



Probiotics: an Antibiotic Replacement Strategy for Healthy Broilers and Productive Rearing

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Abstract

Pathogens develop resistance to antibiotics at a rate much faster than the discovery of new antimicrobial compounds. Reports of multidrug-resistant bacteria isolated from broilers, and the possibility that these strains may spread diseases amongst humans, prompted many European countries to ban the inclusion of antibiotics in feed. Probiotics added to broiler feed controlled a number of bacterial infections. A combination of *Enterococcus faecium*, *Pediococcus acidilactici*, *Bacillus animalis*, *Lactobacillus salivarius* and *Lactobacillus reuteri* decreased the colonisation of *Campylobacter jejuni* and *Salmonella* Enteritidis in the gastro-intestinal tract (GIT) of broilers, whereas *Bacillus subtilis* improved feed conversion, intestinal morphology, stimulated the immune system and inhibited the colonisation of *Campylobacter jejuni*, *Escherichia coli* and *Salmonella* Minnesota. *Lactobacillus salivarius* and *Pediococcus parvulus* improved weight gain, bone characteristics, intestinal morphology and immune response, and decreased the colonisation of *S. Enteritidis*. *Lactobacillus crispatus*, *L. salivarius*, *Lactobacillus gallinarum*, *Lactobacillus johnsonii*, *Enterococcus faecalis* and *Bacillus amyloliquefaciens* decreased the *Salmonella* count and led to an increase in lysozyme and T lymphocytes. Probiotics may also improve feed digestion through production of phytases, lipases, amylases and proteases or stimulate the GIT to secrete digestive enzymes. Some strains increase the nutritional value of feed by production of vitamins, exopolysaccharides and antioxidants. Bacteriocins, if produced, regulate pathogen numbers in the GIT and keep pro-inflammatory and anti-inflammatory reactions in balance.

Keywords Probiotics · Poultry · Productive rearing

Introduction

The last 70 years of research on poultry has largely been devoted to genetic selection and improvement of feed compositions [1]. Less progress has been made on the control of microbial infections caused by *Staphylococcus*, *Pseudomonas*, *Escherichia*, *Salmonella*, *Streptococcus*, *Campylobacter*, *Yersinia*, *Clostridium*, *Bacillus*, *Mycobacterium*, *Klebsiella*, *Enterococcus* and *Proteus* spp. [2, 3]. Broilers are more susceptible to bacterial infections during the first few weeks when the immune system is not fully developed [4]. Furthermore, developing of a stable gut microbiome may take as long as 8 weeks [5]. The longer the delay in reaching bacterial homeostasis, the higher the risk of contracting bacterial infections [6]. To minimise the chances

of contracting bacterial infections, broilers are housed in confined facilities [7]. In many cases, farmers are still supplementing feed with antibiotics [8]. The irresponsible, non-therapeutic, use of antibiotics in poultry (and other animal) feed has led to a rapid increase in bacterial species resistant to antibiotics [9, 10]. Staphylococci associated with poultry farms were reported resistant to oxacillin and tetracycline [3]. Strains of *Pseudomonas* isolated from poultry in Ghana were resistant to cephalosporins, carbapenems, penicillins, quinolones, monobactam and aminoglycoside [11]. *Pseudomonas aeruginosa* strains isolated from poultry in Nigeria were resistant to β -lactams, tetracycline, tobramycin, nitrofurantoin and sulfamethoxazole-trimethoprim [12], whereas strains isolated from chickens in Pakistan showed resistance towards ceftriaxone, meropenem, ciprofloxacin, erythromycin and colistin [13]. *Escherichia coli* strains isolated from broilers were resistant to tetracycline, sulfamethoxazole, streptomycin, trimethoprim, ciprofloxacin, ampicillin, spectinomycin, nalidixic acid, chloramphenicol, neomycin and gentamicin [14]. Similar reports of multi-drug resistance have been reported for *Salmonella* [15], *Streptococcus* [16],

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Campylobacter [17, 18], *Yersinia* [19, 20], *Clostridium* [21–23], *Bacillus* [24], *Mycobacterium* [25], *Enterococcus* [26, 27] and *Proteus* spp. [28].

To control the spreading of antibiotic-resistant bacteria, health authorities in many European countries have now restricted the use of antibiotics in poultry feed [29]. Sweden was the first to ban the use of antimicrobials for growth promotion (year 1986) and prophylactic use (year 1988) [30]. Denmark, The Netherlands, the UK and other European Union countries followed suit [30]. In 2011, these countries banned the use of all essential antibiotics as prophylactic agents [31]. Despite these regulations, at least 60% of all antibiotics currently produced are used in livestock production, including poultry [32, 33].

Since the banning of antibiotics, probiotics [34, 35], prebiotics [34], synbiotics [36, 37], phytobiotics [38], enzymes [39, 40], essential oils [41, 42] and fatty acids [43] were proposed as possible alternatives. Probiotics are live microorganisms that, when administered in adequate cell numbers, confer a health benefit to the host. *Enterococcus faecium*, combined with *Pediococcus acidilactici*, *Bacillus animalis*, *Lactobacillus salivarius* and *Lactobacillus reuteri* (commercially available as PoultryStar®) decreased the colonisation of *Campylobacter jejuni* and *Salmonella* Enteritidis [44, 45]. A probiotic containing *Bacillus subtilis* (CLOSTAT™) improved feed conversion, intestinal morphology, enhanced the immune response, and inhibited gastro-intestinal tract (GIT) colonisation by *C. jejuni*, *Escherichia coli* and *Salmonella* Minnesota [46–49]. A combination of *L. salivarius* and *Pediococcus parvulus* (Floramax® B11) improved weight gain, bone characteristics, intestinal morphology and immune response, and decreased the colonisation of *S. Enteritidis* [50, 51]. Neveling and co-authors [52] have shown that a combination of *Lactobacillus crispatus*, *L. salivarius*, *Lactobacillus gallinarum*, *Lactobacillus johnsonii*, *Enterococcus faecalis* and *Bacillus amyloliquefaciens* inhibited the colonisation of *Salmonella* in the GIT of broilers. Broilers treated with the multi-species probiotic had higher levels of lysozyme in their serum and higher T lymphocyte responses compared to control birds.

The use of probiotics in poultry feed is, however, still far from being common practice. While the murine in vivo model is perhaps the most commonly used for mechanistic studies of probiotics, only a few reports have been published on the role these microorganisms play in keeping the GIT of poultry healthy [53–55], let alone the interactions amongst gut microorganisms and communication between probiotics and broiler gut epithelial cells [56]. Findings from these studies are difficult to compare. Apart from insufficient evidence-based trials, none of the methods used by the different authors have been standardised. In-depth research needs to be done on the gut microbiota of broilers and the inter-microbial- and microbiota-host interactions.

This review assesses the benefits of probiotic use as feed additives for broilers and discusses the mechanisms by which probiotics improve health and growth performance. The role of probiotics in digestion, followed by modulation of the microbiome and physiology of broilers are reviewed. The effect probiotics might have on meat quality is also addressed.

Role of Probiotics in Digestion

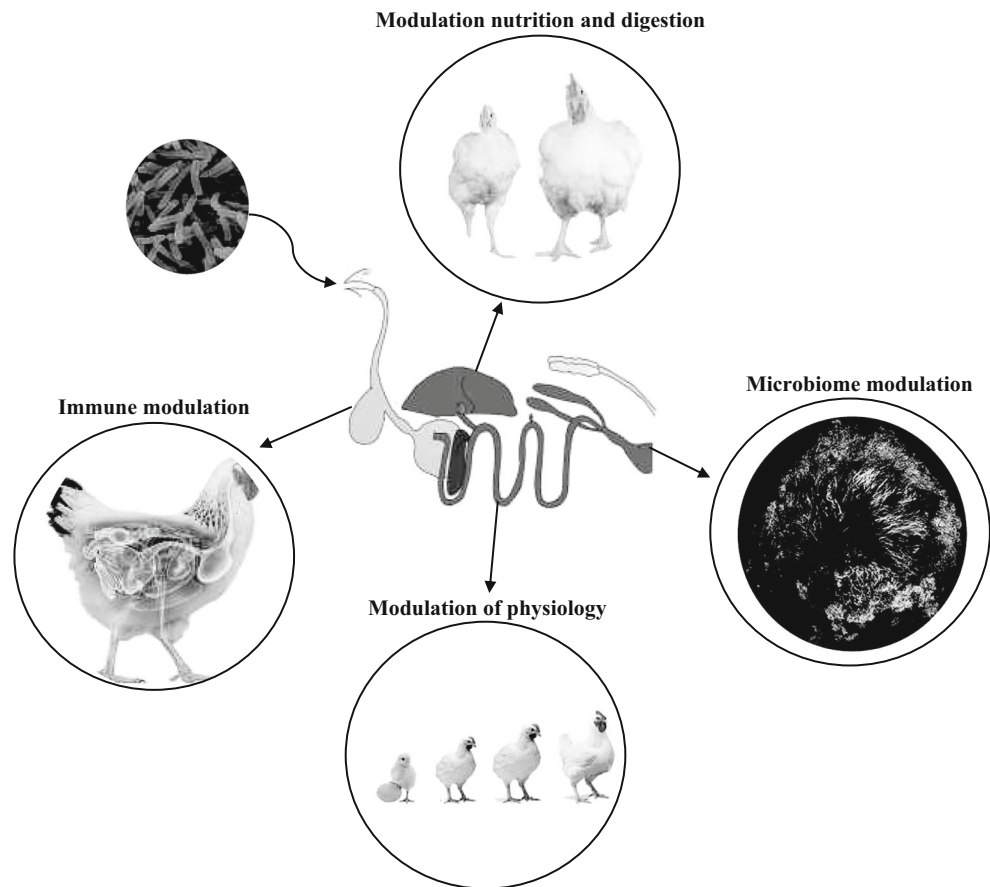
Probiotics convert less degradable compounds into more digestible forms, increasing the nutritional value of feed. This, in turn, causes a shift in the gut microbial population, alters the composition of the gut microbiota and modulates the immune system (Fig. 1). The digestion of feed is improved through production of hydrolytic enzymes (phytases, lipases, amylases or proteases) that increase nutrient uptake [57, 58], or by stimulating the host to increase the secretion of digestive enzymes [59]. Probiotics may also increase the nutritional value of feed by the production of vitamins, exopolysaccharides and antioxidants (Fig. 2). Some probiotics may also regulate the metabolism of cholesterol [60, 61]. Nutrients are digested and absorbed in the proventriculus and small intestine. Undigested carbohydrates and proteins are fermented in the ileum and colon [57, 62]. This is an important step in digestion, as undigested nutrients is a predisposing factor for dysbiosis [63]. Genera from the *Firmicutes* phylum (to which most probiotics belong), in addition to members of the *Actinobacteria* and *Verrucomicrobium* phyla, may aid in the digestion of amino acids [64].

Enzymatic reactions in the colon provides additional energy in the form of lactate, acetate, propionate and butyrate [65]. These short-chain fatty acids (SCFAs), produced by probiotic bacteria, are also converted to lipids and cholesterol [66]. Although SCFAs serve as additional energy, only a small proportion (up to 25%) is recovered [67].

Some strains of *Bifidobacterium* and *Lactobacillus* spp. synthesise vitamin K, biotin, cobalamin, folates, nicotinic acid, pantothenic acid, pyridoxine, riboflavin and thiamine [68]. These vitamins are essential micronutrients and play an important role in growth and reproduction. However, it is unclear if these bacterially produced vitamins are absorbed in the GIT [69].

Exopolysaccharides (EPS), as concluded with studies on other animals, not only protect cells against dehydration, phagocytosis, predation, bacteriophages, antibiotics and other toxic compounds [70], but facilitates adhesion of the cells to the GIT and supports biofilm formation [71]. Exopolysaccharides produced by probiotic bacteria modulate the innate and adaptive immune systems [72, 73], display antitumor properties [74], lower cholesterol levels [75], inhibit pathogens, promote the growth of beneficial bacteria and act as antioxidants [76].

Fig. 1 General modes of action by which probiotics improve broiler health and performance



Anti-oxidative enzymes, such as glutathione S-transferase, glutathione reductase, glutathione peroxidase, superoxide dismutase and catalase, produced by probiotics scavenge reactive oxygen species (ROS) or prevents the formation thereof [77]. Strains of *Lactobacillus acidophilus*, *Lactobacillus plantarum*, *Bifidobacterium longum* and *Pediococcus pentosaceus* have been shown to possess antioxidative activity [77]. This is important, as ROS, often produced during stress, has a pronounced negative effect on the health and performance of broilers [78]. The presence of ROS may also lead to the development of chronic diseases and lipid peroxidation that may result in hyperlipidaemia and hyperglycaemia [79]. This results in an increase in the level of pro-atherogenic lipoproteins and a decrease in HDL (high-density lipoproteins) cholesterol levels [80].

Some probiotic bacteria regulate cholesterol homeostasis by hydrolysing bile acids, which prevents reabsorption in the intestine, indirectly lowering cholesterol levels [81, 82]. Members of the genera *Lactobacillus*, *Bifidobacterium*, *Enterococcus*, *Clostridium* and *Bacteroides* frequently produce bile salt hydrolase [83]. The exact mechanism by which cholesterol is utilised is poorly understood. Probiotics administered to broilers decreased cholesterol levels in meat and blood [81, 82]. Probiotic bacteria may assimilate cholesterol during growth, bind cholesterol to their cell surfaces,

incorporate cholesterol into the cell membrane, deconjugate bile via bile salt hydrolase (BSH) or co-precipitate cholesterol together with deconjugated bile [see review 84].

Microbiome Modulation

Interactions between the microbiota and host are important for development, health, nutrition and digestion and food safety [85]. Detrimental changes in the microbiota composition (dysbiosis) disrupts mutualistic interactions and can lead to disease [86]. Probiotics restore microbiota homeostasis by inhibiting pathogens and by promoting the growth of beneficial bacteria [87]. Immune activation or chronic inflammation induced by pathogens contributes to decreased health and growth in poultry. Probiotics and their metabolites induce shifts in the microbiota composition, from a predominant pathogenic microbiota to more beneficial microbiota [88]. This is primarily done by competitively excluding pathogens from the mucosal surface [89], competing for nutrients [90] and production of exopolysaccharides and antimicrobial compounds such as SCFAs, H_2O_2 and antimicrobial peptides (Fig. 3). Probiotics such as lactic acid bacteria (LAB) can also produce SCFAs (lactate, acetate, isovalerate, butyrate and propionate) which penetrate the cell wall of susceptible bacteria

Fig. 2 Modes of action by which probiotics modulate broiler nutrition and digestion

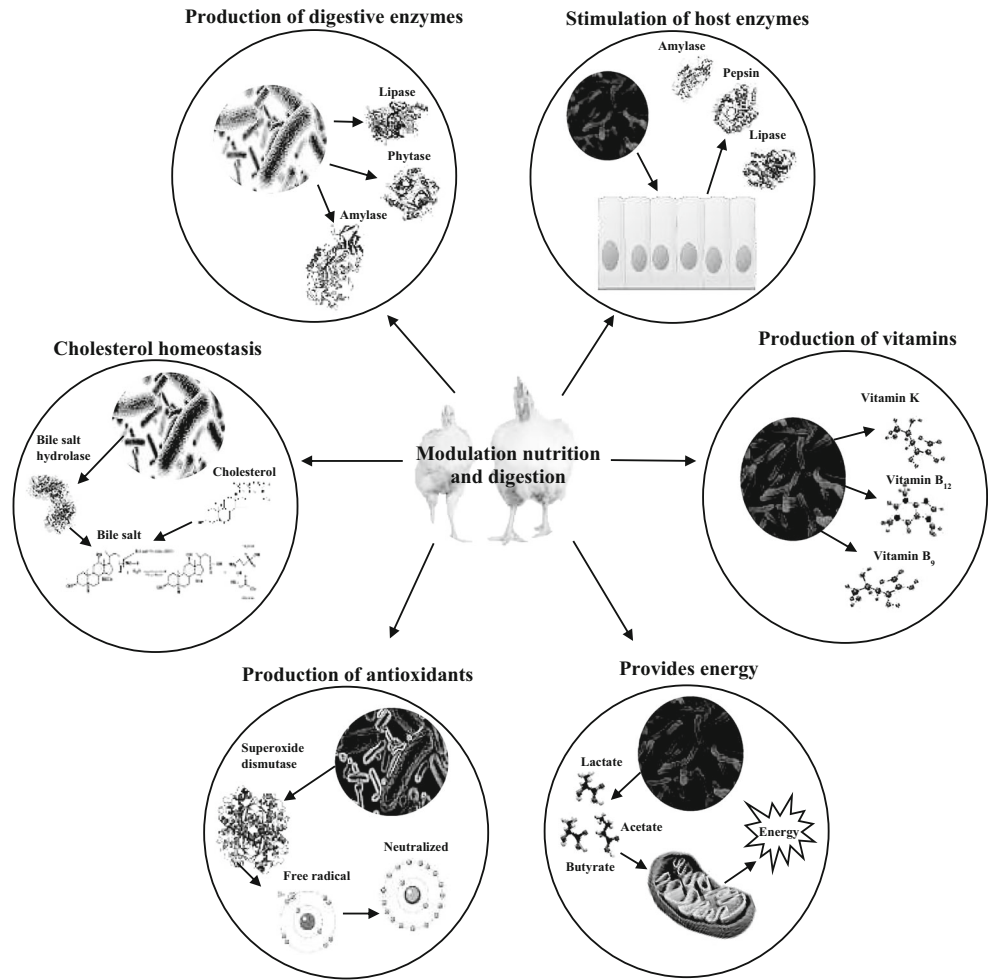
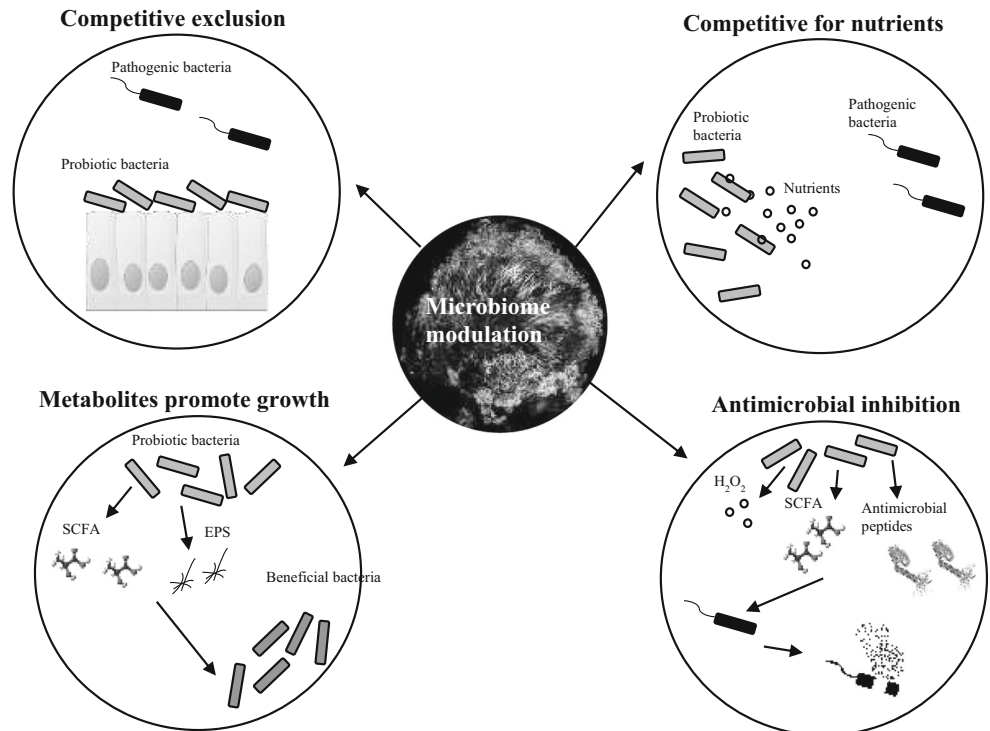


Fig. 3 Modes of action by which probiotics modulate the GIT microbiota composition



[91], decreasing the pH of the cytoplasm [92], leading to disruption of physiological reactions and the denaturation of proteins, enzymes, and nucleic acids [93], and subsequently cell death. Hydrogen peroxide produced by probiotic bacteria reacts with cellular components in the cytoplasm of susceptible prokaryotic cells, leading to apoptosis and destruction of the cell membrane [94].

Some probiotic bacteria produce antimicrobial peptides that may modulate the composition of the microbiota [95–100]. Bacteriocins are small, bacterially produced peptides that possess antimicrobial activity [101, 102]. The antimicrobial activity of bacteriocins has been well studied in vitro, but only a few reports of these peptides rendering protection against pathogens in the GIT have been published [103–105]. Some bacteriocins may also function as “colonizing peptides”, facilitating the introduction and dominance of the producing strain into an already occupied niche, as growth promoters regulating the microbiota composition [see review 106], or as signalling molecules that are used to communicate with other bacteria, or host cells [107]. In Gram-negative bacteria, (*N*-acyl) homoserine lactone typically serves as a signal molecule, while in Gram-positive bacteria, peptides, including some bacteriocins, are used as signalling molecules [see reviews 108, 109]. The production of EPS is equally important in modulating the gut microbiota, as shown with EPS-producing probiotic strains stimulating the growth of *Lactobacillus* and *Bifidobacterium* spp. [110, 111].

Although the microbiota of a broiler has been studied [112–114], much more research is needed to determine the role probiotics play in modulating the microbiota. First, we need to understand the role different members of the microbiota play in host physiology. This may be a challenge, since the gut microbiota of broilers is affected by a number of host and environmental factors [115]. Furthermore, little is known about how the avian microbiota develops, the factors responsible for early-life changes, and how these shifts in microbial populations relate to changes in metabolic functioning, health and growth [115]. It is, however, well known that probiotics assist in the establishment of a mature gut microbiota [116]. The potential role of probiotics in early-life includes the reduction of pathogen cell numbers, and an increase in feed conversion and growth [116]. Addressing these questions requires the developing of more advanced GIT investigation procedures and ultra-sensitive detection systems.

Modulation of Intestinal Barrier Integrity

With modulation of the gut microbiota, physiological changes in broilers are noted. These include changes in the integrity of the gut wall, the rate at which cells undergo apoptosis and, most probably, changes in the type of signals reaching the central nervous system via the enteric nerves (Fig. 4). All of

these physiological alterations are observed as changes in bone health and meat quality [117, 118].

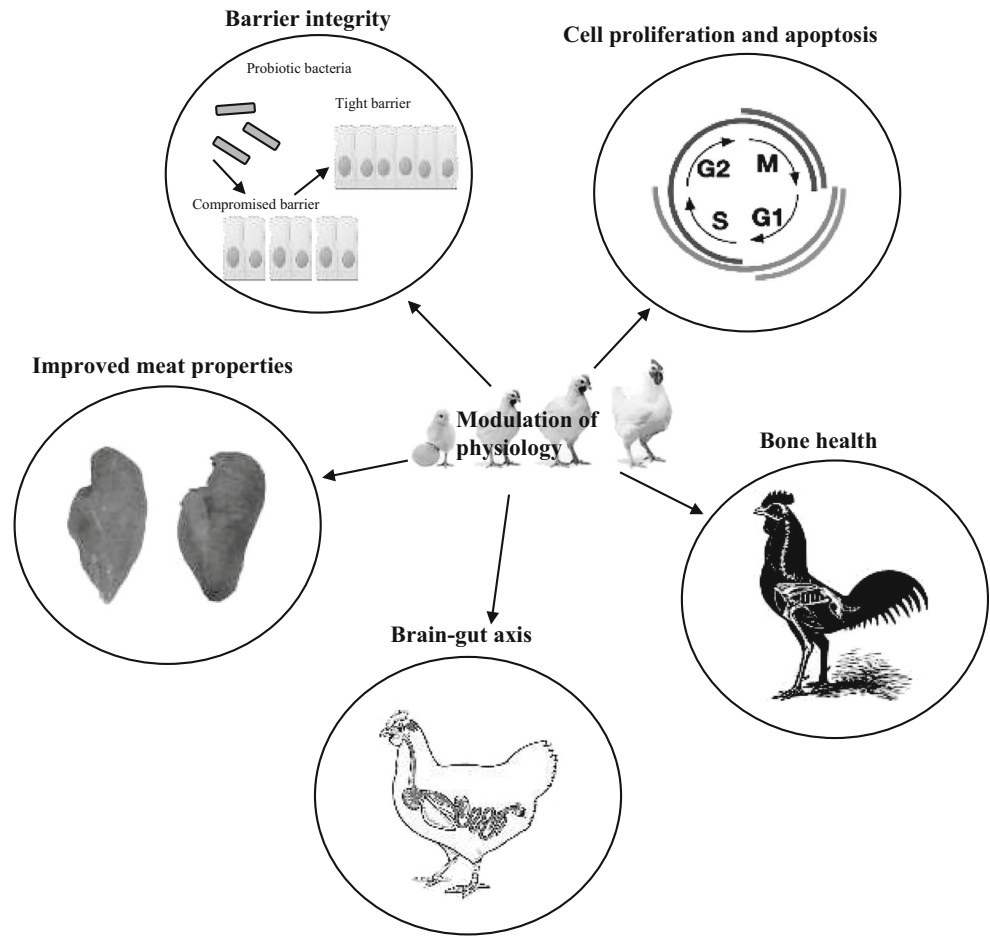
The epithelial monolayer forms a barrier between the luminal content and the interstitial tissue, which prevents diffusion of microbial cells and toxins from the lumen into the circulatory system and the tissue or organs [119]. Maintaining the integrity of the gut barrier is thus critical to prevent the translocation of pathogens into the intestinal lumen [120]. Probiotic bacteria regulate epithelial permeability by modulating tight junction (TJ) proteins (occludin, zonula occludens, claudins and junction adhesion molecules), which in turn inhibits pathogen colonisation, modulates cell proliferation and apoptosis, and controls mucin production [121–124]. Increased permeability leads to mucosal barrier dysfunction; however, probiotics can normalise TJ protein expression and localisation, restoring barrier integrity [125–127].

A balance exists between epithelial cell proliferation, differentiation and apoptosis [128]. Probiotics can modulate cell kinetics by regulating homeostasis of cell proliferation and apoptosis [88]. In addition, an increase in apoptosis over cell proliferation leads to increased susceptibility to pathogenic infections [129]. Numerous researchers have shown that probiotics regulate transcription of genes related to cell proliferation and apoptosis [88, 130, 131]. In the GIT of broilers, probiotic administration has been shown to regulate villus cell proliferation [132–134]. The structure and architecture of villi are important as their morphology regulates the capacity of nutrients absorption and defence responses against pathogens [132]. Probiotics regulate cytokine and oxidant-induced epithelial apoptosis by proteins p75 and p40 which activates anti-apoptotic Akt in a phosphatidylinositol-3'-kinase (PI3K)-dependent manner and inhibiting pro-apoptotic p38/MAPK activation [135, 136]. Reduced apoptosis improves barrier integrity and increases resistance to bacterial invasion [137]. In addition, numerous other studies have shown that probiotics decrease cell apoptosis by differentially expressing apoptosis inhibitor-related proteins, i.e. HIAP2/cIAP, TLR-2, COX2 and PGE2 proteins [138, 139].

Probiotic bacteria also contribute to intestinal barrier integrity by modulating mucin production [140–142]. Mucins are the major protein component coating the GIT. Probiotics normalise intestinal integrity through the restoration of the mucus layer by adjusting the mucin monosaccharide composition, mucus layer thickness and mucin gene expression [140–143]. The structural and functional properties of mucins influence bacterial adhesion to the mucosal surface. In broilers, probiotics modulate intestinal mucin monosaccharide compositions, subsequently influencing the GIT microbiota composition [140–142].

Until recently, the role probiotics play in bone health remained largely unknown [117, 118, 144]. Detrimental microbiota shifts in the GIT microbiota leads to dysbiosis which decreases bone density [145]. Probiotics are capable of

Fig. 4 Modes of action by which probiotics modulate broiler physiology



modulating bone mineralisation and development by impacting multiple aspects [147]. Probiotics modulate bone health by impacting nutrient acquisition important for bone growth (Ca^{2+} and P^{3-}), modulate barrier integrity and immune responses, and by the production of serotonin or oestrogen-like molecules [147, 148]. Probiotics indirectly modulate bone health by regulating microbiota health leading to increased intestinal absorption of minerals (Ca^{2+} and P^{3-}) important for bone health [149]. In addition, probiotics can also regulate bone health by neuroendocrine signalling pathways inducing intestinal cells to produce endocrine factors such as incretins, oestrogen-like molecules and serotonin which acts as signals for bone cells [147, 148]. In broilers, probiotics increase bone thicknesses and improve the mineral content and bone break-age strength [146].

Wang and co-workers [150] have shown that heat-stressed broilers fed a probiotic strain of *B. subtilis* coped better than broilers on a diet without the probiotic. Birds that received the probiotic spent less time in wing spreading, panting, squatting, drinking, sleeping, dozing and sitting. The probiotic-treated birds also had lower concentrations of hepatic IL-6, heat shock protein HSP70, cecal IgA and IgY.

Probiotics and Meat Quality

The role probiotics play in improving broiler carcass meat characteristics is highly debated. Some studies have reported improved meat attributes such as water-holding capacity, tenderness, lipid oxidation stability, sensory properties and microbial safety [151–155]. On the contrary, others have noted that there are no synergistic effects of probiotics on meat quality [156, 157]. Disagreements might be due to the differences in experimental conditions or probiotic compositions used. Probiotic administration in broilers improves the chemical, nutritional and sensorial characteristics of meat by increasing protein and free amino acid content, decreasing fat content and improving sensory properties [158, 159]. In broilers infected with pathogens, probiotic administration decreased pathogen-induced gut permeability [160].

Conclusions

Probiotics have a positive effect on feed digestion, regulation of GIT microbial communities, physiology and the immune system. However, in-depth research is required to understand

the molecular changes brought about by probiotics and the crosstalk between probiotics, pathogens and epithelial cells. This will have to include metagenomic, proteomic and metabolomics studies. Elucidating these unknowns will provide greater insight into the role probiotics play in improving broiler health and growth.

Compliance with Ethical Standards

Conflict of Interest The authors declare that they have no competing interest.

References

- Havenstein GB, Ferket PR, Qureshi MA (2003) Growth, livability, and feed conversion 1957 versus 2001 broilers when fed representative 1957 and 2001 broiler diets. *Poult Sci* 82(10):1500–1508
- Martin JL, Fyfe M, Doré K, Buxton JA, Pollari F, Henry B, Middleton D, Ahmed R, Jamieson F, Ciebin B, McEwen SA (2004) Increased burden of illness associated with antimicrobial resistant *Salmonella enterica* serotype Typhimurium infections. *J Infect Dis* 189(3):377–384
- Agyare C, Boamah VE, Zumbi CN, Osei FB (2018) Antimicrobial resistance - a global threat. Antibiotic use in poultry production and its effects on bacterial resistance. IntechOpen. <https://doi.org/10.5772/intechopen.79371>
- Ohimain EI, Ofongo RTS (2012) The effect of probiotic feed supplementation on chicken health and gut microflora: a review. *Int J Anim Vet Adv* 4:135–114
- Kabir SML (2009) The role of probiotics in the poultry industry. *Int J Mol Sci* 10(8):3531–3546
- Ahmad I (2006) Effect of probiotics on broilers performance. *Int J Poult Sci* 5(6):593–597
- Čermák L, Skřivanová E (2016) Influence of pasture rearing on the cecal bacterial microbiota in broiler chickens. *Sci Agric Bohem* 47:124–128
- Mehdi Y, Létourneau-Montminy M, Gaucher M, Chorfi Y, Suresh G, Rouissi T, Brar SK, Côté C, Ramirez AA, Godbouth S (2018) Use of antibiotics in broiler production: global impacts and alternatives. *Anim Nutr* 4(2):170–117
- Madigan MT, Martinko JM, Bender KS, Buckley FH, Stahl DA (2014) Brock biology of microorganisms, 14th edn. Illinois, Pearson International, p 1006
- Laxminarayan R, Duse A, Wattal C, Zaidi AKM, Wertheim HFL, Sumpradit N, Vlieghe E, Hara GL, Gould IM, Goossens H, Greko C, So AD, Bigdeli M, Tomson G, Woodhouse W, Ombaka E, Peralta AQ, Qamar FN, Mir F, Kariuki S, Bhutta ZA, Coates A, Bergstrom R, Wright GD, Brown ED, Cars O (2013) Antibiotic resistance -the need for global solutions. *Lancet Infect Dis* 13: 1057–1098
- Odoi H (2016) isolation and characterization of multi-drug resistant *Pseudomonas aeruginosa* from clinical, environmental and poultry litter sources in Ashanti region of Ghana (MPhil thesis). Kumasi: Kwame Nkrumah University of Science and Technology
- Aniokette U, Iroha CS, Ajah MI, Nwakaeze AE (2016) Occurrence of multi-drug resistant gram-negative bacteria from poultry and poultry products sold in Abakaliki. *J Agric Sci Food Technol* 2:119–124
- Sharma S, Galav V, Agrawal M, Faridi F, Kumar B (2017) Multi-drug resistance pattern of bacterial flora obtained from necropsy samples of poultry. *J Anim Health Prod* 5:165–171
- Adelowo OO, Fagade OE, Agersø Y (2014) Antibiotic resistance and resistance genes in *Escherichia coli* from poultry farms, Southwest Nigeria. *J Infect Dev Ctries* 8:1103–1112
- Medeiros MAN, de Oliveira DCN, Rodrigues DP, de Freitas DRC (2011) Prevalence and antimicrobial resistance of *Salmonella* in chicken carcasses at retail in 15 Brazilian cities. *Pan American J Public Health* 30:555–560
- Nomoto R, Tien LHT, Sekizaki T, Osawa R (2013) Antimicrobial susceptibility of *Streptococcus gallolyticus* isolated from humans and animals. *Jpn J Infect Dis* 66:334–336
- Rożynek E, Dzierżanowska-Fangrat K, Korsak D, Konieczny P, Wardak S, Szych J, Jarosz M, Dzierżanowska D (2008) Comparison of antimicrobial resistance of *Campylobacter jejuni* and *Campylobacter coli* isolated from humans and chicken carcasses in Poland. *J Food Prot* 71:602–607
- Nguyen TNM, Hotzel H, Njeru J, Mwituria J, El-Adawy H, Tomaso H, Neubauer H, Hafez HM (2016) Antimicrobial resistance of *Campylobacter* isolates from small scale and backyard chicken in Kenya. *Gut Pathog* 8:1–9
- Dallal MMS, Doyle MP, Rezadehbash M, Dabiri H, Sanaei M, Modarresi S, Bakhtiari R, Sharify K, Taremi M, Zali MR, Sharifi-Yazdi MK (2010) Prevalence and antimicrobial resistance profiles of *Salmonella* serotypes, *Campylobacter* and *Yersinia* spp. isolated from retail chicken and beef, Tehran, Iran. *Food Control* 21(4): 388–392
- Zadernowska A, Chaje W (2017) Prevalence, bio film formation and virulence markers of *Salmonella* sp. and *Yersinia enterocolitica* in food of animal origin in Poland. *LWT Food Sci Technol* 75:552–556
- Osman KM, Elhariri M (2013) Antibiotic resistance of *Clostridium perfringens* isolates from broiler chickens in Egypt. *Rev Sci Technol* 32(2):841–850
- Park JY, Kim S, Oh JY, Kim HR, Jang I, Lee HS, Kwon YK (2015) *Poult Sci* 94:1158–1164
- Fan YC, Wang CL, Wang C, Chen TC, Chou CH, Tsai HJ (2016) Incidence and antimicrobial susceptibility to *Clostridium perfringens* in premarket broilers in Taiwan. *Avian Dis* 60(2): 444–449
- Floriștean V, Cretu C, Carp-Cărare M (2007) Bacteriological characteristics of *Bacillus cereus* isolates from poultry. *Bulletin of University of Agricultural Sciences and Veterinary Medicine Cluj-Napoca* 64:1–2
- Reza M, Lijon M, Khatun M, Islam M (2015) Prevalence and antibiogram profile of *Mycobacterium* spp. in poultry and its environments. *J Adv Vet Anim Res* 2(4):458
- Kolář M, Pantůček R, Bardoň J, Vágnerová I, Typovská H, Válka I, Doškař J (2002) Occurrence of antibiotic-resistant bacterial strains isolated in poultry. *Vet Med (Praha)* 47(2–3):52–59
- Vignaroli C, Zandri G, Aquilanti L, Pasquaroli S, Biavasco F (2011) Multidrug-resistant enterococci in animal meat and faeces and co-transfer of resistance from an *Enterococcus durans* to a human *Enterococcus faecium*. *Curr Microbiol* 62(5):1438–1447
- Nahar A, Siddiquee M, Nahar S, Anwar KS, Ali SI, Islam S (2014) Multidrug resistant *Proteus mirabilis* isolated from chicken droppings in commercial poultry farms. Bio-security concern and emerging public health threat in Bangladesh. *J Biosafety Health Educ* 2(2):120–125
- Salyers AA, Amabile-Cuevas CF (1997) Why are antibiotic resistance genes so resistant to elimination? *Antimicrob Agents Chemother* 41(11):2321–2325
- Cogliani C, Goossens H, Greko C (2011) Restricting antimicrobial use in food animals: lessons from Europe. *Microbe* 6:274–279

31. Maron DF, Smith TJS, Nachman KE (2013) Restrictions on antimicrobial use in food animal production: an international regulatory and economic survey. *Glob Health* 9:48
32. Van Boeckel TP, Brower C, Gilbert M, Grenfell BT, Levin SA, Robinson TP, Teillant A, Laxminarayan R (2015) Global trends in antimicrobial use in food animals. *Proc Natl Acad Sci* 112:5649–5654
33. Van Boeckel TP, Gandra S, Ashok A, Caudron Q, Grenfell BT, Levin SA, Laxminarayan R (2014) Global antibiotic consumption 2000 to 2010: an analysis of national pharmaceutical sales data. *Lancet Infect Dis* 14:742–750
34. Adhikari PA, Kim WK (2017) Overview of prebiotics and probiotics: focus on performance, gut health and immunity- a review. *Ann Anim Sci* 17:949–966
35. Wang H, Ni X, Qing X, Liu L, Lai J, Khalique A, Li G, Pan K, Jing B, Zeng D (2017) Probiotic enhanced intestinal immunity in broilers against subclinical necrotic enteritis. *Front Immunol* 8:1592
36. Dunislawaska A, Slawinska A, Stadnicka K, Bednarczyk M, Gulewicz P, Jozefiak D, Siwek M (2017) Synbiotics for broiler chickens- *In vitro* design and evaluation of the influence on host and selected microbiota populations following *in ovo* delivery. *PLoS One* 12:e0168587
37. Pandey KR, Naik SR, Vakil BV (2015) Probiotics, prebiotics and synbiotics- a review. *J Food Sci Technol* 52:7577–7587
38. Gheisar MM, Kim IH (2018) Phytobiotics in poultry and swine nutrition - a review. *Ital J Anim Sci* 17:92–99
39. Kiarie E, Romero LF, Nyachoti CM (2013) The role of added feed enzymes in promoting gut health in swine and poultry. *Nutr Res Rev* 26:71–88
40. Yang Y, Iji PA, Choct M (2009) Dietary modulation of gut microflora in broiler chickens: a review of the role of six kinds of alternatives to in-feed antibiotics. *Worlds Poult Sci J* 65:97–114
41. Alciçek A, Bozkurt M, Cabuk M (2004) The effect of a mixture of herbal essential oils, an organic acid or a probiotic on broiler performance. *SA J Anim Sci* 34:217–222
42. Brenes A, Roura E (2010) Essential oils in poultry nutrition: main effects and modes of action. *Anim Feed Sci Technol* 158:1–14
43. Huff GR, Famell MB, Huff WE, Rath NC, Solis De Los Santos F, Donoghue AM (2010) Bacterial clearance, heterophil function, and hematological parameters of transport-stressed Turkey poults supplemented with dietary yeast extract. *Poult Sci* 89:447–456
44. Ghareeb K, Awad WA, Mohnl M, Porta R, Biarnés M, Böhm J, Schatzmayr G (2012) Evaluating the efficacy of an avian-specific probiotic to reduce the colonisation of *Campylobacter jejuni* in broiler chickens. *Poult Sci* 91:1825–1832
45. Sterzo EV, Paiva JB, Mesquita AL, Freitas Neto OC, Berchieri S (2007) Organic acids and/or compound with defined microorganisms to control *Salmonella enterica* serovar Enteritidis experimental infection in chickens. *Rev Bras Cienc Avic* 9:69–73
46. Teo AY, Tan H-M (2007) Evaluation of the performance and intestinal gut microflora of broiler fed on corn-soy diets supplemented with *Bacillus subtilis* PB6 (CloSTAT™). *J Appl Poult Res* 16:296–303
47. Melegy T, Khaled NF, El-Bana R, Abdellatif H (2011) Effect of dietary supplementation of *Bacillus subtilis* PB6 (CLOSTAT™) on performance, immunity, gut health and carcass traits in broilers. *J Am Sci* 7:891–898
48. Lourenco MC, Kuritz LN, Westphal P, Muniz E, Pickler L, Santin E (2012) Effects of *Bacillus subtilis* in the dynamics of infiltration of immunological cells in the intestinal mucosa of chickens challenged with *Salmonella* Minnesota. *Int J Poult Sci* 11:630–634
49. Abudabos AM, Alyemni AH, Al Marshad MBA (2013) *Bacillus subtilis* PB6 based probiotic (CloSTAT™) improves intestinal morphological and microbiological status of broiler chickens under *Clostridium perfringens* challenge. *Int J Agric Biol* 15:978–982
50. Gutierrez-Fuentes CG, Zuñiga-Orozco LA, Vicente JL, Hernandez-Velasco X, Menconi A, Kuttappan VA, Kallapura G, Latorre J, Layton S, Hargis BM, Téllez G (2013) Effect of a lactic acid bacteria based probiotic, Floramax-B1®, on performance, bone qualities, and morphometric analysis of broiler chickens: an economic analysis. *Biol Syst* 2:113
51. Prado-Rebolledo OF, de Jesus D-MJ, Macedo-Barragan RJ, Garcia-Márquez LJ, Morales-Barrera JE, Latorre JD, Hernandez-Velasco X, Tellez G (2017) Evaluation of a selected lactic acid bacteria-based probiotic on *Salmonella enterica* serovar Enteritidis colonisation and intestinal permeability in broiler chickens. *Avian Pathol* 46:90–94
52. Neveling DP, Van Emmenes L, Ahire JJ, Pieterse E, Smith C, Dicks LMT (2019) Effect of a multi-species probiotic on the colonisation of *Salmonella* in broilers. *Prob Antimicrob Prot*:1–10 published online. <https://doi.org/10.1007/s12602-019-09593-y>
53. Bouzaine T, Dauphin RD, Thonart PH, Urdaci MC, Hamdi M (2005) Adherence and colonization properties of *Lactobacillus rhamnosus* TB1, a broiler chicken isolate. *Lett Appl Microbiol* 40(5):391–396
54. Rocha TS, Baptista AAS, Donato TC, Milbradt EL, Okamoto AS, Rodrigues JCZ, Coppola MP, Andreatti Filho RL (2012) Evaluation of *in vitro* and *in vivo* adhesion and immunomodulatory effect of *Lactobacillus* species strains isolated from chickens. *Poult Sci* 91(2):362–369
55. Sherman PM, Ossa JC, Johnson-Henry K (2009) Unraveling mechanisms of action of probiotics. *Nutr Clin Pract* 24(1):10–14
56. Rehman HU, Vahjen W, Awad WA, Zentek J (2007) Indigenous bacteria and bacterial metabolic products in the gastrointestinal tract of broiler chickens. *Arch Anim Nutr* 61:319–335
57. Flint HJ, Scott KP, Duncan SH, Louis P, Forano E (2012) Microbial degradation of complex carbohydrates in the gut. *Gut Microbes* 3(4):289–306
58. Wang Y, Gu Q (2010) Effect of probiotic on growth performance and digestive enzyme activity of Arbor Acres broilers. *Res Vet Sci* 89(2):163–167
59. Hmani H, Daoud L, Jlidi M, Jalleli K, Ali MB, Brahim AH, Bargui M, Dammak A, Ali MB (2017) A *Bacillus subtilis* strain as probiotic in poultry: selection based on *in vitro* functional properties and enzymatic potentialities. *J Ind Microbiol Biotechnol* 44(8):1157–1166
60. Wealleans AL, Walsh MC, Romero LF, Ravindran V (2017) Comparative effects of two multi-enzyme combinations and a *Bacillus* probiotic on growth performance, digestibility of energy and nutrients, disappearance of non-starch polysaccharides, and gut microflora in broiler chickens. *Poult Sci* 93(12):4287–4297
61. Zhang ZF, Kim IH (2014) Effects of multistrain probiotics on growth performance, apparent ileal nutrient digestibility, blood characteristics, cecal microbial shedding, and excreta odor contents in broilers. *Poult Sci* 93(2):364–370
62. LeBlanc JG, Chain F, Martín R, Bermúdez-Humarán LG, Courau S, Langella P (2017) Beneficial effects on host energy metabolism of short-chain fatty acids and vitamins produced by commensal and probiotic bacteria. *Microb Cell Factories* 16:79
63. Carding S, Verbeke K, Vipond DT, Corfe BM, Owen LJ (2015) Dysbiosis of the gut microbiota in disease. *Microb Ecol Health Dis* 26:26191
64. Sonnenburg JL, Xu J, Leip DD, Chen CH, Westover BP, Weatherford J, Buhler JD, Gordon JI (2005) Glycan foraging *in vivo* by an intestine-adapted bacterial symbiont. *Science* 307(5717):1955–1959
65. den Besten G, van Eunen K, Groen AK, Venema K, Reijngoud D-J, Bakker BM (2013) The role of short-chain fatty acids in the

- interplay between diet, gut microbiota, and host energy metabolism. *J Lipid Res* 54(9):2325–2340
66. Fushimi T, Suruga K, Oshima Y, Fukiharuru M, Tsukamoto Y, Goda T (2006) Dietary acetic acid reduces serum cholesterol and triacylglycerols in rats fed a cholesterol-rich diet. *Br J Nutr* 95(5): 916–924
 67. Bolton W, Dewar WA (1965) The digestibility of acetic, propionic and butyric acids by the fowl. *Br Poult Sci* 6(2):103–105
 68. LeBlanc JG, Milani C, de Giori GS, Sesma F, van Sinderen D, Ventura M (2013) Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr Opin Biotechnol* 24(2):160–168
 69. Eck P, Friel J (2013) Should probiotics be considered as vitamin supplements? *Vitam Miner* 2:e124
 70. Roberts IS (1996) The biochemistry and genetics of capsular polysaccharide production in bacteria. *Annu Rev Microbiol* 50:285–315
 71. Ruas-Madiedo P, Gueimonde M, Margolles A, de los Reyes-Gavilán CG, Salminen S (2006) Exopolysaccharides produced by probiotic strains modify the adhesion of probiotics and enteropathogens to human intestinal mucus. *J Food Prot* 69(8): 2011–2015
 72. Quinteiro-Filho WM, Brisbin JT, Hodgins DC, Sharif S (2015) *Lactobacillus* and *Lactobacillus* cell-free culture supernatants modulate chicken macrophage activities. *Res Vet Sci* 103:170–1753
 73. Apata DF (2008) Growth performance, nutrient digestibility and immune response of broiler chicks fed diets supplemented with a culture of *Lactobacillus bulgaricus*. *J Sci Food Agric* 88:1253–1258
 74. Nishimura J (2014) Exopolysaccharides produced from *Lactobacillus delbrueckii* subsp. *bulgaricus*. *Adv Microbiol* 4(14):1017–1020
 75. Maeda H, Zhu X, Suzuki S, Suzuki K, Kitamura S (2004) Structural characterization and biological activities of an exopolysaccharide kefiran produced by *Lactobacillus kefiranofaciens* WT-2B(T). *J Agric Food Chem* 52(17):5533–5538
 76. Fernandez F, Hinton M, Van Gils B (2002) Dietary mannan-oligosaccharides and their effect on chicken caecal microflora in relation to *Salmonella* Enteritidis colonisation. *Avian Pathol* 31(1):49–58
 77. Kumar M, Kumar A, Nagpal R, Mohania D, Behare P, Verma V, Kumar P, Poddar D, Aggarwal PK, Henry CJ, Jain S, Yadav H (2010) Cancer-preventing attributes of probiotics: an update. *Int J Food Sci Nutr* 61(5):473–496
 78. Mishra B, Jha R (2019) Oxidative stress in the poultry gut: potential challenges and interventions. *Front Vet Sci* 6:60
 79. Tangvarasittichai S (2015) Oxidative stress, insulin resistance, dyslipidemia and type 2 diabetes mellitus. *World J Diabetes* 6(3):456–480
 80. Stancu CS, Toma L, Sima AV (2012) Dual role of lipoproteins in endothelial cell dysfunction in atherosclerosis. *Cell Tissue Res* 349(2):433–446
 81. Shokryazdan P, Jahromi MF, Liang JB, Ramasamy K, Sieo CC, Ho YW (2017) Effects of a *Lactobacillus salivarius* mixture on performance, intestinal health and serum lipids of broiler chickens. *PLoS ONE* 12(5):e0175959
 82. Yalçın S, Eser H, Yalçın S, Yalçın SS (2016) Effects of probiotics on some blood parameters and serum IgG in broilers. *J Pediatr Gastr Nutr* 63(1S):S57
 83. Begley M, Hill C, Gahan CGM (2006) Bile salt hydrolase activity in probiotics. *Appl Environ Microbiol* 72(3):1729–1738
 84. Ooi L-G, Liang M-T (2010) Cholesterol-lowering effects of probiotics and prebiotics: a review of in vivo and in vitro findings. *Int J Sci* 11(6):2499–2522
 85. Oakley BB, Morales CA, Line J, Berrang ME, Meinersmann RJ, Tillman GE, Wise MG, Siragusa GR, Hiatt KL, Seal BS (2013) The poultry-associated microbiota: network analysis and farm-to-fork characterizations. *PLoS ONE* 8:e57190
 86. Frank D, Zhu W, Sartor RB, Li E (2011) Investigating the biological and clinical significance of human dysbioses. *Trends Microbiol* 19(9):427–434
 87. Hemarajata P, Versalovic J (2013) Effects of probiotics on gut microbiota: mechanisms of intestinal immunomodulation and neuromodulation. *Ther Adv Gastroenterol* 6(1):39–51
 88. Howarth GS, Wang H (2013) Role of endogenous microbiota, probiotics and their biological products in human health. *Nutrients* 5(1):58–81
 89. Patterson JA, Burkholder KM (2003) Application of prebiotics and probiotics in poultry production. *Poult Sci* 82(4):627–631
 90. O'Toole P, Cooney J (2008) Probiotic bacteria influence the composition and function of the intestinal microbiota. *Interdiscip Perspect Infect Dis* 17525
 91. Ríos-Covián D, Ruas-Madiedo P, Margolles A, Gueimonde M, de los Reyes-Gavilán CG (Salazar N, 2016) Intestinal short chain fatty acids and their link with diet and human health. *Front Microbiol*:185
 92. Mani-López E, García HS, López-Malo A (2012) Organic acids as antimicrobials to control *Salmonella* in meat and poultry products. *Food Res Int* 45(2):713–721
 93. Alakomi H-L, Puupponen-Pimiä R, Aura A-M, Helander IM, Nohynek L, Oksman-Caldentey K-M, Saarela M (2007) Weakening of *Salmonella* with selected microbial metabolites of berry-derived phenolic compounds and organic acids. *J Agric Food Chem* 55(10):3905–3912
 94. Linley E, Denyer SP, McDonnell G, Simons C, Maillard JY (2012) Use of hydrogen peroxide as a biocide: a new consideration of its mechanisms of biocidal action. *J Antimicrob Chemother* 67(7):1589–1596
 95. Daneshmand A, Kermanshahi H, Sekhavati MH, Javadmanesh A, Ahmadian M (2019) Antimicrobial peptide, cLF36, affects performance and intestinal morphology, microflora, junctional proteins, and immune cells in broilers challenged with *E. coli*. *Sci Rep* 9: 14176
 96. Hu F, Gao X, She R, Chen J, Mao J, Xiao P, Shi R (2017) Effects of antimicrobial peptides on growth performance and small intestinal function in broilers under chronic heat stress. *Poult Sci* 96(4): 798–806
 97. Eijsink VGH, Axelsson L, Diep DB, Havarstein LS, Holo H, Nes IF (2002) Production of class II bacteriocins by lactic acid bacteria; an example of biological warfare and communication. *Antonie Van Leeuwenhoek* 81(1–4):639–654
 98. O'Shea EF, Cotter PD, Stanton C, Ross RP, Hill C (2011) Production of bioactive substances by intestinal bacteria as a basis for explaining probiotic mechanisms: bacteriocins and conjugated linoleic acid. *Int J Food Microbiol* 152(3):189–205
 99. Spinler JK, Taweechotipatr M, Rognerud CL, Ou CN, Tumwasorn S, Versalovic J (2008) Human derived probiotic *Lactobacillus reuteri* demonstrate antimicrobial activities targeting diverse enteric bacterial pathogens. *Anaerobe* 14(3): 166–171
 100. Zheng J, Gänzle MG, Lin XB, Ruan L, Sun M (2015) Diversity and dynamics of bacteriocins from human microbiota. *Environ Microbiol* 17(6):2133–2143
 101. Cotter PD, Hill C, Ross RP (2005) Bacteriocins: developing innate immunity for food. *Nat Rev Microbiol* 3(10):777–788
 102. Klaenhammer TR (1993) Genetics of bacteriocins produced by lactic acid bacteria. *FEMS Microbiol Rev* 12(1–3):39–85
 103. Kierończyk B, Pruszyńska-Oszmałek E, Świątkiewicz S, Rawski M, Długosz J, Engberg EM, Józefiak D (2016) The nisin improves broiler chicken growth performance and interacts with

- salinomycin in terms of gastrointestinal tract microbiota composition. *J Anim Feed Sci* 25:309–316
104. Ustundag AO, Ozdogan M (2019) Effects of bacteriocin and organic acid on growth performance, small intestine histomorphology, and microbiology in Japanese quails (*Coturnix coturnix japonica*). *Trop Anim Health Prod* 51:2187
 105. Proctor A, Phillips GJ (2019) Differential effects of bacitracin methylene disalicylate (BMD) on the distal colon and cecal microbiota of young broiler chickens. *Front Vet Sci* 6:144
 106. Riley MA, Wertz JE (2002) Bacteriocin diversity: ecological and evolutionary perspectives. *Biochimie* 84(5–6):357–364
 107. Gobetti M, De Angelis M, Di Cagno R, Minervini F, Limitone A (2007) Cell-cell communication in food related bacteria. *Int J Food Microbiol* 120(1–2):34–45
 108. Fuqua C, Greenberg EP (2002) Listening in on bacteria: acyl-homoserine lactone signalling. *Nat Rev Mol Cell Biol* 3:685–695
 109. Hegarty JW, Guinane CM, Ross RP, Hill C, Cotter PD (2016) Bacteriocin production: a relatively unharmed probiotic trait? *F1000Res* 5:2587
 110. Pourabedin M, Chen Q, Yang M, Zhao X (2017) Mannan- and xylooligosaccharides modulate caecal microbiota and expression of inflammatory-related cytokines and reduce caecal *Salmonella* Enteritidis colonisation in young chickens. *FEMS Microbiol Ecol* 93(1):fw226
 111. Pourabedin M, Guan L, Zhao X (2015) Xylo-oligosaccharides and virginiamycin differentially modulate gut microbial composition in chickens. *Microbiome* 3:15
 112. Borda-Molina D, Seifert J, Camarinha-Silva A (2016) Current perspectives of the chicken gastrointestinal tract and its microbiome. *Comput Struct Biotechnol J* 16:131–139
 113. Johnson TJ, Youmans BP, Noll S, Cardona C, Evans NP, Kamezos TP, Ngunjiri JM, Abundo MC, Lee C (2018) A consistent and predictable commercial broiler chicken bacterial microbiota in antibiotic-free production displays strong correlations with performance. *Appl Environ Microbiol* 84(12):e00362–e00318
 114. Jurburg SD, Brouwer MSM, Ceccarelli D, van der Goot J, Jansman AJM, Bossers A (2019) Patterns of community assembly in the developing chicken microbiome reveal rapid primary succession. *Microbiology Open* 8:e821
 115. Sanders ME (2016) Probiotics and microbiota composition. *BMC Med* 14(1):82
 116. Patterson JA (2011) The commensal microbiota. In: Callaway TR, Ricke SC (eds) *Direct-fed microbials and prebiotics for animals*. Springer, New York, pp 3–11
 117. Yana FF, Wang WC, Cheng HW (2018) *Bacillus subtilis* based probiotic improved bone mass and altered brain serotonergic and dopaminergic systems in broiler chickens. *J Funct Foods* 49:501–509
 118. Yan F, Murugesan G, Cheng H (2019) Effects of probiotic supplementation on performance traits, bone mineralization, cecal microbial composition, cytokines and corticosterone in laying hens. *Animal* 13(1):33–41
 119. Anderson JM, Van Itallie CM (1995) Tight junctions and the molecular basis for regulation of paracellular permeability. *Am J Phys* 269(4 Pt 1):G467–G475
 120. Rao RK, Samak G (2013) Protection and restitution of gut barrier by probiotics: nutritional and clinical implications. *Curr Nutr Food Sci* 9(2):99–107
 121. Corridoni D, Pastorelli L, Mattioli B, Locovei S, Ishikawa D, Arseneau KO, Chieppa M, Cominelli F, Pizarro TT (2012) Probiotic bacteria regulate intestinal epithelial permeability in experimental ileitis by a TNF-dependent mechanism. *PLoS ONE* 7:e42067
 122. Harhaj NS, Antonetti DA (2004) Regulation of tight junctions and loss of barrier function in pathophysiology. *Int J Biochem Cell Biol* 36(7):1206–1237
 123. Howarth GS (2010) Probiotic-derived factors: probiotaceuticals? *J Nutr* 140(2):229–230
 124. Yan F, Polk DB (2011) Probiotics and immune health. *Curr Opin Gastroenterol* 27(6):496–501
 125. Hernandez-Patlan D, Solis-Cruz B, Pontin KP, Hernandez-Velasco X, Merino-Guzman R, Adhikari B, López-Arellano R, Kwon YM, Hargis BM, Arreguin-Nava MA, Tellez-Isaias G, Latorre JD (2019) Impact of a *Bacillus* direct-fed microbial on growth performance, intestinal barrier integrity, necrotic enteritis lesions, and ileal microbiota in broiler chickens using a laboratory challenge model. *Front Vet Sci* 6:108
 126. Anderson RC, Cookson AL, McNabb WC, Park Z, McCann MJ, Kelly WJ, Roy NC (2010) *Lactobacillus plantarum* MB452 enhances the function of the intestinal barrier by increasing the expression levels of genes involved in tight junction formation. *BMC Microbiol* 10:316
 127. Wang L, Li L, Lv Y, Chen Q, Feng J, Zhao X (2018) *Lactobacillus plantarum* restores intestinal permeability disrupted by *Salmonella* infection in newly-hatched chicks. *Sci Rep* 8:2229
 128. Llewellyn A, Foey A (2017) Probiotic modulation of innate cell pathogen sensing and signalling events. *Nutrients* 9(10):E1156
 129. Prisciandaro LD, Geier MS, Butler RN, Cummins AG, Howarth GS (2011) Evidence supporting the use of probiotics for the prevention and treatment of chemotherapy-induced intestinal mucositis. *Crit Rev Food Sci Nutr* 51(3):239–247
 130. Zhang L, Li N, Caicedo R, Neu J (2005) Alive and dead *Lactobacillus rhamnosus* GG decrease tumor necrosis factor- α -induced interleukin-8 production in Caco-2 cells. *J Nutr* 135(7):1752–1756
 131. Ko JS, Yang HR, Chang JY, Seo JK (2007) *Lactobacillus plantarum* inhibits epithelial barrier dysfunction and interleukin-8 secretion induced by tumor necrosis factor- α . *World J Gastroenterol* 13(13):1962–1965
 132. Sun Y, Rajput IR, Arain MA, Li Y, Baloch DM (2016) Oral administration of *Saccharomyces boulardii* alters duodenal morphology, enzymatic activity and cytokine production response in broiler chickens. *Anim Sci J* 88(8):1204–1211
 133. Wang H, Ni X, Qing X, Zeng D, Luo M, Liu L, Li G, Pan K, Jing B (2017) Live probiotic *Lactobacillus johnsonii* BS15 promotes growth performance and lowers fat deposition by improving lipid metabolism, intestinal development, and gut microflora in broilers. *Front Microbiol* 8:1073
 134. Beski SSM, Al-Sardary SYT (2015) Effects of dietary supplementation of probiotic and synbiotic on broiler chickens hematology and intestinal integrity. *Int J Poult Sci* 14(1):31–36
 135. Yan F, Polk DB (2002) Probiotic bacterium prevents cytokine-induced apoptosis in intestinal epithelial cells. *J Biol Chem* 277(52):50959–50965
 136. Yan F, Cao H, Cover TL, Whitehead R, Washington MK, Polk DB (2007) Soluble proteins produced by probiotic bacteria regulate intestinal epithelial cell survival and growth. *Gastroenterology* 132(2):562–575
 137. Hausmann M (2010) How bacteria-induced apoptosis of intestinal epithelial cells contributes to mucosal inflammation. *Int J Inflamm*:574568
 138. Khailova L, Mount Patrick SK, Arganbright KM, Halpern MD, Kinouchi T, Dvorak B (2010) Bifidobacterium bifidum reduces apoptosis in the intestinal epithelium in necrotizing enterocolitis. *Am J Physiol Gastrointest Liver Physiol* 299(5):G1118–G1127
 139. Dykstra NS, Hyde L, Adawi D, Kulik D, Ahme S, Molin G, Jeppsson B, Mackenzie A, Mack DR (2011) Pulse probiotic

- administration induces repeated small intestinal *MUC3* expression in rats. *Pediatr Res* 69(3):206–211
140. Aliakbarpour HR, Chamani M, Rahimi G, Sadeghi AA, Qujeq D (2012) The *Bacillus subtilis* and lactic acid bacteria probiotics influences intestinal mucin gene expression, histomorphology and growth performance in broilers. *Asian Australas J Anim Sci* 25(9):1285–1293
 141. Majidi-Mosleh A, Sadeghi AA, Mousavi SN, Chamani M, Zarei A (2017) Effects of *in ovo* infusion of probiotic strains on performance parameters, jejunal bacterial population and mucin gene expression in broiler chicken. *Braz J Poultry Sci* 19:97–102
 142. Tsirtsikos P, Fegeros K, Balaskas C, Kominakis A, Mountzouris KC (2012) Dietary probiotic inclusion level modulates intestinal mucin composition and mucosal morphology in broilers. *Poult Sci* 91(8):1860–1868
 143. O’Callaghan J, Butto LF, Macsharry J, Nally K, O’Toole PW (2012) Influence of adhesion and bacteriocin production by *Lactobacillus salivarius* on the intestinal epithelial cell transcriptional response. *Appl Environ Microbiol* 78(15):5196–5203
 144. Quach D, Britton RA (2017) Gut microbiota and bone health. *Adv Exp Med Biol* 1033:47–58
 145. Sylvester FA (2017) Inflammatory bowel disease: effects on bone and mechanisms. *Adv Exp Med Biol* 1033:133–150
 146. Mutuş R, Kocabağlı N, Alp M, Acar N, Eren M, Gezen ŞŞ (2006) The effect of dietary probiotic supplementation on tibial bone characteristics and strength in broilers. *Poult Sci* 85(9):1621–1625
 147. Lavoie B, Lian JB, Mawe GM (2017) Regulation of bone metabolism by serotonin. *Adv Exp Med Biol* 1033:35–46
 148. Ramsey W, Isaacs CM (2017) Intestinal incretins and the regulation of bone physiology. *Adv Exp Med Biol* 1033:13–33
 149. Christakos S, Veldurthy V, Patel N, Wei R (2017) Intestinal regulation of calcium: vitamin D and bone physiology. *Adv Exp Med Biol* 1033:3–12
 150. Wang WC, Yan FF, Hu JY, Amen OA, Cheng HW (2018) Supplementation of *Bacillus subtilis*-based probiotic reduces heat stress-related behaviors and inflammatory response in broiler chickens. *J Anim Sci* 96(5):1654–1666
 151. Bai K, Huang Q, Zhang J, He J, Zhang L, Wang T (2017) Supplemental effects of probiotic *Bacillus subtilis* fmbJ on growth performance, antioxidant capacity, and meat quality of broiler chickens. *Poult Sci* 96(1):74–82
 152. Khan AZ, Kumbhar S, Liu Y, Hamid M, Pan C, Nido SA, Parveen F, Huang K (2018) Dietary supplementation of selenium-enriched probiotics enhances meat quality of broiler chickens (*Gallus gallus* domesticus) raised under high ambient temperature. *Biol Trace Elem Res* 182:328
 153. Ebeid TA, Fathi MM, Al-Homidan I, Ibrahim ZH, Al-Sagan AA (2019) Effect of dietary probiotics and stocking density on carcass traits, meat quality, microbial populations and ileal histomorphology in broilers under hot-climate conditions. *Anim Prod Sci* 59:1711–1719
 154. Aksu Mİ, Karaolu M, Esenbuğa N, Kaya M, Macit M, Ockerman HW (2005) Effect of a dietary probiotic on some quality characteristics of raw broiler drumsticks and breast meat. *J Muscle Foods* 16(4):306–317
 155. Cramer TA, Kim HW, Chao Y, Wang W, Cheng HW, Kim YHB (2018) Effects of probiotic (*Bacillus subtilis*) supplementation on meat quality characteristics of breast muscle from broilers exposed to chronic heat stress. *Poult Sci* 97(9):3358–3368
 156. Yang X, Zhang B, Guo Y, Jiao P, Long F (2010) Effects of dietary lipids and *Clostridium butyricum* on fat deposition and meat quality of broiler chickens. *Poult Sci* 89(2):254–260
 157. Kim HW, Yan FF, Hu JY, Cheng HW, Kim YHB (2016) Effects of probiotics feeding on meat quality of chicken breast during postmortem storage. *Poult Sci* 95(6):1457–1464
 158. Zhang ZF, Zhou TX, Ao X, Kim IH (2012) Effects of β -glucan and *Bacillus subtilis* on growth performance, blood profiles, relative organ weight and meat quality in broilers fed maize-soybean meal based diets. *Livest Sci* 150(1–3):419–424
 159. Abdulla NR, Zamri ANM, Sabow AB, Kareem KY, Nurhazirah S, Ling FH, Sazili AQ, Loh TC (2017) Physico-chemical properties of breast muscle in broiler chickens fed probiotics, antibiotics or antibiotic-probiotic mix. *J Appl Anim Res* 45(1):64–70
 160. Liu X, Yan H, Xu Q, Yin C, Zhang K, Wang P, Hu J (2012) Growth performance and meat quality of broiler chickens supplemented with *Bacillus licheniformis* in drinking water. *Asian Australas J Anim Sci* 25(5):682–689

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