporales/Rousoellaceae	Liu et al. (2014)
<i>Seriascoma didymospora</i>	Bamboo	On dead culms	Thailand	Pleosporales/Occultibambusaceae	This study
<i>Sphaerulina bambusicola</i>	<i>Bambusa</i> sp.	On dead culms	China	Capnodiales/Mycosphaerellaceae	Zhao and Zhao (2012)
<i>Tetraploa</i> sp.	Bamboo	On dead culms	Japan	Pleosporales/Tetraplophaeriaceae	Tanaka et al. (2009)
<i>Tetraplophaeria nagasakiensis</i>	Bamboo	On dead culms	Japan	Pleosporales/Tetraplophaeriaceae	Tanaka et al. (2009)
<i>T. sasicola</i>	<i>Sasa senanensis</i>	On dead culms	Japan	Pleosporales/Tetraplophaeriaceae	Tanaka et al. (2009)
<i>Triplophaeria acuta</i>	<i>Sasa nipponica</i> ?	On dead culms	Japan	Pleosporales/Tetraplophaeriaceae	Tanaka et al. (2009)
<i>T. cylindrica</i>	<i>Sasa kurilensis</i>	On dead culms	Japan	Pleosporales/Tetraplophaeriaceae	Tanaka et al. (2009)
<i>T. a maxima</i>	<i>Sasa kurikensis</i>	On dead culms	Japan	Pleosporales/Tetraplophaeriaceae	Tanaka et al. (2009)
<i>Triplophaeria</i> sp.	<i>Sasa kurilensis</i>	On dead culms	Japan	Pleosporales/Tetraplophaeriaceae	Tanaka et al. (2009)
<i>T. yezoensis</i>	<i>Sasa palmata</i>	On dead culms	Japan	Pleosporales/Tetraplophaeriaceae	Tanaka et al. (2009)
<i>Tubeufia javanica</i>	Bamboo	On dead sheath	Thailand	Tubeufiales/Tubeufiaceae	Boonmee et al. (2014); This study
<i>T. longiseta</i>	Bamboo	On dead culms	Thailand	Tubeufiales/Tubeufiaceae	This study
<i>Vamsapriya bambusicola</i>	Bamboo	On dead culms	Thailand	Xylariales/Xylariaceae	Dai et al. (2014c); This study
<i>V. indica</i>	Bamboo	On dead culms	Thailand, India	Xylariales/Xylariaceae	Dai et al. (2014c); Gawas and Bhat (2005)
<i>V. khunkonensis</i>	Bamboo	On dead culms	Thailand	Xylariales/Xylariaceae	Dai et al. (2014c)
<i>Versicolorisporium triseptatum borealis</i>	<i>Pleioblastus chino</i> ; <i>Sasamorpha borealis</i>	On dead culms	Japan	Pleosporales/Occultibambusaceae	Hatakeyama et al. (2008)
<i>Xylaria bambusicola</i>	Bamboo	On dead culms	Thailand	Xylariales/Xylariaceae	This study

*Vasariaceae*) are *Dothideomycetes*. Twenty three fungal species, belonging to 16 genera (*Anthostomella*, *Arthrinium*, *Astrocystis*, *Cercophora*, *Daldinia*, *Eutypa*, *Hypoxylon*, *Leptospora*, *Myrothecium*, *Nectria*, *Neoanthostomella*, *Peroneutypa*, *Phaeoacremonium*, *Pleurophragmium*, *Vamsapriya* and *Xylaria*), of seven families (*Apiosporaceae*, *Diatrypaceae*, *Lasiosphaeriaceae*, *Nectriaceae*, *Stachybotryaceae*, *Togniniaceae* and *Xylariaceae*) and of one genus, *incertae sedis* belongs to *Sordariomycetes*.

Based on our research, it was observed that most fungi on bamboo are not pathogens. Only *Mendoglia* species are epiphytic on living bamboo culms whereas the other 42 species are saprobic, and it is rare to find diseased bamboo in Thailand. Fungi associated with bamboo are also interesting. For example, *Phaeoacremonium sphinctrophorum* was originally reported as human skin disease (subcutaneous cyst), diagnosed in North America (Mostert et al. 2006). However, we found it growing on a dead bamboo culm in Thailand. According to the literature cited in this paper (Table 2) and based on our own research, most fungi have been recorded from bamboo culms and only a few are isolated from leaves and sheaths. However, fungi occurring on shoots, roots, or inflorescences were not covered in this study. In this paper, we linked sexual and asexual morphs of *Arthrinium*, *Bambusicola*, *Neodeightonia*, *Phaeoacremonium*, *Occultabambusa*, *Rousoella* and *Vamsapriya* by culture sporulation and molecular data. Thus, this work contributes to the effort to designate one name for each fungus (Hawksworth 2012; Wingfield et al. 2012).

Hitherto information on the association of fungi with bamboo substrates has been incomplete (Hyde et al. 2002a). More than 1400 fungal species from bamboo have been recorded (Hyde et al. 2002a), however, very few taxa have molecular data in GenBank. Therefore, the existing species need to be recollected, epitypified and sequenced. Hyde et al. (2002a) stated that most fungal species on bamboo are recorded from temperate regions, while, more genera of bamboo occur in tropical regions. Thus we believe that there are numerous taxa of bambusicolous fungi waiting for recognition and description in tropical areas. A list of bambusicolous fungi published since Hyde et al. (2002a) is provided here.

**Acknowledgments** This work was supported by the Key Laboratory of Yunnan Province Universities of the Diversity and Ecological Adaptive Evolution for Animals and plants on YunGui Plateau. Dong Qin Dai, Nalin N. Wijayawardene and Wen Jing Li thank to Mushroom Research Foundation (MRF), Chiang Rai Province, Thailand for providing Postgraduate Scholarships. Mae Fah Luang University, Chiang Rai Province, Thailand is acknowledged for the financial support to Dong Qin Dai. Kevin D. Hyde is grateful to the Chinese Academy of Sciences, project number 2013T2S0030, for the award of Visiting Professorship for Senior International Scientists at Kunming Institute of Botany, research grant from the Biodiversity Research and Training Program (BRT R253012) and The Thailand Research Fund (BRG 5280002). We would like to thank Jun Bo Yang, Germplasm Bank of

Wild Species in Southwest China, Kunming Institute of Botany, Chinese Academy of Science, Kunming 650201, Yunnan, China and Molecular Biology Center in Germplasm Bank of Wild Species, for the help of molecular work. Rungtiwa Phookamsak expresses sincere appreciations to The Royal Golden Jubilee Ph. D. Program (PHD/0090/2551) under the Thailand Research Fund for financial support. Dong Qin Dai is grateful to Alan J.L. Phillips and Eric H.C. McKenzie for their valuable suggestions.

## References

- Abarca GH, Ruiz RC, Arias RM, Saikawa M, Stadler M (2007) Anamorphic fungi from submerged plant material: *Acumispora verruculosa*, *Pleurophragmium aquaticum* and *P. miniumbonatum*. *Mycotaxon* 101:89–97
- Acero FJ, González V, Sánchez-Ballesteros J, Rubio V, Checa J, Bills GF, Salazar O, Platas G, Peláez F (2004) Molecular phylogenetic studies on the *Diatrypaceae* based on rDNA-ITS sequences. *Mycologia* 96: 249–259
- Adamčík S, Cai L, Chakraborty D, Chen XH, Cotter HVT, Dai DQ, Dai YC, Das K, Deng CY, Ghobad-Nejhad M (2015) Fungal biodiversity profiles 1–10. *Cryptogam Mycol* 36:121–166
- Alias SA, Jones EBG, Kuthubutheen AJ (1994) *Fasciatispora lignicola* sp. nov. (*Amphisphaeriaceae*, Ascomycotina) from drift mangrove wood. *Mycotaxon* 52:77–82
- Aptroot A (1995) A monograph of *Didymosphaeria*. *Stud Mycol* 37:1–161
- Ariyawansa HA, Tanaka K, Thambugala KM, Phookamsak R, Tian Q, Camporesi E, Hongsanan S, Monkai J, Wanasinghe DN, Mapook A (2014) A molecular phylogenetic reappraisal of the *Didymosphaeriaceae* (= *Montagnulaceae*). *Fungal Divers* 68:69–104
- Ariyawansa HA, Hyde KD, Jayasiri SC, Buyck B, Chethana KWT, Dai DQ, Dai YC, Daranagama DA, Jayawardena RSLücking R, Ghobad-Nejhad M, Niskanen T, Thambugala KM, Voigt K, Zhao RL, Li GJ, Doilom M, Boonmee S, Yang ZL, Cai Q, Cui YY, Bahkali AH, Chen J, Cui BK, Chen YY, Monika CD, Dissanayake AJ, Ekanayaka AH, Hashimoto A, Hongsanan S, Jones EBG, Larsson E, Li WJ, Li QR, Liu JK, Luo ZL, Maharachchikumbura SSN, Mapook A, McKenzie EHC, Norphanphou C, Konta S, Pang KL, Perera RH, Phookamsak R, Phukhamsakda C, Pinruan U, Randrianjohany E, Singtripop C, Tanaka K, Tian CM, Tibpromma S, Abdel-Wahab MA, Wanasinghe DN, Wijayawardene NN, Zhang JF, Zhang H, Abdel-Aziz FA, Wedin M, Westberg M, Ammirati JF, Bulgakov TS, Lima DX, Callaghan TM, Callac P, Chang CH, Coca LF, Dal-Forno M, Dollhofer V, Fliegerová K, Greiner K, Griffith GW, Ho HM, Hofstetter V, Jeewon R, Kang JC, Wen TC, Kirk PM, Kytövuori I, Lawrey JD, Xing J, Li H, Liu ZY, Liu XZ, Liimatainen K, Lumbsch HT, Matsumura M, Moncada B, Moncada S, Parnmen S, de Azevedo Santiago ALCM, Sommai S, Song Y, de Souza CAF, de Souza-Motta CM, Su HY, Suetrong S, Wang Y, Wei SF, Yuan HS, Zhou LW, Réblová M, Fournier J, Camporesi E, Luangsa-ard JJ, Tazanathai K, Khonsanit A, Thanakitpipattana D, Somrithipol S, Diederich P, Millanes AM, Common RS, Stadler M, Yan JY, Li XH, Lee HW, Nguyen TTT, Lee HB, Battistin E, Marsico O, Vizzini A, Vila J, Ercole E, Eberhardt U, Simonini G, Wen HA, Chen XH (2015) Fungal diversity notes 111–252—taxonomic and phylogenetic contributions to fungal taxa. *Fungal Divers*:27–274
- Bahl J (2006) Molecular evolution of three morphologically similar families in the Xylariomycetidae (*Apiosporaceae*, *Clypeosphaeriaceae*, *Hyponectriaceae*). The University of Hong Kong (Pokfulam, Hong Kong), <http://hdl.handle.net/10722/51007>

- Bahl J, Jeewon R, Hyde KD (2005) Phylogeny of *Rosellinia captribulensis* sp. nov. and its allies (*Xylariaceae*). *Mycologia* 97: 1102–1110
- Bamboo Botanicals (2016) <http://www.bamboobotanicals.ca/index.html>
- Barr ME (1976) *Buergenerula* and the *Physosporrellaceae*. *Mycologia* 68: 611–621
- Barr ME (1987) Prodomus to class loculoascomycetes. Publ. by the author, Amherst
- Barr ME (1990) Prodomustononlichenized, pyrenomycetous members of the class Hymenoascomycetes. *Mycotaxon* 39:43–184
- Barr ME, Cannon PF (1994) *Calosphaeriales*, *Clavicipitales*, *Coryneliales*, *Diaporthales*, *Diatrypales*, *Halosphaeriales*, *Hypocreales*, *Meliolales*, *Ophiostomatales*, *Phyllachorales*, *Sordariales*, *Trichosphaeriales*, and *Xylariales*. In: Hawksworth DL (ed) *Ascomycetes systematics. Problems and perspectives in the nineties*. Nato ASI Series. Series a: life sciences, vol 269. Plenum Press, New York and London, pp 371–378
- Berkeley MJ, Broome CE (1874) Enumeration of the fungi of Ceylon. Part II. *Bot J Linn Soc* 14:29–141
- Berkeley MJ, Curtis MA (1853) Exotic fungi from the Schweinitzian Herbarium, principally from Surinam. *J Acad Natl Sci Phila* 2: 277–294, N.S
- Berlese (1902) *Icon. Fung* 3:80
- Berlese AN (1968) *Icones fungorum omnium hucusque cognitorum*. (1900–1905). Vol. I. *Bibliotheca Mycologica* 16 A: 1–243 pp. J. Cramer reprint
- Boonmee S, Zhang Y, Chomnunti P, Chukeatirote E, Tsui CKM, Bahkali AH, Hyde KD (2011) Revision of lignicolous *Tubeufiaceae* based on morphological reexamination and phylogenetic analysis. *Fungal Divers* 51:63–102
- Boonmee S, Rossman AY, Liu JK, Li WJ, Dai DQ, Bhat DJ, Jones EBG, McKenzie EHC, Xu JC, Hyde KD (2014) *Tubeufiales*, ord. nov., integrating sexual and asexual generic names. *Fungal Divers* 68: 239–298
- Bystriakova N, Kapos V, Lysenko I, Stapleton CMA (2003) Distribution and conservation status of forest bamboo biodiversity in the Asia-Pacific Region. *Biodivers Conserv* 12:1833–1841
- Cai L, Zhang K, McKenzie EHC, Hyde KD (2004) *Linocarpon bambusicola* sp. nov. and *Dictyochaeta curvispora* sp. nov. from bamboo submerged in freshwater. *Nova Hed* 78(3–4):439–445
- Cai L, Jeewon R, Hyde KD (2006) Molecular systematics of *Zopfiella* and allied genera: evidence from multi-gene sequence analyses. *Mycol Res* 110:359–368
- Carmaràn CC, Romero AI, Giussani LM (2006) An approach towards a new phylogenetic classification in *Diatrypaceae*. *Fungal Divers* 23: 67–87
- Castlebury LA, Rossman AY, Sung G-H, Hyten AS, Spatafora JW (2004) Multigene phylogeny reveals new lineages for *Stachybotrys chartarum*, the indoor air fungus. *Mycol Res* 108:864–872
- Cesati V, De Notaris G (1863) Schema di classificazione degle sferiacei italici aschigeri piu' o meno appartenenti al genere Sphaeria nell'antico significato attribuitoglide Persono. *Comm Soc Crittog Ital* 1(4):177–420
- Chardón CE (1939) *Bol Soc Venez Ci Nat* 5(40):335 (235)–368 (268)
- Chareprasert S, Abdelghany MT, El-sheikh HH, Ahmed AF, Khalil AMA, Sharples GP, Sihanonth P, Soliman HG, Suwannasai N, Whalley AS (2012) *Xylariaceae* on the fringe. *Prog Mol Subcell Biol* 53:229–241
- Chaverri P, Salgado C, Hirooka Y, Rossman A, Samuels G (2011) Delimitation of *Neonectria* and *Cylindrocarpon* (*Nectriaceae*, *Hypocreales*, *Ascomycota*) and related genera with *Cylindrocarpon*-like anamorphs. *Stud Mycol* 68:35–56
- Chen Y, Ran SF, Dai DQ, Wang Y, Hyde KD, Wu YM, Jiang YL (2016) *Mycosphere Essays* 2. *Myrothecium*. *Mycosphere* 7:64–80
- Cooke MC (1879) *New Zealand fungi*. *Grevillea* 8(46):54–68
- Cooke MC (1883) *Hypoxylon* and its allies. *Grevillea* 11(60):121–140
- Costantin J (1888) *Les mucédinées simples*. Librairie Paul Klincksieck, Paris, France
- Crous PW, Groenewald JZ (2013) A phylogenetic re-evaluation of *Arthrinium*. *IMA Fungus* 4:133–154
- Crous PW, Gams W, Wingfield MJ, van Wyk PS (1996) *Phaeoacremonium* gen. nov. associated with wilt and decline diseases of woody hosts and human infections. *Mycologia* 88:786–796
- Crous PW, Slippers B, Wingfield MJ, Rheeder J, Marasas WFO, Philips AJL, Alves A, Burgess T, Barber P, Groenewald JZ (2006a) Phylogenetic lineages in the *Botryosphaeriaceae*. *Stud Mycol* 55: 235–253
- Crous PW, Groenewald JZ, Wingfield MJ (2006a) *Anthostomella eucalyptorum*. *Fungal Planet*, no. 1
- Crous PW, Shivas RG, Quaedvlieg W, van der Bank M, Zhang Y, Summerell BA, Guarro J, Wingfield MJ, Wood AR, Alfenas AC, Braun U, Cano-Lira JF, García D, Marin-Felix Y, Alvarado P, Andrade JP, Armengol J, Assefá A, den Breeÿen A, Camele I, Chewangkoon R, De Souza JT, Duong TA, Esteve-Raventós F, Fournier J, Frisullo S, García-Jiménez J, Gardinnet A, Gené J, Hernández-Restrepo M, Hirooka Y, Hospenthal DR, King A, Lechat C, Lombard L, Mang SM, Marbach PAS, Marinowitz S, Marin-Felix Y, Montañón-Mata NJ, Moreno G, Perez CA, Pérez Sierra AM, Robertson JL, Roux J, Rubio E, Schumacher RK, Stchigel AM, Sutton DA, Tan YP, Thompson EH, van der Linde E, Walker AK, Walker DM, Wickes BL, Wong PTW, Groenewald JZ (2014a) *Fungal planet description sheets*: 214–280. *Persoonia* 32: 184–306
- Crous PW, Wingfield MJ, Schumacher RK, Summerell BA, Giraldo A, Gené J, Guarro J, Wanasinghe DN, Hyde KD, Camporesi E, Jones EBG, Thambugala KM, Malysheva EF, Malysheva VF, Acharya K, Álvarez J, Alvarado P, Assefá A, Barnes CW, Bartlett JS, Blanchette RA, Burgess TI, Carlavilla JR, Coetzee MPA, Damm U, Decock CA, den Breeÿen A, de Vries B, Dutta AK, Holdom DG, Rooney-Latham S, Manjón JL, Marinowitz S, Mirabolfathy M, Moreno G, Romberg MK, Shivas RG, Stalpers JA, Stielow B, Stukely MJC, Swart WJ, Tan YP, van der Bank M, Wood AR, Zhang Y (2014b) *Fungal planet description sheets*: 281–319. *Persoonia* 33:212–289
- Crous PW, Müller MM, Sánchez RM, Giordano L, Nchinotti MV, Anderson FE, Groenewald JZ (2015) Resolving *Tiarospora* spp. allied to *Botryosphaeriaceae* and *Phacidiaceae*. *Phytotaxa* 202: 073–093
- Cruz KS, Cortez VG (2015) *Hypoxylon* (*Xylariaceae*, *Ascomycota*) from Western Paraná, Brazil. *Braz J Bot*:889–901
- D'Souza MA, Bhat DJ (2012) A new species of *Pleurophragmium* from India. *Mycotaxon* 119:477–482
- Dai DQ, Bhat DJ, Liu JK, Chukeatirote E, Zhao RL, Hyde KD (2012) *Bambusicola*, a new genus from bamboo with asexual and sexual morphs. *Cryptogam Mycol* 33:363–379
- Dai DQ, Wijayawardene NN, Bhat DJ, Chukeatirote E, Bahkali AH, Zhao RL, Xu JC, Hyde KD (2014a) *Pustulomyces* gen. nov. accommodated in *Diaporthaceae*, *Diaporthales*, as revealed by morphology and molecular analyses. *Cryptogam Mycol* 35:63–72
- Dai DQ, Wijayawardene NN, Bhat DJ, Chukeatirote E, Zhao RL, Wang Y, Bahkali AH, Hyde KD (2014b) The phylogenetic placement of *Eriosporella bambusicola* sp. nov. in *Capnodiales*. *Cryptogam Mycol* 35:41–49
- Dai DQ, Bahkali AH, Li QR, Bhat DJ, Wijayawardene NN, Li WJ, Chukeatirote E, Zhao RL, Xu JC, Hyde KD (2014c) *Vamsapriya* (*Xylariaceae*) re-described, with two new species and molecular sequence data. *Cryptogam Mycol* 35:339–357
- Dai DQ, Bahkali AH, Li WJ, Bhat DJ, Zhao RL, Hyde KD (2015) *Bambusicola loculata* sp. nov. (*Bambusicolaceae*) from bamboo. *Phytotaxa* 213:122–130
- Dai DQ, Bahkali AH, Ariyawansa HA, Li WJ, Bhat JD, Zhao RL, Mortimer PE, Xu JC, Hyde KD (2016) *Neokalmusia didymospora* sp. nov. (*Didymosphaeriaceae*) from bamboo. *Sydowia* (accepted)

- Daranagama DA, Camporesi E, Tian Q, Liu XZ, Chamyuang S, Stadler M, Hyde KD (2015) *Anthostomella* is polyphyletic comprising several genera in *Xylariaceae*. Fungal Divers:203–238
- De Hoog GS (1985) Taxonomy of the *Dactylaria* complex. IV. *Dactylaria*, *Neta*, *Subulispora* and *Scolecobasidium*. Stud Mycol 26:1–60
- DeScenzo RA, Engel SR, Gomez G, Jackson EL, Munkvold GP, Weller J, Irelan NA (1999) Genetic analysis of *Eutypa* strains from California supports the presence of two pathogenic species. Phytopathology 89:884–893
- Dharmananda S (2004) Bamboo as medicine, Institute for Traditional Medicine, Portland, Oregon, ITM Online
- DiCosmo F, Michaelides J, Kendrick B (1980) *Myrothecium tongaense* anam. - sp. nov. Mycotaxon 12:219–224
- Dissanayake AJ, Jayawardena RS, Boonmee S, Thambugala KM, Senanayake IC, Tain Q, Mapook A, Yan JY, Li YM, Li XH, Chukeatirote E, Hyde KD (2014) The status of family *Myriangiaceae* (*Dothideomycete*). Phytotaxa 176:219–237
- Doilom M, Dissanayake AJ, Phillips AJL, Boonmee S, Liu JK, Bhat DJ, Taylor JE, Bahkali AH, McKenzie EHC, Hyde KD (2016) Microfungi on *Tectona grandis* (teak) in Northern Thailand. Fungal Divers, accepted
- Doveri F (2008) A bibliography of *Podospora* and *Schizothecium*, a key to the species, and a description of *Podospora dasypogon* newly recorded from Italy. Pagine di Micologia 29:61–159
- Dransfield S, Widjaja EA (1995) Bamboos. Backhuys, Leiden (Netherlands)
- Dubey R, Neelima A (2014) *Goosiomycetes bambusicola*-A new cheirosporous anamorphic species from Western Ghats, India. Curr Res Environ Appl Mycol 4:211–216
- Edward JC, Singh KP, Tripathi SC, Sinha MK, Ranade K (1972) Fungi associated with moribund branches of *Rosa* species. Sydowia 26(1–6):266–271
- Eriksson O, Yue JZ (1998) Bambusicolous pyrenomycetes, an annotated check-list. Myconet 1(2):25–78
- Essakhi S, Mugnai L, Crous PW, Groenewald JZ, Surico G (2008) Molecular and phenotypic characterisation of novel *Phaeoacremonium* species isolated from esca diseased grapevines. Persoonia 21:119–134
- Faurel L, Schotter G (1965) Notes mycologiques. IV. *Champignons coprophiles* du Sahara central et notamment de la Tefedest. Revue Mycol 30(3):141–165
- Francis SM (1975) *Anthostomella* Sacc. (Part 1). Mycol Pap 139:1–97
- Fuckel L (1870) Symbolae mycologicae. Beiträge zur Kenntniss der Rheinischen Pilze. Jb Nassau Ver Naturk 23–24:1–459
- Gawas P, Bhat DJ (2005) *Vamsapriya indica* gen. et sp. nov., a bambusicolous, synnematosus fungus from India. Mycotaxon 94:149–154
- Graham AB, Johnston PR, Weir BS (2009) Three new *Phaeoacremonium* species on grapevines in New Zealand. Australas Plant Pathol 38(5):505–513
- Gramaje D, Mostert L, Groenewald JZ, Crous PW (2015) *Phaeoacremonium*: from esca disease to *phaeohyphomycosis*. Fungal Biol 119:759–783
- Gratani L, Crescente MF, Varone L, Fabrini G, Digiulio E (2008) Growth pattern and photosynthetic activity of different bamboo species growing in the Botanical Garden of Rome. Flora-Morphol Distrib Funct Ecol Plants 203:77–84
- Hall T (2004) BioEdit. Ibis Therapeutics, Carlsbad, CA, 92008, USA. (<http://www.mbio.ncsu.edu/BioEdit/bioedit.html>) (18 Mar 2005)
- Hansford CG (1957) Australian Fungi. IV. New species and revisions (cont'd). Proc Linnean Soc NSW 82:209–229
- Hashimoto A, Sato G, Matsuda T, Hirayama K, Hatakeyama S, Harada Y, Shirouzu T, Tanaka K (2015) Molecular taxonomy of *Dinemasporium* and its allied genera. Mycoscience 56:86–101
- Hatakeyama S, Tanaka K, Harada Y (2005) Bambusicolous fungi in Japan (5): three species of *Tetraploa*. Mycoscience 46:196–200
- Hatakeyama S, Tanaka K, Harada Y (2008) Bambusicolous fungi in Japan (7): a new coelomycetous genus, *Versicolorisporium*. Mycoscience 49:211–214
- Hawksworth DL (2012) Managing and coping with names of pleomorphic fungi in a period of transition. Mycosphere 3(2):143–155; IMA Fungus 3:15–24
- Hawksworth DL, Eriksson OE (1986) The names of accepted orders of Ascomycetes. Syst Ascom 5:175–184
- Hawksworth DL, Kirk PM, Sutton BC, Pegler DN (1995) Ainsworth & Bisby's Dictionary of the Fungi, 8th edn. CABI International, Wallingford
- Hennings P (1904) Fungi Amazonici a cl. Ernesto Ule collecti II. Hedwigia 43:242–273
- Hidayat I, Meeboon J, To-Anun C (2007) *Anthostomella* and *Fasciatispora* species (*Xylariaceae*) from palms including *F. ujungkulonensis* sp. nov. Mycotaxon 102:347–354
- Hino I, Katumoto K (1960) Icones fungorum bambusicolorum japonicorum. The Fuji Bamboo Garden, Kobe
- Hirooka Y, Rossmann AY, Samuels GJ, Lechat C, Chaverri P (2012) A monograph of *Allantonectria*, *Nectria*, and *Pleonectria* (*Nectriaceae*, *Hypocreales*, *Ascomycota*) and their pycnidial, sporodochial, and synnematosus anamorphs. Stud Mycol 71:1–210
- Hsieh HM, Ju YM, Rogers JD (2005) Molecular phylogeny of *Hypoxylon* and closely related genera. Mycologia 97:844–865
- Hsieh HM, Lin CR, Fang MJ, Rogers JD, Fournier J, Lechat C, Ju YM (2010) Phylogenetic status of *Xylaria* subgenus *Pseudoxylaria* among taxa of the subfamily *Xylarioideae* (*Xylariaceae*) and phylogeny of the taxa involved in the subfamily. Mol Phylogenet Evol 54:957–969
- Hudson J, Zhou J, Chen J, Harris L, Yip L, Tpwewer GHN (1994) Hypocrellin, from *Hypocrella bambuase*, is phototoxic to human immunodeficiency virus. Photochem Photobiol 60:253–255
- Huelsenbeck JP, Ronquist F (2001) MRBAYES: Bayesian inference of phylogenetic trees. Bioinformatics 17:754–755
- Huhndorf SM, Miller AN (2011) A molecular re-appraisal of taxa in the Sordariomycetidae and a new species of *Rimaconus* from New Zealand. Stud Mycol 68:203–210
- Huhndorf SM, Miller AN, Fernández FA (2004) Molecular systematics of the *Sordariales*: the order and the family *Lasiosphaeriaceae* re-define. Mycologia 96:368–387
- Hyde KD (1991) A new amphispheariaceous fungus from intertidal fronds of *Nypa fruticans*. Trans Mycol Soc Jpn 32:265–271
- Hyde KD (1994) Fungi from rachides of *Livistona* in the western province of Papua New Guinea. Bot J Linn Soc 116:315–324
- Hyde KD (1995) Fungi from palms XVII. The genus *Fasciatispora*, with notes on *Amphisphaerella*. Nova Hed 61(1–2):249–268
- Hyde KD (1996) Fungi from palms. XXVI. The genus *Anthostomella*, with ten new species. Nova Hedwigia 62:273–340
- Hyde KD (1997) The genus *Rousoëlla*, including two new species from palms in Cuyabeno, Ecuador. Mycol Res 101:609–616
- Hyde KD, Eriksson OE, Yue JZ (1996) *Rousoëlla*, an ascomycete genus of uncertain relationships with a *Cytoplea* anamorph. Mycol Res 100:1522–1528
- Hyde KD, Fröhlich J, Taylor JE (1998) Fungi from palms. XXXVI. Reflections on unitunicate ascomycetes with apiospores. Sydowia 50:21–80
- Hyde KD, Zhou D, McKenzie E, Ho W, Dalisay T (2002a) Vertical distribution of saprobic fungi on bamboo culms. Fungal Divers 11:109–118
- Hyde KD, Zhou D, Dalisay T (2002b) Bambusicolous fungi: a review. Fungal Divers 9:1–14
- Hyde KD, Jones EBG, Liu JK, Ariyawansa H, Boehm E, Boonmee S, Braun U, Chomnunti P, Crous PW, Dai DQ, Diederich P, Dissanayake A, Doilom M, Doveri F, Hongsanan S, Jayawardena R,

- Lawrey JD, Li YM, Liu YX, Lücking R, Monkai J, Muggia L, Nelsen MP, Pang KL, Phookamsak R, Senanayake I, Shearer CA, Suetrong S, Tanaka K, Thambugala KM, Wijayawardene NN, Wikee S, Wu HX, Zhang Y, Aguirre-Hudson B, Alias SA, Aptroot A, Bahkali AH, Bezerra JL, Bhat DJ, Camporesi E, Chukeatirote E, Gueidan C, Hawksworth DL, Hirayama K, Hoog SD, Kang JC, Knudsen K, Li WJ, Li XH, Liu ZY, Mapook A, McKenzie EHC, Miller AN, Mortimer PE, Phillips AJL, Raja HA, Scheuer C, Schumm F, Taylor JE, Tian Q, Tibpromma S, Wanasinghe DN, Wang Y, Xu JC, Yan JY, Yacharoen S, Zhang M (2013) Families of *Dothideomycetes*. *Fungal Divers* 63:1–313
- Hyde KD, Nilsson RH, Alias SA, Ariyawansa HA, Blair JE, Cai L, de Cock AWAM, Dissanayake AJ, Glockling SL, Goonasekara ID, Gorczak M, Hahn M, Jayawardena RS, van Kan JAL, Laurence MH, Lévesque CA, Li XH, Liu JK, Maharachchikumbura SSN, Manamgoda DS, Martin FN, McKenzie EHC, McTaggart AR, Mortimer PE, Nair PVR, Pawłowska NJ, Rintoul TL, Shivas RG, Spies CFJ, Summerell BA, Taylor PWJ, Terhem RB, Udayanga D, Vaghefi N, Walther G, Wilk M, Wrzosek M, Xu JC, Yan JY, Zhou N (2014) One stop shop: backbone trees for important phytopathogenic genera: I (2014). *Fungal Divers* 67:21–125
- Idris MAM, Mohamad A (2002) Bamboo shoot utilization in peninsular Malaysia: a case study in Pahang. *J Bamboo Rattan* 1:141–155
- INBAR (2016) International Network for Bamboo and Rattan, <http://www.inbar.int/>
- Index Fungorum (2016) <http://www.indexfungorum.org/Names/Names.asp>
- Jaklitsch WM, Voglmayr H (2012) Phylogenetic relationships of five genera of *Xylariales* and *Rosasphaeria* gen. nov. (*Hypocreales*). *Fungal Divers* 52:75–98
- Jaklitsch WM, Fournier J, Dai DQ, Hyde KD, Voglmayr H (2015) *Valsaria* and the *Valsariales*. *Fungal Divers* 73:159–202
- Janssen JJ (1991) Mechanical properties of bamboo. Kluwer Academic Publishers
- Jayasiri SC, Hyde KD, Ariyawansa HA, Bhat DJ, Buyck B, Cai L, Dai YC, Abd-Elsalam KA, Ertz D, Hidayat I, Jeewon R, Jones EBG, Bahkali AH, Karunarathna SC, Liu JK, Luangsa-ard JJ, Lumbsch HT, Maharachchikumbura SSN, McKenzie EHC, Moncalvo JM, Ghobad-Nejhad M, Nilsson H, Pang KL, Pereira OL, Phillips AJL, Raspé O, Rollins AW, Romero AI, Etayo J, Selçuk F, Stephenson SL, Suetrong S, Taylor JE, Tsui CKM, Vizzini A, Abdel-Wahab MA, Wen TC, Boonmee S, Dai DQ, Daranagama DA, Dissanayake AJ, Ekanayaka AH, Fryar SC, Hongsanan S, Jayawardena RS, Li WJ, Perera RH, Phookamsak R, de Silva NI, Thambugala KM, Tian Q, Wijayawardene NN, Zhao RL, Zhao Q, Kang JC, Promputtha I (2015) The Faces of Fungi database: fungal names linked with morphology, phylogeny and human impacts. *Fungal Divers* 74:3–18
- Jiang SX, Zhang QS, Jiang SH (2002) On Structure, production, and market of bamboo-based panels in China. *J For Res* 13:151–156
- Jiang YL, Wang HF, Pan HQ, Zhang TY (2014) *Myrothecium* (*Hyphomycetes*): three new species, one new variety and a key to species and varieties of the genus known from soils in China. *Mycosystema* 33:7–14
- Ju YM, Rogers JD (1990) *Astrocystis* reconsidered. *Mycologia*: 342–349
- Ju YM, Rogers JD (1996) A revision of the genus *Hypoxylon*. *Mycologia* Memoir no.20. APS Press, St. Paul, p 365
- Ju YM, Rogers JD (1999) The *Xylariaceae* of Taiwan (excluding *Anthostomella*). *Mycotaxon* 73:343–440
- Ju YM, Tzean SS (1985) Investigations of *Xylariaceae* in Taiwan II. The teleomorph of *Xylaria*. *Trans Mycol Soc Repub China* 1:103–128
- Ju YM, Rogers JD, Huhndorf SM (1996) *Valsaria* and notes on *Endoxylina*, *Pseudothyridaria*, *Pseudovalsaria*, and *Rousoella*. *Mycotaxon* 58:419–481
- Ju YM, Rogers JD, San Martín F (1997) A revision of the genus *Daldinia*. *Mycotaxon* 61:243–293
- Ju YM, Vasilyeva L, Rogers JD (1999) *Daldinia singularis* sp. nov. from Eastern Russia and notes on some other taxa. *Mycotaxon* 71:405–412
- Ju YM, Hsieh HM, Rogers JD, Fournier J, Jaklitsch WM, Courtecuisse R (2012) New and interesting penzigioid *Xylaria* species with small, soft stromata. *Mycologia* 104:766–776
- Kamal (2010) *Cercosporoid* fungi of India. – vii + 351 p., Bishen Singh Mahendra Pal Singh Publication, Dehradun
- Kapoor JN, Gill HS (1961) Notes on Indian ascomycetes. *Indian Phytopathol* 14:149–153
- Katoh K, Standley DM (2013) MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol* 30:772–780
- Kelchner SA, Group BP (2013) Higher level phylogenetic relationships within the bamboos (*Poaceae: Bambusoideae*) based on five plastid markers. *Mol Phylogenet Evol* 67:404–413
- Kirk PM, Cannon PF, Minter DW, Stalpers JA (2008) Dictionary of the Fungi, 10th edn. CABI, Wallingford
- Kirschner R, Yang ZL, Zhao Q, Feng B (2009) *Ovipoculum album*, a new anamorph with gelatinous cupulate bulbiferous conidiomata from China and with affinities to the *Auriculariales* (*Basidiomycota*). *Fungal Divers* 43:55–65
- Kornerup A, Wanscher HH (1978) Methuen Handbook of color. London
- Krug J, Cain R (1972) Additions to the genus *Arnium*. *Can J Bot* 50:367–373
- Kruys A, Huhndorf SM, Miller AN (2015) Coprophilous contributions to the phylogeny of *Lasiosphaeriaceae* and allied taxa within *Sordariales* (*Ascomycota*, *Fungi*). *Fungal Divers* 70:101–113
- Kuhnert E, Fournier J, Peršoh D, Luangsa-ard JJ, Stadler M (2014) New *Hypoxylon* species from Martinique and new evidence on the molecular phylogeny of *Hypoxylon* based on ITS rDNA and  $\beta$ -tubulin data. *Fungal Divers* 64:181–203
- Kuhnert E, Surup F, Sir EB, Lambert C, Hyde KD, Hladki AI, Romero AI, Stadler M (2015) Lenormandins A–G, new azaphilones from *Hypoxylon lenormandii* and *Hypoxylon jaklitschii* sp. nov., recognised by chemotaxonomic data. *Fungal Divers* 71:165–184
- Læssøe T, Spooner B (1993) *Rosellinia* & *Astrocystis* (*Xylariaceae*): new species and generic concepts. *Kew Bull* 49(1):1–70
- Lee S, Crous PW (2003) New species of *Anthostomella* on fynbos from South Africa. *Mycol Res* 107:360–370
- Li XM, Gao J, Yue YD (2009) Studies on systematics, biology and bioactive substance of *Shiraia bambusicola*. *For Res Beijing* 22: 279–284
- Lin CS (2004) Bamboo culture bamboo furniture. *Furnit Inter Decor* 7:20
- Linaldeddu BT, Deidda A, Scanu B, Franceschini A, Serra S, Berraf-Tebbal A, Zouaoui Boutiti M, Ben Jamâa MJ, Phillips AJL (2015) Diversity of *Botryosphaeriaceae* species associated with grapevine and other woody hosts in Italy, Algeria and Tunisia, with descriptions of *Lasiodiplodia exigua* and *Lasiodiplodia mediterranea* sp. nov. *Fungal Divers* 71:201–214
- Liu YJ, Whelen S, Hall BD (1999) Phylogenetic relationships among ascomycetes: evidence from an RNA polymerase II subunit. *Mol Biol Evol* 16:1799–1808
- Liu JK, Phookamsak R, Jones EBG, Zhang Y, Ko-Ko TW, Hu HL, Boonmee S, Doilom M, Chukeatirote E, Bahkali AH (2011) *Astrosphaeriella* is polyphyletic, with species in *Fissuroma* gen. nov., and *Neoastrisphaeriella* gen. nov. *Fungal Divers* 51:135–154
- Liu JK, Phookamsak R, Doilom M, Wikee S, Li YM, Ariyawansa HA, Boonmee S, Chomnunti P, Dai DQ, Bhat JD, Romero AI, Zhuang WY, Monkai J, Jones EBG, Chukeatirote E, Ko-Ko WT, Zhao YC, Wang Y, Hyde KD (2012a) Towards a natural classification of *Botryosphaeriales*. *Fungal Divers* 57:149–210

- Liu Y, Liu Z, Wongkaew S (2012b) Developing characteristics and relationships of *Shiraia bambusicola* with Bamboo. *Songklanakarini J Sci Technol* 34:17–22
- Liu JK, Phookamsak R, Dai DQ, Tanaka K, Jones EBG, Xu JC, Chukeatirote E, Hyde KD (2014) *Rousoellaceae*, a new pleosporalean family to accommodate the genera *Neorousoella* gen. nov., *Rousoella* and *Rousoellopsis*. *Phytotaxa* 181:1–33
- Liu JK, Hyde KD, Gareth EBG, Ariyawansa HA, Bhat DJ, Boonmee S, Maharachchikumbura S, McKenzie EHC, Phookamsak R, Phukhamsakda R, Abdel-Wahab MA, Buyck B, Chen J, Chethana KWT, Singtripop C, Dai DQ, Dai YC, Daranagama DA, Dissanayake AJ, Doliom M, Fan LX, Goonasekara D, Hirayama K, Hongsanan S, Jayasiri SC, Jayawardena RS, Karunarathna SC, Li WJ, Mapook A, Norphanphoun C, Pang KL, Perera RH, Peršoh D, Pinruan U, Senanayake IC, Somrithipol S, Satinee S, Tanaka K, Thambugala KM, Tian Q, Tibpromma S, Udayanga D, Wijayawardene NN, Wanasinghe D, Abdel-Aziz FA, Adamčík S, Bahkali AH, Boonyuen N, Bulgakov T, Callac P, Chomnunti P, Greiner K, Hashimoto A, Hofstetter V, Kang JC, Li XH, Liu ZY, Matumura M, Mortimer PE, Rambold R, Randrianjohany E, Sato G, Indrasutdhi VS, Verbeken A, Brackel W, Wang Y, Wen TC, Xu JC, Yan JY, Zhao RL, Camporesi E (2015) Fungal diversity notes 1–110: taxonomic and phylogenetic contributions to fungal species. *Fungal Divers* 72:1–197
- Lombard L, van der Merwe NA, Groenewald JZ, Crous PW (2015) Generic concepts in *Nectriaceae*. *Stud Mycol* 80:189–245
- Lorenzo LE, Havrylenko M (2001) The genera *Armium* and *Podospora* from Argentina. *Mycologia* 93:1221–1230
- Lu BS, Hyde KD (2000) A world monograph of *Anthostomella*. *Fungal Divers Res Ser* 4:1–376
- Lumbsch HT, Huhndorf S (2007) Outline of Ascomycota. *Myconet* 13: 1–99
- Lumbsch HT, Huhndorf SM (2010) *Myconet* Volume 14. Part one. Outline of *Ascomycota*—2009. Part Two. Notes on Ascomycete Systematics. Nos. 4751–5113. *Fieldiana Life Earth Sci* 1:1–64
- Lundqvist N (1972) Nordic *Sordariaceae* s. lat. *Symb Bot Ups* 20:1–374
- Luttrell ES (1975) Centrum development in *Didymosphaeria sadasivanii* (*Pleosporales*). *Am J Bot* 62:186–190
- Ma LG, Xia JW, Ma YR, Zhang XG (2014) Three new species of *Pleurophragmium* from Yunnan, China. *Mycotaxon* 127:213–219
- Maharachchikumbura SS, Hyde KD, Jones EBG, McKenzie EHC, Huang SK, Abdel-Wahab MA, Daranagama DA, Dayaratne M, D'souza MJ, Goonasekara ID, Hongsanan S, Jayawardena RS, Kirk PM, Konta S, Liu JK, Liu ZY, Norphanphoun C, Pang KL, Perera RH, Senanayake IC, Shang Q, Shenoy BD, Xiao YP, Bahkali AH, Kang JC, Somrithipol S, Suetrong S, Wen TC, Xu JC (2015) Towards a natural classification and backbone tree for *Sordariomycetes*. *Fungal Divers* 72:199–301
- Matsushima T (1989) *Matsushima Mycological Memoirs* 6. Matsushima Fungus Collect. Kobe, Japan, pp 1–100
- Melo R, Miller A, Maia L (2015) The genus *Podospora* (*Lasiosphaeriaceae*, *Sordariales*) in Brazil. *Mycosphere* 6:201–215
- Miller JH (1938) Studies in the development of two *Myriangium* species and the systematic position of the order *Myriangiales*. *Mycologia* 30:158–181
- Miller JH (1940) The genus *Myriangium* in North America. *Mycologia* 32:587–600
- Miller AN, Huhndorf S (2001) Neotropical ascomycetes 10. New and interesting *Cercophora* species. *Sydowia* 53:211–226
- Miller AN, Huhndorf SM (2004) A natural classification of *Lasiosphaeria* based on nuclear LSU rDNA sequences. *Mycol Res* 108:26–34
- Miller AN, Huhndorf SM (2005) Multi-gene phylogenies indicate ascomal wall morphology is a better predictor of phylogenetic relationships than ascospore morphology in the *Sordariales* (Ascomycota, Fungi). *Mol Phylogenet Evol* 35:60–75
- Mohan C (2002) Diseases of bamboos in Asia: an illustrated manual. International Network for Bamboo and Rattan
- Mostert L, Crous PW, Groenewald JZ, Gams W, Summerbell R (2003) *Togninia* (*Calosphaeriales*) is confirmed as teleomorph of *Phaeoacremonium* by means of morphology, sexual compatibility, and DNA phylogeny. *Mycologia* 95:646–659
- Mostert L, Groenewald JZ, Summerbell RC, Gams W, Crous PW (2006) Taxonomy and pathology of *Togninia* (*Diaporthales*) and its *Phaeoacremonium* anamorphs. *Stud Mycol* 54:1–115
- Müller E, von Arx JA (1962) Die Gattungen der didymosporenen Pyrenomyceten. *Beiträge zur kryptogamenflora der Schweiz* 11(2):1–922
- Nannfeldt JA (1932) Studien über die morphologie und systematik der nichtlichenisierten inoperculaten Discomyceten. *Nova Acta Regiae Societatis Scientiarum Upsaliensis* 8(2):1–368
- Nitschke TRJ (1869) Grundlage eines Systems der Pyrenomyceten. *Verh Naturhist Vereines Preuss Rheinl* 26:70–77
- Nylander (1854) *Sitzungsber. Niederrhein Ges Natur-Heilk Bonn* 2:9
- Nylander JAA (2004) MrModeltest 2.0. Program distributed by the author. Evolutionary Biology Centre, Uppsala University
- Page RDM (1996) TreeView: an application to display phylogenetic trees on personal computers. *Comput Appl Biosci* 12:357–358
- Pažoutová S, Follert S, Bitzer J, Keck M, Surup F, Šrůtka P, Holuša J, Stadler M (2013) A new endophytic insect-associated *Daldinia* species, recognised from a comparison of secondary metabolite profiles and molecular phylogeny. *Fungal Divers* 60:107–123
- Peláez F, González V, Platas G, Sánchez BJ, Rubio V (2008) Molecular phylogenetic studies within the *Xylariaceae* based on ribosomal DNA sequences. *Fungal Divers* 31:111–134
- Penzig AJO, Saccardo PA (1897) Diagnoses fungorum novorum in insula Java collectorum. Ser. II. *Malpighia* 11:491–530
- Peters TF (1987) Transitions in engineering: Guillaume Henri Dufour and the early 19th century cable suspension bridges. Birkhäuser Verlag
- Petrini LE, Müller E (1986) Haupt- und Nebenfruchtformen europäischer *Hypoxylon*-Arten (*Xylariaceae*, *Sphaeriales*) und verwandter Pilze. *Mycol Helv* 1:501–627
- Petrini O, Candoussau F, Petrini L (1989) Bambusicolous fungi collected in southwestern France 1982–1989. *Mycol Helv* 3:263–279
- Phillips AJL, Alves A, Pennycook SR, Johnston PR, Ramaley A, Akulov A, Crous PW (2008) Resolving the phylogenetic and taxonomic status of dark-spored teleomorph genera in the *Botryosphaeriaceae*. *Persoonia* 21:29–55
- Phillips AJL, Alves A, Abdollahzadeh J, Slippers B, Wingfield MJ, Groenewald JZ, Crous PW (2013) The *Botryosphaeriaceae*: genera and species known from culture. *Stud Mycol* 76:51–167
- Phookamsak R, Liu JK, Manamgoda DS, Wanasinghe DN, Ariyawansa H, Mortimer PE, Chukeatirote E, McKenzie EHC, Hyde KD (2014) Epitypification of two bambusicolous fungi from Thailand. *Cryptogam Mycol* 35:239–256
- Phookamsak R, Norphanphoun C, Tanaka K, Dai DQ, Luo ZL, Liu JK, Su HY, Bhat DJ, Bahkali AH, Mortimer PE, Xu JC, Hyde KD (2015) Towards a natural classification of *Astrosphaeriella*-like species; introducing *Astrosphaeriellaceae* and *Pseudoastrosphaeriellaceae* fam. nov. and *Astrosphaeriellopsis*, gen. nov. *Fungal Divers* 74:143–197
- Pidoplichko NM, Kirilenko TS (1969) New species of the genus *Myrothecium* Tode. *Mykrobiologichnyi Zhurnal Kiev* 31:158–163
- Pinho DB, Junior JH, Nicoli A, Junior BTH, Bragança CAD, Pereira OL (2012) Phylogenetic placement of the genus *Anhellia* and the description of *A. nectandrae* sp. nov. *Mycologia* 104:1291–1298
- Pöldmaa K (2011) Tropical species of *Cladobotryum* and *Hypomyces* producing red pigments. *Stud Mycol* 68:1–34
- Pongcharoen W, Rukachaisirikul V, Phongpaichit S, Rungjindamai N, Sakayaroj J (2006) Pimarane Diterpene and Cytochalasin Derivatives from the Endophytic Fungus *Eutypella scoparia* PSU-D44. *J Nat Prod* 69:856–858

- Pratibha J, Bhat DJ (2008) New and unusual hyphomycetes from Mahabaleshwar, India. *Mycotaxon* 2008:423–431
- Punithalingam E (1969) Studies on *Sphaeropsidales* in culture. *Mycol Pap* 119:1–24
- Raciborski M (1900) Parasitische Algen und Pilze Java's. 3:1–49
- Rannala B, Yang Z (1996) Probability distribution of molecular evolutionary trees: a new method of phylogenetic inference. *J Mol Evol* 43:304–311
- Réblová M (2009) Teleomorph of *Rhodoveronaea* (*Sordariomycetidae*) discovered and re-evolution of *Pleurophragmium*. *Fungal Divers* 36:129–139
- Rehm H (1901) Beiträge zur Pilzflora von Südamerika. XII. *Sphaeriales*. *Hedwigia* 40:100–124
- Rehm H (1907) *Ascomycetes novi*. *Ann Mycol* 5:516–546
- Rehm H (1913) *Ascomycetes philippinenses*. 11. *Philipp J Sci Sect C Bot* 8:251–263
- Rehm H (1914) *Ascomycetes philippinenses*. VI. *Leafl Philipp Bot* 6: 22572281
- Rehner S (2001) Primers for elongation factor 1- $\alpha$  (EF1- $\alpha$ ). <http://ocid.NACSE.ORG/research/deephyphae/EF1primer.pdf>
- Rodrigues KF, Petrini O (1997) Biodiversity of endophytic fungi in tropical regions. In: Hyde KD (ed) *Biodiversity of Tropical Microfungi*. Hong Kong University Press, Hong Kong, pp 57–69
- Rogers JD (1986) Provisional keys to *Xylaria* species in continental United States. *Mycotaxon* 26:85–97
- Rogers JD, Ju YM (1998) Keys to the *Xylariaceae* (excluding *Anthostomella*) of the British Isles. *Bot J Scotl* 50:153–160
- Rogers JD, Ju YM (2012) The *Xylariaceae* of the Hawaiian Islands. *North Am Fungi* 7:1–35
- Rogers JD, Samuels GJ (1986) *Ascomycetes of New Zealand* 8. *Xylaria*. *N Z J Bot* 24:615–650
- Rogers JD, Miller AN, Vasilyeva LN (2008) Pyrenomycetes of the Great Smoky Mountains National Park. VI. *Kretzschmaria*, *Nemania*, *Rosellinia* and *Xylaria* (*Xylariaceae*). *Fungal Divers* 29:107–116
- Rolshausen P, Mahoney NE, Molyneux RJ, Gubler WD (2006) A reassessment of the species concept in *Eutypa lata*, the causal agent of *Eutypa* dieback of grapevine. *Phytopathology* 96:369–377
- Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19:1572–1574
- Rossmann AY, Crous PW, Hyde KD, Hawksworth DL, Aptroot A, Bezerra JL, Bhat DJ, Boehm E, Braun U, Boonmee S, Camporesi E, Chomnunti P, Dai DQ, D'souza MJ, Dissanayake A, Jones EBG, Groenewald JZ, Hernández-Restrepo M, Hongsanan S, Jaklitsch WM, Jayawardena LWJ, Kirk PM, Lawrey JD, Mapook A, McKenzie EHC, Monkai J, Phillips AJL, Phookamsak R, Raja HA, Seifert KA, Senanayake I, Slippers B, Suetrong S, Tanaka K, Taylor JE, Thambugala KM, Tian Q, Tibpromma S, Wanasinghe DN, Wijayawardene NN, Wikee S, Woudenberg JHC, Wu HX, Yan J, Yang T, Zhang Y (2015) Recommended names for pleomorphic genera in *Dothideomycetes*. *IMA Fungus* 6:507–523
- Saccardo PA (1875) *Conspectus generum Pyrenomycetum Italicorum*. *Atti Acad Sci Veneto-Trentino-Istria* 4:101–141
- Saccardo PA (1878) *Fungi Veneti novi vel critici vel mycologiae Venetae addendi*. Series VIII. *Michelia* 1(2):239–275
- Saccardo PA (1886) *Sylloge hyphomycetum*. *Syll Fung* 4:1–807
- Saccardo PA (1891) *Sylloge Fungorum* 9:1045
- Saccardo PA, Paoletti G (1888) *Mycetes Malacenses*. *Funghi della penisola di Malacca raccolti nel 1885 dell. Ab Benedetto Scortechini* 6:387–428
- Samuels GJ, McKenzie EHC, Buchanan DE (1981) *Ascomycetes of New Zealand* 3. Two new species of *Aptospora* and their *Arthrimum* anamorphs on bamboo. *N Z J Bot* 19(2):137–149
- Sato G, Tanaka K, Hosoya T (2008) Bambusicolous fungi in Japan (8): a new species of *Pseudolachnella* from Yakushima Island, southern Japan. *Mycoscience* 49:392–394
- Schmidt JC, Kunze G (1817) *Mykologische Hefte* 1:1–109
- Schoch CL, Shoemaker RA, Seifert KA, Hambleton S, Spatafora JW, Crous PW (2006) A multigene phylogeny of the *Dothideomycetes* using four nuclear loci. *Mycologia* 98:1041–1052
- Schoch CL, Crous PW, Groenewald JZ, Boehm EWA, Burgess TI, de Gruyter J, de Hoog GS, Dixon LJ, Grube M, Gueidan C, Harada Y, Hatakeyama S, Hirayama K, Hosoya T, Huhndorf SM, Hyde KD, Jones EBG, Kohlmeyer J, Kruys A, Li YM, Lücking R, Lumbsch HT, Marvanová L, Mbachou JS, McVay AH, Miller AN, Mugambi GK, Muggia L, Nelsen MP, Nelson P, Owensby CA, Phillips AJL, Phongpaichit S, Pointing SB, Pujade-Renaud V, Raja HA, Rivas Plata E, Robbertse B, Ruibal C, Sakayaroj J, Sano T, Selbmann L, Shearer CA, Shirouzu T, Slippers B, Suetrong S, Tanaka K, Volkmann-Kohlmeyer B, Wingfield MJ, Wood AR, Woudenberg JHC, Yonezawa H, Zhang Y, Spatafora JW (2009) A class-wide phylogenetic assessment of *Dothideomycetes*. *Stud Mycol* 64:1–15
- Seaver FJ (1912) The genus *Lasio-sphaeria*. *Mycologia* 4:115–124
- Senanayake IC, Maharachchikumbura SSN, Hyde KD, Bhat JD, Jones EBG, McKenzie EHC, Dai DQ, Daranagama DA, Dayaratne MC, Goonasekara ID, Konta S, Li WJ, Shang QJ, Stadler M, Wijayawardene NN, Xiao YP, Norphanphoun C, Li Q, Liu XZ, Bahkali AH, Kang JC, Wang Y, Wen TC, Wendt L, Xu JC, Camporesi E (2015) Towards unraveling relationships in *Xylariomycetidae* (*Sordariomycetes*). *Fungal Divers* 73:73–144
- Sharma R, Kulkarni G, Sonawane MS, Shouche YS (2014) A new endophytic species of *Arthrimum* (*Apisporaceae*) from *Jatropha podagrica*. *Mycoscience* 55:118–123
- Shenoy BD, Jeewon R, Hyde KD (2005) *Oxydothis bambusicola*, a new ascomycete with a huge subapical ascus ring found on bamboo in Hong Kong. *Nova Hedwigia* 80(3–4):511–518
- Shimokawa T, Ishida M, Yoshida S, Nojiri M (2009) Effects of growth stage on enzymatic saccharification and simultaneous saccharification and fermentation of bamboo shoots for bioethanol production. *Bioresour Technol* 100:6651–6654
- Shirouzu T, Harada Y (2004) Bambusicolous fungi in Japan (2): *Phialosporostilbe gregarioclava*, a new anamorphic fungus from *Sasa*. *Mycoscience* 45:390–394
- Silvestro D, Michalak I (2011) raxmlGUI: a graphical front-end for RAxML. *Org Divers Evol* 12:335–337
- Singh SM, Yadav LS, Singh PN, Hapat R, Sharma R, Singh SK (2012) *Arthrimum rasikravindrii* sp. nov. from Svalbard, Norway. *Mycotaxon* 122:449–460
- Singhal P, Bal LM, Satya S, Sudhakar P, Naik SN (2013) Bamboo shoots: a novel source of nutrition and medicine. *Crit Rev Food Sci Nutr* 53: 517–534
- Sir EB, Kuhnert E, Surup F, Hyde KD, Stadler M (2015) Discovery of new mitorubrin derivatives from *Hypoxylon fulvo-sulphureum* sp. nov. (*Ascomycota*, *Xylariales*). *Mycol Prog* 14:1–12
- Sivanesan A (1983) Studies on *Ascomycetes*. *Trans Br Mycol Soc* 81(2): 313–332
- Slippers B, Boissin E, Phillips AJL, Groenewald JZ, Lombard L, Wingfield MJ, Postma A, Burgess T, Crous PW (2013) Phylogenetic lineages in the *Botryosphaeriales*: a systematic and evolutionary framework. *Stud Mycol* 76:31–49
- Smith G, Hyde KD (2001) Fungi from palms. XLIX. *Astrocystis*, *Biscogniauxia*, *Cyanopulvis*, *Hypoxylon*, *Nemania*, *Guestia*, *Rosellinia* and *Stilbohypoxylon*. *Fungal Divers* 7:89–127
- Smith GJ, Liew EC, Hyde KD (2003) The *Xylariales*: a monophyletic order containing 7 families. *Fungal Divers* 13:175–208
- Song X, Zhou GM, Jiang H, Yu SQ, Fu JH, Li WZ, Wang WF, Ma ZH, Peng CH (2011) Carbon sequestration by Chinese bamboo forests and their ecological benefits: assessment of potential, problems, and future challenges. *Environ Rev* 19:418–428
- Spegazzini C (1912) *Mycetes argentinenses* (Series VI). *An Mus Nac Hist Nat B Aires* 23:1–146

- Stadler M, Kuhnert E, Peršoh D, Fournier J (2013) The *Xylariaceae* as model example for a unified nomenclature following the “One Fungus-One Name” (1F1N) concept. *Mycology* 4:5–21
- Stadler M, Læssøe T, Fournier J, Decock C, Schmieschek B, Tichy HV, Peršoh D (2014) A polyphasic taxonomy of *Daldinia* (*Xylariaceae*). *Stud Mycol* 77:1–143
- Stamatakis A (2006) RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22(21):2688–2690
- Starbäck K (1899) The *Myriangiales* Bihang till Kongliga Svenska Vetenskaps Academien Handlingar 25:3–37
- Subramanian CV, Natarajan K (1972) *Tretophragmia* Gen. Nov. Proc Indian Natl Sci Acad 39(B):549–552
- Summerbell RC, Gueidan C, Schroers HJ, de Hoog GS, Starinck M, Arocha-Rosete Y, Guarro J, Scott JA (2011) Acremonium phylogenetic overview and revision of *Gliomastix*, *Trichothecium* and *Sarocladium*. *Stud Mycol* 68:139–162
- Suwannasai N, Rodtong S, Thienhirum S, Whalley AJ (2005) New species and phylogenetic relationships of *Hypoxylon* species found in Thailand inferred from the internal transcribed spacer regions of ribosomal DNA sequences. *Mycotaxon* 94:033–324
- Suwannasai N, Martín MP, Phosri C, Sihanonth P, Whalley AJS, Spouge JL (2013) Fungi in Thailand: a case study of the efficacy of an ITS barcode for automatically identifying species within the *Annulohypoxylon* and *Hypoxylon* genera. *PLoS One* 8(2):e54529. doi:10.1371/journal.pone.0054529
- Swofford DL (2002) PAUP: phylogenetic analysis using parsimony, version 4.0 b10. Sinauer Associates, Sunderland
- Sydow H (1938) Neue oder bemerkenswerte australische Micromyceten - III. *Ann Mycol* 36:295–313
- Sydow H, Sydow P (1913) Enumeration of Philippine fungi with notes and descriptions of new species. 1: Micromycetes. *Philipp J Sci Sect C Bot* 8:265–285
- Sydow H, Sydow P (1914a) Diagnosen neuer Philippinischer Pilze. *Ann Mycol* 12(6):545–576
- Sydow H, Sydow P (1914b) Enumeration of Philippine fungi with notes and descriptions of new species. 11. *Philipp J Sci Sect C Bot* 8:475–508
- Sydow H, Sydow P (1917) Beitrag zur Kenntniss der Pilzflora der Philippinen-Inseln. *Ann Mycol* 15(3–4):165–268
- Tai FL, Wei CT (1933) Notes on Chinese fungi. III. *Sinensia* 4:83–128
- Tamura K, Stecher G, Peterson D, Filipiński A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol* 30:2725–2729
- Tanaka K, Harada Y (2003a) Pleosporales in Japan (1): the genus *Lophiostoma*. *Mycoscience* 44:85–96
- Tanaka K, Harada Y (2003b) Pleosporales in Japan (3). The genus *Massarina*. *Mycoscience* 44:173–185
- Tanaka K, Harada Y (2004) Bambusicolous fungi in Japan (1): four *Phaeosphaeria* species. *Mycoscience* 45:377–382
- Tanaka K, Harada Y (2005a) Bambusicolous fungi in Japan (4): a new combination, *Astrosphaeriella aggregata*. *Mycoscience* 46:114–118
- Tanaka K, Harada Y (2005b) Bambusicolous fungi in Japan (6): *Katumotoa*, a new genus of *phaeosphaeriaceae* ascomycetes. *Mycoscience* 46:313–318
- Tanaka K, Harada Y, Barr ME (2005) Bambusicolous fungi in Japan (3): a new combination, *Kalmusia scabriscora*. *Mycoscience* 46:110–113
- Tanaka K, Hirayama K, Yonezawa H, Hatakeyama S, Harada Y, Sano T, Shirouzu T, Hosoya T (2009) Molecular taxonomy of bambusicolous fungi: *Tetraplospheariaceae*, a new pleosporalean family with *Tetraploa*-like anamorphs. *Stud Mycol* 64:175–209
- Tanaka K, Hirayama K, Yonezawa H, Sato G, Toriyabe A, Kudo H, Hashimoto A, Matsumura M, Harada Y, Kurihara Y, Shirouzu T, Hosoya T (2015) Revision of the *Massarineae* (*Pleosporales*, *Dothideomycetes*). *Stud Mycol* 82:75–136
- Tang AMC, Jeewon R, Hyde KD (2009) A re-evaluation of the evolutionary relationships within the *Xylariaceae* based on ribosomal and protein-coding gene sequences. *Fungal Divers* 34:127–155
- Taylor JE, Hyde KD (2003) Microfungi of tropical and temperate palms. *Fungal Divers Press* 12:1–459
- Taylor JE, Fröhlich J, Hyde KD (2001) *Lasiosphaeria* and a similar new genus from palms. *Mycoscience* 42:369–377
- Tewari DN (1993) A monograph on bamboo. International book distributors
- Thambugala KM, Hyde KD, Tanaka K, Tian Q, Wanasinghe DN, Ariyawansa HA, Jayasiri SC, Boonmee S, Camporesi E, Hashimoto A (2015) Towards a natural classification and backbone tree for *Lophiostomataceae*, *Floricolaceae*, and *Amorosiaceae* fam. nov. *Fungal Divers* 74:199–266
- Theissen F, Sydow H (1918) Vorentwürfe zu den Pseudosphaeriales. *Ann Mycol* 16:1–34
- Tode HJ (1790) *Fungi Mecklenburgenses Selecti*. 1:1–47
- Trouillas FP, Pitt WM, Sosnowski MR, Huang R, Peduto F, Loschiavo A, Savocchia S, Scott ES, Gubler WD (2011) Taxonomy and DNA phylogeny of *Diatrypaceae* associated with *Vitis vinifera* and other woody plants in Australia. *Fungal Divers* 49:203–223
- Trouillas FP, Hand FP, Inderbitzin P, Gubler WD (2015) The genus *Cryptosphaeria* in the western United States: taxonomy, multilocular phylogeny and a new species, *C. multicontinentalis*. *Mycologia* 107:1304–1313
- Tulasne LR, Tulasne C (1863) *Selecta Fungorum Carpologia*. 2:1–319. Parisii, Imperatoris Jussu, In Imperiali Typographeo Excudebatur, Paris
- Tulasne LR, Tulasne C (1865) *Selecta Fungorum Carpologia: Nectrii-Phacidiei-Pezizei*. 3
- Tulloch M (1972) The genus *Myrothecium*. *Mycol Pap* 130:1–42
- Udagawa S, Awao T (1984) A new *Myrothecium* species from Japan, a rhizonic acid producer. *Mycotaxon* 20:381–387
- Vasilyeva LN, Stephenson SL (2004) Pyrenomycetes of the Great Smoky Mountains National Park. I. *Diatrype* Fr. (*Diatrypaceae*). *Fungal Divers* 17:191–201
- Vasilyeva LN, Stephenson SL (2006) Pyrenomycetes of the Great Smoky Mountains National Park. III. *Cryptosphaeria*, *Eutypa* and *Eutypella* (*Diatrypaceae*). *Fungal Divers* 22:243–254
- Verma RK, Sharma N, Soni KK, Jamaluddin (2008) Forest fungi of central India. International Book Distributing Co, Lucknow
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *J Bacteriol* 172:4238–4246
- Von Arx JA, Müller E (1975) A re-evaluation of the bitunicate Ascomycetes with keys to families and genera. *Stud Mycol* 9:1–159
- von Höhnel F (1909) Fragmente zur Mykologie: VIII. Mitteilung (Nr. 354 bis 406). Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften Math.-naturw. Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften Math.-naturw Klasse Abt I 118:1157–1246
- Whalley MA, Whalley AJS, Jones EBG (1996) *Camillea selangorensis* sp. nov. from Malaysia. *Sydowia* 48:145–151
- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (eds) *PCR protocols: a guide to methods and applications*, p 315–322
- Wijayawardene DNN, McKenzie EHC, Hyde KD (2012) Towards incorporating anamorphic fungi in a natural classification checklist and notes for (2011). *Mycosphere* 3:157–228
- Wijayawardene NN, Crous PW, Kirk PM, Hawksworth DL, Boonmee S, Braun U, Chomnunti P, Dai DQ, D’souza MJ, Diederich P, Dissanayake A, Doilom M, Hongsanan S, Jones EBG, Groenewald JZ, Jayawardena R, Lawrey JD, Liu JK, Lücking R, Madrid H, Manamgoda DS, Muggia L, Nelsen MP, Phookamsak R, Suetrong S, Tanaka K, Thambugala KM, Wikee S, Zhang Y,



- Aptroot A, Ariyawansa HA, Bahkali AH, Bhat JD, Gueidan C, De Hoog GS, Knudsen K, McKenzie EHC, Miller AN, Mortimer PE, Wanasinghe DN, Phillips AJL, Raja HA, Slippers B, Shivas RS, Taylor JE, Wang Y, Woudenberg JHC, Piątek M, Cai L, Jaklitsch WM, Hyde KD (2014) Naming and outline of *Dothideomycetes*–2014 including proposals for the protection or suppression of generic names. *Fungal Divers* 69:1–55
- Wijayawardene NN, Hyde KD, Wanasinghe DN, Papizadeh M, Camporesi E, Bhat DJ, McKenzie EHC, Phillips AJL, Diederich P, Tanaka K, Li WJ, Tangthirasun N, Phookamsak R, Dai DQ, Dissanayake AJ, Weerakoon G, Maharachchikumbura SSN, Goonasekara I, Hashimoto A, Matsumura M, Wang Y (2016) Taxonomy and phylogeny of dematiaceous coelomycetes. *Fungal Divers*, accepted
- Wingfield MJ, De Beer ZW, Slippers B, Wingfield BD, Groenewald JZ, Lombard L, Crous PW (2012) One fungus one name promotes progressive plant pathology. *Mol Plant Pathol* 13:604–613
- Worapong J, Sun J, Newcombe G (2009) First report of *Myrothecium roridum* from a gymnosperm. *North Am Fungi* 4:1–6
- Wu YM, Jiang YL, Ma YN, Zhang TY (2015) Two new species of *Myrothecium* from the Qinghai-Tibet Plateau Area, China. *Mycotaxon* 129:403–406
- Xu MQ, Dai YC, Fan SH, Jin LX, Lu Q, Tian GZ, Wang LF (2006) Records of bamboo diseases and the taxonomy of their pathogens in China (I). *For Res* 19:692–699
- Xu MQ, Dai YC, Fan SH, Jin LX, Lu Q, Tian GZ, Wang LF (2007) Records of bamboo diseases and the taxonomy of their pathogens in China (II). *For Res* 20:45–52
- Zhang LQ, Wang XG (1999) Fungus Resource of Bamboos in China. *J Bamboo Res* 18:66–72
- Zhang ZY, Zhang X (2000) Potentials of bamboo in traditional Chinese medicine and development of health products. *World Sci Technol* 3: 54–56
- Zhang XM, Zhuang WY (2003) Re-examinations of *Bionectriaceae* and *Nectriaceae* (*Hypocreales*) from tropical China on deposit in HMAS. *Nova Hed* 76(1–2):191–200
- Zhang N, Castlebury LA, Miller AN, Huhndorf SM, Schoch CL, Seifert KA, Rossman AY, Rogers JD, Kohlmeyer J, Sung G-H (2006) An overview of the systematics of the *Sordariomycetes* based on a four-gene phylogeny. *Mycologia* 98:1076–1087
- Zhang Y, Wang HK, Fournier J, Crous PW, Jeewon R, Pointing SB, Hyde KD (2009) Towards a phylogenetic clarification of *Lophiostoma/Massarina* and morphologically similar genera in the *Pleosporales*. *Fungal Divers* 38:225–251
- Zhang M, Wu HY, Wang ZY (2010) Taxonomic studies of *Helminthosporium* from China 5. Two new species from Hunan and Sichuan Province. *Mycotaxon* 113:95–99
- Zhang Y, Crous PW, Schoch CL, Hyde KD (2012) *Pleosporales*. *Fungal Divers* 53:1–221
- Zhao GC, Zhao RL (2012) The higher microfungi from forests of Yunnan Province. book:1–572
- Zhao P, Luo J, Zhuang WY (2011) Practice towards DNA barcoding of the nectriaceae fungi. *Fungal Divers* 46:183–191
- Zhao KN, Yin GH, Zhao GZ, Cao AX (2014) *Embryonispora*, a new genus of hyphomycetes from China. *Mycotaxon* 126:77–81
- Zhaxybayeva O, Gogarten JP (2002) Bootstrap, Bayesian probability and maximum likelihood mapping: exploring new tools for comparative genome analyses. *BMC Genomics* 3(1):4
- Zhou D, Hyde KD (2001) Host-specificity, host-exclusivity, and host-recurrence in saprobic fungi. *Mycol Res* 105:1449–1457
- Zhou D, Hyde KD (2002) Fungal succession on bamboo in Hong Kong. *Fungal Divers* 10:213–227
- Zhou BZ, Fu MY, Xie JZ, Yang XS, Li ZC (2005) Ecological functions of bamboo forest: research and application. *J For Res* 16:143–147