

Strain-Related Differences in the Immune Response: Relevance to Human Stroke

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Abstract There are significant differences in the immune response and in the susceptibility to autoimmune diseases among rodent strains. It would thus be expected that the contribution of the immune response to cerebral ischemic injury would also differ among rodent strains. More importantly, there are significant differences between the immune responses of rodents and humans. All of these factors are likely to impact the successful translation of immunomodulatory therapies from experimental rodent models to patients with stroke.

Keywords Stroke · Immunology · EAE · Strain · Immune response

There is a variability among infarct size and stroke outcome in different strains of mice and rats subjected to similar ischemic insults [1–6]. Much of the literature suggests that there are differences in cerebrovascular anatomy/cerebral blood flow among these strains that leads to the variability in infarct size [2, 7–10]. Little attention has been given to systemic factors that might influence infarct development and functional recovery. Increasing data show that the immune system plays a critical role in the response to cerebral ischemic injury. Following the onset of cerebral ischemia, neutrophils, monocytes, and lymphocytes infiltrate the brain [11, 12]. There is also a marked systemic inflammatory response immediately after stroke, which is paradoxically accompanied by a depression in cellular immune responses that predispose to infection

[13, 14]. Despite the defect in lymphocyte responses, humoral and cellular immune responses to central nervous system (CNS) antigens can be detected in the weeks to months after stroke [15–24]. Understanding how the immune response affects stroke outcome and how it can be manipulated to improve outcome is an active area of research, and the results from this line of research would seemingly be dependent upon the immunologic background in which these phenomena are studied. This review will highlight differences in the immune response among rodent strains and discuss the implications for stroke research and its translation into the clinical arena.

Strain Differences—Lessons from EAE

Rats

Susceptibility to experimental allergic encephalomyelitis (EAE) by active immunization with myelin basic protein (MBP) differs significantly among rodent strains (Table 1). Lewis rats and Dark Agouti (DA) rats are highly susceptible to EAE while strains such as Fischer (F344), Brown Norway (BN), Wistar-Kyoto (WKY), and Piebald Vireo Glaxo (PVG) are poorly susceptible [25–28]. Even among EAE-susceptible Lewis rats, the relative ease of inducing EAE may differ between commercial vendors [29]. And while Wistar and Sprague-Dawley rats are capable of developing EAE, they are used less commonly than Lewis and DA rats to study this autoimmune disease as well as other immunologically mediated diseases [30–36]. The susceptibility of rats to autoimmune diseases of the peripheral nervous system parallels that of EAE, with Lewis rats being relatively susceptible to experimental allergic neuritis (EAN) and BN rats being relatively resistant [37, 38]. Wistar and Sprague-Dawley rats are also susceptible to EAN but develop less severe disease than

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Table 1 Relative propensities for developing inflammatory/autoimmune disease among commonly used rodent strains

	Mouse strains	Rat strains
High susceptibility	SJL/J C3H/He	Lewis Dark Agouti (DA)
Intermediate susceptibility	C57BL/6 CBA	Sprague-Dawley (SD) Wistar Buffalo
Poor susceptibility	DBA/2 B10.S BALB/c CD1	Fischer (F344) Brown Norway (BN) Wistar-Kyoto (WKY) Piebald Virol Glaxo (PVG)

Lewis rats [37]. Because Lewis and DA rats are more prone to develop EAE and EAN, as well as autoimmune diseases that do not affect the nervous system, it suggests that the mechanisms predisposing these strains to autoimmunity are systemic and not unique to the nervous system [39–42].

There appear to be fundamental differences in the immune response among rat strains that contribute to the differing susceptibilities to EAE. For instance, susceptible Lewis rats express more major histocompatibility complex (MHC) II on their astrocytes than resistant strains like BN [27, 43]. The relative numbers of leukocyte subsets also varies among strains; in comparison to BN rats, Lewis rats have more mast cells in the CNS (and PNS) [44]. And EAE-resistant F344 rats have increased numbers of MBP specific CD8+ regulatory T cells compared to EAE-susceptible Lewis rats [45]. Further, cytokine profiles differ among strains and correlate with the ability to induce EAE. Within astrocytes, gene expression for pro-inflammatory cytokines like tumor necrosis factor (TNF)- α is associated with increased susceptibility to EAE, while production of immunomodulatory cytokines like transforming growth factor (TGF)- β 1 is associated with protection from EAE [46, 47]. EAE-resistant and EAE-susceptible rats also differ in the production of TH1 and TH17 cytokines, with susceptible rats expressing more interferon (IFN)- γ , IL-17, and IL-12 in the draining lymph nodes following immunization [48, 49]. Other factors may also contribute to the susceptibility or resistance to EAE, including IL-2 production (lower levels are protective) [50] and endogenous nitric oxide (higher levels are protective) [51].

Given the interplay between the brain, the immune system, and the endocrine system, it is not surprising that there are differences in the neuroendocrine response to stress that influence the susceptibility to EAE. Among wild-type rats, behavioral characteristics like aggression (“attack latency time”) predict susceptibility to EAE [52]. And among inbred rats, EAE resistant strains (PVG, F344, and BN) exhibit higher levels of endogenous steroid hormones, including corticosterone, than EAE-susceptible strains [27, 53–55]. As might be expected, hormonal manipulations affect susceptibility to

EAE, with adrenalectomy rendering the normally EAE-resistant PVG rats susceptible to severe disease [55]. Further, blockade of endogenous glucocorticoids with RU-486 leads to worsened severity of EAE in strains that are already susceptible (i.e., Lewis rats) [56].

EAE-susceptible rat strains also differ with regards to the relative antigenicity of different myelin epitopes [57], and strains resistant to active induction of EAE (i.e., by immunization) may be susceptible to EAE induction by adoptive transfer of lymphocytes [58]. Both of these factors (the antigen/epitope used to induce EAE and whether the disease was induced by active immunization or adoptive transfer) affect the clinical presentation and the neuropathology of the disease [58–60].

Mice

The SJL/J and C3H/He strains of mice are highly susceptible to EAE following immunization with MBP, while the DBA/2, B10.S, and BALB/c strains are relatively resistant (Table 1) [61–63]. Like Lewis rats, SJL mice express more MHC II on their astrocytes than EAE-resistant strains (BALB/c) [43]. That the differences in susceptibility are due (at least in part) to inherent differences in the immune response is demonstrated by the fact that the normally resistant BALB/c mice can be rendered susceptible to active induction of EAE by inhibition of cytotoxic T lymphocyte antigen-4 (CTLA-4), which prevents an inhibitory signal from being delivered to the T cell [64]. The cytokine profiles of mouse strains, like rat strains, also predict the susceptibility to EAE. In general, EAE-susceptible mouse strains tend to have a TH1-type phenotype, while EAE resistant strains have a TH2/TREG phenotype [65–67]. And again, paralleling the situation in rats, neuroendocrine responses play a role in determining the susceptibility of mice to EAE. For example, EAE susceptible SJL/J mice, similar to Lewis rats, have a blunted response of the hypothalamic-pituitary-adrenal axis to stress, contributing to EAE susceptibility [68].

C57BL/6 mice are resistant to the active induction of EAE following immunization with MBP but are susceptible to EAE induced by the myelin oligodendrocyte glycoprotein 35–55 peptide (MOG_{35–55}); CD1 mice, on the other hand, are resistant to EAE induced with MOG_{35–55} [69]. The difference in susceptibility to EAE following immunization with MOG_{35–55} in these strains appears to be related, in part, to the fact that C57BL/6 mice have a higher percentage of CD4+ T cells that produce IFN- γ and IL-17, as well as increased systemic IL-17 [69]. Increased numbers of regulatory B and T cells may also contribute to the resistance of CD1 mice to EAE [69].

EAE is a model of multiple sclerosis (MS). It is an imperfect model, but it has provided insights into both the systemic and CNS immune responses in rodents. Another rodent model of MS results from injection of Theiler’s murine

encephalomyelitis virus (TMEV) into the brains of susceptible mouse strains. Resistant strains are able to clear the virus while susceptible strains become persistently infected. The persistent infection leads to a chronic demyelinating disease that is used as a surrogate of MS [70]. The susceptibility to TMEV among mouse strains resembles that of the susceptibility to EAE. The disease is most severe in SJL mice, of intermediate severity in CBA and C3H/He strains, and least severe in C57BL/6 mice [71]. BALB/c mice are resistant to the neuropathology of TMEV [72]. During CNS infection with TMEV, there are increases in TREG and CD45r(+) B cells, delays in viral elimination and increases in IL-10 messenger RNA (mRNA) in susceptible SJL mice compared to C57BL/6 mice [73]. Further, the astrocytes of susceptible strains appear to produce more IL-1 α than the astrocytes of resistant strains [74]. And as is the case with EAE, the susceptibility to TMEV correlates with the ease of induction of MHC II on astrocytes [75].

Age and Gender Considerations in EAE

The susceptibility to both EAE and TMEV decreases with age [76–81]. The age-related decline in susceptibility to EAE and TMEV is related to the senescence of the immune response that occurs with aging [80, 82]. Sex and sex hormones also affect the susceptibility to EAE, TMEV, and other autoimmune diseases [83–88]. These facts highlight the need for using age- and sex-appropriate models when studying the immune response and also suggest that the relative benefits of immunomodulatory therapies are likely to vary depending on age and sex.

Psychiatric Disorders: Immune and Strain-Related Issues

Numerous psychiatric disorders are posited to have an immunologic basis. Chief among these disorders is depression [89]. Given strain-related differences in the immune response, it is reasonable to assume that there would be strain related differences in the modeling of psychiatric disorders. For instance, administration of LPS leads to long-term depressive-like behavior in C57BL/6 mice but not in CD1 mice [90]. And in comparison to C57BL/6 mice, BALB/c mice have increased immune activation following LPS injection with an increase in depressive behavior [91]. And following an LPS challenge, the microglia of high-anxiety inbred mice (DBA/2J and 129S2/Sv) are polarized to an M1 phenotype relative to mice without anxiety [92]. We also showed that there were strain-related differences in “fatigue” and “depression” after stroke with Lewis rats displayed depressive-like behavior while Wistar and SD rats displayed fatigue-like behavior [93]. These examples just skim the surface of a robust literature

showing an association between the immune response and psychiatric disease, and thus, the relationship between animal strains and the modeling of psychiatric disorders.

The Immunology of Stroke: Strain-Related Issues

Based on the literature that shows fundamental differences in the immune response among rodent strains, one would expect that the immune response following ischemic brain injury would also differ among strains. The relative benefit of immunomodulatory interventions for the treatment of stroke would thus be expected to differ as well. When studying immune mechanisms in stroke, one should ideally evaluate both EAE-susceptible and EAE-resistant strains to best understand the true contribution of the immune response to stroke outcome and the likelihood that modulating that response would be of benefit. For instance, we showed that the long-term behavioral outcomes among Lewis, Sprague-Dawley, and Wistar rats were quite different despite similar infarct volumes at 24 h after middle cerebral artery occlusion (MCAO) [94]. At 1 month after MCAO, the cytokine profiles among these three strains differed with less circulating IL-1 α in Sprague-Dawley rats and more circulating IL-10 in Lewis rats [93]; many of the behaviors assessed at this time point correlated significantly with these cytokine levels.

Wistar and Sprague-Dawley rats, strains with intermediate susceptibility to EAE, are arguably the most common rat strains used in stroke research. Very few stroke studies are done in EAE-resistant F344 rats (with some authors reporting that the strain is unsuitable for the filament model of MCAO) [95]. Our group has used EAE-susceptible Lewis rats almost exclusively for studies that address the post-ischemic immune response. Upon noticing variation in stroke outcomes after switching vendors, we began to take note of differences in what is supposed to be an inbred rat strain. For instance, there were significant differences in baseline temperatures as well as differences in the magnitude of temperature changes following MCAO (Fig. 1). At least one plausible explanation for such differences among inbred animals between vendors is that of differences in the gut microbiome [96]. Given that the gut microbiome plays an important role in driving the immune response and triggering autoimmune disease [97–101], vendor differences in diet and the microbiological milieu may significantly affect the post-ischemic immune response as well as responses to immunomodulatory therapy.

With limited exceptions, virtually all of the studies examining immunologic changes after stroke in mice, as well as the response to immunologic therapies, have been done in the C57BL/6 strain, which has intermediate susceptibility to EAE. One of the obvious reasons for using C57BL/6 mice is the ability to create transgenic animals from this genetic background. What this means practically, however, is that almost

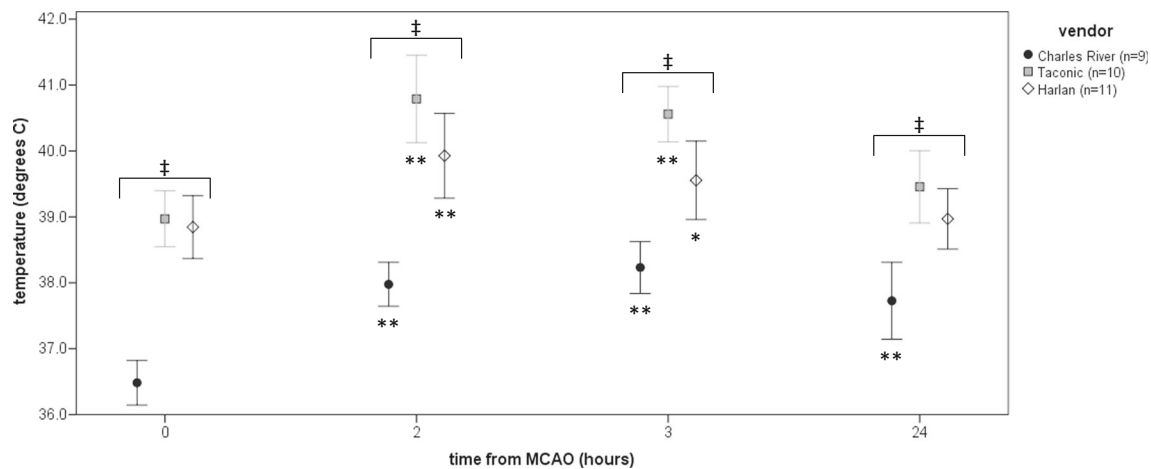


Fig. 1 Temperatures after MCAO. Data are plotted as the mean (\pm standard deviation). Differences in temperature between vendors is assessed by ANOVA and noted by ‡ ($p < 0.001$). Differences from

baseline temperatures after MCAO among animals from a single vendor is assessed by paired *t* test and noted by * $p < 0.05$) and ** $p < 0.01$

all of what we know about the role of the immune system in stroke is derived from a single line of inbred mice. And a recent study showed that despite similar infarct volumes after MCAO in C57BL/6 and FVB mice (in which EAE can be reproducibly induced [102]), there were significant differences in neurological outcome and the immune response between these strains [103]. That the infarct volumes were similar between these strains is important, given that the volume of ischemic tissue injury will undoubtedly affect the nature and strength of the immune response. Relative to FVB mice, C57BL/6 mice had a more robust increase in leukocytes within the brain at 24 h after MCAO. And within C57BL/6 mice the post-ischemic leukocyte infiltration of the brain was composed mostly of lymphocytes, while in FVB mice it was composed mostly of neutrophils and myeloid cells. In addition, the systemic effects of stroke differed by strain, with a decrease in splenic T cells in FVB mice, but not C57BL/6 mice, 24 h after MCAO. These data again highlight the need to evaluate different strains of mice when studying the contribution of the immune response to ischemic brain injury.

Susceptibility to Infection

In a mouse model of stroke, sympathetically mediated suppression of TH1-type immune responses was associated with a predisposition to infection [104]. These data were generated in SV129/J and C57BL/6 J mice. Subsequent studies showed that SV129 mice are much more susceptible to infection following MCAO than either C57BL/6 or BALB/C mice [105]. As it turns out, mouse strains differ markedly in their resistance and response to infection in general, independent of stroke. For instance, given identical pneumococcal infections of the respiratory tract, BALB/c mice demonstrate no bacteremia or death, CBA/Ca, C3H/He, and SJL mice develop severe bacteremia with 100 % mortality, and C57BL/6 and

DBA/2 strains develop only modest bacteremia with approximately 50 % mortality [106]. One would thus expect that not only the susceptibility to infection following stroke would differ among different strains, but that the clinical manifestation of those infections might differ as well.

As mentioned, the increased risk of infection following stroke appears to be, at least in part, mediated by the sympathetic nervous system (SNS) [104, 107]. That the SNS can affect the immune response has been appreciated for some time [108, 109]. The effect of sympathetic activation on the immune response also appears to be strain dependent in that chemical sympathectomy increases mitogen-induced lymphocyte proliferation in EAE resistant DBA/2 mice but not in EAE susceptible C57BL/6 mice [110]. Further, systemic epinephrine levels differ with the strain as well as the age of rat, suggesting that modulation of the sympathetic response may affect the immune response, the susceptibility to infection, and the chance of developing autoimmunity/sustained inflammation after stroke in a strain- and age-dependent fashion [111, 112].

Of Mice and Men

Highly conserved molecular motifs in pathogens, termed pathogen-associated molecular patterns (PAMPs), activate the innate immune response through toll-like receptors (TLRs). TLR-4, for example, is activated by endotoxin/lipopolysaccharide (LPS), a component of the Gram-negative bacterial cell wall. The sensitivity of TLR4 to endotoxin/LPS is vastly different among rodents and humans, with the immune system of rodents favoring “tolerance” to immunologic threats while that of humans favors “resistance” to similar threats [113, 114]. For example, administration of endotoxin intravenously at doses of 15 $\mu\text{g}/\text{kg}$ leads to a severe

systemic response with shock in humans [115, 116]. In mice, on the other hand, the median lethal dose of endotoxin/LPS is in the range of 10–12 mg/kg [117, 118]. And the dose endotoxin/LPS needed to elicit similar systemic levels of IL-6 is about 200-fold higher in mice than in humans [119]. Not surprisingly, the response to endotoxin in rodents is also strain dependent [120]. Similarly, genetic variations in humans (like single nucleotide polymorphisms [SNPs]) modulate the responsiveness of TLR4 to endotoxin and the course of infection/sepsis [121–124].

TLRs are also activated by danger-associated molecular patterns (DAMPs) released from injured cells. The post-ischemic inflammatory response is a sterile response that is undoubtedly set in motion by DAMPs from necrotic tissue. These DAMPs include high-mobility group box (HMGB)-1, adenosine, ATP, uric acid, heat-shock proteins, heparin sulfate, hyaluronan fragments, and DNA [125, 126]. Based on what is known about the differences in the responsiveness to PAMPs, it would not be surprising if there were similar differences in the relative responsiveness to DAMPs between rodents and humans as well as among rodent strains. Additionally, genetic variation among humans would likely influence the responsiveness of the innate immune system to these endogenous DAMPs.

The dissimilarity in the response to sterile injury between rodents and humans is highlighted by the fact there is essentially no correlation between the genomic profiles of mice and humans subjected to blunt force trauma or burn injury [127]. Likewise, the correlation between gene expression in human volunteers treated with LPS and mice treated with LPS is essentially random [127]. These observations highlight the fact that the response to both PAMPs and DAMPs differ significantly between humans and rodents.

Another important difference between the immune system of *Homo sapiens* and rodents is illustrated by the difference in the composition of circulating leukocytes. In C57BL/6 mice, for instance, the majority of circulating cells are lymphocytes (75–90 %) while only 20–40 % of circulating leukocytes in humans are lymphocytes [128, 129]. In humans, most WBCs in circulation are neutrophils (50–70 %), and these neutrophils are not only more numerous than in rodents but also have different properties than rodent neutrophils [114].

Inhibition of α -4 for the Treatment of Stroke

Lymphocytes and monocytes express the integrins α 4 β 1 (CD49d/CD29) and α 4 β 7 (CD49d/CD103), both of which are important for cell trafficking [130]. CD49d/CD29 is also known as very late activation antigen-4 (VLA-4), and CD49d/CD103 is also known as lymphocyte-Peyer's patch adhesion molecule-1 (LPAM-1). Binding of lymphocytes and monocytes to the endothelium occurs through the interaction of

the α 4 integrins with either vascular cell adhesion molecule-1 (VCAM-1) or mucosal addressin cell adhesion molecule-1 (MAdCAM-1). In 2001, we showed that inhibition of the α 4 integrin with blocking antibodies (TA-2) decreased infarct volume and improved outcome at 48 h after MCAO in Lewis rats [131]. Treatment with the same antibody was shown to decrease infarct volume at 24 h after MCAO in both Sprague-Dawley and spontaneously hypertensive rats (SHR) [132]. Subsequent studies showed that inhibition of VLA-4 (with a different antibody) in C57BL/6 J mice improved outcome in moderately severe stroke [133], although subsequent studies in C57BL/6 mice with the same antibody showed no benefit [134]. While the difference in outcomes in these studies may have been due to chance alone, it is also clear that not all C57BL/6 mice are the same and there may be important genetic and behavioral differences between substrains [135–137]. In a recent “multi-center study” using young male C57BL/6 mice, inhibition of CD49d was found to decrease infarct volume and improve outcome in mild stroke (permanent cortical ischemia induced by electrocoagulation) but not in severe stroke (transient MCAO induced by an intraluminal filament) [138]. Based on inherent differences in the immune response within a species (not to mention between species), the outcomes of this study may have been different if the strokes were performed in different mouse strains (ones more predisposed to inflammation/autoimmune disease), mice with a different gut microbiome, older mice, or female mice. In particular, female mice are noted to express less VLA-4 in the spleen and brain after MCAO compared to male rats [139].

Of note, rodents are known to express VLA-4 on neutrophils as well as on monocytes and lymphocytes; inhibition of α 4 thus blocks the migration of all of these cell types in rodents [140, 141]. The influx of PMNs (as well as lymphocytes and monocytes) into the rodent brain following stroke would thus be inhibited by antibodies that block VLA-4 [142]. In contrast to rodents, there are relatively more circulating PMNs in humans, yet these PMNs do not express appreciable α 4 [143, 144]. Inhibition of α 4 may, therefore, not prevent the influx of PMNs into the human brain after stroke, suggesting that the effect/benefit of VLA-4 inhibition might differ in rodents and humans.

Summary

Characterizing the immunology of ischemic stroke in a single rodent strain provides information only about the immunologic consequences of stroke in that rodent strain. Predicting the clinical response of patients to an immunological intervention from studies done in a single strain of inbred mice or rats would, therefore, seem destined to fail. Not only are there critical differences between the immune responses of different rodent strains, there are even larger differences between the

immune response of rodents and *Homo sapiens* [114, 145–147]. And as opposed to inbred rodent strains, humans are genetically distinct from each other and have a wide range of genetic polymorphisms that influence the immune response, which suggests that the efficacy of a given intervention might differ from person to person. In summary, these observations suggest that the use of animal models for understanding the development of the post-ischemic inflammatory response has serious limitations and that translation of effective immunomodulatory therapies from rodents to humans is likely to be unreliable. While rodent models of stroke may never adequately address what happens in humans, it would seem that, at a minimum, studies of immunologic therapies for stroke be conducted in strains with different propensities for the development of inflammatory/immunologic diseases.

Compliance with Ethical Standards

Conflict of Interest The authors declare that they have no competing interests.

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