

# Unraveling the AM fungal community for understanding its ecosystem resilience to changed climate in agroecosystems

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### Abstract

The changing global climate affects the agroecosystem making it challenging to achieve the world's sustainable development goals. Among the facets of belowground microbial communities, the arbuscular mycorrhizal fungi (AMF) hold an important place. They represent the most common symbiont phylum colonizing more than 80% of the plant families and are likely to be affected by global climate change. These fungi facilitate plant's mineral acquisition, improving growth and protecting them from biotic and abiotic stresses. The elevated carbon dioxide  $(eCO<sub>2</sub>)$  level, temperature, increased nitrogen and phosphorus deposition influences the plant phenology and AMF functioning through changes in diversity and community composition of AMF. The interaction effects of soil management practices due to climate change affect the system productivity and perturb mineral cycling. Understanding the carbon and nitrogen cycling of an agro-ecosystem and its associated AMF communities concerning ecosystem productivity is the need of the hour. Plant-fungal associations require a more environment resilient approach to ameliorate the effect of anthropogenic changes in carbon and nitrogen cycles. Since AMF communities alter due to local environmental conditions and land-use changes, the most adapted community may help in predicting the mycorrhizal responses to chemical fertilizers,  $eCO<sub>2</sub>$ , temperature and drought. In this review, we aimed at investigating (i) the diversity and community composition of AMF in relation to the change in crop and soil management practices, and (ii) how the adapted AMF communities may perform in maintaining the ecosystem resilience of these agroecosystems under climate change conditions. Hence, AMF-plant symbiosis can be effectively integrated into global climate change models. Eventually, the ecosystem resilience will be better understood to exploit the resident AMF communities to offset some of the detrimental effects of anthropogenic environmental change.

Keywords AMF community  $\cdot$  climate change  $\cdot$  global warming  $\cdot$  Glomalin

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# 1 Introduction

Global climate change caused by anthropogenic activities such as the intensification of agriculture, fossil fuel combustion and excess use of chemical fertilizers has increased the emissions of nitrogenous gases such as nitrous oxide, oxides of nitrogen, ammonia and,  $CO<sub>2</sub>$ . Due to change in precipitation regimes, for instance the current level of  $CO<sub>2</sub>$  has increased from 280 ppm of preindustrial regimes to 400 ppm, which has contributed to global warming effects (Betts et al. [2016;](#page-10-0) Behera et al. [2013;](#page-10-0) IPCC [2013;](#page-11-0) Weltzin et al. [2003](#page-14-0)). In general, global climate change is known to affect terrestrial plants, microorganisms, and change the composition of soil communities which are involved in the functioning of various soil ecosystems (Pugnaire et al. [2019](#page-13-0)) As per the IPCC report ([2014](#page-11-0)), the expected global climate change includes an increase in temperature and changes in rainfall regimes which shift the range of species, altering plant's distribution and associated rhizospheric microbial communities. Ultimately, the shifting of adapted associations under global climate change will modify the outcome of plant-soil microbe interactions.

Arbuscular mycorrhizal fungi (AMF), are the most common association forming a symbiotic relationship with roots of more than 80% terrestrial plant species (Classen et al. [2015](#page-10-0); Smith and Read [2008\)](#page-13-0). AMF are obligate symbionts providing nutrients to the plants in exchange for carbon (C) and are mostly affected due to global climate change. These fungi are functionally important and offer a wide range of benefits to their hosts including enhanced nutrient uptake (Bagyaraj et al. [2015;](#page-10-0) Smith et al. [2011](#page-13-0)), pathogen resistance (Cameron et al. [2013\)](#page-10-0), heavy metal tolerance (Hildebrandt et al. [2007](#page-11-0)), soil C sequestration (Rillig et al. [2004](#page-13-0); Wang et al. [2017;](#page-14-0) Parihar et al. [2020](#page-12-0); Solaiman [2014\)](#page-14-0), improved water relations (Auge, [2001](#page-9-0)) and drought alleviation (Mathimaran et al. [2017\)](#page-12-0) while being influence and affecting other soil biotas (Drigo et al. [2010](#page-11-0); Nuccio et al. [2013;](#page-12-0) Gupta et al. [2018](#page-11-0)). Therefore, the research on AMF in relation to environmental changes is imperative, because AMF may help to either mitigate or intensify climate change under future ecological conditions.

AMF distribution is primarily influenced by abiotic factors (drought, rainfall, flood,  $eCO<sub>2</sub>$  and temperature), and the biotic environment (Rasmussen et al. [2018;](#page-13-0) Vályi et al. [2016](#page-14-0)). AMF play an essential role in ecosystem resilience by facilitating species-species interactions between the nurse and facilitated plants (Song et al. [2010;](#page-14-0) Cheng et al. [2012](#page-10-0); Montesinos-Navarro et al. [2019\)](#page-12-0) and nutrient changes in soils (van der Heijden, [2008\)](#page-14-0). Global climate change has direct and indirect effects on AMF communities due to the shift in plant phenology (Augé et al., [2001;](#page-9-0) Rillig et al. [2002a;](#page-13-0) Singh et al. [2010](#page-13-0)). As per earlier reports, higher temperature and low moisture levels alter the root-associated microbial community composition, which was found to increase plant growth, and influence AMF colonization (Augé [2001;](#page-9-0) Gavito et al. [2000,](#page-11-0) [2002\)](#page-11-0). Seasonal fluctuations in rainfall patterns (Hazard et al. [2013;](#page-11-0) Alguacil et al. [2015](#page-9-0)) and temperature (Dumbrell et al. [2011\)](#page-11-0) also influence the AMF development in the soil. AMF diversity and colonization pattern were greatly affected by phenological soybean growth stages, and the rhizosphere was found to be mainly dominated by *Glomus* followed by Acaulospora and Gigaspora species (Buade et al. [2020](#page-10-0)). In addition, the levels of available nitrogen (N; Velázquez et al. [2018\)](#page-14-0), phosphorus (P; Zhao et al. [2017](#page-15-0)), soil pH (Melo et al. [2017\)](#page-12-0), soil moisture (Deepika and Kothamasi [2015](#page-10-0)) and organic matter (Wang et al. [2015\)](#page-14-0) may also influence the AMF community. Although many reports are available on mycorrhizal responses to global climate change (Begum et al. [2020](#page-10-0); Saleh et al. [2020;](#page-13-0) Del Buono [2021](#page-10-0); Wang et al. [2021](#page-14-0)), the information on the anthropogenic component of global climate change concerning changed crop and soil management practices on the diversity and community composition of AMF is scanty. The diversity and species composition of plant communities can also influence the abundance, diversity and species composition of AMF communities (Oehl et al. [2003;](#page-12-0) Landis et al.  $2004$ ). The higher temperature and  $CO<sub>2</sub>$ , droughts, floods, nutrient level of N and P shift the distribution of AMF community, cooperate with the newly evolved species, and established synergistic interaction for sustaining the agroecosystems. Therefore, AMF are an essential component for predicting the terrestrial ecosystems functioning in relation to climate change (Drigo et al. [2008](#page-11-0)). AMF are ecologically important and may have an enormous impact on ecosystem responses (Fig. [1](#page-2-0)). They directly influence the individual plants and indirectly influence the plant processes such as plant dispersal and community interactions (Johnson et al. [2013a](#page-12-0); Treseder, [2016](#page-14-0)). The selection of appropriate local native AMF and their production in sufficient quantity and improved quality are critical issues for their application in agriculture, particularly under global climate change (Berruti et al. [2016](#page-10-0)). Native AMF strains appear to be physiologically, ecologically and genetically more adaptable to the changing environment and could be more stable to their host plants (de Oliveira et al. [2017](#page-10-0); Hart et al. [2018](#page-11-0)). However, previous research suggested AMF communities were not resilient (Gavito et al. [2008](#page-11-0)), but later on it was validated that AMF communities of tropical dry forest ecosystem are resilient and prove to land-use change and management practices (Carrillo and Gavito [2018](#page-10-0)). Hence, it becomes pertinent to study the AMF community composition and identify the well-adapted resilient AMF communities to the changed climatic conditions for sustaining the productivity under different agroecosystems (Table [1\)](#page-3-0).

The present review provides a comprehensive up-to-date knowledge on the AMF communities against global climate change underlying drivers especially, rising  $CO<sub>2</sub>$ , higher temperature, N deposition, drought and precipitation. Further, deciphering the resilient and dominant AMF communities may determine the plant community structure under global climate change scenarios for mitigating the adverse effects of global climate change.

# 2 Global climate change and global warming are the key drivers of AMF community and resilience to its functioning

Since, global climate change affects AMF, the functional outcome of any change in AMF communities also influences the mechanisms through which global climate change operates. Typically,  $eCO<sub>2</sub>$  enrichment generally increases the biomass of roots and mycorrhizal fungi (Rillig et al. [2002a](#page-13-0); Drigo et al.

<span id="page-2-0"></span>

Fig. 1 The effect of elevated CO<sub>2</sub>, higher temperature, drought warming on plants –microbe interaction, AMF and extraradical mycelium (modified from Compant et al. [2010](#page-10-0))

[2008\)](#page-11-0). It is also likely that different AMF species may have a magnitude of variable plant responses (Hart et al. [2018;](#page-11-0) Gupta et al. [2019\)](#page-11-0). Therefore, most efficacious strains need to be selected under  $eCO<sub>2</sub>$  and temperature conditions. The higher rainfall alters the AMF community composition directly or indirectly due to change in plant traits related to plant nutrition and water uptake (Deveautour et al. [2018](#page-10-0)). Changes in the AMF community have been observed in different  $C_3$  and  $C_4$ genotypes as  $C_4$  plants, which have higher assimilation rates and use more  $CO<sub>2</sub>$  due to kranz anatomy. However, climate change, soil warming or drought stress indirectly affect plants associated with AMF (Auge et al. [2015\)](#page-10-0). The plants under  $eCO<sub>2</sub>$  decrease their allocation of N-rich metabolites and increase the C rich metabolites allocation to root exudates (Tarnawski and Aragno, [2006](#page-14-0)). AMF may also be indirectly influenced by global climate change (temperature, rainfall) as it resulted in altered soil properties such as pH and nutrient availability and associated physico-biochemical properties which are the key drivers affecting the AMF communities (Fig. 1). The  $eCO<sub>2</sub>$  level in the atmosphere influences AMF symbiosis and is crucial for predicting the soil ecosystem changes. Over the past two decades, various studies have been conducted to examine the effect of  $eCO<sub>2</sub>$  [Evidence from open top chamber (OTC) and free air  $CO<sub>2</sub>$  enrichment (FACE) experimental studies] on changes in AMF community composition and diversity in a range of systems/practices under different ecosystems (Grover et al. [2015](#page-11-0)). It is reported that eCO2 brought changes in ecosystem productivity, plant community structure and species composition (Bazzaz [1990](#page-10-0)).

Such changes in the plant community are associated with the AMF community changes (van der Heijden et al. [1998](#page-14-0)). However, the results are contrasting and variable due to a change in AMF strains and their responses are variable due to the altered conditions (Gavito et al. [2000\)](#page-11-0). It has been shown that plants allocate more C from their photosystem to the growing hyphae and roots under  $eCO<sub>2</sub>$ , which resulted in increased AMF colonization in comparison with ambient  $CO<sub>2</sub>$ . Moreover,  $eCO<sub>2</sub>$  concentrations help in organic matter degradation, thus generating net C sources rather than Csinks, which indirectly help mycorrhiza growth (Cheng et al. [2012;](#page-10-0) Phillips et al. [2012;](#page-13-0) Parihar et al. [2020\)](#page-12-0). The potential for mitigating rising  $CO<sub>2</sub>$  levels through increased allocation and storage in mycorrhizas and other belowground C pools is currently debated (Phillips et al. [2012;](#page-13-0) Verbruggen et al. [2013\)](#page-14-0). However, many studies have been carried out where AMF has been shown to enhance C sink in terms of producing higher amounts of recalcitrant copious sticky glycoproteins called "glomalin" which remains as part of stable C pool (Rillig et al. [2001](#page-13-0), [2004](#page-13-0); Wang et al. [2017;](#page-14-0) Parihar et al. [2020\)](#page-12-0). AMF influence agroecosystem directly through mycelium effects by modulating host physiology and resource capture, and indirectly through changes in plant and soil microbial community composition. In addition, the direct effects of the AMF on agroecosystem via mycelium and its products e.g., glomalin becomes significantly important in performing ecosystem services (Rillig [2004a](#page-13-0), [b](#page-13-0)) The atmospheric elevated partial pressure of  $CO<sub>2</sub>$  (pCO<sub>2</sub>), a higher level of C brought out to adaptive AMF strains, which shift AMF community



<span id="page-3-0"></span>Table 1 Recent reports on the impact of agricultural management practices on AMF abundance and community composition under different agroecosystems

<sup>a</sup> Genera and families as presented in Wijayawardene et al. [\(2020\)](#page-14-0)

composition towards higher  $pCO<sub>2</sub>$ . Under such conditions, native strains play a crucial role in increasing the N concentration in hosts and maintaining the ecosystem resilience (Jarrell and Beverly  $1981$ ; Gamper et al.  $2005$ ) e.g., eCO<sub>2</sub> brought changes in the AMF community composition by an increased ratio of Glomeraceae to Gigasporaceae members (Cotton et al. [2015](#page-10-0)). The members of Glomeraceae dominate in the composition of the AMF communities in vineyards of Oregon State (Schreiner and Mihara [2009\)](#page-13-0), Piedmont (Italy; Balestrini et al. [2010](#page-10-0)) and Burgundy (France; Bouffaud et al.  $2016$ ). Primarily, it has been assumed that  $eCO<sub>2</sub>$  increases the C supply to some specific AMF taxa, which resulted in increased abundance along with reduced community evenness, and richness, where intraspecific and interspecific variations were also reported among isolated such as Glomus claroideum and G. intraradices (Cotton et al. [2015](#page-10-0)). Contrarily, an increase in the AMF richness arising from C-limitation has also been suggested. Surprisingly, the bulk of studies detected no significant effect of  $eCO<sub>2</sub>$  on AMF richness (Antoninka et al. [2011;](#page-9-0) Cotton et al. [2015;](#page-10-0) Mueller and Bohannan [2015](#page-12-0)). Mostly, the impact of  $eCO<sub>2</sub>$  on AMF is observed mainly on evenness and not on the AMF community diversity.

### 2.1 Impact of climate change and global warming on AMF under FACE and OTC conditions

Predicting the productivity potential of agroecosystem's at  $eCO<sub>2</sub>$  levels is necessary to understand the interactive effects of  $eCO<sub>2</sub>$  with AMF partners and plants uptake of nutrients and growth (Fig. [1\)](#page-2-0). Jakobsen et al. [\(2016](#page-11-0)) worked out the effects of  $eCO<sub>2</sub>$  up to 900 ppm under FACE on the growth of two host plants e.g., Medicago truncatula and Brachypodium distachyon differing in P utilization efficiency on AMF symbiosis. They found that the application of AMF at ambient and  $eCO<sub>2</sub>$  increased mycorrhizal formation and the growth of Medicago truncatula plants at low P conditions Also reported AMF development and function is likely more influenced by the temperature component of climate change than by its  $(CO<sub>2</sub>)$ ) component. The  $eCO<sub>2</sub>$  had small effects on P acquisition. However, it enhanced the conversion of tissue P into biomass, which indicates no increase of P fertilizers for sustaining the growth of these plants. Similarly, Gamper et al. ([2004\)](#page-11-0) assessed the impact of  $eCO<sub>2</sub>$  with the partial pressure of  $CO<sub>2</sub>$ and N under FACE in a seven-year well-fertilized grass and legume monocultures for AMF symbiosis and nutrient availability. They showed that  $eCO<sub>2</sub>$  at a low level of N fertilization marginally increased the AMF root colonization but significantly augmented the production of smaller spores in both the hosts in monocultures. However, larger AMF spores were different in both the hosts. The higher N fertilization decreased the AMF formation in Lolium perenne but not in Trifolium repens. A ten years experiment of the interaction effect of  $eCO<sub>2</sub> \times N$  reported a significant effect of both on AMF (Staddon et al. [2004\)](#page-14-0). However, the  $eCO<sub>2</sub>$  and limited N decreased root colonization but increased extraradical hyphae. Understanding of C fluxes in  $C_3$  plants to AMF and subsequent transfer to other rhizosphere microbial communities and in C cycling at  $eCO<sub>2</sub>$  are scarce. Drigo et al. ([2010](#page-11-0)) proposed a conceptual model based on a stable isotope tracer study. They observed that under  $eCO<sub>2</sub>$  (up to 700 ppm) plant-assimilated C is rapidly transferred to AMF, releasing slowly to the resident niche bacterial and fungal rhizosphere populations. This study helped to understand the C-flow paths in soils and predicts the sustainability of terrestrial ecosystems at  $eCO<sub>2</sub>$ . The  $eCO<sub>2</sub>$  influences on the AMF community dynamics were further assessed in soybean free air concentration enrichment monitored for three growing seasons over five years. It was observed that  $eCO<sub>2</sub>$  consistently favored the dominance of Rhizophagus irregularis (Rhizoglomus irregulare) and not the Gigasporaceae (Cotton et al. [2015](#page-10-0)).

Other studies conducted under OTC at  $eCO<sub>2</sub>$  and temperature were found to positively influence diversity and community composition of AMF, where Glomus spores were found to be most abundant. However, there was a clear difference in the AMF species richness observed at the altered conditions (Asha et al.  $2017$ ). When the ambient  $CO<sub>2</sub>$  level was increased to 550ppm, a concomitant increase of Glomus species (up to 93% of AMF taxa) over the ambient  $CO<sub>2</sub>$  (up to 62%) was detected (Klironomos et al. [2005](#page-12-0)). The relative abundance of sequences putatively identified as Rhizoglomus irregulare was also shown to increase from an average of 53% under ambient  $CO<sub>2</sub>$  to 73% under  $eCO<sub>2</sub>$  (550ppm; Cotton et al.  $2015$ ) and an increase in the  $CO<sub>2</sub>$  levels to 560ppm lead to a fourfold increase in the spores of Glomus clarum (Wolf et al. [2003\)](#page-14-0).

# 2.2 Impact of temperature on diversity and community composition of AMF

Increased temperatures may generally promote the plant growth by enhancing the photosynthesis, which indirectly supports the development of AMF through the nutrient exchange (Mathur et al. [2018](#page-12-0)). The selection of ecosystem resilient AMF strains under the changing global climate scenario for promoting the growth is the need of an hour (Fig. [1](#page-2-0)). It is known that AMF form colonization with most plants and respond to plant growth at higher temperatures (Graham et al. [1982;](#page-11-0) Fitter et al. [2000](#page-11-0)). AMF ameliorate the effects of temperature stress due to global climate change on plants by increasing growth and nutrient content (Pischl and Barber [2016\)](#page-13-0). However, these effects do not extend and confer tolerance to plants against herbivorous organisms. The temperature may also significantly alter the structure of the AMF hyphal network, where low temperature (cooled soil) allocate more vesicles (storage) and high temperature (warmed soil)

contributes to more extensive extraradical hyphal networks (growth) (Hawkes et al. [2008\)](#page-11-0). The detection of a direct effect of temperature on the extraradical mycelium was investigated in a compartment study to assess the differences in nutrient uptake through AMF species (Jakobsen et al. [1992\)](#page-11-0). This study provided a better understanding of rhizospheric C-allocation and the increased respiration of the extraradical hyphae when the soil temperature is high (Heinemeyer et al. [2006\)](#page-11-0) in conferring the tolerance to plants. The temperature responses of AMF have not been investigated intensively along with  $eCO<sub>2</sub>$ . The effect of temperature such as extremely low or high temperatures in a given region can considerably reduce the AMF population because no adaptation found in these AMF isolates (Funneliformis mosseae) in the soils (Addy et al. [1994;](#page-9-0)Gavito and Azcón-Aguilar, [2012](#page-11-0)).Therefore, AMF adapts and confer resilience to themselves against the changed climatic conditions (Al-Karaki et al. [2004\)](#page-9-0). Region-specific AMF strains and their communities need to be identified for their exploitation in sustaining the agroecosystems.

# 2.3 Impact of nitrogen and phosphorus fertilizers on diversity and community composition of AMF

The continuous application of N and P fertilizers created environmental problems such as reduced biodiversity and suppressed ecosystem functionalities (Socolow [1999\)](#page-13-0). To overcome the ecological issues, beneficial microorganisms in increasing the growth, nutrient availability and sustaining the agroecosystem's productivity are being promoted. In addition, the response of plants to  $eCO<sub>2</sub>$ , the availability of both N (Daepp et al. [2001](#page-10-0)) and P (Campbell and Sage [2002\)](#page-10-0) need to be assessed to understand the contribution of AMF in acquiring these nutrients. The exploitation of native AMF forming association with most terrestrial plants is gaining importance in order to provide nutrients and protection even under environmental stress. The effects of N enrichment on AMF are mediated by soil P availability, where N enrichment of P-limited soils was found to increase the biomass of AMF. In contrast, N enrichment of P-rich soils declined the AMF biomass (Johnson et al. [2003\)](#page-12-0). Resource availability affects the plant community composition and, through interactive effects of N,  $CO<sub>2</sub>$ , and plant diversity, it governs the AMF communities (LeBauer and Treseder [2008\)](#page-12-0). Higher N concentrations increase plant productivity, but decrease the C availability to AMF (Johnson et al. [2003](#page-12-0)). The impacts of N and rainfall on AMF communities at  $eCO<sub>2</sub>$ were assessed in a seven-year field study. It was observed that N application reduced AMF, whereas rainfall increased AMF abundance. The interaction of both N and precipitation mainly altered the AMF community structure than AMF phylogenetic structure (Chen et al. [2017](#page-10-0)). Cavagnaro et al. ([2007](#page-10-0)) reported that inoculation of AMF in tomato plants grown at  $eCO<sub>2</sub>$  showed higher contents of N, P and Zn, when compared to plants grown at under ambient  $CO<sub>2</sub>$ . However, the response of AMF on N uptake was found to vary with host plants and AMF species used (Chen et al. [\(2007\)](#page-10-0). For example, AMF colonization enhanced N uptake under  $eCO<sub>2</sub>$  in *Plantago lanceolata*, but not in Festuca arundinacea plants. The P use efficiency in plants is being modulated through P transformation at  $eCO<sub>2</sub>$  condition. Ainsworth et al. [\(2003\)](#page-9-0) showed a higher photosynthetic rate and plant growth at  $eCO<sub>2</sub>$ , mediated and metabolized through concentration of the Rubisco enzyme to facilitate C assimilation by autotrophic organisms. It is known that  $eCO<sub>2</sub>$  increases the Rubisco concentration to metabolize more inorganic P to transform into organic P. Organic P is needed to synthesize Rubisco which is a major constituent of rRNA involved in the synthesis of Rubisco enzyme (Veneklaas et al.  $2012$ ). The eCO<sub>2</sub> increases the grain yield due to an increased P assimilation in the grain (Buddrick et al.  $2014$ ). Hence, it is likely that  $eCO<sub>2</sub>$  levels increase the Puse efficiency by accumulating a higher proportion of P in plant tissue to further help in the photosynthesisassociated metabolisms and assimilation activities. The AMF diversity and community vary with the type of fertilization. The application of organic manure in maize is regarded as the most critical factor influencing AMF community composition, followed by N and P fertilizer inputs. In another recent study on coffee agroecosystem concentrating on identifying the AMF communities in conventional and organic systems, AMF were affected due to interactive effects of N fertilization with light shade (Aldrich-Wolfe et al. [2020\)](#page-9-0). The higher correlation of AMF community composition in organic manure applied maize rhizosphere activates more AMF species to interact with other species thereby regulating the AMF symbiotic system in maintaining the soil health and productivity of crops (Zhu et al. [2016](#page-15-0)). There are not many studies available concerning the diversity and community composition of AMF in relation to N and P fertilization at  $eCO<sub>2</sub>$ . Therefore, further work is needed in this area to harness the potential of stabilized native AMF communities in rationalizing the fertilizer uses without compromising crop productivity.

### 2.4 Impact of precipitation/rainfall on diversity and community composition of AMF

The mycorrhizal soils had significantly more water stable aggregates and substantially higher extraradical hyphal densities than the non-mycorrhizal soils. However, comparative water relations of both mycorrhizal and non-mycorrhizal soils have not extensively studied (Auge [2001\)](#page-9-0). Variations in water regimes/precipitation influence the soil microbial community (Clark et al. [2009\)](#page-10-0). Changes in rainfall distribution pattern can directly influence the plant communities, diversity and productivity, which alters AMF abundance,

functioning and communities at regional scale (Zhang et al. [2016](#page-14-0); Johnson et al. [2004](#page-11-0)). Precipitation alters the root architecture which changes the soil environment and shifts in AMF community associated with changes in root traits (Deveautour et al. [2018\)](#page-10-0). Zhang et al. ([2016](#page-14-0)) examined the direct effect of a gradient of rainfall (50 to 400 mm) on AMF communities associated with dominant plant species in a Tibetan alpine steppe. Among the 31 AMF operational taxonomic units identified, AMF community composition varied significantly among sites but did not differ among plant species. Further, they showed that precipitation directly affected the AMF hyphal length density, and indirectly influenced AMF species richness suggesting the role of water precipitation in driving the AMF communities at the regional scale. The magnitude of AMF responses to confer tolerance in plants against the water stresses differs with the AMF species-plant combinations (Zhang et al. [2016\)](#page-14-0). Under water stress, plants acclimatize native AMF species to survive and proliferate extensive hyphae to acquire more water. It has been shown that during water stress some AMF species may become inefficient in forming colonization in plants (Deepika and Kothamasi [2015](#page-10-0)). However, under congenial conditions, efficient AMF, which require moreC fluxes, may lose their abilities due to insufficient C supply from the host plants (Johnson, [2010](#page-12-0)). The impacts of drought on AMF abundance and diversity have been reported in a number of studies (McHugh and Schwartz et al. [2015](#page-12-0); Martínez-García et al. [2012\)](#page-12-0) On the other hand, higher rainfall conditions also altered the AMF community composition in several studies (Sun et al. [2013;](#page-14-0) Li et al. [2015;](#page-12-0) Gao et al. [2016](#page-11-0); Deveautour et al. [2018](#page-10-0)). Deepika and Kothamasi ([2015](#page-10-0)) studied that under flood conditions, the AMF community composition with Sorghum vulgare resulted in lowest diversity mainly dominated by two Acaulospora phylotypes. An increase of rainfall and relative humidity increased the AMF colonization (Pande and Tarafdar [2004](#page-12-0)). Changes in AMF formation and an increase in the abundance of one species were observed under increased temperature and precipitation (Sun et al. [2013\)](#page-14-0), and phylogenetic clustering of communities have also been reported (Chen et al. [2017](#page-10-0)). It was concluded that the plant community influenced the AMF and was closely related with the composition of the AMF community. Rainfall has been observed as a significant driver for AMF spore density, especially of Acaulosporaceae, Diversisporaceae and Glomeraceae, which were positively correlated with abiotic factors. In contrast, members of the Gigasporaceae showed a negative correlation with these climatic variables (Tchabi et al. [2008,](#page-14-0) Al-Yahya'ei et al. [2011](#page-9-0)). More studies related to AMF abundance, community composition and associated ecological functions are needed under global climate, change and models covering all the factors of production functions such as rainfall, temperature, seasons etc (Cotton et al. [2018](#page-10-0)).

# 2.5 Impact of plant genotypes (C3 and C4 plants) on diversity and community composition of AMF

Plant species are critical in regulating the AMF symbiotic efficacy in  $C_3$  and  $C_4$  plants. Due to climate change, plants are likely to induce changes in physiology and root exudation. Compared to some commonly grown  $C_3$  plants, greater mycorrhizal symbiotic efficacy has been reported in  $C_4$  plants (Hetrick [1991](#page-11-0); Wilson and Harnett [1997](#page-14-0)). In the case of  $C_4$ plants, grasses are categorized as obligate AMF hosts (Hetrick [1991\)](#page-11-0).  $C_4$  plants often benefit more from AMF associations than  $C_3$  plants, partly due to their higher photosynthetic efficiency and greater nutrient demands (Hoeksema et al. [2010\)](#page-11-0). Increasing  $CO<sub>2</sub>$  is generally known to enhance plant C fixation, particularly in  $C_3$  plants, and also increase the proportion of fixed C allocated to plant roots (Ainsworth and Long [2005;](#page-9-0) Drigo et al.  $2010$ ). Since C<sub>4</sub> plants have kranz type anatomy that can facilitate two-fold  $CO<sub>2</sub>$  fixation hence are more receptive to AMF under  $eCO<sub>2</sub>$  and temperature. The AMF mediated drought resistance varies among  $C_3$  and  $C_4$  plant functional groups (Yamori et al. [2014\)](#page-14-0). Single or mixed AMF groups differed in their response, where  $C_3$  plants were positively influenced by Rhizoglomus irregulare and compared to other species, C<sub>4</sub> plants benefitted from *Funneliformis mosseae*. Many studies have observed variable responses of AMF to drought conditions, where  $C_3$  species showed higher water use efficiency than  $C_4$  species (Edwards et al. [2010;](#page-11-0) Worchel et al. [2013](#page-14-0); Augé et al. [2015\)](#page-10-0). The mycorrhizal symbiotic efficiency in  $C_3$  and  $C_4$  plants vary depending on the plant species involved. Among C<sub>4</sub> plants, grasses are highly mycorrhizal and exhibited higher mycorrhizal dependency (98 to 99%) than  $C_3$  plants (Hetrick [1991\)](#page-11-0). However AMF colonization seems often to be a rather poor indicator for plant growth promotion (Sharma et al. [1996\)](#page-13-0).

# 3 Significance of AMF resilience and its functioning in relation global climate change

Global climate change alters the functioning of individual AMF and well-adapted communities. Climate-changing parameters such as  $eCO<sub>2</sub>$ , temperature, precipitation etc., are known to affect terrestrial plants associated microbial communities including AMF. Previous studies on global climate change with AMF species and their functioning advocates that plant species and abiotic factors are jointly involved in driving the AMF community required to perform various soil eco-functions (Gamper et al. [2005;](#page-11-0) Klironomos et al. [2005\)](#page-12-0). Together, these studies suggest that  $eCO<sub>2</sub>$  concentrations may make AMF communities more effective in some ways, but less effective in another way (Table [1](#page-3-0)).

#### 3.1 Alleviates drought stress

There are many reports available, where the role of AMF symbiosis in the alleviating drought stress have been spelled out (Mathimaran et al. [2017;](#page-12-0) Auge [2001](#page-9-0); Ruiz-Lozano [2003](#page-13-0); Wu et al. [2013](#page-14-0); Sharma et al. [2020](#page-13-0)). AMF confers drought tolerance to plants through various mechanisms such as increased extended hyphal network to absorb more water (Mathimaran et al. [2017\)](#page-12-0), improved plant root architecture and soil properties (Wu et al. [2012](#page-14-0)), regulating the physiology of plants to perform functions (Yang et al. [2014](#page-14-0)), regulating plant gene expressions such as aquaporins (Li et al. [2013](#page-12-0); Barzana et al. [2014](#page-10-0)), and osmotic adjustments in plants such enhanced levels of proline, reactive oxygen species (ROS), trehalose sugars (Kaya et al. [2009;](#page-12-0) Ocon et al. [2007](#page-12-0); Sharma et al. [2020\)](#page-13-0). Under drought conditions the interactive effects of plant genotype and AMF richness offset the damage caused due to drought and led to high barley crop yield (Sendek et al. [2019\)](#page-13-0). Recently, Chareesri et al. [\(2020](#page-10-0)) showed that under drought, besides increasing the photosynthetic parameters e.g., higher levels of chlorophyll fluorescence in AMF inoculated rice plants were revealed. AMF also increased the levels of abscisic acid (ABA) and indole acetic acid (IAA) in plants under drought. The authors argued that higher response in AMF inoculated rice plants under drought might have been mediated through nutrients and also through the regulation of plant hormones, especially IAA and contributed to a better recovery and resulted in high rice grain yields. This becomes highly relevant under the changing climate, where drought is the critical factor limiting the productivity of the majority of crops. Hence, establishing colonization with resilient AMF species could be essential to regulate and stimulate various mechanisms conferring the drought tolerance in crop plants.

# 3.2 Stimulation of other plant growth promoting microorganisms (PGPM) and soil biota

Global climate change is altering the plant's behavior and belowground AMF species distributions and interactions with other microbial communities. These beneficial rhizosphere microbes improve crop's nutrient status as described above, facilitate and mobilize soil resources and processes required for sustaining the growth of plants especially under change climate. It is known that environment change through  $eCO<sub>2</sub>$ , temperature and drought, alter the root exudates composition and induces alterations in the activity, function, abundance and structure of the soil microbial community associated with plants or plant metabolites (Rangel-Castro et al. [2005](#page-13-0); Rasche et al. [2009](#page-13-0)). Shifting of resident AMF species under elevated atmospheric  $CO<sub>2</sub>$  conditions are linked with changes in the active bacterial and fungal rhizosphere communities.

Alterations in root exudates include changes in the availability of chemo-attracting or signal compounds as well as a

different C/N ratio or nutrient availability (Bansal and Mukerji [1994;](#page-10-0) Kandeler et al. [2006](#page-12-0)). The extent of AMF-derived benefits depends on the compatibility of the soil environments including microbes from where the communities were selected and identified. Numerous studies have been conducted on the combined inoculation of AMF with other plant growth promoting bacteria (PGPB) such as Rhizobium meliloti, and resulted in higher biomass and grain yield under drought conditions (Panwar [1993;](#page-12-0) Zoppellari et al. [2014](#page-15-0) Valdenegro et al. [2001,](#page-14-0) Mortimer et al. [2013](#page-12-0); Sharma et al. [2016\)](#page-13-0). In a recent review, Sharma et al. [\(2020\)](#page-13-0) highlighted the role of the tripartite symbiosis of legumes with AMF-rhizobium in conferring the drought tolerance through the trehalose-based mechanism. The community changes at the  $eCO<sub>2</sub>$  may substantially affect the diversity and activities of rhizospheric microbes. Therefore, a stabilized resilient combination of AMF (Glomus clarum) with PGPB Bacillus sp. strains may improve the drought, salt tolerance in plants by increasing chlorophyll, proline and soil aggregates (Kohler et al. [2009](#page-12-0), Grover et al. [2015;](#page-11-0) Diagne et al. [2020\)](#page-10-0). However, studies related to combination of AMF with PGPB particularly at  $eCO<sub>2</sub>$  are lacking. Although AMF and other PGPB including endophytes are being used as biofertilizers, phytostimulants, bioprotectants in agriculture (Welbaum et al. [2004;](#page-14-0) Lugtenberg and Kamilova [2009](#page-12-0)), the knowledge on how exactly plantassociated microbial communities are being influenced either directly under environmental change or indirectly through changed plant physiology are lacking (Drigo et al. [2008\)](#page-11-0). Therefore, a better understanding of the selection and performance of appropriate ecosystem resilient soil microbiome is needed to mitigate the impacts of global climate change on plant productivity.

# 3.3 Changes in mycorrhizal dependence and nutrient availability

In general, much work has been carried out in different agroecosystems on how mycorrhizal species are changing their nutritional contribution in plants and gaining significance as potential biofertilizers. However, the information regarding the contribution of AMF to the plant's nutritional requirement under changed climatic conditions is inadequate. AM colonization stimulates plant nutrient acquisition. AMF infect and form a symbiosis with the roots of plants and improves plant growth by supplying N, P, K, Ca and micronutrients including zinc (Zn) and S, as a strategy to obtain essential nutrients from plants (Gupta and Abbott [2020\)](#page-11-0). Thus, AMF provide nutritional support to the plants even under unfitting conditions inside the root cells. AMF produce fungal structures such as arbuscules, which assist in exchanging of inorganic minerals and organic C and P compounds (Li et al. [2016;](#page-12-0) Prasad et al. [2017;](#page-13-0) Gupta and Abbott [2020\)](#page-11-0). Under P-limited conditions, mycorrhizal association improves P supply (Bucher [2007,](#page-10-0)

Bagyaraj et al. [2015\)](#page-10-0). For example, Pi uptake rate was markedly enhanced in the AMF-colonized maize, strawberry and onion plants (Garcés-Ruiz et al. [2017](#page-11-0); Sharma and Adholeya, [2000](#page-13-0) and [2004\)](#page-13-0). It is evident that inoculation with AMF and enforcing the AMF symbiosis can significantly enhance the concentration of various macro and micro-nutrients, which leads to increased photosynthates production and hence increased biomass accumulation (Tchabi et al. [2010](#page-14-0); Chen et al. [2017](#page-10-0); Mitra et al. [2019;](#page-12-0) Gupta et al. [2019](#page-11-0)). Apart from the macronutrients, AMF association has been reported to increase the phyto-availability of micronutrients like zinc and copper (Cu; Smith and Read [2008\)](#page-13-0). AMF inoculation in tomato plants increased leaf area, N, P, K and Ca contents, reflecting enhanced plant growth (Balliu et al. [2015\)](#page-10-0), in chickpea improved growth and increased levels of protein, iron Fe, and Zn (Pellegrino and Bedini [2014\)](#page-13-0) and in soybean enhanced nodulation and physiological status (Sharma et al. [2012,](#page-13-0) [2016\)](#page-13-0). Increased N in colonized plants by AMF- results in higher chlorophyll contents, as chlorophyll molecules can effectively trap N (De Andrade et al. [2015](#page-10-0)). It was recently being reported that inoculation with native AMF cause significant alterations in the N contents of crop plants (Turrini et al. [2018\)](#page-14-0). Under the conditions of ambient and elevated  $CO<sub>2</sub>$ , a substantial improvement in the accumulation C and N and the assimilation of N due to AMF inoculation has been reported (Zhu et al. [2016\)](#page-15-0).

# 3.4 Improved soil aggregation, stability and soil sequestration

AMF activities vary with the way plants behave under the changed environment and soil conditions. Elevated atmospheric  $CO<sub>2</sub>$  leads to C fixation by plants and more significant transfer of C to growing roots in the soil and stands as one of the most significant factors in AMF functioning. Plants invest additional C in their AMF partner, and AMF through their extended hyphal network facilitates C transport and take part in C cycling thus becoming ecologically crucial in sustaining the productivity of agroecosystems (Parihar et al. [2020\)](#page-12-0).

AMF produce large amounts of copious recalcitrant glycoproteins known as "glomalin" which are hydrophobic, insoluble and significantly important in stabilizing soil aggregates (Wright and Upadhyaya [1998](#page-14-0), Rillig [2004a](#page-13-0)). The greater stability of aggregates leads to larger amounts of protected organic C in terrestrial ecosystems (Wright and Upadhyaya [1998](#page-14-0); Singh [2012;](#page-13-0) Wang et al. [2015](#page-14-0); Gupta [2020](#page-11-0); Gupta and Abbott [2020\)](#page-11-0). AMF influence soil aggregate stability and long-term C and N storage through glomalin (Wright and Upadhyaya [1998](#page-14-0)). Glomalin also responds to plant species and land-use changes (Rillig et al. [2002b,](#page-13-0) [2003](#page-13-0)). However, glomalin concentration in the soil varies and is responsive to AMF species, crop and soil management practices, e.g. crop rotations, tillage, etc. (Agnihotri et al. [2018\)](#page-9-0), and factors of global climate change e.g., elevated atmospheric  $CO<sub>2</sub>$ , warming (Rillig et al. [2002a\)](#page-13-0). Rillig et al.  $(2001)$  $(2001)$ demonstrated the role of AMF-mediated glomalin in soil aggregate water stability. They suggested that glomalin should be used as a useful indicator in assessing the impacts of global climate change based on altered aggregate stability. Nautiyal et al. [\(2019\)](#page-12-0) recently showed that the glomalin concentrations and soil aggregation varied across different cropping systems. They observed that glomalin content (total and easily extractable) was positively correlated with soil organic C. Long back, Rillig et al. [\(1999\)](#page-13-0) observed that when compared to ambient  $CO<sub>2</sub>$  the glomalin production and soil aggregation increased at higher  $CO<sub>2</sub>$  indicating the role of AMF in capturing C. Thus, under  $eCO<sub>2</sub>$ , the soils can retain large amounts of C through the production of glomalin, which can sequester C and may help in mitigating the effects of  $eCO<sub>2</sub>$  (Zhang et al. [2015\)](#page-14-0). To simulate  $eCO<sub>2</sub>$  in different agroecosystems, systematic long-term field studies are needed to establish and predict the contribution of AMF in sequestering soil C.

The adoption of appropriate agriculture management practices can contribute to the mitigation of climate change. Conservation agriculture, for example zero tillage (ZT) practice has been shown to increase soil organic carbon and improve soil quality and glomalin pool (Gathala et al. [2011;](#page-11-0) Agnihotri et al. [2021](#page-9-0)). The C sequestration varies with the cropping system and tillage practices (Aryal et al. [2015](#page-9-0)). This may result in changes of the microbial community composition, e.g. AMF biomass. In one of the long-term field trial in India (over 20 years) where maize–wheat and cotton–wheat production systems under zero tillage system showed more Csequestration than the rice-wheat system under the same practice indicating the role of crop sequences and soil management practices in C cycling. Since AMF are ecologically crucial in maintaining C balance and C storage for the future (Parihar et al. [2020](#page-12-0)). Furthermore, an assay for extraction and quantification of higher recovery of glomalin in soil and roots should be optimized to decipher the role of AMF in C cycling and soil processes (Holatko et al. [2021\)](#page-11-0).

### 4 Conclusions and future prospects

The impact of anthropogenic changes on the current and future global environment is unprecedented; therefore, a holistic and comprehensive approach is required to mitigate these adverse effects. AMF are widespread and highly responsive to the changing environment. This review provides an understanding of the impacts of climatic change and associated crop and soil management practices on AMF symbiosis for ecosystem resilience. Given the massive importance of these fungi in ecosystems, there is a need to include them in global climate change models. The use of real-time data in the models will help to formulate strategies for sustaining the ecosystem

<span id="page-9-0"></span>services and food grain production for the future. The shreds of evidence from climate change studies indicate the shaping up of the AMF communities. However, native AMF are not sufficiently considered, both fully exploited in sustainable crop and soil management practices.

Soil and crop management practices, e.g. doses of fertilizers, use of  $C_3$  or  $C_4$  plants, need to be customized to adopt AMF favoring systems to eventually help in sustaining the productivity of agroecosystems through mitigating the effects of climate change. However, to arrive at a logical conclusion, long-term field studies across the agroecosystems are needed in different climatic situations to elucidate the potential of AMF in mitigating the anthropogenic perturbations and sustaining the belowground ecosystem services systematically. Such studies will help to generate data on the contribution of AMF and associated practices for predicting their role in sustaining the productivity in different agroecosystems for the future.

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### **Declarations**

Ethics approval This article does not contain any studies with human participants or animals performed by any of the author.

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### References

- Addy HD, Schaffer GF, Miller MH, Peterson RL (1994) Survival of the external mycelium of a VAM fungus in frozen soil over winter. Mycorrhiza 5:1–5
- Al-Yahya'ei MN, Oehl F, Vallino M, Lumini E, Redecker D, Wiemken A, Bonfante P (2011) Unique arbuscular mycorrhizal fungal communities uncovered in date palm plantations and surrounding desert habitats of Southern Arabia. Mycorrhiza 21(3):195-209. [https://doi.](https://doi.org/10.1007/s00572-010-0323-5) [org/10.1007/s00572-010-0323-5](https://doi.org/10.1007/s00572-010-0323-5)
- Aryal JP, Sapkota TB, Jat ML, Bishnoi DK (2015) On-farm economic and environmental impact of zero-tillage wehat: A case of Northwest India. Exp Agric 51(1):1–16. [https://doi.org/10.1017/](https://doi.org/10.1017/S001447971400012X) [S001447971400012X](https://doi.org/10.1017/S001447971400012X)
- Agnihotri R, Bharti A, Ramesh A, Prakash A, Sharma MP (2021) Glomalin related protein and C16:1ω5 PLFA associated with AM fungi as potential signatures for assessing the soil C sequestration under contrasting soil management practices. Eur. J. Soil Biol. 103: 103286. <https://doi.org/10.1016/j.ejsobi.2021.10328>
- Agnihotri R, Maheshwari H, Ramesh A, Praksh A, Sharma MP (2018) Glomalin, the AM fungi secreted soil protein: role in sequestering soil carbon and heavy metal reduction (pages 105-114). In: Rakshit A, Sarkar B, Abhilash PC (eds) Soil Amendments for Sustainability: Challenges and Perspectives, 1st edn. CRC Press, Taylor Francis group, Boca Raton, p 428. <https://doi.org/10.1201/9781351027021>
- Ainsworth EA, Davey PA, Hymus GJ, Osborne CP, Rogers A, Blum H et al (2003) Is stimulation of leaf photosynthesis by elevated carbon dioxide concentration maintained in the long term? A test with Lolium perenne grown for 10 years at two nitrogen fertilization levels under Free Air CO<sub>2</sub> Enrichment (FACE). Plant Cell Environ 26(5):705–714
- Ainsworth EA, Long SP (2005) What have we learned from 15 years of free-air  $CO<sub>2</sub>$  enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising  $CO<sub>2</sub>$ . New Phytol 165:351-372
- Aldrich-Wolfe L, Black KL, Hartmann EDL et al (2020) Taxonomic shifts in arbuscular mycorrhizal fungal communities with shade and soil nitrogen across conventionally managed and organic coffee agroecosystems. Mycorrhiza 30:513–527
- Alguacil MM, Torrecillas E, Lozano Z, Roldán A (2015) Arbuscular mycorrhizal fungi communities in a coral cay system (Morrocoy, Venezuela) and their relationships with environmental variables. Sci Total Environ 505:805–813
- Alguacil MM, Torrecillas E, Lozano Z, Torres MP, Roldán A (2014) Prunus persica crop management differentially promotes arbuscular mycorrhizal fungi diversity in a tropical agro-ecosystem. PLoS One 9(2):e88454. <https://doi.org/10.1371/journal.pone.0088454>
- Al-Karaki G, McMichael B, Zak J (2004) Field response of wheat to arbuscular mycorrhizal fungi and drought stress. Mycorrhiza 14: 263–269
- Antoninka A, Reich PB, Johnson NC (2011) Seven years of carbon dioxide enrichment, nitrogen fertilization and plant diversity influence arbuscular mycorrhizal fungi in a grassland ecosystem. New Phytol 192:200–214
- Asha HN, Nirmalnath PJ, Sagarkar MA, Venkatesh H (2017) Impact of elevated  $CO<sub>2</sub>$  and/or temperature on the am fungal diversity in groundnut rhizosphere under open top chamber facility. Int J Curr Microbiol App Sci 6(1):882–895. [https://doi.org/10.20546/ijcmas.](https://doi.org/10.20546/ijcmas.2017.601.104) [2017.601.104](https://doi.org/10.20546/ijcmas.2017.601.104)
- Auge RM (2001) Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. Mycorrhiza 11:3–42
- Augé RM, Stodola AJ, Tims JE et al (2001) Moisture retention properties of a mycorrhizal soil. Plant Soil 230:87–97. [https://doi.org/10.1023/](https://doi.org/10.1023/A:1004891210871) [A:1004891210871](https://doi.org/10.1023/A:1004891210871)
- <span id="page-10-0"></span>Augé RM, Toler HD, Saxton AM (2015) Arbuscular mycorrhizal symbiosis alters stomatal conductance of host plants more under drought than under amply watered conditions: a meta-analysis. Mycorrhiza 25:13–24. <https://doi.org/10.1007/s00572-014-0585-4>
- Bagyaraj DJ, Sharma MP, Maiti D (2015) Phosphorus nutrition of crops through arbuscular mycorrhizal fungi. Curr Sci 108:1288–1293
- Balestrini R, Magurno F, Walker C, Lumini E, Bianciotto V (2010) Cohorts of arbuscular mycorrhizal fungi (AMF) in Vitis vinifera, a typical Mediterranean fruit crop. Environ Microbiol Rep 2:594–604. <https://doi.org/10.1111/j.1758-2229.2010.00160.x>
- Balliu A, Sallaku G, Rewald B (2015) AMF Inoculation enhances growth and improves the nutrient uptake rates of transplanted, salt-stressed tomato seedlings. Sustainability 7:15967–15981
- Bansal M, Mukerji KG (1994) Positive correlation between VAMinduced changes in root exudation and mycorrhizosphere mycoflora. Mycorrhiza 5:39–44
- Barzana G, Aroca R, Bienert GP, Chaumont F, Ruiz-Lozano JM (2014) New insights into the regulation of aquaporins by the arbuscular mycorrhizal symbiosis in maize plants under drought stress and possible implications for plant performance. Mol Plant Microbe Interact 27:349–363
- Baltruschat H, Santos V, Silva D, Schellenberg I, Deubel A, Sieverding E, Oehl F (2019) Unexpectedly high diversity of arbuscular mycorrhizal fungi in fertile Chernozem croplands in Central Europe. Catena 182:104–135
- Bazzaz FA (1990) The response of natural ecosystems to the rising global CO2 levels. Ann Rev Ecol Syst 21:167–196
- Begum N, Ahanger MA, Zhang L (2020) AMF inoculation and phosphorus supplementation alleviates drought induced growth and photosynthetic decline in Nicotiana tabacum by up-regulating antioxidant metabolism and osmolyte accumulation. Environ Exp Bot 176: 104088. <https://doi.org/10.1016/j.envexpbot.2020.104088>
- Behera SN, Sharma M, Aneja VP et al (2013) Ammonia in the atmosphere: a review on emission sources, atmospheric chemistry and deposition on terrestrial bodies. Environ Sci Pollut Res 20:8092– 8131
- Berruti A, Lumini E, Balestrini R, Bianciotto V (2016) Arbuscular Mycorrhizal Fungi as Natural Biofertilizers: Let's Benefit from Past Successes. Front. Microbiol. 6:1559. [https://doi.org/10.3389/](https://doi.org/10.3389/fmicb.2015.01559) [fmicb.2015.01559](https://doi.org/10.3389/fmicb.2015.01559)
- Betts RA, Jones CD, Knight JR et al (2016) El Nino and a record CO<sub>2</sub> rise. Nat Clim Change 6:806–810
- Bouffaud ML, Bernaud E, Colombet A, van Tuinen D, Wipf D, Redecker D (2016) Regional-scale analysis of arbuscular mycorrhizal fungi: the case of Burgundy vineyards. J Int Sci Vigne Vin 50:1–8. [https://](https://doi.org/10.20870/oeno-one.2016.50.1.49) [doi.org/10.20870/oeno-one.2016.50.1.49](https://doi.org/10.20870/oeno-one.2016.50.1.49)
- Buade R, Chourasiya D, Prakash A, Sharma MP (2020) Changes in Arbuscular mycorrhizal fungal community structure in soybean rhizosphere soil assessed at different growth stages of soybean. Agric Res. <https://doi.org/10.1007/s40003-020-00481-4>
- Bucher M (2007) Functional biology of plant phosphate uptake at root and mycorrhizae interfaces. New Phytol 173:11–26. [https://doi.org/](https://doi.org/10.1111/j.1469-8137.2006.01935.x) [10.1111/j.1469-8137.2006.01935.x](https://doi.org/10.1111/j.1469-8137.2006.01935.x)
- Buddrick O, Jones OAH, Cornell HJ, Small DM (2014) The influence of fermentation processes and cereal grains in wholegrain bread on reducing phytate content. J Cereal Sci 59:3–8
- Cameron DD, Neal AL, van Wees SCM et al (2013) Mycorrhiza-induced resistance: more than the sum of its parts? Trends Plant Sci 18:539– 545
- Campbell CD, Sage RF (2002) Interactions between atmospheric  $CO<sub>2</sub>$ concentration and phosphorus nutrition on the formation of proteoid roots in white lupin (Lupinus albus L.). Plant Cell Environ 25:1051– 1059
- Carrillo M, Gavito M (2018) Arbuscular mycorrhizal fungal spore communities of a tropical dry forest ecosystem show resilience to land-

use change. Fungal Ecol. 32. [https://doi.org/10.1016/j.funeco.2017.](https://doi.org/10.1016/j.funeco.2017.11.006) [11.006](https://doi.org/10.1016/j.funeco.2017.11.006)

- Castillo CG, Borie F, Oehl F, Sieverding E (2016) Arbuscular mycorrhizal fungi biodiversity: prospecting in Southern-Central zone of Chile. A review. J. Soil Sci. Plant Nutr. 16(2):400–422
- Cavagnaro TR, Sokolow SK, Jackson LE (2007) Mycorrhizal effects on growth and nutrition of tomato under elevated atmospheric carbon dioxide. Funct Plant Biol 34:730–736
- Chareesri A, De Deyn GB, Sergeeva L et al (2020) Increased arbuscular mycorrhizal fungal colonization reduces yield loss of rice (Oryza sativa L.) under drought. Mycorrhiza 30:315–328
- Chen YL, Xu ZW, Xu TL et al (2017) Nitrogen deposition and precipitation induced phylogenetic clustering of arbuscular mycorrhizal fungal communities. Soil Biol Biochem 115:233–242
- Chen B, Xiao X, Zhu YG, Smith FA, Xie ZM, Smith SE (2007) The arbuscular mycorrhizal fungus Glomus mosseae gives contradictory effects on phosphorus and arsenic acquisition by Medicago sativa Linn. Sci Total Environ 379(2-3):226–234
- Cheng L, Booker FL, Tu C, Burkey KO, Zhou L, Shew HD et al (2012) Arbuscular mycorrhial fungi increase organic carbon decomposition under elevated CO<sub>2</sub>. Science 337:1084-1087
- Clark NM, Rillig MC, Nowak RS (2009) Arbuscular mycorrhizal fungal abundance in the Mojave Desert: seasonal dynamics and impacts of elevated  $CO<sub>2</sub>$ . J Arid Environ 73(9):834–843F
- Classen AT, Sundqvist MK, Henning JA, Newman GS, Moore JAM, Cregger MA, Moorhead LC, Patterson CM (2015) Direct and indirect effects of climate change on soil microbial and soil microbialplant interactions: What lies ahead? Ecosphere 6(8):130
- Compant S, Van Der Heijden MGA, Sessitsch A (2010) Climate change effects on beneficial plant–microorganism interactions. FEMS Microbiol Ecol 73:197–214
- Cotton TEA, Fitter AH, Miller RM, Dumbrell AJ, Helgason T (2015) Fungi in the future: interannual variation and effects of atmospheric change on arbuscular mycorrhizal fungal communities. New Phytol 205:1598–1607. <https://doi.org/10.1111/nph.13224>
- Cotton TEA (2018) Arbuscular mycorrhizal fungal communities and global change: an uncertain future. FEMS Microbiol. Ecol. [https://](https://doi.org/10.1093/femsec/fiy179) [doi.org/10.1093/femsec/fiy179](https://doi.org/10.1093/femsec/fiy179)
- Cui J, Bai L, Liu X, Jie W, Cai B (2018) Arbuscular mycorrhizal fungal communities in the rhizosphere of a continuous cropping soybean system at the seedling stage. Braz J Microbiol. 49(2):240–247. <https://doi.org/10.1016/j.bjm.2017.03.017>
- Daepp M, Nösberger J, Lüscher A (2001) Nitrogen fertilization and developmental stage alter the response of Lolium perenne to elevated CO2. New Phytol 150:347–358
- De Andrade SAL, Domingues AP, Mazzafera P (2015) Photosynthesis is induced in rice plants that associate with arbuscular mycorrhizal fungi and are grown under arsenate and arsenite stress. Chemosphere 134:141–149. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.chemosphere.2015.04.023) [chemosphere.2015.04.023](https://doi.org/10.1016/j.chemosphere.2015.04.023)
- De Oliveira JRG, de Resende GM, de Melo NF, Yano-Melo AM (2017) Symbiotic compatibility between arbuscular mycorrhizal fungi (autoctone or exotic) and three native species of the Caatinga in different phosphorus levels. Acta Sci Biol Sci 39:59–69
- Deepika S, Kothamasi D (2015) Soil moisture a regulator of arbuscular mycorrhizal fungal community assembly and symbiotic phosphorus uptake. Mycorrhiza 25:67–75
- Del Buono D (2021) Can biostimulants be used to mitigate the effect of anthropogenic climate change on agriculture? It is time to respond. Sci Total Environ 751:141763. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2020.141763) [2020.141763](https://doi.org/10.1016/j.scitotenv.2020.141763)
- Deveautour C, Donn S, Power SA et al (2018) Experimentally altered rainfall regimes and host root traits affect grassland arbuscular mycorrhizal fungal communities. Mol Ecol 27:2152–2163
- Diagne N, Ndour M, Djighaly PI, Ngom D, Ngom MCN, Ndong G, Svistoonoff S, Cherif-Silini H (2020) Effect of Plant Growth

<span id="page-11-0"></span>Promoting Rhizobacteria (PGPR) and Arbuscular Mycorrhizal Fungi (AMF) on Salt Stress Tolerance of Casuarina obesa (Miq.). Front. Sustain. Food Syst 4:601004. [https://doi.org/10.3389/fsufs.](https://doi.org/10.3389/fsufs.2020.601004) [2020.601004](https://doi.org/10.3389/fsufs.2020.601004)

- Drigo B, Kowalchuk GA, van Veen JA (2008) Climate change goes underground: effects of elevated atmospheric CO2 on microbial community structure and activities in the rhizosphere. Biol Fert Soils 44(5):667–679. <https://doi.org/10.1007/s00374-008-0277-3>
- Drigo B, Pijl AS, Duyts H et al (2010) Shifting carbon flow from roots into associated microbial communities in response to elevated atmospheric CO<sub>2</sub>. P Natl Acad Sci 107:10938-10942
- Dumbrell AJ, Ashton PD, Aziz N, Feng G, Nelson M, Dytham C, Fitter AH, Helgason T (2011) Distinct seasonal assemblages of arbuscular mycorrhizal fungi revealed by massively parallel pyrosequencing. New Phytol 190:794–804
- Edwards EJ, Osborne CP, Strömberg CAE, Smith SA (2010) The origins of C4 grasslands: integrating evolutionary and ecosystem science. Science 328:587–591. <https://doi.org/10.1126/science.1177216>
- Fitter AH, Heinemeyer A, Staddon PL (2000) The impact of elevated  $CO<sub>2</sub>$  and global climate change on arbuscular mycorrhizas: a mycocentric approach. New Phytol 147:179–187
- Gamper H, Hartwig UA, Leuchtmann A (2005) Mycorrhizas improve nitrogen nutrition of Trifolium repens after 8 yr of selection under elevated atmospheric CO<sub>2</sub> partial pressure. New Phytol 167:531-542. [https://doi.org/10.1111/j.1469-8137.2005.01440.x2005.](https://doi.org/10.1111/j.1469-8137.2005.01440.x2005.01440.x) [01440.x](https://doi.org/10.1111/j.1469-8137.2005.01440.x2005.01440.x)
- Gamper H, Peter M, Jansa J, Liischer A, Hartwig UA, Leuchtmann A (2004) Arbuscular mycorrhizal fungi benefit from 7 years of free air CO2 enrichment in well-fertilized grass and legume monocultures. Global Change Biol 10:189–199
- Gao C, Kim YC, Zheng Y et al (2016) Increased precipitation, rather than warming, exerts a strong influence on arbuscular mycorrhizal fungal community in a semiarid steppe ecosystem. Botany 94:459–469
- Garcés-Ruiz M, Calonne-Salmon M, Plouznikoff K, Misson C, Navarrete-Mier M, Cranenbrouck S et al (2017) Dynamics of short-term phosphorus uptake by intact mycorrhizal and nonmycorrhizal maize plants grown in a circulatory semi-hydroponic cultivation system. Front Plant Sci 8:1471. [https://doi.org/10.3389/](https://doi.org/10.3389/fpls.2017.01471) [fpls.2017.01471](https://doi.org/10.3389/fpls.2017.01471)
- Gathala MK, Ladha J, Kumar V, Saharawat YS, Kumar V, Sharma PK et al (2011) Tillage and crop establishment affects sustainability of South Asian rice–wheat system. Agron J 103(4):961–971
- Gavito M, Castillo D, González-Monterrubio C, Vieira-Hernández T, Martinez-Trujillo M (2008) High compatibility between arbuscular mycorrhizal fungal communities and seedlings of different land use types in a tropical dry ecosystem. Mycorrhiza. 19:47–60. [https://doi.](https://doi.org/10.1007/s00572-008-0203-4) [org/10.1007/s00572-008-0203-4](https://doi.org/10.1007/s00572-008-0203-4)
- Gavito ME, Azcón-Aguilar C (2012) Temperature stress in arbuscular mycorrhizal fungi: a test for adaptation to soil temperature in three isolates of Funneliformis mosseae from different climates. Agric. Food Sci 21(1):2–11. <https://doi.org/10.23986/afsci.4994>
- Gavito ME, Bruhn D, Jakobsen I (2002) Phosphorus uptake by arbuscular mycorrhizal hyphae does not increase when the host plant grows under atmospheric CO<sub>2</sub> enrichment. New Phytol 154: 751–760. <https://doi.org/10.1046/j.1469-8137.2002.00404.x>
- Gavito ME, Curtis PS, Mikkelsen TN, Jakobsen I (2000) Atmospheric CO<sub>2</sub> and mycorrhiza effects on biomass allocation and nutrient uptake of nodulated pea (Pisum sativum L.) plants. J Exp Bot 51:1931– 1938
- Graham JH, Leonard RT, Menge JA (1982) Interaction of light intensity and soil temperature with phosphorus inhibition of vesiculararbuscular mycorrhiza formation. New Phytol 91(4):683–690
- Grover M, Maheswari M, Desai S, Gopinath KA, Venkateswarlu B  $(2015)$  Elevated CO<sub>2</sub>: Plant associated microorganisms and carbon sequestration. Appl Soil Ecol 95:73–85
- Gupta MM (2020) Arbuscular mycorrhizal fungi the potential soil health indicators. In: Giri B, Verma A (eds) Soil Health. Springer, Cham, pp 183–195
- Gupta MM, Abbott LK (2020) Exploring economic assessment of the arbuscular mycorrhizal symbiosis. Symbiosis. [https://doi.org/10.](https://doi.org/10.1007/s13199-020-00738-0) [1007/s13199-020-00738-0](https://doi.org/10.1007/s13199-020-00738-0)
- Gupta MM, Chourasiya D, Sharma MP (2019) Diversity of arbuscular mycorrhizal fungi in relation to sustainable plant production systems. In: Satyanarayana T, Johri BN, Das SK (eds) Microbial diversity in ecosystem sustainability and biotechnological applications volume 1. Microbial Diversity in Normal and Extreme Environments, Springer Singapore, pp 167–186
- Gupta MM, Aggarwal A (2018) From Mycorrhizosphere to Rhizosphere Microbiome: The Paradigm Shift. In Root Biology, pp 487–500
- Hart MM, Antunes PM, Chaudhary VB, Abbott LK (2018) Fungal inoculants in the field: is the reward greater than the risk? Funct Ecol 32(1):126–135 Hashem A, Alqarawi AA, Radhakrishnan R, Al-Arjani ABF, Aldehaish HA, Egamberdieva D, & Abd\_Allah EF. (2018). Arbuscular mycorrhizal fungi regulate the oxidative system, hormones and ionic equilibrium to trigger salt stress tolerance in Cucumis sativus L Saudi J. Biol. Sci. 25(6):1102-1114
- Hawkes CV, Hartley IP, Ineson P, Fitter AH (2008) Soil temperature affects carbon allocation within arbuscular mycorrhizal networks and carbon transport from plant to fungus. Global Change Biol 14(5):1181–1190
- Hazard C, Gosling P, van der Gast CJ et al (2013) The role of local environment and geographical distance in determining community composition of arbuscular mycorrhizal fungi at the landscape scale. ISME J 7:498–508. <https://doi.org/10.1038/ismej.2012.127>
- Heinemeyer A, Ineson P, Ostle N et al (2006) Respiration of the external mycelium in the arbuscular mycorrhizal symbiosis shows strong dependence on recent photosynthates and acclimation to temperature. New Phytol 171:159–170
- Hetrick BAD (1991) Mycorrhizas and root architecture. Experientia 47(4):355–362. <https://doi.org/10.1007/BF01972077>
- Hildebrandt U, Regvar M, Bothe H (2007) Arbuscular mycorrhiza and heavy metal tolerance. Phytochemistry 68(1):139–146
- Hoeksema JD, Bala Chaudhary V, Gehring CA, Collins Johnson N, Karst J, Koide RT et al (2010) A meta-analysis of context-dependency in plant response to inoculation with mycorrhizal fungi. Ecol Lett 13: 394–407. <https://doi.org/10.1111/j.1461-0248.2009.01430.x>
- Holátko J, Brtnický M, Kučerík J, Kotianová M, Elble J, Kintl A, Kynický J, Benada O, Datta R, Jansa J (2021) Glomalin – Truths, myths, and the future of this elusive soil glycoprotein (Perspectives Paper). Soil Biol Biochem 153:108116
- IPCC (2014) In: Core Writing Team, Pachauri RK, Meyer LA (eds) Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change. IPCC, Geneva, Switzerland, p 151
- IPCC, Climate Change (2013) The physical science basis: working group I contribution to the fifth assessment report of the intergovernmental panel on climate change. Cambridge University Press
- Jakobsen I, Abbott LK, Robson AD (1992) External hyphae of vesiculararbuscular mycorrhizal fungi associated with Trifolium subterraneum L. 2. Hyphal uptake of 32P at defined distances from roots. New Phytol 120(4):509–516
- Jakobsen I, Smith SE, Smith FA, Watts-Williams SJ, Clausen SS, Gronlund M (2016) Plant growth responses to elevated atmospheric CO2 are increased by phosphorus sufficiency but not by arbuscular mycorrhizas. J Exp Botany 67(21):6173–6186
- Jarrell WM, Beverly RB (1981) The dilution effect in plant nutrition studies. Adv Agron 34:197–224
- Johnson D, Vandenkoornhuyse PJ, Leake JR, Gilbert L, Booth RE, Grime JP et al (2004) Plant communities affect arbuscular

<span id="page-12-0"></span>mycorrhizal fungal diversity and community composition in grassland microcosms. New Phytol 161:503–515

- Johnson NC (2010) Resource stoichiometry elucidates the structure and function of arbuscular mycorrhizas across scales. New Phytol 185: 631–647
- Johnson NC, Angelard C, Sanders IR et al (2013a) Predicting community and ecosystem outcomes of mycorrhizal responses to global change. Ecol Lett 16:140–153. <https://doi.org/10.1111/ele.12085>
- Johnson JM, Houngnandan P, Aboubacry K, Kadidia S, Marc N et al (2013b) Diversity patterns of indigenous arbuscular mycorrhizal fungi associated with rhizosphere of cowpea (Vigna unguiculata (L.) Walp.) In Benin, West Africa. Pedobiologia. 56:121–128. <https://doi.org/10.1016/j.pedobi.2013.03.003>
- Johnson NC, Rowland DL, Corkidi L, Egerton-Warburton LM, Allen EB (2003) Nitrogen enrichment alters mycorrhizal allocation at five mesic to semiarid grasslands. Ecology 84:1895–1908
- Kandeler E, Mosier AR, Morgan JA, Milchunas DG, King JY, Rudolph S, Tscherko D (2006) Response of soil microbial biomass and enzyme activities to the transient elevation of carbon dioxide in a semiarid grassland. Soil Biol Biochem 38(8):2448–2460
- Kaya C, Ashraf M, Sonmez O, Aydemir S, Tuna AL, Cullu MA (2009) The influence of arbuscular mycorrhizal colonization on key growth parameters and fruit yield of pepper plants grown at high salinity. Sci Hortic 121:1–6
- Klironomos JN, Allen MF, Rillig MC et al (2005) Abrupt rise in atmospheric  $CO<sub>2</sub>$  overestimates community response in a model plant– soil system. Nature 433:621–624
- Kohler J, Caravaca F, Alguacil MDM, Roldan A (2009) Elevated CO<sub>2</sub> increases the effect of an arbuscular mycorrhizal fungus and a plantgrowth-promoting rhizobacterium on structural stability of a semiarid agricultural soil under drought conditions. Soil Biol Biochem 41:1710–1716
- Landis FC, Gargas A, Givnish TJ (2004) Relationships among arbuscular mycorrhizal fungi, vascular plants and environmental conditions. New Phytol 164:493–504
- LeBauer DS, Treseder KK (2008) Nitrogen limitation of net primary productivity in terrestrial ecosystems is globally distributed. Ecology 89:371–379. <https://doi.org/10.1890/06-2057.1>
- Li T, Hu YJ, Hao ZP, Li H, Chen BD (2013) Aquaporin genes GintAQPF1 and GintAQPF2 from Glomus intraradices contribute to plant drought tolerance. Plant Signal Behav 8 (5) :e24030
- Li X, Zhu T, Peng F et al (2015) Inner Mongolian steppe arbuscular mycorrhizal fungal communities respond more strongly to water availability than to nitrogen fertilization. Environ Microbiol 17: 3051–3068
- Li X, Zeng R, Liao H (2016) Improving crop nutrient efficiency through root architecture modifications. J Integr Plant Biol 58:193–202. <https://doi.org/10.1111/jipb.12434>
- Lugtenberg B, Kamilova F (2009) Plant-growth-promoting rhizobacteria. Annu Rev Microbiol 63:541–556
- Martínez-García LB, de Dios MJ, Pugnaire FI (2012) Impacts of changing rainfall patterns on mycorrhizal status of a shrub from arid environments. Eur J Soil Biol 50:64–67
- Mathimaran N, Sharma MP, Mohan Raju B, Bagyaraj DJ (2017) Mycosphere Essay 17 Arbuscular mycorrhizal symbiosis and drought tolerance in crop plants. Mycosphere 8(3):361–376
- Mathur S, Sharma MP, Jajoo A (2018) Improved photosynthetic efficacy of maize (Zea mays) plants with arbuscular mycorrhizal fungi (AMF) under high temperature stress. J Photochem Photobiol B 180:149–154
- Maurer C, Ruedy M, Chervet A, Sturny WG, Flisch R, Oehl F (2014) Diversity of arbuscular mycorrhizal fungi in field crops using no-till and conventional tillage practices. Agrar forschung Schweiz 5(10): 398–405
- McHugh TA, Schwartz E (2015) Changes in plant community composition and reduced precipitation have limited effects on the structure of

soil bacterial and fungal communities present in a semiarid grassland. Plant Soil 388:175–186

- Medina J, Cornejo P, Borie F, Meyer S, Palenzuela J, Vieira HEE, Ferreira ACA, Silva GA, Sánchez-Castro I, Oehl F (2014) Corymbiglomus pacificum, a new Glomeromycete from a saline lakeshore in Chile. Mycotaxon 127:173–183
- Medina J, Meier S, Rubio R, Curaqueo G, Borie F, Aguilera P, Oehl F, Cornejo P (2015) Arbuscular mycorrhizal status of pioneer plants from the mouth of lake Budi, Araucanía Region, Chile. J Soil Sci Plant Nutr 15(1):142–152. [https://doi.org/10.4067/S0718-](https://doi.org/10.4067/S0718-95162015005000012) [95162015005000012](https://doi.org/10.4067/S0718-95162015005000012)
- Melo CD, Luna S, Krüger C et al (2017) Arbuscular mycorrhizal fungal community composition associated with Juniperus brevifolia in native Azorean forest. Acta Oecol 79:48–61
- Melo CD, Walker C, Krüger C et al (2019) Environmental factors driving arbuscular mycorrhizal fungal communities associated with endemic woody plant Picconiaazorica on native forest of Azores. Ann Microbiol 69:1309–1327. [https://doi.org/10.1007/s13213-019-](https://doi.org/10.1007/s13213-019-01535-x) [01535-x](https://doi.org/10.1007/s13213-019-01535-x)
- Mitra D, Navendra U, Panneerselvam U, Ansuman S, Ganeshamurthy AN, Divya J (2019) Role of mycorrhiza and its associated bacteria on plant growth promotion and nutrient management in sustainable agriculture. Int J Life Sci Appl Sci 1:1–10
- Montesinos-Navarro A, Valiente-Banuet A, Verdú M (2019) Processes underlying the effect of mycorrhizal symbiosis on plant-plant interactions. Fungal Ecol 40:98–106
- Mortimer PE, Le Roux MR, Perez-Fernandez MA, Benedito VA, Kleinert A, Xu JC, Valentine AJ (2013) The dual symbiosis between arbuscular mycorrhiza and nitrogen fixing bacteria benefits the growth and nutrition of the woody invasive legume Acacia cyclops under nutrient limiting conditions. Plant Soil 366:229–241
- Mueller RC, Bohannan BJM (2015) Shifts in the phylogenetic structure of arbuscular mycorrhizal fungi in response to experimental nitrogen and carbon dioxide additions. Oecologia 179:175–185
- Nautiyal P, Rajput R, Pandey D, Arunachalam K, Arunachalam A (2019) Role of glomalin in soil carbon storage and its variation across land uses in temperate Himalayan regime. Biocatal Agric Biotechnol 21: 101311
- Njeru EM, Avio L, Bocci G, Sbrana C, Turrini A, Bàrberi P, Giovannetti M, Oehl F (2015) Contrasting effects of cover crops on 'hot spot' arbuscular mycorrhizal fungal communities in organic tomato. Biol Fertil Soils 51:151–166
- Nuccio EE, Hodge A, Pett-Ridge J et al (2013) An arbuscular mycorrhizal fungus significantly modifies the soil bacterial community and nitrogen cycling during litter decomposition. Environ Microbiol 15: 1870–1881
- Ocon A, Hampp R, Requena N (2007) Trehalose turnover during abiotic stress in arbuscular mycorrhizal fungi. New Phytol 174:879–891
- Oehl F, Sieverding E, Mäder P, Dubois D, Ineichen K, Boller T, Wiemken A (2004) Impact of long-term conventional and organic farming on the diversity of arbuscular mycorrhizal fungi. Oecologia 138(4):574–583
- Oehl F, Sieverding E, Ineichen K, Mäder P, Boller T, Weimken A (2003) Impact of land use intensity on the diversity of arbuscular mycorrhizal fungi in agroecosystems of Central Europe. Appl Environ Microbiol 69:2816–2824
- Pande M, Tarafdar JC (2004) Arbuscular mycorrhizal fungal diversity in neem-based agroforestry systems in Rajasthan. Appl Soil Ecol 26: 233–241. <https://doi.org/10.1016/j.apsoil.2003.12.009`>
- Panwar JD (1993) Response of VAM and Azospirillum inoculation to water status and grain yield in wheat under water stress condition. Indian J Plant Physiol 36:41–43
- Parihar M, Rakshit A, Meena VS, Gupta VK, Rana K, Choudhary M, Tiwari G, Mishra PK, Pattanayak A, Bisht JK, Jatav SS, Khati P, Jatav H (2020) The potential of arbuscular mycorrhizal fungi in C

<span id="page-13-0"></span>cycling: a review. Arch Microbiol 202:1581–1596. [https://doi.org/](https://doi.org/10.1007/s00203-020-01915-x) [10.1007/s00203-020-01915-x](https://doi.org/10.1007/s00203-020-01915-x)

- Pellegrino E, Bedini S (2014) Enhancing ecosystem services in sustainable agriculture: biofertilization and biofortification of chickpea (Cicer arietinum L.) by arbuscular mycorrhizal fungi. Soil Biol Biochem 68:429–439. <https://doi.org/10.1016/j.soilbio.2013.09.030>
- Pellegrino E, Gamper HA, Ciccolini V, Ercoli L (2019) Forage rotations conserve diversity of arbuscular mycorrhizal fungi and soil fertility. Front Microbiol 10:2969. [https://doi.org/10.3389/fmicb.2019.](https://doi.org/10.3389/fmicb.2019.02969) [02969](https://doi.org/10.3389/fmicb.2019.02969)
- Pereira CMR, Silva DKA, Ferreira ACA, Goto BT, Maia LC (2014) Diversity of arbuscular mycorrhizal fungi in Atlantic forest areas under different land uses. Agric Ecosystem Environ 185:245–252
- Phillips RP, Meier IC, Bernhardt ES, Grandy AS, Wickings K, Finzi AC (2012) Roots and fungi accelerate carbon and nitrogen cycling in forests exposed to elevated  $CO<sub>2</sub>$ . Ecol Lett 15:1042–1049
- Pischl PH, Barber NA (2016) Plant responses to arbuscular mycorrhizae under elevated temperature and drought. J Plant Ecol 10:692–701
- Prasad R, Bhola D, Akdi K, Cruz C, Sairam KVSS, Tuteja N et al (2017) Introduction to mycorrhiza: historical development. In: Varma A, Prasad R, Tuteja N (eds) Mycorrhiza - Function, Diversity. State of the Art. Springer, Cham, pp 1–7
- Prates Júnior P, Moreira BC, da Silva MCS, Veloso TGR, Stürmer SL et al (2019) Agroecological coffee management increases arbuscular mycorrhizal fungi diversity. PLoS One 14(1):e0209093. [https://doi.](https://doi.org/10.1371/journal.pone.0209093) [org/10.1371/journal.pone.0209093](https://doi.org/10.1371/journal.pone.0209093)
- Pugnaire FI, Morillo JA, Peñuelas J, Reich PB, Bardgett RD, Gaxiola A et al (2019) Climate change effects on plant-soil feedbacks and consequences for biodiversity and functioning of terrestrial ecosystems. Sci Adv. 5(11). <https://doi.org/10.1126/sciadv.aaz1834>
- Ramírez-Gómez M, Pérez-Moncada U, Serralde-Ordoñez D, Peñaranda-Rolón A, Roveda-Hoyos G, Rodriguez A (2019) Diversity of arbuscular mycorrhizal fungi communities associated with cape gooseberry (Physalis peruviana L.) crops. Agron Colomb 37(3): 239–254. <https://doi.org/10.15446/agron.colomb.v37n3.74008>
- Rangel-Castro JI, Killham K, Ostle N, Nicol GW, Anderson IC, Scrimgeour CM, Ineson P, Meharg A, Prosser JI (2005) Stable isotope probing analysis of the influence of liming on root exudate utilization by soil microorganisms. Environ Microbiol 7(6):828–838
- Rasche F, Lueders T, Schloter M, Schaefer S, Buegger F, Gattinger A, Hood-Nowotny RC, Sessitsch A (2009) DNA-based stable isotope probing enables the identification of active bacterial endophytes in potatoes. New Phytol 181(4):802–807
- Rasmussen PU, Hugerth LW, Blanchet FG, Andersson AF, Lindahl BD, Tack AJ (2018) Multiscale patterns and drivers of arbuscular mycorrhizal fungal communities in the roots and root-associated soil of a wild perennial herb. New Phytol 220(4):1248–1261
- Rillig MC (2004a) Arbuscular mycorrhizae, glomalin, and soil aggregation. Can J Soil Sci 84:355–363
- Rillig MC (2004b) Arbuscular mycorrhizae and terrestrial ecosystem processes. Ecol Lett 7:740–754. [https://doi.org/10.1111/j.1461-](https://doi.org/10.1111/j.1461-0248.2004.00620.x2004.00620.x) [0248.2004.00620.x2004.00620.x](https://doi.org/10.1111/j.1461-0248.2004.00620.x2004.00620.x)
- Rillig MC, Field CB, Allen MF (1999) Soil biota responses to long-term atmospheric  $CO<sub>2</sub>$  enrichment in two California annual grasslands. Oecologia 119:572–577
- Rillig MC, Ramsey PW, Morris S et al (2003) Glomalin, an arbuscularmycorrhizal fungal soil protein, responds to land-use change. Plant Soil 253:293–299
- Rillig MC, Wright SF, Shaw MR, Field CB (2002a) Artificial climate warming positively affects arbuscular mycorrhizae but decreases soil aggregate water stability in an annual grassland. Oikos. 97(1): 52–58. <https://doi.org/10.1034/j.1600-0706.2002.970105.x>
- Rillig MC, Wright SF, Eviner VT (2002b) The role of arbuscular mycorrhizal fungi and glomalin in soil aggregation: comparing effects of five plant species. Plant Soil 238:325–333
- Rillig MC, Wright SF, Nichols KA et al (2001) Large contribution of arbuscular mycorrhizal fungi to soil carbon pools in tropical forest soils. Plant Soil 233:167–177
- Ruiz-Lozano JM (2003) Arbuscular mycorrhizal symbiosis and alleviation of osmotic stress. New perspectives for molecular studies. Mycorrhiza 13:309–317
- Saleh AM, Abdel-Mawgoud M, Hassan AR et al (2020) Global metabolic changes induced by arbuscular mycorrhizal fungi in oregano plants grown under ambient and elevated levels of atmospheric CO2. Plant Physiol Biochem 151:255–263. [https://doi.org/10.](https://doi.org/10.1016/j.plaphy.2020.03.026) [1016/j.plaphy.2020.03.026](https://doi.org/10.1016/j.plaphy.2020.03.026)
- Schreiner RP, Mihara K (2009) The diversity of arbuscular mycorrhizal fungi amplified from grapevine roots (Vitis vinifera L.) in Oregon vineyards is seasonally stable and influenced by soil and vine age. Mycologia 101:599–611
- Sendek A, Karakoç C, Wagg C et al (2019) Drought modulates interactions between arbuscular mycorrhizal fungal diversity and barley genotype diversity. Sci Rep 9:9650
- Sharda KW, Rodrigues BF (2008) Ecology of arbuscular mycorrhizal fungi associated with Carica papaya. In agro-based ecosystem of goa, india Trop. Subtrop. Agroecosystems 8:265–278
- Sharma MP, Adholeya A (2000) Enhanced growth and productivity following inoculation with indigenous AM fungi in four varieties of onion (Allium cepa L.) in an alfisol. Biol Agric Hortic 18(1):1–14
- Sharma MP, Adholeya A (2004) Effect of arbuscular mycorrhizal fungi and phosphorus fertilization on the post vitro growth and yield of micropropagated strawberry grown in a sandy loam soil. Can J Bot 82:322–328
- Sharma MP, Gaur A, Bhatia NP, Adholeya A (1996) Growth responses and dependence of Acacia nilotica var. cupriciformis on the indigenous arbuscular mycorrhizal consortium of a marginal wasteland soil. Mycorrhiza 6(5):441–446
- Sharma MP, Grover M, Chourasiya D, Bharti A, Agnihotri R et al (2020) Deciphering the role of trehalose in tripartite symbiosis among rhizobia, arbuscular mycorrhizal fungi, and legumes for enhancing abiotic stress tolerance in crop plants. Front Microbiol 11:509919
- Sharma MP, Jaisinghani K, Sharma SK et al (2012) Effect of native soybean rhizobia and am fungi in the improvement of nodulation, growth, soil enzymes and physiological status of soybean under microcosm conditions. Agric Res 1(4):346–351. [https://doi.org/10.](https://doi.org/10.1007/s40003-012-0038-2) [1007/s40003-012-0038-2](https://doi.org/10.1007/s40003-012-0038-2)
- Sharma MP, Sandeep S, Sharma S, Aketi R, Bhatia (2016) Coinoculation of resident AM fungi and soybean rhizobia enhanced nodulation, yield, soil biological parameters and saved fertilizer inputs in Vertisols under microcosm and field conditions. Soybean Research 14:39–53
- Sharmah D, Jha DK (2014) Diversity of arbuscular mycorrhizal fungi in undisturbed forest, slash-and-burn field, and monoculture forest of Indo-Burma megadiverse region. Braz J Bot 37:339–351. [https://](https://doi.org/10.1007/s40415-014-0075-0) [doi.org/10.1007/s40415-014-0075-0](https://doi.org/10.1007/s40415-014-0075-0)
- Singh BK, Bardgett RD, Smith P, Reay DS (2010) Microorganisms and climate change: terrestrial feedbacks and mitigation options. Nat Rev Microbiol 8:779–790
- Singh PK (2012) Role of glomalin related soil protein produced by arbuscular mycorrhizal fungi: A review. Agric Sci Res J 2:119–125
- Smith S, Read D (2008) Mycorrhiza symbiosis, 3rd edn. Academic Press, San Diego
- Smith SE, Jakobsen I, Grønlund M, Smith FA (2011) Roles of arbuscular mycorrhizas in plant phosphorus nutrition: interactions between pathways of phosphorus uptake in arbuscular mycorrhizal roots have important implications for understanding and manipulating plant phosphorus acquisition. Plant Physiol 156:1050–1057
- Socolow RH (1999) Nitrogen management and the future of food: Lessons from the management of energy and carbon. Proc Natl Acad Sci, USA 96:6001–6008
- <span id="page-14-0"></span>Solaiman ZM (2014) Contribution of arbuscular mycorrhizal fungi to soil carbon sequestration. In mycorrhizal fungi: Use in Sustainable Agriculture and Land Restoration (Editor ZM Solaiman). Soil Biology 41. [https://doi.org/10.1007/978-3-662-45370-4\\_18](https://doi.org/10.1007/978-3-662-45370-4_18) Springer-Verlag Berlin Heidelberg
- Song YY, Zeng RS, Xu JF, Li J, Shen X (2010) Interplant communication of tomato plants through underground common mycorrhizal networks. PLoS One 5:e13324
- Staddon A, Gregersen R, Jakobsen I (2004) The response of two Glomus mycorrhizal fungi and a fine endophyte to elevated atmospheric CO2, soil warming and drought. Global Change Biol 10:1909–1921
- Sun X, Su Y, Zhang Y et al (2013) Diversity of arbuscular mycorrhizal fungal spore communities and its relations to plants under increased temperature and precipitation in a natural grassland. Chin Sci Bull 58:4109–4119
- Tarnawski S, Aragno M (2006) The influence of elevated  $CO<sub>2</sub>$  on diversity, activity and biogeochemical function of rhizosphere and soil bacterial communities. In: Nösberger J, Long SP, Norby RJ et al (eds) Managed ecosystems and  $CO<sub>2</sub>$ -case studies, processes and perspectives. Ecological studies series, Vol 187. Springer, Berlin, pp 393–409
- Tchabi A, Coyne D, Hountondji F, Lawouin L, Wiemken A, Oehl F (2008) Arbuscular mycorrhizal fungal communities in sub-Saharan savannas of Benin, West Africa, as affected by agricultural land use intensity and ecological zone. Mycorrhiza. 18:181–195
- Tchabi A, Coyne D, Hountondji F, Lawouin L, Wiemken A, Oehl F (2010) Efficacy of indigenous arbuscular mycorrhizal fungi for promoting white yam (Dioscorea rotundata) growth in West Africa. Appl Soil Ecol 45(2):92–100. [https://doi.org/10.1016/j.apsoil.2010.](https://doi.org/10.1016/j.apsoil.2010.03.001) [03.001](https://doi.org/10.1016/j.apsoil.2010.03.001)
- Treseder KK (2016) Model behavior of arbuscular mycorrhizal fungi: predicting soil carbon dynamics under climate change. Botany 94: 417–423
- Turrini A, Bedini A, Loor MB, Santini G, Sbrana C, Giovannetti M et al (2018) Local diversity of native arbuscular mycorrhizal symbionts differentially affects growth and nutrition of three crop plant species. Biol Fertil Soils 54:203–217. [https://doi.org/10.1007/s00374-017-](https://doi.org/10.1007/s00374-017-1254-5) [1254-5](https://doi.org/10.1007/s00374-017-1254-5)
- Valdenegro M, Barea JM, Azcon R (2001) Influence of arbuscularmycorrhizal fungi, Rhizobium melilotistrains and PGPR inoculation on the growth of Medicago arborea used as model legume for revegetation and biological reactivation in a semi-arid mediterranean area. Plant Growth Regul 34:233–240
- Vályi K, Mardhiah U, Rillig MC, Hempel S (2016) Community assembly and coexistence in communities of arbuscular mycorrhizal fungi. ISME J 10(10):2341–2235
- Van der Heijden MGA (2008) The unseen majority: soil microbes as drivers of plant diversity and productivity in terrestrial ecosystems. Ecol Lett 11:651
- Van der Heijden MGA, Klironomos JN, Ursic M et al (1998) Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. Nature 396(6706):69–72
- Velázquez MS, Fabisik JC, Abarca CL et al (2018) Colonization dynamics of arbuscular mycorrhizal fungi (AMF) in Ilex paraguariensis crops: Seasonality and influence of management practices. J King Saud Univ Sci. <https://doi.org/10.1016/j.jksus.2018.03.017>
- Vieira LC, Silva DK, Escobar IEC, Silva JM, Moura IA, Oehl F, Silva GA (2020) Changes in an arbuscular mycorrhizal fungi community along an environmental gradient. Plants 9(1):52. [https://doi.org/10.](https://doi.org/10.3390/plants9010052) [3390/plants9010052](https://doi.org/10.3390/plants9010052)
- Veneklaas EJ, Lambers H, Bragg J et al (2012) Opportunities for improving phosphorus-use efficiency in crop plants. New Phytol 195:306– 320
- Verbruggen E, Veresoglou SD, Anderson IC, Caruso T, Hammer EC, Kohler J et al (2013) Arbuscular mycorrhizal fungi - short-term

liability but long-term benefits for soil carbon storage? New Phytol 197:366–368

- Vilcatoma-Medina C, Kaschuk G, Zanette F (2018) Colonization and spore richness of arbuscular mycorrhizal fungi in Araucaria nursery seedlings in Curitiba, Brazil. Int J Agron:5294295–5294296. [https://](https://doi.org/10.1155/2018/5294295) [doi.org/10.1155/2018/5294295](https://doi.org/10.1155/2018/5294295)
- Wang J, Ma S, Wang GG et al (2020) Arbuscular mycorrhizal fungi communities associated with wild plants in a coastal ecosystem. J For Res 32:683–695. <https://doi.org/10.1007/s11676-020-01127-5>
- Wang W, Zhong Z, Wang Q, Wang H, Fu Y, He X (2017) Glomalin contributed more to carbon, nutrients in deeper soils, and differently associated with climates and soil properties in vertical profiles. Scientific Reports 7:1–13. <https://doi.org/10.1038/s41598-017->
- Wang C, Gu Z, Cui H et al (2015) Differences in Arbuscular Mycorrhizal Fungal Community Composition in Soils of three land use types in subtropical hilly area of southern China. PloS One 10:e0130983
- Wang J, Zhang J, Wang C et al (2021) Precipitation exerts a strong influence on arbuscular mycorrhizal fungi community and network complexity in a semiarid steppe ecosystem. Eur J Soil Biol 102: 103268. <https://doi.org/10.1016/j.ejsobi.2020.103268>
- Welbaum G, Sturz AV, Dong Z, Nowak J (2004) Fertilizing soil microorganisms to improve productivity of agroecosystems. Crit Rev Plant Sci 23:175–193
- Weltzin JF, Loik ME, Schwinning S, Williams DG et al (2003) Assessing the response of terrestrial ecosystems to potential changes in precipitation. Bio Science 53:941–952
- Wijayawardene NN, Hyde KD, Al-Ani LK, Tedersoo L et al (2020) Outline of Fungi and fungus like taxa. Mycosphere 11:1060–1456. <https://doi.org/10.5943/mycosphere/11/1/8>
- Wilson GWT, Harnett DC (1997) Effects of mycorrhizae on plant growth and dynamics in experimental tallgrass prairie. Am J Bot 84:478– 482. <https://doi.org/10.2307/2446024>
- Wolf J, Johnson NC, Rowland DL et al (2003) Elevated CO<sub>2</sub> and plant species richness impact arbuscular mycorrhizal fungal spore communities. New Phytol 157:579–588
- Worchel ER, Giauque HE, Kivlin SN (2013) Fungal symbionts alter plant drought response. Microb Ecol 65:671–678. [https://doi.org/10.](https://doi.org/10.1007/s00248-012-0151-6) [1007/s00248-012-0151-6](https://doi.org/10.1007/s00248-012-0151-6)
- Wright SF, Upadhyaya A (1998) A survey of soils for aggregate stability and glomalin, a glycoprotein produced by hyphae of arbuscular mycorrhizal fungi. Plant Soil 198:97–107
- Wu QS, He XH, Zou YN, Liu CY, Xiao J, Li Y (2012) Arbuscular mycorrhizas alter root system architecture of Citrus tangerine through regulating metabolism of endogenous polyamines. Plant Growth Regul 68:27–35
- Wu QS, Srivastava AK, Zou YN (2013) AMF induced tolerance to drought stress in citrus: a review. Sci Hortic 164:77–87
- Yamori W, Hikosaka K, Way DA (2014) Temperature response of photosynthesis in C3, C4, and CAM plants: temperature acclimation and temperature adaptation. Photosynth Res 119:101–117
- Yang Y, Li X, Kong X, Ma L, Hu X (2014) Transcriptome analysis reveals diversified adaptation of Stipa purpurea along a drought gradient on the Tibetan Plateau. Funct Integr Genomics 15:295–307
- Yang C, Hamel C, Schellenberg MP, Perez JC, Berbara RL (2010) Diversity and functionality of arbuscular mycorrhizal fungi in three plant communities in semiarid Grasslands National Park, Canada. Microb Ecol. 59(4):724–733. [https://doi.org/10.1007/s00248-009-](https://doi.org/10.1007/s00248-009-9629-2) [9629-2](https://doi.org/10.1007/s00248-009-9629-2) Epub 2010 Jan 15
- Zhang J, Tang X, He X, Liu J (2015) Glomalin-related soil protein responses to elevated  $CO<sub>2</sub>$  and nitrogen addition in a subtropical forest: Potential consequences for soil carbon accumulation. Soil Biol Biochem 83:142–149. <https://doi.org/10.1016/j.soilbio.2015.01.023>
- Zhang J, Wang F, Che R et al (2016) Precipitation shapes communities of arbuscular mycorrhizal fungi in Tibetan alpine steppe. Sci Rep 6: 23488. <https://doi.org/10.1038/srep23488>
- <span id="page-15-0"></span>Zhang R, Mu Y, Li X, Li S, Sang P, Wang X, Wu H, Xu N (2020) Response of the arbuscular mycorrhizal fungi diversity and community in maize and soybean rhizosphere soil and roots to intercropping systems with different nitrogen application rates. Sci Total Environ 740(20):139810. [https://doi.org/10.1016/j.scitotenv.](https://doi.org/10.1016/j.scitotenv.2020.139810) [2020.139810](https://doi.org/10.1016/j.scitotenv.2020.139810)
- Zhao H, Li X, Zhang Z et al (2017) Species diversity and drivers of arbuscular mycorrhizal fungal communities in a semi-arid mountain in China. PeerJ 5. <https://doi.org/10.7717/peerj.4155>
- Zhu X, Song F, Liu S, Liu F (2016) Arbuscular mycorrhiza improve growth, nitrogen uptake, and nitrogen use efficiency in wheat grown

under elevated CO<sub>2</sub>. Mycorrhiza. 26:133-140. [https://doi.org/10.](https://doi.org/10.1007/s00572-015-0654-3) [1007/s00572-015-0654-3](https://doi.org/10.1007/s00572-015-0654-3)

Zoppellari F, Malusa E, Chitarra W, Lovisolo C, Spanna F, Bardi L (2014) Improvement of drought tolerance in maize (Zea mays L.) by selected rhizospheric microorganisms. Italian J of Agrometeorol 19:5–18

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