ORIGINAL ARTICLE



# **Valorization of pineapple rind for bromelain extraction using microwave assisted technique: optimization, purifcation, and structural characterization**

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**Abstract** Huge amount of waste is generated by the pineapple processing industries which raises concerns regarding its safe disposal into the environment. This ever-increasing problem of waste management can be solved by the valorization of pineapple by-products to high-value compounds. The extraction of proteolytic enzyme, bromelain from pineapple rind using green techniques can help to overcome the drawbacks associated with conventional methods. In the present study, the extraction of bromelain from pineapple rind using microwave assisted technique resulted in considerable amount of proteolytic activity (127.8 U/mL) and protein content (2.55 mg/mL). The optimized extraction conditions were found as 200 W microwave power, 1:5 solid/ liquid ratio and after treatment time of 10 min. Highest specifc activity (512 U/mg) of bromelain was obtained after using gel fltration chromatography. FTIR result confrmed the presence of functional groups in bromelain, whereas, XRD analysis indicated the semi-crystalline nature of bromelain. The results indicated MAE as an efective green technique for the extraction of bromelain from pineapple rind. The proteolytic action of the extracted bromelain makes it a suitable functional ingredient for its applications in bakery, dairy, and seafood processing industries.

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**Keywords** Bromelain · Pineapple rind · Microwave assisted extraction · Valorization · Green techniques

### **Abbreviations**

- MAE Microwave assisted extraction
- RSM Response surface methodology
- BBD Box-Behnken design
- SEM Scanning electron microscopy
- XRD X-ray difraction
- FTIR Fourier transform infrared spectroscopy

# **Introduction**

Pineapple is a perennial herb known as *Ananas comosus* and belongs to the botanical family Bromeliaceae. After citrus, bananas and mangoes, the most important tropical fruit produced globally is pineapple in terms of its production and yield. The leading producers of pineapple are Thailand, Brazil, Costa Rica, China, Indonesia, Philippines, India, Nigeria, Colombia, Mexico (FAOSTAT [2020](#page-10-0)). Four varieties of pineapple are available around the world including Abacaxi, Smooth Cayenne, Queen, and Red Spanish (Huang et al. [2021\)](#page-10-1). Pineapple has been utilized in fruit processing industries as a main ingredient in fruit juice concentrates, jellies, squash, jams, essence, and pickles. Besides having an acceptable taste and favor, pineapple is rich source of essential nutrients, vitamins, minerals, dietary fber, and, antioxidants. The rich composition of pineapple in nutritive substances provides various health benefts like prevention of colon cancer, immunomodulatory, anti-infammatory, and antimicrobial activity.

Pineapple fruit is composed of 30% pulp, where, 70% of it is discarded in the form of peel, crown, core, trimmings, and residue (Nakthong et al. [2017\)](#page-11-0). Therefore, a large amount

of waste is generated by the pineapple processing industries which is often discarded into the environment without any further treatment. This may lead to serious environmental hazards due to emission of greenhouse gases during decomposition. This instigated for the valorization of pineapple by-products for its utilization as a source of proteins, and other bioactive compounds including a valuable proteolytic enzyme, bromelain. The bioactive compounds have been gaining importance for the development and commercialization of nutraceuticals and innovative functional food products, especially in the post-pandemic era (Galanakis [2023](#page-10-2)). The food ingredients and active compounds have been popularly known for health-promoting actions such as anti-infammatory, anti-microbial, and antiviral effects (Galanakis et al. [2020\)](#page-10-3).

Since pineapple is considered as the popular source of the protease enzyme, bromelain, it is known to be of the most important tropical fruit (Ketnawa et al. [2010](#page-10-4)). Bromelain is found in the rind, leaves, stem, crown, tubes, and bark of pineapple fruit (Mohan et al., [2016\)](#page-11-1). Extensive applications of bromelain have been reported in pharmaceutical, food, and detergent industries. Some of the important areas of utilization of bromelain in the food industries include meat tenderization, baking, brewing, and, protein hydrolysates production (Ketnawa et al. [2012\)](#page-10-5). Along with this, action of bromelain results in gentle peeling of skin, therefore, it has been used as an active ingredient in cosmetic products (Ramli et al., [2017](#page-11-2)).

The adopted extraction and purifcation strategy for bromelain should be cheap, high yielding and speedy (Gupta et al. [2004](#page-10-6)). In previous studies, the conventional approach for bromelain extraction involved cell disruption technique using homogenization and crushing of pineapple rind followed by fltration/centrifugation to obtain crude bromelain extract (Colletti et al. [2021](#page-10-7)). This process is time consuming, has high chances of presence of impurities, and results in lower extraction yield. Moreover, conventional isolation and purifcation techniques are considered obsolete due to their low purifcation potential (Soares et al. [2012\)](#page-11-3). These problems can be circumvented with the use of emerging green technologies like microwave-assisted extraction (MAE) (Yan et al. [2020](#page-10-8)). MAE is an innovative technology which involves the use of microwave energy causing molecular motion by ionic conduction and dipole rotation (da Rosa et al. [2019](#page-10-9)). MAE leads to increase in temperature and pressure which causes changes in cell structure and increased penetration of solvent into the sample matrix.

The present study was carried out using popular commercial variety of pineapple i.e., 'Kew, which is mainly grown (70%) around the world (TFNET News Compilation, [2016\)](#page-11-4). Extraction of crude bromelain from pineapple rind was done using MAE technique. Further, optimization, purifcation (using dialysis and gel fltration chromatography method) as well as structural characterization was also carried out.

#### **Materials and methods**

## **Materials**

The pineapple rind was procured from the fruit processors in local markets of Longowal, Punjab, India. The pineapple rind was then dried using cabinet dryer (40 $\pm$ 1 °C) for 6 h and grounded to make fne powder. The pineapple rind powder was stored in the refrigerator (4 °C) till further analysis. All the chemicals and enzymes used in this experimental work were of analytical grade.

#### **Proximate analysis**

The proximate analysis of pineapple rind powder was carried out to analyze moisture, protein, fat, crude fber, ash content following the standard protocol (AOAC [2007\)](#page-10-10). Kjeldahl method was used for determination of nitrogen content and multiplied with factor 6.25 for conversion to protein. The available carbohydrates were determined by subtraction of percentage sum of other proximate constituents from 100.

## **Microwave assisted extraction of crude bromelain**

#### *Extraction using distilled water*

Preparation of crude bromelain extract from pineapple rind powder was carried out using microwave assisted extraction method (2450 MHz, Samsung Model MS23F301TAK/ TL, Malaysia) and distilled water as the solvent (Ketnawa et al. [2012](#page-10-5)). The oven was modifed for condensation of vapours produced in the sample during extraction (Kashyap et al. [2022\)](#page-10-11). The pineapple rind powder (5 g) was added to distilled water (50 mL) and microwave treatment was given to it. The mixture was then centrifuged (5000 g, 10 min), filtered, and finally the supernatant was stored at  $20^{\circ}$  C.

#### *Extraction using ethanol*

Preparation of crude bromelain extract from pineapple rind powder was carried out using microwave assisted extraction with ethanol as solvent (Huang et al. [2021\)](#page-10-1). The pineapple rind powder (5 g) was added to ethanol (50 mL) and microwave treatment was given to it. The mixture was then centrifuged (5000 g, 10 min), fltered, and fnally the supernatant was stored at 20<sup>°</sup>C.

#### *Extraction using acetone*

Preparation of crude bromelain extract from pineapple rind powder was carried out using microwave assisted extraction with acetone as solvent (Huang et al. [2021](#page-10-1)). The pineapple rind powder (5 g) was added to acetone (50 mL) and microwave treatment was given to it. The mixture was then centrifuged (5000 g, 10 min), fltered, and fnally the supernatant was stored at 20° C.

#### **Extraction using phosphate bufer**

Preparation of crude bromelain extract from pineapple rind powder was carried out using microwave assisted extrac-tion with phosphate buffer as solvent (Gul et al., [2021](#page-10-12)). The pineapple rind powder (5 g) was added to acetone (50 mL) and microwave treatment was given to it. The mixture was then centrifuged (5000 g, 10 min), fltered, and fnally the supernatant was stored at 20° C.

## **Extraction using NaOH and HCl solution**

Preparation of crude bromelain extract from pineapple rind powder was carried out using microwave assisted extraction with diferent NaOH and HCl solutions (0.6 M) as solvent (Wen et al.  $2021$ ). The pineapple rind powder (5 g) was added to NaOH (0.6 M, 50 mL) and HCl solutions (0.6 M, 50 mL) and microwave treatment was given to it. The mixture was then centrifuged (5000 g, 10 min), fltered, and finally the supernatant was stored at  $20^{\circ}$  C.

Based on the preliminary trials, effect of solid to liquid ratio (1:3–1:10 g/mL), microwave power (100–300 W), and treatment time (5–10 min) was observed on the proteolytic activity and protein content of crude bromelain extracted from pineapple rind. The process optimization was carried out using response surface methodology.

# **Purifcation of crude bromelain extract**

#### *Ammonium sulphate precipitation*

Initial purifcation of crude bromelain was carried out using ammonium sulphate precipitation method (Gautam et al. [2010\)](#page-10-13). The process involved, pinch by pinch addition of ammonium sulphate salt  $(6.6 \text{ g})$  to crude extract  $(15 \text{ mL})$ with continuous stirring (45 min) on a magnetic stirrer and kept overnight at 4 °C. It was then centrifuged (10000 g, 10 min) and the collected pellet was re-dissolved in phosphate buffer  $(0.01 \text{ M}, 10 \text{ mL})$  which was utilized for subsequent analysis.

#### *Dialysis*

Crude bromelain extract after initial purifcation by ammonium sulphate precipitation was subjected to dialysis for further purifcation (Gautam et al. [2010\)](#page-10-13). The crude extract was dissolved in tris HCl buffer (10 mL, 10 mM) and placed in a dialysis bag. The bag was suspended in beaker (500 mL) containing phosphate bufer-NaCl solution (100 mM), the setup was kept in refrigerator and the solution was changed after every 6 h until there is no precipitation. The dialysis process was continued for almost 48 h and the obtained extract was stored for further analysis.

#### *Gel fltration chromatography*

Gel fltration chromatography was used for the purifcation of bromelain with Sephadex G-75 resin (Costa et al. [2014](#page-10-14)). The resin was packed into a glass column (200 mm long, 17.5 mm in diameter) and one and a half volume of column was equilibrated with acetate buffer. The resin was left in contact with the sample for one hour after application. Following an elution with acetate buffer (1 M, pH 4.5) at a flow rate of 0.7 mL/min, the eluent was collected and once no more protein was eluted, the column was washed using acetate buffer  $(5 \times 10^{-3} \text{ M}, \text{pH } 4.5)$ . After purification, the enzyme was kept frozen at -20 °C.

#### **Determination of bromelain activity**

The determination of bromelain activity was done using casein as substrate (Mala et al. [2021](#page-10-15)). The solution containing enzyme (500  $\mu$ L) was mixed with casein (0.65% (w/v), 2.5 mL) in potassium phosphate buffer (0.05 M, pH 7.5) and then kept in a water bath (37  $\degree$ C, 10 min). The reaction was stopped when trichloroacetic acid (2.5 mL, 110 mM) was added. This was followed by centrifugation (5000 g, 10 min) after 20 min and the obtained supernatant was then mixed with sodium carbonate solution (2.5 mL, 500 mM) and folin ciocalteau phenol reagent (500 µL, 20%, v/v). Incubation was done at room temperature for 30 min and the absorbance was red at 660 nm using UV–vis spectrophotometer (DR 5000, HACH Germany).

The proteolytic activity (units/mL) was calculated by using following formula:

 $\mu$ mol tyrosine equivalent released  $\times$  total volume of assay (in mL) Volume of enzyme used in mL time of assay in  $(min) \times volume$  used in calorimetric determination  $(mL)$ 

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## **Specifc activity and purifcation fold**

Specifc activity of bromelain was determined as the ratio of the total volumetric activity and protein concentration in the sample (Ye et al., 2021). The result of specifc activity was expressed as (U/mg) activity units per mg of protein (Coelho et al. [2013](#page-10-16)).

Specific activity (U/mg) =  $\frac{\text{Enzyme activity (U/mL)}}{\text{Concentration of protein (mg/mL)}}$ 

Purifcation fold of enzyme was calculated as ratio of specifc activity after purifcation to the initial specifc activity.

Purification fold  $=$   $\frac{\text{Specific activity after purification}}{\text{Initial specific activity}}$ 

## **Experimental design and data analysis**

The effect of various process variables (solid/liquid ratio 1:3–1:10, microwave power 100–300 W, and time 5–15 min) on proteolytic activity and protein content of bromelain extracted from pineapple rind was determined by Response Surface Methodology (RSM) using Box-Behnken design (BBD) (Table [1](#page-3-0)). Design Expert 11.0.7.1 software (Statease Inc., Minneapolis, USA) was used for data analysis and study of regression equation coefficients. Verification of model was done by determination of  $\mathbb{R}^2$ . Validation of model was done using analysis of variance (ANOVA) and

the model's ft value was determined by statistical testing of each term with F-values of  $p \le 0.05$ .

# **Structural characterization of purifed bromelain extract**

#### *Fourier transform infrared analysis*

Fourier transform infrared (FTIR) analysis was carried out for identifcation of the specifc components of the bromelain based on their functional groups. A difuse refectance accessory was used to carry out spectral analysis for all samples in an FTIR instrument (Perken Elmer Spectrum, RX-I, USA) for detection of characteristic chemical functional groups. The spectrum was then recorded with absorbance mode  $(4 \text{ cm}^{-1} \text{ resolution})$  in the range of 600–4000 cm<sup>-1</sup> (Nor et al. [2015\)](#page-11-6).

## *X‑ray difraction analysis*

Difraction pattern of bromelain was determined using X-ray difraction (XRD) analysis using PAN-analytic-X'pert PRO MRD (Almelo, Netherlands). At 40 kV and 25 mA tube current, measurements were done using Cu line as radiation source. Difraction angle ranging from 3.0 to 45.0 °2θ was used for scanning of the samples at the scanning rate of  $2^{\circ}$  min<sup>-1</sup> (Sharma and Sharma, [2018](#page-11-7)).

<span id="page-3-0"></span>**Table 1** Box-Behnken design matrix for extraction of crude bromelain from pineapple rind by using microwave assisted extraction



## **Results and discussion**

The proximate analysis of pineapple rind as well as extraction of bromelain from pineapple rind was carried out using microwave assisted extraction. The process optimization as well as purifcation and structural elucidation of the extracted bromelain was also performed which has been discussed in this section.

## **Proximate analysis of pineapple rind**

The results of proximate analysis of pineapple rind indicated moisture and carbohydrate content of  $86.66 \pm 0.07\%$  and  $18.65 \pm 0.22\%$ , respectively. The other proximate components included protein  $(3.98 \pm 0.015\%)$ , ash  $(8.62 \pm 0.33\%)$ , fat  $(0.98 \pm 0.03\%)$  and crude fiber  $(12.98 \pm 0.16\%)$ . The proximate analysis of pineapple rind obtained in the current study was comparable to the earlier reported value of pineapple rind (Upadhyay et al. [2010\)](#page-11-8). The fat, ash and fber content of pineapple rind found in the current study was higher than the fat  $(0.63 \pm 0.03\%)$ , ash  $(6.80 \pm 0.28\%)$  and crude fiber  $(13.96 \pm 0.14\%)$  content reported in previous studies on pineapple rind (Aruna [2019](#page-10-17)).

# **Efect of microwave assisted technique on the extraction of bromelain from pineapple rind**

Extraction of bromelain from pineapple rind was carried out using microwave assisted technique which has been presented in Fig. [1](#page-4-0). Extraction of bromelain from pineapple rind using microwave assisted extraction indicated the highest proteolytic activity (98.89 U/mL) and protein content (2.35 mg/mL) obtained using ethanol as the solvent. It was followed by HCl, distilled water, phosphate buffer and acetone and NaOH. In the previous studies on microwave assisted extraction of crude bromelain, the highest proteolytic activity and protein content was found to be 92.78 U/ mL and 2.55 mg/mL, respectively (Mala et al. [2021](#page-10-15)). Based on the preliminary trials, process optimization of microwave assisted extraction of bromelain from pineapple rind was done using response surface methodology. The diferent extraction parameters including solid/ liquid ratio (1:3 to 1:10), microwave power (100 to 300 W), and time (5 to 15 min) were evaluated to study their efect on proteolytic activity and protein content of bromelain. BBD was chosen to study the interactive efect of diferent process variables.

# **Extraction of crude bromelain from pineapple rind for the determination of proteolytic activity by using microwave assisted extraction**

MAE technique was used to extract crude bromelain from pineapple rind by varying process parameters (solid/liquid



<span id="page-4-0"></span>**Fig. 1 a** Proteolytic activity of crude bromelain extract obtained using MAE technique **b**. Protein content of crude bromelain extract obtained using MAE technique

ratio (1:3 to 1:10), microwave power (100 to 300 W), and treatment time (5 to 15 min)) for the determination of proteolytic activity. The interactive efects of extraction parameters involved in the microwave assisted extraction of bromelain from pineapple rind on the proteolytic activity were presented by response surface plots (Fig. [2\)](#page-5-0). A signifcant effect  $(p \le 0.05)$  of the extraction parameters (solid/liquid ratio, microwave power, and treatment time) was observed on the proteolytic activity of bromelain extracted from pineapple rind (Table [2a](#page-5-1)).

Interactive effect of solid/ liquid ratio and time on extraction of crude bromelain from pineapple rind has been depicted in Fig. [2](#page-5-0)a. The interactive efect of time and solid/ liquid ratio indicated that with an increase in treatment time from 5 to 10 min and solid/ liquid ratio from 1:3 to 1:5, an increase in proteolytic activity was observed (127.8 U/mL), however, with further increase in time  $(15 \text{ min})$  and solid/ liquid ratio (1:10), proteolytic activity was found to decrease (72.19 U/mL). This may be due to degradation of thermolabile compounds on exposure to microwave irradiation for longer time duration which resulted in reduced proteolytic activity (Doulabi et al. [2020](#page-10-18)).



<span id="page-5-0"></span>**Fig. 2** Interactive efect of (**a**) solid/liquid ratio and time; (**b**) power and solid/liquid ratio; (**c**) power and time on the proteolytic activity of bromelain obtained from pineapple rind using MAE

<span id="page-5-1"></span>**Table 2** ANOVA for response surface reduced quadratic model for proteolytic activity of bromelain extract obtained from pineapple rind



 $R^2$ =0.9886, Adjusted  $R^2$ =0.9740, Predicted  $R^2$ =0.8583, Adequate Precision=22.9004, Coefficient of variation  $(\%) = 6.28$ 

Interactive effect of power and solid/ liquid ratio on extraction of crude bromelain from pineapple rind has been depicted in Fig. [2](#page-5-0)b. The interactive effect of solid/liquid ratio and power indicated that with increase in microwave power from 100 to 200 W, and solid/ liquid ratio from 1:3 to 1:5, an increase in proteolytic activity (127.8 U/mL) was observed, however, with further increase in power (300 W) and solid/ liquid ratio (1:10), proteolytic activity was found to decrease (49.46 U/mL). This might be as the solid particles caused an uneven absorption of microwave radiation which led to lower proteolytic activity (Behere et al. [2020](#page-10-19)).

Interactive efect of power and time on extraction of crude bromelain from pineapple rind has been depicted in Fig. [2](#page-5-0)c. The interactive effect of treatment time and power indicated that with an increase in power from 100 to 200 W and treatment time from 5 to 10 min, an increase in proteolytic activity (127.8 U/mL) was observed, however, with further increase in power (300 W) and time (15 min), proteolytic activity (70.99 U/mL) was found to decrease. This may be due to degradation of the enzyme on prolonged exposure to microwave irradiation (Mandal and Mandal [2010](#page-11-9)).

# **Extraction of crude bromelain from pineapple rind for the determination of protein content by using microwave assisted extraction**

MAE technique was used for the extraction of crude bromelain from pineapple rind by varying process parameters (solid/liquid ratio (1:3 to 1:10), microwave power  $(100 \text{ to } 180 \text{ W})$ , and treatment time  $(5 \text{ to } 15 \text{ min})$  for the

<span id="page-6-1"></span>**Table 3** ANOVA for response surface reduced quadratic model for protein content of bromelain extract obtained from pineapple

rind

determination of protein content. The interactive effects of extraction parameters involved in the microwave assisted extraction of bromelain from pineapple rind on the protein content were presented by response surface plots (Fig. [3](#page-6-0)). A significant effect ( $p \le 0.05$ ) of the extraction parameters (solid/liquid ratio, microwave power, and time) was observed on the protein content of bromelain extracted from pineapple rind (Table [3](#page-6-1)).

Interactive effect of time and solid/ liquid ratio on extraction of crude bromelain from pineapple rind has been depicted in Fig. [3](#page-6-0)a. The interactive efect of solid/ liquid ratio and time indicated that with an increase in time from 5 to 10 min and solid/ liquid ratio from 1:3 to 1:5, an increase in protein content was observed (2.55 mg/mL), however, with further increase in time (15 min) and solid/ liquid ratio (1:10), protein content was found to decrease (1.84 mg/ml). This may be due to degradation in protein caused by prolonged microwave irradiation time and absorption of microwave radiation by the solid particles which lead to lower protein recovery (Mala et al. [2021](#page-10-15); Behere et al. [2020\)](#page-10-19).

Interactive effect of power and solid/ liquid ratio on extraction of crude bromelain from pineapple rind has been depicted in Fig. [3b](#page-6-0). The interactive efect of solid/ liquid ratio and power indicated that with an increase in power from 100 to 200 W and solid/ liquid ratio from 1:3 to 1:5, an increase in protein content (2.55 mg/ mL) was observed, however, with further increase in solid/liquid ratio (1:10) and power (300 W), protein content was found to decrease (1.77 mg/mL). This might be due to excessive swelling of the plant matrix caused by large amount of solvent, which



<span id="page-6-0"></span>**Fig. 3** Interactive efect of (**a**) solid/liquid ratio and time; **b** power and solid/liquid ratio; **c** power and time on the protein content of bromelain obtained from pineapple rind using MAE



 $R^2$ =0.9863, Adjusted  $R^2$ =0.9687, Predicted  $R^2$ =0.8865, Adequate Precision=21.2520, Coefficient of variation  $(\%) = 3.51$ 

absorbs more microwave energy and generate excessive heat. The additional heat may cause thermal degradation of extracted solutes (Behere et al., [2020\)](#page-10-19).

Interactive efect of power and time on extraction of crude bromelain extract from pineapple rind has been depicted in Fig. [3](#page-6-0)c). The interactive efect of time and power indicated that with an increase in power from 100 to 200 W and treatment time from 5 to 10 min, an increase in protein content (2.55 mg/ mL) was observed, however, with further increase in power (300 W) and time (15 min), protein content (1.95 mg/ mL) was found to decrease, which may be due to the degradation of protein at high microwave power. Similar effect of microwave power was observed in the previous studies on paprika where a decrease in the fnal protein content was observed at higher microwave power (Kiss et al., [2000\)](#page-10-20). The results of the optimization of diferent process variables (temperature, treatment time, and liquid to solid ratio) on the extraction of bromelain using microwave assisted extraction revealed that the highest proteolytic activity (127.8 U/ml) and protein content (2.55 mg/mL) of bromelain was obtained with 1:5 solid/liquid ratio, microwave power of 200 W after microwave irradiation time of 10 min. The present study involves lower solvent consumption (1:5) and lesser irradiation time (10 min) which makes the process more economical, whereas, higher value of solid to liquid ratio (1:20) and irradiation time (40 min) was observed in the previous reported studies on extraction of protein from pineapple peels (Bansod et al. [2023](#page-10-21)).

# **Purifcation of crude bromelain extracted from pineapple rind**

#### *Ammonium sulphate precipitation*

Purifcation of crude bromelain extracted from pineapple rind using ammonium sulphate precipitation resulted in proteolytic activity of 127.8 U/mL. Proteolytic activity of bromelain obtained in present study using ammonium sulphate (50% saturation) was found to be slightly higher than the previously reported value of proteolytic activity (120.05 U/mL) obtained using ammonium sulphate (60% saturation) precipitation method (Gul et al., 2021). This might be due to the diference in saturation range of ammonium sulphate used in the present study. Purifcation of enzyme using ammonium sulphate precipitation is a cost-efective procedure as it may help reduce the number of steps in purifcation of biomolecules (Soares et al. [2011](#page-11-10)).

#### *Dialysis*

Purifcation of crude bromelain extracted from pineapple rind using dialysis resulted in proteolytic activity of 150.29 U/mL. The proteolytic activity of bromelain obtained after purifcation using dialysis was higher as compared to that of crude bromelain activity. This might be as the structural integrity of the purifed bromelain was maintained by this purifcation as well as due to the presence of other proteins in the purifed enzyme extract (Bhagavathy et al., [2019](#page-10-22)).

#### *Gel fltration chromatography*

Gel fltration chromatography was carried out for the purifcation of crude bromelain using Sephadex G-75 resin. Purifcation of crude bromelain extracted from pineapple rind using gel fltration chromatography resulted in proteolytic activity of 36.01 U/mL and specifc activity of 512 U/mg with purifcation-fold of 10.26. Value of specifc activity (512 U/mg) reported in the present study was lower than the previously reported specifc activity (729.16 U/mg) of bromelain where Sephadex G-50 was used as resin in gel fltration chromatography performed in combination with ion-exchange chromatography (Ilyas et al., [2018](#page-10-23)).

# **Structural characterization of bromelain extract obtained from pineapple rind**

Structural characterization of purifed bromelain extract obtained from pineapple rind using microwave assisted extraction was carried out using Fourier transform infrared spectroscopy and X-ray difraction analysis which has been discussed in this section (Fig. [4\)](#page-8-0).

#### *Fourier transform infrared spectroscopy*

FTIR spectroscopy of control and purifed bromelain extract obtained from pineapple rind using microwave assisted extraction has been depicted in Fig. [4](#page-8-0)a-a'. Characteristic peaks of bromelain were observed at 3271.15 cm−1 (N–H stretching vibration of secondary amide), 2927.90 cm−1 (aliphatic C-H stretching), 1416.78 cm<sup>-1</sup> (C = C stretching), and 2928.01 cm−1 (methylene asymmetric C-H stretching) 1644.23 cm−1 (C=C stretching), and 923.75 cm−1 (third overtone C-H stretching). The wavelength in the range 3338- 3380 cm−1 spectrum showed the presence of N–H stretching vibrations. The bands near 1179–1149  $cm^{-1}$ , 1255– 1290 cm<sup>-1</sup>, and, 1517–1587 cm<sup>-1</sup>, were assigned to C-N stretching vibration frequencies (Sharma and Sharma, [2018](#page-11-7)). Similar peaks of bromelain were reported in previous studies (Nor et al. [2015](#page-11-6); Sharma and Sharma 2018; Soares et al. [2012](#page-11-3)).

#### *X‑ray difraction analysis*

X-ray diffraction (XRD) analysis of purified bromelain extract obtained from pineapple rind using microwave assisted extraction has been presented in Fig. [4](#page-8-0)b. The



<span id="page-8-0"></span>**Fig. 4** FTIR spectroscopy of bromelain (control), **a** FTIR spectroscopy of purifed bromelain extract obtained from pineapple rind using MAE **b**. XRD analysis of bromelain as contro, XRD analysis of purifed bromelain extract obtained from pineapple rind using MAE



**Fig. 4** (continued)

bromelain extract showed the presence of sharp peaks between 10–23° 2θ and 5–25° 2θ, respectively. The difused peaks displayed by difractogram indicated the amorphous behaviour of bromelain. Intense peaks were observed at 20°, 35° and 40° 2θ, whereas low intensity peaks were observed at 5° and 10° 2θ. The results indicated that bromelain had semi-crystalline nature with intense peaks at 18.72°, 19.54°, 20.34° and 21.04° 2θ and low intensity peaks observed at 12.47°, 16.18° and 23.82° 2θ (Sharma and Sharma, 2018).

# **Conclusion**

Microwave assisted extraction has been found as an efective green technique for the extraction of bromelain from pineapple rind with potential to be utilized at industrial level. The optimized extraction conditions which resulted in highest proteolytic activity (127.8 U/mL) and protein content (2.55 mg/mL) of bromelain extract were 1:5 solid/liquid ratio, 200 W microwave power and treatment time of 10 min. Purifcation after gel fltration chromatography resulted in highest specific activity (512 U/mg). FTIR analysis confirmed the presence of functional groups in bromelain, whereas, XRD analysis depicted semi-crystalline nature of extracted bromelain. The results indicated pineapple rind as a chief source of proteolytic enzyme, bromelain, and protein. The employment of MAE technique for the extraction of bromelain can be an efective valorization strategy for value addition of pineapple waste while maintaining sustainability of the process. The extracted bromelain fnds applications in several food industries for meat tenderization, making of protein hydrolysate, cheese, and fish sauce production.

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#### **Declarations**

**Confict of interest** The authors declare no confict of interest regarding the publication of this paper.

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