METHODS AND RESOURCES ARTICLE

Environmental DNA metabarcoding reveals the biological community structure in Poyang Lake, China

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Abstract

Poyang Lake is the largest freshwater lake in China, and its unique ecosystem plays an important role in maintaining biodiversity. However, the biodiversity of Poyang Lake is facing a serious threat as a result of human activities. Species investigation is the basis of biodiversity protection. In order to improve the water ecological monitoring system and achieve efficient and non-invasive species monitoring, environmental DNA metabarcoding was used in this study to assess biodiversity in diferent habitats in Poyang Lake. A total of 45 species including 31 fsh species and six bivalve species were detected in water samples collected from 29 sampling sites in six habitats in Poyang Lake (Jiangxi section of the mainstream of the Yangtze River, channel connecting Poyang Lake and the Yangtze River, main lake area of Poyang Lake, Nanjishan Nature Reserve, Junshan Lake and Qinglan Lake). The species were detected through a standardized process involving water sample collection, fltration, environmental DNA extraction, genetic marker amplifcation, high-throughput sequencing and bioinformatics analysis. The 31 fsh species, which included 11 alien species, were mainly cyprinids and lake dwellers. Alpha diversity analysis indicated a decline in biodiversity in Poyang Lake habitats and a serious need for water ecology conservation. Beta diversity analysis revealed signifcant diferences in the biotic community structures of the six habitats in Poyang Lake. The results align with those obtained with more traditional methods, and hence, environmental DNA metabarcoding can be used as an alternative biodiversity monitoring tool for rapid detection of the diversity and spatial distribution of organisms (especially fsh). This technique provides a new toolkit for biodiversity monitoring and aquatic ecological conservation in Poyang Lake area.

Keywords Environmental DNA · Biodiversity · Habitat degradation · Freshwater

Introduction

The loss of biodiversity is one of the most serious environmental crises facing the world, and the biodiversity decline in freshwater populations is even greater than that in marine and terrestrial ecosystems (Valentini et al. [2016\)](#page-11-0). The development of rapid and efective tools to monitor biodiversity fuctuations is the focus of scientifc research on conservation and management strategies. Traditional survey methods have been invaluable for monitoring aquatic biodiversity

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¹ School of Life Sciences, Nanchang University, Nanchang 330031, People's Republic of China and developing management and conservation strategies, but they also have many shortcomings, such as inefficiency, selectivity, destructiveness or strict reliance on declining taxonomic expertise (Wheeler et al. [2004](#page-11-1)).

The recently developed environmental DNA metabarcoding technology can directly extract DNA from environmental samples (such as water, sediment, soil, etc.), apply universal primers for target groups, and identify multiple target species in environmental samples through polymerase chain reaction (PCR) amplifcation combined with high-throughput sequencing (Taberlet et al. [2012\)](#page-10-0). There is no need to collect target organisms. The advantages of non-destructive sampling and high detection sensitivity compensate for the defciencies of traditional morphological monitoring. The environmental DNA metabarcoding technology has great application potential for biodiversity assessment (Thomsen and Willerslev [2015](#page-11-2)). In recent years, environmental DNA metabarcoding has

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been widely used for fsheries management and diversity monitoring in freshwater ecosystems (Ruppert et al. [2019](#page-10-1)). A variety of amphibians, fsh, mammals, insects and crustaceans have been found in environmental DNA biodiversity surveys in hundreds of ponds, streams and rivers in the USA (Thomsen et al. [2012\)](#page-10-2). Evans et al. ([2017\)](#page-10-3) used three pairs of common fish primers to detect all fish species previously identifed in a reservoir using traditional methods as well as 11 previously uncaptured fsh species. Zhang et al. ([2020\)](#page-11-3) systematically evaluated the infuence of spatial sampling design on the fsh community structure in three lakes of diferent sizes based on environmental DNA technology, and the results confrmed that shoreline sampling was equally effective.

Located on the south bank of the middle and lower reaches of the Yangtze River, Poyang Lake is the largest freshwater lake in China and one of only three lakes connected to the Yangtze River (Zhang and Li [2007](#page-11-4)). Water flows into Poyang Lake from five major rivers; the Ganjiang, Fuhe, Xinjiang, Raohe and Xiushui Rivers. The water then passes into the Yangtze River in the Hukou region after being regulated and stored in the lake (Zhang [1993\)](#page-11-5). In recent years the increase in human activities, especially the overexploitation and utilization of freshwater resources (e.g. reclaiming land from lakes, sand mining, damming rivers, dike breeding, overfshing, etc.,) has caused serious damage to the freshwater ecosystem in Poyang Lake. These activities place the lake at risk of degradation and have seriously affected the biodiversity (especially fish species), resulting in signifcant changes in the community structure (Hu et al. [2011;](#page-10-4) Chen et al. [2012\)](#page-10-5). The biodiversity and ecological balance of Poyang Lake have attracted close attention from the state and Federal government. In recent years, relevant measures have been taken to ban fshing in Poyang Lake (Xu et al. [2020](#page-11-6)).

There is an urgent need to establish a rapid, effective, and environmentally friendly monitoring system for biodiversity conservation and ecological restoration in Poyang Lake. Baseline data can provide a scientifc foundation for the formulation and implementation of fshery management and ecological protection policies.

In this study, environmental DNA metabarcoding technology was used for the frst time to investigate the structure and constitutive mechanisms of biological communities in diferent habitats in Poyang Lake. This study can provide the required baseline data to enable better understanding of the degradation of the lake fshery habitat and the evolution of biodiversity patterns. It can also provide important data for the restoration of the lake habitat and biodiversity, and provide a theoretical framework for the conservation and adaptive management of important fshery lakes.

Materials and methods

Study area

Poyang Lake is a seasonal lake formed by water from the Yangtze River and five other rivers, with obvious flood and drought rhythms. It covered an area of about 2000 km^2 at the time of sampling. We divided Poyang Lake into six habitats (Jiangxi section of the mainstream of the Yangtze River, channel connecting Poyang Lake and the Yangtze River, main lake area of Poyang Lake, Nanjishan Nature Reserve, Junshan Lake and Qinglan Lake) (Fig. [1](#page-2-0)).

Environmental DNA sampling

In this study, samples were collected from Poyang Lake in April 2019. According to the geographical environment, hydrology and habitat characteristics of Poyang Lake, a total of 29 sampling sites were set up in six habitats (Fig. [1\)](#page-2-0). A 1 L surface water sample was collected from each site in a sterile plastic bottle and stored at -4 °C before fltration. We conducted a study on the vertical distribution diferences for the environmental DNA diversity of fsh in Poyang Lake, and the results showed that there were no signifcant diferences (unpublished). Three duplicate samples were collected at each site. To reduce contamination between the sites, the surveyor followed a sterile protocol, e.g. wearing sterile gloves at each site, and the equipment was sterilized between each site. We flled the sample bottles with sterilized water to ensure there was no contamination on site or in the bottles.

Water sample fltration and environmental DNA extraction

The water was fltered within 24 h through a 0.45-μm mixed cellulose flter membrane (Jinteng, China) with an oil-free vacuum pump (Rocker 300, Taiwan). In order to assess the possible presence of exogenous DNA contamination, a feld blank control was used during fltration. The membranes were folded and preserved in sterile 1.5 mL centrifuge tubes at -20 °C. Environmental DNA was extracted from the samples and blanks using the DNeasy Blood & Tissue Kit (Qiagen, Germany) according to the method described by Zhang et al. (Zhang et al. [2020](#page-11-3)). Three extractions were performed for each site. A blank membrane was used as the negative control.

Fig. 1 Sampling sites at six habitats in Poyang Lake basin. Jiangxi section of the mainstream of the Yangtze River (A1–A3), channel connecting Poyang Lake and the Yangtze River (B1–B8), main lake area of Poyang Lake (C1–C8), Nanjishan Nature Reserve (D1–D3), Junshan Lake (E1–E5), and Qinglan Lake (F1–F2)

Amplifcation and high‑throughput sequencing

Mitochondrial cytochrome b degenerate primers L14912- CYB: 5'-TTCCTAGCCATACAYTAYAC-3' (Y =C or T) and H15149-CYB: 5'-GGTGGCKCCTCAGAAGGAC ATTTGKCCYCA-3' $(K = G \text{ or } T, Y = C \text{ or } T)$ were amplifed by PCR with barcodes for an eight-base sequence unique to each sample (Miya and Nishida [2000](#page-10-6)). The product size was approximately 285 bp and the target site of the primer pair was a conserved region widely found in vertebrates (Minamoto et al. [2012\)](#page-10-7). PCR reactions were performed in triplicate in a 25 µL mixture comprising 5 μL of $5 \times$ reaction buffer, 5μ L of $5 \times$ GC buffer, 2μ L of a deoxynucleotide (dNTP, 2.5 mM) mixture, 1 μL of forward and reverse primers (10 uM), 50 ng of DNA template, 0.25 μL of Q5 High-Fidelity DNA (2 U/μL) Polymerase (New England Biolabs, USA), and molecular biologygrade water added up to 25 μL. For all samples, PCR was

performed as follows: 98 °C for 2 min, followed by 30 cycles of 98 °C for 15 s, 55 °C for 30 s, and 72 °C for 30 s, with a final extension at 72 °C for 5 min. Molecular biology-grade water was used as a template for the negative control in each PCR reaction. Amplicons were extracted from 2% agarose gels and purifed using the AxyPrep DNA Gel Extraction Kit (Axygen Biosciences, Union City, CA, US) according to the manufacturer's instructions. All negative controls had no target bands, indicating that there was no exogenous DNA contamination during sampling, fltration, DNA extraction and PCR amplifcation. Pairedend sequencing was performed on an Illumina MiSeq. The Mova-Seq-PE250 sequencing strategy was used. Lowquality sequences were removed using QIIME software according to the following conditions: (1) sequences less than 150 bp in length, (2) sequences with an average Phred quality score $<$ 20, (3) sequences containing ambiguous bases other than N, (4) sequences with base mismatches for the 5' end primers > 1 , and (5) sequences containing single nucleotide repeats > 8 bp (Caporaso et al. [2010\)](#page-10-8). Sequence alignment was performed using the UCLUST tool in QIIME software (Edgar [2010](#page-10-9)). Clusters with highquality sequence consistency over 97% were identifed as operational taxonomic units (OTUs). The sequence with the highest abundance was selected from each OTU, screened against GenBank using the Basic Local Alignment Search Tool (BLAST), and assigned to the taxon with the highest total score (Chariton et al. [2010\)](#page-10-10). Finally, OTUs with relative abundance values below 0.001% were removed (Bokulich et al. [2013\)](#page-10-11). The relative abundance of all species at each site was estimated with $pi = N_i/N$, where N_i is the number of reads for species *i* and *N* is the total read for all species in the sample.

Data analysis

The parameters used for species annotation were identity value \geq 97% and e-value < 10⁻¹⁰. OTUs that aligned to the same species were merged. If there was an OTU that could not be annotated at the species level, the analysis was carried out one level higher. The number of valid sequences of species in each sample was counted in an Excel table. A species was considered to be present at the sampling point if it was detected in at least two duplicate samples. Sequencing produced a total of 2.98 million paired end reads, with 2.51 million reads remaining after quality fltering. The average number of reads per site after fltering was 60,185.

The alpha diversity for each sample was calculated using four diversity indices: Shannon–Wiener index $(H' = -\sum_{i=1}^{s} (p_i \text{In} P_i)$, Pielou index (($J' = H'/\text{In} S$)), Simpson index $\overline{(C} = \sum_{i=1}^{s} [Ni(Ni-1)/N(N-1)]$, and Margalef index $(D = (S − 1)/lnN)$, where *S* is the total number of species in each sample, pi is the relative abundance, N_i is the number of reads for species *i* and *N* is the total read for all species in the sample. The diferences in biodiversity between habitats were compared via non-parametric tests using IBM® SPSS® 25.0. The Kruskal–Wallis test was used to evaluate whether the distributions of multiple biodiversity indices were signifcantly diferent. Beta diversity refers to the diferences or similarities in community composition among diferent groups of samples, determined through inter-group comparative analysis. Beta diversity was assessed using Principal Component Analysis (PCA), Nonmetric Multidimensional Scaling (NMDS) and cluster analysis. For NMDS, multivariate analysis of variance (Adonis) was used to evaluate the diferences in community structure in diferent habitats.

Results

Major taxa at the class level based on environmental DNA metabarcoding

In the spring of 2019, Pisces species accounted for 68.9% of all species identifed in Poyang Lake, followed by Bivalves at 13.3%. The Pisces species in the Yangtze River habitat (A1–A3) accounted for about 70% of the total. Other classes included Mammalia, Bivalvia and Insecta (Fig. [2\)](#page-4-0). The number of classes found at each sampling site in the channel connecting Poyang Lake and the Yangtze River (B1–B8) varied considerably. The samples collected at the B1 site contained Pisces and Mammals, with the former accounting for about 65% of all species. The samples collected at the B2, B3, B7 and B8 sites contained Pisces, Mammalia and Mastigophora. Four or more classes were identifed at B4, B5 and B6. Among the sampling sites in the main lake area of Poyang Lake, fve classes were identifed at C1 and C6, and three classes were identifed at C2, C3, C4 and C5. Oligotricla was only found at C7. In Nanjishan Nature Reserve (D1–D3), Pisces species were absolutely dominant and the sample from D2 only contained fshes. In Junshan Lake and Qinglan Lake habitats, Pisces accounted for more than 50% of the species identifed (Fig. [2](#page-4-0)).

Species composition and abundance

A total of 45 species were identifed in the six habitats investigated, including 31 species of Pisces and six species of Bivalves (Table [1\)](#page-5-0). Among the habitats, the largest number of species (28 species) was identifed in the channel connecting Poyang Lake and the Yangtze River, while the lowest number was identifed in Qinglan Lake (10 species). Among the Pisces species identifed, *Cyprinus carpio* were found in all habitats. Three fsh species, *Tachysurus fulvidraco*, *Hypophthalmichthys nobilis* and *Cyprinus rubrofuscus*,

Fig. 2 Class-level diversity of major taxa across the surveyed sites in Poyang Lake. Jiangxi section of the Yangtze River (A1–A3), channel connecting Poyang Lake and the Yangtze River (B1–B8), main lake area of Poyang Lake (C1–C8), Nanjishan Nature Reserve (D1–D3), Junshan Lake (E1–E5), Qinglan Lake (F1–F2)

were identified in at least four habitats. Some fishes, such as *Myxocyprinus asiaticus, Gobiobotia flifer*, *Rutilus rutilus*, *Mugilogobius myxodermus*, *Oreonectes platycephalus*, and *Microphysogobio tafangensis,* were identifed in only one habitat. Among the Bivalves identifed, *Solenaia oleivora* and *Nodularia douglasiae* were found in four habitats, *Arconaia lanceolata* and *Acuticosta chinensis* were found in three habitats, and *Lamprotula leai* and *Limnoperna fortunei* were found in only two habitats. Human sequences were present in all habitats. The amplifed sequence contained a signifcant proportion of human sequences, which was normal because there were traces of human activity in the water. The primers used in this study need human sequenceinhibiting primers to be added during amplifcation. These primers were not added in our study, which was why the human sequences were present in all habitats.

In this study, 31 fish species from eight orders, 11 families and 24 genera were identifed, including 20 indigenous species belonging to eight families and 15 genera as well as 11 exotic species from fve families and 10 genera (Table [1](#page-5-0)). Among them, 70.97% were limnicolous fshes, 16.13% were river–lake migratory fshes, 9.68% were river-sea migratory fishes and 3.23% were potamophilous fishes. Among the fish sequences detected, 94.62% were for limnicolous, 5.61% for river–lake migratory fshes, 0.15% for river-sea migratory fshes, and 0.06% for potamophilous fshes. The Pisces species included the orders Cypriniformes, Siluriformes, Clupeiformes, Beloniformes, Osmeriformes, Gymnotiformes, Cyprinodontiformes and Perciformes, with Cypriniformes comprising the largest number of species at 19.

The highest abundance of *C. carpio* and *Fundulus notatus* was found in the Jiangxi section of the mainstream of the Yangtze River (Fig. [3](#page-6-0)). *C. carpio* and *T. fulvidraco* were most abundant in the channel connecting Poyang Lake and the Yangtze River habitat. Only *C. carpio* showed high abundance in both the main lake area of Poyang Lake and Nanjishan Nature Reserve. The species with the highest abundance in the Junshan Lake habitat were *C. carpio* and *Xenocypris davidi*. *Hypophthalmichthys nobilis* was the most abundant in Qinglan Lake.

Alpha diversity analysis of diferent habitats

The Shannon-Weiner diversity index for the diferent habitats ranged from 0.45 to 1.70. The highest diversity index was found at the Jiangxi section of the mainstream of the Yangtze River, followed by the channel connecting Poyang Lake and the Yangtze River. The lowest diversity value was found in Qinglan Lake (Fig. [4](#page-7-0)). The Pielou Index ranged from 0.20 to 0.61, with the highest value in the Jiangxi section of the mainstream of the Yangtze River and the lowest in Qinglan Lake. The Simpson index ranged from 0.22 to 0.84, with the highest value in Qinglan Lake and the lowest in the Jiangxi section of the mainstream of the Yangtze River. The Margalef index ranged from 1.04 to 2.62, with the highest value in the channel connecting Poyang Lake and the Yangtze River and the lowest in Nanjishan Nature Reserve. There was no significant difference $(P=0.416)$ in the diversity indices among diferent habitats (Fig. [4](#page-7-0)).

Beta diversity analysis in diferent habitats based on environmental DNA metabarcoding

PCA showed that the habitats in the channel connecting Poyang Lake and the Yangtze River, the main lake area of Poyang Lake, Nanjishan Nature Reserve and Junshan Lake had relatively similar species compositions. The Jiangxi section of the mainstream of the Yangtze River and Qinglan **Table 1** Species identifed through environmental DNA metabarcoding of water samples collected from six habitats in Poyang Lake

A—Jiangxi section of the mainstream of the Yangtze River, B—channel connecting Poyang Lake and the Yangtze River, C—main lake area of Poyang Lake, D—Nanjishan Nature Reserve, E—Junshan Lake, F— Qinglan Lake. HG represents ecotype

LT limnicolous, *MRL* river–lake migratory, *RM* river-sea migratory, *RT* potamophilous

a Indicates species not previously detected in Poyang Lake

Fig. 3 Log-scaled percentage heat map at species level. Horizontal coordinates indicate sampling sites, vertical coordinates indicate species names. Red indicates high abundance. Blue indicates low abundance

Lake were very diferent from the other habitats (Fig. [5a](#page-8-0)). After standardized conversion of the species abundance in the 29 sampling sites, clustering was performed based on the Euclidean distance and ward minimum connections. The results showed that the 29 sampling sites were clustered into three branches with a Euclidean distance $D=17.2$. Branch I included all sampling sites in the Jiangxi section of the mainstream of the Yangtze River. Branch II included 24 sampling sites in four habitats, specifcally the channel connecting Poyang Lake and the Yangtze River, the main lake area of Poyang Lake, Nanjishan Nature Reserve and Junshan Lake. The sampling sites at Qinglan Lake were part of branch III (Fig. [5](#page-8-0)b).

In this study, NMDS analysis was conducted on 5 groups of samples (Qinglan Lake was deleted due to the small number of sample points) at the species level. The NMDS stress value was 0.12, indicating a good result. The overall community structure was signifcantly diferent among habitats $(R^2=0.486, P<0.001)$ (Fig. [6](#page-8-1)). Adonis analysis showed that there were signifcant diferences between the Jiangxi section of the mainstream of the Yangtze River and the main lake area of Poyang Lake (R^2 =0.017, P=0.025) and between the main lake area of Poyang Lake and Junshan Lake $(R^2=0.020,$ $P=0$).

Fig. 4 Diferences in biodiversity index among diferent habitats. A—Jiangxi section of the mainstream of the Yangtze River, B—channel connecting Poyang Lake and the Yangtze River, C—main lake

area of Poyang Lake, D—Nanjishan Nature Reserve, E—Junshan Lake, F—Qinglan Lake

Discussion

Fish community structures in Poyang Lake based on environmental DNA

A total of 31 fsh species were detected in this study using environmental DNA metabarcoding technology. The majority of fsh were Cyprinidae (16 species), accounting for 51.61% of all fsh species identifed, which was consistent with the results obtained using traditional methods (Fang et al. [2016](#page-10-12); Zhang and Li [2007\)](#page-11-4). This confrms a previous report indicating that the majority of fsh in Poyang Lake were Cyprinidae, and that limnicolous fsh were the most abundant (Yang et al. [2015](#page-11-7)). In addition, 11 exotic species were detected in this study distributed in four orders. Cypriniformes were the most abundant, possibly due to their strong adaptability. The detection of the rare fsh *Coilia nasus* in the channel connecting Poyang Lake and the Yangtze River as well as in the main lake area of Poyang Lake indicated that environmental DNA metabarcoding is efective for the detection of endangered species. The second-class state protected animal *M. asiaticus* was detected in the Junshan Lake habitat due to artifcial cultivation of the species over a large surface area in this habitat. *Cyprinus carpio* was the dominant fsh species in the Jiangxi section of the mainstream of the Yangtze River, the channel connecting Poyang Lake and the Yangtze River, the main lake area of Poyang Lake and Nanjishan Nature Reserve, as determined with environmental DNA metabarcoding. The dominant fsh species caught using traditional methods are *C. carpio* and *Carassius auratus* (Wang et al. [2016](#page-11-8); He et al. [2016](#page-10-13); Xiong [2018](#page-11-9); Hu et al. [2005](#page-10-14)). The dominant species observed in Qinglan Lake and Junshan Lake were closely related to the cultivated species at the time because dike breeding is performed in these two lakes.

Even with environmental DNA metabarcoding approaches, valuable input from taxonomic experts is often required (Evans and Lamberti [2018](#page-10-15)). Firstly, environmental DNA metabarcoding can only determine the presence of species based on the genetic information available in environmental samples. The technique cannot capture information about the population size, age structure, physiological status and growth developmental stage of the target species (Shu et al. [2020](#page-10-16)). Secondly, environmental DNA metabarcoding relies on the integrity of molecular databases, which can lead to false negatives when the target species sequence is missing from the database (Cristescu and Hebert [2018](#page-10-17)). In addition, the bias of PCR may lead to the failure of environmental DNA detection for some low abundance species (Carew et al. [2013\)](#page-10-18) and biomass estimation errors

Fig. 5 Principal component analysis and clustering analysis in six habitats, **a** Sample of principal component analysis. Treat 1: Jiangxi section of the mainstream of the Yangtze River, Treat 2: channel connecting Poyang Lake and the Yangtze River, Treat 3: main lake area of Poyang Lake, Treat 4: Nanjishan Nature Reserve, Treat 5: Junshan Lake, Treat 6: Qinglan Lake, **b** Sample hierarchical clustering tree

based on OTU levels. Jiangxi section of the mainstream of the Yangtze River (A1–A3), channel connecting Poyang Lake and the Yangtze River (B1–B8), main lake area of Poyang Lake (C1–C8), Nanjishan Nature Reserve (D1–D3), Junshan Lake (E1–E5), and Qinglan Lake (F1–F2)

(Elbrecht and Leese [2015](#page-10-19), [2017](#page-10-20)). Therefore, in actual practice, traditional survey methods should be combined with environmental DNA detection technology to monitor aquatic biodiversity more comprehensively and efficiently. These methods will play a more active role in the construction of aquatic ecological civilizations (Ge et al. [2020](#page-10-21)).

Alpha diversity of biological communities in Poyang Lake

Higher Shannon-Weiner index values indicate higher diversity in a community. A Shannon-Weiner diversity index between 1.5 and 3.5 indicates the community has a high level of biodiversity (Magurran [1988](#page-10-22)). In this study, the Shannon-Weiner diversity indices for the habitats in the Jiangxi section of the mainstream of the Yangtze River, the channel connecting Poyang Lake and the Yangtze River and the Junshan Lake in Poyang Lake were between 1.5 and 3.5 (Fig. [4\)](#page-7-0). This means the biodiversity in these habitats was high. The other habitats had low diversity, especially the Qinglan Lake habitat. The high biodiversity in the Jiangxi section of the mainstream of the Yangtze River and the channel connecting Poyang Lake and the Yangtze River was due to their special geographical location. The spatial and temporal continuity of rivers and lakes is an important reason for

Fig. 6 Nonmetric multidimensional scaling (NMDS) ordination of the community compositions of all environmental DNA samples colored according to the habitat. A—Jiangxi section of the mainstream of the Yangtze River, B—channel connecting Poyang Lake and the Yangtze River, C—main lake area of Poyang Lake, D— Nanjishan Nature Reserve, E—Junshan Lake, F—Qinglan Lake. $(0.01 < {}^*P \le 0.05, 0.001 < {}^*P \le 0.01, {}^*{}^*P \le 0.001)$

their high biodiversity. Some scholars believe that the convergence of rivers can help increase fsh diversity (Fernandas et al. [2004;](#page-10-23) Röpke et al. [2015](#page-10-24)). For the channel connecting Poyang Lake and the Yangtze River, seasonal inundation makes the foodplain a rich habitat for aquatic animals. Vegetation along the lakeside and in the water can provide shelter for aquatic animals as well as spawning grounds for *C. carpio, C. auratus* and other fishes during the flood season (Qian et al. [2002](#page-10-25)). Due to the temperature gradient and vortex formed at the confuence, nutrients, wood debris and organic matter accumulate in these areas, which is conducive to the growth of phytoplankton and zooplankton and provide a rich source of food for aquatic animals (Gayoso and Podesta [1996](#page-10-26)). Due to the frequent dike aquaculture activities in Qinglan Lake, fshery farming has damaged the biological community structure of the water body, severely afecting the biological diversity. Aquatic plants were seriously damaged in Junshan Lake due to a large number of crab farms in the early twentieth century. Subsequently, the lake underwent a decade-long ecological restoration. Junshan Lake has a high Shannon-Weiner diversity index and Pielou index, which is attributed to the 10 years of ecological restoration. The biodiversity level of habitats in which dike farming was prohibited showed a decreasing trend due to overfshing. For example, the Shannon-Weiner diversity index recorded using traditional methods in the channel connecting Poyang Lake and the Yangtze River was greater than 2 in 2012 (He et al. [2016\)](#page-10-13). Data based on net fshing in 2014 in Hukou indicated a Shannon-Weiner diversity index of 2.59 (Wang et al. [2016](#page-11-8)). Survey data from 2009 showed that there were 42 freshwater mussel species in Poyang Lake (Xiong et al. [2011](#page-11-10)). Survey data from 2016 to 2017 showed that there were 24 mussel species in Poyang Lake (Li et al. [2019\)](#page-10-27). The species richness of mussels in Poyang Lake decreased signifcantly. The situation for water ecological protection is grim with the continuous decline of biodiversity in Poyang Lake. In an effort to improve ecological protection, a 10-year fshing ban will be imposed on the waters of the mainstream of the Yangtze River and the Hukou region in Poyang Lake from January 1, 2021.

Beta diversity analysis in six habitats

PCA and clustering tree analysis showed that the habitats in the channel connecting Poyang Lake and the Yangtze River, the main lake area of Poyang Lake, Nanjishan Nature Reserve and Junshan Lake had relatively similar community structures (Fig. [5\)](#page-8-0). However, the community structures in these four habitats were obviously diferent from those in the Jiangxi section of the mainstream of the Yangtze River and Qinglan Lake (Fig. [5\)](#page-8-0). This may be related to the fact that the Jiangxi section of the mainstream of the Yangtze River is located at the confuence of Poyang Lake and the river, resulting in a higher Shannon-Weiner diversity index and Pielou index compared to the other habitats. Due to frequent aquaculture and lack of timely ecological restoration, the community structure in the Qinglan Lake habitat is quite diferent from those of the other habitats. The species composition and abundance heat maps for all habitats were different (Table [1,](#page-5-0) Fig. [3\)](#page-6-0) and there were significant differences in community structure among the habitats (Fig. [6](#page-8-1)). These results are of great signifcance for the protection, restoration and scientifc management of fshery lakes in China. They can provide a foundation for decision-making by government management departments.

Conservation strategies for biodiversity in Poyang Lake

The current fshing ban policy is conducive to the restoration of biodiversity in Poyang Lake. Based on the results of this study, the following suggestions are proposed. Firstly, carry out artifcial proliferation and release to curb the decline of fshery resources. Secondly, strengthen pollution control and strictly control the pollution of lakes. Thirdly, formulate a scientifc and reasonable sand mining plan. Fourthly, regular monitoring and early warning systems should be established, such as by increasing seasonal sampling and developing monitoring plans.

Conclusions

Environmental DNA metabarcoding technology has great application potential for the monitoring and conservation of biodiversity. This study represents the frst application of environmental DNA metabarcoding for the assessment of biodiversity in Poyang Lake and demonstrates the feasibility of using environmental DNA metabarcoding to detect species composition and the distribution of organisms (especially fsh) in diferent habitats. The results of high-throughput sequencing showed a decreasing trend in the biodiversity and characteristics of the habitat community structure in Poyang Lake. Although environmental DNA metabarcoding cannot completely replace traditional fsh survey methods, it can serve as an important complementary tool for rapid determination of the diversity and spatial distribution of organisms in water bodies. It can reduce potential damage to the ecosystem compared to traditional monitoring techniques, shorten the survey period, improve the detection efficiency, and provide reliable data to enable a rapid response for the aquatic ecological protection.

Author contributions CZ and XW: conceived and designed the research; JC and TG: contributed to feld sampling, fltration and environmental DNA extraction in the laboratory. CZ and JC: wrote the manuscript, CZ and SO: contributed to the Discussion section and, subsequently, to various iterations of the manuscript, all authors reviewed the manuscript before submission.

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Data availability The datasets analyzed during the current study are available from the corresponding author on a reasonable request.

Declarations

Conflict of interest The authors declare that they have no known competing fnancial interests or personal relationships that could have appeared to infuence the work reported in this paper.

References

- Bokulich NA, Subramanian S, Faith JJ, Gevers D, Gordon JI, Knight R, Mills DA, Caporaso JG (2013) Quality fltering vastly improves diversity estimates from Illumina amplicon sequencing. Nat Methods 10:57–59. <https://doi.org/10.1038/nmeth.2276>
- Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD, Costello EK, Fierer N, Peña AG, Goodrich JK, Gordon JI, Ley RE, Lozupone CA, McDonald D, Muegge BD, Pirrung M, Reeder J, Sevinsky JR, Turnbaugh PJ, Walters WA, Widmann J, Yatsunenko T, Zaneveld J, Knight R (2010) QIIME allows analysis of high-throughput community sequencing data. Nat Methods 7:335–336.<https://doi.org/10.1038/nmeth.f.303>
- Carew ME, Pettigrove VJ, Metzeling L, Hofmann AA (2013) Environmental monitoring using next generation sequencing: rapid identifcation of macroinvertebrate bioindicator species. Front Zool 10:45.<https://doi.org/10.1186/1742-9994-10-45>
- Chariton AA, Court LN, Hartley DM, Collof MJ, Hardy CM (2010) Ecological assessment of estuarine sediments by pyrosequencing eukaryotic ribosomal DNA. Front Ecol Environ 8:233–238. <https://doi.org/10.1890/090115>
- Chen W, Zhang Y, Zhao C, Wang C (2012) Species composition and biodiversity of fsh community in Hukou section of the Yangtze River. Resour Environ Yangtze Basin. 21:684–691
- Cristescu ME, Hebert PDN (2018) Uses and misuses of environmental DNA in biodiversity science and conservation. Annu Rev Ecol Evol Syst 49:209–230. [https://doi.org/10.1146/annurev-ecols](https://doi.org/10.1146/annurev-ecolsys-110617-062306) [ys-110617-062306](https://doi.org/10.1146/annurev-ecolsys-110617-062306)
- Edgar RC (2010) Search and clustering orders of magnitude faster than BLAST. Bioinformatics 26:2460–2461. [https://doi.org/10.1093/](https://doi.org/10.1093/bioinformatics/btq461) [bioinformatics/btq461](https://doi.org/10.1093/bioinformatics/btq461)
- Elbrecht V, Leese F (2015) Can DNA-based ecosystem assessments quantify species abundance? Testing primer bias and biomasssequence relationships with an innovative metabarcoding protocol. PLoS ONE 10:e0130324. [https://doi.org/10.1371/journal.](https://doi.org/10.1371/journal.pone.0130324) [pone.0130324](https://doi.org/10.1371/journal.pone.0130324)
- Elbrecht V, Leese F (2017) Validation and development of COI metabarcoding primers for freshwater macroinvertebrate bioassessment. Front Env Sci Switz 5:11. [https://doi.org/10.3389/fenvs.](https://doi.org/10.3389/fenvs.2017.00011) [2017.00011](https://doi.org/10.3389/fenvs.2017.00011)
- Evans NT, Lamberti GA (2018) Freshwater fsheries assessment using environmental DNA: a primer on the method, its potential, and

shortcomings as a conservation tool. Fish Res 197:60–66. [https://](https://doi.org/10.1016/j.fishres.2017.09.013) [doi.org/10.1016/j.fshres.2017.09.013](https://doi.org/10.1016/j.fishres.2017.09.013)

- Evans NT, Li YY, Renshaw MA, Olds BP, Deiner K, Turner CR, Jerde CL, Lodge DM, Lamberti GA, Pfrender ME (2017) Fish community assessment with eDNA metabarcoding: effects of sampling design and bioinformatic fltering. Can J Fish Aquat Sci 74:362–1374. <https://doi.org/10.1139/cjfas-2016-0306>
- Fang C, Chen W, Zhou H, Zhang Y, Fu P, He G, Wu B, Wang S (2016) Fish resources of Poyang Lake and its utilization proposal. Jiangsu J Agr Sci 44:233–243. [https://doi.org/10.15889/j.issn.1002-1302.](https://doi.org/10.15889/j.issn.1002-1302.2016.09.067) [2016.09.067](https://doi.org/10.15889/j.issn.1002-1302.2016.09.067)
- Fernandas CC, Podos J, Lundberg JG (2004) Amazonian ecology: tributaries enhance the diversity of electric fshes. Science 305:1960– 1962. <https://doi.org/10.1126/science.1101240>
- Gayoso AM, Podesta GP (1996) Surface hydrography and phytoplankton of the Brazil-Malvinas currents confuence. J Plankton Res 18:941–951.<https://doi.org/10.1093/plankt/18.6.941>
- Ge Y, Yan Y, Cheng Q (2020) Environmental DNA and its application in aquatic biodiversity. Fishery Inform Strat 35:55–62. [https://doi.](https://doi.org/10.13233/j.cnki.fishis.2020.01.008) [org/10.13233/j.cnki.fshis.2020.01.008](https://doi.org/10.13233/j.cnki.fishis.2020.01.008)
- He G, Fang C, Chen W, Fu P, Zhou H, Zhang Y, Wu B, Wang S (2016) Fish community structure and diversity in Pingfeng section of the channel connecting the Poyang Lake and the Yangtze River. Jiangxi Fishery Sci 4:3–6. [https://doi.org/10.3969/j.issn.1006-](https://doi.org/10.3969/j.issn.1006-3188.2016.04.002) [3188.2016.04.002](https://doi.org/10.3969/j.issn.1006-3188.2016.04.002)
- Hu M, Wu Z, Liu Y (2011) Fish diversity and community structure in Hukou area of Lake Poyang. J Lake Sci 23:246–250
- Hu M, Wu Z, Zhou H, Zhang A, Song W, Zhang C (2005) The fsheries characters and resource status of Nanjishan Natural Reserve in Poyang Lake. Resour Environ Yangtze Basin 14:561–565
- Li K, Liu X, Zhou Y, Xu Y, Lv Q, Ouyang S, Wu X (2019) Temporal and spatial changes in macrozoobenthos diversity in Poyang Lake Basin, China. Ecol and Evol 9:6353–6365. [https://doi.org/](https://doi.org/10.1002/ece3.5207) [10.1002/ece3.5207](https://doi.org/10.1002/ece3.5207)
- Magurran AE (1988) Ecological diversity and its measurement. Princeton University Press, NewJersey
- Minamoto T, Yamanaka H, Takahara T, Honjo MN, Kawabata ZI (2012) Surveillance of fsh species composition using environmental DNA. Limnology 13:193–197. [https://doi.org/10.1007/](https://doi.org/10.1007/s10201-011-0362-4) [s10201-011-0362-4](https://doi.org/10.1007/s10201-011-0362-4)
- Miya M, Nishida M (2000) Use of mitogenomic information in teleostean molecular phylogenetics: a tree-based exploration under the maximum-parsimony optimality criterion. Mol Phylogenet Evol 17:437–455.<https://doi.org/10.1006/mpev.2000.0839>
- Qian X, Huang C, Wang Y, Xiong F (2002) The status quo of fshery resources of Lake Poyang and its environmental monitoring. Acta Hydrobiol Sin 26:612–617. [https://doi.org/10.3321/j.issn:1000-](https://doi.org/10.3321/j.issn:1000-3207.2002.06.006) [3207.2002.06.006](https://doi.org/10.3321/j.issn:1000-3207.2002.06.006)
- Röpke CP, Amadio SA, Winemiller KO, Zuanon J (2015) Seasonal dynamics of the fsh assemblage in a foodplain lake at the confuence of the Negro and Amazon Rivers. J Fish Biol 89:194–212. <https://doi.org/10.1111/jfb.12791>
- Ruppert KM, Kline RJ, Rahman MS (2019) Past, present, and future perspectives of environmental DNA (eDNA) metabarcoding: a systematic review in methods, monitoring, and applications of global eDNA. Glob Ecol Conserv 17:e00547. [https://doi.org/10.](https://doi.org/10.1016/j.gecco.2019.e00547) [1016/j.gecco.2019.e00547](https://doi.org/10.1016/j.gecco.2019.e00547)
- Shu L, Lin J, Xu Y, Cao T, Feng J, Peng Z (2020) Investigating the fish diversity in Erhai Lake based on environmental DNA metabarcoding. Acta Hydrobiol Sin 44:1080–1086. [https://doi.org/10.](https://doi.org/10.7541/2020.125) [7541/2020.125](https://doi.org/10.7541/2020.125)
- Taberlet P, Coissac E, Hajibabaei M, Rieseberg LH (2012) Environmental DNA. Mol Ecol 21:1789–1793. [https://doi.org/10.1111/j.](https://doi.org/10.1111/j.1365-294X.2012.05542.x) [1365-294X.2012.05542.x](https://doi.org/10.1111/j.1365-294X.2012.05542.x)
- Thomsen PF, Willerslev E (2015) Environmental DNA-an emerging tool in conservation for monitoring past and present biodiversity.

Biol Conserv 183:4–18. [https://doi.org/10.1016/j.biocon.2014.](https://doi.org/10.1016/j.biocon.2014.11.019) [11.019](https://doi.org/10.1016/j.biocon.2014.11.019)

- Thomsen PF, Kielgast J, Iversen LL, Wiuf C, Rasmussen M, Gilbert MTP, Orlando L, Willerslev E (2012) Monitoring endangered freshwater biodiversity using environmental DNA. Mol Ecol 21:2565–2573. [https://doi.org/10.1111/j.1365-294X.2011.](https://doi.org/10.1111/j.1365-294X.2011.05418.x) [05418.x](https://doi.org/10.1111/j.1365-294X.2011.05418.x)
- Valentini A, Taberlet P, Miaud C, Civade R, Herder J, Thomsen PF, Bellemain E, Besnard A, Coissac E, Boyer F, Gaboriaud C, Jean P, Poulet N, Roset N, Copp GH, Geniez P, Pont D, Argillier C, Baudoin JM, Peroux T, Crivelli AJ, Olivier A, Acqueberge M, Brun ML, Møller PR, Willerslev E, Dejean T (2016) Next-generation monitoring of aquatic biodiversity using environmental DNA metabarcoding. Mol Ecol 25:929–942. [https://doi.org/10.](https://doi.org/10.1111/mec.13428) [1111/mec.13428](https://doi.org/10.1111/mec.13428)
- Wang S, Duan X, Chen W, Zhang H, Fu P, He G, Wu B (2016) Status and changes of fsh resources in the Hukou area of Poyang Lake. Freshwater Fish 46:50–55. [https://doi.org/10.3969/j.issn.1000-](https://doi.org/10.3969/j.issn.1000-6907.2016.06.009) [6907.2016.06.009](https://doi.org/10.3969/j.issn.1000-6907.2016.06.009)
- Wheeler QD, Raven PH, Wilson EO (2004) Taxonomy: impediment or expedient? Science 303:285. [https://doi.org/10.1126/science.](https://doi.org/10.1126/science.303.5656.285) [303.5656.285](https://doi.org/10.1126/science.303.5656.285)
- Xiong G (2018) Preliminary survey on fsh resources in Wetland Inland Lakes of Poyang Lake. Jiangsu J Agr Sci 46:186–190. [https://doi.](https://doi.org/10.15889/j.issn.1002-1302.2018.21.048) [org/10.15889/j.issn.1002-1302.2018.21.048](https://doi.org/10.15889/j.issn.1002-1302.2018.21.048)
- Xiong L, Ouyang S, Chen T, Qi T, Wu X (2011) Diversity patterns of freshwater mussels in poyang lake area. J Nanchang Univ (nat Sci) 35:288–295
- Xu N, Xiong M, Shao K, Que Y, Li J (2020) Preliminary study on environmental dna metabarcoding for detecting biodiversity in the middle and lower reaches of the Yangtze River. Res Environ Sci 33:1187–1196. [https://doi.org/10.13198/j.issn.1001-6929.](https://doi.org/10.13198/j.issn.1001-6929.2020.03.06) [2020.03.06](https://doi.org/10.13198/j.issn.1001-6929.2020.03.06)
- Yang S, Li M, Zhu Q, Wang M, Liu H (2015) Spatial and temporal variations of fsh assemblages in Poyang Lake. Resour Environ Yangtze Basin 24:54–64
- Zhang B (1993) The hydrological features and the renovative strategy of the Poyang Lake. Resour Environ Yangtze Basin 2:36–42
- Zhang T, Li Z (2007) Fish resources and fshery utilization of Lake Poyang. J Lake Sci 19:434–444. [https://doi.org/10.18307/2007.](https://doi.org/10.18307/2007.0412) [0412](https://doi.org/10.18307/2007.0412)
- Zhang S, Lu Q, Wang Y, Wang X, Zhao J, Yao M (2020) Assessment of fish communities using environmental DNA: effect of spatial sampling design in lentic systems of diferent sizes. Mol Ecol Resour 20:242–255. <https://doi.org/10.1111/1755-0998.13105>

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