



Effect of soil amendments on soil respiration in the midland agroecological environment, Ethiopia

Genetu Fekadu¹ · Enyew Adgo² · Derege Tsegaye Meshesha² · Atsushi Tsunekawa³ · Nigussie Haregeweyn⁴ · Fei Peng^{5,6} · Mitsuru Tsubo³ · Temesgen Mulualem² · Simeneh Demissie² · Birhanu Kebede⁷ · Gizachew Ayalew Tiruneh⁸

Received: 10 June 2023 / Accepted: 24 February 2024 / Published online: 27 March 2024
© The Author(s), under exclusive licence to Springer-Verlag GmbH Germany, part of Springer Nature 2024

Abstract

The effects of soil amendments on soil respiration (SR) rate have not yet been fully evaluated. In the Upper Blue Nile Basin, Ethiopia, we investigated seasonal variations in SR rates between and within soil amendments. We set up 24 plots with 8 treatments—polyacrylamide (PAM), gypsum (G), biochar (B), lime (L), control (C), PAM + L, PAM + G, and PAM + B—in 3 replicates. Soil temperature, moisture, and SR data were collected monthly. Data were evaluated by paired *t*-test and nonparametric repeated-measures ANOVA. The SR rates were significantly lower ($P < 0.05$) in PAM + B ($3.43 \pm 0.55 \mu\text{mol m}^{-2} \text{s}^{-1}$) than in B ($4.23 \pm 0.61 \text{ CO}_2 \mu\text{mol m}^{-2} \text{s}^{-1}$) and PAM + L ($4.14 \pm 0.57 \mu\text{mol m}^{-2} \text{s}^{-1}$). The SR rate differed significantly in all plots between the wet and dry seasons. The relationships between SR rate and soil properties were not significant, although there was a non-significant positive association with soil organic carbon, total nitrogen, soil pH, and clay content. There was a significant ($P < 0.01$) association between soil moisture and SR in all treatments. PAM + B offers a practical means of enhancing carbon storage efficiency. Comprehensive studies should be conducted in a variety of agroecological settings to determine optimal techniques to reduce SR emissions.

Keywords Semi-arid · Biochar · Soil moisture · Polyacrylamide · Upper Blue Nile basin

Introduction

Land degradation leading to soil erosion reduces agricultural production and threatens food security worldwide (Powlson et al. 2011). In Ethiopia, large-scale soil erosion has occurred as population growth has turned forests, pastures, and even hillsides into farmland without sustainable management. Soil erosion is a major problem in many parts of Ethiopia (Haregeweyn et al. 2015). The eastern and northern regions of the Ethiopian Highlands suffer significant damage through land degradation (Hurni 1988; Shiferaw 2011). Other potential contributing factors are the increased use of steep slopes for agricultural purposes and reduced vegetation cover (Amede et al. 2001); biophysical factors (Hurni et al. 2010); conversion of natural ecosystems to cultivated ecosystems (Seyoum 2016); anthropogenic factors (Nyssen et al. 2007); and increasing influence on resources (Guyassa and Raj 2013). National soil erosion rates are estimated at up to $220 \text{ t ha}^{-1} \text{ y}^{-1}$, with a mean of $37 \text{ t ha}^{-1} \text{ y}^{-1}$ (Tamene et al. 2022). Soil loss reduces agricultural productivity by depleting soil nutrients, robbing large amounts of soil

✉ Genetu Fekadu
genetufekadu@gmail.com

- ¹ Department of Natural Resources Management, University of Gondar, Gondar, Ethiopia
- ² Department of Natural Resources Management, Bahir Dar University, Bahir Dar, Ethiopia
- ³ Arid Land Research Center, Tottori University, Tottori, Japan
- ⁴ International Platform for Dryland Research and Education, Tottori University, Tottori, Japan
- ⁵ Key Laboratory of Desert and Desertification, Northwest Institute of Eco-Environment and Resources, Chinese Academy of Sciences, Lanzhou, China
- ⁶ Beiluhe Observation and Research Station On Frozen Soil Engineering and Environment in Qinghai-Tibet Plateau, Chinese Academy of Sciences, Lanzhou, China
- ⁷ Faculty of Civil and Water Resource Engineering, Bahir Dar Institute of Technology, Bahir Dar University, Bahir Dar, Ethiopia
- ⁸ Department of Natural Resources Management, Debre Tabor University, Debre Tabor, Ethiopia

organic matter (SOM), and altering soil properties (Hailelassie et al. 2006).

A number of soil amendment approaches have been tested and used to avert soil erosion in Ethiopia and elsewhere (Mamedov et al. 2021; Mulualem et al. 2021, 2022; Awad et al. 2012; Kebede et al. 2022; Albalasmeh et al. 2021; Asghari et al. 2011). Polyacrylamide (PAM), biochar (B), lime (L), and gypsum (G) are common amendments used to modify soil properties in order to reduce soil erosion. PAM stabilizes soil aggregates and improves cohesiveness (Mamedov et al. 2021), reduces soil nutrient depletion, and increases nutrient utilization (Mulualem et al. 2022). As a result, CO₂ emissions are reduced (Awad et al. 2012). In acidic soils, the use of PAM with biochar and lime increased the yield of teff (Mulualem et al. 2021). PAM is more effective than other soil additives in improving the physical quality of sandy loam soils (Albalasmeh et al. 2021; Asghari et al. 2011). The addition of PAM increased water infiltration and decreased soil erosion (Kebede et al. 2022; Sojka and Entry 2000), and it promoted the growth and survival of some fungal and bacterial species that bind soil aggregates (Caesar-Tonthat et al. 2008).

Lime is often used to reduce soil acidity; applying lime alone or in combination with other additives such as gypsum is a viable way to improve soil health and crop yield (Bossolani et al. 2020). Biochar improves soil quality (Smith et al. 2010). It improved the properties of heavily weathered soil and reduced soil loss (Jien and Wang 2013). Chen et al. (2018) detailed its long-term effects on soil physiochemical properties. One long-term benefit may be reducing additional carbon (C) mineralization in compost (Jien et al. 2015). Using biochar to improve soil moisture and nutrient retention is beneficial for plant growth (Sales et al. 2022).

Soil amendments can increase crop yield (Aina et al. 2018). The improvement of SOM is one of the best management strategies for ensuring soil sustainability in subtropical and tropical regions (Jien et al. 2015). Organic fertilizers improve the content of microbial biomass, the content of SOM, the structure of microbial communities, and the activity of enzymes involved in the degradation of OM (Nett et al. 2012). Organic additives improve soil nutrient status and chemical properties (Angelova et al. 2013). Cow dung solids can improve soil physical and chemical properties (Loper et al. 2010). The co-use of organic fertilizers with mineral fertilizers increases crop yields, reduces mineral fertilizer consumption, and conserves soil resources (Bayu et al. 2006).

Soil respiration (SR) is the major mechanism of C exchange between the soil surface and the atmosphere (Sheng et al. 2010) and largely determines C accumulation in ecosystems. SR is an important indicator of soil fertility and a measure of soil quality (Staben et al. 1997). Good land management reduces CO₂ emissions (Abegaz et al. 2020;

Lemma et al. 2021), mitigates climate change (Mekonnen and Getahun 2020), and increases soil organic carbon (SOC) content. The type and proportion of applied OM additives (Ray et al. 2020) and SOC distribution (Menichetti et al. 2013) have the greatest influence on soil respiration. Compared with OM alone and OM + gypsum, gypsum alone reduced cumulative respiration within the top 5 cm of soil (Wong et al. 2009). The addition of digestive-rich organic additives to the soil improved microbial richness and respiration (Holatko et al. 2021). Biochar reduces heterotrophic soil respiration by enhancing recalcitrant fraction of carbon (Li et al. 2018). Therefore, understanding the regulation of SR is important, as small changes in SR rates can have large effects on atmospheric CO₂ emissions and soil C sequestration (Bowden et al. 2004). Furthermore, understanding the sensitivity of the terrestrial C cycle to climate change requires knowledge of the mechanisms that control the release of CO₂ from the soil via SR to the atmosphere (Savage et al. 2008). However, to the best of our knowledge, the effect of soil modification on SR rates has not been adequately assessed in Ethiopia. Hence, soil respiration study on soil amendments could inspire different institutions to provide various options for reducing carbon emissions by implementing amendments practices in different land uses and enhancing carbon sequestration in the soil. This study was designed to identify optimal soil amendments that minimize SR and increase SOC storage. The objectives were: (1) to assess differences in SR rates between soil amendments; (2) to evaluate SR rates between the wet and dry seasons; and (3) to pinpoint soil-related and environmental factors (soil moisture and temperature) that contribute to SR.

Materials and methods

Study site

The study was conducted in the Aba Gerima catchment (11° 38' 0"–11° 40' 30" N, 37° 29' 30"–37° 31' 0"), Bahir Dar Zuri District, which was chosen as representative of the midland agroecological zone of the Upper Blue Nile basin, northwestern Ethiopia (Fig. 1).

We collected climate data from the Ethiopian National Meteorological Agency station in Bahir Dar spanning 21 years, 2000 to 2020. The average annual rainfall in the study area was 1486 mm, and the average monthly temperature ranged from 18 to 23 °C (Fig. 2). The main rainy season (growing season) starts in June and ends in September (Mihretie et al. 2021). According to FAO's soil classification system (2006), Leptosols are the predominant soil type here (Getahun 2016). Teff (*Eragrostis tef* (Zucc.) Trotter), finger millet (*Eleusine coracana* Gaertn.), maize (*Zea mays* L.),

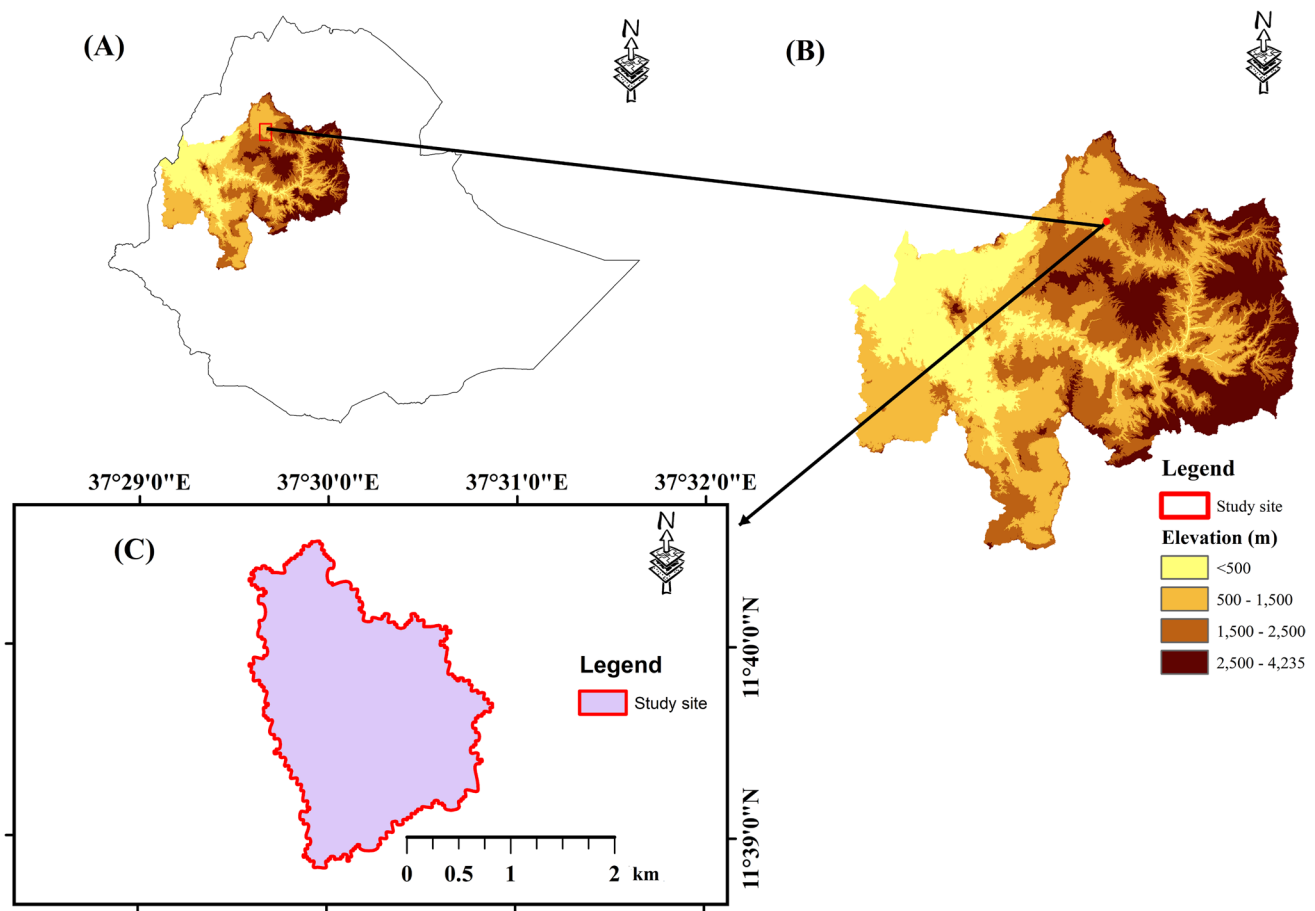


Fig. 1 The study site in **A** Ethiopia, **B** Upper Blue Nile basin, and **C** Aba Gerima watershed

and lupin (*Lupinus albus* L.) are the most commonly grown crops (Ebabu et al. 2019).

Setup of experimental plots

We used 8 treatments—control (C), gypsum (G), biochar (B), lime (L), polyacrylamide (PAM), PAM + B, PAM + G, and PAM + L—with 3 replications in a randomized complete block design with 24 plots. We used lime (CaCO_3) with 98% neutralizing value for the experiment. We did not analyze the purity of gypsum due to lack of finance and time. The source of Gypsum and lime was from local distributors or suppliers. JICA (Japan international cooperation agency) imported anionic PAM from Japan. It comprised, by weight, 50%, 22%, 20% and 8% of C, O, N and H, respectively. Biochar was made from wood charcoal from stem and branches of *Acacia decurrens*. An *acacia decurrens* with a diameter of 0.1–0.2 m were cut in different sizes for burning, arranged on soil (0.5–1 m height, 2–3 m diameter), and were set on fire for burning. The pile was covered by a layer of corn debris, and soil to avoid complete burning of biomass into ash. The burning process took on 3–5 days on average.

The pyrolysis temperature for preparation of biochar was 400–450 °C as described by Geng et al. (2022). After burning this, the charcoal was taken out and manually crushed and sieved to get uniform sizes (< 4 mm diameter) by mixing the biochar before application on experimental plots. For the study, we used blocking to minimize the effect of slope differences among the treatments. The average slope for experimental plots was 10% for experimental site (Muluaem et al. 2021). The size of the plot was 1.3 m width by 4.5 m length (5.85 m²). The perimeter of each plot was bordered by a 0.35 m metal sheet, of which 0.15 m was inserted into the ground to delineate nutrients from plot surroundings. Runoff trenches with trapezoidal cross-sections (2.5 m length and 1 m width on the upper, 1.5 m length and 0.5 m width on the foot, and 0.6 m deep) were dug beneath each plot and lined with a geo-membrane plastic sheet to avoid water loss by infiltration. In 2018, we applied PAM ($\text{CH}_2=\text{CH}-\text{CONH}_2$)_n at 40 kg ha⁻¹, biochar at 8 t ha⁻¹, gypsum ($\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$) at 5 t ha⁻¹, and lime (CaCO_3) at 4 t ha⁻¹. In 2019, we reapplied only PAM at 20 kg ha⁻¹. The experimental field was hoed and the full rates of all amendments except PAM were broadcasted manually 20–25 days before sowing (July) and

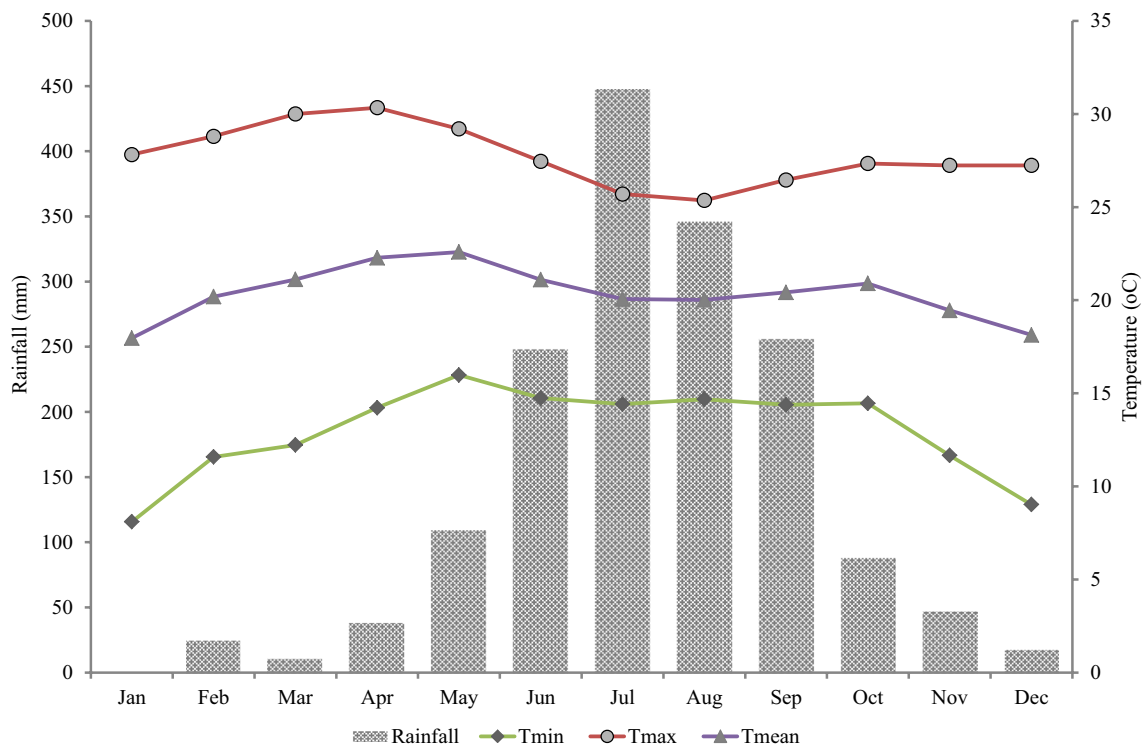


Fig. 2 Rainfall and temperature distribution in the study area

immediately mixed into the upper 0.15–0.20 m of the soil in the first year of the experiment (2018), whereas the granular (solid) PAM was applied to the soil surface (0.3–0.5 m) by hand in two applications, half at the start of the rainy season (i.e., 20–25 days before sowing) and the other half immediately after teff sowing in both study seasons. Moreover, in 2019 we applied only PAM by half rate. The ratings values were chosen from the literature (Kebede et al. 2020; Mulualem et al. 2021). The test plots were hand-plowed 3 times a year: during the dry season (May), at the beginning of the wet season (June), and at the end of July, when the teff was sown. During the test period, all plots were planted with teff, which is the most important staple crop in Ethiopia (Hunegnaw et al. 2021), even though it is low-yielding, unprofitable, and resource-intensive (Mihretie et al. 2022).

SR measurement and data collection

Data on SR, soil moisture, and soil temperature were collected from October 2019 to August 2021, with interruption from April to August 2020 due to COVID-19. Later, we started data collection on September 2020 until August 2021. Our interest was to compare two years soil respiration data however; due to interruption we analyzed the merged soil respiration data. SR was measured monthly in each test plot (Liang et al. 2019; Powers et al. 2018; Xiao et al. 2021). A PVC pipe collar (19 cm diameter, 11 cm

height) was carefully inserted 5 cm into the soil in each plot (Fekadu, et al. 2023a, b, c). Soil respiration data were collected between 09:00 and 12:00 on rainless mornings (Fekadu et al. 2023a, b, c; Jiang et al. 2013; Sheng et al. 2010). SR was measured for 90 s (Table 1), and the average SR of 90 readings for each collar was used.

Soil sampling and analysis

Soil sampling was collected at 0–20 cm soil depth from each experimental plot using hand auger. For every experimental

Table 1 Parameters and methods of soil analysis or tools used

Parameter	Method or instrument
Available P	Sodium bicarbonate method (Olsen et al. 1954)
Organic C and total N	CN Corder (Macro Corder JM1000CN, J-Science Lab, Kyoto, Japan)
pH (1:2.5 soil:water)	Peech (1965)
Soil bulk density	Core sample method, oven-dried at 105 °C for 24 h
Texture	Hydrometer method (Bouyoucos 1962)
Soil respiration	LI-8100 automated soil CO ₂ flux system (LI-8100, LI-COR, Lincoln, NE, USA)
Soil moisture (M_s)	Gravimetric method
Soil temperature (T_s)	Omega probe 6000-09TC (LI-COR)

plot, we collected four samples in north, south, east and west direction of the experimental plot, and later we mixed four soil samples of the experimental plot and took 200 g soil for further physico-chemical analysis. The time for soil sampling data collection was after crop harvest in February 2020. Soil samples were air-dried at room temperature, ground, and sieved through a 2-mm sieve for soil physical and chemical analyses (Table 1). All plots had a clay-loam soil texture (Table 2).

Statistical analysis

One-way ANOVA was used to assess the effects of soil amendments on soil properties. Tukey's HSD test at $P < 0.05$ was used to detect significant differences between mean values of soil properties in several comparative tests between treatments in experiments. SR data were not normally distributed, so they were analyzed by the nonparametric Friedman's test, followed by Wilcoxon's post hoc analysis to isolate means with $P < 0.05$. Significance values for some tests were adjusted by Bonferroni's correction. A paired t -test analysis was used to determine the significance of differences between seasons (wet and dry) among treatments. Pearson's correlation analysis was used to assess the relationships of SR with soil properties, soil temperature, and soil moisture (Gomez and Gomez 1983).

Results and discussion

Effects of soil amendments on soil respiration

The amendments altered the soil physico-chemical properties. PAM + L increased soil pH by (6.05 ± 0.09) relative

to the control (5.35 ± 0.10) ; Table 2), and soil pH was significantly higher ($P < 0.01$) in PAM + L than in other plots (C, G, PAM and PAM + G). This is due to the lime, which increases the Ca^{2+} content at cation exchange sites, where H^+ can be displaced. Lime application is well known to increase soil pH. Dissolution of lime significantly reduced exchangeable Al^{3+} and increased exchangeable Ca^{2+} (Li et al. 2018). Liming generally creates a more favorable environment for microbial activity and can lead to the net mineralization of organic forms of P (Ameyu 2019).

Available phosphorus (P_{av}) was lowest in the gypsum-treated plots (G) and highest in PAM + B (Table 2). P_{av} was significantly higher ($P < 0.001$) in PAM + B than in G only. The elevated P_{av} was associated with the improvement in soil pH (Table 2; $r = 0.75^{**}$; Table 5), and the decreased loss of P_{av} increased P_{av} in PAM + B. Our results are supported by similar studies (Li et al. 2019; Muluaem et al. 2021) showing that soil amendment with B and PAM reduces runoff, sedimentation, and washout, and that PAM improves soil cohesion and water-holding capacity (Muluaem et al. 2021). Furthermore, the PAM + B and B plots had significantly higher SOC concentrations ($P < 0.001$) than the control (C) and G plots. Biochar alters soil microbial abundance, activity, and community structure (Hardy et al. 2019), enhancing SOM degradation and SOC levels. Increased SR by microorganisms is thought to promote SOM degradation and improve soil fertility (Anjum and Khan 2021). Sorption to biochar-mediated aggregates is one of the most important reasons for the stabilization of SOC. A study in a maize field in the Central Plains of China showed that biochar additives increased SOC (Zhang et al. 2012).

However, TN, soil bulk density (SBD), and sand, silt, and clay contents did not change significantly among soil amendments during the study (Table 2). In contrast, Abdulrahman

Table 2 Physico-chemical properties (mean \pm SE) of topsoil samples (0–20 cm): all are clay loam

Amendment	P_{av} (ppm)	SOC (%)	TN (%)	pH (H_2O) 1:2.5	BD (g cm^{-3})	Clay (%)	Sand (%)	Silt (%)
B	$13.5 \pm 0.41^{\text{ab}}$	$1.84 \pm 0.04^{\text{a}}$	0.16 ± 0.01	$5.89 \pm 0.03^{\text{abcd}}$	1.29 ± 0.03	36 ± 2.67	32 ± 1.65	32 ± 1.46
C	$10.5 \pm 0.96^{\text{ab}}$	$1.51 \pm 0.04^{\text{b}}$	0.14 ± 0.01	$5.35 \pm 0.10^{\text{cd}}$	1.30 ± 0.03	37 ± 1.76	31 ± 1.65	32 ± 1.46
G	$8.6 \pm 0.32^{\text{b}}$	$1.52 \pm 0.04^{\text{b}}$	0.14 ± 0.01	$5.28 \pm 0.04^{\text{d}}$	1.31 ± 0.03	34 ± 1.76	33 ± 1.65	33 ± 1.46
L	$15.1 \pm 0.56^{\text{ab}}$	$1.64 \pm 0.04^{\text{ab}}$	0.15 ± 0.01	$5.91 \pm .06^{\text{abc}}$	1.29 ± 0.03	35 ± 3.06	33 ± 1.65	32 ± 1.46
PAM	$13.1 \pm 0.12^{\text{ab}}$	$1.62 \pm 0.04^{\text{ab}}$	0.14 ± 0.01	$5.38 \pm .08^{\text{bcd}}$	1.24 ± 0.03	35 ± 1.15	31 ± 1.65	34 ± 1.46
PAM+B	$16.3 \pm 1.14^{\text{a}}$	$1.87 \pm 0.04^{\text{a}}$	0.16 ± 0.01	$6.00 \pm 0.19^{\text{ab}}$	1.23 ± 0.03	36 ± 0.67	30 ± 1.65	34 ± 1.46
PAM+G	$10.2 \pm 0.90^{\text{ab}}$	$1.59 \pm 0.04^{\text{ab}}$	0.15 ± 0.01	$5.34 \pm 0.04^{\text{cd}}$	1.25 ± 0.03	35 ± 2.96	31 ± 1.65	34 ± 1.46
PAM+L	$14.3 \pm 1.11^{\text{ab}}$	$1.66 \pm 0.04^{\text{ab}}$	0.15 ± 0.01	$6.05 \pm 0.09^{\text{a}}$	1.23 ± 0.03	35 ± 1.15	31 ± 1.65	34 ± 1.46
<i>P</i> value	***	***	ns	**	ns	ns	ns	ns

Treatment levels within a column that doesn't share the same letters are significantly different

ns non-significant, B biochar, C control, G Gypsum, L lime, PAM Polyacrylamide, P_{av} available phosphorus, SOC soil organic carbon, TN Total nitrogen, BD bulk density, ($n = 3$)

** , ***Indicate the ANOVA p value is significant at $P < 0.01$, and $P < 0.001$, respectively, by Tukey test

et al. (2020) showed that B and PAM had a significant effect on SBD.

Paired *t*-test analysis comparing treatments with the control revealed a significantly increased mean SR in B, L, and PAM+L only (Table 3). These increases may be related to the improvement of pH, P_{av} , and SOC (Table 2). Improvement of soil property could have direct impact on soil microbial population. As soil property improved in the soil, the microbial activate for decomposition of soil organic matter becomes faster. This ultimately enhances soil respiration. Various studies have reported the function of soil amendments on soil properties; biochar affects soil physical and chemical properties (Chintala et al. 2013; Gross et al. 2021; Kookana et al. 2011; Layek et al. 2022; Li et al. 2017; Nigussie et al. 2012). Polyacrylamide and biochar also improve properties of soil (Alkhasha et al. 2018). PAM alone improves the structural stability of the soil (Mamedov et al. 2021). Biochar and lime improve the characteristics of acidic soils (Wu et al. 2020). Increased inputs of TN and SOC from the soil substrate were associated with an increase in SR (Wang et al. 2013).

Friedman's ANOVA revealed a significant difference ($\chi^2 = 28.86$, d.f. = 7, $P = 0.0003$) in the effects of soil amendments on SR (Table 4). Soil respiration was significantly less in PAM+B ($3.43 \pm 0.55 \mu\text{mol m}^{-2} \text{s}^{-1}$) than in B ($4.23 \pm 0.61 \text{CO}_2 \mu\text{mol m}^{-2} \text{s}^{-1}$) and PAM+L ($4.14 \pm 0.57 \mu\text{mol m}^{-2} \text{s}^{-1}$; $P < 0.05$; Table 4). The reason could be due to the addition of soil amendments in the soil mainly polyacrimide and biochar. Our study is supported by Awad et al. (2017) who studied that, PAM+B inhibits

Table 3 Paired *t*-test analysis of soil respiration between control and each soil amendment

No	Amendment	CO ₂ flux ($\mu\text{mol m}^{-2} \text{s}^{-1}$)	<i>P</i> value ($n = 18$)
1	C	3.60 ^b	0.0361
	B	4.19 ^a	
2	C	3.60 ^a	0.3931
	G	3.94 ^a	
3	C	3.60 ^b	0.0472
	L	4.20 ^a	
4	C	3.60 ^a	0.6008
	PAM	3.47 ^a	
5	C	3.60 ^a	0.3839
	PAM+B	3.38 ^a	
6	C	3.60 ^a	0.7338
	PAM+G	3.72 ^a	
7	C	3.60 ^b	0.0067
	PAM+L	4.09 ^a	

Different letters in the same column for paired treatments indicated significant difference at $P < 0.05$. See the section "Setup of experimental plots" for treatment codes

Table 4 ANOVA results of soil respiration in different soil amendments

Soil amendment	Min	Median	Max	Mean \pm SE	MR
	CO ₂ flux ($\mu\text{mol m}^{-2} \text{s}^{-1}$)				
C	0.59	1.69	16.01	3.64 ± 0.54	4.57 ^{ab}
B	0.40	2.06	17.04	4.23 ± 0.61	5.88 ^a
G	0.45	1.87	21.97	4.00 ± 0.66	4.53 ^{ab}
L	0.34	2.09	21.39	4.23 ± 0.67	5.11 ^{ab}
PAM	0.40	1.86	14.60	3.53 ± 0.53	4.46 ^{ab}
PAM+B	0.42	1.94	14.99	3.43 ± 0.55	3.79 ^b
PAM+G	0.31	2.41	14.83	3.70 ± 0.44	5.33 ^{ab}
PAM+L	0.70	2.39	19.52	4.14 ± 0.57	5.93 ^a
χ^2	28.86				
d.f	7				
<i>P</i> -value	0.0003				

Treatment levels with the same letter within a column are not significantly different at $P < 0.05$ ($n = 18$). *SE* standard error of the mean, *MR* mean rank. See the section "Setup of experimental plots" for treatment codes

microbial biomass C and reduces plant residue decomposition. Biochar reduces the organic C mineralization rate (Geng et al. 2022). PAM+B could be an excellent tool to reduce soil loss (Lee et al. 2015). The SOC (mean \pm SE) was slightly higher in PAM+B (1.87 ± 0.04) than in B (1.84 ± 0.04) and PAM+L (1.66 ± 0.04), though statistically not significant (Table 2). PAM+B similarly increased SOM (Abulaiti et al. 2022). The addition of PAM and B improved soil properties (Alkhasha et al. 2018), mainly because PAM significantly improves soil cohesion without increasing the rate of decomposition (Awad et al. 2013), and B significantly increases soil C sequestration and SOM (Hua et al. 2014). On the other hand, Awad et al. (2016) found that B and PAM, both individually and in combination, had no significant effect on total CO₂ emissions owing to their very slow rates of decomposition. Using B alone as a soil amendment increased SR (Li et al. 2021) and soil CO₂ production (Smith et al. 2010). Biochar addition improved soil microbial abundance, soil enzyme activity, community structure, and microbial biomass C (Hardy et al. 2019; Oladele 2019; Zhang et al. 2014). However, Šlapáková et al. (2018) found that it did not increase SR.

SR was significantly higher in PAM+L than in PAM+B (Table 4). The reason could be related to change in soil pH. Addition of lime may enhance the microbial activity and then soil respiration in the soil. As soil pH increases until optimum (5.5–6.5), more CO₂ is released into the atmosphere owing to increased soil microbial activity (Yusnaini et al. 2021). Liming increased the contribution of respiration to total C loss (Andersson and Valeur 1994). Lime in the soil is principally added to reduce soil pH (Mululem et al. 2021). Moreover, addition of lime in the soil

is a soil-forming process which involves the deposition and accumulation of calcium carbonate in soil, resulting in hardening of the soil. Among long-term no-till soils, limed soils had a higher SR and greater microbial C biomass than unlimed soils, attributed to the higher soil pH (Fuentes et al. 2006), which may increase soil turnover and organic mineralization (Rowley et al. 2018).

SR in each amendment in dry and wet seasons

We compared SR between the rainy season (June–August) and the dry season (December–February). Means differed significantly between seasons in all plots (Fig. 3). This seasonal difference in SR may be due to that in soil moisture. In all plots, soil moisture was significantly higher ($P < 0.05$) during the wet season, and soil temperature was significantly higher ($P < 0.05$) during the dry season (Fig. S1). This suggests that at our study site, soil moisture limits SR more than soil temperature does, especially during the dry season. Hashimoto et al. (2004) similarly reported that SR is higher during the rainy season. In addition, changes in soil moisture affect the activity of soil microbes (Orchard and Cook 1983).

The effect of soil moisture on SR can be assessed seasonally to represent environmental changes affecting their interaction (Jeong et al. 2018). In general, SR increases as soil moisture increases. Soil moisture was significantly correlated with SR, soil enzymes, and microbial biomass C (Tomar and Baishya 2020). Emissions in the wet season are significantly higher than those in the dry season owing to improved hydrothermal conditions, which are more favorable for soil microbes and the survival of plant roots (Cui et al. 2020; Orchard and Cook 1983; Rottassana et al. 2021;

Tomar and Baishya 2020). Boonriam et al. (2021) reported that SR in the rainy season is double that in the dry season in Thailand. Here, during the dry season, SR was similar among treatments (Fig. 3). Lacking enough water, drier soils cannot use available energy substrates (Orchard and Cook 1983).

Factors contributing to SR in various treatments

The rate of CO₂ release from the soil is used as a measure of microbial activity (Orchard and Cook 1983). It is determined mainly by the soil and by environmental parameters such as soil moisture and soil temperature. We found no significant correlation ($P > 0.05$) of SR with any soil property. However, we found positive associations ($P > 0.05$) with clay content ($r = 0.21$), soil pH ($r = 0.05$), TN ($r = 0.12$), and SOC ($r = 0.02$; Table 5).

Soil moisture was significantly ($P < 0.01$) related to SR in all treatments. The coefficients of the correlation between SR and soil moisture ranged from 0.42 to 0.64 (Table S1). Li et al. (2021) similarly showed that SR was positively correlated with soil moisture. Microbial respiration is linearly related to soil moisture (Cook and Orchard 2008). The influence of soil moisture on estimating SR in semi-arid habitats has also been reported (Meena et al. 2020).

There was no association with soil temperature during the dry season, but the daily variation of SR was positively correlated with that of soil temperature during the rainy season (Adachi et al. 2009). Soil temperature was significantly negatively correlated with SR in all plots (Table S1). Soil moisture but not soil temperature determines the incidence of SR (Hashimoto et al. 2004). In our case, soil moisture

Fig. 3 Pairwise comparison of SR (mean ± SE) between wet (W) and dry (D) seasons within treatments. Paired treatments with the different letters in the bars are significantly different at $P < 0.05$

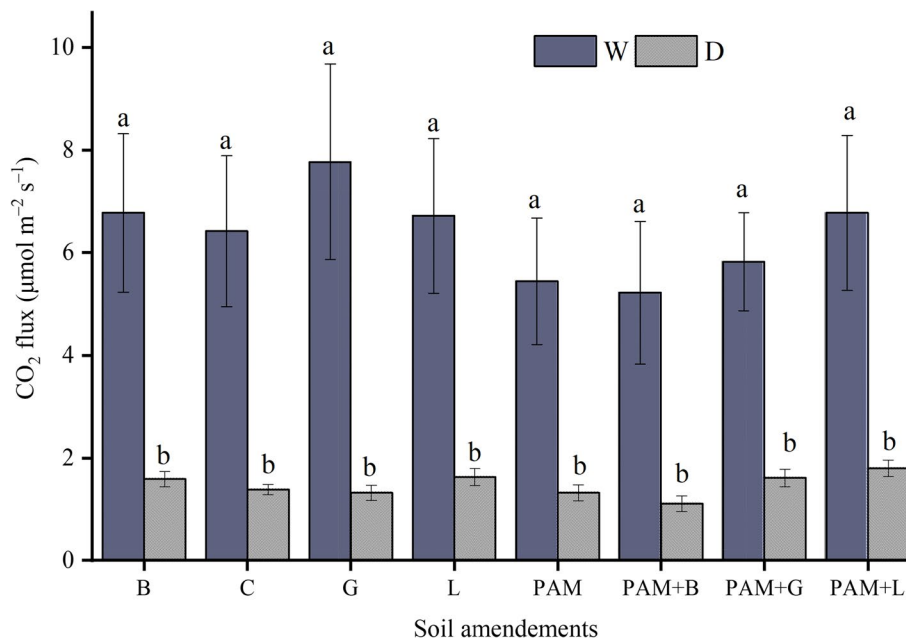


Table 5 Pearson's correlations of soil respiration with soil factors

	SR	P_{av}	SOC	TN	pH	SBD	Sand	Silt	Clay
SR	1								
P_{av}	-0.095 ns	1							
SOC	0.015 ns	0.600**	1						
TN	0.118 ns	0.296 ns	0.498**	1					
pH	0.051 ns	0.745**	0.568**	0.486*	1				
SBD	-0.065 ns	-0.235 ns	-0.279 ns	-0.357 ns	-0.117 ns	1			
Sand	0.048 ns	-0.016 ns	-0.224 ns	-0.315 ns	-0.076 ns	0.377 ns	1		
Silt	-0.329 ns	-0.018 ns	0.099 ns	0.155 ns	0.043 ns	0.282 ns	-0.297 ns	1	
Clay	0.211 ns	0.029 ns	0.126 ns	0.165 ns	0.036 ns	-0.559**	-0.676**	-0.503**	1

SR soil respiration ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$), P_{av} available P (mg kg^{-1}), SOC soil organic C (%), TN total N (%), SBD soil bulk density (g cm^{-3}), ns not significant ($n=3$)

* $P < 0.05$, ** $P < 0.01$

played an important role in influencing the change in SR relative to soil temperature.

Conclusions

Applications of soil amendments will have positive impact for improving soil properties so as to increase crop production. The applied soil amendment practices contributed to soil respiration in different ways during the study period. Soil respiration was significantly lower ($P < 0.05$) in PAM + B ($3.43 \pm 0.55 \mu\text{mol m}^{-2} \text{ s}^{-1}$) than in other treatments, including the controls, and was significantly higher ($P < 0.05$) in B ($4.23 \pm 0.61 \text{ CO}_2 \mu\text{mol m}^{-2} \text{ s}^{-1}$) and PAM + L ($4.14 \pm 0.57 \mu\text{mol m}^{-2} \text{ s}^{-1}$). Higher soil respiration values were observed in the wet season than in the dry season in all plots. This was mainly due to the influence of soil moisture effect on soil respiration. Since we focused on some soil amendments practices, further studies with different rates are required to assess the effect of soil amendments on soil respiration which is not yet studied to have more understanding on the relationship between soil respiration and soil amendments. More detailed studies are needed in a variety of agroecological settings to determine the best soil amendment methods that minimize CO_2 emissions.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s12665-024-11522-4>.

Acknowledgements This study was supported by the Science and Technology Research Partnership for Sustainable Development (SATREPS, grant number JPMJSA1601) and the Japan Science and Technology Agency (JST) / Japan International Cooperation Agency (JICA). We thank the staff of the SATREPS project at Bahir Dar University, Ethiopia.

Author contributions Genetu Fekadu: conceptualization, methodology, data collection, writing–draft manuscript, review editing, and

visualization. Enyew Adgo, Derege Tsegaye Meshesha, Nigussie Haregeweyn, Fei Peng, Atsushi Tsunekawa, and Mitsuru Tsubo: writing–review and editing, visualization, supervision, funding acquisition. Temesgen Mulualem, Simeneh Demissie, Birhanu Kebede and Gizachew Ayalew Tiruneh: writing–review and editing.

Funding This study was funded by the Science and Technology Research Partnership for Sustainable Development (grant number JPMJSA1601), and the Japan International Cooperation Agency (JICA)/ Japan Science and Technology Agency (JST).

Availability of data and materials The data analyzed during the current study are available from the corresponding author on rational request.

Declarations

Conflict of interest The authors declare no competing interests.

Ethical approval, consent to participate, consent publish Not applicable since no humans or animals were involved in this study.

References

- Abdulrahman MK, Al-Wazzan FA, Al-Jawadi LM (2020) Effect of polyacrylamide and biochar on calcareous soil moisture content and maize production under drip irrigation. *Plant Arch* 20(2):9505–9515
- Abegaz A, Tamene L, Abera W, Yaekob T, Hailu H, Nyawira SS et al (2020) Soil organic carbon dynamics along chrono-sequence land-use systems in the highlands of Ethiopia. *Agr Ecosyst Environ* 300(April):106997. <https://doi.org/10.1016/j.agee.2020.106997>
- Abulaiti A, She D, Liu Z, Sun X, Wang H (2022) Application of biochar and polyacrylamide to revitalize coastal saline soil quality to improve rice growth. *Environ Sci Pollut Res*. <https://doi.org/10.1007/s11356-022-23511-w>
- Adachi M, Ishida A, Bunyavejchewin S, Okuda T, Koizumi H (2009) Spatial and temporal variation in soil respiration in a seasonally dry tropical forest, Thailand. *J Trop Ecol* 25:531–539. <https://doi.org/10.1017/S026646740999006X>
- Aina OE, Olowoyo JO, Mugivhisa LL, Amoo SO (2018) Effect of different soil amendments on growth performance and levels of

- copper and zinc in *Lycopersicon esculentum*. *Nat Environ Pollut Technol* 17(1):255–259
- Albalasmeh AA, Hamdan EH, Gharaibeh MA, Hanandeh AE (2021) Improving aggregate stability and hydraulic properties of Sandy loam soil by applying polyacrylamide polymer. *Soil Tillage Res* 206:1–10. <https://doi.org/10.1016/j.still.2020.104821>
- Alkhasha A, Al-omran A, Aly A (2018) Effects of biochar and synthetic polymer on the hydro-physical properties of sandy soils. *Sustainability* 10:1–18. <https://doi.org/10.3390/su10124642>
- Amede T, Belachew T, Geta E (2001) Reversing the degradation of arable land in the Ethiopian Highlands. International center for research in agroforestry
- Ameu T (2019) A review on the potential effect of lime on soil properties and crop productivity improvements. *J Environ Earth Sci* 9(2):17–23. <https://doi.org/10.7176/JEES>
- Andersson S, Valeur I (1994) Influence of lime on soil respiration, leaching of DOC, and C/S relationships in the mor humus of a haplic podsol. *Environ Int* 20(1):81–88
- Angelova VR, Akova VI, Artinova NS, Ivanov KI (2013) The effect of organic amendments on soil chemical characteristics. *Bulg J Agr Sci* 19(5):958–971
- Anjum, Khan A (2021) Decomposition of soil organic matter is modulated by soil amendments. *Carbon Manag* 12(1):37–50. <https://doi.org/10.1080/17583004.2020.1865038>
- Asghari S, Abbasi F, Neyshabouri MR (2011) Effects of soil conditioners on physical quality and bromide transport properties in a sandy loam soil. *Biosys Eng* 109(1):90–97. <https://doi.org/10.1016/j.biosystemseng.2011.02.005>
- Awad YM, Blagodatskaya E, Ok YS, Kuzyakov Y (2012) Effects of polyacrylamide, biopolymer, and biochar on decomposition of soil organic matter and plant residues as determined by ¹⁴C and enzyme activities. *Eur J Soil Biol* 48:1–10. <https://doi.org/10.1016/j.ejsobi.2011.09.005>
- Awad YM, Blagodatskaya E, Ok YS, Kuzyakov Y (2013) Effects of polyacrylamide, biopolymer and biochar on the decomposition of ¹⁴C-labelled maize residues and on their stabilization in soil aggregates. *Eur J Soil Sci*. <https://doi.org/10.1111/ejss.12034>
- Awad YM, Pausch J, Ok YS, Kuzyakov Y (2016) Interactive effects of biochar and polyacrylamide on decomposition of maize rhizodeposits: implications from ¹⁴C labeling and microbial metabolic quotient. *J Soils Sediments*. <https://doi.org/10.1007/s11368-016-1576-1>
- Awad YM, Lee SS, Ok YS, Kuzyakov Y (2017) Effects of biochar and polyacrylamide on decomposition of soil organic matter and ¹⁴C-labeled alfalfa residues. *J Soils Sediments* 17:611–620. <https://doi.org/10.1007/s11368-016-1368-7>
- Bayu W, Rethman NFG, Hammes PS, Alemu G (2006) Effects of farmyard manure and inorganic fertilizers on sorghum growth, yield, and nitrogen use in a semi-arid area of Ethiopia. *J Plant Nutr* 29(2):391–407. <https://doi.org/10.1080/01904160500320962>
- Boonriam W, Suwanwaree P, Hasin S, Archawaakom T, Chanonmuang P, Yamada A (2021) Seasonal changes in spatial variation of soil respiration in dry evergreen forest, Sakaerat biosphere reserve, Thailand. *ScienceAsia* 47S:112–119. <https://doi.org/10.2306/scienceasia1513-1874.2021.S009>
- Bossolani JW, Crusciol CAC, Merloti LF, Moretti LG, Costa NR, Tsai SM, Kuramae EE (2020) Long-term lime and gypsum amendment increase nitrogen fixation and decrease nitrification and denitrification gene abundances in the rhizosphere and soil in a tropical no-till intercropping system. *Geoderma* 375:1–12. <https://doi.org/10.1016/j.geoderma.2020.114476>
- Bowden RD, Davidson E, Savage K, Arabia C, Steudler P (2004) Chronic nitrogen additions reduce total soil respiration and microbial respiration in temperate forest soils at the Harvard Forest. *For Ecol Manage* 196(1):43–56. <https://doi.org/10.1016/j.foreco.2004.03.011>
- Bouyoucos GJ (1962) Hydrometer method improved for making particle size analyses of soils. *Agronomy J* 54(5):464–465. <https://doi.org/10.2134/agronj1962.00021962005400050028x>
- Caesar-tonthat TC, Busscher WJ, Novak JM, Gaskin JF, Kim Y (2008) Effects of polyacrylamide and organic matter on microbes associated to soil aggregation of Norfolk loamy sand. *Appl Soil Ecol* 40:240–249. <https://doi.org/10.1016/j.apsoil.2008.04.008>
- Chen J, Sun X, Zheng J, Zhang X, Liu X, Bian R et al (2018) Biochar amendment changes temperature sensitivity of soil respiration and composition of microbial communities 3 years after incorporation in an organic carbon-poor dry cropland soil. *Bio Fertil Soils* 54:175–188. <https://doi.org/10.1007/s00374-017-1253-6>
- Chintala R, Mollinedo J, Schumacher TE, Malo DD, Julson JL (2013) Effect of biochar on chemical properties of acidic soil. *Arch Agron Soil Sci*. <https://doi.org/10.1080/03650340.2013.789870>
- Cook FJ, Orchard VA (2008) Relationships between soil respiration and soil moisture. *Soil Biol Biochem* 40(5):1013–1018. <https://doi.org/10.1016/j.soilbio.2007.12.012>
- Cui Y, Feng J, Liao L, Yu R, Zhang X, Liu Y et al (2020) Controls of temporal variations on soil respiration in a tropical lowland rainforest in Hainan Island, China. *Trop Conserv Serv* 13:1–14. <https://doi.org/10.1177/1940082920914902>
- Ebabu K, Tsunekawa A, Haregeweyn N, Adgo E, Meshesha DT, Aklog D et al (2019) Effects of land use and sustainable land management practices on runoff and soil loss in the Upper Blue Nile basin, Ethiopia. *Sci Total Environ* 648:1462–1475. <https://doi.org/10.1016/j.scitotenv.2018.08.273>
- Fekadu G, Adgo E, Meshesha DT, Tsunekawa A, Haregeweyn N, Peng F, Tsubo M, Masunaga T et al (2023a) Seasonal and diurnal soil respiration dynamics under different land management practices in the sub - tropical highland agroecology of Ethiopia. *Environ Monit Assess* 195(1):1–17. <https://doi.org/10.1007/s10661-022-10705-5>
- Fekadu G, Adgo E, Meshesha DT, Tsunekawa A, Haregeweyn N, Peng F, Mulualem T et al (2023b) Spatiotemporal variations of soil respiration under different land uses and their control in North-western Ethiopia. *Model Earth Syst Environ* 9:1–13. <https://doi.org/10.1007/s40808-023-01842-y>
- Fekadu G, Adgo E, Meshesha DT, Tsunekawa A, Haregeweyn N, Peng F, Tsubo M, Tassew A et al (2023c) Dry-season autotrophic and heterotrophic soil respiration in contrasting agroecological settings. *Acta Agric Scand Sect B -Soil Plant Sci* 73(1):54–66. <https://doi.org/10.1080/09064710.2023.2189613>
- Fuentes JP, Bezdicek DF, Flury M, Albrecht S, Smith JL (2006) Microbial activity affected by lime in a long-term no-till soil. *Soil Tillage Res* 88:123–131. <https://doi.org/10.1016/j.still.2005.05.001>
- Geng N, Kang X, Yan X, Yin N, Wang H, Pan H et al (2022) Biochar mitigation of soil acidification and carbon sequestration is influenced by materials and temperature. *Ecotoxicol Environ Saf* 232:113241. <https://doi.org/10.1016/j.ecoenv.2022.113241>
- Getahun M (2016) Soil characterization, classification and mapping of three twin watersheds in the Upper Blue Nile basin (Aba Gerima, Guder and Dibatie). Bahir Dar, Ethiopia
- Gomez KA, Gomez AA (1983) Statistical procedures for agricultural research, 2nd edn. Wiley
- Gross A, Bromm T, Glaser B (2021) Soil organic carbon sequestration after biochar application: a global meta-analysis. *Agronomy* 11:1–21. <https://doi.org/10.3390/agronomy11122474>
- Guyassa E, Raj AJ (2013) Assessment of biodiversity in cropland agroforestry and its role in livelihood development in dryland areas: a case study from Tigray region, Ethiopia. *J Agric Technol* 9(4):829–844
- Hailelassie A, Priess JA, Veldkamp E, Lesschen JP (2006) Smallholders' soil fertility management in the central highlands of Ethiopia: implications for nutrient stocks, balances and sustainability

- of agroecosystems. *Nutr Cycl Agroecosyst* 75:135–146. <https://doi.org/10.1007/s10705-006-9017-y>
- Hardy B, Sleutel S, Dufey JE, Cornelis JT (2019) The long-term effect of biochar on soil microbial abundance, activity and community structure is overwritten by land management. *Front Environ Sci* 7(July):1–14. <https://doi.org/10.3389/feenvs.2019.00110>
- Haregeweyn N, Tsunekawa A, Nyssen J, Poesen J, Tsubo M, Tsegaye Meshesha D et al (2015) Soil erosion and conservation in Ethiopia: a review. *Prog Phys Geogr* 39(6):750–774. <https://doi.org/10.1177/0309133315598725>
- Hashimoto S, Tanka N, Suzuki M, Inoue A, Takizawa H, Kosaka I et al (2004) Soil respiration and soil CO₂ concentration in a tropical forest, Thailand. *J for Res* 9:75–79. <https://doi.org/10.1007/s10310-003-0046-y>
- Holátko J, Hammerschmidt T, Kintl A, Danish S, Skarpa P, Latal O et al (2021) Effect of carbon-enriched digestate on the microbial soil activity. *PLoS ONE* 16(7):1–13. <https://doi.org/10.1371/journal.pone.0252262>
- Hua L, Lu Z, Ma H, Jin S (2014) Effect of biochar on carbon dioxide release, organic carbon accumulation, and aggregation of soil. *Environ Prog Sustain Energy* 33(3):941–946. <https://doi.org/10.1002/ep.11867>
- Hunegnaw Y, Alemayehu G, Ayalew D, Kassaye M (2021) Effects of soil amendments on selected soil chemical properties and productivity of tef (*Eragrostis tef* [Zucc.] Trotter) in the highlands of northwest Ethiopia. *Open Agric* 6(1):702–713. <https://doi.org/10.1515/opag-2021-0048>
- Hurni H (1988) Degradation and conservation of the resources in the Ethiopian highlands. *Mt Res Dev* 8(2):123–130
- Hurni H, Abate S, Bantider A, Debele B, Ludi E, Portner B et al (2010) Land degradation and sustainable land management in the highlands of Ethiopia. In: Hurni H, Wiesmann U (eds) *Global change and sustainable development: a synthesis of regional experiences from research partnerships*. Geographica Bernesia, pp 187–207
- Jeong S, Eom J, Park J, Chun J, Lee J (2018) Effect of precipitation on soil respiration in a temperate broad-leaved forest. *J Ecol Environ* 42(10):4–11. <https://doi.org/10.1186/s41610-018-0071-6>
- Jiang H, Deng Q, Zhou G, Hui D, Zhang D, Liu S et al (2013) Responses of soil respiration and its temperature/moisture sensitivity to precipitation in three subtropical forests in southern China. *Biogeosciences* 10(6):3963–3982. <https://doi.org/10.5194/bg-10-3963-2013>
- Jien SH, Wang CS (2013) Effects of biochar on soil properties and erosion potential in a highly weathered soil. *CATENA* 110:225–233. <https://doi.org/10.1016/j.catena.2013.06.021>
- Jien SH, Wang CC, Lee CH, Lee TY (2015) Stabilization of organic matter by biochar application in compost-amended soils with contrasting pH values and textures. *Sustainability (switzerland)* 7(10):13317–13333. <https://doi.org/10.3390/su71013317>
- Kebede B, Tsunekawa A, Haregeweyn N, Mamedov AI, Tsubo M, Fenta AA et al (2020) Effectiveness of polyacrylamide in reducing runoff and soil loss under consecutive rainfall storms. *Sustainability (switzerland)* 12(4):1–18. <https://doi.org/10.3390/su12041597>
- Kebede B, Tsunekawa A, Haregeweyn N, Tsubo M, Mulualet T, Mamedov AI et al (2022) Effect of Polyacrylamide integrated with other soil amendments on runoff and soil loss: case study from northwest Ethiopia. *Int Soil Water Conserv Res* 10(3):487–496. <https://doi.org/10.1016/j.iswcr.2021.12.001>
- Kookana RS, Sarmah AK, Zwieten LV, Krull E (2011) Biochar application to soil: agronomic and environmental benefits and unintended consequences (1st ed., vol. 112). *Adv Agron*. <https://doi.org/10.1016/B978-0-12-385538-1.00003-2>
- Layek J, Narzari R, Hazarika S, Das A, Rangappa K, Devi S et al (2022) Prospects of biochar for sustainable agriculture and carbon sequestration: an overview for eastern Himalayas. *Sustainability* 14:1–19. <https://doi.org/10.3390/su14116684>
- Lee SS, Shah HS, Awad YM, Kumar S, Ok YS (2015) Synergy effects of biochar and polyacrylamide on plants growth and soil erosion control. *Environ Earth Sci* 74(3):2463–2473. <https://doi.org/10.1007/s12665-015-4262-5>
- Lemma B, Williams S, Paustian K (2021) Long term soil carbon sequestration potential of smallholder croplands in southern Ethiopia with DAYCENT model. *J Environ Manag*. <https://doi.org/10.1016/j.jenvman.2021.112893>
- Li Y, Hu S, Chen J, Müller K, Li Y, Fu W et al (2017) Effects of biochar application in forest ecosystems on soil properties and greenhouse gas emissions: a review. *J Soils Sediments*. <https://doi.org/10.1007/s11368-017-1906-y>
- Li Y, Li Y, Chang SX, Yang Y, Fu S, Jiang P (2018) Biochar reduces soil heterotrophic respiration in a subtropical plantation through increasing soil organic carbon recalcitrancy and decreasing carbon-degrading microbial activity. *Soil Biol Biochem* 122:173–185. <https://doi.org/10.1016/j.soilbio.2018.04.019>
- Li F, Liang X, Niyungeko C, Sun T, Liu F, Arai Y (2019) Effects of biochar amendments on soil phosphorus transformation in agricultural soils. *Adv Agron*. <https://doi.org/10.1016/bs.agron.2019.07.002>
- Li D, Zhou J, Zhang Y, Sun T, An S, Jia H (2021) Effects of amendments on physicochemical properties and respiration rate of soil from the arid region of Northwest China. *Sustainability* 13(10):5332. <https://doi.org/10.3390/su13105332>
- Liang Y, Cai Y, Yan J, Li H (2019) Estimation of soil respiration by its driving factors based on multi-source data in a sub-alpine meadow in North China. *Sustainability*. <https://doi.org/10.3390/su11123274>
- Loper S, Shober AL, Wiese C, Denny GC, Stanley CD, Gilman EF (2010) Organic soil amendment and tillage affect soil quality and plant performance in simulated residential landscapes. *HortScience* 45(10):1522–1528. <https://doi.org/10.21273/hortsci.45.10.1522>
- Mamedov AI, Tsunekawa A, Haregeweyn N, Tsubo M, Fujimaki H, Kawai T et al (2021) Soil structure stability under different land uses in association with polyacrylamide effects. *Sustainability* 13:1–17. <https://doi.org/10.3390/su13031407>
- Meena A, Hanief M, Dinakaran J, Rao KS (2020) Soil moisture controls the spatio-temporal pattern of soil respiration under different land use systems in a semi-arid ecosystem of Delhi, India. *Ecol Process* 9(1):1–13. <https://doi.org/10.1186/s13717-020-0218-0>
- Mekonnen M, Getahun M (2020) Soil conservation practices contribution in trapping sediment and soil organic carbon, Minizr watershed, northwest highlands of Ethiopia. *J Soils Sediments* 20(5):2484–2494. <https://doi.org/10.1007/s11368-020-02611-5>
- Menichetti L, Ekblad A, Kätterer T (2013) Organic amendments affect $\delta^{13}C$ signature of soil respiration and soil organic C accumulation in a long-term field experiment in Sweden. *Eur J Soil Sci* 64(5):621–628. <https://doi.org/10.1111/ejss.12077>
- Mihretie FA, Tsunekawa A, Haregeweyn N, Adgo E, Tsubo M, Masunaga T et al (2021) Agro-economic evaluation of alternative crop management options for teff production in midland agro-ecology, Ethiopia. *Agriculture* 11:1–20. <https://doi.org/10.3390/agriculture11040298>
- Mihretie FA, Tsunekawa A, Haregeweyn N, Adgo E, Tsubo M, Masunaga T et al (2022) Exploring teff yield variability related with farm management and soil property in contrasting agro-ecologies in Ethiopia. *Agric Syst* 196:103338. <https://doi.org/10.1016/j.agsy.2021.103338>
- Mulualet T, Adgo E, Meshesha DT, Tsunekawa A, Haregeweyn N, Tsubo M et al (2021) Examining the impact of polyacrylamide and other soil amendments on soil fertility and crop yield in

- contrasting agroecological environments. *J Soil Sci Plant Nutr* 21(3):1817–1830. <https://doi.org/10.1007/s42729-021-00482-4>
- Mulualem T, Adgo E, Meshesha DT, Tsunekawa A, Haregeweyn N, Tsubo M et al (2022) Dual benefits of polyacrylamide and other soil amendments: mitigation of soil nutrient depletion and improvement of use-efficiency in midland agro-ecology, Ethiopia. *Land Degrad Dev*. <https://doi.org/10.1002/ldr.4367>
- Nett L, Ruppel S, Ruehlmann J, George E, Fink M (2012) Influence of soil amendment history on decomposition of recently applied organic amendments. *Soil Sci Soc Am J* 76(4):1290–1300. <https://doi.org/10.2136/sssaj2011.0190>
- Nigusie A, Kissi E, Misganaw M, Ambaw G (2012) Effect of biochar application on soil properties and nutrient uptake of lettuces (*Lactuca sativa*) grown in chromium polluted soils. *Am-Eurasian J Agric Environ Sci* 12(3):369–376
- Nyssen J, Poesen J, Moeyersons J, Haile M, Deckers J (2007) Dynamics of soil erosion rates and controlling factors in the Northern Ethiopian Highlands – towards a sediment budget. *Earth Surf Process Landf*. <https://doi.org/10.1002/esp>
- Oladele SO (2019) Effect of biochar amendment on soil enzymatic activities, carboxylate secretions and upland rice performance in a sandy clay loam Alfisol of Southwest Nigeria. *Sci Afr* 4:e00107. <https://doi.org/10.1016/j.sciaf.2019.e00107>
- Olsen SR, Cole CW, Watanabe FS, Dean LA (1954) Estimation of available phosphorus in soils by extraction with sodium bicarbonate. Circular, vol 939. USDA, Washington, DC, pp 119
- Orchard VA, Cook FJ (1983) Relationship between soil respiration and soil moisture. *Soil Biol Biochem* 15(4):447–453
- Peech M (1965) Hydrogen-ion activity. In: Black CA et al (eds) *Methods of soil analysis, part 2*, vol 60. American Society of Agronomy, pp 914–926
- Powers M, Kolka R, Bradford J, Palik B, Jurgensen M (2018) Forest floor and mineral soil respiration rates in a northern minnesota red pine chronosequence. *Forests* 16(9):1–15. <https://doi.org/10.3390/f9010016>
- Powlson DS, Gregory PJ, Whalley WR, Quinton JN, Hopkins DW, Whitmore AP et al (2011) Soil management in relation to sustainable agriculture and ecosystem services. *Food Policy* 36(SUPPL. 1):72–87. <https://doi.org/10.1016/j.foodpol.2010.11.025>
- Ray RL, Griffin RW, Fares A, Elhassan A, Awal R, Woldesenbet S, Risch E (2020) Soil CO₂ emission in response to organic amendments, temperature, and rainfall. *Sci Rep* 10(1):1–14. <https://doi.org/10.1038/s41598-020-62267-6>
- Rodtassana C, Unawong W, Yaemphum S, Chanthorn W, Chawchai S, Nathalang A et al (2021) Different responses of soil respiration to environmental factors across forest stages in a Southeast Asian forest. *Ecol Evol* 11:15430–15443. <https://doi.org/10.1002/ece3.8248>
- Rowley MC, Grand S, Verrecchia ÉP (2018) Calcium-mediated stabilisation of soil organic carbon. *Biogeochemistry* 137:27–49. <https://doi.org/10.1007/s10533-017-0410-1>
- Sales BK, Bryla DR, Trippe KM, Scagel CF, Strik BC, Sullivan DM (2022) Biochar as an alternative soil amendment for establishment of northern highbush blueberry. *HortScience* 57(2):277–285. <https://doi.org/10.21273/HORTSCI16257-21>
- Savage K, Davidson EA, Richardson AD (2008) A conceptual and practical approach to data quality and analysis procedures for high-frequency soil respiration measurements. *Funct Ecol* 22(6):1000–1007. <https://doi.org/10.1111/j.1365-2435.2008.01414.x>
- Seyoum B (2016) Assessment of soil fertility status of Vertisols under selected three land uses in Girar Jarso District of North Shoa Zone, Oromia National Regional State, Ethiopia. *Environ Syst Res* 5(1):1–16. <https://doi.org/10.1186/s40068-016-0069-y>
- Sheng H, Yang Y, Yang Z, Chen G, Xie J, Guo J, Zou S (2010) The dynamic response of soil respiration to land-use changes in subtropical China. *Glob Chang Biol* 16:1107–1121. <https://doi.org/10.1111/j.1365-2486.2009.01988.x>
- Shiferaw A (2011) Estimating soil loss rates for soil conservation planning in the borena woreda of South wollo highlands, Ethiopia. *J Sustain Dev Africa* 13(1):87–107
- Šlapáková B, Jeřábková J, Vofříšek K, Tejnecký V, Drábek O (2018) The biochar effect on soil respiration and nitrification. *Plant, Soil Environ* 64(3):114–119. <https://doi.org/10.17221/13/2018-PSE>
- Smith JL, Collins HP, Bailey VL (2010) The effect of young biochar on soil respiration. *Soil Biol Biochem* 42(12):2345–2347. <https://doi.org/10.1016/j.soilbio.2010.09.013>
- Sojka RE, Entry JA (2000) Influence of polyacrylamide application to soil on movement of microorganisms in runoff water. *Environ Pollut* 108:405–412
- Staben ML, Bezdicsek DF, Fauci MF, Smith JL (1997) Assessment of soil quality in conservation reserve program and wheat-fallow soils. *Soil Sci Soc Am J* 61(1):124–130. <https://doi.org/10.2136/sssaj1997.03615995006100010019x>
- Tamene L, Abera W, Demissie B, Desta G, Woldearegay K, Mekonnen K (2022) Soil erosion assessment in Ethiopia: a review. *J Soil Water Conserv* 77(2):1–14. <https://doi.org/10.2489/jswc.2022.00002>
- Tomar U, Baishya R (2020) Seasonality and moisture regime control soil respiration, enzyme activities, and soil microbial biomass carbon in a semi-arid forest of Delhi, India. *Ecol Process*. <https://doi.org/10.1186/s13717-020-00252-7>
- Wang W, Zeng W, Chen W, Zeng H, Fang J (2013) Soil respiration and organic carbon dynamics with grassland conversions to woodlands in temperate China. *PLoS ONE* 8(8):1–10. <https://doi.org/10.1371/journal.pone.0071986>
- Wong VNL, Dalal RC, Greene RSB (2009) Carbon dynamics of sodic and saline soils following gypsum and organic material additions: a laboratory incubation. *Appl Soil Ecol* 41:29–40. <https://doi.org/10.1016/j.apsoil.2008.08.006>
- Wu S, Zhang Y, Tan Q, Sun X, Wei W, Hu C (2020) Biochar is superior to lime in improving acidic soil properties and fruit quality of Satsuma mandarin. *Sci Total Environ*. <https://doi.org/10.1016/j.scitotenv.2020.136722>
- Xiao HB, Shi ZH, Li ZW, Chen J, Huang B, Yue ZJ, Zhan YM (2021) The regulatory effects of biotic and abiotic factors on soil respiration under different land-use types. *Ecol Ind* 127:1–9. <https://doi.org/10.1016/j.ecolind.2021.107787>
- Yusnaini S, Niswati A, Aini SN, Arif MAS, Dewi RP, Rivaie AA (2021) Changes in soil respiration after application of in situ soil amendment and phosphate fertilizer under soybean cultivation at Ultisol South Lampung, Indonesia. *IOP Conf Ser: Earth Environ Sci* 724(1):1–7. <https://doi.org/10.1088/1755-1315/724/1/012002>
- Zhang A, Liu Y, Pan G, Hussain Q, Li L, Zheng J, Zhang X (2012) Effect of biochar amendment on maize yield and greenhouse gas emissions from a soil organic carbon poor calcareous loamy soil from Central China Plain. *Plant Soil* 351:263–275. <https://doi.org/10.1007/s11104-011-0957-x>
- Zhang QZ, Dijkstra FA, Liu XR, Wang YD, Huang J, Lu N (2014) Effects of biochar on soil microbial biomass after four years of consecutive application in the north China Plain. *PLoS ONE* 9(7):1–8. <https://doi.org/10.1371/journal.pone.0102062>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.