



Mitigation of salinity stress in plants using plant growth promoting bacteria

Komal K. Bhise¹ · Padma B. Dandge²

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Abstract

Salinity is one of the major constraints prevailing in environment that affects not only plant growth but also agriculture productivity and soil fertility. Salinity stress causes nutritional and hormonal imbalance, ion toxicity, oxidative and osmotic stress and increased susceptibility of plant to diseases. Increasing use of chemical based fertilizers creates demand for environmental friendly and ecological compatible alternative in agriculture. Plant growth promoting bacteria (PGPB) have tremendous ability to mitigate salinity stress and ameliorate plant growth, playing vital role in food security by increasing agriculture crop productivity. PGPB inoculation under salinity promotes plant growth by several traits including 1-aminocyclopropane 1 carboxylate (ACC) deaminase activity, synthesis of plant hormones (such as indole acetic acid (IAA), gibberellic acid (GA), abscisic acid (ABA), cytokinin) and exopolysaccharides. PGPB alleviates salinity stress in plants by providing nutrients, maintaining high potassium and sodium ratio, increasing accumulation of osmolytes, enhanced photosynthesis and activity of antioxidant enzymes. The present review highlights and discusses current knowledge on effect of salinity stress on plant growth, PGPB mechanisms resulting in mitigation of salinity stress and increasing plant growth. It assesses morphological, biochemical, physiological and molecular changes induced in plants suffering from salinity when supplemented with PGPB.

Keywords Agriculture · Salinity stress · Environmental friendly · Plant growth promoting bacteria

1 Introduction

Stress is any external factor that has negative effects on plant growth and development (Foyer et al. 2016). One of the major abiotic stresses that limit the agricultural yield is salinity stress (Nadeem et al. 2016). The adverse effect of high concentration of minerals such as Na⁺ and/or Cl⁻ on plant growth is known as salinity stress (Munns 2005). Agriculture field is largest financial source for most of the world's population and soil salinity is a huge problem in soil under irrigation. The world's soil in hot and dry region is commonly saline and carries low agriculture potential, in such areas crops grows under irrigation condition, results in secondary soil salinization, affecting worldwide 20% of irrigated land (Glick et al. 2007a). In India salinity affected soil is about 7 billion hectares (Patel et al.

2011), in Maharashtra about 33,200 ha land is suffering from salinity and in case of Kolhapur district the different talukas are adversely affected by salinity (Naik 2014).

Salts are present as electrically charged ions in the soil due to soil weathering and insufficient rainfall (Shrivastava and Kumar 2015). Soil salinity is measured in terms of electric conductivity (EC) which is the capability of the substance to conduct electricity (AE 2001). The saline soil has EC of saturation extract above 4 dS m⁻¹ and 15% of exchangeable sodium. The total yield of many crops is reduced at this EC while many crops shows decrease in yield at the lower EC (Jamil et al. 2011). Around 40% of world's land has been affected by salinity (Jadhav et al. 2010). The problem of salinity is becoming more severe and by the 2050 an estimated of around 50% of land would be affected by salinity stress (Jamil et al. 2011). In addition every year salinity stressed soil is increasing by 10% because of several primary and secondary reasons. Salinity in soil may be natural that is primary and anthropogenic also known as secondary. The primary causes of salinity are, weathering of rocks, entry of brackish water in soil and ocean salt carried in wind and rains (Manchanda and Garg 2008). The secondary causes include, overuse of chemical fertilizers, industrial effluent entry in agricultural soil, use

✉ Padma B. Dandge
pbd_biochem@unishivaji.ac.in

¹ Department of Microbiology, Shivaji University, Kolhapur, Maharashtra 416004, India

² Department of Biochemistry, Shivaji University, Kolhapur, Maharashtra 416004, India

of low quality ground water for soil irrigation and irrigation without proper drainage system (Parihar et al. 2015). The other factors includes, soil amendments i.e., gypsum that results in increase in salt level of soil (Amacher et al. 2000).

The research on mitigation of salinity stress and soil pollution is prime important to increase crop productivity in order to feed worldwide increasing population. Escalating global hunger and salinity stress creates lucrative interest in sustainable environmental practices (Rashid et al. 2016). Intensive research is going on to ameliorate stress condition, improve plant productivity and protect the plants from phytopathogens using beneficial soil microorganisms known as PGPB (Berg and Martinez 2015; Egamberdieva et al. 2015; Bhise et al. 2017a, 2017b). PGPB promotes the plant growth by nitrogen fixation, phosphate and potassium solubilization as well as synthesis of plant hormones, siderophore, hydrogen cyanide, ammonia and ACC deaminase enzyme (Bhise et al. 2017a; Gontia-Mishra et al. 2017). Soil salinity is scourge for plant growth and crop productivity. Salinity stressed plants suffers from nutritional imbalance, ions toxicity, high osmotic and oxidative stress, reduced photosynthetic rates and decreased productivity. In the view of severity of salinity and PGPB importance in agriculture field the present review discuss the effect of salinity on plant growth status, the key role of PGPB in mitigating salinity stress in plants and promoting its growth and productivity.

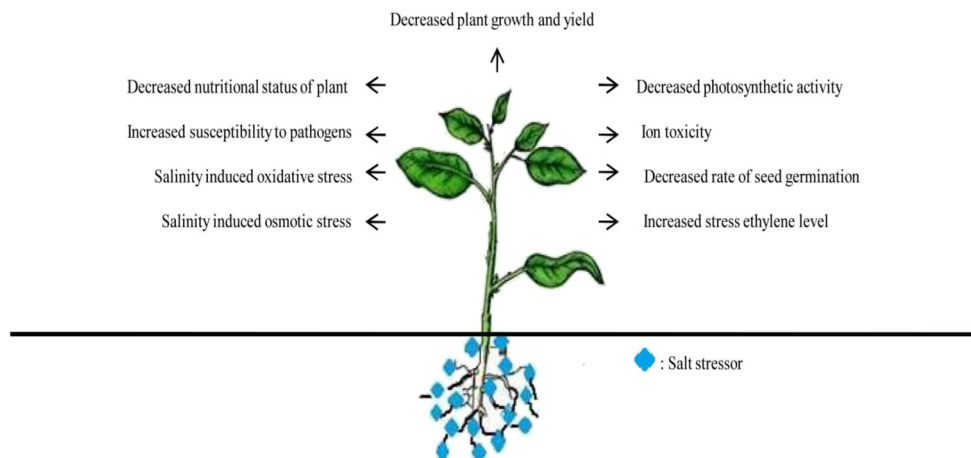
2 Effects of salinity stress on plant growth

Salinity stress affects morphological, biochemical and physiological processes of plant growth. Different crops exhibit different response to salinity; mostly all crops shows growth however total yield is reduced (Singh and Chatrath 2001). The overall effect of salinity stress on plant growth is shown in Fig. 1. Excess of salt in the soil leads to accumulation of soluble salt in the root zone which results in osmotic pressure and disturbs the growth of plants by affecting water and ions

uptake (Tester and Davenport 2003). The most vital phases of plant growth cycle is seed germination and seedling growth (Kaveh et al. 2011) which determines the total yield of crops. Generally low salt concentration induces dormancy state and reduces germination (Khan and Weber 2006), while high salt increases time required for germination and decreases percentage of germination (Rouhi et al. 2011; Ansari and Zadeh 2012). Salinity stress results in accumulation of high concentration of Na^+ and Cl^- which affects other vital elements presence and can decrease plant ability to access as well uptake minerals plus essential nutrients and its distribution in plants (Carmen and Roberto 2011), resulting in nutritional imbalance which reduces physiological activity and plant growth (Moradi and Ismail 2007). The accumulated Na^+ in plant tissue increases formation of reactive oxygen species (ROS) such as superoxide (O_2^-), single oxygen ($^1\text{O}^2$), hydroxyl radical (OH^\cdot), hydrogen peroxide (H_2O_2) (Ahmad et al. 2011) and inhibit photosynthesis. ROS causes lipid peroxidation, protein oxidation, DNA damage, inactivation of enzymes (Islam et al. 2015) plus chlorophyll degradation (Verma and Mishra 2005). Salinity results in accumulation of sodium and chlorine ions in chlorophyll and chloroplast hence photosynthesis is inhibited as these are important components in photosynthesis (Zhang et al. 2005). In salinity condition the considerably reduced efficiency of photosystem II, CO_2 assimilation rate and electron transport chain has been noticed by Stepien and Klobus (2005), Photosystem II is relatively sensitive to salinity stress (Allakhverdiev et al. 2000). Plants exposed to salinity stress produce an increases level of ethylene or stress hormone which actually causes damages to plants (Abeles and Heggestad 1973). Presence of environmental stresses is the major issue that disturbs rhizosphere functioning and influence agriculture productivity (Singh 2015).

Agriculture soil in arid and semi-arid regions is reduced by 1–2% every year by salinity stress (Kafi and Khan 2008). About 70% reductions in the yield of main crops such as wheat, rice, maize and barley are reported (Acquaah 2007), which is found to be not enough to deal with food demand of

Fig. 1 Effect of salinity on plant growth



worldwide increasing population in 2050 (Ray et al. 2013). The costs related to salinity stress are enormous, estimated at US\$12 billion per year and it is increasing (Qadir et al. 2008; Dodd and Pérez-Alfocea 2012). In order to meet food demand of worldwide increasing population, around 50% increase in yield of key crops is crucial (Godfray et al. 2010).

3 Plant growth promoting bacteria

PGPB are present in the soil zone surrounding the plant root known as rhizosphere (Walker et al. 2003). Plant rhizosphere is rich in PGPB due to presence of root exudates, which acts as attractant for number of organisms present in soil (Singh and Strong 2016). Plant rhizosphere is helpful in facilitating nutrients and water to plants and provides continuous benefits to microbes present in rhizosphere, which promotes plant growth. The plant and microbe interaction in rhizosphere determines plant growth and soil fertility. Plant provides nutrients to PGPB in return PGPB endorse growth of plant through several plant growth promoting traits (Patel et al. 2015).

3.1 Role of PGPB in salinity stress amelioration in plants

The main challenge in agriculture field is development of technologies especially for stressed soil which endow agriculture sustainability and increase crop yield (Gepstein and Glick 2013; Hamilton et al. 2016). Currently there has been interest in the use of beneficial organism known as PGPB in the agriculture field, which improves plant growth and performance through several physiological, biochemical and molecular pathway (Palacios and Bashan 2014).

Under non stress condition PGPB enhance the plant growth by common mechanisms however in stress condition some PGPB may be not able to perform efficiently because of the incapability to survive in stress condition. However some PGPB are well known for amelioration of many stresses including salinity stress (Forni et al. 2017). The improved plant growth by PGPB under stress condition is due to the some mechanisms such as lowering stress induced ethylene level by ACC deaminase activity, (Glick et al. 2007a; Saharan and Nehra 2011), improving plant nutrition through nitrogen fixation, phosphate solubilization and siderophore synthesis (Etesami and Beattie 2017; Etesami 2018), by synthesizing exopolysaccharides that decrease accumulation of sodium ions in plant root by binding to excessive sodium ions and preventing their translocation to plant leaves (Qin et al. 2016; Etesami and Beattie 2017), upregulating plant defense enzymes (catalase (CAT), glutathione reductase (GR), superoxide dismutase (SOD), ascorbate peroxidase (APX)) to have protection against oxidative stress (Islam et al. 2016; Qin et al. 2016), maintaining high K^+/Na^+ ratio by regulating

expression of ion transporters hence protects against ion toxicity (Islam et al. 2016; Etesami 2018; Pérez-Alfocea et al. 2010), increasing accumulation of osmolytes such as proline, glycine betain and sugars (Creus et al. 2004), preserving high photosynthetic activity and stomatal conductance (del Amor and Cuadra-Crespo 2012) and altering hormonal status of plant (Arora et al. 2012).

Promotion of salinity stressed plant growth by plant growth promoting traits of PGPB is shown in Fig. 2. Amelioration of salinity stress and alleviation of plant growth has been observed in many plants after PGPB inoculation such as wheat (Egamberdieva 2009), rice (Nautiyal et al. 2013), cotton (Yao et al. 2010), oat (Sapre et al. 2018), chickpea (AbdAllah et al. 2018), chili pepper (Park et al. 2013), licorice (Egamberdieva et al. 2016), mung beans (Patel et al. 2015), mint (Bharti et al. 2014) and chinese cabbage (Lee et al. 2016). Table 1 shows development of salt tolerance in crops inoculated with PGPB with several plant growth promoting traits.

3.1.1 Reduction in stress ethylene level

Ethylene is gaseous plant hormone, widely known as stress hormone and produced by all plants (Mayak et al. 2004). Ethylene plays a key role in plant growth and causes physiological changes in plant at molecular level at low concentration (Glick et al. 2007b) and is regulated by biotic and abiotic stress factors (HarDOIM et al. 2008). An elevated level of ethylene is synthesized when plants are exposed to different stress condition that causes increased damages to plants (Abeles and Heggstad 1973). The ethylene level is elevated under stress condition due to enhanced production of ACC, an immediate precursor of ethylene biosynthesis (Zapata et al. 2007). For maintaining normal plant growth, reduction in stress-induced ethylene is necessary that will alleviate effect of salinity on plant growth (Glick 2004; Etesami and Beattie 2017).

PGPB facilitate the plant growth in saline condition by producing key enzyme known as ACC deaminase which catabolise ACC to ammonia and α -ketobutyrate (Glick et al. 2007a). The reduced level of ACC results in lowering stress ethylene concentration in plant root vicinity and helpful for promoting plant growth. Although salinity stress has been associated with loss of ACC deaminase activity in some PGPB (Upadhyay et al. 2009) however some PGPB maintains ACC deaminase activity even in the presence of salinity and helps the plants to tolerate stress by reducing stress ethylene level (Mayak et al. 2004). Bhise et al. (2017a) isolated plant growth promoting salt tolerant organism *Enterobacter cloacae* strain KBPD from salt affected soil of Kolhapur, MS, India. Following inoculation of *V. radiata* by strain KBPD, significant improvement in plant growth was observed even in presence of 50, 100 and 150 mM NaCl stress. Similarly *Hartmannibacter diazotrophicus* E19T exhibited ACC

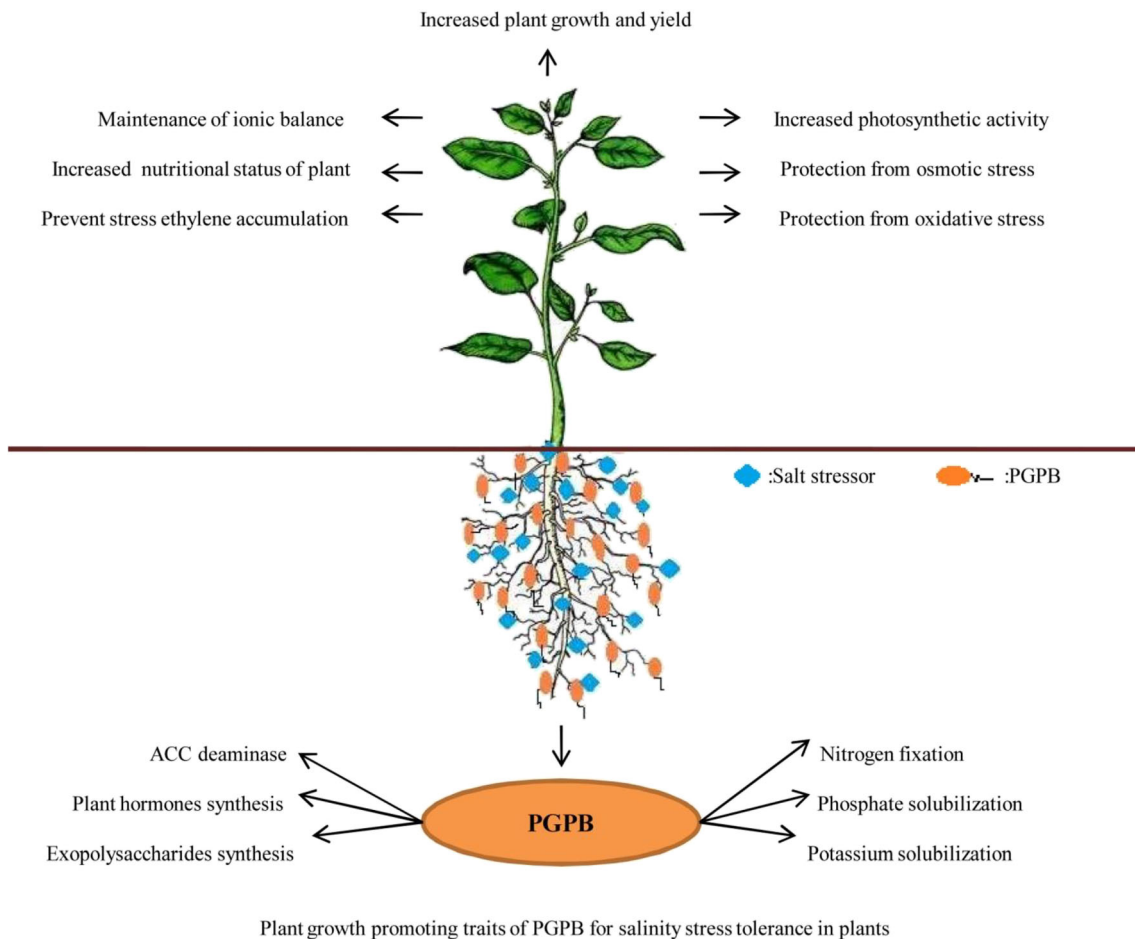


Fig. 2 Salinity stress alleviation using PGPB

deaminase activity, isolated from salinity affected rhizosphere of *Plantago winteri*. When strain E19T added to 200 mM salinity stressed barley seedlings a decreased ethylene emission and significant improved root, shoot dry weight and water contents in root was observed (Suarez et al. 2015). Likewise, *P. fluorescens* with ACC deaminase activity when added to cucumber plant suffering from salinity stress (7 and 10 dS m⁻¹) alleviation in salinity condition was noticed (Nadeem et al. 2016). Furthermore *Klebsiella* sp. strain IG 3 with expression of ACC deaminase activity and IAA production in salinity stress, when inoculated to oat (*Avena sativa*) under NaCl stress (100 mM) considerable improvement in shoot and root length, shoot and root dry weight and relative water content were observed (Sapre et al. 2018). Furthermore stress ethylene level increased under salinity decreases seed germination rate and plant root development (Belimov et al. 2001; Saravanakumar and Samiyappan 2007). Inoculation of salinity affected canola seeds (*Brassica napus* L.) by ACC deaminase producing *P. fluorescens* and *P. putida* significantly increased rate of germination (Jalili et al. 2009) and promoted plant growth. Salinity amelioration in plants by PGPB with

ACC deaminase activity has been reported (Gontia-Mishra et al. 2014; Gontia-Mishra et al. 2017; Singh et al. 2015; Siddikee et al. 2011, 2012; Bacilio et al. 2016).

3.1.2 Maintenance of nutritional and ionic balance

Salinity affects plant growth by nutritional imbalance as it increases sodium and decreases calcium and potassium ions uptake. Maintenance of sodium ion exclusion and potassium ion acquisition has been in relation with salt tolerance in plants (Hauser and Horie 2010). Inoculation of salinity suffering plant by *B. subtilis* can down regulate high affinity potassium ion transporters (HKT1) that reduces excessive absorption of sodium ions by plant root thereby maintains ionic status of plant even in stress (Qin et al. 2016). Furthermore, these PGPB triggers HKT1 induction in inoculated plant shoot thereby facilitating recirculation of sodium ions from shoot to root (Zhang et al. 2008). A reduced sodium ion uptake was noticed in plant tissue when supplied with *B. subtilis* by regulating HKT1 which controls uptake of sodium ions (Zhang et al. 2008). *Bacillus subtilis* GB03 produces volatile

Table 1 Development of salt tolerance in crops inoculated with PGPB having different plant growth promoting traits

Name of organism	Plant growth promoting traits	Plant	Effect of bacteria on plant growth	Reference
<i>Pantoea agglomerans</i> strain KL	ACC deaminase activity, phosphate solubilisation, Ammonia and exopolysaccharide production	Rice	Increased root and shoot length, fresh and dry weight, chlorophyll content, reduction in proline and MDA at 100 mM salinity stress	Bhise and Dandge 2019
<i>Enterobacter sp.</i> P23	ACC deaminase activity, phosphate solubilisation, IAA, siderophore and HCN production	Rice	Increase in root and shoot length, fresh and dry weight, chlorophyll content, total protein, total sugar, decrease in proline, MDA and ethylene level at 150 mM salinity	Sarkar et al. 2018
<i>Curtobacterium albidum</i> SRV4 strain	ACC deaminase activity, nitrogen fixation, IAA, HCN and exopolysaccharide production	Paddy plant	Increase in plant height, dry weight, chlorophyll content, carotenoid content, antioxidant enzyme activity, potassium ion uptake and decrease in sodium ion uptake at 100, 200 and 300 mM salinity	Vimal et al. 2018
<i>Enterobacter sp.</i> EN-21	ACC deaminase activity	Sugarcane	Increase in plant length, fresh and dry weight, chlorophyll content and potassium ion uptake and reduction in proline, MDA and sodium ion uptake at 200 mM salinity	Kruasuwan et al. 2018
<i>Rhodopseudomonas palustris</i> G5	Nitrogen fixation, phosphate and potassium solubilization and IAA, 5-aminolevulinic acid synthesis	Cucumber	Increase in shoot and root length, fresh and dry weight, total chlorophyll, soluble sugar content, SOD, POD and PPO activities and decrease in MDA at 4% salt stress	Ge and Zhang 2019
<i>Enterobacter cloacae</i> strain KBPD	ACC deaminase activity, phosphate solubilisation, IAA, siderophore, ammonia, HCN and exopolysaccharide production	<i>Vigna radiata</i>	Increase in root and shoot length, fresh and dry weight, chlorophyll content and reduction in proline at 50, 100 and 150 mM salinity	Bhise et al. 2017a
<i>Chryseobacterium gleum</i> sp. SUK	ACC deaminase activity, IAA, siderophore, ammonia, hydrogen cyanide production	Wheat	Improvement in root and shoot length, fresh and dry weight, chlorophyll, proteins, amino acids, phenolics, flavonoids and potassium ion content and decreased level of proline and sodium ion at 100 mM salt stress	Bhise et al. 2017b
<i>Enterobacter cloacae</i> ZNP-3	ACC deaminase, phosphate solubilization, IAA, HCN production	Wheat	Increase in root and shoot length, fresh and dry weight, chlorophyll content and potassium ion uptake and reduction in MDA, sodium ion uptake and H ₂ O ₂ and O ₂ ⁻ at 150, 175 and 200 mM salinity stress	Singh et al. 2017
<i>Pseudomonas. Fluorescens</i> 002	ACC deaminase activity and IAA production	Maize	Increase in root length, number of lateral root, dry root mass at 150 mM salinity	Zerrouk et al. 2016
<i>Pseudomonas fluorescens, Bacillus megaterium</i> and <i>Variovorax paradoxus</i>	ACC deaminase activity, IAA, siderophore and exopolysaccharide production	Cucumber	Increase in root and shoot length, total biomass and chlorophyll content at salinity stress of 7 and 10 dS m ⁻¹	Nadeem et al. 2016
<i>Serratia sp.</i> SL-12	ACC deaminase activity, phosphate solubilisation, IAA, siderophore and HCN production	Wheat	Increase in shoot and root length, fresh and dry weight, chlorophyll content, total soluble sugar, potassium and calcium ion uptake, decrease in sodium ion uptake, proline and MDA at 150, 175 and 200 mM salinity	Singh and Jha 2016a
<i>Bacillus cereus</i> Pb25	ACC deaminase activity, phosphate solubilisation, IAA and siderophore production	Mung beans	Increase in fresh and dry weight of shoot, root and nodule, chlorophyll content, potassium, phosphorous, nitrogen level and reduced sodium ion and MDA level at salinity stress of 9 dS m ⁻¹	Islam et al. 2016
<i>Paenibacillus xylanexedens</i> and <i>Enterobacter cloacae</i>	ACC deaminase activity and IAA production	Canola	Increase in root length at 100 mM salinity	Yaish et al. 2015
<i>Pseudomonas fluorescens</i> YsS6 and <i>Pseudomonas migulae</i> 8R6	ACC deaminase activity	Tomato	Increase in shoot fresh and dry weight, chlorophyll content, decrease in sodium ion content at 165 and 185 mM salt stress	Ali et al. 2014
<i>Streptomyces sp. strain</i> PGPA39	ACC deaminase activity, Phosphate solubilisation and IAA production	Tomato	Increase in dry plant biomass, chlorophyll content and reduction in proline content at 180 mM salinity	Palaniyandi et al. 2014
<i>Bacillus pumilus</i> strain DH-11	ACC deaminase activity, phosphate solubilisation, IAA and siderophore production	Potato	Increase in plant length, photochemical efficiency and antioxidant enzyme activity at 200 mM salinity	Gururani et al. 2013
<i>Rhizobium</i> and <i>Pseudomonas</i>	ACC deaminase activity and IAA production	Mung beans	Increase in total dry matter of seedling and salt tolerance index of plant at salinity stress of 6 and 4 dS m ⁻¹	Ahmad et al. 2013

organic compounds which prevent excessive accumulation of sodium ions in plant over control by regulating HKT1 in plant shoot and root exposed to salinity stress (Yang and Kloepper 2009). The salt tolerance in *B. napus* is may be due to the, increased activity of plasma membrane Na⁺/H⁺ exchanger ensuring enhanced sodium ion extrusion from plant root, activation of H⁺-ATPase and maintenance of negative membrane potential ensuring increased potassium retention and

decreased sensitivity of K⁺-permeable channels of plant root to ROS (Chakraborty et al. 2016). A very strong correlation between salinity tolerance in plants and K⁺ retention have been reported for plants such as *Brassica* species (Chakraborty et al. 2016), *Hordeum vulgare* (Wu et al. 2015) and *Triticum aestivum* (Cuin et al. 2012). Supplementation of *Azotobacter chroococcum* reported to decrease sodium and increase calcium, magnesium and potassium ion level in

plants (Rojas-Tapias et al. 2012). Similarly in other reports also decrease in sodium and increase in potassium ion concentration was noticed in plants upon PGPB inoculation (Bano and Fatima 2009; Kohler et al. 2009). The PGPB supplemented salinity stressed plants maintains adequate level of potassium ions to alleviate salinity originated sodium ion toxicity (Wang et al. 2016). A remarkably increased level of P, N, K⁺, Mg²⁺ and Ca²⁺ has been noticed in roots and leaves of radish when seeds were pretreated with *Pseudomonas fluorescens* and *Bacillus subtilis* (Mohamed and Goma 2012). Inoculation of salt affected strawberry plant by *Kocuria erythromyxa* EY43 alleviates detrimental effects of salinity stress by increasing uptake of micro and macronutrients such as Mg, Mn, S, Cu, Fe, P, K and N plus decreases sodium ion concentration in root and leaves (Karlidag et al. 2013). An increase in phosphate contents of salinity stressed wheat plants was noticed when supplied with *B. aquimaris* (Upadhyay and Singh 2015). PGPB with nitrogen fixation, phosphate and potassium solubilization ability helps the plants in mobilization of these nutrients from soil and makes accessible form of nutrients available for luxuriant plant growth even under stress condition.

Some PGPB have an ability to assist the plant growing in salt stressed environment from detrimental effects of accumulated sodium ions by producing exopolysaccharides (Upadhyay and Singh 2015; Kasim et al. 2016). The produced exopolysaccharides binds to Na⁺ and reduces its availability, resulting in low sodium ions uptake hence maintains high K⁺/Na⁺ ratio (Upadhyay and Singh 2015). Inoculation of *B. subtilis* strain GB03 improves K⁺/Na⁺ in white clover (*Trifolium repens* L.) root and shoot in presence of 50, 100 and 150 mM salinity (Han et al. 2014). Similarly the exopolysaccharide synthesis by *Enterobacter* sp. P23 plays an important role in reducing sodium ion concentration in rice seedlings by binding to excess Na⁺ thus alleviate salinity stress (Sarkar et al. 2018). These results corroborates with Upadhyay et al. (2011) and Ashraf et al. (2004) for the wheat crops. Furthermore the PGPB produced exopolysaccharide binds to soil particles and forms macroaggregates plus microaggregates and enhance salinity stress resistance because of the improvement in soil structure (Sandhya et al. 2009). Exopolysaccharide production helps the inoculated PGPB to adhere to plant root which helps to exert beneficial effect of PGPB on plant and makes the PGPB as an ideal inoculum to salt suffering soil (Bhise et al. 2017a).

3.1.3 Protection from salinity induced oxidative stress

Salinity affected plants produces ROS which damages nucleic acid, proteins and lipids and are generated by plasma membrane, cell wall, peroxisome, mitochondria and chloroplast (Das and Roychoudhury 2014). The prolonged salinity decreases ROS scavenging enzymes such as CAT, SOD and

APX and increases lipid peroxidation (Halo et al. 2015). Salt tolerance can be developed in plants by increasing activity of antioxidant enzymes (Hernández et al. 2003). Bacterization of salinity affected plant by PGPB mitigates stress by increasing ROS scavenging enzyme activity (Bianco and Defez, 2009). Kadmiri et al. (2018) showed increase in activity of POD of wheat under 5% salt stress when bacterized with *P. fluorescens* Ms-01 and *P. fluorescens* Ms-01 + *A. brasilense* DSM1690 in comparison with uninoculated plants, indicating role of PGPB in protecting plants against salinity stimulated oxidative stress. Moreover increase in enzyme activity of CAT and POD were reported by Islam et al. (2016) when 9 dS m⁻¹ salt stressed *Vigna radiata* supplied with *Bacillus cereus* Pb25. Similar results of difference in antioxidant enzyme activity of salt stressed plants and bacterized salt stressed plants were reported by Kohler et al. (2010) for lettuce plants supplied with *Pseudomonas mendocin*, Gururani et al. (2013) for *Solanum tuberosum* added with *Bacillus* strains and Egamberdieva et al. (2017) in tomato plants with *P. extremorientalis* TSAU20. The increase in enzyme activity of ROS scavenging enzyme in salt stressed plants with PGPB inoculation may be because of the PGPB effect on antioxidant enzyme encoding genes (Gururani et al. 2013).

Lipid peroxidation causes increased level of malondialdehyde (MDA), indicates damaged structural integrity of cell membrane however bacterial inoculation to salinity stressed plant reduces MDA accumulation and assist the plant to recover from salinity induced toxic effects (Wu et al. 2014). Furthermore, PGPB supplementation to salinity affected plants enhances level of defensive enzymes and phenol which develops resistance in plants against salinity (Sharma and Sharma 2017). A decrease in MDA level was noticed by Sarkar et al. (2018) in salinity stressed rice seedlings in presence of bacterial inoculation of *Enterobacter* sp. P23. Similarly, *Pseudomonas pseudoalcaligenes* promotes growth of salinity affected rice GJ-17 by reducing lipid peroxidation (Jha and Subramanian 2014). These results corroborates with Singh and Jha (2016a) who reported reduced MDA level in wheat plants upon inoculation of *Serratia* sp. SL-12 and Sapre et al. (2018) in oat plant upon *Klebsiella* sp. IG 3 inoculation. The lower malondialdehyde content in bacterized salinity affected plants was reported by other studies also (Bharti et al. 2014; Barnawal et al. 2014; Jha and Subramanian 2014).

3.1.4 Protection from salinity induced osmotic stress

In salinity condition soil-water potential changes which decreases water uptake by plant. Under such condition plant accumulates compatible solutes such as proline, trehalose, glycine betaine and some organic solutes, playing vital role in plant protection by osmotic adjustment that enhances water uptake, diluting toxic ion concentration and limiting water

loss (Ashraf et al. 2013; Slama et al. 2006). Proline accumulation in salt stress protects the plants by osmotic pressure adjustment and stabilization of plant functional units like proteins, enzymes and electron transport system-complex II (Ashraf and Foolad 2007; Mäkelä et al. 2000). Egamberdieva et al. (2017) observed increased level of proline when salt stressed plants were added with *P. extremorientalis* TSAU20. The salinity stressed *Acacia gerrardii* inoculated with *Bacillus subtilis* enhanced the proline level resulting in salt tolerance by maintaining water balance in plant tissue (Hashem et al. 2016). This result corroborates with Kruasuwan and Thamchaipenet (2018) for sugarcane in presence of *Enterobacter* sp. EN-21. The *Bacillus pumilus* and *P. pseudoalcaligenes* inoculation to rice plants increased concentration of glycine betaine and develops salt tolerance (Jha et al. 2011). Furthermore addition of *Dietzia natronolimnaea* strain STR1 confers salt tolerance in wheat plants and exhibited elevated level of proline and antioxidants, enabling wheat plant to survive in stress (Bharti et al. 2016).

3.1.5 Improvement in photosynthesis

Salinity tolerance in plants is positively correlated with photosynthetic contents. Inoculation of salinity affected plants with PGPB increases photosynthetic activity and improves plant growth (Kang et al. 2014a). Salinity suffering plant having decreased synthesis of photosynthetic pigment when supplemented with *Bacillus subtilis* increased synthesis of chlorophyll a, b and carotenoid was observed (Abeer et al. 2015). Increase in chlorophyll content in salinity stressed plants provided with PGPB inoculum is reported in *Arachis hypogaea* (Shukla et al. 2012), *Mentha arvensis* (Bharti et al. 2014), *Avena sativa* (Sapre et al. 2018), *Vigna radiata* (Bhise et al. 2017a), *Triticum aestivum* (Singh and Jha 2016b) and sugarcane (Kruasuwan and Thamchaipenet, 2018). The PGPB inoculation to plants stimulates photosynthetic pigment synthesis might because of the increased uptake of nitrogen, phosphorus and potassium. Dawwam et al. (2013) demonstrated mitigation in salinity induced toxicity on photosynthetic machinery of *C. arietinum* by inoculation with *B. subtilis* through promotion of pigment synthesis and its associated components (Abd Allah et al. 2018). Similarly chlorophyll a contents of cotton plant exposed to 0.7% salinity stress was less than non stressed plants while inoculation with *K. oxytoca* Rs-5 significantly improved chlorophyll a concentration. This increase may be because of the increased plant biomass as dry matter accumulation was significantly increased so the leaf biomass (Wu et al. 2014). Inoculation of 150 mM NaCl stressed chickpea (*Cicer arietinum*) with ACC deaminase and IAA producing *Pantoea dispersa* PSB3 ameliorate salinity stress by reducing sodium ion uptake, electrolyte leakage and improving potassium ion uptake and chlorophyll content (Panwar et al. 2016).

3.1.6 Regulation of phytohormones

Salinity stress disturbs plant hormone balance hence hormone homeostasis under salinity condition might be the possible mechanism involved in phytohormone-originated tolerance in plants (Alqarawi et al. 2014; Iqbal and Ashraf 2013). The ability of plant to acclimatize to stress depends on their interaction with microbes that has IAA, GA and cytokinin producing ability (Berg et al. 2013). Phytohormones regulate tolerance in plants against salinity (Ryu and Cho 2015) and develop plant protective response to stress (Raghavan et al. 2006). These phytohormones affect proliferation of cell in root system and increases water and nutrient uptake due to overproduction of root hairs and lateral roots (Arora et al. 2013). Salinity stress reduces IAA content in plants thus causes stomatal closure (Dunlap and Binzel 1996) and disruption in cell wall extension and plasticity (Ribaut and Pilet 1994). PGPB with IAA production ability stimulate endogenous synthesis of IAA hence compensate salinity induced IAA reduction in plants (Liu et al. 2013). Kadmiri et al. (2018) investigated role of IAA producing *A. brasilense* DSM1696 and *P. fluorescens* Ms-01 on salinity stress alleviation in wheat crops. Author showed significant improvement in plant height and weight in presence of 5% salinity stress when added with studied organisms indicating role of IAA in plant growth promotion. Furthermore, IAA produced by PGPB stimulates ACC deaminase activity through signalling cascade that hydrolyze ACC into α -ketobutyrate and ammonia (Glick 2005) hence help the plant grow under salinity stress by reducing stress ethylene level. *P. aureantiaca* TSAU22 and *P. extremorientalis* TSAU20 with IAA production ability when inoculated to *Sylebum marianum* mitigates salinity stress (Egamberdieva et al. 2013). Asim et al. (2013) noticed reduced IAA level in salinity affected soybean seedling while PGPB inoculation improved phytohormone level in plant. IAA changes root morphology resulting in improved plant growth and yield. *A. brasilense*, a salt tolerant organism isolated from salinity affected soil of Algeria near Mediterranean coast produced IAA even in salt stress. Supplementation of durum wheat plants by *A. brasilense* ameliorated salinity stress and promoted plant growth (Nabti et al. 2007, 2010). Sarkar et al. (2018) demonstrated rice seed germination of 48% in 150 mM salinity stress and 76% in same salinity stress when inoculated with IAA producing *Enterobacter* sp. P23, indicating alleviation in salinity stress in rice seed due to bacterial inoculation. The role of IAA in salinity stress tolerance and plant growth promotion has been identified in several studies (Ali and Abbas 2003; Kaya et al. 2013, 2010; Saba et al. 2013).

Salinity stress reduces GA synthesis in plants while PGPB inoculation triggers endogenous level of GA (Shahzad et al. 2016) and activates salinity stress mitigation strategies to prevent damages (Kang et al. 2014b). Gibberellic acids influences many processes of plant growth and development includes cell

division, cell elongation, stem growth, meristem size of root and leaf (Guo et al. 2015; Martínez et al. 2016) fruit setting and seed germination (Hedden and Phillips 2000). GA mitigates salinity induced adverse effects on water use efficiency and water plant relationship (Yamaguchi 2008). GA application improved water use efficiency in tomato plants in saline soil (Maggio et al. 2010). In addition, inoculation of salinity affected soyabean by GA producing *Pseudomonas putida* H-2-3 mitigates salinity stress (Kang et al. 2014c). Under salinity, GA increases magnesium and nitrogen level in root and leaves (Tuna et al. 2008). GA ameliorates salinity stress in mung beans by increasing protein synthesis, reducing sugars and activity of antioxidant enzymes and by reducing ribonuclease activity (Mohammed 2007). An increased endogenous level of GA in PGPB such as *Acinetobacter calcoaceticus* SE370, *Burkholderia cepacia* SE4 and *Promicromonospora* spp. SE188 inoculated cucumber plants over control plants improved plant growth under salinity (Kang et al. 2014a). IAA and GA producing *Bacillus subtilis* and *Pseudomonas fluorescens* when used for pretreatment of radish seed results in increase in salinity tolerance (Mohamed and Gomaa 2012). PGPB with IAA and GA producing ability promotes plant growth and seed germination under salinity (Kang et al. 2014a, 2014b) by enhancing α -amylase activity in seed, resulting in starch solubilization into simple sugars (Kim et al. 2006).

There are few reports on the role of bacterial synthesized ABA on its status in plant under salinity stress. However PGPB changes ABA status of plants and signaling pathways mediated by ABA that may results in enhanced salt tolerance in plants. Bharti et al. (2016) showed salinity tolerance in wheat plants by changing ABA signaling cascade when inoculated with *Dietzia natronolimnaea* STR1. In addition auxin and ABA production by *Bacillus amyloliquefaciens* RWL-1 induce salinity tolerance in rice plant (Shahzad et al. 2017). ABA mitigates inhibitory effect of salinity on plant growth due to accumulation of calcium and potassium, proline and sugar in root that helps to neutralize inhibitory effects of sodium and chlorine ions (Jaschke et al. 1997; Popova et al. 1995). Halotolerant bacteria with phytohormones such as IAA, GA and ABA producing ability when inoculated to soyabean plants alleviated salinity stress and improved shoot and root length and dry biomass under salinity condition (Naz et al. 2009). Research work carried out on IAA, GA and ABA showed that these hormones mitigated salinity stress and improved plant growth when PGPB *P. chlororaphis* and *P. extremorientalis* inoculated in salinity stressed common beans (Egamberdieva 2011; Jha and Subramanian 2013).

PGPB inoculation to salt stressed plant can manipulate cytokinins concentration of plant by shifting cytokinins homeostasis or by producing cytokinins (Glick 2012; Pallai et al. 2012; Kapoor and Kaur 2016). An increased level of cytokinin in plant shoot decreases ethylene level and leaf senescence

in tomato plant hence supports plant growth under salinity condition (Ghanem et al. 2010). In plant growth cytokinins acts as master regulator and known to mitigate salinity stress in plants (Fahad et al. 2015). However, the actual role of cytokinin in salinity tolerance in plant is not clear.

3.1.7 Proteogenomic level

Proteomic approach provides information regarding expression of whole proteins and their interaction in cell (Paul et al. 2006). Salt tolerant plants have altered activity of proteins. Studies on molecular mechanism regulating response of plants against salinity mainly focus on transcription changes (Brumós et al. 2009). Kandasamy et al. (2009) showed expression of 23 proteins during study of plant growth promotional property of *Pseudomonas fluorescens* strain KH-1 in rice leaf sheaths. Salt stressed plants and bacterized salt stressed plants express variation in antioxidative processes, response against pathogens and proteins of photosynthesis (Cheng et al. 2012). Salt stressed plants in presence of bacteria express increase in proteins related to deoxyribonucleic acid (DNA) synthesis hence protects *Arabidopsis* plants from salinity (Arimura et al. 2004). Furthermore, plants inoculated with PGPB provide protection against biotic and abiotic stress by increasing chitinase and 20S proteasome activity (Cheng et al. 2012).

Gene expression and their role in salt tolerance can be studied by transcriptomic analysis. Genomic and transcriptomic are used to characterize number of transcription factor and genes. Supplementation of *Burkholderia phytofirmans* PsJN in medium increases genes such as ROS scavenging (APX2), responsive-to-dessication 29B (RD29B) and relative-to-dessication 29A (RD29A) and detoxification (Glyoxalase 7) and reduces lipoyxygenase-2 in salinity affected *Arabidopsis* plants (Pinedo et al. 2015). Similarly *Medicago truncatula* inoculation by bacteria alleviates salinity stress by reducing ethylene-signaling genes expression (Bianco and Defez, 2009). PGPB inoculation increases gene expression encoding for APX, CAT, DHAR (Dehydroascorbate reductase) and GR (Glutathione reductase) (Habib et al. 2016).

3.2 Advantages of using PGPB

Agriculture field provides food security to worldwide increasing human population. Now farmers are facing major problems such as increasing soil salinity stress, declined soil fertility and overall crop yield. Improvement in soil fertility and crop yield is main aim of most of the countries. PGPB present in soil acts as powerful agents in maintaining soil fertility, enhancing plant growth and nutrient acquisition, constituting a key resource for sustainable agricultural practices. The term bio-fertilizer mostly refers to use of PGPB to increase the

nutrient contents of plants by combination of various activities (Whipps 2001).

Use of PGPB as biofertilizer has several advantages in agriculture field which includes, 1) PGPB as a bioinoculant restore and reclaim wasteland by converting nonfertile soil into fertile soil 2) Reduces the use of chemical based fertilizers that consequently leading to reduce the production cost. The usual annual production cost of nitrogen fertilizer is USD 30 million that would be reduced to 1 million by the replacement of chemical fertilizers by biofertilizers (Herridge 2008). 3) Decreases the pollution problems and land degradation arises because of the use of chemical based fertilizers and constitutes environmentally friendly approach (Stefan et al. 2008). 4) Maintains cation exchange capacity and structure of soil. 5) Significantly improves plant growth, quality and yield by controlling growth of phytopathogens thus ensures food security. Other advantages include safety, targeted activity, effective in small amount and self multiplication.

4 Conclusion

Salinity is one of the major constraints for agriculture yield, economy and food security. Salinity affects plant growth at morphological, biochemical, physiological and molecular level. Now it's inevitable to have promising agriculture practice that helps plant to withstand salinity stress and also increases agricultural productivity. PGPB can be considered as ecofriendly and cost effective approach to combat salinity stress and improve crop yield. Under salinity condition, ACC deaminase secretion by PGPB reduces stress ethylene accumulation and significantly improves plant growth. PGPB promotes the plant growth by enriching soil with vital nutrients such as nitrogen, phosphorous and potassium. Salinity induced decrease in hormones level is compensated by secretion of plant hormones such as IAA, GA, ABA and cytokinin by PGPB. Exopolysaccharide produced by PGPB maintains ionic balance of plant by preventing transport of accumulated sodium to plant root by binding with the excess sodium ions. Bacterization of salt stressed plants by PGPB enhances antioxidants, which alters redox status of plants thus improving photosynthesis efficiency as well as accumulate compatible solutes such as proline, which avoids stress induced damages by regulating cellular activities. The present reviews focal point is key role of PGPB in salt stress tolerance and promotion of plant growth and development. Considering this scenario future research is needed to develop bacterial consortia to saline soil having multiple plant growth promoting traits expressing PGPB. Understanding of interaction between PGPB present in consortia and consortia plus plant

will pave means to exploit much more benefits from PGPB for escalating salinity stress mitigation in plants.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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