Spatial distribution characteristics of bacterial community structure and gene abundance in sediments of the Bohai Sea

Quanrui Chen[1, 2], Jingfeng Fan[1, 3*], Jie Su[1], Hongxia Ming[1], Zhihao Sun[1, 2], Mengfei Li[1, 2], Xiaohui Zhao[1, 2], Yantao Wang[1, 2], Yingxue Zhang[1, 2], Huizhen Zhang[1], Yuan Jin[3], Xiaowan Ma[3], Bin Wang[2]

[1] National Marine Environmental Monitoring Center, Dalian 116023, China

[2] College of Aquaculture and Life, Dalian Ocean University, Dalian 116023, China

[3] Fourth Institute of Oceanography, Ministry of Natural Resources, Beihai 536000, China

Abstract

This study investigated differences in the community structure and environmental responses of the bacterial community in sediments of the Bohai Sea. Illumina high-throughput sequencing technology and real-time PCR were used to assay the bacterial 16S rRNA genes in the surface sediments of 13 sampling stations in the Bohai Sea. The results showed that sediments at the majority of the 13 sampling stations were contaminated by heavy metal mercury. The main phyla of bacteria recorded included Proteobacteria (52.92%), Bacteroidetes (11.76%), Planctomycetes (7.39%), Acidobacteria (6.53%) and Chloroflexi (4.97%). The genus with the highest relative abundance was *Desulfobulbus* (4.99%), which was the dominant genus at most sampling stations, followed by *Lutimonas* and *Halioglobus*. The main factors influencing bacterial community structure were total organic carbon, followed by depth and total phosphorus. The content of lead, cadmium, chromium, copper and zinc had a consistent effect on community structure. Arsenic showed a negative correlation with bacterial community structure in most samples, while the impact of mercury on community structure was not significant. The bacterial community in sediment samples from the Bohai Sea was rich in diversity and displayed an increase in diversity from high to low latitudes. The data indicated that the Bohai Sea had abundant microbial resources and was rich in bacteria with the potential to metabolize many types of pollutants.

Key words: Bohai Sea sediments, bacterial community structure, high-throughput sequencing, 16S rRNA gene

1 Introduction

Coastal ecosystems are extremely important, both in terms of their ecological significance and their economic value to humans (Lv et al., 2016). Sediments comprise an important part of this ecosystem and are subject to interactions between geological, hydrodynamic, biological and chemical factors (Köster and Meyer-Reil, 2001). Offshore sediments are both a source and sink for a large number of pollutants, including nutrients, heavy metals, and petroleum or pesticide residues (Lofrano et al., 2017). Microorganisms participate in various biogeochemical cycles in sediments such as carbon fixation (Jing et al., 2018), methanogens (Tian et al., 2014), nitrification and denitrification (Fan et al., 2011), and biotransformation for a variety of organic pollutants (Wada et al., 2012). Microbial communities are known to be impacted by various ecological factors (Wang et al., 2018), with human activities also producing changes in sediment microbial communities (Yang et al., 2000). Studies of microbial diversity in marine sediments are critical to understanding the evolutionary and ecological processes that affect global microbial diversity, and the response of marine ecosystems to environmental change.

As a semi-open sea in northern China, a large number of population centers have become established on the shores of the Bohai Sea. The GDP of surrounding cities has grown rapidly with the development of industry and agriculture. Recent studies have reported that the Bohai Bay is polluted by heavy metals to varying degrees (Zhou et al., 2017; Li et al., 2017). Heavy metals have also been reported in the nutrient salts of the Bohai Sea. Microbial communities are often considered to have better adaptability and tolerance because they have metabolic pathways that can utilize various resources (Dai et al., 2016; Marshall et al., 2018). Changes in the microbial community under the influence of long-term ecological factors and the potential for selective exposure to different heavy metal contaminants in the environment remains unclear. In this study, the Bohai Sea was selected as the research area. Real-time quantitative PCR and high-throughput sequencing of bacterial 16S rRNA genes were used to study the community abundance and structure of bacteria in the sediments, and the influence of ecological factors was explored. The interrelationship between bacterial communities and ecosystems in sediments aids our understanding of the geographical distribution patterns of marine microbes and the niche differentiation of various microbial communities.

2 Materials and methods

2.1 *Sample collection*

The collection of experimental samples was carried out in late May 2017. A total of 13 stations (Fig. 1), were located throughout the Liaodong Bay, the Huanghe (Yellow) River Estuary, the periphery of the Laizhou Bay and the central part of the Bohai Sea. The average depth of the sampling stations was 20 m. The sediments at 0–10 cm of the surface layer were collected using a grab sampler. Three replicate samples were collected at each station. The three samples from each station were mixed together and dispensed into two labelled sterile valve bags, shipped back to the laboratory and stored at –20°C for future analysis.
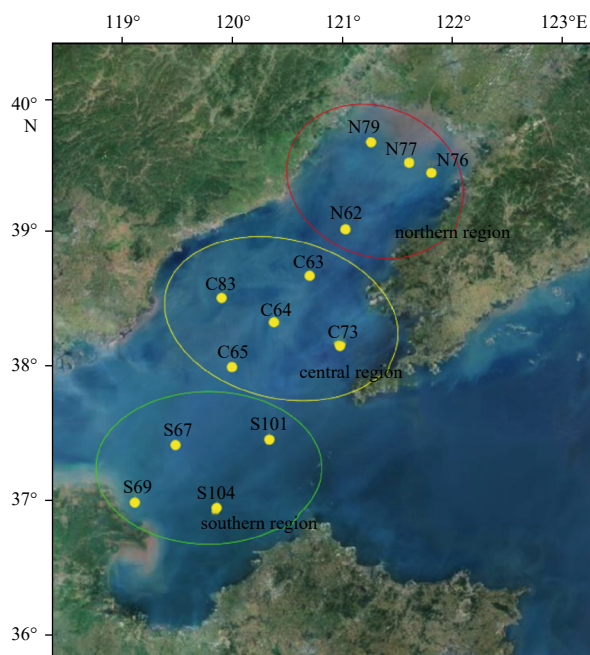
Fig. 1. Map of sample collection stations in the Bohai Sea, China.

2.2 *Determination of environmental factors and concentration of heavy metals*

Station depth was measured at each sampling site and the particle size of the sediment was measured using a LS13320 laser diffraction particle size analyzer (Beckman Coulter, USA). Total organic carbon (TOC), total phosphorus (TP) and total nitrogen (TN) in the sediment, as well as heavy metals including cadmium (Cd), chromium (Cr), copper (Cu), lead (Pb), zinc (Zn), mercury (Hg), arsenic (As), were determined following "Marine Monitoring Specification Part 5: Sediment Analysis GB 17378.5–2007"(General Administration of Quality Supervision, Inspection and Quarantine of the People's Republic of China and Standardization Administration of China, 2008). Determination of nutrient salts the nitrates and nitrous acid salt and ammonia nitrogen content in the sediments were determined by the method of "Measurement of Soil Ammonium, Nitrite and Nitrate Chloride Solution Extraction-spectrophotometry HJ 634–2012" (Ministry of Environmental Protection of the People's Republic of China, 2012).

2.3 *DNA extraction and high-throughput sequencing*

Total bacterial DNA was extracted from sediment samples using the DNeasy PowerSoil Kit (Qiagen, Germany). After the sediment was mixed, 0.35 g of each sample was taken for total DNA extraction, according to the kit extraction instructions. This was carried out in triplicate. After extraction, the quality of the DNA was tested by 1.5% agarose gel electrophoresis. Three DNA samples were then combined into one and stored at –80°C.

The extracted total DNA template was subjected to two-step PCR amplification using a V4-V5 region-specific primer of the bacterial 16S rRNA gene to construct a 16S rRNA gene library (Liu et al., 2008). The first step uses specific primer amplification: 515F (5′-GTGCCAGCMGCCGCGGTAA-3′) and 926R (5′-CCGT-CAATTCMTTTGAGTTT-3′). PCR amplification conditions: 5× buffer, 10 μL; dNTP (10 mmol/L), 1 μL; Phusion ultra-fidelity DNA polymerase 1 U; 1 μL of each primer (10 μmol/L); 5 μL of total DNA template; and finally supplemented with 50 μL ddH₂O. The initial PCR procedures were: 94°C for 2 min; follow by 25 cycles of denaturation at 94°C for 30 s; 56°C annealing for 30 s; 72°C for 30 s; and final extension at 72°C for 5 min. The first PCR product was electrophoresed using a 2% detection agarose gel and recovered using the AxyPrep DNA Gel Extraction Kit (Axygen, USA). The second PCR amplification was then performed, to add the linker barcode required for sequencing the Illumina high throughput sequencing platform to the target fragment. After the product was purified, the quantitative control was carried out. Finally, the 16S rRNA gene v4-v5 region was sequenced by Tiny-Gene Bio-Tech (Shanghai) Co., Ltd. through the Illumina MiSeq platform using 2×300 bp end pairing sequencing.

2.4 *Data statistics and analysis*

Using the sequencing data, the sample was first distinguished according to the barcode, and the optimized sequence was obtained by splicing, quality control and filtering. The optimized sequence was then clustered using Mothur (version 1.39.5) and USEARCH programs, these sequences were clustered to OTUs (operational taxonomic units) at 97% sequence identity (furthest neighbor method), and then the classification level of OTU in each sample was classified according to the species level. Mothur and R language were used to plot the dilution curve and abundance level map of OTUs, and multi-sample similarity analysis, principal component analysis and alpha diversity were completed by OTU classification level results. Environmental factors and RDA analysis between heavy metals and samples were performed using CANOCO 5.0 software, and SPSS 23.0 for correlation analysis between various factors.

The heavy metal pollution assessment was calculated by the geoaccumulation index (I_{geo}) method (Muller, 1969), and the calculation equation is as follows:

$$I_{geo} = \log_2 \left[\frac{\rho_n}{k \times B_n} \right]. \tag{1}$$

where ρ_n represents the concentration of metal (n) in the sample, B_n is the background value of the element in the sediment, k is used to correct the background value of the region, and the sediment environment is usually taken as 1.5 (Müller and Förstner, 1976).

The original sequence of the bacterial 16S rRNA gene obtained by high-throughput sequencing in this study has been registered in the NCBI database, and the gene sequence accession number is SRR6323824 to SRR6323836 (13 samples).

3 Results

3.1 *Spatial distribution of environmental factors*

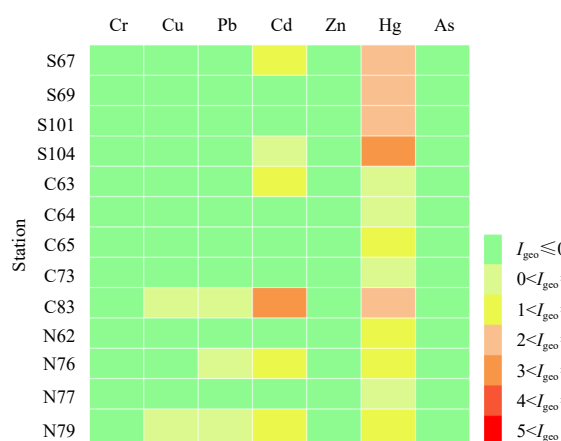The environmental factors of sediment samples from 13 sta-

Table 1. Environmental factor parameters

Station	NO$_3^-$/μg·g^{-1}	NO$_2^-$/μg·g^{-1}	NH$_4^+$/μg·g^{-1}	TOC/%	TP/μg·g^{-1}	TN/μg·g^{-1}	depth/m	Grid size (Mz, Φ)
S67	421	29.9	153	0.66	143	130	23	6.63
S69	513	27.6	173	0.43	386	122	10	5.76
S101	482	59	140	0.36	174	93.6	27	5.64
S104	256	29.9	121	0.53	348	135	17.5	6.11
C63	217	68.9	96.2	0.49	255	97.3	28	6.22
C64	149	85.3	101	0.24	181	91.6	19.8	4.99
C65	228	70.7	104	0.39	208	93.6	19	4.64
C73	183	103	68.3	0.62	221	145	34	6.21
C83	330	83.4	93.3	0.92	269	157	23	7.37
N62	185	75	84.9	0.36	212	95.8	26	5.43
N76	350	75.4	112	0.84	290	114	13	6.3
N77	256	67.4	72	0.53	219	100	12.7	5.81
N79	138	58.5	197	0.75	245	111	12.5	6.52

tions in the Bohai Sea are shown in Table 1. Nitrate content in the sediments of the Bohai Sea region was between 138–513 μg/g and the ammonia nitrogen content was 72–197 μg/g. Nitrate and ammonia nitrogen were highest in the southern region, with lowest values recorded in the central region. Nitrous acid salt content ranged from 27.6–103 μg/g in the central region. TP was higher near sampling Stas S69 and S104. Overall TP values were highest in the southern region, followed by the northern region, and were lowest in the central region. Highest values for TN, TOC and grid size were recorded at sampling Sta. C83 near Qinhuangdao. The sampling stations ranged in depth from 10 to 34 m, and the central region was significantly deeper than other areas. According to the on-site measurement data, there was no significant difference in salinity and pH between stations, so it was not included in the analysis.

3.2 *Evaluation of heavy metal pollution in sediments*

According to the results of the geoaccumulation index method in the sediment (Fig. 2), heavy metals such as Cr, Zn and As in the sediment of the 13 sampling sites in the Bohai Sea could be classified as unpolluted ($I_{geo} \leqslant 0$). Cu was present at low pollution levels at sampling Stas N79 and C83 ($0 < I_{geo} \leqslant 1$), while Pb was also recorded as slightly polluting at three sampling stations ($0 < I_{geo} \leqslant 1$). Six sampling stations were found to be moderately polluted by Cd. In particular, Sta. N79 near the coast of Jinzhou in the Liaodong Bay was highly polluted. Hg was detected at all

sampling stations. Stations S104 and S101 in the Laizhou Bay area had the highest concentrations of Hg. The degree of Hg pollution was highest in the southern region, followed by the northern region, and was lowest in the central region. Stations N77, C64 and C73 had low levels of heavy metal pollution, with only mild contamination of Hg. Sampling Sta. C83 was affected by a variety of heavy metal pollutants.

3.3 *Bacterial community structure*

The gene abundance of the bacterial 16S rRNA gene V4–V5 region from the 13 sampling stations was determined by real-time PCR (qPCR) as shown in Fig. 3. The gene abundance of the 16S rRNA gene V4–V5 region in the sediment at sampling stations in the Bohai Sea ranged from 8.83×10^9 to 6.71×10^{10} copy/g, with an average abundance of 3.17×10^{10} copy/g. The gene abundance in the northern region was found to be lower than that in the southern and central regions. Sampling Sta. C63 in the central region, located close to the northern region, had the lowest gene abundance. The sediments of sampling Stas C83 and S104 were rich in bacterial 16S abundance. The only ecological factor that showed a significant correlation with the gene abundance at each sampling station was TN ($P<0.05$).

Samples from the 13 sampling sites in the Bohai area were sequenced by high-throughput sequencing of the bacterial 16S rRNA gene V4–V5 region of the extracted total DNA. As shown in Table 2, the number of optimized sequences obtained was 22

Fig. 2. Heat map showing the geoaccumulation index (I_{geo}) in sediments collected at sampling stations.
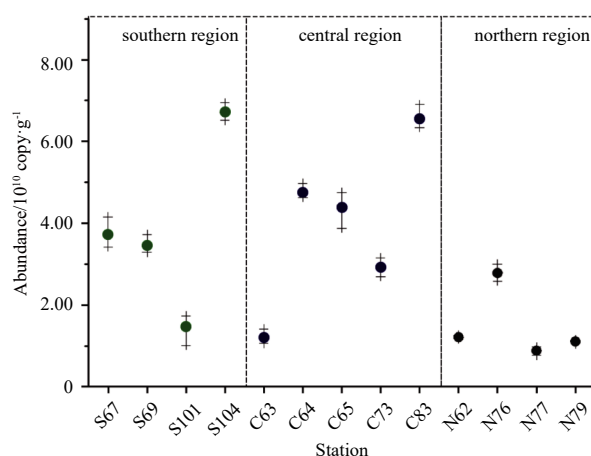
Fig. 3. Gene abundance of the bacterial 16S rRNA gene V4-V5 region in each station.

514–32 385. All sequences were clustered by 97% similarity to obtain 3 041 different OTUs. The number of OTUs at each sampling station ranged from 1 585 to 2 182, and the highest value was recorded at Sta. S67. This indicated that the sediments at this site had rich bacterial diversity, while Sta. S69 near the Huanghe River Estuary had the lowest number of OTUs (1 585). A maximum of 1 937 OTUs was recorded in the southern region, which was significantly higher than that in the northern region. The coverage of all samples was above 97.7%, indicating that the sample library coverage was high, and most of the bacterial 16S rRNA gene sequences were detected at all sites. The sequencing results were well representative and reliable. The Chao1 index showed that Sta. S67 had the highest total number of species, while the Shannon index showed that Sta. C83 had the highest bacterial diversity. The Shannon index of all stations was between 5.59 and 6.24. The Shannon index value was highest for the southern region, followed by the northern region, with the lowest value obtained for the central region, meaning that the relative average bacterial diversity in the central region was lower than that in the southern and northern regions. The Chao1 index showed a significant correlation with the Shannon index between stations ($P<0.05$).

The Venn diagram shows the number of common and unique OTUs in each sediment sample. As shown in Fig. 4a, 2 439 OTUs were common to all three regions. The southern and central regions had 222 common OTUs, which is higher than the overlap of the northern and central and northern and southern regions. There were 56 unique OTUs in the northern region, indicating that the northern region had relatively more endemic microbial species than the central or southern regions. The petal diagram in Fig. 4b shows the number of OTUs shared between the 13 sampling stations. The core microbiome analysis found 100% of the OTUs covering the 13 samples. The number of core OTUs was 540. Station S69, near the mouth of the Huanghe River in the southern region, had the highest number of unique OTUs (21), followed by Sta. C63 in the central region, which had 14 unique OTUs.

3.4 *Bacterial community species composition*

The OTUs of the bacterial communities in the sediments of the 13 sampling stations were clustered into 36 phyla, 49 classes, 77 orders, 109 families, 181 genera and 198 species. The main taxonomic results are shown in Fig. 5a, in which Proteobacteria dominated the sediment samples from the Bohai Sea, with an av-

Table 2. High-throughput sequencing results and diversity indices between each station

Station	Reads	OTU	Ace	Chao1	Shannon	Simpson	Coverage/%
S67	28 770	2 182	2 547	2 520	6.13	0.008 2	98.2
S69	22 563	1 585	2 029	2 079	5.83	0.009 9	97.9
S101	32 385	1 963	2 299	2 328	5.94	0.008 7	98.6
S104	22 514	2 017	2 395	2 397	6.1	0.008 6	97.7
C63	26 467	2 011	2 371	2 347	6.07	0.008 8	98.2
C64	24 761	1 812	2 245	2 208	5.59	0.016 1	97.9
C65	25 325	1 825	2 282	2 248	5.78	0.009 8	98.0
C73	28 309	1 838	2 345	2 370	5.62	0.014 7	98.1
C83	24 577	2 117	2 473	2 505	6.24	0.007 2	98.0
N62	24 563	1 727	2 135	2 113	5.78	0.010 8	98.1
N76	26 143	2 065	2 457	2 428	6.07	0.009 2	98.0
N77	24 527	1 892	2 325	2 279	5.92	0.011 3	97.9
N79	26 312	1 873	2 257	2 239	5.93	0.009 3	98.2
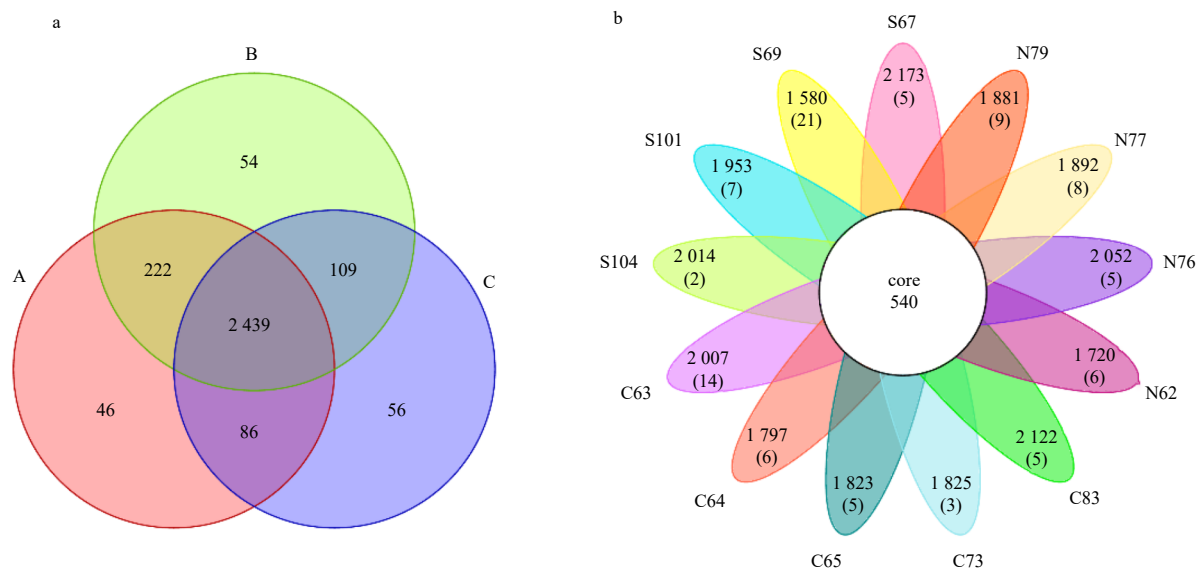
Fig. 4. OTU distribution Venn diagram (a) and petal picture (b). A, B and C represent the Venn diagram southern region, the central region and the northern region Petal picture, respectively.

erage relative abundance of 52.92%. The average relative abundance of Bacteroidetes was 11.76%. The relative abundance of Planctomycetes was 7.39%, Acidobacteria had a relative abundance of 6.53%, while Chloroflexi had a relative abundance of 4.97%. The remaining bacteria had a relative abundance of less than 4.0%. In the central region, at Sta. S69, Bacteroidetes were most abundant (22.29%), and were significantly more abundant here than other sampling stations. A total of 77 bacterial orders (including unclassified parts) were detected at the 13 sampling stations. The main orders are shown in Fig. 5b. Xanthomonadales were dominant at all sampling stations (average relative abundance of 12.29%), followed by Desulfobacterales (9.23%), Flavobacteriales (6.41%) and Desulfuromonadales (2.83%). Station S69 had the lowest relative abundance of Desulfobacterales (3.59%) and the highest relative abundance of Flavobacteriales (14.07%).

As shown in Fig. 6, at the genus level, the *Desulfobulbus* of the δ-Proteobacteria was the only genus with a relative abundance higher than 1%. *Desulfobulbus* had a relative abundance of 4.99%, followed by the genus *Lutimonas* (0.88%) of the family Flavobacteriia, the genus *Halioglobus* (0.84%) of the γ-proteobacteria, and the genus *Lutibacter* of the family Flavobacteriia (0.83%). *Nitrosomonas*, a functional microbe involved in the nitrogen cycle, was recorded at various sampling stations, with an average relative abundance of (0.13%). The genus *Nitrospira* had an average relative abundance of 0.37%. In addition, *Blastopirellula* (0.39%) and *Rhodopirellula* (0.30%) of the Planctomycetacia, which are involved in anaerobic ammonium oxidation was detected at various stations, and some genera (*Streptococcus*, *Bacillus* and *Kangiella*) that may have a mercury reductase (*merA*) gene were also detected. Unclassified genera accounted for 78.13% of all sequences.
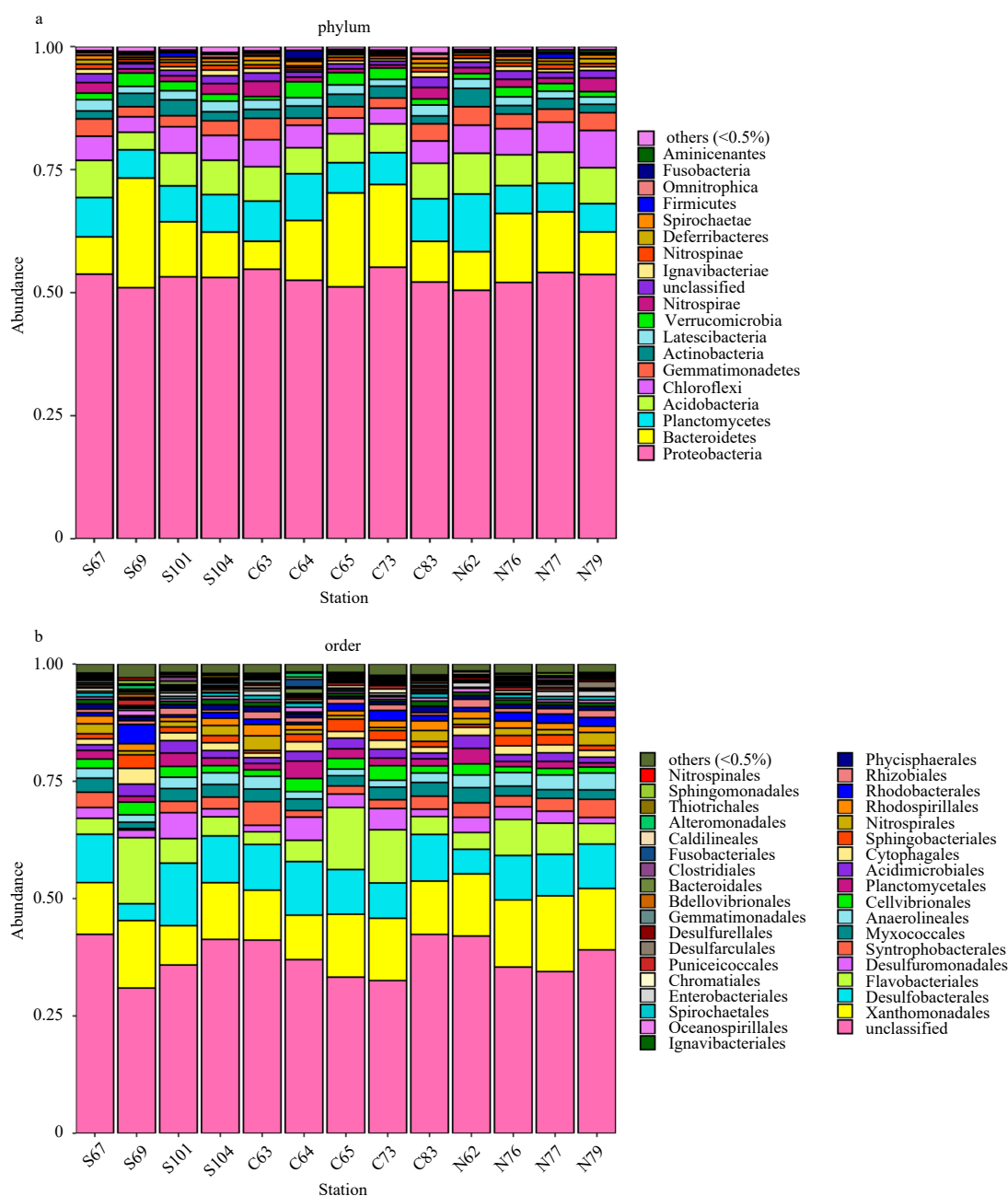
Fig. 5. Histogram of community structure of bacteria at the phylum level (a) and order level (b).
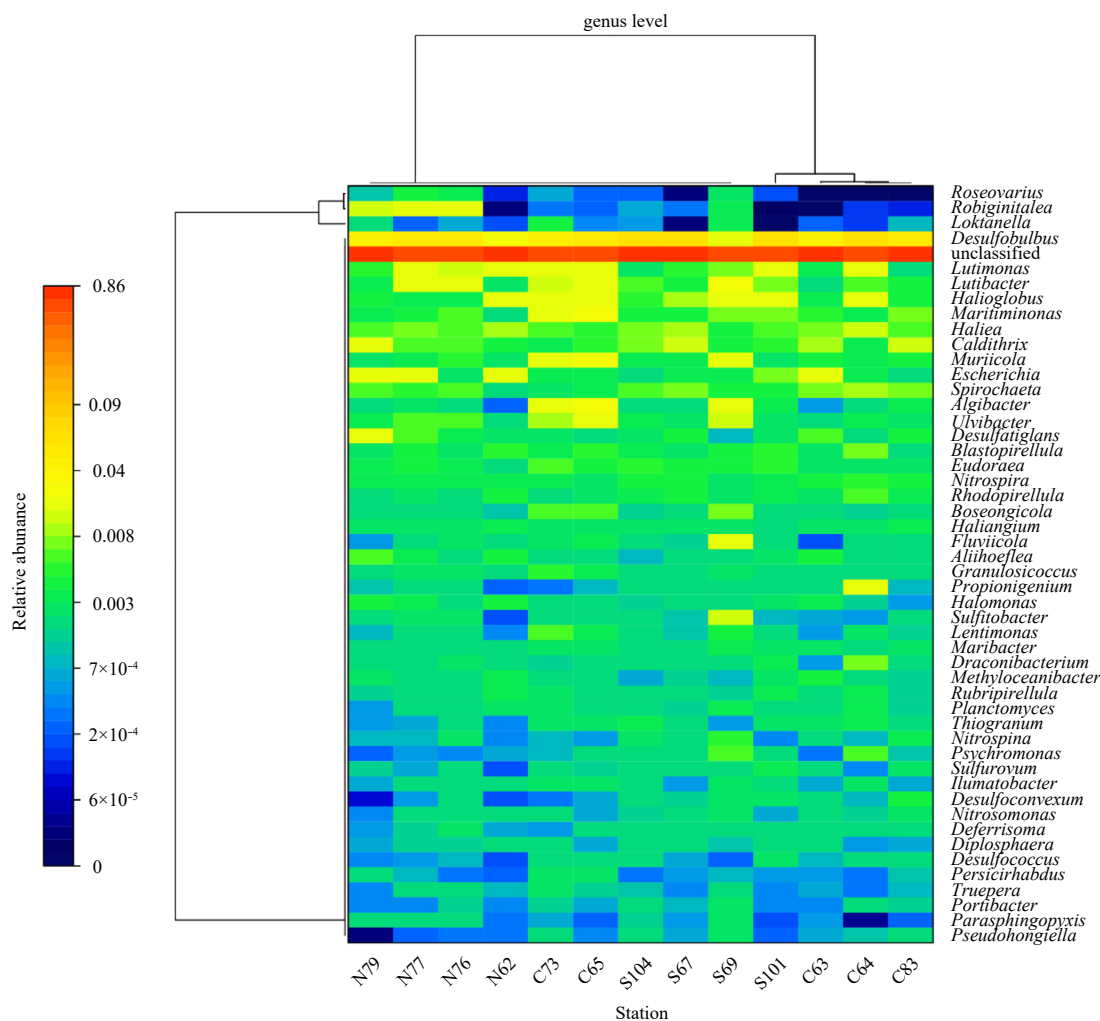
Fig. 6. Heat map of the community composition of bacteria at the genus level.

3.5 *Correlation analysis between bacterial community structure and ecological factors*
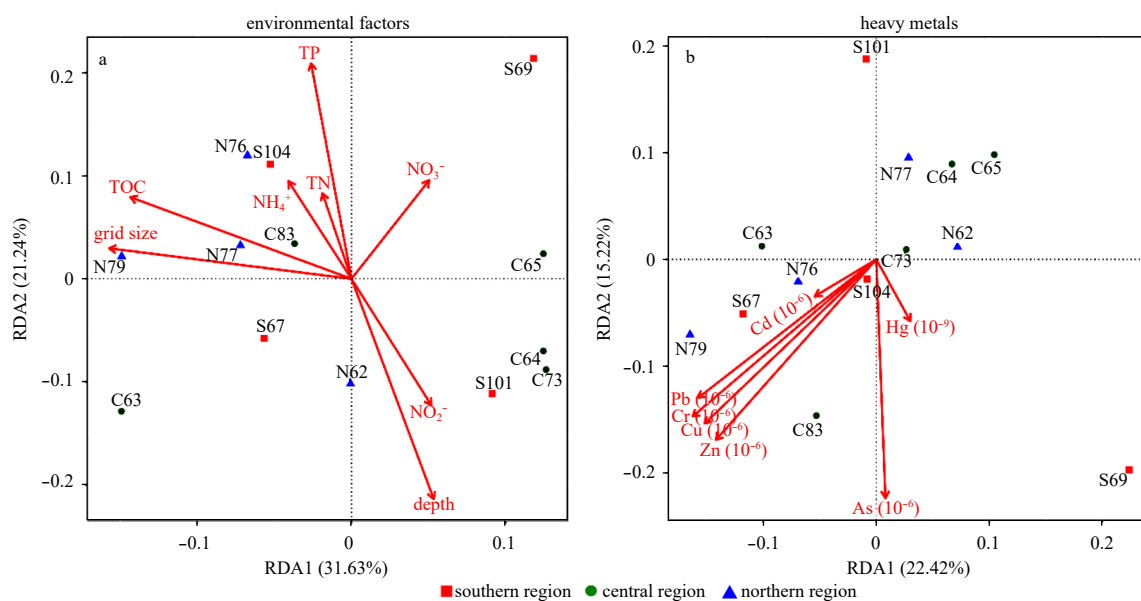
3.5.1 *Redundancy analysis of environmental factors and community structure*

Some of the environmental factors of the sediments in the Bohai Sea region varied greatly and were influenced by bacterial community structure. Redundancy analysis of environmental factors and the bacterial community structure at each sampling station was carried out using CANOCO 5.0 software (Fig. 7a). The degree of interpretation of the sorting axes RDA1 and RDA2 were 31.63% and 21.24%, respectively. The composition of the bacterial community at Sta. S69 was quite different from the other sampling stations, and was mainly affected by the positive correlation of nitrate content. The environmental factors that were positively correlated with the community structure at Stas N76, N77 and N79 were TOC and sediment size, followed by ammonia nitrogen, TN and TP. Stations C62, C64, C73 and S101 had a significant positive correlation with sediment depth and a significant negative correlation with TOC and grid size. For all environmental factors, TOC had the highest significant correlation with bacterial community structure (*P*=0.027, *P*<0.05), while there was no significant correlation with ammonium (*P*=0.738), nitrate (*P*=0.394) and nitrite (*P*=0.269) and the bacterial community in

the sediments of the Bohai Sea. The results showed that the main factor influencing bacterial community structure in the sediments of the Bohai Sea was the TOC content in the sediment, followed by depth and TP.

3.5.2 *Redundancy analysis of heavy metals and community structure*

Redundancy analysis of heavy metal content and bacterial community structure at each sampling station was also carried out with CANOCO 5.0 software (Fig. 7b). The degree of interpretation of the sorting axes RDA1 and RDA2 were 22.42% and 15.22%, respectively. It can be seen that the sequence of influence of heavy metals on the microbial community structure in the sediment was Zn, Cr, As, Cu, Pb, Hg and Cd. Correlation with bacterial communities showed consistency in the effects of Pb, Cd, Cr, Cu and Zn. There was a positive correlation between the bacterial community at Stas C83 and S104 and heavy metals (*P*<0.05). The sediments at these two sampling stations were heavily contaminated by heavy metals. The negative correlation between Stas C65, C64 and C73 and most heavy metals were not significantly associated with Hg. The bacterial community structure at Sta. S101 was significantly negatively correlated with As. In the majority of samples the content of arsenic was negatively correlated with bacterial community structure; however, the in-

Fig. 7. Redundancy analyses of environmental factors and community structure (a), and heavy metals and community structure (b).

fluence of Hg on community structure was not significant. Although the pollution of Hg and Cd was the most obvious in each sediment samples, the effects of these two heavy metals on the bacterial community structure were not significant, this result may be the stress response of bacterial community structure to heavy metals, and is a long-term accumulation and evolution process, but this conclusion needs to be further confirmed by metagenomics data in the future.

4 Discussion

4.1 *Bacterial community structure diversity*

The bacterial community structure in marine sediments drives a variety of biogeochemical cycles. The study of the potential distribution patterns of these microorganisms and the mechanisms by which their community structure changes in the environment has always received much attention (Shade, 2018). The results of the current study showed that a large number of bacterial OTUs (1 585–2 182) were detected in sediments of the Bohai Sea. Diversity was highest in the southern and central regions, with bacterial 16S rRNA gene abundance showing the same trend. In studies of the Liaohe Estuary of the Bohai Sea, using high-throughput sequencing of the bacterial 16S rRNA gene V3-V4 region our laboratory found that the diversity of bacterial communities in the sediments farther from the shore was lower than estuarine, freshwater and mixed areas. A total of 32 phyla were recorded in the previous studies, lower than the 36 phyla in the current study. The diversity of sedimentary bacterial communities in the northern region of the current study coincided with the previous results from the Liaohe Estuary (Zhang et al., 2018). A relatively high number of OTUs (2 590–3 777) has been reported from the sediments of the Liaodong Bay in the Bohai Sea (Zheng et al., 2014). The number of OTUs obtained by 454 pyrosequencing from three sediment deposits in the Bohai Sea ranged from 3 371–5 224 (Liu et al., 2015). The main reason for the difference was that intertidal sediments had more abundant dissolved oxygen and stronger bioturbation than the distant sea sediments, and compared with Illumina sequencing, 454 pyrosequencing results were more prone to higher mismatch

rates and the emergence of artificial amplification products (Gomez-Alvarez et al., 2009; Quince et al., 2009).

A variety of taxa were detected at all sampling stations; however, Proteobacteria were the most abundant and extensive in sediments. These findings were consistent with results from the Hangzhou Bay (Su et al., 2018), Jiaozhou Bay (Zhao et al., 2014) and the Changjiang (Yangtze) River Estuary (Feng et al., 2009). In the current study, the average relative abundances of γ-proteobacteria, δ-proteobacteria and α-proteobacteria were 24.6%, 22.9% and 4.7%, respectively. Bacterial community structure was similar to that reported from previous studies of coastal marine sediments (Guan et al., 2014; Lv et al., 2016). At Sta. S69 near the Huanghe River Estuary, γ-proteobacteria and α-proteobacteria accounted for high relative abundances of 30.2% and 8.5%, respectively, while δ-proteobacteria accounted for only 10.6%. Studies have found that the input of freshwater at the Changjiang River Estuary impacts the microbial community in the estuarine sediments (Guo et al., 2018). The relative abundance indicated that the bacterial community structure in S69 sediments with continuous freshwater and sediment input from the Huanghe River Estuary was significantly different from that in marine sediments. Higher levels of sediment in the southern region originated from the Huanghe River, and the impact of freshwater input on bacterial community structure provided a possible explanation for this difference. The current study also revealed that there were a large number of diverse microbial resources in the semi-open seas of the Bohai Sea.

The distribution pattern of bacteria was mainly driven by two factors, geographical distance and environmental variables. There was an obvious negative correlation between the distribution of bacterial communities and geographical distance (Martiny et al., 2011). Geographical barriers, such as ocean currents, water masses, and depth (pressure) (Brown et al., 2009; Zhang et al., 2014), constrain bacterial spread, and so the effects of constructing bacterial communities at small spatial scales such as the Bohai Sea were significant. Studies have found that Firmicutes occupy an important role in coastal sediments (Zinger et al., 2011). However, the average abundance of Firmicutes in sediments of the Bohai Sea was found to be 0.35%, far lower than the

relative abundance of Bacteroidetes. Bacteroidetes accounted for 11.8% of the surface sediment in the Bohai Sea, and may play a role in the degradation of molecular organic matter in the environment (Sinkko et al., 2013). The discovery of Chloroflex (5.0%) at the 13 sampling stations means that sediments in the Bohai Sea were subjected to oil pollution, as it has been noted in previous studies that the bacteria within this phylum play a key role in petroleum degradation (Woese, 1987). Therefore, although the Bohai Sea is a geographically small area, it comprises various ecosystems, and the microbial community diversity of the area is a rich resource, which warrants further study. The microbial processes lay a solid foundation for revealing the geochemical cycles in which microbes participate.

4.2 *Analysis of dominant flora under different pollutants*

In addition to environmental factors, the presence of pollutants (including oil pollution and heavy metal pollution) at various concentrations in marine sediments can also have significant effects on bacterial community structure (Wang and Tam, 2012). Therefore, the analysis of bacterial community structure is of great significance, because it can indirectly reflect multiple changes in the marine environment. Desulfuromonadales of δ-proteobacteria, detected in the current study, were often found in an environment under metal stress, which means that they have metal reducing ability (Cabrera et al., 2006; Vandieken et al., 2012). In the current study, Desulfuromonadales had an average abundance of 2.83% overall; however, the relative abundance of Desulfuromonadales at Stas S69, S104, C63, C83 and N79, which are more seriously polluted by heavy metals, was lower than others. Stations S101, C64 and C73 had relatively unpolluted sediments. Xanthomonadales was the dominant order, accounting for 12.28% of all orders. This order is generally detectable in marine sediments; however, these organisms use various other carbon substrates as carbon sources, and are classified as a general hydrocarbon degrading microorganisms (Gutierrez, 2017). Therefore, it is possible that the high abundance in the northern region may be due to a higher oil content in the sediment than in the central and southern regions. Desulfobacterales in the δ-Proteobacteria were the second most abundant order, accounting for an average relative abundance of 9.23%. Desulfobacterales may play a role in the sulfate reduction process, indicating that sulfate reduction processes are common in sediments of the Bohai Sea. These sediments are mainly affected by human activities. These results may reveal that the organic pollution caused by the industrial and agricultural development of the surrounding coastal cities in the Bohai Sea environment was significant and different, with contamination of regional sediments significantly affecting patterns of microbial community composition.

Desulfobulbus was the most abundant genus at most sampling stations, as shown in Fig. 8. All members of this genus can reduce sulfate and thiosulfate to hydrogen sulfide gas (Kuever et al., 2015). Among the dominant species, *Lutimonas*, *Halioglobus* and *Caldithrix* were significantly associated with Cr, Cu, Pb, Cd and Zn (P<0.05). Some *Lutimonas* have been found to contain catalase and oxidase, and have the ability to reduce nitrate to nitrite (Kim et al., 2014). In the ocean, the genus *Halioglobus* is found to reduce nitrate to nitrogen (Park et al., 2012). *Caldithrix* is a common genus in the ocean, which is associated with anaerobic respiration with molecular hydrogen or acetate as an electron donor and nitrate as an electron acceptor, especially in thermophilic, halophilic and sulfide-rich environments. It can be present in large amounts in petroleum-containing sediments and participates in the nitrogen cycle, forming ammonium salts
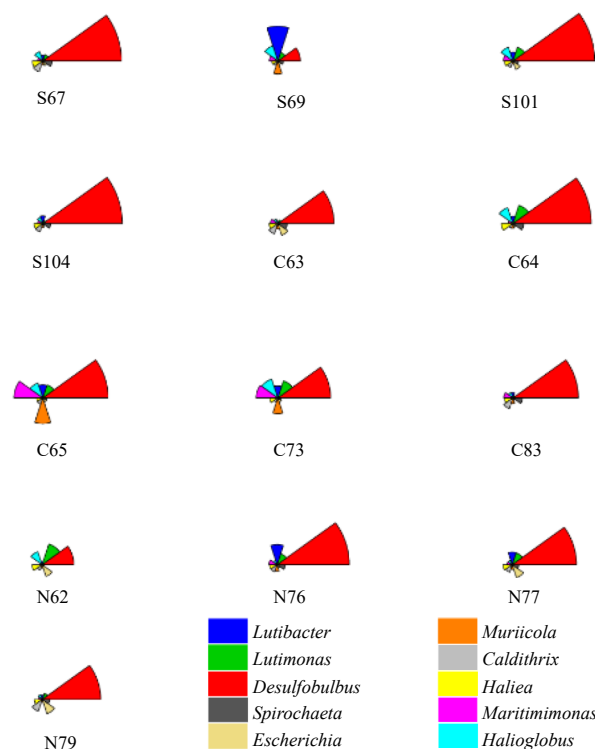
Fig. 8. Star map analysis of the ten genera with the highest relative abundance.

(Alauzet and Jumas-Bilak, 2014; Miroshnichenko et al., 2003). In the current study, it was also found that *Caldithrix* was significantly correlated with sediment particle size (*P*<0.01) and TOC (*P*<0.05). At Stas S104 and S101, which were most polluted by Hg, the relative abundance of the *Eudoraea* genus was significantly higher than that of other stations. As one of the dominant genera, it is speculated that the genus may have a better Hg tolerance. Some studies found that the *Eudoraea* genus has no effect on nitrates, but can use various carbon compounds as the sole carbon source (Alain et al., 2008). The sediment of sampling Sta. 83 was found to be polluted by various heavy metals. The relative abundance of *Desulfoconvexum* was found to be four times higher at Sta. 83 than the average relative abundance of other stations, and was found to be abundant in the southern region where heavy metal pollution was relatively serious. It was significantly correlated with nitrate (*P*<0.05). Within this genus, some species were found to be able to utilize organic matter in a variety of environments and eventually completely oxidized to carbon dioxide (Könneke et al., 2013).

5 Conclusions

(1) The main groups of bacteria in the sediments of the Bohai Sea included Proteobacteria, Bacteroidetes, Planctomycetes, Acidobacteria and Chloroflexi. At the genus level, *Desulfobulbus* was most abundant, followed by *Lutimonas*, *Halioglobus* and *Caldithrix*, with abundance significantly related to the presence of heavy metals.

(2) Using real-time PCR and high-throughput sequencing of bacterial 16S rRNA genes in sediment samples from 13 sampling stations in the Bohai Sea, community structure diversity and abundance of bacteria in the sediment were found to increase from the northern region to the central region, and further increase in the southern region.

(3) In the Bohai Sea sediments were mainly polluted by Hg, with highest levels of Hg pollution in the southern region, particularly at Stas S101 and S104, where the Hg the Hg concentration was extremely high, an issue which deserves attention. In addition, sediments in the sea around Qinhuangdao were also affected by a variety of heavy metals.

(4) The main factors influencing bacterial community structure in the sediments of the Bohai Sea were TOC content in the sediment, followed by the depth and TP. The correlation between heavy metals and bacterial communities showed the consistency of the effects of Pb, Cd, Cr, Cu and Zn on the community, while the effect of Hg on the community structure was not significant. Arsenic content also showed a negative correlation with bacterial community structure in the majority of samples.

References

Alain K, Intertaglia L, Catala P, et al. 2008. *Eudoraea adriatica* gen. nov., sp. nov., a novel marine bacterium of the family *Flavobacteriaceae*. International Journal of Systematic and Evolutionary Microbiology, 58(10): 2275–2281, doi: 10.1099/ijs.0.65446-0

Alauzet C, Jumas-Bilak E. 2014. The phylum deferribacteres and the genus *Caldithrix*. In: Rosenberg E, DeLong E F, Lory S, et al., eds. The Prokaryotes. Berlin, Heidelberg: Springer, 595–611

Brown M V, Philip G K, Bunge J A, et al. 2009. Microbial community structure in the North Pacific Ocean. The ISME Journal, 3(12): 1374–1386, doi: 10.1038/ismej.2009.86

Cabrera G, Pérez R, Gómez J M, et al. 2006. Toxic effects of dissolved heavy metals on *Desulfovibrio vulgaris* and *Desulfovibrio* sp. strains. Journal of Hazardous Materials, 135(1–3): 40–46, doi: 10.1016/j.jhazmat.2005.11.058

Dai Tianjiao, Zhang Yan, Tang Yushi, et al. 2016. Identifying the key taxonomic categories that characterize microbial community diversity using full-scale classification: a case study of microbial communities in the sediments of Hangzhou Bay. FEMS Microbiology Ecology, 92(10): fiw150, doi: 10.1093/femsec/fiw150

Fan Jingfeng, Chen jiaying, Chen Liguang, et al. 2011. Research on denitrifying bacteria quantification and diversity in Liaohe Estuary sediments. Haiyang Xuebao (in Chinese), 33(3): 94–102

Feng Biwei, Li Xiaoran, Wang Jinhui, et al. 2009. Bacterial diversity of water and sediment in the Changjiang estuary and coastal area of the East China Sea. FEMS Microbiology Ecology, 70(2): 236–248, doi: 10.1111/j.1574-6941.2009.00772.x

General Administration of Quality Supervision, Inspection and Quarantine of the People's Republic of China, Standardization Administration of China. 2008. GB 17378.5-2007 The specification for marine monitoring—Part 5: sediment analysis (in Chinese). Beijing: Standards Press of China, 1–104

Gomez-Alvarez V, Teal T K, Schmidt T M. 2009. Systematic artifacts in metagenomes from complex microbial communities. The ISME Journal, 3(11): 1314–1317, doi: 10.1038/ismej.2009.72

Guan Xiangyu, Zhu Lingling, Li Youxun, et al. 2014. Composition and variation of sediment bacterial and *nirS*-harboring bacterial communities at representative sites of the Bohai Gulf coastal zone, China. World Journal of Microbiology and Biotechnology, 30(4): 1291–1300, doi: 10.1007/s11274-013-1553-4

Guo Xingpan, Lu Dapei, Niu Zuoshun, et al. 2018. Bacterial community structure in response to environmental impacts in the intertidal sediments along the Yangtze Estuary, China. Marine Pollution Bulletin, 126: 141–149, doi: 10.1016/j.marpolbul.2017.11.003

Gutierrez T. 2017 Aerobic Hydrocarbon-degrading *Gammaproteobacteria*: *Xanthomonadales*. In: McGenity T, ed. Taxonomy, Genomics and Ecophysiology of Hydrocarbon-Degrading Microbes. Cham: Springer, 1–15

Jing Xiaoyan, Gou Honglei, Gong Yanhai, et al. 2018. Raman-activated cell sorting and metagenomic sequencing revealing carbon-fixing bacteria in the ocean. Environmental Microbiology, 20(6): 2241–2255, doi: 10.1111/1462-2920.14268

Kim Y O, Park S, Nam B H, et al. 2014. Description of *Lutimonas halo-*

cynthiae sp. nov., isolated from a golden sea squirt (*Halocynthia aurantium*), reclassification of *Aestuariicola saemankumensis* as *Lutimonas saemankumensis* comb. nov. and emended description of the genus *Lutimonas*. International Journal of Systematic and Evolutionary Microbiology, 64(6): 1984–1990

Könneke M, Kuever J, Galushko A, et al. 2013. *Desulfoconvexum algidum* gen. nov., sp. nov., a psychrophilic sulfate-reducing bacterium isolated from a permanently cold marine sediment. International Journal of Systematic and Evolutionary Microbiology, 63(3): 959–964

Köster M, Meyer-Reil L A. 2001. Characterization of carbon and microbial biomass pools in shallow water coastal sediments of the southern Baltic Sea (Nordrügensche Bodden). Marine Ecology Progress Series, 214: 25–41, doi: 10.3354/meps214025

Kuever J, Rainey F A, Widdel F. 2015. Desulfobulbus. In: Whitman W B, DeVos P, Chun J, et al., eds. Bergey's Manual of Systematics of Archaea and Bacteria. Hoboken: John Wiley & Sons, Inc, 1–6

Li Hongjun, Ye Sheng, Ye Jinqing, et al. 2017. Baseline survey of sediments and marine organisms in Liaohe Estuary: heavy metals, polychlorinated biphenyls and organochlorine pesticides. Marine Pollution Bulletin, 114(1): 555–563, doi: 10.1016/j.marpolbul.2016.09.002

Liu Zongzhi, DeSantis T Z, Andersen G L, et al. 2008. Accurate taxonomy assignments from 16S rRNA sequences produced by highly parallel pyrosequencers. Nucleic Acids Research, 36(18): e120, doi: 10.1093/nar/gkn491

Liu Jiwen, Liu Xiaoshou, Wang Min, et al. 2015. Bacterial and archaeal communities in sediments of the north Chinese marginal seas. Microbial Ecology, 70(1): 105–117, doi: 10.1007/s00248-014-0553-8

Lofrano G, Libralato G, Minetto D, et al. 2017. In situ remediation of contaminated marinesediment: an overview. Environmental Science and Pollution Research, 24(6): 5189–5206, doi: 10.1007/s11356-016-8281-x

Lv Xiaofei, Ma Bin, Yu Junbao, et al. 2016. Bacterial community structure and function shift along a successional series of tidal flats in the Yellow River Delta. Scientific Reports, 6: 36550, doi: 10.1038/srep36550

Marshall I P G, Karst S M, Nielsen P H, et al. 2018. Metagenomes from deep Baltic Sea sediments reveal how past and present environmental conditions determine microbial community composition. Marine Genomics, 37: 58–68, doi: 10.1016/j.margen.2017.08.004

Martiny J B H, Eisen J A, Penn K, et al. 2011. Drivers of bacterial β-diversity depend on spatial scale. Proceedings of the National Academy of Sciences of the United States of America, 108(19): 7850–7854, doi: 10.1073/pnas.1016308108

Ministry of Environmental Protection of the People's Republic of China. 2012. HJ 634-2012 Soil-determination of ammonium, nitrite and nitrate by extraction with potassium chloride solution-spectrophotometric methods (in Chinese). Beijing: China Environmental Press, 1–18

Miroshnichenko M L, Kostrikina N A, Chernyh N A, et al. 2003. *Caldithrix abyssi* gen. nov., sp. nov., a nitrate-reducing, thermophilic, anaerobic bacterium isolated from a Mid-Atlantic Ridge hydrothermal vent, represents a novel bacterial lineage. International Journal of Systematic and Evolutionary Microbiology, 53(1): 323–329, doi: 10.1099/ijs.0.02390-0

Muller G. 1969. Index of geoaccumulation in sediments of the Rhine River. Geojournal, 2(3): 108–118

Müller G, Förstner U. 1976. Schwermetalle in den Sedimenten der Elbe bei Stade: veränderungen seit 1973. Naturwissenschaften, 63(5): 242–243, doi: 10.1007/BF00610914

Park S, Yoshizawa S, Inomata K, et al. 2012. *Halioglobus japonicus* gen. nov., sp. nov. and *Halioglobus pacificus* sp. nov., members of the class *Gammaproteobacteria* isolated from seawater. International Journal of Systematic and Evolutionary Microbiology, 62(8): 1784–1789

Quince C, Lanzén A, Curtis T P, et al. 2009. Accurate determination of microbial diversity from 454 pyrosequencing data. Nature Methods, 6(9): 639–641, doi: 10.1038/nmeth.1361

Shade A. 2018. Understanding microbiome stability in a changing world. Msystems, 3(2): e00157-17, doi: 10.1128/mSystems. 00157-17

Sinkko H, Lukkari K, Sihvonen L M, et al. 2013. Bacteria contribute to sediment nutrient release and reflect progressed eutrophication-driven hypoxia in an organic-rich continental sea. PLoS One, 8(6): e67061, doi: 10.1371/journal.pone.0067061

Su Zhiguo, Dai Tianjiao, Tang Yushi, et al. 2018. Sediment bacterial community structures and their predicted functions implied the impacts from natural processes and anthropogenic activities in coastal area. Marine Pollution Bulletin, 131: 481–495, doi: 10.1016/j.marpolbul.2018.04.052

Tian Qi, Wang Jia, Fan Xiaolei, et al. 2014. Methanogenic activity and methanogen diversity in marine gas field sediments. Environmental Science (in Chinese), 35(6): 2322–2327

Vandieken V, Pester M, Finke N, et al. 2012. Three manganese oxide-rich marine sediments harbor similar communities of acetate-oxidizing manganese-reducing bacteria. The ISME Journal, 6(11): 2078–2090, doi: 10.1038/ismej.2012.41

Wada M, Suzuki S, Nara T, et al. 2012. Microbial community respiration and structure of dead zone sediments of Omura Bay, Japan. Journal of Oceanography, 68(6): 857–867, doi: 10.1007/s10872-012-0136-6

Wang Haitao, Gilbert J A, Zhu Yongguan, et al. 2018. Salinity is a key factor driving the nitrogen cycling in the mangrove sediment. Science of the Total Environment, 631–632: 1342–1349, doi: 10.1016/j.scitotenv.2018.03.102

Wang Yafen, Tam N F Y. 2012. Natural attenuation of contaminated marine sediments from an old floating dock Part II: changes of sediment microbial community structure and its relationship with environmental variables. Science of the Total Environment, 423: 95–103, doi: 10.1016/j.scitotenv.2012.01.066

Woese C R. 1987. Bacterial evolution. Microbiological Reviews, 51(2): 221–271

Yang Yonghua, Yao Jian, Hua Xiaomei. 2000. Effect of pesticide pollution against functional microbial diversity in soil. Journal of Microbiology (in Chinese), 20(2): 23–25, 27

Zhang Huizhen, Chang Yongkai, Chen Quanrui, et al. 2018. Community structure analysis of Archaea and bacteria in sediments of Liaohe Estuary. Haiyang Xuebao (in Chinese), 40(6): 113–130

Zhang Yao, Zhao Zihao, Dai Minhan, et al. 2014. Drivers shaping the diversity and biogeography of total and active bacterial communities in the South China Sea. Molecular Ecology, 23(9): 2260–2274, doi: 10.1111/mec.12739

Zhao Yangguo, Feng Gong, Bai Jie, et al. 2014. Effect of copper exposure on bacterial community structure and function in the sediments of Jiaozhou Bay, China. World Journal of Microbiology and Biotechnology, 30(7): 2033–2043, doi: 10.1007/s11274-014-1628-x

Zheng Binghui, Wang Liping, Liu Lusan. 2014. Bacterial community structure and its regulating factors in the intertidal sediment along the Liaodong Bay of Bohai Sea, China. Microbiological Research, 169(7–8): 585–592, doi: 10.1016/j.micres.2013.09.019

Zhou Kan, Fan Jie, Liu Hanchu. 2017. Spatiotemporal patterns and driving forces of water pollutant discharge in the Bohai Rim Region. Progress in Geography (in Chinese), 36(2): 171–181, doi: 10.18306/dlkxjz.2017.02.004

Zinger L, Amaral-Zettler L A, Fuhrman J A, et al. 2011. Global patterns of bacterial beta-diversity in seafloor and seawater ecosystems. PLoS One, 6(9): e24570, doi: 10.1371/journal.pone.0024570