ORIGINAL ARTICLE



Diversity of phytoplankton in some domestic wastewater-fed urban fish pond ecosystems of the Chota Nagpur Plateau in Bankura, India

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Received: 6 September 2017 / Accepted: 14 May 2018 / Published online: 22 May 2018 © The Author(s) 2018

Abstract

The present limnological investigation is conducted to study the relationship between phytoplankton abundance and five important physicochemical factors in urban wastewater-fed seven fish ponds of Chota Nagpur Plateau area. A total number of 43 phytoplankton taxa representing four classes, namely Cyanophyceae (7), Chlorophyceae (29), Bacillariophyceae (5) and Euglenophyceae (2), are thriving in these ponds which may suggest that different nutrient-rich wastewater supports the diversity and abundance of the phytoplankton. Different values of diversity indices, results of post hoc analysis and rarefaction curve are depicted spatial variations of phytoplankton abundance and physicochemical factors. From the Principal Component Analysis, out of 43 phytoplankton species, 23 important species are extracted. The canonical correspondence analysis presents that most of the phytoplankton species densities are associated with higher values of the physicochemical variables in these ponds. Correspondingly, in the present study, Algal Genus Pollution Index (AGPI) is employed to study the water quality of seven sites. From the AGPI score, it is revealed that Site 4 has probable high organic pollution and Site 2 and Site 3 have moderate organic pollution. Therefore, long-term intensive studies and proper management are necessary to protect these ponds toward eutrophication and degradation, because these ponds not only act as a safeguard of livelihoods but also contribute significantly at local level food and water security and economic prosperity.

Keywords Phytoplankton · Physicochemical factors · Diversity indices · Principle component · Post hoc analysis · Canonical correspondence analysis

Introduction

The information about phytoplankton composition and abundance provides knowledge about the trophic status of wetlands, which will aid to evaluate the possible or optimal usage of them. In an aquatic ecosystem, the base of the food chain is formed by the phytoplankton (Tas and Gonulal 2007; Saravanakumar et al. 2008) and they are responsible for more than 40% of Earth's photosynthetic production (Schmidt 2000). Therefore, variations in phytoplankton community structure alter the productivity of aquatic system where different organisms exist interdependently. Thus, phytoplankton not only acts as a water quality indicator but

also is useful for biomonitoring of lentic freshwater bodies, where multidimensional biological spectrum exists and ecological disturbance caused by alteration of different physicochemical factors (Ponmanickam et al. 2007; Shekhar et al. 2008). These physicochemical factors of aquatic system mostly depend on the discharge of domestic sewage, agricultural runoff water and other anthropogenic activities (Pal et al. 2014).

Several wetland characteristics, which may be spatial, temporal, physical, chemical and biological, have impaction phytoplankton assemblages. Under different irradiance and temperature, algal species show photosynthetic optima (Wehr and Sheath 2003) and species-specific photosynthetic response to light intensity depends on temperature (Wetzel 2001). Moreover, depending upon the temperature the CO₂ availability from the water–gas exchange and the bacterial metabolism are affected, resulting in the rate of photosynthesis altered (Prescott 1962). Both pH and conductivity are correlated with the water carbonate cycle. pH of an aquatic system mostly depends



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on the presence of carbonate, bicarbonate, carbonic acid, and free CO₂ in water (Wetzel 2001). Deviations in CO₂ concentration levels due to photosynthesis and respiration can alter the pH of the water. Also, CO₂ uptake by algae increases the pH. It is noted that upon different pH level, phytoplankton species have a different tolerance range. Phytoplankton assemblage is greatly influenced by pH in association with the salinity of the water (Sridhar et al. 2006; Saravanakumar et al. 2008). Bacillariophyceae dominates in slightly higher pH in large lakes in South India (Senthilkumar and Sivakumar 2008), whereas coccal green algae and desmids dominate in high altitude lakes having a less acidic pH (Das and Keshri 2012, 2013). Depending upon the concentration of total dissolved solids (TDS), i.e., an increase in different ions—especially bicarbonates, and carbonates in lake water, increase the electrical conductance (Goswami et al. 2010; Pal et al. 2013, 2018). Wetzel (2001) have considered that phytoplankton diversity is inversely correlated with TDS and conductivity. Badsi et al. (2012) have reported that freshwater phytoplankton adapts to the low salinity. However, depending upon the tolerance level and adaptive capability the pulses of phytoplankton abundance change with the fluctuation of salinity in the freshwater ecosystem.

Further, notwithstanding the chances for competitive exclusion, many species of phytoplankton can coexist owing to their distinctive needs and adaptations. From this, Hutchinson (1961) gives the concept of "paradox of the plankton" and proposes that several niches continually occur in the aquatic ecosystem which allows for phytoplankton diversity. It is not certain that the individual characteristics of an aquatic ecosystem influence community structure, but rather the amalgamation of certain characteristics. Thus, Wetzel (2001) stated that for particular quantification of the relationship with phytoplankton assemblages, a wide variety of characteristics of the aquatic system may not suffice but the associations are valuable for constructing general correlations.

Bankura district is situated in the eastern part of India and is a part of the lower edge of the Chota Nagpur Plateau. The Chota Nagpur Plateau area is internationally famous for coal, bauxite and iron mining. The soil is not fertile enough and of laterite type mixed with sand and rock, red in color. Several ponds and drains are there in this district, but most of them remain dry in the summer. Most of the ponds are used for household purposes and pisciculture practices in monsoon. Furthermost, these ponds receive domestic sewage and depending upon the character and amount of sewage the characteristics of ponds alter. Ali et al. (1999) examined that composite wastewater which contains heavy metals causes the formation of phytoplankton bloom in lakes. Phytoplankton assemblage of water body beside the agricultural field and industrial area in West Bengal, India, is greatly

influenced by agricultural and industrial wastes (Ghosh and Keshri 2011; Das et al. 2011, 2015).

Though various studies have indicated the influence of different abiotic factors on phytoplankton communities as a whole (Mukhopadhyay et al. 1997; Chakraborty and Das 2004; Chakraborty et al. 2004; Chattopadhyay and Banerjee 2007), no work has been done yet in this region. Thus, an assessment of phytoplankton community structure and five very important abiotic factors is done to understand the numerous functional properties of phytoplankton species. Moreover, this information can be cast off to assess which, if any, of the factors have an impact on phytoplankton community structure. The five abiotic factors and phytoplankton population structure were tested statistically to define whether any relationships are present. Moreover, due to short life span, easy to quantitative determination and rapid response to pollutants algae act as a bioindicator of water quality changes. Thus, in the present study, the water pollution index (AGPI) is used to detect, if any, organic pollution extant in the water bodies of these areas.

Materials and methods

Study area

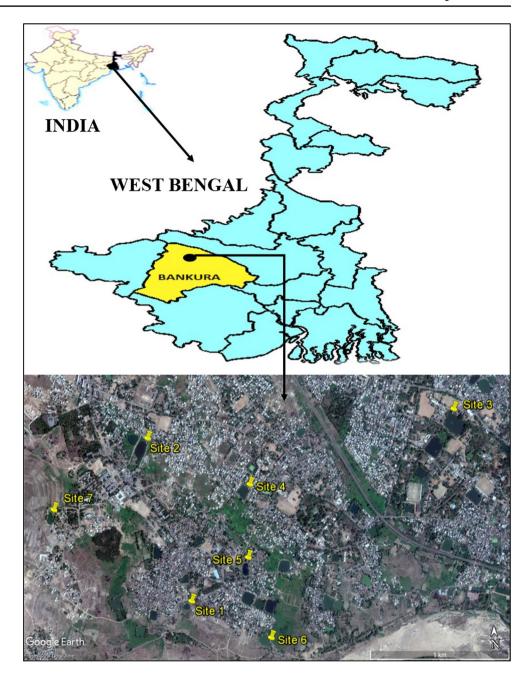
In the Eastern part of the Chota Nagpur Plateau within Damodar and Darakeswar river basin and in the periphery of Bankura Town, seven study sites were selected for the present study (Fig. 1). This area has undulating landform, interspersed rocky hillocks, laterite soil with slope from west to east. From March to early June due to hot westerly winds, the temperature laid between 37 and 46 °C and for this reason the little amount of water remains in all water bodies including the rivers also. In the monsoon (July to September), the average rainfall ranged between 1200 and 1400 mm, whereas in dry winter (October to February) the temperature varied between 8 and 20 °C. The domestic sewage-fed seven selected sites were used for pisciculture and household purposes like bathing, washing clothes and kitchen utensils. The location, area and brief description of all sites are presented in Table 1.

Physicochemical analysis of water

The sampling of the present study was carried out once in a month from January 2017 to July 2017. Water samples were collected in airtight polyvinyl chloride (PVC) bottles. The pH, temperature (TEMP), conductivity (COND), total dissolved solids (TDS) and salinity (SAL) were measured by Eutech PCS Tester-35 Multiparameter testing equipment.



Fig. 1 Map showing the study sites (Sites 1–7; Google Earth, 2017) nearby Bankura town region (India and West Bengal maps are not in scale.)



Phytoplankton collection, identification and analysis

Samples were collected from the different parts of the water bodies. Four samples were collected from each water body per month. One liter of water sample was collected from each collecting point, and 10% Lugols' Iodine solution was mixed instantly on the spot in 100:1 ratio. All collections were done before 10 am in the morning. Sampling bottles were kept in the laboratory for about 6 h. Then, the 10-mL precipitation was collected from the amber bottle. From the precipitated samples, all workout has been done and 5% formalin solution was used to preserve it for the future reference. The identification and quantitative analysis of phytoplankton (PHYTO) samples were done by using Carl Zeiss Axiostar microscope with photomicrometry Nikon camera attachment. The phytoplankton was identified by several monographs, viz. Turner (1892), Husted (1930), Desikachary (1959), Prescott (1962) and Philipose (1967).

Statistical analysis

To show phytoplankton community structure, the diversity indices like Shannon–Wiener diversity index $[H'(log_e)]$, Simpson's dominance index (D_{SIPM}) and Pielou's evenness index (J') were used. It was noted that the Shannon–Wiener



Table 1 A brief description of seven selected study sites

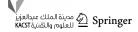
Sites	Site location	Area (ha)	Description
Site 1	23°13′35.27″N, 87° 2′27.82″E	0.57	Over 20% area covered with Eichornia crassipes, Pistia stratiotes, Spirodela polyrhiza and Wolffia globosa, whereas in the bank, Colocasia esculenta and Polygonum plebeium are dominated
Site 2	23°14′12.04″N, 87° 2′17.71″E	1.44	20% area covered mostly by <i>E. crassipes</i> and <i>Salvinia molesta</i> . Also, dense marginal macrophytes dominated by <i>Alternanthera sessilis</i> , <i>Commelina bengalensis</i> , and <i>Eclipta alba</i> are present
Site 3	23°14′16.40″N, 87° 3′34.66″E	2.14	70% of the bank is concreted. Rest of the bank area wetland macrophytes like <i>C. esculenta</i> , <i>P. plebeium</i> , <i>A. sessilis</i> , <i>C. bengalensis</i> are present. A little amount of <i>E. crassipes</i> , <i>P. stratiotes</i> present in open water
Site 4	23°14′0.00″N, 87° 2′43.00″E	0.92	Water body with a dense <i>Azolla pinnata</i> on surface water and <i>Najas graminea</i> , <i>Hydrilla verticillata</i> in under water. <i>A. sessilis</i> , <i>P. plebeium</i> , and <i>Ipomoea aquaica</i> are mostly present in bank region
Site 5	23°13′45.00″N, 87°2′42.00″E	1.19	Dominated by <i>E. crassipes</i> , <i>Lemna minor</i> , <i>P. barbatum</i> , <i>S. polyrhiza</i> over the 40% of the surface water. In the bank, the dominated macrophytes are <i>I. aquaica</i> , <i>A. sessilis</i> , <i>C. benghalensis</i> , <i>Centella asiatica</i>
Site 6	23°13′26.64″N, 87° 2′47.90″E	0.81	Clear surface water and presence of <i>Ceratophyllum demersum</i> in under water and <i>Marsilea minuta</i> , <i>C. benghalensis</i> , <i>I. aquaica</i> are present in the bank
Site 7	23°13′56.99″N, 87° 1′54.03″E	0.52	Dominated by <i>Nymphoides hydrophylla</i> . In bank region, dense mat <i>Hygrophila spinosa</i> , <i>Marsilea minuta</i> is observed

diversity index was more subtle to rare species and Simpson's index gave more prominence to common species, while Pielou's index gave the indication about the species homogeneity (Adhikari et al. 2017). Beta diversity was measured to know about the differences in species composition among the study sites. Spatially constrained individual-based rarefaction was used to estimate the species richness that was directly comparable to the areas that differed in spatial extent. All these analyses were calculated by using PAST (version 3.15, 2017) software.

In any ecosystem, the physicochemical factors influence the biotic interactions and in most cases, univariate analyses were inadequate to assess interactions. Thus, multivariate analyses were practically used to understand the role of physicochemical factors in influencing biotic community from different ecosystems. Here the selection of methods for statistical analyses was done following Quinn and Keough (2002). Principal component analysis (PCA) was executed with Kaiser normalization, and Varimax rotation was used as a dimension-reducing technique and to comment on the group or groups of important phytoplankton species in those study sites. Forty-three phytoplankton species of seven study sites were subjected to PCA to reduce the number of dimensions. Here first three components of PCA were taken into consideration, and factor loadings (FL)>0.7 at any component were considered to be significant. Canonical correspondence analysis (CCA) was used to evaluate the relationship, if any, between abiotic factors and phytoplankton abundance of seven study sites. Twenty-three phytoplankton species were shortlisted (based on PCA) from 7 study sites, in order to evaluate how different community was structured by their environmental characteristic. The significance of the CCA axes and respective environment variables was evaluated by the Monte Carlo tests with 999 unrestricted permutations (TerBraak and Prentice, 1988). Post hoc analysis was performed to know about the significant difference in abiotic factors and phytoplankton abundance within each site. The two-dimensional hierarchical cluster analyses were known as dendrogram, and by linkage methods, this dendrogram was constructed. In this method, groups were fused according to the distance between their nearest members. Thus, to show the affinity between study sites, the dendrograms were constructed based on abiotic factors and phytoplankton abundance. Statistical analyses were performed using Statistica for Windows 8.0 (StatSoft Inc. 2007) software and PAST (version 3.15, 2017) software. The Graphs were prepared by using Origin Pro 2016.

Results and discussion

Overall, 43 phytoplankton species are identified, of which 7 species belong to the class Cyanophyceae, 14 belongs to class Chlorophyceae, and 5 belongs to class Bacillariophyceae and 2 species of Euglenophyceae. The abundance of *Oscillatoria limosa* is the highest in Site 1, Site 3 and Site 6, while *Chlorella vulgaris* in Site 2, *Merismopedia minima*, *Anabaena circinalis* in Site 5, *Spirogyra maxima* in Site 7 are most abundant. It is noted that under the family Euglenophyceae two species like *Euglena acus* and *Phacus acuminatus* are only observed at Site 4 ("Appendix 1"). Considering all sites the percentage contribution of Chlorophyceae is the



highest trailed by Cyanophyceae and Bacillariophyceae. The same trends are followed in sitewise calculation except Site 3 and Site 4 where Cyanophyceae is more abundant than Chlorophyceae (Fig. 2). The trend of phytoplankton density (number mL^{-1}) is Site 4> Site 5> Site 3> Site 1> Site 7> Site 6> Site 2, while Site 4> Site 5> Site 7> Site

3> Site 2> Site 6> Site 1 is the trend of number of phytoplankton species assemblage. The hierarchical cluster analysis (Single Linkage Euclidean Distance) of this study based on phytoplankton abundance placed Site 2, Site 3, Site 6 and Site 7 in a close cluster, and Site 1 and Site 5 were distantly clustered with them, whereas Site 4 is an out group (Fig. 3a).

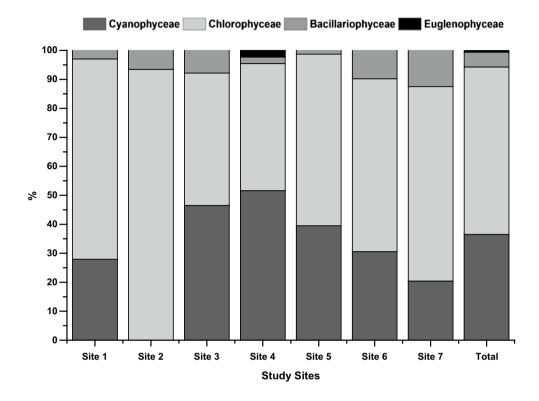


Fig. 2 Percentage distribution of Phytoplankton families in each site and altogether seven sites

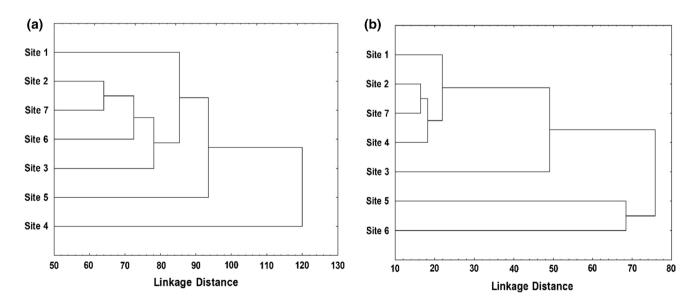


Fig. 3 Hierarchical cluster analysis (single linkage Euclidean distances) of seven different sites depending on phytoplankton assemblage (a) and physicochemical factors (b)



Table 2 Spatial changes in diversity indices: Shannon—Wiener general diversity index $H'(\log_e)$, Simpson's dominance index (D_{SIPM}) and Pielou's evenness index (J')

	$H'(\log_e)$	D_{SIMP}	J'
Site 1	1.546	0.770	0.782
Site 2	2.014	0.835	0.749
Site 3	2.113	0.846	0.689
Site 4	2.541	0.903	0.705
Site 5	2.259	0.861	0.684
Site 6	1.690	0.788	0.774
Site 7	2.390	0.899	0.839

That means the phytoplankton species composition of Site 4 is different from all other sites.

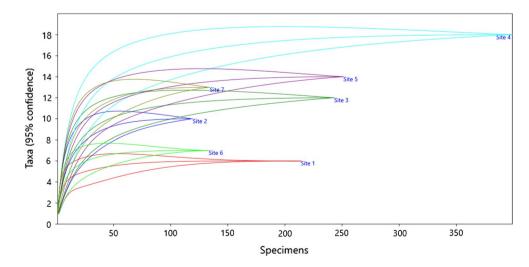
Sitewise values of diversity indices, viz. Shannon–Wiener diversity index $[H'(\log_e)]$, Simpson's dominance index (D_{SIPM}) and Pielou's evenness index (J'), are depicted in Table 2. The species diversity $H'(\log_e)$ and D_{SIPM} values are high in Site 4, Site 5, Site 7 and Site 3 and low at Site 1 and Site 2. But except Site 7, J' showed a reverse trend. It is noted that $H'(\log_e)$ is more sensitive to rare species, whereas D_{SIPM} gives importance to the common species. That means here in Site 4, Site 5, Site 7 and Site 3 high phytoplankton diversity reflects the dominance of common phytoplankton species, while the opposite phenomenon is shown in Site 1 and Site 2. However, low values of species dominance reflect high evenness of species distribution. In

the present study due to high J' values in Site 1, Site 2 and Site 6 cause low D_{SIPM} values. However, due to the difference in species abundance and composition, the value of beta diversity (Bw) of Site 1 is different with Site 3, Site 4, Site 5 and Site 6, while very similar with Site 2 and Site 7(Table 3). However, except Site 1, the species composition of Site 2 is highly similar to other sites. Site 3 is more similar with Site 5 than Site 4 and Site 7, while with Site 6 low similarity is found. Though the values of $H'(\log_e)$ and D_{SIPM} of Site 7 are very close to Site 3, Site 4, Site 5 and Site 6, the lower similarity of species composition is noticed from the Bw values. The individualbased spatially constrained rarefaction curve gives more idea about species richness of samples that differ in the area and sampling effort (Chiarucci et al. 2009). Though the overall diversity of seven sites does not differ much, the species accumulation curves are constructed, which calculated the expected number of species in a subsample drawn randomly from a single representative sample from an assemblage (Fig. 4). The individual-based rarefaction curves amply pointed out that Site 4 followed by Site 5 and Site 3 has higher species richness than other sites, and this result corroborates with the Bw values. However, the post hoc analysis (Tukey's HSD) gives better understanding regarding spatial differences in phytoplankton assemblages at p < 0.05 level (Fig. 5).

Table 3 Whittaker beta diversity (B_w) within seven study sites

	Site 1					
Site 2	1.00	Site 2				
Site 3	0.67	0.91	Site 3			
Site 4	0.75	0.93	0.73	Site 4		
Site 5	0.60	0.83	0.85	0.63	Site 5	
Site 6	0.69	0.88	0.58	0.76	0.81	Site 6
Site 7	1.00	0.74	0.76	0.81	0.78	0.700

Fig. 4 Individual-based rarefaction curves calculated on the basis of seven study sites under study





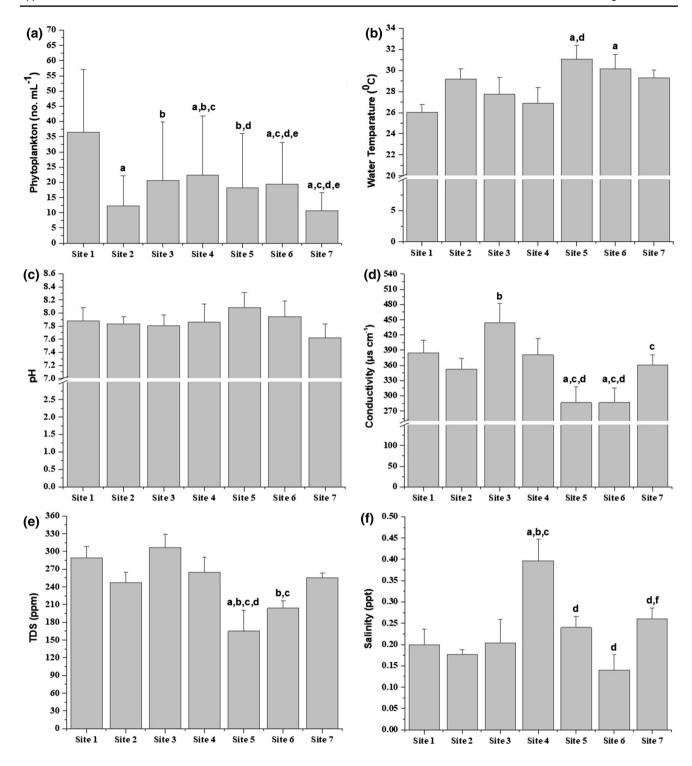


Fig. 5 Phytoplankton assemblage and physicochemical factors at seven selected study sites. By using post hoc analysis (Tukey's HSD), the site which differs significantly (p < 0.05) with others sites, marked as follows: a Site 1, b Site 2, c Site 3, d Site 4, e Site 5, f Site 6

It is very important to monitor physicochemical factors in the aquatic system for better understanding about their influences on the distribution of aquatic diversity (Das et al. 2012, 2015). In the present study, the five most important physicochemical factors are assessed and sitewise

graphically represented in Fig. 5. The mean water temperature (WT), which is an important factor to control aquatic life, varies between 25 and 31 °C from different sites. Significant spatial differences in WT (p < 0.05) are noted between Site 1 and Site 5, Site 6 and Site 4 with Site 5. Though



the recorded temperature range is good for plankton growth (Wassie and Melese 2017), it can be noted that increase of 1 °C WT increases the ionic mobility and solubility of salts, which proliferate conductivity 2–4% (Miller et al. 1988) and effects on phytoplankton assemblage. The water pH is slightly basic, i.e., 7.5–8, in seven study sites, and this pH range not only is better for phytoplankton but also is healthy for aquatic life including fish and fall (Oso and Fagbuaro 2008). TDS is the sum of all ion particles smaller than 2 microns, and its value beyond the safe limit of 500 ppm (Hem 1985; EPA 2012) causes the cell to swell or shrink and affect aquatic organism to move in the water column. The TDS of all the sites, which is mostly contributed by the domestic sewage, lies between 136 and 303 ppm, and these values are very conductive for the aquatic life (Mohamed et al. 2009). The conductivity (COND) is a measure of a solution's conductive abilities that affect the ecological feasibility of shallow freshwater ponds, viz. the present study sites. Actually COND is a primary indicator of alteration in aquatic system and it depends on natural flooding, evaporation and pollution load. It is noted that an increase in water COND decreases the phosphorus availability to the phytoplankton that results in decline of phytoplankton populations (Chouyyok et al. 2010; López-Flores et al. 2014). The average COND in seven sites is 272–431 μS cm⁻¹ which is lower than safe limits (2000 µS cm⁻¹) of freshwater lakes criteria (Kemker 2015), and it indicates healthy pond ecosystem. Significant spatial differences (p < 0.05) for both TDS and COND are shown same for Site 1 with Site 5 and Site 6; Site 3 with Site 5 and Site 6; Site 4 with Site 5. Except these, the TDS of Site 2 significantly differs with Site 5 and Site 5 with Site 7; likewise COND of Site 2 significantly differ with Site 3, Site 3 with Site 7 and Site 4 with Site 6. Nielsen et al. (2003) has suggested that salinity of freshwater lakes above 1ppt creates adverse effect on aquatic biota; especially some studies (Redden and Rukminasari 2008; Flöder et al. 2010; Larson and Belovsky 2013) indicate that high salinity in aquatic system can decrease phytoplankton concentration as well as it affects the species composition. But here in the seven sites, the salinity ranges from 0.13 to 0.39 ppt, which is required to maintain physiological process as well as the growth of the phytoplankton. Post hoc analysis at p < 0.05shows that the salinity levels of Site 4 were significantly different with other six sites, and it may be due to higher sewage input in Site 4 than other sites (Fig. 5). It is also revealed from the present study that by mixing of domestic sewage and agricultural runoff, which contains numerous luxuriant nutrients and different salts of sodium, potassium, calcium, and magnesium, presently maintained the favorable condition for phytoplankton production (Fig. 6).

The species dynamics of phytoplankton have changed with the alteration of physicochemical factors. Each species

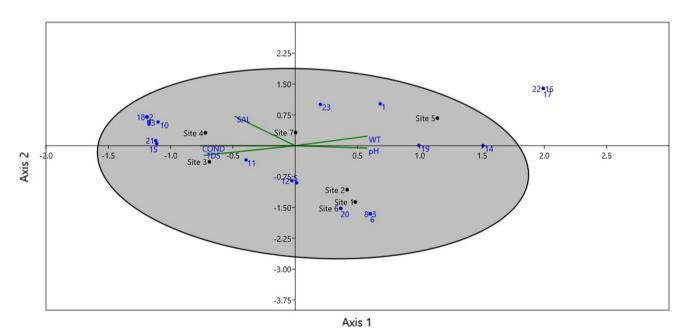


Fig. 6 Canonical correspondence analysis (CCA) ordination diagram showing scatter plot for selected phytoplankton, environmental variables and seven study sites. Vector lines represent the relationship of significant environmental variables to the ordination axes; their length is proportional to their relative significance. ($1=Anabaena\ circinalis$, $2=Ankistrodesmus\ falcatus$, $3=Chlorella\ vulgaris$, $4=Chroococcus\ limneticus$, $5=Closterium\ parvulum$, 6=Dimorpho-

coccus lunatus, 7=Euglena acus, 8=Eunotia pectinalis, 9=Gonium pectorale, 10=Merismopedia minima, 11=Mougeotia punctata, 12=Nitella mucronata, 13=Oocystis elliptica, 14=Oocystis pusilla, 15=Oscillatoria major, 16=Pediastrum duplex, 17=Pediastrum tetras, 18=Phacus acuminatus, 19=Scenedesmus obliquus, 20=Spirogyra gracilis, 21=Spirogyra weberi, 22=Spirulina major, 23=Tetraedron trigonum)



respond differently to the environmental factors. So, to find out this relationship different canonical correspondence analysis (CCA) is done. But, before that, the factor analysis is performed to identify the principle component within the long list of phytoplankton at seven study sites in the Chota Nagpur Plateau. The significant presence of phytoplankton is recognized depending upon factor loadings (FL)>0.7 of the first three components that explained 68.26% of phytoplankton species and out of a total of 43 species of phytoplankton only 23 phytoplankton species are shortlisted as significant species (Table 4). Within this 23 phytoplankton species, 5 are from Cyanophyceae, 17 from Chlorophyceae, only 1 from Bacillariophyceae and no representation of Euglenophyceae found.

Canonical correspondence analysis (CCA) exhibits that the densities of most of the phytoplankton species are associated with higher values of the physicochemical variables. The result of the test of significance using the Monte Carlo test on CCA to highlight the phytoplankton species-environmental variable relations is depicted in Table 5. The eigenvalues and the percentages of variance in axis 1 are found to be higher than axis 2, which corroborates the findings of Liu et al. (2010) and Sharma et al. (2016). Between physicochemical factors and phytoplankton species, the eigenvalue for axis 1 (0.5550) explains 38.47% correlation, while axis 2 (0.416) explicates 29.06% correlation. A. circinalis, O. pusilla, P. duplex, P. tetras, S. obliquus and S. major indicate that they are positively correlated with the higher value of WT and pH. The TDS and COND are positively correlated with each other (Pal et al. 2013; Das et al. 2015), and with them O. major, S. weberi has a positive relationship. The distribution of A. falcatus, C. limneticus, E. acus, G. pectoral, M. minima, O. elliptica, P. acuminatus is found to be positively dependant on salinity. However, some phytoplankton species, viz. Chlorella vulgaris, Closterium parvalum, Dimorphococcus lunatus, Eunotia pectinalis, Nitella mucronata are found to be least affected by the variation of measured physicochemical factors.

Conclusion

The physicochemical and biological characters change in the lentic ecosystem due to the discharge of wastes, which increase the concentration of different chemicals (Roy Goswami et al. 2011, 2013; Pal et al. 2014). From the present study, it is apparent that a respectable number of phytoplankton species thrived in these wastewater-fed ponds, when added to these ponds harbor different floating and marginal macrophytes. That means this urban wastewater, which contained several nutrients, not only enriches the phytoplankton diversity and abundances in these ponds but also is helpful for pisciculture practices

Table 4 Factor loadings of phytoplankton species from seven different sites

Ankistrodesmus falcatus - 0.984 0.014 0.075 Aulosira implexa - 0.646 0.654 0.250 Bulbochaete satigera 0.293 - 0.233 - 0.182 Chara braunii 0.319 - 0.348 - 0.044 Chlorella vulgaris 0.039 0.087 - 0.908 Chroococcus limneticus - 0.984 0.014 0.075 Cladophora glomerata - 0.338 0.015 0.368 Closterium parvulum 0.245 - 0.098 - 0.877 Closterium ehrenbergii 0.145 - 0.528 0.335 Coeleastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.558 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 <	Phytoplankton species	PC 1	PC 2	PC 3
Aulosira implexa - 0.646 0.654 0.250 Bulbochaete satigera 0.293 - 0.233 - 0.182 Chara braunii 0.319 - 0.348 - 0.044 Chlorella vulgaris 0.039 0.087 - 0.908 Chroococcus limneticus - 0.984 0.014 0.075 Cladophora glomerata - 0.338 0.015 0.368 Closterium parvulum 0.245 - 0.098 - 0.877 Closterium ehrenbergii 0.145 - 0.528 0.335 Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Euglena decus - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Meri	Anabaena circinalis	- 0.283	0.816	0.258
Bulbochaete satigera 0.293 - 0.233 - 0.182 Chara braunii 0.319 - 0.348 - 0.044 Chlorella vulgaris 0.039 0.087 - 0.908 Chroococcus limneticus - 0.984 0.014 0.075 Cladophora glomerata - 0.338 0.015 0.368 Closterium parvulum 0.245 - 0.098 - 0.877 Closterium ehrenbergii 0.145 - 0.528 0.335 Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Eunotia pectinalis 0.039 0.087 - 0.908 Merismopedia minima - 0.984 0.014 0.075 <td< td=""><td>Ankistrodesmus falcatus</td><td>- 0.984</td><td>0.014</td><td>0.075</td></td<>	Ankistrodesmus falcatus	- 0.984	0.014	0.075
Chara braunii 0.319 - 0.348 - 0.044 Chlorella vulgaris 0.039 0.087 - 0.908 Chroococcus limneticus - 0.984 0.014 0.075 Cladophora glomerata - 0.338 0.015 0.368 Closterium parvulum 0.245 - 0.098 - 0.877 Closterium ehrenbergii 0.145 - 0.528 0.335 Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Eunotia pectinalis 0.039 0.087 - 0.908 Merismopedia minima - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335	Aulosira implexa	-0.646	0.654	0.250
Chlorella vulgaris 0.039 0.087 - 0.908 Chroococcus limneticus - 0.984 0.014 0.075 Cladophora glomerata - 0.338 0.015 0.368 Closterium parvulum 0.245 - 0.098 - 0.877 Closterium ehrenbergii 0.145 - 0.528 0.335 Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554	Bulbochaete satigera	0.293	-0.233	- 0.182
Chroococcus limneticus - 0.984 0.014 0.075 Cladophora glomerata - 0.338 0.015 0.368 Closterium parvulum 0.245 - 0.098 - 0.877 Closterium ehrenbergii 0.145 - 0.528 0.335 Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitzschia sigma 0.198 - 0.313 0.033 <t< td=""><td>Chara braunii</td><td>0.319</td><td>-0.348</td><td>- 0.044</td></t<>	Chara braunii	0.319	-0.348	- 0.044
Cladophora glomerata - 0.338 0.015 0.368 Closterium parvulum 0.245 - 0.098 - 0.877 Closterium ehrenbergii 0.145 - 0.528 0.335 Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.333 Mougeotia punctata - 0.938 - 0.033 0.039 Mitzalia mucronata 0.258 - 0.113 - 0.844	Chlorella vulgaris	0.039	0.087	- 0.908
Closterium parvulum 0.245 - 0.098 - 0.877 Closterium ehrenbergii 0.145 - 0.528 0.335 Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.333 Morismopedia minima - 0.938 - 0.033 0.039 Morismopedia minima - 0.938 - 0.011 - 0.554 Nitzla mucronata 0.258 - 0.113 - 0.844	Chroococcus limneticus	- 0.984	0.014	0.075
Closterium ehrenbergii 0.145 - 0.528 0.335 Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.984 0.014 0.075 Merismopedia minima - 0.988 - 0.033 0.039 Mougeotia punctata 0.258 - 0.113 - 0.844 Nitzschia amphibia - 0.107 - 0.294 0.053 Nit	Cladophora glomerata	-0.338	0.015	0.368
Coelastrum microporum 0.237 0.439 0.457 Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis pusilla 0.263 0.879 0.380 Oscillat	Closterium parvulum	0.245	- 0.098	- 0.877
Coleochaete islamii 0.397 0.411 0.024 Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia amphibia - 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis pusilla 0.263 0.879 0.380 Oscilla	Closterium ehrenbergii	0.145	- 0.528	0.335
Cymbella acuta 0.349 - 0.559 0.072 Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia amphibia - 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscil	Coelastrum microporum	0.237	0.439	0.457
Desmidium baileii 0.145 - 0.528 0.335 Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 P	Coleochaete islamii	0.397	0.411	0.024
Dimorphococcus lunatus 0.039 0.087 - 0.908 Euglena acus - 0.984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0.908 Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum tetras 0.215 0.879 0.258 Pha	Cymbella acuta	0.349	- 0.559	0.072
Euglena acus - 0,984 0.014 0.075 Eunotia pectinalis 0.039 0.087 - 0,908 Gonium pectorale - 0,984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0,938 - 0.033 0.039 Mougeotia punctata - 0,814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0,844 Nitzschia sigma 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0,984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum tetras 0.215 0.879 0.258 Phacus acu		0.145	- 0.528	0.335
Eunotia pectinalis 0.039 0.087 — 0.908 Gonium pectorale — 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 — 0.528 0.335 Merismopedia minima — 0.938 — 0.033 0.039 Mougeotia punctata — 0.814 0.071 — 0.554 Nitella mucronata 0.258 — 0.113 — 0.844 Nitzschia sigma 0.198 — 0.303 — 0.625 Oedogonium fragile 0.404 — 0.532 0.016 Oedogonium aquaticum 0.293 — 0.233 — 0.182 Oocystis elliptica — 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 — 0.549 0.565 Oscillatoria major — 0.766 — 0.332 0.285 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus — 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 — 0.570 <	Dimorphococcus lunatus	0.039	0.087	- 0.908
Gonium pectorale - 0.984 0.014 0.075 Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia amphibia - 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570	Euglena acus	- 0.984	0.014	
Hyalotheca dissiliens 0.145 - 0.528 0.335 Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia amphibia - 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214	Eunotia pectinalis	0.039	0.087	- 0.908
Merismopedia minima - 0.938 - 0.033 0.039 Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia amphibia - 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus obliquus 0.320 0.744 - 0.490	Gonium pectorale	- 0.984	0.014	0.075
Mougeotia punctata - 0.814 0.071 - 0.554 Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia amphibia - 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus obliquus 0.320 0.744 - 0.490	Hyalotheca dissiliens		- 0.528	0.335
Nitella mucronata 0.258 - 0.113 - 0.844 Nitzschia amphibia - 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus bijugatus 0.039 0.087 - 0.908 Spirogyra gracilis - 0.141 0.033 0.792	Merismopedia minima	- 0.938	- 0.033	0.039
Nitzschia amphibia - 0.107 - 0.294 0.053 Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus bijugatus 0.039 0.087 - 0.908 Scenedesmus obliquus 0.320 0.744 - 0.490 Spirogyra gracilis - 0.141 0.033 0.792	Mougeotia punctata	- 0.814	0.071	- 0.554
Nitzschia sigma 0.198 - 0.303 - 0.625 Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus bijugatus 0.039 0.087 - 0.908 Scenedesmus obliquus 0.320 0.744 - 0.490 Spirogyra gracilis - 0.141 0.033 0.792 Spirogyra maxima 0.314 - 0.616 0.182 <th< td=""><td>Nitella mucronata</td><td>0.258</td><td>- 0.113</td><td>- 0.844</td></th<>	Nitella mucronata	0.258	- 0.113	- 0.844
Oedogonium fragile 0.404 - 0.532 0.016 Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus bijugatus 0.039 0.087 - 0.908 Spirogyra gracilis - 0.141 0.033 0.792 Spirogyra weberi - 0.828 - 0.286 0.259 Spirogyra maxima 0.314 - 0.616 0.182 Spirogyra maxima 0.314 - 0.616 0.182 Sy	Nitzschia amphibia	-0.107	- 0.294	0.053
Oedogonium aquaticum 0.293 - 0.233 - 0.182 Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus bijugatus 0.039 0.087 - 0.908 Scenedesmus obliquus 0.320 0.744 - 0.490 Spirogyra gracilis - 0.141 0.033 0.792 Spirogyra maxima 0.314 - 0.616 0.182 Spirogyra maxima 0.314 - 0.616 0.182 Spirolina major 0.215 0.879 0.258 Syned	Nitzschia sigma	0.198	- 0.303	- 0.625
Oocystis elliptica - 0.984 0.014 0.075 Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus bijugatus 0.039 0.087 - 0.908 Scenedesmus obliquus 0.320 0.744 - 0.490 Spirogyra gracilis - 0.141 0.033 0.792 Spirogyra weberi - 0.828 - 0.286 0.259 Spirogyra maxima 0.314 - 0.616 0.182 Spirulina major 0.215 0.879 0.258 Synedra acus - 0.465 0.056 0.670 Tetraedron tr	Oedogonium fragile	0.404	-0.532	0.016
Oocystis pusilla 0.263 0.879 0.380 Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus bijugatus 0.039 0.087 - 0.908 Scenedesmus obliquus 0.320 0.744 - 0.490 Spirogyra gracilis - 0.141 0.033 0.792 Spirogyra weberi - 0.828 - 0.286 0.259 Spirogyra maxima 0.314 - 0.616 0.182 Spirulina major 0.215 0.879 0.258 Synedra acus - 0.465 0.056 0.670 Tetraedron trigonum - 0.704 0.602 0.238 Eigenvalue </td <td>Oedogonium aquaticum</td> <td>0.293</td> <td>- 0.233</td> <td>- 0.182</td>	Oedogonium aquaticum	0.293	- 0.233	- 0.182
Oscillatoria limosa 0.296 - 0.549 0.565 Oscillatoria major - 0.766 - 0.332 0.285 Pediastrum duplex 0.215 0.879 0.258 Pediastrum tetras 0.215 0.879 0.258 Phacus acuminatus - 0.984 0.014 0.075 Pleurotaenium trabecula 0.186 0.698 - 0.570 Rhizoclonium lapponicum - 0.310 0.132 0.214 Scenedesmus bijugatus 0.039 0.087 - 0.908 Scenedesmus obliquus 0.320 0.744 - 0.490 Spirogyra gracilis - 0.141 0.033 0.792 Spirogyra weberi - 0.828 - 0.286 0.259 Spirogyra maxima 0.314 - 0.616 0.182 Spirulina major 0.215 0.879 0.258 Synedra acus - 0.465 0.056 0.670 Tetraedron trigonum - 0.704 0.602 0.238 Eigenvalue 12.608 8.772 7.972	Oocystis elliptica	- 0.984	0.014	0.075
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Spirulina major 0.215 0.879 0.258 Synedra acus - 0.465 0.056 0.670 Tetraedron trigonum - 0.704 0.602 0.238 Eigenvalue 12.608 8.772 7.972	Spirogyra weberi	- 0.828	- 0.286	0.259
Synedra acus - 0.465 0.056 0.670 Tetraedron trigonum - 0.704 0.602 0.238 Eigenvalue 12.608 8.772 7.972				0.182
Tetraedron trigonum - 0.704 0.602 0.238 Eigenvalue 12.608 8.772 7.972	Spirulina major			0.258
Eigenvalue 12.608 8.772 7.972	•			0.670
				0.238
Cumulative % 29.321 49.721 68.262	=			
	Cumulative %	29.321	49.721	68.262

Average extraction: principal components with Kaiser normalization. Factor loadings > 0.7 in each component were considered as significant and are marked with bold faces



Table 5 Monte Carlo test of canonical correspondence analysis (CCA) using overall mean values to indicate phytoplankton—environment relationship at selected study sites

Axes	1	2	3	4	Total inertia
Eigenvalues	0.550	0.416	0.292	0.173	1.431
Cumulative percentage variance of species—environment relation	38.47	67.53	87.91	100.0	

Summary of Monte Carlo test (999 permutations under reduced model)

Test of significance of first canonical axis: eigenvalue = 0.550

p value = 0.006

Test of significance of all canonical axes: trace = 1.431

p value = 0.001

Table 6 Pollution index of algal genera according to Palmer (1969) at seven study sites

Algal Genus	Pollution index	Site 1	Site 2	Site 3	Site 4	Site 5	Site 6	Site 7
Cyanophyceae			1		1		,	
Oscillatoria	5	5	_	5	5	_	5	_
Chlorophyceae								
Ankistrodesmus	2	2	_	_	2	_	_	_
Chlorella	3	_	3	_	_	_	_	_
Closterium	1	_	1	1	_	_	_	1
Scenedesmus	4	_	4	_	_	4	_	4
Bacillariophyceae								
Nitzschia	3	_	3	3	3	_	3	3
Synedra	2	2	_	2	2	2	_	_
Euglenophyceae								
Euglena	5	_	_	_	5	_	_	_
Phacus	2	_	_	_	2	_	_	_
Total score		9	11	11	19	6	8	8

Following numerical values for pollution classification: 0–10 = lack of organic pollution; 10–15 = moderate pollution; 15–20 = probable high organic pollution; 20 or more = high organic pollution

and it will surely improve the local economy. Also as per Palmer's (1969) Algal Genus Pollution Index (Table 6), it is revealed from the present study that Site 4 has probable high organic pollution, Site 2 and Site 3 have moderate organic pollution, and rest of sites have lack of organic pollution. However, necessary precautions and management like control of floating macrophytes density, nutrient loading within these ponds and regular monitoring are needed to check eutrophication and these will support a host of planktonic populations as well as the pisciculture practice. However, intensive long-term studies are important to protect these productive systems from rapid urbanization and industrialization.

Acknowledgements The authors express their deep sense of gratitude to Principal, Bankura Sammilani College, West Bengal, for providing

laboratory facilities. The last author thankfully acknowledges University Grants Commission (UGC) (Grant No. (BSR)BL/15-16/0006), Government of India, for Dr. D.S. Kothari Postdoctoral Fellowship.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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 Table 7
 Phytoplankton diversity and density (number mL^{-1}) in selected seven study sites

Phytoplankton species	Site 1	Site 2	Site 3	Site 4	Site 5	Site 6	Site 7
Cyanophyceae							'
Anabaena circinalis	_	_	_	42 ± 6	73.7 ± 10.3	_	13.3 ± 5.1
Aulosira implexa	_	_	_	22.3 ± 5.1	20 ± 5.6	_	_
Chroococcus limneticus	_	_	_	8.3 ± 3.1	_	_	_
Merismopedia minima	_	_	_	75 ± 5.6	_	_	15 ± 5.6
Oscillatoria limosa	60 ± 6.2	_	70.3 ± 9.61	_	_	41.7 ± 10.1	_
Oscillatoria major	_	_	45 ± 19.9	60.3 ± 8.1	_	_	_
Spirulina major	_	_	_	_	7.3 ± 3.8	_	_
Chlorophyceae							
Ankistrodesmus falcatus	_	_	_	24.67 ± 4.2	_	_	_
Bulbochaete satigera	_	_	_	_	_	_	17.3 ± 0.6
Chara braunii	_	_	_	_	_	5.3 ± 3.5	3.7 ± 3.1
Chlorella vulgaris	_	37 ± 6	_	_	_	_	_
Cladophora glomerata	56.3 ± 11.1	_	_	28 ± 4.4	_	_	_
Closterium parvulum	_	5.7 ± 2.31	_	_	_	_	5 ± 5.3
Closterium ehrenbergii	_	_	3.3 ± 2.1	_	_	_	_
Coelastrum microporum	43 ± 5.2	_	_	_	22 ± 2.7	_	_
Coleochaete islamii	_	_	_	_	3 ± 2.7	_	3.7 ± 2.5
Desmidium baileii	_	_	24.7 ± 2.9	_	_	_	_
Dimorphococcus lunatus	_	14.3 ± 3.1	_	_	_	_	_
Gonium pectorale	_	_	_	27 ± 6.6	_	_	_
Hyalotheca dissiliens	_	_	15 ± 4.6	_	_	_	_
Mougeotia punctata	_	18.3 ± 7.5	_	23 ± 3.6	_	_	_
Nitella mucronata	_	4 ± 2	_	_	_	_	4 ± 3
Oedogonium fragile	_	_	6.3 ± 1.5	_	_	16.3 ± 3.1	16.7 ± 5.1
Oedogonium aquaticum	_	_	_	_	_	_	16 ± 5.6
Oocystis elliptica	_	_	_	5 ± 2.7	_	_	_
Oocystis pusilla	5 ± 3.6	_	_	_	14 ± 3.5	_	_
Pediastrum duplex	_	_	_	_	33.7 ± 5.7	_	_
Pediastrum tetras	_	_	_	_	18.3 ± 9.7	_	_
Pleurotaenium trabecula	_	4.7 ± 3.1	_	_	4±3.6	_	_
Rhizoclonium lapponicum	_	_	_	19.3 ± 1.5	13.7 ± 8.0	33.7 ± 8.1	_
Scenedesmus bijugatus	_	15 ± 3.6	_	-	-	_	_
Scenedesmus obliquus	_	15.3 ± 4	_	_	18.7 ± 4.6	_	7.7 ± 0.6
Spirogyra gracilis	44 ± 6.2	_	23.7 ± 2.3	24.3 ± 8	21.7 ± 5.5	26 ± 3.6	_
Spirogyra weberi	_	_	13.7 ± 4.2	22.3 ± 0.6	_	_	_
Spirogyra maxima	_	_	26.7 ± 10.1	_	_	_	19 ± 5.3
Tetraedron trigonum	_	_	_	3 ± 1	2.3 ± 2.3	_	_
Bacillariophyceae				<u> </u>	2.0 <u>2</u> 2.0		
Cymbella acuta	_	_	11.7 ± 4.2	_	_	_	14.7 ± 8.3
Eunotia pectinalis	_	2.7 ± 1.5	_	_	_	_	-
Nitzschia amphibia	_	_	_	3.3 ± 1.5	_	9.7 ± 3.2	2.7 ± 3.1
Nitzschia sigma	_	5.3 ± 2.3	-2.67 ± 1.5	- -	_	3.6 ± 3.1	, <u>_</u>
Synedra acus	6.3 ± 2.3	- 2.3	5 ± 2.7	- 6±1	-3.33 ± 1.2	- -	_
Euglenophyceae	0.5 <u>.</u> 2.5		J <u>. 2</u> .1	<u></u>	5.55 <u>+</u> 1.2		
Euglena acus	_	_	_	2.7 ± 1.6	_	_	_
Phacus acuminatus				6.3 ± 4			



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Appendix 1

See Table 7.

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