



Hydrochemical characteristics and quality assessment of shallow groundwater under intensive agriculture practices in arid region, Qena, Egypt

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Abstract

Monitoring groundwater quality and analyzing geochemical processes are very crucial in dry environments, especially in areas with intense agriculture and inadequate irrigation management systems. A total of forty-one groundwater samples were collected from the study area and analyzed for major ions chemistry. The results of the geochemical analyses are used to investigate the geochemical processes and make groundwater assessments. The hydrogeochemical characteristics of the groundwater samples revealed a highly mineralized $\text{Na}^+ - \text{Cl}^-$ water type, implying that the majority of the samples are chemically unfit for human consumption. Based on the water quality parameters analyzed for the groundwater samples collected in this study, such as sodium adsorption ratio (SAR), soluble sodium percentage (SSP), residual sodium carbonate (RSC), magnesium hazard (MH), permeability index (PI), and Kelley's ratio (KR), the suitability of groundwater samples for irrigation purposes is good to moderately good. Also they may reflect some alkaline hazard threats to the soil as indicated by the SSP and KR. The salinity hazard is recognized as high and expected to increase with the expansion of irrigation; hence, special salinity control management for groundwater in the study area is required. Nitrate is widely recognized as a contaminant in the study area. The nitrate concentrations range from 0.1 to 256.8 mg/L, with more than 30% of the samples exceeding the safe drinking water limit of 50 mg/L. The main concern is that the average nitrate concentration will likely continue to increase in the groundwater of the study area. Groundwater monitoring, in terms of both quality and quantity, is essential for sustainable groundwater management.

Keywords Nile Valley aquifer · Groundwater quality · Irrigation water · Hydrochemistry · Nitrate pollution · Water quality index · Egypt

Introduction

In recent years, Egypt has experienced serious water scarcity. The biggest challenge facing Egypt's water resources system is population growth and rapid economic development. The River Nile constitutes the principal permanent source of fresh water in Egypt and supplies nearly all of the country's water need, including those of agriculture and industry (Wahba et al. 2018). The current actual available

water resources in Egypt are about 55.5 billion m^3 /year from the River Nile water, and supplies around 95% of the annual fresh water demand in Egypt (SEAM 2005; Omar and Moussa 2016). Rainwater is very limited in Egypt and is concentrated in a very narrow strip on the coastal parts of the country. Groundwater constitutes the second important source of freshwater in Egypt and represents about 12% of the total water supply in Egypt (Abo-El-Fadl 2013). The groundwater resource in Egypt comprises two main major aquifers, the transboundary Nubian Sandstone Aquifer System (NSAS) and the Nile Valley Aquifer System (NVAS). The Nubian Aquifer is the deepest and oldest sedimentary rocks, which are often early to late Cretaceous in age (Hess et al. 1987). The Nubian Aquifer covers about 82% of the area of Egypt as well as south-east Libya, north-east Chad, and north Sudan with a total area of about 2.2 million square kilometers (CEDARE 2001; Hesse et al. 1987). The Nubian

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Aquifer is heavily exploited in Egypt, particularly in the Western Desert, where deep drilling wells have been carried out in recent decades (El Tahlawi et al. 2008; Mohammed 2015). The Nubian Aquifer is located in the study area beneath the shallow Nile Valley Aquifer, but it is not widely utilized. The Nile Valley Aquifer is also one of the major aquifers in Egypt. It lies beneath the majority of the Nile valley, close to the River Nile banks, and constitutes a highly productive groundwater source. The Nile Valley Aquifer is shallow, and its deposits are often Tertiary to Quaternary deposits (El Tahlawi et al. 2008; Mohammed et al. 2016). The Nile Valley Aquifer is our main focus in this study.

For decades, Egypt has pursued a nationwide program of land reclamation and new settlements in order to address the overpopulation problem and to expand agricultural land (Wolff 1993; Adriansen 2009; Ashour et al. 2009; Mohammed et al. 2016). Converting arid or desert areas to agricultural land is referred to as land reclamation. This is primarily accomplished by extending water canals from existing agricultural land or drilling groundwater wells. Groundwater resources are viewed as the major sustainable water supply in many of the reclamation projects in Egypt. New lands are also reclaimed on a small scale by residents on the desert fringes outside the old lands. They mostly are relying on groundwater for irrigation and residential purposes because of the lack of water from the River Nile. As groundwater abstraction increases due to urban and agricultural expansion, changes in groundwater quantity and quality often occur, especially in the shallow aquifer systems in arid regions (Ambast et al. 2006; Ayotte et al. 2015; Li et al. 2015). The study area is an excellent example of a vast tract of reclaimed land that is held by the government, the business sector, and individuals.

Groundwater contamination due to over-exploitation, intensive irrigation, and human activities has been reported in many regions of the world (Han 1998; El-Naqa and Al-Shayeb 2009; Nel et al. 2009; McCallum et al. 2010; Rattray 2015; Li et al. 2016). Apparently, most groundwater contains dissolved salts and trace elements as a result of the natural weathering of the earth's surface and aquifer rock-forming minerals. Agriculture has become a critical cause and a source of groundwater pollution that might alter the soil physical properties and structures and leave the water unsuitable or less valuable for other water uses (Scanlon et al. 2007; Pastén-Zapata et al. 2014; Mohammed et al. 2016). Agricultural return flow water could be of a significant component contributing to the groundwater recharge particularly in the case of intensively irrigated areas. Irrigation return flow water could also transmit pollutants that produced from the frequently applied pesticides, fertilizers, and manure to the agricultural fields (Pulido-Bosch et al.

2000; Weitzman et al. 2021). Irrigation return flow significantly increases the salinity, nitrate, and total phosphorus concentrations in groundwater (Pearce and Schumann 2001; Widory et al. 2004; Foster et al. 2018; Park et al. 2018).

The hydrogeochemical characterization and assessment of groundwater quality have become a serious concern in recent decades as public awareness of the importance of groundwater quality protection rises. A various literature on groundwater quality has been published and recognized in many parts of the world (Li et al. 2013; Ogunfowokan et al. 2013; Alaya et al. 2014; Brindha et al. 2014; Wu et al. 2014; Shammi et al. 2016; Yehia et al. 2017; Tolera et al. 2020). Pollution of groundwater has an impact on water quality, human health, and economic development (Schiavo et al. 2006). In the developing countries, about 80% of diseases are directly related to poor drinking water quality and unsanitary conditions (UNESCO 2007; Das and Nag 2015). As a result, determining the quality of groundwater is critical for determining the acceptability of water for a specific use.

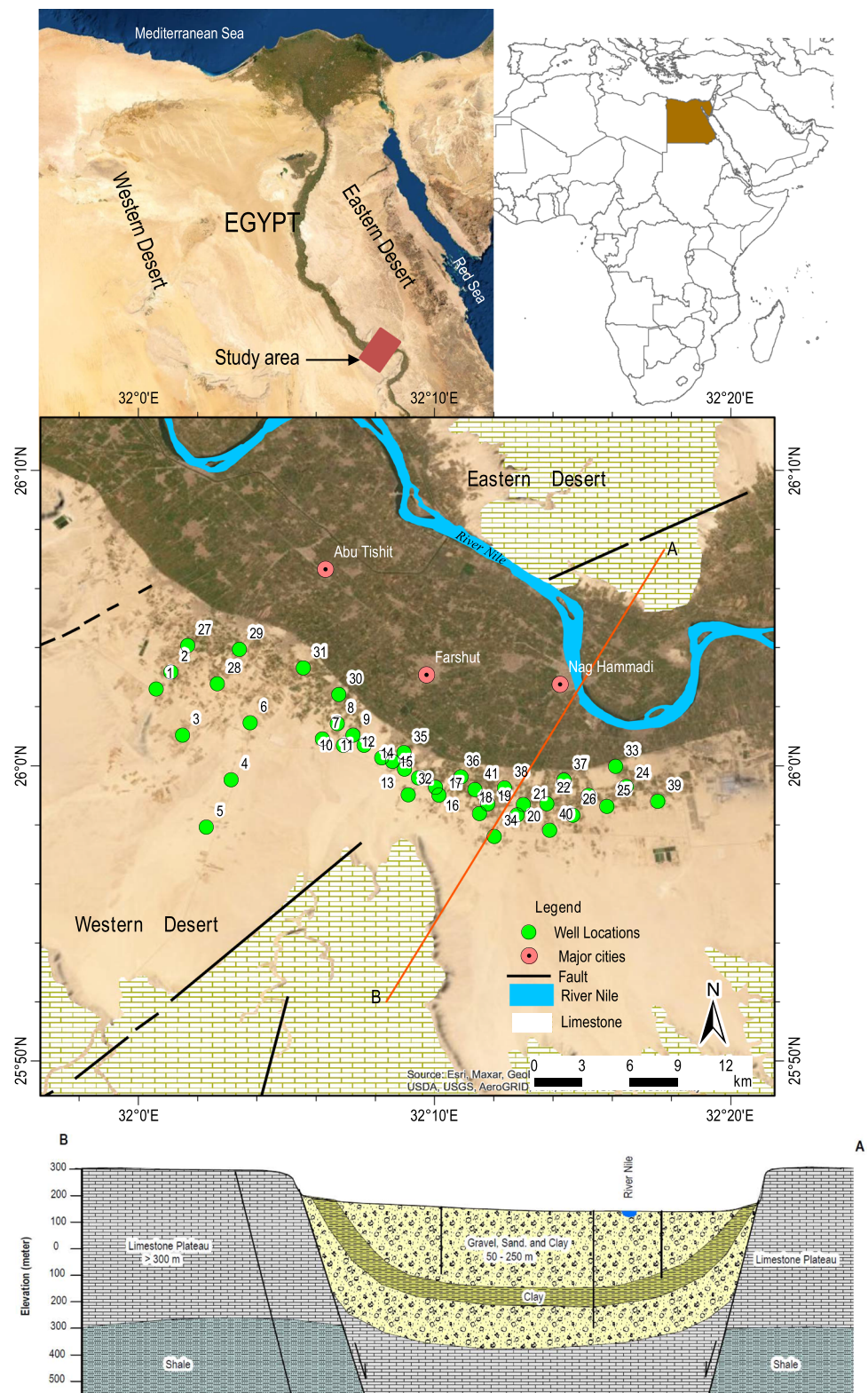
A basic knowledge of water quality is very useful for irrigation management and is an important consideration in the design and operation of the irrigation system in reclaimed lands. Therefore, water quality issues and their management need to be well studied. The objective of this paper is to employ hydrochemical approaches to get a broad picture of groundwater suitability for domestic and irrigation purposes. Also, being expected to provide information might be quite valuable in terms of groundwater sustainability in an intensive agricultural area.

Study area

Description

The study area is part of the Nile Valley and is located to the north of Qena city in Upper Egypt. The study area extends from the River Nile main channel to the high land of the limestone plateau in the Western Desert of Egypt. It is located between latitudes of 25°45' to 26°20' N and longitudes of 31°45' to 32°30' E as shown in Fig. 1. The study area includes both the old lands in the Nile Valley and the new reclaimed lands outside the Nile Valley on the desert fringes. The topography of the study area increases in elevation gradually from the River Nile main channel toward the limestone plateau (Mohammed 2005). The area is characterized by arid climatic conditions with extremely hot and dry summers and little to no rain in winter. Sporadic rain events occasionally occur, but are very rare. The average annual rainfall is about 0.01 mm (Qena Metrological Station 2001–2004).

Fig. 1 Overview figure of the study area showing: world location of Egypt, physical map of Egypt shows the location of the study area, and detailed map of the study area shows the samples and the cross section (A–B) locations. (Source: Free ArcGIS base map sources)



Geology

The geomorphological units in the study area and throughout the Nile Valley can be distinguished into: (1) The young

alluvial plain, which occupies the flood plain of the recent Nile, bordering the River Nile main channel on the east and west banks (Abdel-Samie 2000; Mohammed 2005). The young alluvial plain is almost flat and gently slopes from

south to north. The surface of such plain is underlain by silt and clay deposits. (2) The old alluvial plain, which bounds the young alluvial plain on both sides of the Nile Valley and is underlain by mixed sands and gravels that have commonly developed into terraces that represent the former Nile levels during the Pleistocene period. The terraces are not typically preserved in a single location since they are frequently removed by water erosion (Ball 1910; Said 1962; Abdel-Samie 2000; Mohammed 2005). (3) The calcareous structural plateau, which is made up of tertiary limestone. The plateau has an irregular surface and rises to more than 450 m above sea level at the study area. The surface of the plateau is dissected by complex drainage patterns which drain toward the River Nile (Abdel-Samie 2000; Mohammed 2005).

The stratigraphic succession in the study area is generally composed of a variety of sedimentary rocks belonging to a wide range of ages. The top portion of the sedimentary section is dominated by carbonate rocks belonging to the Eocene age. The lower portion is mainly Mesozoic clastics, the Nubian Sandstone formation. The surface of the study area is overlain by Pliocene and Quaternary fluvial silty clay, sand, and gravel (Said 1991; Awad et al. 1997). Near the River Nile bank, the area is covered by the Nile mud and silt with little admixture of sand (Mohammed 2005).

The geological structure of the study area is part of the entire Nile Valley, which is mostly influenced by wrench faults that run more or less parallel to either the Gulf of Suez or the Gulf of Aqaba direction (Youssef 1968). The majority of the faults are normal fault-oriented NW – SE and NE – SW. They are notably affecting the Eocene limestone with downthrow toward the Nile Valley. They create the major graben, which contains the River Nile and its valley (Fig. 1) (Said 1962). There are also large joints associated with these faults (Said 1962). The existing complex structural elements may cause a hydraulic connection between deep aquifers and the quaternary aquifer in the study area.

Hydrogeology

Qena region represents an area of most extensive groundwater resources in Upper Egypt, where significant amounts of groundwater have been exploited and used in reclamation. In recent years, farmers in the old cultivated and new reclaimed lands have started to dig new wells to alleviate surface water shortages and the tail ends of irrigation canals. Tubewells have also been the only source of irrigation water in small-scale reclamation schemes. In most cases, wells are constructed on the initiative of farmers. Farmers share their wells either individually or in groups. In addition, large-scale desert reclamation projects that rely on groundwater have been expanded and developed (Awad et al. 1997; Mohammed 2013; Mohammed et al. 2016).

The Nile Valley Aquifer System represents the main source of groundwater in the Nile Valley plains of the Qena area. It mainly consists of graded sand and gravels, overlain in the majority of the region by clays and silts of the Holocene age. The sand and gravel water bearing unit varies in thickness from one place to another, with a range from 20 to 300 m, with maximum thickness near the center of the Nile Valley and decreasing towards its edges, and underlain by impermeable compact Pliocene plastic clays (Awad et al. 1997; Mohammed et al. 2016). The silt and clay layer has variable permeability and acts at many locations as a semi-confining layer. Its thickness ranges from 1 to 24 m, and it is missing at the desert fringes where sand and gravel constitute an unconfined aquifer (Idris and Nour 1990; Campos 2009; Mohammed et al. 2016). The hydraulic conductivity of this aquifer varies from 30 to 100 m/day with a regional average of 50 m/day (RIGW 1988; Awad et al. 1997). Recharge of the aquifer system is mainly by the vertical infiltration of the irrigation return flow water and seepage from irrigation canals. Recharge from rainfall precipitation is generally of minor significance. Another possible source of recharge is the vertical flow from deeper aquifers (most notably the Nubian Aquifer). Discharge of the aquifer system is either through the lateral outflow to the River Nile or vertically through pumping for irrigation (RIGW and IWACO 1986; Hamza et al. 1999; Mohammed et al. 2016). The regional groundwater flow of the Nile Valley Aquifer takes a longitudinal movement parallel to the River Nile flow. Transversal groundwater flow takes place from the reclaimed lands toward the cultivated lands in the flood plain, and then near to the River Nile channel, groundwater drains into the River Nile (RIGW 1986; Awad et al. 1997; Hamza et al. 1999; Mohammed et al. 2016). The groundwater levels were measured when possible in various producing wells in the study area. Under the cultivated lands, the groundwater depth is less than 10 m and increases toward the desert fringes where the reclaimed lands to reach about 46 m.

Agricultural activities

Agriculture is the primary use of lands within the study area. More than 81% of the inhabited Nile Valley is under cultivation in the Qena province, where the study area is part of (SEAM 2005). The study area is characterized by intensive agriculture activities, where multiple crops planted throughout the year. The principal crops grown are wheat, sugarcane, clover, corn, fava beans, and vegetables. The main agricultural challenges in the study area include the high cost of reclamation, limited water supplies, and a relatively rapid population growth rate. Groundwater is the major source for irrigation in the reclaimed fields, where the old cultivated lands are usually irrigated by irrigation canals that

dissect the alluvial plains. No large irrigation schemes were constructed across the study area, while the irrigation habits comprise both traditional and modern irrigation practices. The vast majority of the agricultural lands are irrigated using the traditional surface (flood) irrigation system. Modern and high water use efficiency irrigation methods like drip and sprinkler are less widespread. The potential impacts of the surface irrigation practices include an increase in return flow rates. Irrigation return flow is the quantity of water which infiltrates back (returns) to the groundwater after irrigation of agricultural fields. Irrigation return flow water could be a significant source of groundwater recharge in shallow aquifers, but it could also transmit nutrients and pollutants, which may result in groundwater quality problems in the aquifer (Mohammed et al. 2016).

Large quantities of commercial fertilizer, together with manure, are applied during the growing seasons. The most regularly used commercial fertilizers are N, P, K, and S fertilizers, which include urea (46.5% N), ammonium nitrate (33.5% N), ammonium sulfate (20.6% N), calcium nitrate (15.5% N), superphosphate (15% P_2O_5), and potassium sulfate (50% K_2O) (FAO 2005). However, N and P fertilizers are the most regularly utilized in the study area, and generally Egypt has one of the highest rates of nitrogen application in the world (FAO 2005). The application rates of these chemical fertilizers are described as extensive to moderate in the study area. Pesticides of various varieties are widely used in the agricultural activities in the study area. Pesticides are used to protect agricultural crops from insects, disease fungus, and weeds. Pesticides, on the other hand, could be easily leached from plants and soil into the shallow groundwater during irrigation.

In the recent few decades, the study area has experienced fast land cover changes (e.g., increased agricultural areas). We mapped the land-cover changes due to agricultural development and urban expansion using the Google

Earth software platform. Google Earth enables users to freely examine satellite imagery with medium, high, and very high spatial resolution. Google Earth has enabled many users wishing to monitor land (foresters, conservationists, land cover, etc.) to bypass the need of remote sensing experts to monitor their land (Sidhu et al. 2018; Floreano and de Moraes 2021). Figure 2 depicts the change in land cover in the study area over a 35-year period. One can visually notice the extensive land-cover changes. A rough estimation of the land-cover changes in the study area indicates that the agricultural and urban areas increased from 33.17 Km^2 in 1985 to 218.14 km^2 in 2020. The continuous expansion in agricultural and urban areas in the study area is reliant on groundwater resources where the surface water (River Nile and irrigation canals) is not available. These agricultural developments, as well as high irrigation rates under arid conditions, are predicted to have an impact on groundwater quality in the study area.

Materials and methods

Sample collection and analysis

Forty-one groundwater samples were collected from the investigated area in March 2018. The sampling plan focused on the reclaimed lands. The locations (latitude and longitude) of the collected samples are located using a handheld global positioning system (GPS) device in the field and presented in Fig. 1. Samples were collected directly from tubewells after sufficient time of pumping to ensure that the water sample represented the original groundwater source rather than water retained in pipes. Samples were collected using new, dry, and clean polyethylene bottles of different sizes. Before collecting the samples, the bottles were washed thoroughly with well water. At each locality, one sample

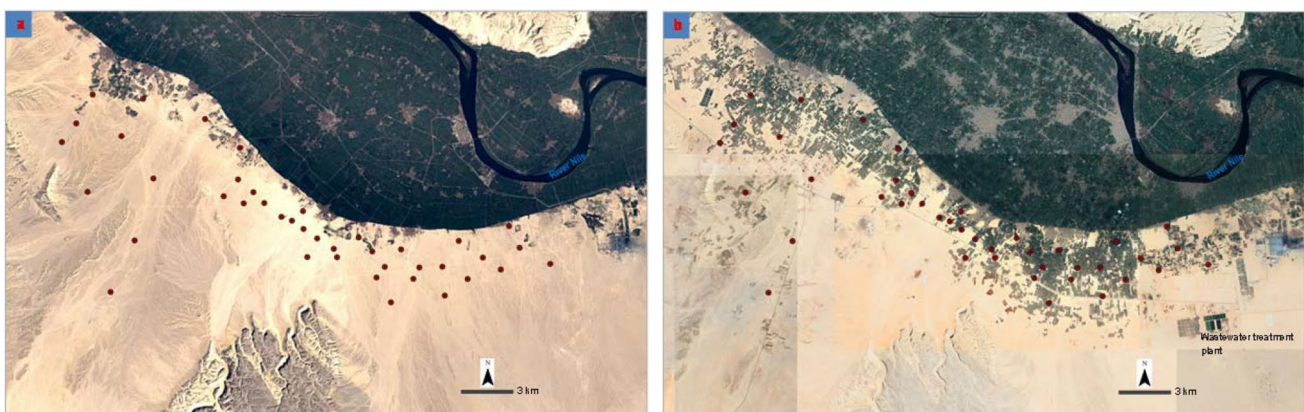


Fig. 2 Map showing the vast changes in the land cover in the study area over a period from 1985 (a) to 2020 (b). Source of the maps are Google Earth free application

bottle was collected without headspace for alkalinity and anion measurements; second sample bottle was collected and filtered on site using a 0.45 micron filter and acidified with nitric acid (HNO₃) to preserve dissolved metals. Cations were analyzed from these acidified bottle samples. Electrical conductivity (EC), hydrogen ion concentration (pH), and field temperature (°C) were measured in the field by using portable meters (JENWAY Model 430). All of the collected sample bottles were carefully labeled, tightly sealed, and stored in a refrigerator at 4 °C.

The chemical analyses of the collected samples were analyzed in the central laboratory of the Company for Drinking Water and Sanitation at Qena, Egypt. Titrations were used to determine alkalinity, hardness, and chloride. Flame photometer instrument (JENWAY PFP7) was used to analyze Na⁺ and K⁺ ions. Sulfate and nitrate levels were analyzed by using spectrophotometric methods (HACH DR 5000). The concentrations of the analyzed chemical constituents are reported in milligrams per liter (mg/L). As required, a milliequivalent per liter (meq/L) concentration is calculated.

As a basic and simple check of the quality of the chemical analysis in the data set, the charge balance error (CEB) is calculated. According to Freeze and Cherry (1979) and Kehew (2001), the percent charge-balance error is calculated based as follows (Eq. 1):

$$\% \text{ Charge Balance Error (CBE)} = \left(\frac{\sum \text{Cations} - \sum \text{Anions}}{\sum \text{Cations} + \sum \text{Anions}} \right) \times 100 \quad (1)$$

where the ion concentration unit is in milliequivalent per liter (meq/L). The % CBE value will have either a positive or negative charge, depending upon whether the cations or anions are larger. The groundwater chemistry results may be good if the CBE is < ±5%, and if the CBE is > ±5%, it will be considered as poor (Hounslow 1995; Rahman et al. 2021a). However, the CBE of up to ±10% is acceptable (Rahman et al. 2021a). The minimum, maximum, and average charge balance errors are −6.7%, 4.9%, and −0.2%, respectively, in the collected samples in this study. Three samples only have charge balance errors of −6.7%, −6.0%, and −5.9%. The charge balance errors were considered to be reliable in this research work.

Water quality assessment methods

Water quality index

Various methodologies were used to better understand the data sets of the water quality parameters. The water quality index (WQI) is a simple and powerful tool for evaluating water quality status. The WQI aims to provide a single value of the water quality status by using the water constituents and their concentrations present in a sample (Abbasi and

Abbasi 2012). The water quality indices (WQIs) have been used as a tool to assess the state of water quality since the 1960s (Horton 1965). Several WQIs have been produced by various researchers and agencies around the world. However, there is no WQI that has been globally preferred.

The WQI used in this work is the weighted arithmetic average method, developed by Brown et al. (1970). The method is one of the most effective tools for the evaluation of groundwater quality and is used for the evaluation of groundwater quality worldwide. This method has been used and described by various researchers throughout the world (Sadat-Noori et al. 2014; Batabyal and Chakraborty 2015; Sutadian et al. 2016; Gradilla-Hernández et al. 2020; Reyes-Toscano et al. 2020). Four common steps have been used in the WQI method:

In the first step: for the purpose of calculation, 11 water quality parameters have been selected (pH, TDS, Total Hardness (TH), Na⁺, K⁺, Ca²⁺, Mg²⁺, HCO₃⁻, Cl⁻, SO₄²⁻, NO₃⁻), and an individual weight (*w_i*) was assigned to each of the 11 parameters based on its importance in the evaluation of water quality. The individual weight assigned ranges from 1 (lowest) to 5 (highest).

In the second step: a relative weight (*W_i*) of each given parameter is calculated using the following Eq. (2):

$$W_i = \frac{w_i}{\sum_{i=1}^n w_i} \quad (2)$$

where *W_i* is the relative weight of the *i*th parameter, *w_i* is the weight of each parameter, and *n* is the number of parameters. Table 1 lists the individual weight and the relative weight of the selected parameters.

In the third step: the quality rating for each parameter is calculated from the following Eq. (3):

Table 1 Individual weight and relative weight of the parameters to calculation of WQI

Parameter	Individual weight (w _i)	Relative weight (W _i)	WHO, (2011)
pH	4	0.1000	8.5
TDS	5	0.1250	1000
TH	4	0.1000	500
Na ⁺	4	0.1000	200
K ⁺	2	0.0500	12
Ca ²⁺	3	0.0750	200
Mg ²⁺	3	0.0750	150
HCO ₃ ⁻	1	0.0250	200
Cl ⁻	4	0.1000	250
SO ₄ ²⁻	5	0.1250	250
NO ₃ ⁻	5	0.1250	50
Total	40	1.0000	

$$q_i = \left(\frac{C_i}{S_i} \right) 100 \tag{3}$$

where q_i is the quality rating, C_i is the concentration of each parameter, and S_i is the concentration of the permissible limits of the World Health Organization (WHO, 2011) (Table 1).

In the fourth step: the sub-index of i th parameter (SI_i) is calculated for each parameter using the following Eq. (4):

$$SI_i = W_i q_i \tag{4}$$

where SI_i is the sub-index of i th parameter, q_i is the quality rating, and W_i is the relative weight. Then the WQI is calculated from the following Eq. (5):

$$WQI = \sum SI_i \tag{5}$$

The computed WQI values are classified into five categories: excellent water, good water, fair water, poor water and unacceptable water for human consumption (Table 2) (Sadat-Noori et al. 2014; Reyes-Toscano et al. 2020).

Irrigation purposes

In order to evaluate the suitability of groundwater in the study area for irrigation uses, multiple irrigation quality indices were calculated, including salinity hazard (EC), sodium adsorption ratio (SAR), soluble sodium percentage (SSP), residual sodium carbonate (RSC), magnesium hazard (MH), permeability index (PI), and Kelley's ratio (KR). These indices can be calculated as follows:

Sodium adsorption ratio (SAR) is computed using the absolute and relative concentrations of main cations (US Salinity Laboratory Staff 1954; Raghunath 1987; Abdel-Satar et al. 2017), with concentrations provided in milliequivalents per liter (Eq. 6).

$$SAR = \frac{Na^+}{\sqrt{\frac{Ca^{2+} + Mg^{2+}}{2}}} \tag{6}$$

Soluble sodium percentage (SSP), commonly known as sodium percentage (Na%), is an estimation of the sodium hazard in irrigation water that expresses the percentage of

sodium out of the total cation concentrations (Wilcox 1955; Todd 1980; Reddy 2012). SSP is calculated using the following Eq. (7) where concentrations are reported in milliequivalents per liter.

$$SSP\% = \frac{Na^+ + K^+}{Ca^{2+} + Mg^{2+} + Na^+ + K^+} \times 100 \tag{7}$$

Residual sodium carbonate (RSC) is used to evaluate the hazard effects of bicarbonate and carbonate on groundwater quality for irrigation usage (Eaton 1950; US Salinity Laboratory Staff 1954; Todd 1980). RSC is calculated using the following Eq. (8) where the concentrations are reported in milliequivalents per liter.

$$RSC = (HCO_3^- + CO_3^{2-}) - (Ca^{2+} + Mg^{2+}) \tag{8}$$

Magnesium hazard (MH), also known as magnesium adsorption ratio (MAR), is used to assess the impact of high magnesium concentrations on irrigation water (Szabolcs and Darab 1964; Todd 1980; Ayers and Westcot 1985; Raghunath 1987). It is calculated using the Eq. (9), with the concentrations expressed in milliequivalents per liter.

$$MH\% = \frac{Mg^{2+}}{Ca^{2+} + Mg^{2+}} \times 100 \tag{9}$$

Permeability index (PI) measures how easily water flows through a medium. The following Eq. (10) is used to calculate PI (Doneen 1961, 1964), where the concentrations are in milliequivalents per liter.

$$PI\% = \frac{Na^+ + \sqrt{HCO_3^{3-}}}{Ca^{2+} + Mg^{2+} + Na^+} \times 100 \tag{10}$$

Kelley's ratio (KR) is comparing the sodium ion to the calcium plus magnesium ions. KR is calculated using the following Eq. (11) (Kelley 1951, 1963), where the concentrations are in milliequivalents per liter.

$$KR = \frac{Na^+}{Ca^{2+} + Mg^{2+}} \tag{11}$$

Results and discussion

Hydrochemical characteristics

The physical and chemical parameters, including pH, the total dissolved solids (TDS), and major ions of the collected groundwater samples are summarized in Table 3. The physiochemical parameters of the groundwater samples in the study area showed considerable variability in close proximity to each other, which might be related to various

Table 2 Classification of water quality index

WQI	Water quality	References
< 50	Excellent	Sadat-Noori et al. (2014)
50–100	Good	Reyes-Toscano et al. (2020)
100–200	Fair	
200–300	Poor	
> 300	Unacceptable	

Table 3 Statistical summary of hydrochemical parameters analyzed in groundwater samples in the study area

Parameter	Unit	Minimum	Maximum	Average	Standard deviation	WHO standard (2011)	No. of samples exceeding WHO standard	Egypt standard (1995, 2007)	No. of samples exceeding Eg. standard
T	°C	26	35	30	2	–	–	–	–
pH		7.4	8.2	7.7	0.18	6.5–8.5	–	6.5–8.5	–
EC	μS/cm	955	8855	2573	1475	–	–	–	–
TDS	mg/L	621	5756	1673	959	1000	30	1000	30
TH	mg/L	160	2310	504	369	500	18	500	18
Ca ²⁺	mg/L	25	448	104	82	200	4	200	4
Mg ²⁺	mg/L	14	286	59	45	150	1	150	1
K ⁺	mg/L	4	14	8	2	12	2	–	–
Na ⁺	mg/L	129	1370	428	245	200	35	200	35
HCO ₃ [−]	mg/L	47	434	268	71	200	32	250	27
Cl [−]	mg/L	110	2710	645	482	250	32	250	32
SO ₄ ^{2−}	mg/L	99	790	293	184	250	15	400	10
NO ₃ [−]	mg/L	0.1	257	53	58	50	12	50	12

geological origins and anthropogenic influences. The chemical composition of the natural groundwater is controlled by numerous factors, including precipitation, anthropogenic inputs, water–rock interaction, the evaporation process, residence time, and mixing (Clark and Fritz 1997; Mook 2000; Dotsika et al. 2010). The concentrations of cations and anions are represented in Fig. 3. The groundwater in the study area is alkaline/basic in character (pH from 7.39 to 8.22). The sources of the alkaline pH could be the levels of the hard water minerals and the effluents from agricultural activities. The TDS of the groundwater samples in the study area ranges from 620 to 5755 mg/L, with an average of 958 mg/L. TDS levels indicate the influence of the dissolution of the halite (NaCl) and gypsum (CaSO₄·2H₂O) of the adjacent carbonate plateau. The dissolution of limestone, dolomite, gypsum, and halite is also the principal source of the Ca²⁺, Mg²⁺, Na⁺, K⁺, CO₃^{2−}, SO₄^{2−}, and Cl[−] content in the groundwater (Somay and Gemic 2012; Pazand et al. 2018). Few boreholes distributed in the study area show a TDS of fresh water (TDS < 1000 mg/L).

Of most of the collected groundwater samples, the Na⁺ and Cl[−] ions had the highest concentrations of all the major elements. The concentration of Na⁺ ranges from 129 to 1370 mg/L, with an average of 428 mg/L, and the concentration of Cl[−] ranges from 110 to 2710 mg/L, with an average of 645 mg/L. Such a wide range of concentrations suggests that multiple sources generate that chemical composition. The Na⁺·Cl[−] relationship in the groundwater samples of the study area shows similarities in concentrations of both Na⁺ and Cl[−] ions. Figure 4 shows the value of Cl[−] as a function of Na⁺ in the groundwater samples in the study area, and there is a strong correlation between them ($R^2 = 0.92$). The average Na⁺/Cl[−] ratio of the groundwater

samples in the study area is 1.18, suggesting the same sources of both Na⁺ and Cl[−] ions, and illustrating the considerable influence of the dissolution of evaporates, such as halite, on groundwater chemistry. In addition, it indicates the less prevalent nature of the ion exchange interaction between divalent cations and sodium in the groundwater geochemical processes.

The major water type and dominating ionic concentrations of the collected groundwater samples are described and plotted on Chadha's diagram (Chadha 1999) as shown in Fig. 5. This diagram clearly shows that the majority of the groundwater samples ($\approx 95\%$) are of Na⁺·Cl[−] water type. Few samples are of Ca²⁺·Mg²⁺·Cl[−] water type.

The Cl[−]/sum anions ratio is a valuable ratio for determining the source of groundwater (Hounslow 1995). When the ratio is > 0.8 and the TDS > 500 mg/L, then the source of groundwater is seawater, brine, or evaporates; when the ratio is > 0.8 and the TDS < 100 mg/L, then the source of groundwater is rainwater; and when the ratio is < 0.8, then the source of groundwater is rock weathering. This ratio is less than 0.8 in almost all of the collected groundwater samples of the study area, which indicate the weathering processes and mineral dissolution from the nearby sedimentary rocks.

The plot of (Ca²⁺ + Mg²⁺) versus (HCO₃[−] + SO₄^{2−}) (Fig. 6) shows that the majority of the samples are close to the 1:1 trend line, indicating that the dominant reactions in the system are the dissolutions of evaporates (Srinivasamoorthy et al. 2014; Pazand et al. 2018). The chloro-alkaline indices (CAIs) are used to assess the ion exchange reaction between groundwater and its host rock (Schoeller 1967). Two chloro-alkaline indices (CAI 1, 2) are used to identify the ion exchange processes and are calculated using the

Fig. 3 Concentrations of major ions in the study area **a** for cations and **b** for anions

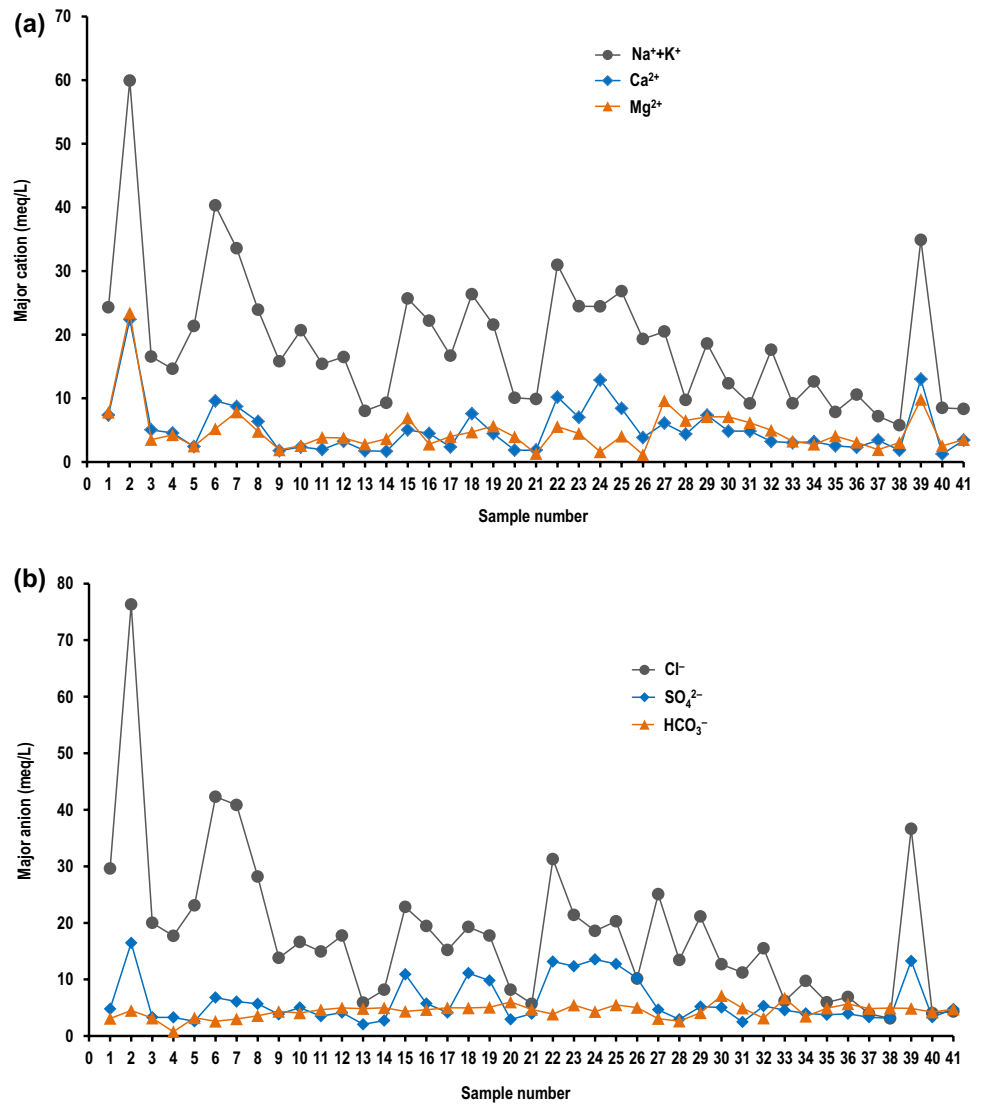


Fig. 4 Plot of Cl^- versus $\text{Na}^+ + \text{K}^+$ in groundwater samples from the study area

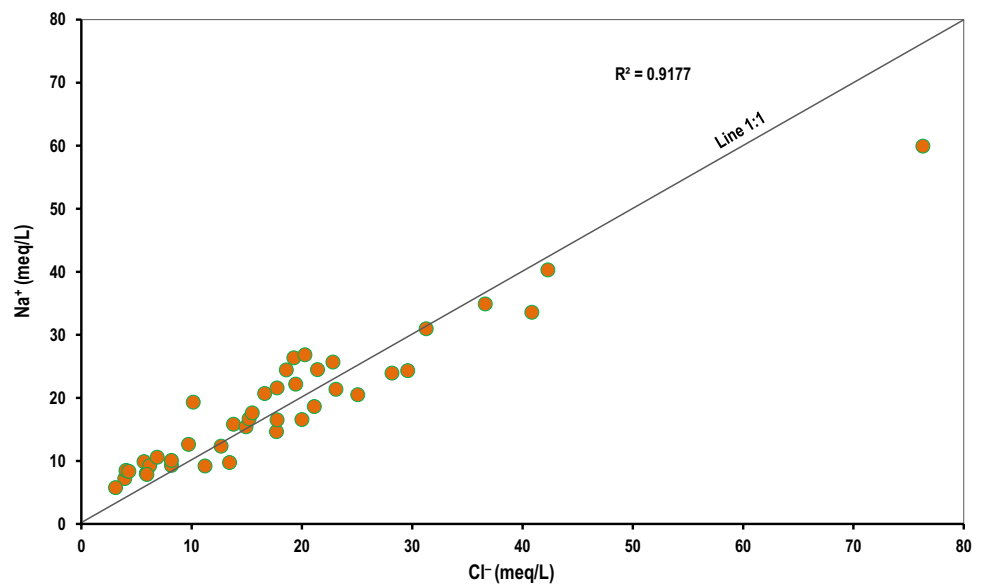


Fig. 5 Chadha's classification diagram explaining the chemical groundwater type of the collected samples (Chadha 1999)

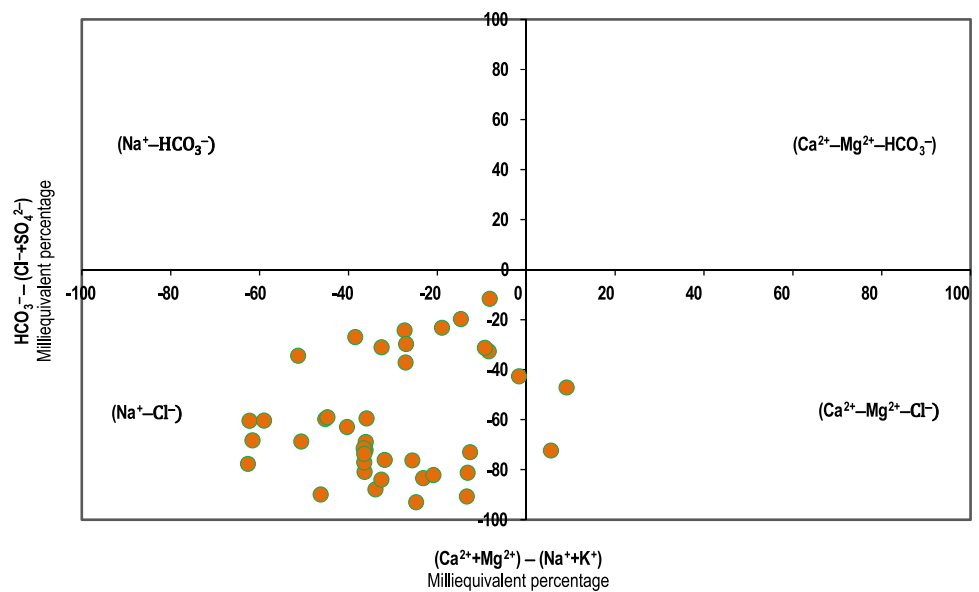
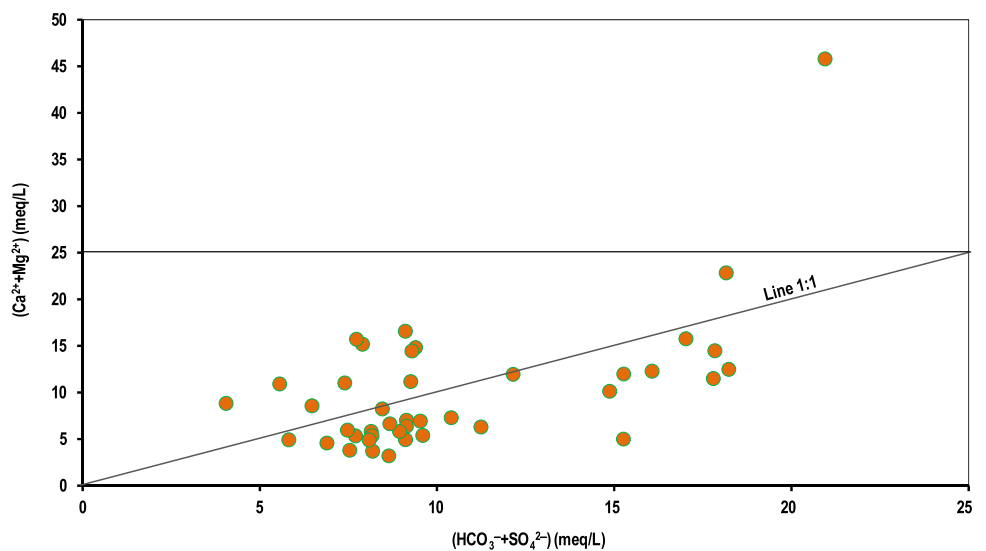


Fig. 6 Plot of (HCO₃⁻+SO₄²⁻) versus (Ca²⁺+Mg²⁺) in groundwater samples from the study area



following Eqs. (12, 13) where the concentrations are in milliequivalents per liter.

$$CAI1 = \frac{Cl^{-}-(Na^{+} + K^{+})}{Cl^{-}} \quad (12)$$

$$CAI2 = \frac{Cl^{-}-(Na^{+} + K^{+})}{SO_4^{2-} + HCO_3^{-} + CO_3^{2-} + NO_3^{-}} \quad (13)$$

When Na⁺ and K⁺ ions in groundwater are exchanged with Ca²⁺ or Mg²⁺ ions that occur in the aquifer materials (direct ion exchange), both indices will be positive and the dissolved concentrations of Ca²⁺ and Mg²⁺ will increase in the groundwater. If the indices are negative, a possible

reverse ion exchange occurs between the Na⁺ and K⁺ ions contained in the aquifer materials and the Ca²⁺ or Mg²⁺ ions dissolved in the groundwater, and then, the concentrations of Na⁺ and K⁺ will increase in the groundwater. The CAI 1 and 2 indices are calculated for the collected groundwater samples from the study area, and both are similar in terms of positive and negative samples. The CAI values are scattered in a small range without any prevalence of direct or reverse ion exchange in the samples (Fig. 7). The CAI results in the study area might be an indication of the different sources of the dissolved ions in groundwater, including dissolution of the aquifer matrix, adjacent rock weathering, agricultural activities, urbanization, and industrial activities.

Fig. 7 Plot of CAI 1 and 2 of groundwater samples from the study area

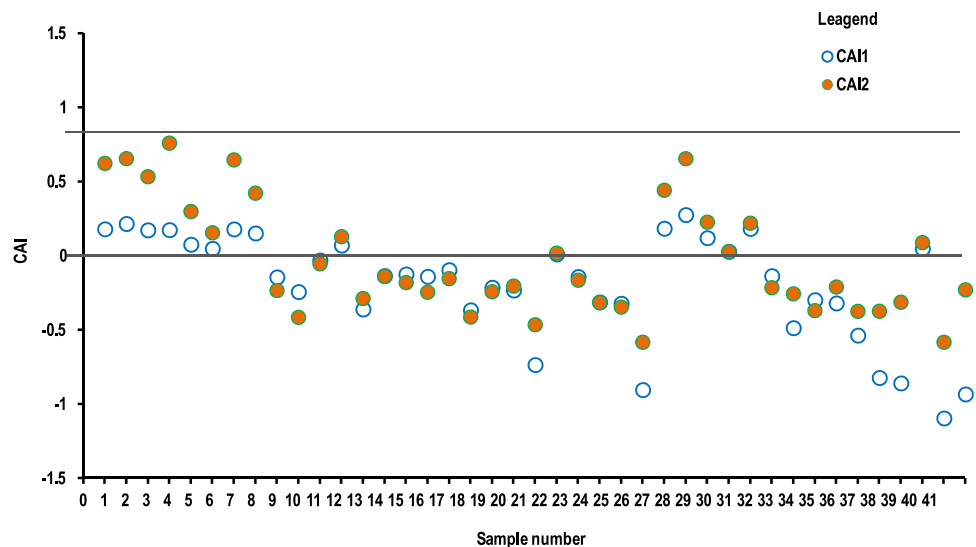


Table 4 WQI for the groundwater samples in the study area

Classification of WQI		Number of samples	References
WQI	Water quality		
<50	Excellent	0	0%
50–100	Good	16	39%
100–200	Fair	20	49%
200–300	Poor	4	10%
>300	Unacceptable	1	2%

Water quality index

The WQI method is widely used for groundwater quality assessment around the world. In this study, the WQI calculated based on the hydrochemical data and the WHO (2011) standards for drinking purpose were considered for the calculation of the WQI. The calculated WQI values ranged from 53.3 to 447.5 with an average of 129.8. Table 4 represents the WQI categories and percentage of samples in each category. The results obtained from the groundwater samples from the study area showed that more than 50% of the samples are fair to poor water quality, while around of 39% are of good water quality (Table 4).

Drinking water quality

The suitability of the groundwater for domestic uses is directly related to different physiochemical parameters and reactions in groundwater. The natural baseline geochemistry of groundwater resulting from interaction with rocks, in addition to anthropogenic sources, could create widespread health and acceptability problems. In the present work, we evaluated the groundwater suitability for domestic uses in the study area through different parameters, as well

as comparing the ion concentrations by the World Health Organization (WHO 2011) guideline values and the Egyptian guideline values (EMH 1995; EHCW 2007).

TDS is one of the main and important parameters in terms of drinking water. The quality of the taste is primarily related to the total amount of solids in groundwater. Based on the TDS classification given by Freeze and Cherry (1979) and Davis and Dewiest (1966), the groundwater samples of the study area are described as fresh and brackish groundwater types, as represented in Table 5. The TDS of brackish water is dominant in the study area, accounting for about 73% of the total samples (Fig. 8). The TDS values and the concentrations of the major ions in groundwater samples are compared with the WHO (2011) (Elumalai et al. 2020) and Egyptian (EMH 1995; EHCW 2007) drinking water guidelines and given in Table 3. High TDS indicates that groundwater may contain elevated levels of ions that are above the primary or secondary drinking water standards, such as elevated levels of nitrate and other trace constituents.

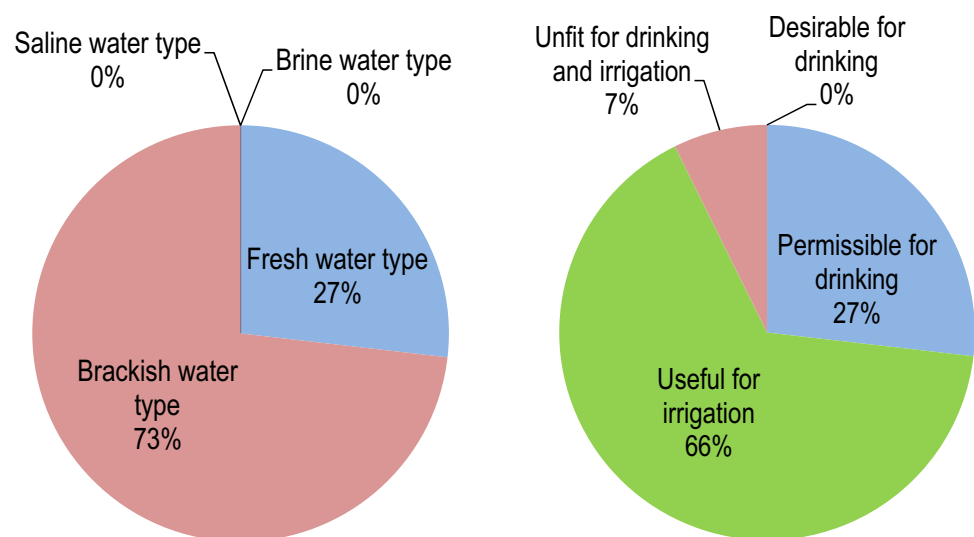
One commonly important aspect of water quality that is mostly caused by the presence of calcium and magnesium ions in water is the water hardness. Calcium carbonate is one of the main causes of hardness, so total hardness is usually expressed as equivalents of calcium carbonate, or mg/L as CaCO₃. Total hardness (TH) can be calculated from individual concentrations of calcium and magnesium in mg/L using the following Eq. (14) (Kehew 2001):

$$TH = 2.5(\text{concentration of Ca}^{2+} \text{ in mg/L}) + 4.1(\text{concentration of Mg}^{2+} \text{ in mg/L}) \tag{14}$$

The total hardness of the collected groundwater samples is calculated using Eq. (14) and is listed in Table 5. The hardness values in the collected groundwater samples from the study area range from 159 to 2291 mg/L

Table 5 Classification of groundwater samples in the study area based on TDS and TH

Component	Severity	Classification	Number of samples	Reference
TDS mg/L	< 1000	Fresh water type	11 27%	Freeze and Cherrey (1979)
	1000–10,000	Brackish water type	30 73%	
	10,000–100,000	Saline water type	0 0	
	> 100,000	Brine water type	0 0	
TDS mg/L	< 500	Desirable for drinking	0 0	Davis and Dewiest (1966)
	500–1000	Permissible for drinking	11 27%	
	1000–3000	Useful for irrigation	27 66%	
	> 3000	Unfit for drinking and irrigation	3 7%	
TH mg/L as CaCO ₃	0–60	Soft	0 0	Durfor and Becker (1964)
	61–120	Moderately hard	0 0	
	121–180	Hard	1 2%	
	> 180	Very hard	40 98%	
TH mg/L as CaCO ₃	0–75	Soft	0 0	Todd and Mayers (2005)
	75–150	Moderately hard	0 0	
	150–300	Hard	14 34%	
	> 300	Very hard	27 66%	

Fig. 8 Classification of groundwater samples based on TDS. Left based on Freeze and Cherrey (1979) and right based on Davis and DeWiest (1966)

as CaCO₃, with an average of 504 mg/L as CaCO₃. The degree of hardness in water is classified in terms of its calcium carbonate concentration according to Durfor and Becker (1964), and Todd and Mays (2005) after Sawyer and McCarty (1967). Based on the previous given classifications of TH, the groundwater samples in the study area range from hard to very hard groundwater (Table 5 and Fig. 9). The high levels of TH in the study area are attributed to the weathering of carbonate rocks from the nearby plateau and the excessive use of lime fertilizers during the agriculture activities. Long-term consumption of hard water can cause many diseases and health dysfunctions.

Livestock and poultry water quality

Domestic animals are an integral part of agricultural societies and provide a valuable source of food. Irrigation water supplies frequently serve as a drinking water source for livestock in the study area. For livestock, fundamentally the same water quality requirements hold as for human consumption. However, animals can drink water with moderately high dissolved solids (EC less than 5 dS/m) (Ayers and Westcot 1985). It is well known that excessively saline water can cause physiological distress or death in livestock. The National Academy of Sciences and National Academy

Fig. 9 Classification of groundwater samples based on TH. Left based on Durfor and Becker (1964) and right based on Todd and Mayers (2005)

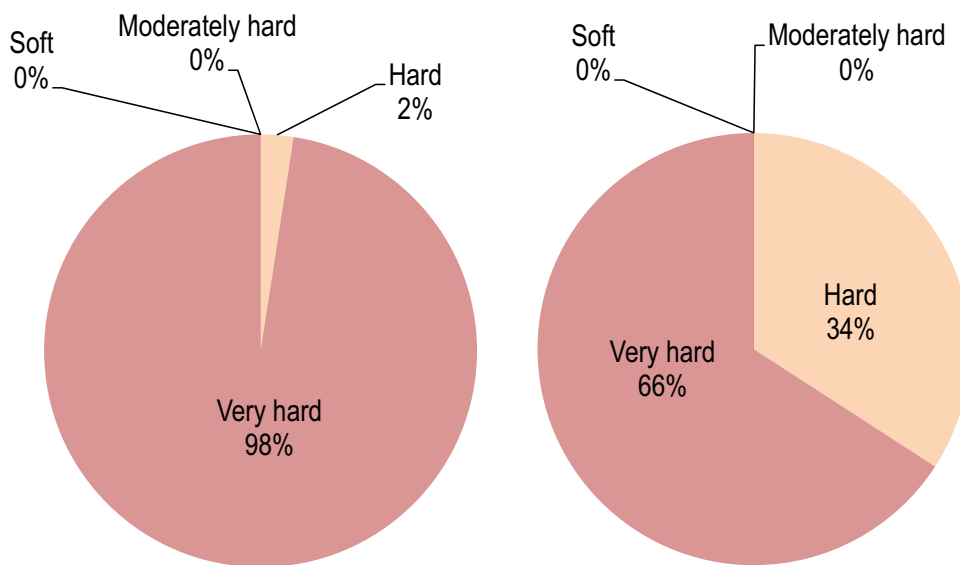


Table 6 Guide to use of saline water for livestock and poultry of groundwater samples in the study area

Parameter	Severity	Hazard and limitation	Number of samples		Reference
TDS mg/L	Less than 1000	Excellent for all classes of livestock and poultry	11	27%	NAS-NAE (1972)
	1000–2999	Very satisfactory for all classes of livestock and poultry	27	66%	
	3000–4999	Satisfactory for livestock	2	5%	
	5000–6999	Not acceptable for poultry	1	2%	
	7000–10,000	Unfit for poultry. Avoid use for all animals if possible	0	0	
	Over 10,000	Not recommended for use under any conditions	0	0	

of Engineering (NAS-NAE 1972) established a guideline for evaluating water for livestock and poultry production (Table 6). It is also recommended by NAS-NAE that water containing a TDS of 3000 mg/L or less should be satisfactory for livestock under almost any circumstance. Based on the preceding criteria, the groundwater in the study area is generally satisfactory for livestock (TDS < 5000 mg/L) (Fig. 10).

Irrigation water quality

The suitability of water for irrigation is determined by its mineral constituents and the type of the plant and soil to be irrigated. These minerals could be derived from natural processes and anthropogenic activities, and they might affect crop yields and soil fertility. It is therefore important to have detailed information concerning the quality of irrigation water and its impact on crops and soils. The suitability of water for irrigation is determined not only by the total amount of salt present but also by the kind of salt. The chemical parameters that are commonly used to determine the suitability of the water for irrigation, as well as their impact on crop productivity and soil quality are discussed below.

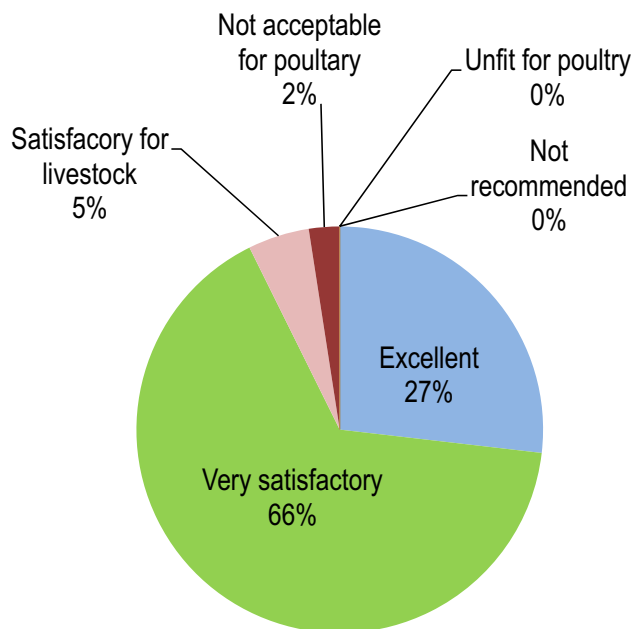


Fig. 10 Classification of groundwater samples for livestock in the study area

Salinity hazard (EC)

A salinity problem occurs when salt accumulates in the crop root zone to the point where the crop is unable to draw enough water from the salty soil solution, causing water stress for an extended period of time. When water uptake is significantly reduced, the plant slows its rate of growth (Ayers and Westcot 1985). Usually, water with a TDS of about 5,000 mg/L is unsuitable for irrigation (Pillsbury and Blaney 1966; Ayers and Westcot 1985). Water is divided into four classes based on the EC in $\mu\text{S}/\text{cm}$ by the US Salinity Laboratory Staff (1954) (Table 7). As shown in Table 7, more than half of the collected groundwater samples from the study area exhibit conductivity values in the C4 class, which indicates a risk of salinity.

Sodium adsorption ratio (SAR)

The sodium adsorption ratio of a soil solution is simply related to the adsorption of sodium by soil. The main problem with high sodium concentration is its effect on soil permeability and water infiltration. Continued use of water with a high SAR leads to a breakdown in the physical structure of the soil. The sodium replaces calcium and magnesium adsorbed on the soil clay minerals and causes dispersion of soil particles. This dispersion results in the breakdown of soil aggregates and causes cementation of the soil under drying conditions, as well as prevents the infiltration of water and air into the soil (Ayers and Westcot 1985). In practice, the SAR value of the water in the soil increases due to an increase in the concentration of all salts (US Salinity Laboratory Staff 1954). The SAR can be calculated using Eq. (6).

Irrigation water is classified into four classes based on SAR values proposed by the US Salinity Laboratory Staff (1954) and shown in Table 7. According to the SAR

Table 7 Irrigation water quality parameters of groundwater samples in the study area

Index	Class	Severity	Hazard and limitation		Number of samples	References
EC $\mu\text{S}/\text{cm}$	C1	<250	Low-salinity water	TDS < 200	0	US Salinity Laboratory Staff (1954)
	C2	250–750	Medium-salinity water	200–500	0	
	C3	750–2250	High-salinity water	500–1500	20	
	C4	> 2250	Very high-salinity water	> 1500	21	
SAR	S1	< 10	Low sodium	No harmful effect	26	US Salinity Laboratory Staff (1954)
	S2	10–18	Medium sodium	Problems on fine texture soils and sodium-sensitive plants	15	
	S3	18–26	High sodium	Harmful effects could be anticipated in most soils	0	
	S4	> 26	Very high sodium	Generally unsatisfactory for irrigation purposes	0	
SSP		<20	Excellent	EC < 250	0	Todd (1980)
		20–40	Good	250–750	0	
		40–60	Permissible	750–2000	11	
		60–80	Doubtful	2000–3000	27	
		> 80	Unsuitable	> 3000	3	
RSC		< 1.25	Generally safe for irrigation		40	US Salinity Laboratory Staff (1954)
		1.25–2.5	Marginal as an irrigation source		1	
		> 2.5	Usually unsuitable for irrigation without amendment		0	
MH		< 50	Suitable for irrigation		17	Szabolcs and Darab (1964)
		> 50	Harmful and unsuitable for irrigation water		24	
PI	I	> 75	Good		17	Doneen (1964) in Das and Nag (2015)
	II	25–75	Moderate		24	
	III	< 25	Unsuitable		0	
KR		< 1	Suitable		2	Kelley (1951), Das and Nag (2015)
		> 1	Unsuitable		39	

classification, more than two-thirds (63%) of the groundwater samples collected from the study area fall into the low sodium hazard (S1 class), while the remaining samples (37%) fall into the S2 class, which groundwater could be used for irrigation with minor limitations for soils and plants (Table 7).

A diagram was proposed by Wilcox (1948) for the classification of irrigation water. This diagram has been widely used. The features of this diagram are based on the previously discussed salinity hazard (EC) and sodium adsorption ratio (SAR). Using the SAR and EC values as coordinates, locate the corresponding point on the diagram. The position of the point determines the quality classification of the water. The Wilcox diagram for the studied groundwater samples is shown in Fig. 11. In accordance with the Wilcox diagram, the groundwater samples in the study area belong to classes C3-S1 (17%), C3-S2 (27%), C3-S3 (5%), C4-S2 (10%), C4-S3 (36%), and C4-S4 (5%), indicating high salinity hazard and low, medium, and high alkali hazard, which might require special management for salinity control and good to very salt tolerance plants should be selected.

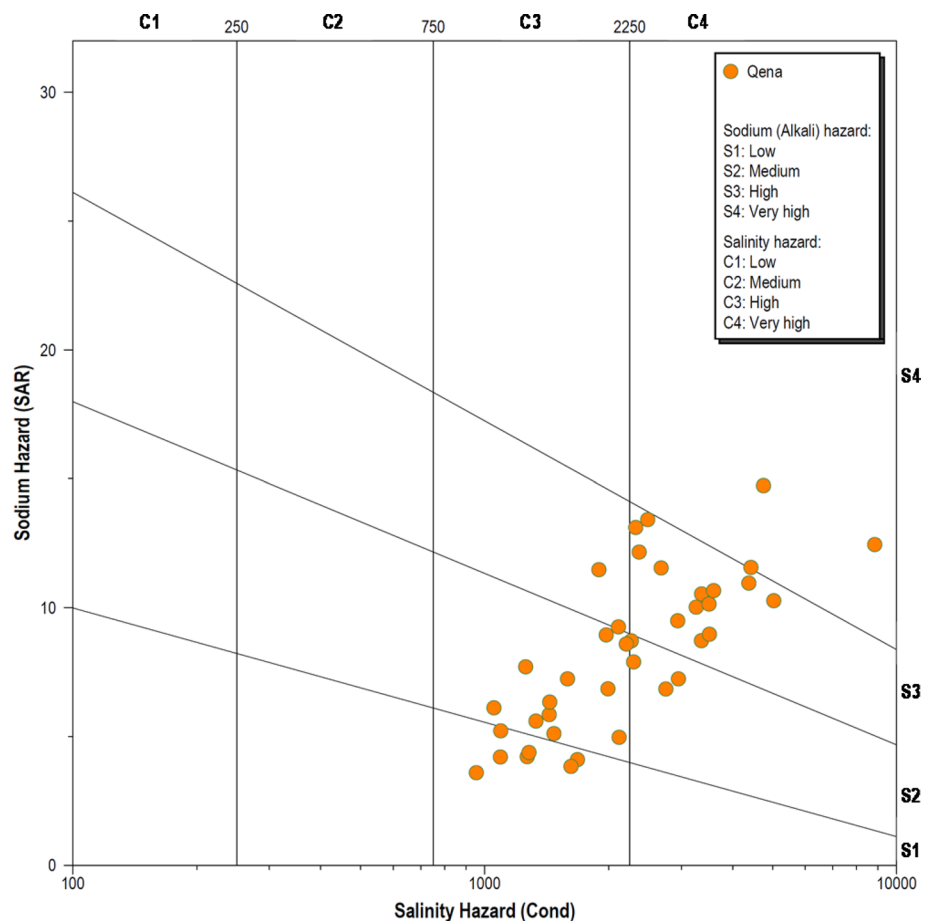
Soluble sodium percentage (SSP)

SSP is also referred to as sodium percentage (Na %). It is an estimation of the proportion of sodium ion in irrigation water like SAR, but it expresses the percentage of sodium out of the total cations. SSP is calculated by using Eq. (7). Todd (1980) classified the irrigation water according to the SSP into five classes (Table 7). According to this classification, the majority of the groundwater samples collected from the study area fall into permissible (27%) to doubtful (66%) categories (Table 7). Similar to SAR, high presence of sodium in irrigation water reacts with soil and causes a reduction in its permeability. Furthermore, frequent irrigation with high Na⁺ water makes the soil plastic and sticky in wet conditions and forms clods and crusts in dry conditions (Punmia and Lal 1981).

Residual sodium carbonate (RSC)

RSC compares the bicarbonate (HCO₃⁻) and carbonate (CO₃²⁻) concentration (also known as carbonates) to Ca²⁺ and Mg²⁺ concentrations. Irrigation water containing high carbonates greatly complicates the management of excessive sodium. When water containing carbonates dries at the soil

Fig. 11 Wilcox diagram showing the suitability of groundwater for irrigation based on EC and SAR in the study area



surface, Ca^{2+} and Mg^{2+} carbonates (lime) are formed. Since Ca^{2+} and Mg^{2+} are no longer dissolved, they do not counteract the negative effects of Na^+ , and problems related to high Na^+ may occur. White lime deposits may also become visible on turf leaves during hot dry periods as carbonates are deposited during evaporation (James 2001). RSC is calculated using Eq. (8). The RCS equation indicates the potential for Ca^{2+} and Mg^{2+} precipitation in the soil and the removal of Ca^{2+} and Mg^{2+} from the soil solution. High carbonate water can have good infiltration properties if the Ca^{2+} and Mg^{2+} levels are also high (James 2001). If water that is high in RSC is repeatedly used, the soil becomes alkaline and is likely to become sodic over time if the water also contains significant amounts of Na^+ . The degree of sodium hazard in groundwater is classified into three classes based on RSC by the US Salinity Laboratory Staff (1954) (Table 7). According to the RSC classification, almost all the groundwater samples (40 samples) in the study area are safe for irrigation ($\text{RSC} < 1.25$).

Magnesium hazard (MH)

In a magnesium-dominated water (ratio of $\text{Ca}^{2+}/\text{Mg}^{2+} < 1$) or a magnesium soil (soil–water ratio of $\text{Ca}^{2+}/\text{Mg}^{2+} < 1$), the potential effect of sodium may be slightly increased. This means a given SAR value will show slightly more damage. The lower the $\text{Ca}^{2+}/\text{Mg}^{2+}$ ratio, the more damaging is the SAR (Ayers and Westcot 1985). The magnesium hazard of irrigation water is calculated using Eq. (9) proposed by Szabolcs and Darab (1964) and could be classified into two classes (Table 7). Based on the MH classification, 41% of the collected water samples in the study area is suitable for irrigation ($\text{MH} < 50\%$), and 59% of the samples are unsuitable for irrigation (Table 7). If the MH value is more than 50%, the soil becomes alkaline and its impact on crop yield will be observed (Chaudhary and Satheeshkumar 2018).

Permeability index (PI)

The SAR, SSP, and RSC are used to evaluate the effect of salts in irrigation water on soil permeability. As is well known, irrigation water with high sodium content is unsuitable for irrigation of agricultural lands. Excess sodium in water produces an undesirable effect by changing soil permeability and water infiltration. The concentrations of Ca^{2+} , Na^+ , Mg^{2+} , and HCO_3^- influence the permeability of the soil profile; therefore, these ions are used to calculate PI values of irrigation water. The PI could be explained as a measure of the extent to which soil structure deteriorates when water is applied, a high ratio indicating a high degree of deterioration (US Salinity Laboratory Staff 1954). Low soil permeability causes poor drainage by impeding the downward movement of water and often results in a surface

crust formation that deters seed germination and emergence (Singh et al. 2020). PI is calculated using Eq. (10). PI is classified into three classes (Doneen 1964) as shown in Table 7. Based on the PI classification and the PI values, the entire groundwater samples collected from the study area fall under classes I and II (PI of 55–90%), which indicates that the water is overall good to moderately good for irrigation purposes.

Kelley's ratio (KR)

KR is an important index for evaluating irrigation water. It compares the Na^+ ion concentration with the Ca^{2+} and Mg^{2+} ion concentrations (Kelley 1951, 1963). KR is calculated using Eq. (11). Water is classified into two categories: water with KR values > 1 , which indicates an excess of Na^+ , and water with KR values < 1 , which is suitable for irrigation (Das and Nag 2015). The groundwater samples in the study area show KR values of more than 1 (95% of samples). According to the KR values, the majority of the collected groundwater samples from the study area are unsuitable for irrigation purposes (Table 7).

Nitrate contamination

Nitrate pollution of groundwater is becoming more prevalent in many parts of the world, including most European and North American countries (Strebel et al. 1989; Korom 1992; Spalding and Exner 1993; Feast et al. 1998; Tesoriero et al. 2000), and poses a serious threat to groundwater-based drinking water supplies. Since the early 1960s, excessive fertilizers and manure application in agriculture have been the primary cause of increased shallow groundwater nitrate (Appelo and Postma 2005). The maximum-allowable nitrate concentration in drinking water is 50 mg NO_3^- and the recommended level is less than 25 mg/L NO_3^- (WHO 2011). Egypt is one of the countries suffering from the elevated nitrate content in their groundwater, which derived from a variety of sources. The Nile Valley Aquifer System, which this study is a part of, has been studied by many researchers, and they have reported the high nitrate levels that exceed the permissible limits of drinking water (Mohammed 2005; Ahmed et al. 2007; Gedamy et al. 2012; Ali and El-Aassar 2018; El-Sayed 2018; Salman et al. 2019; Redwan et al. 2020; Shamruk and Akib 2021).

The nitrate (NO_3^-) concentrations in the groundwater samples in the study area range from 0.1 to 256.8 mg/L, with an average of 53 mg/L. Thirty percent (12 samples) of the groundwater samples had nitrate concentrations higher than the maximum acceptable nitrate concentration for drinking water of 50 mg/L set by WHO (2011). Sixty-three percent of the samples (25 samples) had nitrate levels higher than the recommended range of 25 mg/L. Elevated nitrate contents

are found in the groundwater wells distributed in both agricultural and rural parts of the study area. The widespread use of chemical fertilizers, organic fertilizers (manure), and waste from livestock are considered to be the main sources of nitrate that leaches into groundwater in the agricultural areas, whereas septic systems are the primary source of high nitrate in rural sections of the study area. Another important factor for rising nitrate levels in reclaimed land is the lack of silty soils cover, which allows nitrate formed by the oxidation of N-fertilizers to easily infiltrate into the deeper part of the alluvial aquifer. This finding is in line with research from Egypt (Shamrugh et al. 2001; Mohammed 2005; Salman et al. 2019) as well as other parts of the world (Spalding et al. 1982; Baker 1992; Chowdary et al. 2005; Kundu et al. 2009; Gu et al. 2013; Rahman et al. 2021b). The nitrate contour map (Fig. 12) depicts the spatial distributions of nitrate concentrations in the study area and shows no notable trend of nitrate concentration changes. However, the increase in the nitrate concentrations in the eastern part of the study area is due to intensive agricultural activities in this area, such as newly reclaimed lands, as well as the presence of individual agriculturally related human activities, such as camel and poultry farms and other feedlots.

Health risk of NO_3^- in groundwater

The presence of high nitrate levels in drinking water is believed to be a health risk to humans and animals (Sall and Vanclooster 2009; IARC 2010; WHO 2011). In humans, nitrate and nitrite are treated as probably carcinogenic (IARC 2010). Pregnant women are advised to avoid high nitrate water, because research shows possible connections to birth defects and miscarriages (IARC 2010).

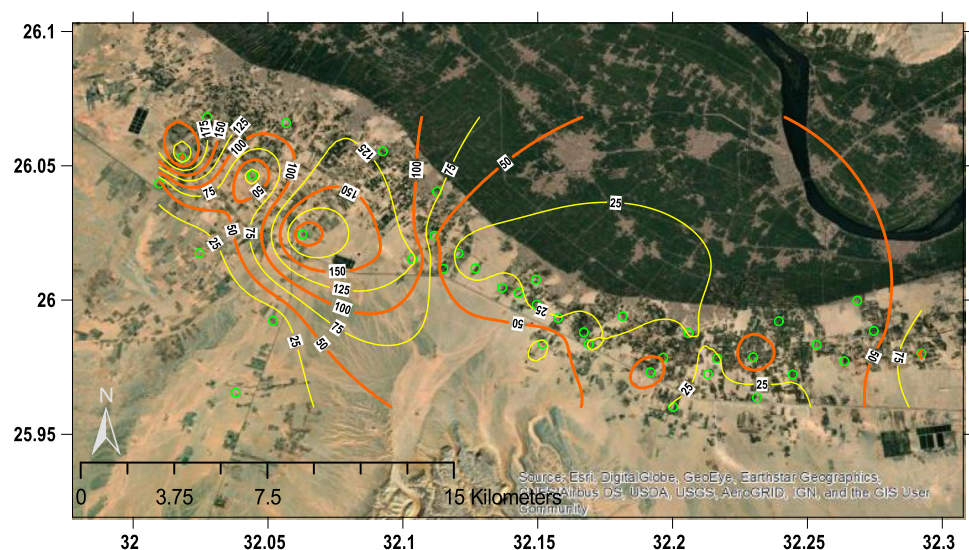
Infants who are given high nitrate water or infant formula mixed with high nitrate water can develop a condition called methemoglobinemia, also known as blue baby syndrome (Kross et al. 1992; WHO 2003; Greer and Shannon 2005). Cases have also been reported in babies who have been fed vegetables irrigated with high levels of nitrate water, particularly carrots and spinach (Simon 1966; Keating et al. 1973). Animals are affected by nitrate poisoning in the same manner as it affects humans and babies.

We conducted a preliminary human health risk assessment from the exposure to nitrate-contaminated groundwater following the USEPA methodology (Sadler et al. 2016; Zhai et al. 2017; Rahman et al. 2021b). In order to perform a health risk assessment, the nitrate intake from water is necessary to estimate. The health risk assessments for nitrate in drinking water are presented for adults and children. The average daily dose (ADD) as mg/kg/day was calculated using the following Eq. (15).

$$\text{ADD} = \frac{C_f \times C_d}{B_w} \quad (15)$$

where C_f is the nitrate concentration in drinking water (mg/L), C_d is the average daily drinking water intake (L/day), and B_w is the body weight (kg). The ADD was calculated based on the NO_3^- concentrations obtained in the samples of the current study, and the average daily water intake, which is recommended to be 2.3 and 1.5 L/day for adults and children, respectively, was based on the literature (Su et al. 2013; Rahman et al. 2021b). Also, the average body weight used in the ADD calculation is 70 and 15 kg for adults and children, respectively, according to USEPA (2002) and Rahman et al. (2021b). The hazard quotient (HQ), which is the ratio of an exposure level to a reference level, is calculated from the following Eq. (16).

Fig. 12 Nitrate concentrations (in mg/L) contour map of the study area



$$HQ = \frac{ADD}{RfD} \tag{16}$$

where ADD values are calculated from Eq. (15), and RfD is the acceptable daily intake (mg/kg/day) (the reference oral dose), where its value is 1.6 mg/kg/day as specified by USEPA (1991). It is presumed that there are no health concerns if the HQ value is less than one (Su et al. 2013).

The estimated ADD and HQ for adults and children in the groundwater samples from the study area are shown in Table 8. The higher the HQ value, the higher the health risks. The HQ value of < 1 often indicates low health risk in the exposed population (Sadler et al. 2016; Rahman et al. 2021b). The HQ value in the samples indicated a high level of risk (HQ > 1) for adults. HQ is more than one in more than 32% of the samples. We also found a significantly higher risk (HQ > 1) for children, with HQ greater than one in nearly 80% of the samples in the study area, implying that the children are more vulnerable to health risks of NO₃⁻ contamination in groundwater. The previous findings showed that the increase in the NO₃⁻ concentration in drinking water could be a potential health risk for humans, particularly infants and children. This study recommends proper management and monitoring of the agricultural activities in the study area to minimize the potential risk of nitrate in groundwater.

Water treatment

The groundwater in the study area is characterized by its high content of TDS (73% of the collected samples are brackish water) and other ion constituents, as described above. Furthermore, about 63% of the samples have nitrate concentrations higher than the recommended level. These constituents degrade water quality and are expected to cause problems with water use in a variety of ways. As a result, groundwater in many parts of the study area is unsuitable for a variety of uses (ex., drinking, irrigation, and livestock). In order to improve the groundwater quality and make it safe for usage, TDS has to be reduced, and dissolved nitrate should be reduced or removed. There is no simple method for lowering TDS and removing nitrate from water. However, to improve the water quality in the study area so that it is suitable for small-scale human activities or projects (e.g., poultry farms, camel farms, livestock, local brick industry, and medicinal and aromatic plants), mixing of water should be considered.

Mixing of water might offer an overall improvement in quality and reduce the potential toxicity problem if an alternative source of good quality water is available. The desired water salinity can be obtained by mixing two water of known salinity. You can select the parameters that will be included in the mixing calculations. The mixed water then generates solution concentrations as a result of the stepwise mixing of specified proportions of two selected water. For mixing, each of the input solutions is multiplied by its mixing fraction, and a new output solution is stoichiometrically calculated (Parkhurst and Appelo 1999).

In this section, a mixing model is conducted between water from different sources as a proposed solution for lowering the chemical content, particularly nitrate levels in the heavily contaminated wells to acceptable limits for usage in various purposes. The water of poor quality wells in the study area will be mixed with good quality water from nearby irrigation canals or the River Nile main channel. This also could be mixed with water from available nearby better quality wells. In this model, the water of wells Ar-6, Ar-22, and Ar-29 collected from the study area is mixed with irrigation canal water (S2) (Table 9). Mixing could take place in house cisterns, the roof tanks, and in pools. Table 10 summarizes the mixing percentages between the mixed sources and changes in the concentrations of the major ions and nitrate.

Table 8 Estimated daily intake (ADD) and hazard quotient (HQ) for adults and children in the study area

Age group	ADD (mg/kg/day)			HQ		
	Minimum	Maximum	Average	Minimum	Maximum	Average
Adults	0.002	8.438	1.753	0.001	5.274	1.095
Children	0.007	25.680	5.334	0.004	16.050	3.334

Table 9 The chemical composition of wells (Ar-6, Ar-22, and Ar-29) and irrigation canal water (S2)

Parameter	Unit	Wells			Irrigation canal
		Ar-6	Ar-22	Ar-29	S2
pH		7.47	7.73	7.65	8.7
Temp	°C	26.9	32	30.9	21.1
TDS	mg/L	3090.6	2847.8	1789.7	152.4
Ca ²⁺	mg/L	192.0	204.0	146.8	20.3
Mg ²⁺	mg/L	63.6	67.7	86.6	8.6
Na ⁺	mg/L	921.6	706.5	422.8	20.7
K ⁺	mg/L	9.0	10.0	9.0	7.0
HCO ₃ ⁻	mg/L	158.4	235.6	250.0	95.2
SO ₄ ²⁻	mg/L	325.2	631.8	249.4	14.4
Cl ⁻	mg/L	1500.0	1110.0	750.0	15.7
NO ₃ ⁻	mg/L	225.0	76.13	117.3	1.5

Table 10 Changes in the chemistry of wells Ar-6, Ar-22, and Ar-29 as a result of mixing with irrigation canal water (S2)

Irrigation canal (S2)		0.00	0.1	0.2	0.3	0.4	0.5	0.6	0.7	0.8	0.9	1.0
Well Ar-6		1.0	0.9	0.8	0.7	0.6	0.5	0.4	0.3	0.2	0.1	0.00
Parameter	pH	7.47	7.6	7.7	7.8	8.0	8.1	8.2	8.3	8.5	8.6	8.7
	Temp	26.9	26.3	25.7	25.2	24.6	24.0	23.4	22.8	22.3	21.7	21.1
	TDS	3090.6	2796.8	2503.0	2209.1	1915.3	1621.5	1327.7	1033.8	740.0	446.2	152.4
	Ca ²⁺	192.0	174.8	157.7	140.5	123.3	106.2	89.0	71.8	54.6	37.5	20.3
	Mg ²⁺	63.6	58.1	52.6	47.1	41.6	36.1	30.6	25.1	19.6	14.1	8.6
	Na ⁺	921.6	831.5	741.4	651.3	561.2	471.1	381.1	291.0	200.9	110.8	20.7
	K ⁺	9.0	8.8	8.6	8.4	8.2	8.0	7.8	7.6	7.4	7.2	7.0
	HCO ₃ ⁻	158.4	152.1	145.8	139.4	133.1	126.8	120.5	114.1	107.8	101.5	95.2
	SO ₄ ²⁻	325.2	294.1	263.0	232.0	200.9	169.8	138.7	107.6	76.6	45.5	14.4
	Cl ⁻	1500.0	1351.6	1203.1	1054.7	906.3	757.9	609.4	461.0	312.6	164.2	15.7
NO ₃ ⁻	225.0	202.7	180.3	158.0	135.6	113.3	90.9	68.5	46.2	23.8	1.5	
Irrigation canal (S2)		0.00	0.1	0.2	0.3	0.4	0.5	0.6	0.7	0.8	0.9	1.0
Well Ar-22		1.0	0.9	0.8	0.7	0.6	0.5	0.4	0.3	0.2	0.1	0.00
Parameter	pH	7.73	7.8	7.9	8.0	8.1	8.2	8.3	8.4	8.5	8.6	8.7
	Temp	32	30.9	29.8	28.7	27.6	26.6	25.5	24.4	23.3	22.2	21.1
	TDS	2847.8	2578.3	2308.7	2039.2	1769.6	1500.1	1230.5	961.0	691.4	421.9	152.4
	Ca ²⁺	204.0	185.6	167.3	148.9	130.5	112.2	93.8	75.4	57.0	38.7	20.3
	Mg ²⁺	67.7	61.8	55.9	50.0	44.1	38.2	32.3	26.4	20.4	14.5	8.6
	Na ⁺	706.5	637.9	569.3	500.8	432.2	363.6	295.0	226.4	157.9	89.3	20.7
	K ⁺	10.0	9.7	9.4	9.1	8.8	8.5	8.2	7.9	7.6	7.3	7.0
	HCO ₃ ⁻	235.6	221.6	207.5	193.5	179.4	165.4	151.3	137.3	123.2	109.2	95.2
	SO ₄ ²⁻	631.8	570.1	508.3	446.6	384.8	323.1	261.4	199.6	137.9	76.1	14.4
	Cl ⁻	1110.0	1000.6	891.1	781.7	672.3	562.9	453.4	344.0	234.6	125.2	15.7
NO ₃ ⁻	76.13	68.7	61.2	53.7	46.3	38.8	31.4	23.9	16.4	9.0	1.5	
Irrigation canal (S2)		0.00	0.1	0.2	0.3	0.4	0.5	0.6	0.7	0.8	0.9	1.0
Well Ar-29		1.0	0.9	0.8	0.7	0.6	0.5	0.4	0.3	0.2	0.1	0.00
Parameter	pH	7.65	7.8	7.9	8.0	8.1	8.2	8.3	8.4	8.5	8.6	8.7
	Temp	30.9	29.9	28.9	28.0	27.0	26.0	25.0	24.0	23.1	22.1	21.1
	TDS	1789.7	1626.0	1462.2	1298.5	1134.8	971.0	807.3	643.6	479.8	316.1	152.4
	Ca ²⁺	146.8	134.2	121.5	108.9	96.2	83.6	70.9	58.3	45.6	32.9	20.3
	Mg ²⁺	86.6	78.8	71.0	63.2	55.4	47.6	39.8	32.0	24.2	16.4	8.6
	Na ⁺	422.8	382.6	342.4	302.2	262.0	221.7	181.5	141.3	101.1	60.9	20.7
	K ⁺	9.0	8.8	8.6	8.4	8.2	8.0	7.8	7.6	7.4	7.2	7.0
	HCO ₃ ⁻	250.0	234.5	219.0	203.5	188.1	172.6	157.1	141.6	126.1	110.6	95.2
	SO ₄ ²⁻	249.4	225.9	202.4	178.9	155.4	131.9	108.4	84.9	61.4	37.9	14.4
	Cl ⁻	750.0	676.6	603.1	529.7	456.3	382.9	309.4	236.0	162.6	89.2	15.7
NO ₃ ⁻	117.3	105.7	94.1	82.5	71.0	59.4	47.8	36.2	24.6	13.1	1.5	

Conclusion

Forty-one groundwater samples were collected in this study from the investigated area in the Nile Valley, north of Qena city. The Nile Valley aquifer represents the major source of water for irrigation and domestic usage in the study area. The hydrochemical investigations show that groundwater in the study area is influenced by

both natural processes and anthropogenic activities. The chemical weathering of the aquifer rock-forming minerals and the adjacent carbonate plateau is one of the most important processes influencing the groundwater chemistry in the study area. The groundwater samples in the study area are characterized by fresh to brackish water with Ca²⁺·Mg²⁺·Cl⁻ and Na⁺·Cl⁻ hydrochemical facies. The majority of the samples are chemically unsuitable for

human consumption according to the permissible limits set by WHO, and they are classified as hard to very hard types.

Based on the water quality parameters examined for the groundwater samples collected in this study, the suitability of groundwater samples for irrigation purposes is good to moderately good. Also, it may reflect some prominent alkaline hazard to the soil as reflected by the SSP and KR. The salinity hazard is recognized as high, and it is expected to increase with the expansion of irrigation.

The results revealed that the NO_3^- concentrations of the samples exceed the safe limit for drinking water set by WHO of 50 mg/L. NO_3^- in the groundwater of the study area is derived from multiple sources, including synthetic N-fertilizers, manure, and rural septic tanks. The human health risk assessments indicate a high level of risk for adults, but a significantly higher health risk for children drinking NO_3^- -contaminated groundwater in the study area. The main concern is that the average NO_3^- concentration will likely continue to increase in the groundwater of the study area. In order to reduce nitrate pollution, excessive chemical fertilizer and manure applications should be prevented, as well as raise awareness of the negative impacts of excessive fertilizers use on the environment. Mixing of water from various sources was carried out in order to reduce the chemical content, particularly nitrate levels, in the highly contaminated wells to tolerable levels. Mixing, when it is possible, can be used as the most cost-effective water treatment method for decreasing chemical concentration below the limitations for various uses.

The salinity hazard is regarded as high, and special management for salinity control is required for groundwater in the study area. It is suggested that modern and high water use efficiency irrigation methods such as drip and sprinkler must be applied in the study area: first to prevent aquifer depletion and, second, to reduce the contamination that might be caused by return flow water. Otherwise, groundwater monitoring, both in terms of quality and quantity, is essential for long-term groundwater management.

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Declarations

Conflict of interest The authors have declared that they have no conflicts of interest.

Data availability All data used in this study are available from the corresponding author upon request.

Ethical approval No animals were involved in this study. No human participants were involved in this study.

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