



# Biogenic production of silver nanoparticles from milk of *Capra aegagrus hircus* and mechanism of antibacterial activity on different bacteria

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## Abstract

In the present study, we report a simple, rapid, cost-effective approach for the white synthesis of silver nanoparticles (AgNPs) using *Capra aegagrus hircus* milk. The formation of AgNPs was visually examined and further investigated using UV–visible spectrophotometer, transmission electron microscopy, scanning electron microscopy with energy dispersive X-ray, Fourier infrared spectroscopy, and X-ray diffractometer. Crystalline lattice indices of AgNPs were performed using the XRD analysis. The diffraction peaks at  $2\theta$  values of  $37.7^\circ$ ,  $46.1^\circ$ ,  $67.4^\circ$ , and  $76.84^\circ$  corresponding to lattice planes (111), (200), (220), and (311), respectively. The obtained AgNPs were spherical in shape with the size between 5 and 50 nm. The antibacterial activity of AgNPs against *Klebsiella* sp. (Accession Number: KC899845), and *Staphylococcus* sp. (Accession Number: KC688883) were evaluated by means of cell growth.

**Keywords** Antibacterial · Goat milk · Silver · Pathogens · White synthesis

## Introduction

Nanoparticles are being considered as an eminent component of the widely accelerating field of nanotechnology exemplifying various real-world applications. In the realms of metal nanoparticles, silver, gold, copper, and zinc oxide have been demonstrated as phenomenon alternative therapeutic agents (Khan et al. 2021). Metallic nanoparticles have attracted considerable scientific interest due to their

unique optical properties of surface plasmon resonance (SPR) and physicochemical properties (Castro et al. 2014). They are classified into different types such as carbon, metal, ceramic, polymeric, semi-conductor, and lipid-based nanoparticles, based on size, and structure properties (Thomas et al. 2015). Among them, silver nanoparticles (AgNPs) are one of the most sought after and greatly investigated metal nanoparticles.

Silver nanoparticles (AgNPs) are one of the most engineered nanoparticles used in several commercial areas, including medical devices, healthcare products, cleaning agents, food storage, packing, and textile coatings (Rolim et al. 2019). Silver nanoparticles (AgNPs) play a significant role in resolving numerous medical problems due to their chemical biocompatibility, inertness, oxidation resistance, and safe use as antibacterial activity (Carlson et al. 2008) against a variety of microorganisms (Mahmoud et al. 2021). The antimicrobial potency of silver nanoparticles increased the usage of silver nanoparticles in wound dressing, drug carriers, and in artificial implantation (Liao et al. 2019). They are also used for environmental applications because of their potent antimicrobial activity against bacteria, viruses, and fungi (Rolim et al. 2019; Ahluwalia et al. 2018).

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To exploit the numerous properties of AgNPs, innovative methods of synthesis are being employed. Chemical and physical modes are widely implemented for the synthesis of AgNPs (Khan et al. 2021; Gurunathan et al. 2015). Eco-friendly green synthesis of nanoparticles is biocompatible, cost effective, less time and energy consuming and it also non toxic compared to the physical and chemical methods (Veeraraghavan et al. 2021). Accordingly, numerous biological resources are being explored for the synthesis of AgNPs including, plants (Aravinthan et al. 2015; Mythili et al. 2018; Sengottaiyan et al. 2016; Ameen et al., 2019; Sampath et al., 2021), bacteria (Ameen et al. 2020), fungi (Mukherjee et al. 2002), algae (Gopu et al., 2021; Valarmathi et al., 2020; Govarthan et al. 2017) oilcakes (Govarthan et al. 2016), organic compound (Govarthan et al. 2014), and even vegetable waste (Mythili et al. 2018). So far, many studies reported for AgNPs as plant extracts, due to medical value owing to the presence of phenolic compounds (polyphenols, tannic acid, flavonoids, terpenoids), amino acids and vitamins (Ebrahiminezhad et al. 2018). But only few studies attempted the biosynthesis of AgNPs using milk and other related bioresources.

Lee et al. (2013) reported that, synthesis of AgNPs using cow milk have potential application in medical and pharmaceutical sciences. Although a wide variety of biological sources have been used to synthesize AgNPs, the use of *Capra aegagrus* (goat) milk is not common. However, there is no report on synthesis of AgNPs using goat milk. In addition, goat milk has various effects of human health considering the total solid, fat, protein, mineral, vitamins, and lactose (Turkmen et al. 2017). In particular, goat milk contains 13.2% total solids, consisting of 4.5% fat, 3.6% protein, 0.8% minerals, and 4.3% lactose (Üçüncü 2013). It has been reported that the goat milk has maximum number of conjugated linoleic acids which play an important role in stimulation of immunity, growth promotion, and prevention of diseases. The most important effect of goat milk proteins is their healing effect on cow milk allergy, the most common food allergy, which causes many deaths in infants (Turkmen 2017). The present study investigated the biological synthesis of silver nanoparticles using fresh goat milk and evaluate the antibacterial activity of the synthesized AgNPs against the human pathogens.

## Materials and methods

### Materials and chemicals

Silver nitrate was purchased from Sigma–Aldrich (St. Louis, MO), nutrient broth was purchased from Hi-media Laboratories Pvt. Ltd (Mumbai, India). *Capra aegagrus* milk was

procured from local goat farm in Mallasamudram, Tamil Nadu, India.

### Synthesis of AgNPs

AgNPs synthesis was carried out according to Lee et al. (2013). Briefly, 4 ml of goat milk was mixed with 96 ml of 1 mM silver nitrate solution. The mixture was then incubated in a rotary shaking incubator at 37 °C until the color changed to dark brown. The reaction precipitates were then filtered and centrifuged at 10,000 rpm for 15 min. the resulting AgNPs were washed with sterile water. The obtained AgNPs was freeze dried at – 80 °C and used for further characterization and antibacterial studies.

### Characterization of AgNPs

The optical absorption spectra of the synthesized AgNPs were observed using UV–visible spectrophotometer (Elico-SL 164) in the range of 200–800 nm. The morphology and size of the biogenic AgNPs were determined by transmission electron microscopy (TEM, FEI Tecnai TF 20 high resolution). The presence of elemental silver in the AgNPs was analyzed by scanning electron micrograph-energy dispersive spectroscopy (SEM–EDS; Jeol JSM 6390). Fourier transform infrared (FT-IR) spectra of the AgNPs were analyzed using Perkin-Elmer FT-IR spectrophotometer (IRAffinity-1S) operated at a resolution of 4 cm<sup>-1</sup>. The spectra were recorded at wavelength ranges from 500 to 4000 cm<sup>-1</sup>. The structural characterization was conducted using X-ray powder diffraction (XPRT-Pro diffractometer using Cu–K $\alpha$  radiation). Sample Scanning was done in the region of  $2\theta$  from 20 to 80° at 0.04°/min with a time constant of 2 s.

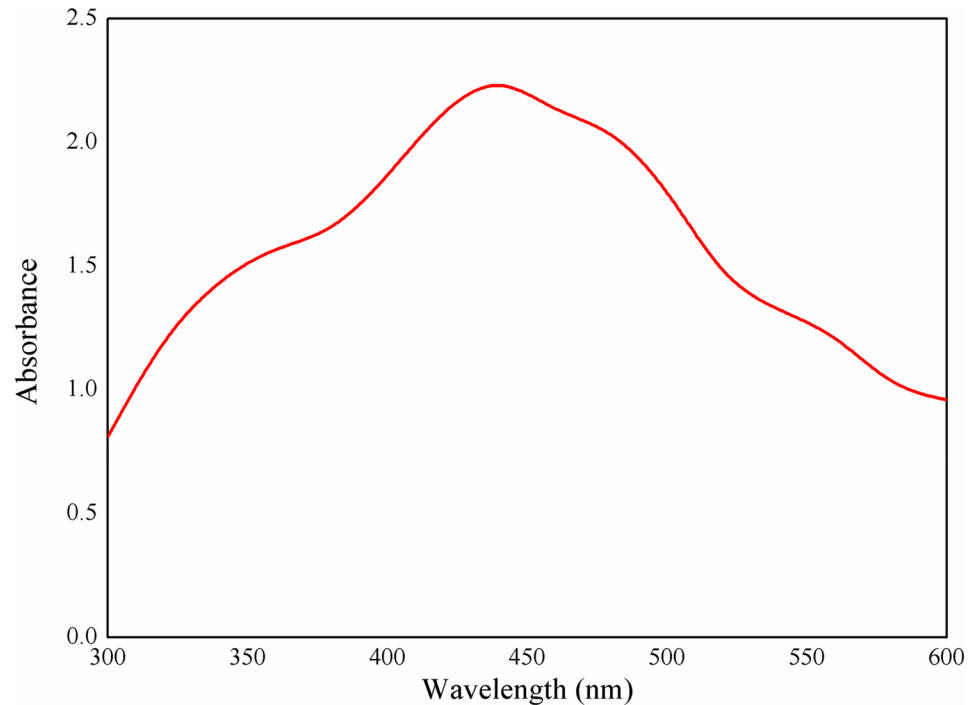
### Antibacterial activity

The antibacterial activity against *Klebsiella* sp. (Accession Number: KC899845), and *Staphylococcus* sp. (Accession Number: KC688883) of AgNPs synthesized from goat milk was evaluated according to Govarthan et al. (2014). Briefly, the strains cultivated in 100 mL of nutrient broth amended with different concentrations (1–5 mM) of AgNPs. The growth of the bacterial strains was indexed by measuring the optical density (at  $\lambda = 600$  nm) at regular intervals (0–48 h) using spectrophotometer. The growth curve was plotted between optical density and time. The flask without AgNPs was used as a control for this experiment.



**Fig. 1** Process of goat milk mediated AgNPs synthesis

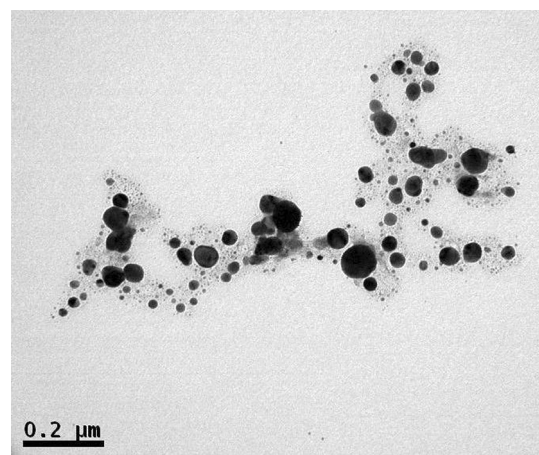
**Fig. 2** Surface plasmon resonance spectra of AgNPs synthesized using goat milk



## Results and discussion

### Characterization of AgNPs

In the present study investigated a white chemistry milk approach-based biosynthesis of AgNPs from *Capra aegagrus* milk. The development AgNPs through the reduction of  $\text{Ag}^+$  into  $\text{Ag}^0$  ions was visually confirmed from colorless to dark brown in the flasks within 5 h. It has been reported that the proteins present in the milk could be responsible for the reduction of  $\text{Ag}^+$  in the mixture. The process of goat milk mediated AgNPs synthesis is shown in Fig. 1. Our previous study obtained after 8 h of reaction, the mixture turned from milky white to dark brown (Lee et al. 2013). The preliminary conformation of AgNPs synthesis was recorded by UV–Vis spectroscopy. The surface plasmon resonance (SPR) was found to be



**Fig. 3** TEM image of synthesized AgNPs

higher at 420 nm and its absorbance intensity increased with the increase of the incubation period (Fig. 2). The SPR at 420–450 nm is corresponding to the silver nanoparticles formation, according to the earlier reports (Singh et al. 2015).

The TEM images of biogenic AgNPs are shown in Fig. 3. The size of the AgNPs was in the range of 5–50 nm. The AgNPs were found to be spherical in shape, well dispersed, and homogeneous in nature. Shape is another important parameter that has an influence on the biological activity of nanomaterials. It was stated that the cellular uptake of spherical shaped nanoparticles was higher than rod-shaped (Barabadi et al. 2021). Further validate the formation of AgNPs, samples were characterized by SEM–EDS and the results are presented in Fig. 4. Strong silver peak was observed at 3 keV, which is reasonable absorption peak of AgNPs which could be the crystallites due to SPR. The results are consistent with previous studies reporting strong absorption of AgNPs approximately at 3 keV (Lee et al. 2013). The agglomerations of the particles on the surface are due to the multilayer coating of the particle leading to the formation of a few huge clusters on the surface.

FT-IR analysis involved in identifying the possible functional groups and/or secondary metabolites involved in AgNPs synthesis. Figure 5 shows the FT-IR spectra of AgNPs synthesized from goat milk. The band at 2929  $\text{cm}^{-1}$  indicates the presence of C–H bonds of alkanes. The bands at 1662 and 1536  $\text{cm}^{-1}$  are characteristic of amide I and amide II bands, respectively. Subsequently,

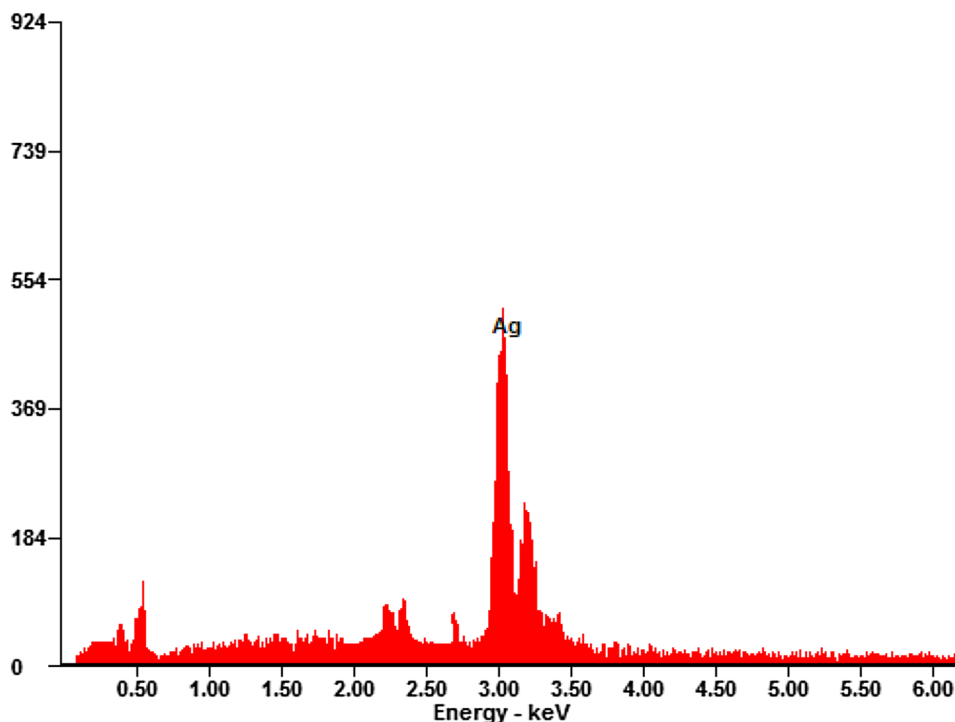
intense peaks were identified at 1404 and 1028  $\text{cm}^{-1}$ , corresponding to C=C and C–O bonds, respectively. The peak recorded at 1662  $\text{cm}^{-1}$  could be due to the C=C stretching associated with the presence of aromatic rings, while the band around 1404  $\text{cm}^{-1}$  could be attributed to aliphatic and aromatic groups in the plane deformation vibrations of methyl and methylene groups (Memon et al. 2018; Zhao et al. 2016).

Crystalline lattice indices of AgNPs were performed using the XRD analysis (Fig. 6). The diffraction peaks at  $2\theta$  values of 37.7°, 46.1°, 67.4°, and 76.84° corresponding to lattice planes (111), (200), (220), and (311), respectively. The observed peaks have been attributed to hexagonal phase of AgNPs (JCPDS no. 41–1402). A lot of unimportant peaks are seen owing to the bioorganic phases of crystallization that are stuck to the synthesized nano particles' surfaces. The XRD pattern strongly confirms the high crystalline nature of the biosynthesized silver nanoparticles. The similar XRD peaks were also found in the earlier studies have been observed the diffraction peaks of AgNPs at lattice planes of (111), (200), (220), and (311), respectively, (Govarthanan et al. 2016; Aravinthan et al. 2015).

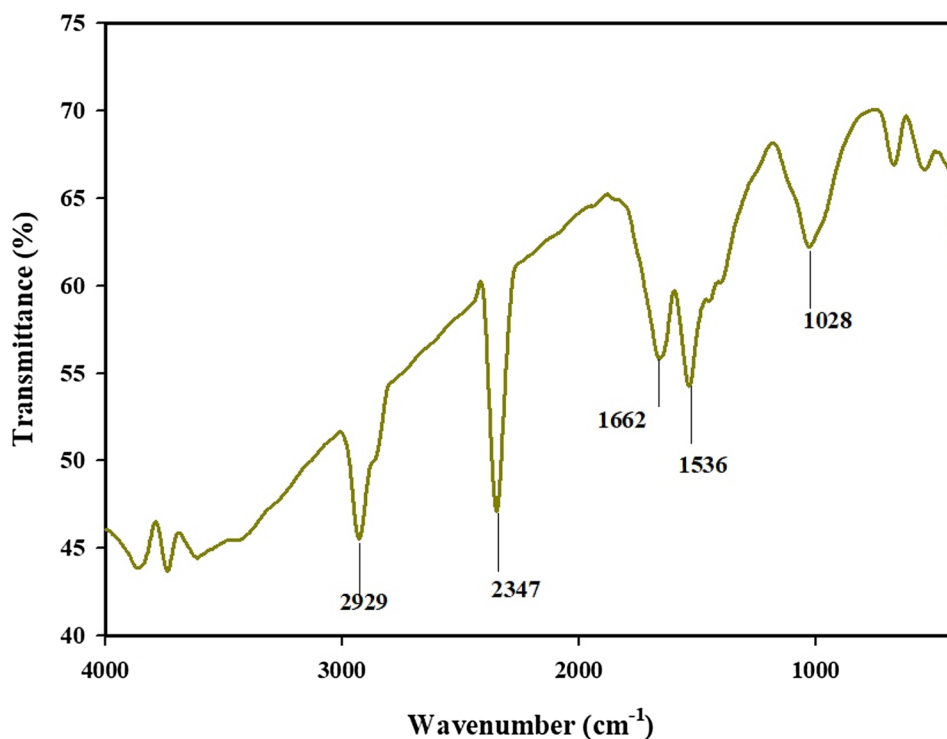
### Antibacterial activity

The antibacterial activity of AgNPs against *Klebsiella* sp. (Accession Number: KC899845), and *Staphylococcus* sp. (Accession Number: KC688883) was evaluated by means of cell growth. Optical density was measured

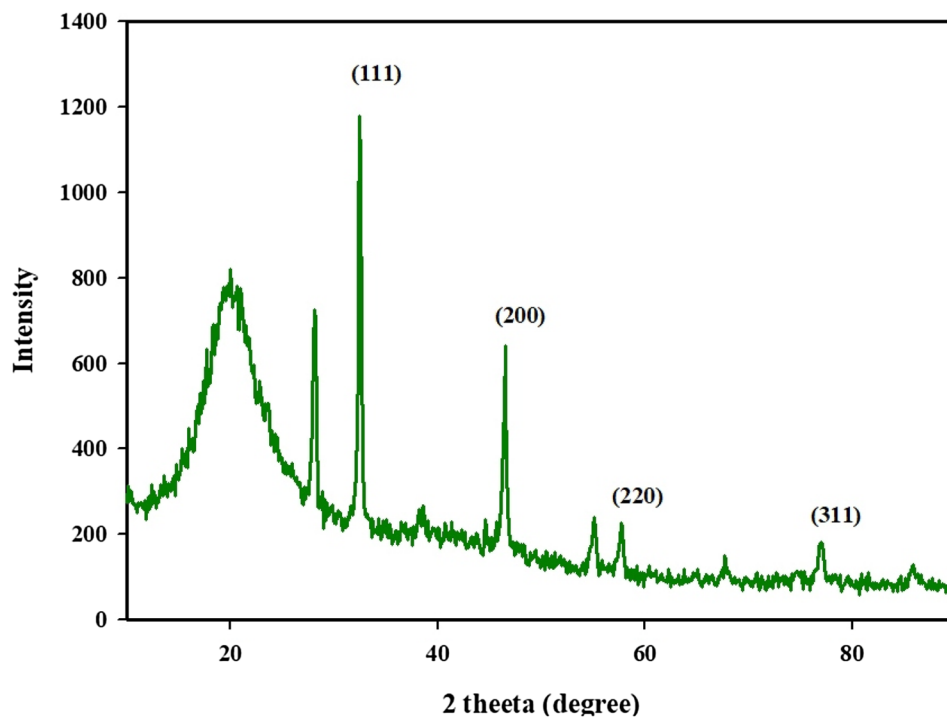
**Fig. 4** SEM–EDX of AgNPs synthesized using goat milk



**Fig. 5** FT-IR analysis of AgNPs synthesized using goat milk



**Fig. 6** XRD of goat milk mediated synthesis of AgNPs



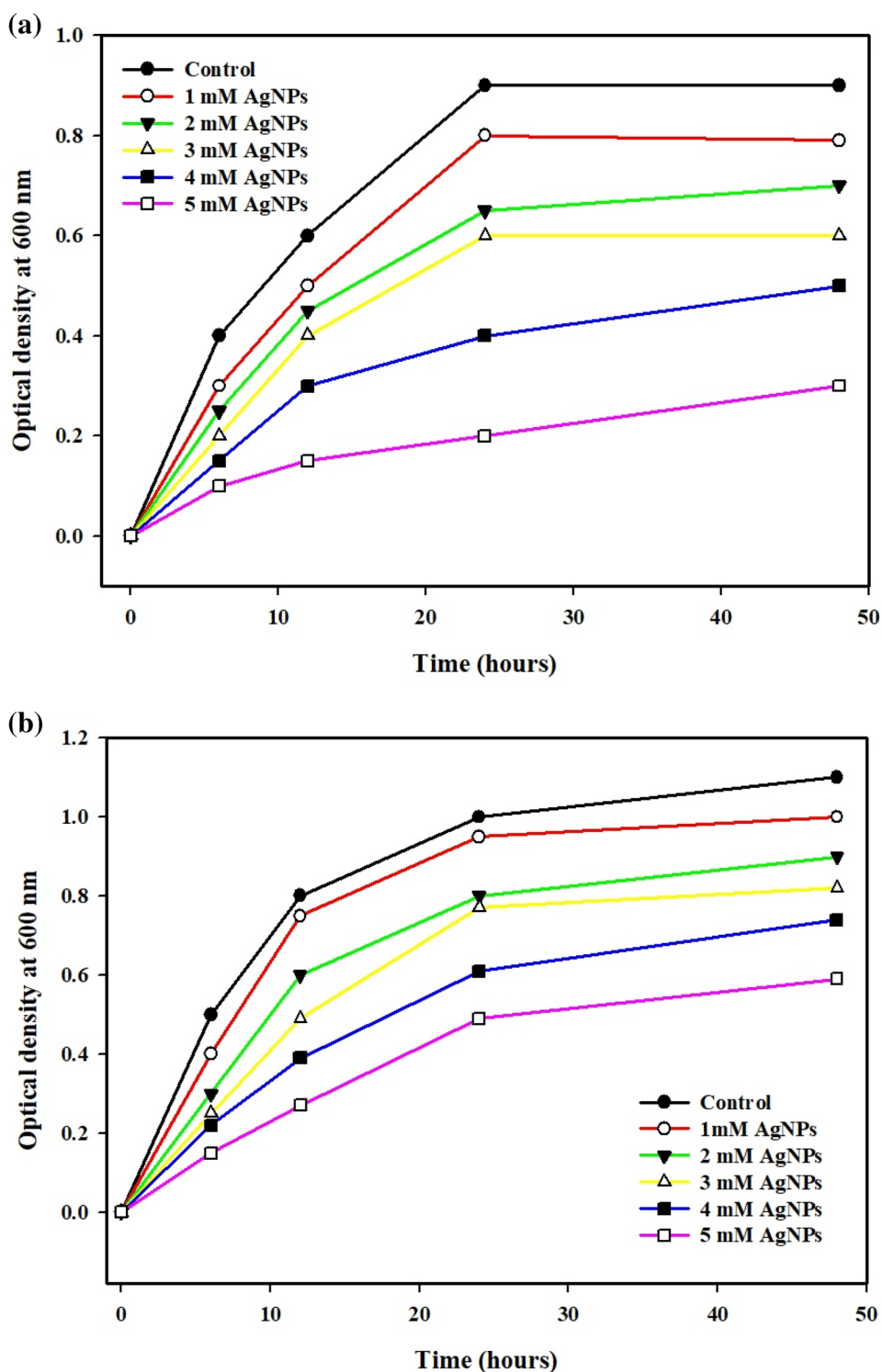
at 600 nm and plotted as a function of time for 48 h at regular intervals with various concentrations (1–5 mM) of AgNPs (Fig. 7a and b). The results showed that the higher concentration (5 mM) of AgNPs effectively encountered the population of cell growth. As expected, the increasing

concentration of AgNPs decreased the growth of bacterial population in the medium (Saxena et al. 2012).

It has previously been reported that nanoparticles with a size smaller than 20 nm can interact more easily with membrane proteins, causing maximum permeability,



**Fig. 7 a, b** Antibacterial activity of AgNPs



which leads to the cell death of bacteria (Deshmukh et al. 2019). The mechanism of antibacterial activity of AgNPs remains unclear to date. Meanwhile, the mechanism of action of silver itself is by disrupting the cell membrane, causing ROS, penetration of cell membrane, and will bind

to DNA and protein. However, based on existing works of literature, the principle of antibacterial mechanism of AgNPs is divided into oxidative stress, metal ion release, and non-oxidative mechanism (Sukweenadhi et al. 2021).

## Conclusion

A simple one step white synthesis of AgNPs is reported along with the bio-reduction as well as biostabilization potentials of *Capra aegagrus* milk. The elemental analysis confirmed the presence of elemental silver in AgNPs. FT-IR analysis confirmed that the functional groups of goat milk have played a significant role for the bio-reduction of Ag<sup>+</sup> ions into AgNPs. The antibacterial activity results clearly demonstrated that AgNPs synthesized by goat milk could be used for the treatment of disease which will cause by *Klebsiella* sp. and *Staphylococcus* sp. Thus, the white synthesized AgNPs can be used as a potential antibacterial agent in near future.

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## Declarations

**Conflict of interest** The authors declare that have no conflict of interests.

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