



Accelerated storage test of betalains extracted from the peel of pitaya (*Hylocereus cacti*) fruit

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Abstract Betalains are nitrogen-containing colorants with antioxidant properties that can be found in plant materials such as pitaya peels. However, thermo-stability of these natural colors may vary with different source, yet few study has reported the rate orders of degradation for pitaya-sourced betalains. In this study, accelerated storage test of betalains, namely betacyanin and betaxanthin, extracted from pitaya peel are investigated by heat treatment of the extract at elevated temperatures. The results show that degradation kinetics of betacyanins and betaxanthins can both fit first-order kinetics and Arrhenius equation with activation energies at -49.2 kJ/mol and -40.0 kJ/mol, respectively. The result of Student's *t*-test indicated that the predicted *k* values are statistically the same as compared to their corresponding experimental values. LSD estimation also showed that *k* value variation tendency of the two betalains appears to be the same at 60 °C or below, while betacyanins tend to degrade faster above 80 °C than betaxanthins due to higher coefficient value of *k* value variation. This result also suggests that the pitaya-sourced betalains tend to degrade gradually even though they are stored under refrigerated condition. However, the betalains showed appreciably lower rate of degradation if they are processed at 60 °C or below.

Keywords Betalains · Betacyanins · Betaxanthins · Arrhenius equation · Degradation kinetics

Introduction

The two categories of betalains, which include betacyanins and betaxanthins, have traditionally been used as food colorants for red-violet (betacyanins) and yellow-orange (betaxanthins) colors (Pavokovic and Krsnik-Rasol 2011). In recent decades, betalains have gained interest from scientific communities due to its health medicinal values (Zhang et al. 2013; Tesoriere et al. 2014; Khan 2016). Despite the health benefits, the thermal instability and molecular degradability drawbacks of natural colorants demonstrated in other studies (Chandran et al. 2014; Priatni and Pradita 2015; Güneser 2016; Rodriguez-Sanchez et al. 2017) also led to the importance of the investigations on the rate orders for betalain degradation.

Betalains, a group of nitrogen-containing colorants, can be extracted from various plant materials, including Caryophyllales, the flowers of Aizoaceae and Portulacaceae, red beetroots, and Cactaceae fruits such as pitayas (Delgado-Vargas et al. 2000; Slavov et al. 2013). In the past decades, betalains from different sources have been identified. The properties of betalains can vary with different plants in terms of betacyanin/betaxanthin ratio, water solubility, range of spectrum covered, and thermostability (Azeredo 2009). Hence, realizing the stability of betalains derived from each plant material is important while proposing a proper storage condition. In addition, the study reported by Rodríguez-Sánchez et al. (2015) also shows that, among the factors that cause molecular break down of betalains, storage temperature is one of the most critical ones.

In recent years, the properties of betalains extracted from the fruit of *Hylocereus cacti*, or pitayas, are found to be more promising than those from other sources (Azeredo 2009). Pitayas, or locally called dragon fruit, are cultivated

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in most tropical regions in the world, especially in Malaysia, Thailand, Vietnam, and Taiwan. Among the different genus of pitayas, white (*Hylocereus undatus*) and red pitayas (*Hylocereus polyrhizus*) are the most commonly grown (Merten 2003; Wiset et al. 2012). The peel of pitayas, representing approximately 18–24% of the whole fruit (Chuck-Hernández et al. 2016), can contain up to 150 mg of betacyanin per 100 g fresh sample (Jamilah et al. 2011). In practice, pitaya peels are discarded while the fruit pulps are processed or consumed. Therefore, recycling of pitaya peel for betalain extraction not only can reduce production cost of the health benefit colorants, but also create a higher economic value for the fruit. However, few studies have reported the rate orders for degradation of pitaya peel derived betalains.

Accelerated storage test (AST) has been used widely to estimate and predict shelf-lives of labile products in food, biology, and pharmaceutical industry (Franks 1994; Presa-Owen et al. 1995). Among the methods of AST, Arrhenius equation is most generally used in the valid assumption for deterioration (or degradation) rate of temperature-dependent compounds, components, or overall food products (King et al. 1998; Reyes and Cisneros-Zevallos 2007; Danisman et al. 2015; Yalçınöz and Erçelebi 2015; Ruiz-Gutiérrez et al. 2015; Priatni and Pradita 2015). Hence, by means of Arrhenius relationship, degradation kinetics of betalains extracted from pitaya peels can be predicted (Ruiz-Gutiérrez et al. 2015; Priatni and Pradita 2015).

In this study, AST of betalains extracted from pitaya peels were carried out to investigate the degradation kinetics of betacyanins and betaxanthins under different storage and processing temperatures. While predicting the rate constant (*k*) of betacyanins and betaxanthins at different storage temperatures, the predicted *k* values under refrigerated and ambient storage conditions were also compared with their corresponding experimental values.

Materials and methods

The procedures to investigate thermostability of betalains extracted from pitaya peels included handling of raw materials, extract preparation, heat treatment, sample analysis, calculation of betacyanin/betaxanthin content, and degradation kinetic studies of betacyanins and betaxanthins.

Handling of raw materials

Fresh pitaya peels from different sources were cleaned and cut into small pieces with an area of approximately 2 cm × 2 cm. The pitaya peel pieces were well-mixed and stored under − 18 °C for not more than 14 days.

Extract preparation

To prepare 10% pitaya peel extract, 100 g pitaya peel was added into a Waring Blender filled with 900 g deionized water and blended for 30 s. Pitaya peel extract was obtained after passing through a 200 μm filter.

Heat treatment

Twenty grams of the pitaya peel extract was added into a 100 mL blue cap bottle and placed within an oil bath at 70, 80, 90, 100, 110, and 120 °C for 1, 3, 5, 10, and 30 min, respectively. For extracts stored under refrigerated (4 °C) and ambient (25 °C) conditions, samples were taken every 2 h for 8 h in total. The contents of betacyanins and betaxanthins were then analyzed.

Sample analysis

Analysis of betacyanin and betaxanthin content was according to the method reported by Castellar et al. (2003), Stintzing et al. (2005), and Sumaya-Martinez et al. (2011) with some modification. After heat treatment, pitaya peel extract samples were centrifuged at 11,200×*g* for 2 min to remove the sediments. The samples were analysed by photospectrometer (Hitachi UV–Vis U-2800, Japan) at 535 nm and 480 nm respectively for analysis of betacyanin and betaxanthin content. Absorbance readings for all samples were made against distilled water as a blank.

Calculation of betacyanin/betaxanthin content

According to Beer Lambert's Law, betacyanin and betaxanthin content can be calculated by using the equation below (Ruiz-Gutiérrez et al. 2015; Priatni and Pradita 2015):

$$\begin{aligned} &\text{Betacyanin or Betaxanthin Content} \\ &= A \times MW \times 1000 \times DF / \epsilon \times l \end{aligned} \quad (1)$$

where *A* is the absorbance at 535 or 480 nm; *MW* is the molecular weight of betanin (550 g/mol) or indicaxanthin (308 g/mol); *DF* is the dilution factor of sample; ϵ is the extinction coefficient of betanin (60,000 L/mol cm) or indicaxanthin (48,000 L/mol cm); *l* is the pathway of light transmittance (1 cm). The contents of betacyanin and betaxanthin are expressed as mg betanin and indicaxanthin equivalent/L extract, respectively.

Degradation kinetic analysis

The degradation kinetics of betacyanin and betaxanthin content were described by fitting first-order kinetic model to the experiment data based on the equation below:

$$w_{(t)} = w_0 \times \exp^{-kt} \tag{2}$$

$$\ln(w_{(t)}/w_0) = -kt \tag{3}$$

where w_0 is the content of betacyanin and betaxanthin before heat treatment; $w_{(t)}$ is the content of betacyanin and betaxanthin at time t (min); k (min^{-1}) is the rate constant at the temperature determined; and t is the time of heat treatment.

Temperature dependence of betacyanin and betaxanthin degradations can be determined by calculating the activation energy (E_a) values based on Arrhenius equation (Chandran et al. 2014):

$$k_{(T)} = A \times \exp^{-E_a/RT} \tag{4}$$

$$\ln k_{(T)} = \ln A - \left(\frac{E_a}{R}\right) \left(\frac{1}{T}\right) \tag{5}$$

where $k_{(T)}$ is the k value (min^{-1}) at temperature T (K); A is the pre-exponential factor; R is the universal gas constant [8.314×10^{-3} kJ/(K mol)]; E_a is the activation energy (kJ/mol); and T (K) is the temperature of heat treatment.

Statistical analyses

Duplicate results were obtained for all experiments performed. Arithmetic means, standard deviations, correlation coefficients, Student t-test, and Fisher Least Significant Difference (LSD) were calculated using Microsoft Excel 2010 and Sigma Plot 10.0. Student t-test and Fisher LSD were respectively used to estimate statistical significance of the predicted k values and the correlation coefficients of betalain k values at different temperatures.

Results and discussion

Thermostability of betalains

The overall result in this study shows that betalains extracted from pitaya peels are found to be thermos-unstable. The content of betalains decreased with increasing time of heat treatment, while the degradation rate also enhanced at elevated temperatures. The degradation kinetics of betacyanins and betaxanthins extracted from pitaya peels are shown in Fig. 1.

Data shown in Fig. 1a, b indicated that degradation of pitaya-peel-based betacyanins and betaxanthins fits first-order kinetics during heat treatment from 70 to 120 °C. It was also found in the comparison between the degradation kinetics of betacyanins and betaxanthins that the k values for both components were approximately 0.008 at 70 °C. However, the value for betacyanin degradation increases 10 times (0.08) as the temperature increased to 120 °C. At

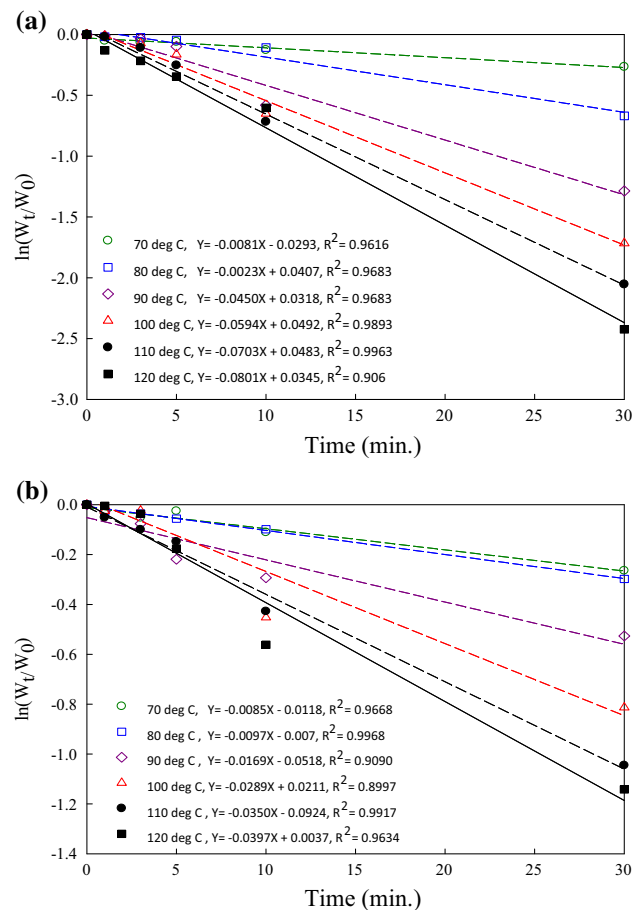


Fig. 1 Degradation kinetics of **a** betacyanins and **b** betaxanthins extracted from pitaya peels

the same temperature, k value for betaxanthins was 0.04, only 5 times increase. The result in Fig. 1 can therefore imply that thermal instability of betacyanins appeared to be higher than that of betaxanthins at elevated temperatures, namely 70–120 °C. The result can also be observed from the residual yellow color of pitaya peel extract after 30 min heat treatment (Table 1).

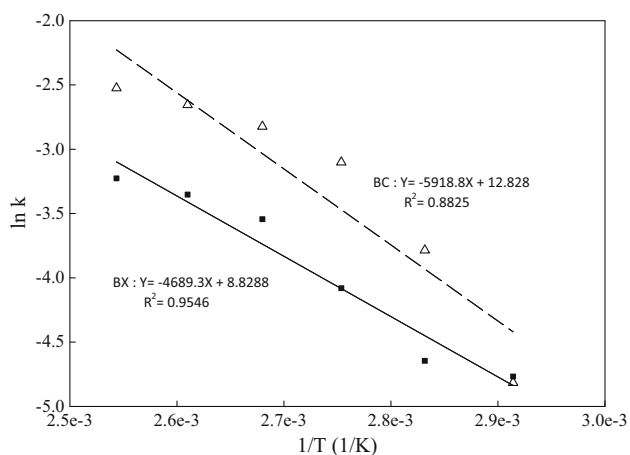
k values and activation energies

To predict the degradation kinetics of pitaya-peel-based betalains stored under different temperatures, regression lines of k values for betacyanins and betaxanthins found at different temperatures were created based on Arrhenius model, namely Arrhenius regression lines (Fig. 2).

In Fig. 2, the activation energies (E_a) for betacyanin and betaxanthin degradation are found to be -49.21 kJ/mol (or -11.76 kcal/mol) and -38.99 kJ/mol (or -9.32 kcal/mol), respectively. E_a for betacyanin found in this study appears to be lower as compared to that reported by Priatni and Pradita (2015), -14.37 kcal/mol, in the investigation for degradation kinetics of betacyanins

Table 1 Appearance of pitaya peel extract samples taken at different times (120 °C heat treatment)

10% pitaya peel extract						
Heat treatment time (min)	Fresh sample	1	3	5	10	30
BC content (mg betanin equivalent/L ext.)	23.39	20.54	18.83	16.54	12.80	2.07
BX content (mg indicaxanthin equivalent/L ext.)	8.02	7.98	7.73	6.72	4.57	2.56

**Fig. 2** Arrhenius regression lines for betacyanins (square) and betaxanthins (triangle)

extracted from red dragon fruit. In addition, the study reported by Chandran et al. (2014) related to heat treatment of beetroot to determine the degradation of visual color also indicated that betaxanthins can show higher stability at elevated temperatures than betacyanin and degradation of both pigments follow first order kinetics and Arrhenius model. In other words, the content of betacyanin and betaxanthin at different storage temperatures can be predicted based on the Arrhenius regression lines.

Predicted and experimental k value comparison

The predicted k values under refrigerated (4 °C) and ambient temperature (25 °C) storage condition of betacyanins and betaxanthins, have been compared with their corresponding experimental values. Statistical significance of these predicted and experimental values is estimated by Student's t-test ($P = 0.95$) as shown in Table 2.

Data presented in Table 2 showed that the predicted k values for storage of betacyanins and betaxanthins under refrigerated (4 °C) and ambient temperature (25 °C)

condition were within the confidence interval, which indicates that no statistically significant difference as compared with its corresponding experimental k value (t value $< t_{0.05}$). This can also designate that there was no significant difference between predicted and actual results at the storage temperatures and time intervals tested.

Variation of k values at different temperature

In Fig. 3a, b, regression coefficients of k value variation at different temperatures have been determined for betacyanin and betaxanthin degradation. It can be seen that the changes of first-order coefficient values for both betacyanin and betaxanthin degradation are both found at 60–80 °C. According to the estimation based on Fisher Least Significant Difference (LSD) at $P = 0.05$, the tendency of the k value variations for both betacyanins and betaxanthins are found to be identical at temperatures below 60 °C. The tendency can also be observed from the identical first-order coefficient ($1.00E-04$) shown in Fig. 3a, b. This also explains why color change can hardly be observed from the pitaya peel extract samples of which heat treatment were carried out at 70 °C (data not shown). Above 80 °C, however, the rates of k value increase appear to be approximately 7 (for betaxanthin) or 15 (for betacyanins) times higher, while betacyanins show appreciably higher growth than betaxanthins. This finding implies a higher color stability of the two colorants at 60 °C or below, suggesting a temperature limit of their application or process (e.g. drying, extraction, sterilization...etc.).

The tendency of k value variation below 60 °C can be attributed to reversibility of Schiff base reactions, which suggested by both Elbe et al. (1981) and Molina et al. (2014) in the stability study of red-beet derived betalains. Results for both studies indicated that the color maintenance of betalains due to degradation reversibility can mostly occur at temperatures below 66 °C (or below 60 °C found in this study). As for the tendency of k value

Table 2 Statistical significance of predicted and experimental k values estimated by student's t-test ($P = 0.95$)

	k value (min^{-1})		Confidence interval of the predicted k value ^a
	Predicted	Experimental	
Betacyanins (4 °C)	1.98E-04	2.04E-04	1.53E-04 to 2.55E-04
Betaxanthins (4 °C)	3.06E-04	2.95E-04	9.07E-05 to 4.99E-04
Betacyanins (25 °C)	8.90E-04	8.54E-04	6.03E-04 to 1.11E-03
Betaxanthins (25 °C)	1.01E-03	6.24E-04	1.33E-03 to 8.37E-05

^aPredicted k value falls within the confidence interval indicates no statistically significant difference as compared with its corresponding experimental k value (t value $< t_{0.05}$)

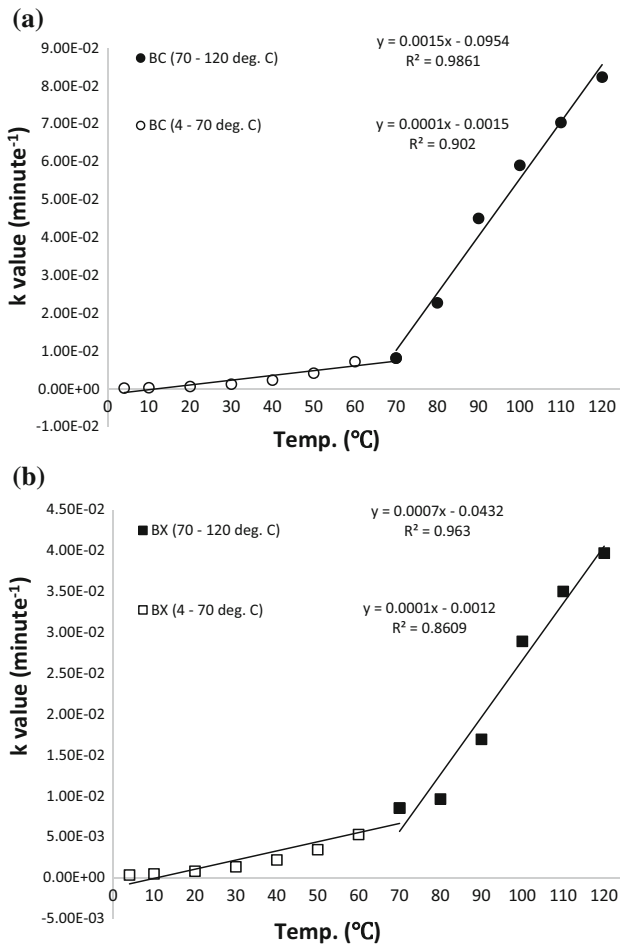


Fig. 3 Regression of k value variation (with temperature) for betacyanin (a) and betaxanthin (b) degradation. Statistical significance of k values for both betacyanins and betaxanthins at 4–120 °C were estimated based on Fisher Least Significant Difference (LSD) at $P = 0.05$

variation above 80 °C, the study of yellow pitaya peel color stability reported by Cejudo-Bastante et al. (2016) also agreed that betalains can suffer a pronounced degree of degradation at 80 °C or higher. The same tendency can also be observed from the data in Fig. 1, which shows that the k values (or slopes) for betacyanins and betaxanthins appear to be similar at 70 °C, while a more significant

difference of k values for the two categories of betalains is found at 80–120 °C.

The main cause of this situation can be due to reversibility and degradability of the broken down molecules of betalains, such as betalamic acid from betacyanins and betaxanthins, cyclo-Dopa from betacyanins, and amines from betaxanthins. These molecules can undergo Schiff base condensation which leads to betanin regeneration under a lower processing temperature. However, subsequent degradation or reaction, such as aldol condensation or Millard reaction, of these broken down molecules can mostly occur at higher processing temperatures (e.g. 80 °C or above found in this study), and hence, leading to loss of color (Woo et al. 2011). The overall finding of the above data also indicates that aqueous solution for both betacyanin and betaxanthin are highly unstable. However, thermos-degradation can be retarded significantly if they are stored under lower temperature.

Conclusion

Betacyanins and betaxanthins are two categories of valuable colorants that can be extracted easily from pitaya peels. Despite the thermos-unstable properties, their degradation fits first order kinetics and Arrhenius model with activation energies of -49.2 kJ/mol (betacyanin) and -40.0 kJ/mol (betaxanthin). In the comparison of predicted and experimental k values based on Student's t-test, no significant difference was found between betacyanins and betaxanthins, indicating that the predicted value can be designated as reliable. The regression coefficients of k value variation for betacyanins and betaxanthins showed that a transition was found at 60–80 °C. According to the result of LSD estimation, the tendency of the k value variations for the two colorants is found to be identical at temperatures below 60 °C. Above 80 °C, the regression coefficients appeared to increase significantly, while betacyanins is expected to be higher than that of betaxanthin, which implies that the colorants can be more stable and increased rate of degradation can be avoided if they are processed below 60 °C. Data presented in this study shows

that both pitaya-sourced aqueous betacyanins and betaxanthins can degrade gradually even though they are stored under refrigerated conditions, yet the appreciably reduced rate constant (k value) of degradation below 60 °C also suggests a tolerance limit of their processing temperature. To improve the stability of betalains, some countermeasures are to be investigated in future studies.

References

- Azeredo HMC (2009) Betalains: properties, sources, applications, and stability—a review. *Int J Food Sci Technol* 44:2365–2376
- Castellar R, Obon JM, Alacid M, Fernandez-Lopez JA (2003) Color properties and stability of betacyanins from opuntia fruits. *J Agric Food Chem* 51:2772–2776
- Cejudo-Bastante MJ, Hurtado N, Delgado A, Heredia FJ (2016) Impact of pH and temperature on the colour and betalain content of Colombian yellow pitaya peel (*Selenicereus megalanthus*). *J Food Sci Technol* 53(5):2405–2413
- Chandran J, Nisha P, Singhal RS, Pandit AB (2014) Degradation of colour in beetroot (*Beta vulgaris* L.): a kinetics study. *J Food Sci Technol* 51(10):2678–2684. <https://doi.org/10.1007/s13197-012-0741-9>
- Chuck-Hernández C, Parra-Saldívar R, Sandate-Flores L (2016) Pitaya (*Stenocereus* spp.). *Encyclopaedia of food and health*. Elsevier, Amsterdam, pp 385–391
- Danisman G, Arslan E, Toklucu A (2015) Kinetic analysis of anthocyanin degradation and polymeric colour formation in grape juice during heating. *Czech J Food Sci* 33(2):103–108
- Delgado-Vargas F, Jimenez AR, Paredes-Lopez O (2000) Natural pigments: carotenoids, anthocyanins, and betalains—characteristics, biosynthesis, processing, and stability. *Crit Rev Food Sci Nutr* 40(3):173–289
- Elbe JH, Schwartz SJ, Hildenbrand BE (1981) Loss and regeneration of betacyanin pigments during processing of red beets. *J Food Sci* 46:1713–1715
- Franks F (1994) Accelerated stability testing of bioproducts: attractions and pitfalls. *Trends Biotechnol* 4:114–117
- Güneser O (2016) Pigment and color stability of beetroot betalains in cow milk during thermal treatment. *Food Chem* 196:220–227
- Jamilah B, Shu CE, Kharidah M, Dzulkifly MA, Noranizan A (2011) Physico-chemical characteristics of red pitaya (*Hylocereus polyrhizus*) peel. *Int Food Res J* 18:279–286
- Khan MI (2016) Plant betalains: safety, antioxidant activity, clinical efficacy, and bioavailability. *Compr Rev Food Sci Food Saf* 15:316–330. <https://doi.org/10.1111/1541-4337.12185>
- King VAE, Lin HJ, Liu CF (1998) Accelerated storage testing of freeze-dried and controlled low-temperature vacuum dehydrated *Lactobacillus acidophilus*. *J Gen Appl Microbiol* 44:161–165
- Merten S (2003) A review of *Hylocereus* production in the United States. *J Prof Assoc Cactus* 5:98–105
- Molina GA, Hernández-Martínez AR, Cortez-Valadez M, García-Hernández F, Estevez M (2014) Effect of tetraethyl orthosilicate (TEOS) on the light and temperature stability of a pigment from *Beta vulgaris* and its potential food industry applications. *Molecules* 19:17985–18002
- Pavokovic D, Krsnik-Rasol M (2011) Complex biochemistry and biotechnological production of betalains. *Food Technol Biotechnol* 49(2):145–155
- Presas-Owen S, Lopez-Sabater MC, Rivero-Urgell M (1995) Shelf-life prediction of an infant formula using an accelerated stability test (Rancimat). *J Agric Food Chem* 43:2879–2882
- Priatni S, Pradita A (2015) Stability study of betacyanin extract from red dragon fruit (*Hylocereus polyrhizus*) peels. *Procedia Chem* 16:438–444
- Reyes LF, Cisneros-Zevallos L (2007) Degradation kinetics and colour of anthocyanins in aqueous extracts of purple- and red-flesh potatoes (*Solanum tuberosum* L.). *Food Chem* 100:885–894
- Rodríguez-Sánchez M, Amaya-Guerra CA, Quintero-Ramos A, Perez-Carrillo E, de Ruiz-Anchondo T, Baez-Gonzalez J, Melendez-Pizarro C (2015) Effect of extrusion cooking on bioactive compounds in encapsulated red cactus pear powder. *Molecules* 20:8875–8892. <https://doi.org/10.3390/molecules20058875>
- Rodríguez-Sánchez JA, Victoria MTC, Barragan-Huerta BE (2017) Betaxanthins and antioxidant capacity in *Stenocereus pruinosus*: stability and use in food. *Food Res Int* 91:63–71
- Ruiz-Gutiérrez MG, Amaya-Guerra CA, Quintero-Ramos A, Pérez-Carrillo E, de Ruiz-Anchondo T, Báez-González J, Meléndez-Pizarro C (2015) Effect of extrusion cooking on bioactive compounds in encapsulated red cactus pear powder. *Molecules* 20:8875–8892
- Slavov A, Karagyzov V, Denev P, Kratchanova M, Kratchanov C (2013) Antioxidant activity of red beet juices obtained after microwave and thermal pretreatments. *Czech J Food Sci* 31(2):139–147
- Stintzing FC, Herbach KM, Mosshammer MR, Carle R, Yi W, Sellappan S, Akoh CC, Bunch R, Felker P (2005) Color, betalain pattern, and antioxidant properties of cactus pear (*Opuntia* spp.) clones. *J Agric Food Chem* 53:442–451
- Sumaya-Martinez MT, Cruz-Jaime S, Madrigal-Santillan E, Garcia-Paredes JD, Carino-Cortes R, Cruz-Cansino N, Valadez-Vega C, Martinez-Cardenas L, Alanis-Garcia E (2011) Betalain, acid ascorbic, phenolic contents and antioxidant properties of purple, red, yellow and white cactus pears. *Int J Mol Sci* 12:6452–6468
- Tesoriere L, Attanzio A, Allegra M, Gentile C, Livrea MA (2014) Inducible nitric oxide synthase (iNOS)-1 activation and NF- κ B-dependent release of inflammatory mediators and prevents the increase of epithelial permeability in IL-1 β -exposed Caco-2 cells. *Br J Nutr* 111:415–423. <https://doi.org/10.1017/S0007114513002663>
- Wiset L, Poomsa-ad N, Srilaong V (2012) Comparisons of antioxidant activity and bioactive compounds of dragon fruit peel from various drying methods. *Int J Biol Biomol Agric Food Biotechnol Eng* 6(10):943–946
- Woo KK, Ngou FH, Ngo LS, Soong WK, Tang PY (2011) Stability of betalain pigment from red dragon fruit (*Hylocereus polyrhizus*). *Am J Food Technol* 6(2):140–148
- Yalçınöz SK, Erçelebi EA (2015) Anthocyanin degradation and colour kinetics of cornelian cherry concentrate. *Br J Appl Sci Technol* 10(4):1–12
- Zhang Q, Pan J, Wang Y, Lubet R, You M (2013) Beetroot red (betanin) inhibits vinyl carbamate- and benzo(a)pyrene-induced lung tumorigenesis through apoptosis. *Mol Carcinog* 52(9):686–691. <https://doi.org/10.1002/mc.21907>

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