Commentary: Regulating proNGF Action: Multiple Targets for Therapeutic Intervention

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Abstract Neurotrophins are initially synthesized as precursor forms that are cleaved to release *C*-terminal mature forms that bind to Trk receptors to initiate survival and differentiative responses. Recent studies suggest that the precursor form of NGF (proNGF) acts as a distinct ligand by binding to a receptor complex of p75 and sortilin to initiate cell death. Induction of proNGF and p75 has been observed in multiple pathological states and injury models in the central nervous system, and blockade of proNGF/p75 interaction is efficacious in limiting neuronal apoptosis. Multiple strategies that may act to limit proNGF action are considered as potential therapeutic targets for future development.

Keywords Neurotrophin \cdot Cell death \cdot p75 \cdot Neuronal injury

Neurotrophins consist of a family of proteins, including nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), neurotrophin 3 (NT-3), and neurotrophin 4 (NT-4/5) with well-characterized differentiative, survival, and synaptic activities in the developing and adult nervous system (Chao 2003). Although neurotrophins are initially synthesized as precursor forms (proneurotrophins), cleavage by intracellular proteases, such as furin or proconvertases, generates carboxyl- terminal mature neurotrophins. Mature neurotrophins bind to Trk receptor tyrosine kinases and with the p75 neurotrophin receptor, a tumor necrosis

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factor (TNFR) superfamily member (Huang and Reichardt 2001; Dechant and Barde 2002; Hempstead 2002). The interaction of mature neurotrophins with Trk receptors initiates many of the differentiative and synaptic activities of mature neurotrophins. Neurotrophins and p75 have also been found to induce apoptosis, detected using genetic models in which mice lacking p75 exhibit impaired sympathetic neuron or retinal ganglion cell death (Bamji et al. 1998; Frade and Barde 1999). However, cell death mediated by p75 in cultured cells required high concentrations of mature neurotrophins (Casaccia-Bonnefil et al. 1996; Yoon et al. 1998; Kenchappa et al. 2006), suggesting that another form of neurotrophins might selectively activate p75. Indeed, we have shown that the precursor form of NGF, or proNGF, can be released by cells and is a specific and selective ligand for p75 that initiates apoptosis (Lee et al. 2001). This unexpected finding suggests that the precursor form is a biologically active ligand, and that the mature and pro-forms of NGF may execute opposing actions. Subsequent studies demonstrate that proNGF preferentially interacts with high affinity to a heteromeric receptor complex of p75 and the type I transmembrane protein sortilin, wherein p75 binds to the mature domain of NGF, and sortilin interacts with the prodomain (Nykjaer et al. 2004). Thus, the specificity of neurotrophin action is dictated both by the form of ligand that is released (pro or mature), and by the differential utilization of receptors, with proNGF preferentially binding to and activating p75 and sortilin, and mature NGF binding to TrkA.

proNGF Effects in Development and Aging

The activity of proNGF in inducing apoptosis has been studied in development, and in aged animals. Although it

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would be attractive to evaluate a mouse model lacking the prodomain of NGF as a means to dissect proNGF and mature NGF actions, this has not been possible as neurotrophin prodomains subserve important functions in protein folding and intracellular trafficking (Suter et al. 1991; Chen et al. 2005). In addition, p75 interacts with all forms of neurotrophins and with multiple co-receptors, including TrkA, TrkB, and TrkC, to modulate neurotrophin activities, and with the Nogo receptor, Lingo-1, and ephrin A to alter axonal guidance (Schecterson and Bothwell 2008). p75 deficient mice have been a valuable tool to assess proNGF actions as neurons cultured from these animals are resistant to proNGF action. However, genetic deletion of p75 is likely to impart multiple and complex phenotypes based on effects of p75 in regulating Trk activity and on axon guidance by coupling with other receptor components. Thus, determining proNGF-specific phenotypes using p75 null animals is difficult. However, a sortilin deficient mouse has been generated and studied to evaluate proNGFinduced developmental apoptosis (Jansen et al. 2007). Consistent with the impaired apoptosis of developing retinal ganglion cells observed in E15.5 embryos deficient in p75 or NGF (48 or 56% reduction, respectively), embryos deficient in sortilin also exhibit reduced retinal ganglion cell death (63% reduction) (Frade and Barde 1998, 1999; Jansen et al. 2007). This result, together with prominent immunoreactivity for proNGF, but not mature NGF in the developing retina, strongly suggests that developmental elimination of post mitotic retinal ganglion cells is mediated by proNGF. In contrast, no reduction in sympathetic ganglion neurons was observed in neonatal sortilin deficient animals, suggesting that other, sortilin-independent mechanisms regulate sympathetic neuron elimination in vivo.

In the uninjured central and peripheral nervous systems, proNGF levels are very low in young adult rodents (Harrington et al. 2004; Jansen et al. 2007). However, proNGF levels are up regulated with advanced age. Specifically, proNGF levels are elevated in peripheral nerves of aged (60-week-old) mice, where it mediates agedependent sympathetic neuron death (Jansen et al. 2007). Although systematic and quantitative analysis of proNGF levels in postmortem human brains from aged but cognitively normal individuals is not available, elevated proN-GF levels have been observed in patients with Alzheimer's disease (Fahnestock et al. 2001; Pedraza et al. 2005). Interestingly, proNGF extracted from these human brains can mediate apoptosis of cultured sympathetic neurons. Further studies, using animal models that develop Alzheimer's-like pathology, may be informative in identifying whether proNGF is mechanistically linked to disease progression.

proNGF Effects in Models of Neuronal Injury

The effects of proNGF have been most extensively characterized in acute injury models in the peripheral and central nervous systems. In spinal cord injury, both proN-GF and p75 expression are induced and maintained for at least 1 week; in a related model of corticospinal motor neuron axotomy, proNGF, p75, and sortilin are all coordinately up regulated for 2 weeks (Brunello et al. 1990; Beattie et al. 2002; Harrington et al. 2004; Jansen et al. 2007). In the corticospinal axotomy model, genetic deletion of p75 or sortilin, or haploinsufficiency of NGF, largely rescues corticospinal neuron cell death. Importantly, infusion of function-blocking antibodies to the prodomain of proNGF also markedly reduces cell death, strongly suggesting that proNGF is an endogenous, inducible, proapoptotic cytokine.

proNGF has also demonstrated pro-apoptotic actions in cultured spinal motor neurons, cells which express p75 and sortilin (Domeniconi et al. 2007). In this study, reactive astrocytes were observed to upregulate proNGF production in response to peroxynitrite, an oxidant and producer of free radicals. Although these in vitro results have not yet been extended to in vivo models, these studies provide a potential therapeutic target for the treatment of motor neuron disease. Astrocytes also appear to be a significant source of the elevated proNGF levels that occur following pilocarpine induced seizures (Volosin et al. 2008). In this model, both proNGF and proBDNF are up regulated by astrocytes, but not microglia. Infusion of function-blocking antibody specific for the prodomain of NGF, following seizure induction, impairs hippocampal neuron death in vivo, suggesting that proNGF is the relevant neurotrophin in mediating the apoptotic effects. Additional studies indicate that proNGF is an apoptotic ligand in basal forebrain cholinergic neurons is aged rodents (Al-Shawi et al. 2008). In addition, injured sciatic neurons express proNGF and this may result in the loss of p75-expressing neurons following transection (Arnett et al. 2007). Further studies in the retina suggest that proNGF is induced in microglia in a model of retinal dystrophy (Srinivasan et al. 2004), and that sortilin and p75 are induced in retinal ganglion cells following elevations in intraocular pressure, suggesting that proNGF may play a role in the retinal neuron death that occurs in this ischemic setting (Wei et al. 2007). Collectively, these diverse models of injury or aging suggest that proNGF may be a potent proapoptotic ligand. However, cell death in each of these models is self-limiting, suggesting that there are endogenous regulatory mechanisms to modulate the actions of proNGF. These potential mechanisms, and their relevance as future therapeutic targets, will be considered below.

Regulation of Processing of proNGF to Mature NGF: Intracellular Conversion

In most adult tissues and in cultured cells, mature NGF is the predominant isoform, present at very low (nanogram) levels (Shetty et al. 2003). These observations pose the question of how proNGF, secreted in injury response states, escapes the mechanisms that normally ensure efficient intracellular conversion of proNGF to mature NGF. Mowla et al. (1999) have demonstrated that in heterologous neuroendocrine cells and hippocampal neurons, proNGF is cleaved efficiently by furin and the mature domain is trafficked to constitutive secretory vesicles, whereas the prodomain remains in the region of the cell body where it may be sorted to lysosomes for degradation. Indeed, secretion of a soluble prodomain has been very difficult to detect by most investigators, although Dicou (2008) has detected prodomain peptides in inflammatory states. These studies suggest that in uninjured cells, efficient conversion of proNGF to mature NGF, and constitutive secretion of mature NGF is the norm. However, the intracellular chaperones that bind to proNGF, and traffic it to the trans-Golgi network where furin cleavage occurs have not been well characterized. One candidate is sortilin, a VpS10p protein that has been well characterized, as described above, as a cell surface co-receptor with p75 for proNGF. However, sortilin has a predominantly intracellular location (McCormick et al. 2008), and has a well characterized role in regulating the intracellular trafficking of proBDNF to regulated secretory vesicles, and other cargo, including sphingomyelinase, to the lysosome (Chen et al. 2005; Ni and Morales 2006). Thus, it is possible that sortilin may promote the trafficking and degradation of the cleaved prodomain to lysosomes, although formal experimental proof of this is lacking. Other chaperones known to bind to the mature domains of neurotrophins, such as carboxypeptidase E that binds to BDNF, do not effectively bind to mature NGF (Lou et al. 2005). Therefore, the intracellular mechanisms that regulate proNGF intracellular trafficking, promote intracellular proNGF to mature NGF conversion, and regulate proNGF release remain to be determined.

Impaired Cleavage of Secreted proNGF

In several injury models in the central nervous system, proNGF expression is detectable for several days to weeks following injury. Surprisingly, little conversion of proNGF to mature NGF is observed in these vivo settings (Beattie et al. 2002; Harrington et al. 2004; Jansen et al. 2007), despite the susceptibility of recombinant proNGF to multiple proteases, including select matrix metalloproteinases (MMPs) and plasmin (Lee et al. 2001; Bruno and Cuello

2006: Althaus and Kloppner 2006). These observations suggest that proteolysis of extracellular proNGF is regulated following in vivo injury, and may involve the coordinate induction of known inhibitors of MMPs and plasmin. These include tissue inhibitors of metalloproteinase (TIMPs), neuroserpin, and alpha-2 macroglobulin, proteins that are transcriptionally regulated and induced in neurodegenerative disease, and with neuronal excitotoxicity (Bruno and Cuello 2006). Indeed, alterations in MMP and TIMP expression have been documented in Huntingtons and Parkinson's diseases (Dzwonek et al. 2004; Jaworski et al. 1999; Lorenzi et al. 2003). The detection of intact proNGF in the cerebral spinal fluid of rodents following spinal injury suggests that inhibitors may also be present in significant amounts, a hypothesis which is awaiting formal evaluation.

Induction of p75

Expression of the p75 receptor has emerged as a key regulatory element in proNGF-induced cell death. In most adult tissues, p75 is expressed at low levels, in contrast to higher and more widespread distribution in development (Yang et al. 2009; Roux and Barker 2002). However, in pathologic states, including seizure, brain injury, ischemia, and excitotoxicity, p75 expression is induced, as noted above. The significant reduction of injury-induced apoptosis observed in p75 deficient mice (Troy et al. 2002; Harrington et al. 2004) underscores the importance of p75 induction in determining cell loss following injury.

The molecular mechanisms that regulate p75 expression, both in development and in injury, remain largely unknown. The p75 promoter resembles a housekeeping gene, with high GC content, multiple Sp1 binding sites, but no TATA or CAAT elements (Sehgal et al. 1988; Patil et al. 1990). A transgenic approach has been undertaken to evaluate p75 transcriptional regulation (Huber and Chao 1995; Carroll et al. 1995). In one study, mice harboring 4 kb of 5' sequence of human p75 and the human p75 cDNA as a minigene exhibited expression by mesenchymal cells during development, mimicking endogenous expression (Huber and Chao 1995). In addition, this p75 minigene was induced following sciatic nerve injury, although expression in uninjured peripheral neurons was lacking. In a second approach, analysis of a 8.4 kb murine p75 promoter, using a lacZ reporter, documented appropriate expression in peripheral neurons and the retina, but no induction in Schwann cells during Wallerian degeneration (Carroll et al. 1995). Collectively, these observations suggest that multiple promoter elements exist, and that injury-response elements that regulate neuronal, but not glial expression, are encoded within the proximal 4 kb of the promoter.

More recent studies have evaluated the role of hypoosmolar stress in inducing p75, as brain edema is a common complication of seizures, and traumatic brain injury (Peterson and Bogenmann 2003; Ramos et al. 2007). In examination of 25 kb upstream of rat p75 promoter, proximal Sp1 elements were found to be critical for p75 induction. Furthermore, p75 transcription appeared to be regulated by the enhanced expression of Sp1, mediated by inhibition of Sp1 degradation in hypo-osmolar states. High levels of expression of Sp1 persist in neurons for at least 14 days following ischemic injury, providing a mechanism by which prolonged induction of p75 may occur. Interestingly, sortilin expression is unaffected by hypo-osmolarity, and indeed the elements that regulate transcription of sortilin are unknown.

Molecular Strategies to Attenuate proNGF Action

As summarized above, studies by multiple laboratories provide mounting evidence that the induction of proNGF and p75 in several pathophysiologically relevant states may result in cellular apoptosis in a p75 and sortilin dependent manner. The low levels of p75 and proNGF in the uninjured central nervous system, and the observations that proNGF and p75 induction occurs over several hours to days, suggests that a window of opportunity exists during which administration of pharmacologic agents to block the induction of ligand and receptors, or their interaction, may attenuate neuronal apoptosis. One successful strategy has been through the identification of small molecular inhibitors that impair the interaction of p75 with its ligands. Through in silico modeling, small molecules have been identified that interact with a p75 structural domain important for mature NGF binding; in addition, these molecules block proNGF actions in cultured neurons (Massa et al. 2006). The development of these, as well as other molecules identified by screening or modeling approaches to impair proNGF/p75/sortilin interactions may provide useful reagents to block proNGF actions. Although the crystallographic structure of p75 with mature NGF is available, as well as the crystallographic structure of sortilin, the structure of the proNGF/p75/sortilin complex has remained elusive (He and Garcia 2004; Quistgaard et al. 2009), but may provide information for the development of antagonists in the future.

Other pharmacological strategies may involve the development of drugs that block the induction of p75 or proNGF. As noted above, the key promoter elements of p75 that direct neuronal and injury-responsive expression remain to be described, and far less is known about NGF transcriptional regulation than related neurotrophins, such as BDNF. However, minocycline treatment of rodents with

spinal cord injury has been shown to attenuate proNGF and p75 induction, suggesting that this approach may be feasible, and could be optimized once the relevant promoter elements have been characterized (Yune et al. 2007).

Lastly, the activation of intracellular or extracellular proteases to specifically cleave proNGF to mature NGF is another attractive target. To this end, a more detailed understanding of the mechanisms that regulate intracellular trafficking of proNGF in injured cells, and permit inefficient intracellular cleavage is needed. In addition, the stability of proNGF in the injured central nervous system suggests that specific protease inhibitors in the local inflammatory environment may prevent efficient extracellular cleavage of proNGF. Thus, quantitative assessment of locally produced proteases and their specific inhibitors in the injured central nervous system may provide candidate molecules for manipulation, and to promote proNGF to mature NGF conversion.

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References

- Al-Shawi R, Hafner A, Olsen J, Chun S, Raxa S, Thrasivoulou C, Lovestone S, Killick R, Simons P, Cowen T (2008) Neurotoxic and neurotrophic roles of proNGF and the receptor sortilin in the adult and ageing nervous system. Eur J NeuroSci 27:2103–2114
- Althaus HH, Kloppner S (2006) Mature pig oligodendrocytes rapidly process human recombinant pro-nerve growth factor and do not undergo cell death. J Neurochem 98:506–517
- Arnett MG, Ryals JM, Wright DE (2007) pro-NGF, sortilin and p75NTR: potential mediator of injury induced apoptosis in the mouse dorsal root ganglion. Brain Res 1183:32–42
- Bamji SX, Majdan M, Pozniak CD, Belliveau DJ, Aloyz R, Kohn J, Causing CG, Miller FD (1998) The p75 neurotrophin receptor mediates neuronal apoptosis and is essential for naturally occurring sympathetic neuron death. J Cell Biol 140:911–923
- Beattie MS, Harrington AW, Lee R, Kim JY, Boyce SL, Longo FM, Bresnahan JC, Hempstead BL, Yoon SO (2002) proNGF induces p75-mediated death of oligodendrocytes following spinal cord injury. Neuron 36:375–386
- Brunello N, Reynolds M, Wrathall JR, Mocchetti I (1990) Increased nerve growth factor receptor mRNA in contused rat spinal cord. Neurosci Lett 118:238–240
- Bruno MA, Cuello AC (2006) Activity-dependent release of precursor nerve growth factor, conversion to mature nerve growth factor and its degradation by a protease cascade. Proc Natl Acad Sci USA 103:6735–6740
- Carroll SL, Schweitzer JB, Holtzman DM, Miller ML, Sclar GM, Milbrandt J (1995) Elements in the 5' flanking sequences of the mouse low-affinity NGF receptor gene direct appropriate CNS, but not PNS, expression in transgenic mice. J Neurosci 15:3342– 3356
- Casaccia-Bonnefil P, Carter BD, Dobrowsky RT, Chao MV (1996) Death of oligodendrocytes mediated by the interaction of nerve growth factor with its receptor p75. Nature 383:716–719
- Chao MV (2003) Neurotrophins and their receptor: a convergence point for many signaling pathways. Nat Rev Neurosci 4:299–309

- Chen ZY, Ieraci A, Teng H, Dall H, Meng CX, Herrera DG, Nykjaer A, Hempstead BL, Lee FS (2005) Sortilin controls the intracellular sorting of brain-derived neurotrophic factor to the regulated secretory pathway. J Neurosci 25:6156–6166
- Dechant G, Barde YA (2002) The neurotrophin receptor p75(NTR): novel functions and implications for diseases of the nervous system. Nat Neurosci 5:1131–1136
- Dicou E (2008) High levels of the proNGF peptides LIP1 and LIP2 in the serum and synovial fluid of rheumatoid arthritis patients: evidence for two new cytokines. J Neuroimmunol 194:143–146
- Domeniconi M, Hempstead BL, Chao MV (2007) pro-NGF secreted by astrocytes promotes motor neuron cell death. Mol Cell Neurosci 34:271–279
- Dzwonek J, Rylski M, Kaczmarek L (2004) Matrix metalloproteinases and their endogenous inhibitors in neuronal physiology of the adult brain. FEBS Lett 567:129–135
- Fahnestock M, Michalski B, Xu B, Coughlin MD (2001) The precursor pro-nerve growth factor is the predominant form of nerve growth factor in brain and is increased in Alzheimer's disease. Mol Cell Neurosci 18:210–220
- Frade JM, Barde YA (1998) Microglial-derived nerve growth factor causes cell death in the developing retina. Neuron 20:35–41
- Frade JM, Barde YA (1999) Genetic evidence for cell death mediated by nerve growth factor and the neurotrophin receptor p75 in the developing mouse retina and spinal cord. Development 126:683– 690
- Harrington AW, Leiner B, Blechschmitt C, Arevalo JC, Lee R, Morl K, Meyer M, Hempstead BL, Yoon SY, Giehl KM (2004) Secreted proNGF is a pathophysiolocial death-inducing ligand after CNS injury. Proc Natl Acad Sci USA 101:6226–6230
- He XL, Garcia KC (2004) Structure of nerve growth factor complexed with the shared neurotrophin receptor p75. Science 304:870–875
- Hempstead BL (2002) The many faces of p75NTR. Curr Opin Neurobiol 12:260–267
- Huang EJ, Reichardt LF (2001) Trk receptors: roles in neuronal signal transduction. Annu Rev Biochem 72:609–642
- Huber LJ, Chao MV (1995) Mesenchymal and neuronal cell expression of the p75 neurotrophin receptor gene occur by different mechanisms. Dev Biol 167:227–238
- Jansen P, Giehl K, Nyengaard JR, Teng K, Lioubinski O, Sjoegaard SS, Breiderhoff T, Gotthardt M, Lin F, Eilers A, Petersen CM, Lewin GR, Hempstead BL, Willnow TE, Nykjaer A (2007) Roles for the pro-neurotrophin receptor sortilin in neuronal development, aging and brain injury. Nat Neurosci 10:1449–1457
- Jaworski J, Biedermann IW, Lapinska J, Szkiarczyk A, Figiel I, Konopka D, Nowicka D, Filipkowski RK, Hetman M, Kowalczyk A, Kaczmarek L (1999) Neuronal excitation-driven and AP-1 dependent activation of tissue inhibitor of metalloproteinase-1 gene expression in rodent hippocampus. J Biol Chem 274:28106–28112
- Kenchappa RS, Zampieri N, Chao MV, Barker PA, Teng HK, Hempstead BL, Carter BD (2006) Ligand-dependent cleavage of the p75 receptor is necessary for NRIF nuclear translocation and apoptosis in sympathetic neurons. Neuron 50:219–232
- Lee R, Kermani P, Teng KK, Hempstead BL (2001) Regulation of cell survival by secreted pro-neurotrophins. Science 294:1945– 1948
- Lorenzi S, Albers DS, LeWitt PA, Chirichigno JW, Hilgenberg SL, Cudkowicz ME, Beal MF (2003) Tissue inhibitors of matrix metalloproteinases are elevated in cerebrospinal fluid of neurodegenerative diseases. J Neurol Sci 207:71–76
- Lou H, Kim SK, Zaitsev E, Snell CR, Lu B, Loh YP (2005) Sorting and activity-dependent secretion of BDNF require interaction of a specific motif with the sorting receptor carboxypeptidase E. Neuron 45:245–255

- Massa SM, Xie Y, Yang T, Harrington AW, Kim ML, Yoon SO, Kraemer R, Moore LA, Hempstead BL, Longo FM (2006) Small, nonpeptide p75NTR ligands induce survival signaling and inhibit proNGF-induced death. J Neurosci 26:5230–5288
- McCormick PJ, Dumaresq-Doiron K, Pluviose AS, Pichette V, Tosato G, Lefrancois S (2008) Palmitoylation controls recycling in lyosomal sorting and trafficking. Traffic 9:1984–1997
- Mowla SJ, Pareek S, Farhadi HF, Petrecca K, Fawcett JP, Seidah NG, Morris SJ, Sossin WS, Murphy RA (1999) Differential sorting of nerve growth factor and brain-derived neurotrophic factor in hippocampal neurons. J Neurosci 19:2069–2080
- Ni X, Morales CR (2006) The lysosomal trafficking of acid sphingomyelinase is mediated by sortilin and mannose 6-phosphate receptor. Traffic 7:889–902
- Nykjaer A, Lee R, Teng KK, Jansen P, Madsen P, Nielsen MS, Jacobsen C, Kliemannel M, Schwarz E, Willnow TE, Hempstead BL, Petersen CM (2004) Sortilin is essential for proNGFinduced neuronal cell death. Nature 427:843–848
- Patil N, Lacy E, Chao MV (1990) Specific neuronal expression of human NGF receptors in the basal forebrain and cerebellum of transgenic mice. Neuron 4:437–447
- Pedraza CE, Podlesniy P, Vidal N, Arevalo JC, Lee R, Hempstead BL, Ferrer I, Iglesias M, Espinet C (2005) pro-NGF isolated from the human brain affected by Alzheimer's disease induces neuronal apoptosis mediated by p75NTR. Am J Pathol 166:533– 543
- Peterson S, Bogenmann E (2003) Osmotic swelling induces p75 neurotrophin receptor (p75NTR) expression via nitric oxide. J Biol Chem 278:33943–33950
- Quistgaard EM, Madsen P, Grøftehauge MK, Nissen P, Petersen CM, Thirup SS (2009) Ligands bind to Sortilin in the tunnel of a tenbladed beta propeller domain. Nat Struct Mol Biol 16:96–98
- Ramos A, Ho WC, Forte S, Dickson K, Boutilier J, Favell K, Barker PA (2007) Hypo-omolar stress induces p75NTR expression by activating Sp1-dependent transcription. J Neurosci 27:1498– 1506
- Roux PP, Barker PA (2002) Neurotrophin signaling through the p75 receptor. Prog Neurobiol 67:203–233
- Schecterson LC, Bothwell M (2008) An all-purpose tool for axon guidance. Sci Signal 1(47):pe50
- Sehgal A, Patil N, Chao M (1988) A constitutive promoter directs expression of the nerve growth factor receptor gene. Mol Cell Biol 8:3160–3167
- Shetty AK, Zaman V, Shetty GA (2003) Hippocampal neurotrophin levels in a kainate model of temporal lobe epilepsy: a lack of correlation between brain-derived neurotrophic factor content and progression of aberrant dentate mossy fiber sprouting. J Neurochem 87:147–159
- Srinivasan B, Roque CH, Hempstead BL, Al-Ubaidi MR, Roque RS (2004) Microglia-derived pronerve growth factor promotes photoreceptor cell death via p75 neurotrophin receptor. J Biol Chem 279:41839–41943
- Suter U, Heymach JV Jr, Shooter EM (1991) Two conserved domains in the NGF propeptide are necessary and sufficient for the biosynthesis of correctly processed and biologically active NGF. EMBO J 10:2395–2400
- Troy CM, Friedman JE, Friedman WJ (2002) Mechanisms of p75mediated death of hippocampal neurons. Roles of caspases. J Biol Chem 277:34295–34302
- Volosin M, Trotter C, Cragnolini A, Kenchappa RS, Light M, Hempstead BL, Carter BD, Friedman WJ (2008) Induction of proneurotrophins and activation of p75NTR-mediated apoptosis via neurotrophin receptor-interacting factor in hippocampal neurons after seizures. J Neurosci 28:9870–9879
- Wei Y, Wang N, Lu Q, Zhang N, Sheng D, Li J (2007) Enhanced protein expressions of sortilin and p75NTR in retina of rat

following elevated intraocular pressure induced retinal ischemia. Neurosci Lett 429:169–174

- Yang J, Siao CJ, Nagappan G, Marinic T, Jing D, McGrath K, Chen ZY, Mark W, Tessarollo L, Lee FS, Lu B, Hempstead BL (2009) Neuronal release of proBDNF. Nat Neurosci 12:113–115
- Yoon SO, Casaccia-Bonefil P, Carter BD, Chao MV (1998) Competitive signaling between TrkA and p75 nerve growth

factor receptors determines cell survival. J Neurosci 18:3273-3281

Yune TY, Lee JY, Jung GY, Kim SJ, Jiang MH, Kim YM, Oh YJ, Markelonis GJ, Oh TH (2007) Minocycline alleviates death of oligodendrocytes by inhibiting pro-nerve growth factor production in microglia after spinal cord injury. J Neurosci 27:7751– 7761