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Sequestering a non-steroidal anti-inflammatory drug using modified orange peels

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Abstract

This study investigates the sorption of a non-steroidal anti-inflammatory drug, ibuprofen, (IBP) using acid activated carbon prepared from orange peel (OPA). OPA was characterized via the use of Fourier transform infrared and scanning electron microscopy techniques. Four isotherm models were utilized to assess the adsorption data: Langmuir, Freundlich, Temkin, and Dubinin–Raduskevich, respectively. It was established that the IBP adsorption onto OPA fitted Langmuir isotherm model most. The optimum monolayer adsorption capacity of OPA was 49.30 mg/g at 50 °C. The adsorption data was subjected to kinetic test using pseudo-first-order and pseudo-second-order, Elovich, and intraparticle diffusion models. The sorption process was best described by pseudo-second-order kinetic model. The mean heat of adsorption, E_a at all temperatures studied was < 8.00 kJmol^{-1}, signifying that the sorption mechanism is physisorption. Thermodynamic study reveals that the adsorption is spontaneous and exothermic. OPA was established to be a viable and effective adsorbent for the sorption of IBP from aqueous medium.

Keywords Adsorption · Orange peel · Ibuprofen · Kinetics · Thermodynamics

Introduction

Endocrine-disrupting chemicals (EDCs) are bioactive compounds that affect living tissues. According to the report of the Environmental Protection Agency of 2015, an endocrine disruptive chemical is a substance which affects the production, secretion, binding, movement, function and/or removal of some natural hormones which are accountable for the upkeep of homeostasis, behavior, reproduction, and growth within the body (Diamanti-Kandarakis et al. 2009; Endocrine Society 2016). The endocrine system comprises of an assembly of glands which secrete hormones into the circulatory system and transports them to specific organs in the body. The major function of hormones in animals is the coordination of chemicals (Hafez et al. 2016). This helps in the regulation of various body functions such as growth, sexual development and reproduction. EDCs can imitate some hormones that occur naturally in the body, according to the National Institute of Environmental Health Sciences, EDC binds to a receptor within the cell, and then interfere with the hormonal secretion or control of the receptor. Abnormalities and reduction in organ activities are some of the resulting effects of such disruptions. The everyday use of chemicals in foods and other daily needs such as soaps, perfumes, and other household products, which has become a norm in the modern society, has brought about the release of persistent substances, many of which introduce EDCs into the surroundings. Available data has confirmed the high risk that these EDCs pose on human metabolism which can eventually lead to metabolic diseases (Swedenborg et al. 2009; Propper 2005). Parabens are examples of antimicrobial

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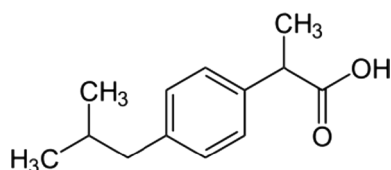
Fig. 1 Structure of ibuprofen

Table 1 Physiochemical properties of ibuprofen

Appearance	Colourless crystalline solid
Boiling point	157 °C
Brand name	Brufen; Motrin; Nurofen; Advil; Nuprin
Density	1.029 g/cm^3
IUPAC name	(RS)-2-(4-(2-Methylpropyl)phenyl)propanoic acid
Melting point	77–78 °C
Molar mass	206.28 g/mol
Molecular formula	$C_{13}H_{18}O_2$
Solubility	Partially soluble in water but soluble in organic solvent

agents that have weak estrogenic activity and are in most cosmetics and food materials (Daughton and Ternes 1999). In this study, ibuprofen is of primary concern as a form of EDC in the environment. The consumption rate of ibuprofen in the world is very high. For example, in UK, it is one of the most highly rated (among top five) consumed drugs, with annual estimation production of several kilotons (Sebastine and Wakeman 2003; National Institute of Environmental Health Sciences 2019).

Ibuprofen is among the important drugs enumerated by the World Health Organization, and so ibuprofen is among the common pharmaceuticals regularly found within reach (National Institute of Environmental Health Sciences 2019). Figure 1 represents ibuprofen structure with a molecular formula of $C_{13}H_{18}O_2$, having an IUPAC nomenclature of 2-{4-(2-methylpropyl) phenyl} propanoic acid. It was initially developed in the year 1960 while investigating for a safer substitute for aspirin. A non-steroidal anti-inflammatory drug (NSAID) is use to suppress pain, fever, arthritis symptoms, and dysmenorrhea. It is identified to possess an antiplatelet effect. Comparison with aspirin or some well-known antiplatelet drugs indicate that it is comparatively mild and short-lived. It acts as vasodilator, having been revealed to enlarge coronary arteries and some other blood vessels. A small dose of ibuprofen is healthy but large dose is usually accompanied by health hazard. The physiochemical properties of ibuprofen are shown in Table.1.

In the world, several kilotons of ibuprofen are produced yearly (Buser et al. 1999) and effluents from pharmaceutical industries producing ibuprofen are released into the environment without adequate treatment. It was stated that the quantity of IBP in the surroundings falls within the range of 10 µg/L and 169 µg/L (Dos Santos et al. 2007). The endocrine disruptive action of IBP has been identified through various studies (Buser et al. 1999). "4-IBAP (4- isobutylacetophenone)" is one of the harmful metabolites of ibuprofen which is known for its hazardous effect on the CNS (central nervous system). Hence, it is of necessity to develop an effective means for sequestering IBP from the environment.

There are different techniques for the uptake of EDCs from wastewater; these include: membrane filtration (Zhang et al. 2014), oxidation processes (Garoma and Matsumoto 2009), ultrasonic treatment (Méndez-Arriaga et al. 2008), electrochemical degrading (Ciriaco et al. 2009), miniature created wetlands (Dordio et al. 2009, 2010), and adsorption (Choi et al. 2005). Among the various clean-up methods available, adsorption method with activated carbon is the most sustainable because it is eco-friendly and is mostly used for EDC removal. Different agricultural wastes materials have been used as adsorbents, among them are: hen feathers (Gupta et al. 2006), wheat husks (Gupta et al. 2007), bottom ash and deoiled soya (Gupta et al. 2009), durian seed (Ahmad et al. 2014a and b), lime peels (Ahmad et al. 2015), banana stalk (Bello et al. 2012).

However, commercially activated carbon is costly and scarce; hence there is a need to investigate the effectiveness of different agricultural wastes or natural substances in removing IBP from industrial effluents.

The most extensively used adsorbents for emerging contaminants adsorption from effluent is activated carbon; this is as a result of their enlarged surface area, high porosity, high level of surface reactivity, high capacity of adsorption and its efficiency in removing a wide range of different forms of adsorbates (Nasuha et al. 2010). Orange peels are usually found in large quantity in soft drinks industries and usually treated as wastes thereby serving as pollutants to the environment. Its component includes: cellulose, pectin, chlorophyll pigments, lignin, hemicellulose and other hydrocarbons with relative low molecular weight. Based on our findings, there is no documented report addressing the use of orange peel activated carbon (OPA) for the adsorption of IBP from wastewater. This research work is focused on the preparation and characterization of orange peel as an activated carbon for the purpose of adsorbing IBP from aqueous media. Effect of operational parameters includes: Contact time, temperature, and initial concentration governing the adsorption were studied. Sorption Isotherm, kinetic and thermodynamic studies were carried out.

Materials and methods

Sample collection and pre-treatment

Orange peel samples were collected from a disposal unit at LAUTECH Ogbomoso where they serve as agricultural wastes. It undergoes washing using distilled water followed by drying to a specific weight. It was later crushed into smaller particles and sieved to 106 μm size. Thereafter, it was stored in a desiccator for further use.

Adsorbent characterization

Fourier transform infrared (FTIR)

FTIR spectroscopic study of orange peel was carried-out using FTIR spectroscopy device (FTIR-2000, Perkin Elmer). This equipment gave spectra revealing the typical functional groups present in both the raw and acid activated orange peel samples.

Scanning electron micrograph

This microscope (SEM) is among the most versatile device used in determining and investigating morphological structures. This equipment was utilized to investigate the porosity level and surface properties of both raw and acid activated orange peel samples. The sample with numerous pores has high affinity for adsorption of adsorbates.

Activated carbon preparation

Thirty grams of orange peel sample was measured into a beaker followed by the addition of 600 cm^3 of H$_3$PO$_4$; it was continuously stirred on a hot plate using a stirrer until the formation of a paste. This was moved into a crucible and then subjected to heating at 500 °C for 1 h 30 min in a furnace. It undergoes cooling followed by washing using de-ionized water until a pH of 6.9 was achieved. It was dried at a temperature of 105 °C for 4 h in an oven. Thereafter, it was crushed to smaller size. The adsorbent was stored in desiccator for further use.

Adsorbate solution preparation

Preparation of the stock solution was done by dissolving ibuprofen (500 mg) in 1 L of ethanol. Working solutions were obtained by serial dilution from the stock solution. Ibuprofen is moderately water soluble (Baccar et al. 2012); however, it is completely soluble in organic solvents

(Manrique and Martinez 2007). Due to this, ethanol (organic solvent) was used as a solvent for the adsorbate so as to ensure complete dissolution.

Adsorption equilibrium experiments

Ten milligrams of the adsorbent was weighed into pre-arranged conical flasks (250 ml) which contains 100 ml solutions of IBP with various initial IBP concentrations (5–25 mg/L). This mixture was then agitated using iso-thermal water bath at 120 revolutions per minute at different temperatures (30, 40, 50 °C) until equilibrium was reached. The solution of the sample was taken at pre-determined time intervals using micro filter syringe for residual concentration determination. Concentrations of IBP solutions were obtained by taking the absorbance readings using UV–Vis spectrophotometer at 240 nm wavelength. Solution temperature, initial IBP concentration and contact time of the process were also examined. The amount of IBP adsorbed and the percentage removed were determined using the equation below:

$$q_e = \frac{(C_e - C_o)}{W} \tag{1}$$

where C_o signifies IBP initial concentrations at liquid-phase. C_e signifies equilibrium concentrations of IBP at liquid-phase measured in (mgL^{-1}). V signifies volume (dm^3). W signifies adsorbent mass used (g). Percentage uptake of IBP was calculated using Eq. 2.

$$\text{Removal}\,(\%)\frac{(C_e - C_o)}{C_o} \times 100 \tag{2}$$

where C_0 signifies initial concentration of IBP at liquid-phase (mgL^{-1}), C_e signifies equilibrium. concentration of Ibuprofen (mgL^{-1}).

Solution temperature effect

The influence of solution temperature on the IBP sorption process was observed at different temperatures: 30 °C, 40 °C and 50 °C, respectively. This was done by adjusting the temperature regulator of the water bath shaker.

Adsorption isotherms

This allows us to determine the process of adsorption at equilibrium point of adsorption. Four models were employed to test the adsorption data.

Langmuir isotherm

This is centered on a single layer and uniform adsorption to the surface of a solid. Langmuir suggested that the distance is inversely proportional to the intermolecular forces, in accordance with Langmuir (Eq. 3).

$$\frac{C_e}{q_e} = \frac{C_e}{q_m} + \frac{1}{K_L q_m} \tag{3}$$

C_e = adsorbate concentration at equilibrium (mgL^{-1}); q_e = adsorbate concentration uptake per unit weight of adsorbent (mg/g); q_m = Adsorbent optimum adsorptive capacity (mg/g); K_L = Langmuir constant (L/mg). Langmuir constant is obtained from the graph of $\frac{c_e}{q_e}$ against C_e. The features of Langmuir expression may be expressed in form of R_L (separator factor) that has no dimension which is expressed as:

$$R_L = \frac{1}{(1 + K_L C_o)} \tag{4}$$

C_o signifies the maximum initial amount of solute. The dimensionless constant (R_L) signifies that the sorption is satisfactory ($0 < R_L < 1$), unsatisfactory ($R_L > 1$), linear ($R_L = 1$), or irreversible ($R_L = 0$).

Freundlich isotherm

This describes sorption process which occurs on nonuniform surface. It suggests that the spots whose binding strength is higher are first occupied and reduction in the strength depends on the filling rate of the sites. The Freundlich isotherm (Freundlich 1906) is:

$$\ln q_e = \ln k_f + \frac{1}{n} \ln C_e \tag{5}$$

where q_e signifies adsorbate concentration uptake per unit weight of adsorbent (mg/g); k_f signifies Freundlich constant (mgg^{-1}), which specify the comparative adsorption capacity of the adsorbent associated with binding energy; C_e signifies adsorbate equilibrium concentration (mg/L); n signifies Freundlich coefficient signifying the deviance from adsorption linearity and is also referred to as heterogeneity factor.

Temkin isotherm

This accounts for the effect of the interaction which exists between the adsorbate and the adsorbent. This isotherm predicts that the heat of adsorption for all molecules in the film is inversely proportional to the coverage surface. This model is expressed below (Temkin and Pyzhev 1940):

$$q_e = B \ln K_t + B \ln C_e \tag{6}$$

where

$$B = \frac{RT}{b} \tag{7}$$

B signifies heat capacity constant (Lmg^{-1}); b signifies Temkin constant; q_e signifies adsorbate concentration uptake at equilibrium (mg/g); C_e signifies amount of adsorbate at equilibrium (mgL^{-1}); T signifies Temperature; R signifies Gas constant (8.314 J/mol K); K_t signifies Equilibrium binding constant (L/mg). The plots of q_e against C_e give a straight line graph.

Dubinin–Radushkevich isotherm

This isotherm accounts for the free energy of adsorption and porosity. It is used in describing the mechanism of adsorption with Gaussian dispersion energy to a uniform surface. It is stated as (Dubinin and Radushkevich 1947):

$$\ln q_e = \ln q_m - \beta \varepsilon^2 \tag{8}$$

where ε can be estimated from

$$\varepsilon = RT \left[1 + \frac{1}{c_e} \right] \tag{9}$$

$$E_a = \frac{1}{\sqrt{2\beta}} \tag{10}$$

where R signifies gas constant (8.314 J/mol/K), T signifies temperature (K), C_e signifies amount of adsorbate at equilibrium (mgL^{-1}). E signifies free energy of adsorption.

Adsorption kinetic studies

It portrays the interaction that exists between the amount of adsorbate in the solution and adsorption rate, and the effect of adsorption capacity in terms of adsorption rate. In this study, four models were engaged to test the adsorption data. These includes: pseudo first-order (Lagergren and Svenska 1898), pseudo-second-order, (Ho and McKay 1999a, b) Elovich (Unganish et al. 1976), and intraparticle diffusion (Weber and Morris 1962), respectively.

Kinetic study validation

Standard deviation Δq_e (%) were employed to authenticate the kinetic models used in testing the adsorption data. The isotherm expression suitability toward the adsorption kinetic data was ascertained via comparing the R^2 values.

$$\Delta q_e(\%) = 100 \frac{\sqrt{(q_{e.\exp} - q_{e.\text{cal}})}}{N - 1} \tag{11}$$

N denotes the number of data points, $q_{e.\exp}$ denotes the experimental value (mg/g) and $q_{e.\text{cal}}$ denotes the calculated value (mg.g^{-1}).

Test of kinetic models

These models were subjected to validation using the sum of squares error (SSE %) aside from the R^2 value. The kinetic adsorption process of IBP onto orange peel activated carbon was investigated at different initial concentrations. The sum of squares error (SSE, %) is expressed as:

$$\text{SSE}(\%) = \sqrt{\Sigma \frac{(q_{e.\exp} - q_{e.\text{calc}})^2}{N}} \tag{12}$$

N denotes the number of data points, $q_{e.\exp}$ denotes the experimental value (mg.g^{-1}) and $q_{e.\text{cal}}$ denotes the estimated value (mg.g^{-1}). A low value of SSE signifies good fit.

Adsorption thermodynamic studies

This study was examined as a function of temperature. Its parameters are $\Delta G°$, $\Delta H°$ and $\Delta S°$, which are essential in defining the spontaneity and the nature of the relationship that exist between the adsorbent and adsorbate. It was determined employing the following equations:

$$\text{In}K_L = \frac{\Delta S°}{R} - \frac{\Delta H°}{\text{RT}} \tag{13}$$

where K_L represents Langmuir constant (g/L) $\Delta S°$ represents entropy change (kJ/mol K). R represents gas constant (J/mol K). $\Delta H°$ represents enthalpy change (kJ/mol). T represents the temperature (K). Therefore, $\Delta H°$ and $\Delta S°$ were gotten from a plot of ln K_L against 1/T. If $\Delta H°$ value is positive,

it depicts an endothermic adsorption exercise; meanwhile, if the value is negative, it shows that the process is exothermic. A nonnegative value of $\Delta S°$ implies a rise in the level of disorderliness at the solution/solid interface which takes place in the course of the adsorption (Bello et al. 2011). In addition, when the value of $\Delta G°$ is negative, it reveals that the sorption is spontaneous at the studied temperature. $\Delta G°$ can be estimated via the expression below:

$$\Delta G° = -\text{RTIn}K_o \tag{14}$$

Arrhenius expression was used to ascertain whether the adsorption is chemical or physical in nature. Physisorption process has E_a which falls within 5 to 40 kJ/mol. Nollet et al. (2003), documented that chemisorption processes possess elevated energy of activation that falls within 40–800 kJ/mol.

Results and discussion

Characterization

Scanning electron microscopy (SEM)

This equipment was employed to detect the pore size distribution of the adsorbent. Figure 2a and b respectively depicts the SEM of OPR and OPAC as shown below. Comparing these figures, OPR has a rough surface with undeveloped pore size, but for OPAC, well-enlarged pores were formed on the surface. This implies that the activation process was effective in opening up the pores of the adsorbent. OPAC has several pore sizes for the uptake of ibuprofen drug molecules. Similar observations were observed in other reported

Fig. 2 a The micrograph of OPR. **b** The micrograph of OPA
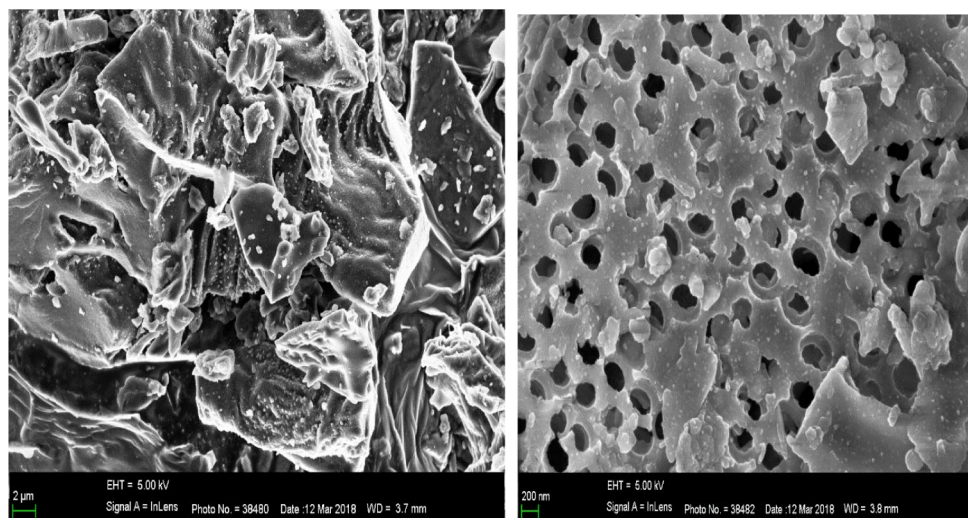
(a) The micrograph of OPR   **(b)** The micrograph of OPA

studies (Bui and Choi 2009; Shashi et al. 2010; Ahmad et al. 2014a and b).

Fourier transform infrared spectroscopy (FTIR)

This technique revealed the functional groups available in the adsorbent. The samples were studied within 400–4000 cm^{-1} on an FTIR spectrophotometer using KBr discs. The FTIR spectra of orange peel raw (OPR) and activated orange peel (OPAC) are shown below (Fig. 3a and b respectively). From these figures, some broad peaks are observed while some bands are shifted to lower and higher wavenumbers. New peaks are also formed as a result of activation process. Table 2 contains the FTIR band assignment of OPAC. A similar band assignment was observed from the studies carried out by Ahmad et al. (2015) and Bello et al. (2012).

Batch adsorption studies

Adsorption isotherms

Isotherm models investigated include: Langmuir, Freundlich, Temkin, and Dubinin–Radushkevich. The isotherm having the highest correlation coefficient (R^2) value fit the adsorption data most. Langmuir theory (Langmuir 1918) suggests that the sorbent surface is homogeneous, and there is no interaction in-between the molecules adsorbed during the formation of a monolayer surface coverage. Table 3 shows the different isotherm parameters calculated at various temperatures. From Table 4, the highest value of q_m is 49.3 mg/g at 50 °C Comparing this value (maximum monolayer adsorption capacities of OPAC) with other adsorbent shows that OPAC is an effective adsorbent (Table 4). The R_L value calculated at various temperatures range from 0.0039–0.140. The values are less than 1, suggesting that the adsorption data was favorable at all IBP concentration range studied (Zhang and Pan 2014).

This model can be used when the adsorption surface is not uniform, and the extent of adsorption increases with concentration (Mittal et al. 2008). The values of n for Freundlich isotherm were all greater than 1, signifying that the adsorption process was favorable for IBP adsorption on OPAC. The $1/n$ values were below 1 at all temperatures, indicating that the adsorbate was favorably adsorbed on the adsorbent (Tunç et al. 2009). The Freundlich constant (k_F) at 323 K is 59.9, which is approximately three times greater when compared with that of 303 K ($k_F = 20.3$). This suggests that a rise in temperature increases the adsorbent sorption capacity. D-R and Langmuir isotherms fit the adsorption data most, judging from R^2 values, these signify that the surface binding energy of the adsorbent was homogeneous, the relationship between the adsorbent and adsorbate was negligible, and no

competition exists between the molecules of IBP adsorbed on the surface of the adsorbent (Gong et al. 2008).

Temkin isotherm constant (b_T) otherwise known as the equilibrium binding factor was calculated from the Temkin isotherm expression. As shown in Table 3, OPAC has the highest equilibrium binding constant at low temperature. The apparent free energy (E), porosity, and the sorption process parameters were investigated using Dubinin–Radushkevich isotherm model. The value of E_a at various working temperatures lies in the range 5.00–7.07 kJ/mol. Since E$_a$ < 40 kJ/mol, the nature of adsorption mechanism is physisorption (Onyango et al. 2004). Comparing the average correlation coefficient (R^2), it was inferred that the adsorption data fits the isotherm in this order: Dubinin–Radushkevich ($R^2 = 0.965$) > Langmuir ($R^2 = 0.960$) > Freundlich ($R^2 = 0.954$) > Temkin ($R^2 = 0.953$). Generally, Langmuir adsorption isotherm is based on single layer coverage of the IBP on the adsorbent surface which depict that the adsorption of IBP by activated carbon follows monolayer formation (Singh et al. 2012).

Separation factor

Langmuir expression may be expressed in form of separation factor, R_L. The R_L value depict the favorability of adsorption; it can be interpreted in this wise; adsorption is irreversible when ($R_L = 0$), adsorption is favorable when ($0 < R_L < 1$) and adsorption is linear when ($R_L = 1$). It was observed in Table 3 that, the values of R_L ranges from 0 to 1 for the adsorption of IBP onto OPAC. This reveals that the adsorption process was favorable at the temperature range studied (Bello et al. 2011).

Adsorption kinetic studies

The kinetic data for IBP adsorption onto OPAC are presented in Table 5. To determine the most suitable kinetic model, correlation coefficient (R^2) values were computed. R^2 values of pseudo-first-order kinetic model ranged from 0.8848 to 0.9347. These values are lower when compared to that of pseudo-second-order kinetic model i.e. 0.999 to 1 (Table 5) suggesting that pseudo-second-order best fit the adsorption data. Similarly, Fig. 4 shows the plot of t/q_t versus t for pseudo-second-order model. The plot gave straight line which passes through the origin; this is an indication of a good fit. Also, considering R^2 values and the relationship between $q_{e, cal.}$ and $q_{e, exp.}$ calculated from the adsorption data, there were good agreements in the values obtained for pseudo-second-order kinetic model. This denotes that pseudo-second-order kinetic model fits the adsorption data most. The kinetic data of uptake of IBP by OPAC at different initial concentrations were subjected to model validity test using sum squares of error (SSE, %) A low value of SSE
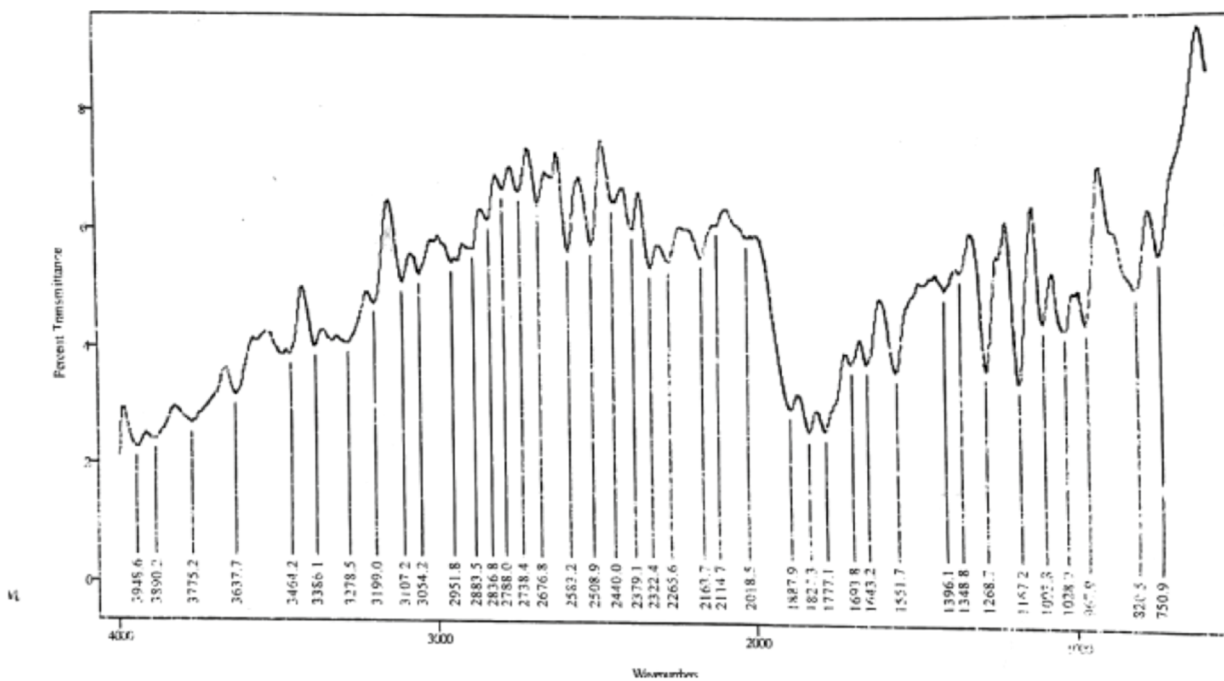
(a) FTIR Spectrum of OPR
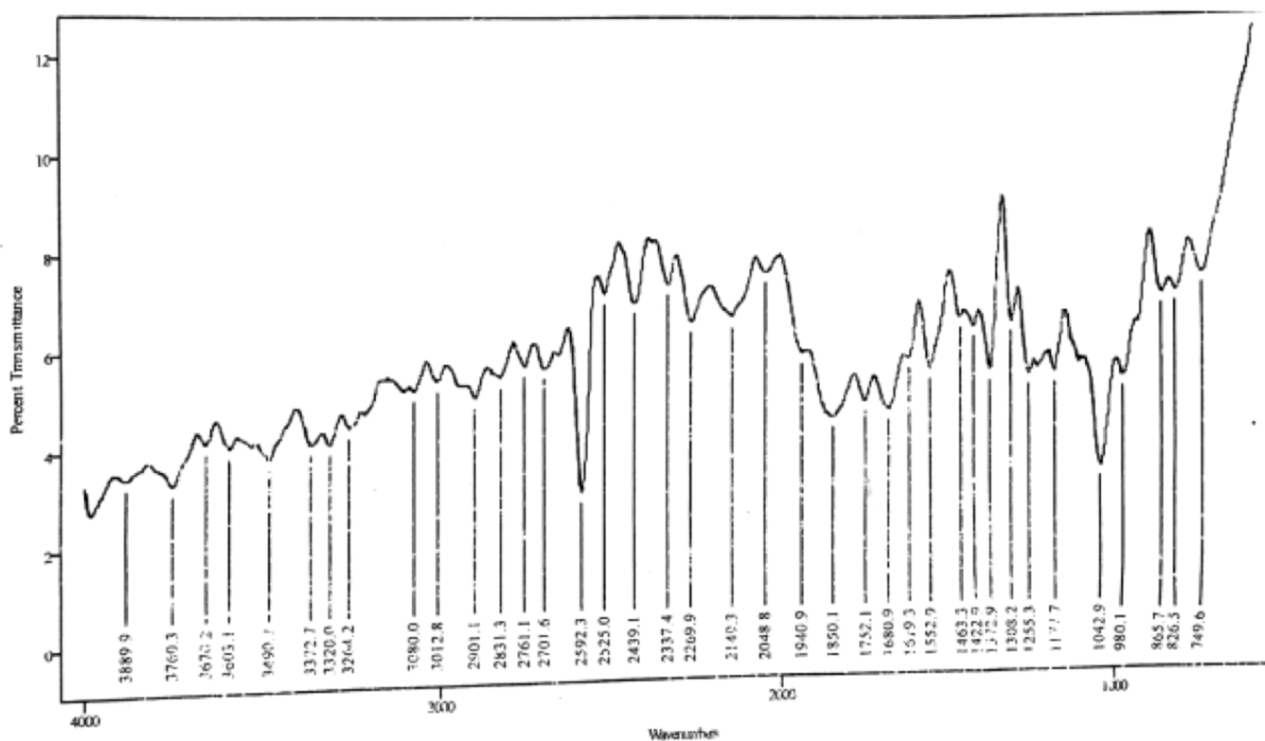
(b) FTIR Spectrum of OPA

Fig. 3 **a** FTIR Spectrum of OPR. **b** FTIR Spectrum of OPA

and greater R^2 value gave the best fit. Table 5 shows that pseudo-second-order kinetic model best describe the adsorption of IBP onto OPAC (Bello et al. 2008). One of the most suitable models for explaining the mechanism of chemisorption is the Elovich equation (Tan et al. 2009). This model assumes that the solid surfaces are actively heterogeneous.

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Table 2 FTIR spectra of raw and acid-activated orange peel (OP) wavenumber (cm-1)

OPR	OPA	Differences	Band assignment
31,072	3490.7	+ 383.5	O–H Stretch
2583.2	2592.3	+ 9.1	C–H Stretch
1777.1	1850.1	+ 73	C=O Stretch
1551.7	1552.9	+ 1.2	C=C Stretch
1268.7	1372.9	+ 104.2	C–H bending
1167.2	1042.9	− 124.3	C–N Stretch
820.5	865.7	+ 45.2	C–H bending

Table 3 Isotherm parameters for IBP adsorption onto OPA at different temperatures

Model	Temperature (K)		
	303	313	323
Langmuir isotherm			
q_m	26.50	38.60	49.30
K_L	5.00	1.58	1.45
R_L	0.004	0.013	0.014
R^2	0.970	0.993	0.917
Freundlich isotherm			
K_f	20.3	25.3	59.9
n	3.10	1.50	0.90
R^2	0.967	0.962	0.933
Temkin isotherm			
K_T	1.37	1.80	3.10
b_T	65.00	40.90	30.40
R^2	0.912	0.996	0.952
B	3.85	8.13	13.67
D-R isotherm			
q_m	21.50	26.00	41.10
R^2	0.945	0.993	0.958
β	1E-06	3E-06	2E-06
E_a (kJ/mol)	7.07	4.08	5.00

Hypothetically, an increase in the temperature of the solution, leads to a decrease in the surface coverage (β), because at higher temperature there is reduction in the available sites for adsorption. Low R^2 values were obtained for the adsorption process ranging from 0.8433 to 0.9679. According to Dogan et al., low R^2 value is an indication of disagreement between the experimental data in Elovich kinetic model. From Table 5, comparing the correlation coefficient values, the manner with which the adsorption process fitted the three kinetic models: pseudo-second-order > pseudo-first-order > Elovich. A series of elementary steps resulted in the mechanism for the adsorption of ibuprofen. The rate controlling step is possibly that of intraparticle diffusion. This is further confirmed from the plots of q_t versus $t^{1/2}$ giving a linear plot that does not pass through the origin. The value of the intercept C shows the relationship between the boundary layer thickness and the boundary layer effect (Ozcan et al. 2005). The boundary layer effect is a measure of the bias or affinity of the adsorbent for IBP.

The rate constant k_{diff}, C and R^2 are shown in Table 5. For intraparticle diffusion model, the correlation coefficients (R^2) are between 0.8385 and 0.9682, which implies that adsorption of IBP onto OPA was intraparticle diffusion model controlled. Figure 5 represents the plot of q_t against $t^{1/2}$ at various initial concentrations. The plot does not pass through the origin; this is suggesting some level of boundary layer diffusion (Wang and Wang 2007).

Thermodynamic studies

This study was undertaken to determine the changes in energy that happen in the process. Thermodynamic parameters investigated include: change in entropy (ΔS^{o}), change in enthalpy (ΔH^{o}), and standard free energy change (ΔG^{o}). The ΔH^{o} and ΔS^{o} values were calculated from the slopes and intercepts (respectively) of the graph of ln k_L against $1/T$ (inverse of temperature, T). The negative value of $\Delta H°$ at a particular temperature indicate the exothermic nature of the adsorption process, likewise the positive value of $\Delta S°$ confirms an increase in the degree of randomness at the

Table 4 Adsorption capacities of IBP on various adsorbents

Adsorbent	Adsorptive capacity (mgg^{-1})	References
Honeycomb-shaped activated carbon	16.73	Dubey et al. (2010)
AC from olive waste cake	9.09	Baccar et al. (2012)
Mesoporous silica SBA-15	0.41	Bui and Choi (2009)
Olive waste cake	12.60	Baccar et al. (2012)
Parthenium hysterophorus	3.289	Sandip et al. (2016)
Artemisia vulgaris-derived mesoporous honeycomb-shaped activated carbon	16.945	Shashi et al. (2010)
Raspberry derived carbon-tubules	20.0	Dubey et.al. (2014)
Activated Orange Peel (OPA)	49.3	This present work

Table 5 Pseudo-first-order, pseudo-second-order Elovich and intraparticle diffusion model parameters for ibuprofen adsorption onto OPA at 30 °C

Kinetic model	Parameters	10 mg/L	20 mg/L	30 mg/L	40 mg/L	50 mg/L
Pseudo-first-order						
	k_1	0.017	0.029	0.031	0.041	0.049
	Q_e (cal) (mg/g)	0.316	0.283	0.288	0.333	0.686
	Q_e (exp) (mg/g)	4.750	9.993	13.962	19.953	24.712
	R^2	0.935	0.919	0.921	0.928	0.885
	SSE (%)	1.980	4.343	6.120	8.770	10.700
Pseudo-second-order						
	k_2	0.311	0.441	0.408	0.418	0.233
	Q_e (cal)	4.720	9.990	13.950	19.960	24.750
	Q_e (exp)	4.750	9.993	13.962	19.953	24.712
	R^2	0.999	1.000	1.000	1.000	1.000
	SSE (%)	0.013	0.001	0.005	0.003	0.017
Elovich						
	α	3.37E+25	4.503E+5	1.44E+67	1.76E+9	4.48E+3
	β	14.140	12.400	11.600	10.760	3.067
	R^2	0.868	0.968	0.931	0.843	0.849
Intraparticle						
	k_{diff}	0.031	0.032	0.034	0.033	0.083
	C	4.393	9.681	13.620	19.647	24.012
	R^2	0.968	0.929	0.893	0.896	0.839

Fig. 4 Plot of pseudo-second-order kinetic model at 30 °C for ibuprofen adsorption onto OPA
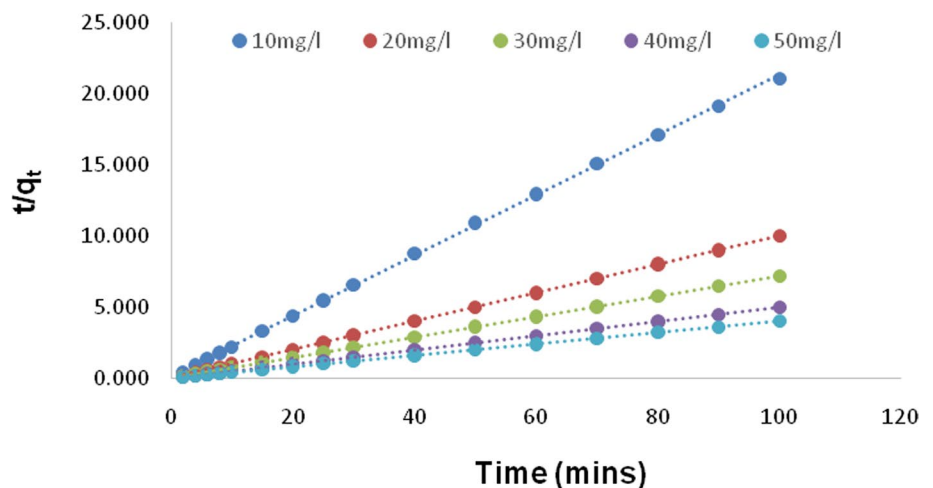
adsorbate/adsorbent interface in the course of the adsorption process. Negative $\Delta G°$ values obtained suggest the spontaneity of the process at the temperatures under consideration. Similarly, the energy of activation E_a was determined from Arrhenius expressions which indicate the plot of ln k_2 against 1/T for IBP sorption onto OPAC. This result is in consonance with that obtained by Ajemba (2014) for malachite green adsorption by activated clay from aqueous solutions.

Thermodynamic parameters are presented in Table 6. From the table, the negative value of $\Delta H°$ depicts the exothermic nature of the process. $\Delta H°$ and ΔS^0 values are 52.76 kJ/mol and 60.19 kJ/mol K, respectively. It was found that as temperature increases, the $\Delta G°$ values decreases; this implies that the force that drives the process decreases at higher temperature, resulting in reduced adsorption. To examine the mechanism governing the sorption process, Arrhenius expression, vis-à-vis activation energy, E_a, was employed. According to Nollet et al. (2003), for a physiosorption process, the energy of activation value range from 5 to 40 kJ/mol; while for chemisorption, the value is between 40 and 800 kJ/mol. For this work, the values of E_a obtained fall within that of a physical adsorption i.e. physisioption process.

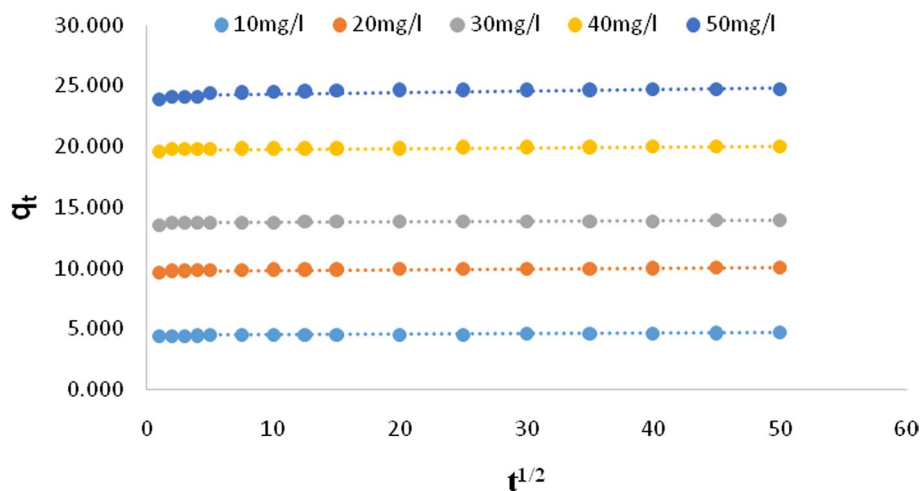**Fig. 5** Plot of intraparticle diffusion model of IBP adsorption onto OPA at 30 °C

Table 6 Thermodynamic parameters of IBP adsorption onto OPA at different temperatures

Temp(K)	ΔG° (kJ/mol)	ΔH° (kJ/mol)	ΔS° (kJ/molK)	Ea (kJ/mol)	A
303	−3487.5	-52.76	60.19	21.75	1634
313	−33,030.7				
323	−33,855.1				

Conclusion

In this research work, an inexpensive adsorbent using orange peel for the sorption of ibuprofen was prepared. The surface chemistry of OPR and OPAC was carried out, and it was found that there is an enlargement in the pore size of the OPAC; this is due to the acid activation resulting in improved performance. The kinetic data fitted pseudo-second-order kinetic model most. Thermodynamic study revealed that the reaction is exothermic and spontaneous. The sorption process followed a physisorption mechanism; hence, OPAC was both viable and effective in the removal of ibuprofen from aqueous solutions.

Compliance with ethical standards

Conflict of interest There is no conflict of interest among the authors.

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