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Diversity and Distribution Patterns of Biofouling Macrobenthos Based on the Ship Navigation Type

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Abstract

Hull fouling by marine organisms is a known major pathway for introducing non-indigenous species, but information on this process is very limited. This study aimed to investigate variations in species diversity and distribution patterns based on navigation type (domestic–international and international) and attachment area. This study investigated the attached macroinvertebrates on six ships in 2021 and 2022. Quadrat sampling $(15 \times 15 \text{ cm})$ was conducted by scuba divers at eight ship attachment areas. Forty species from seven phyla were identifed through qualitative and quantitative surveys, with 17 attached species, including 10 non-indigenous species (NIS). Dominant species included *Balanus trigonus*, *Amphibalanus amphitrite*, and *Bugula neritina*. Cluster and non-metric multidimensional scaling analyses (nMDS) revealed four distinct groups based on species density, with signifcant diferences between domestic–international and international ships. Statistical analyses indicated signifcant diferences in species number, density, richness, and diversity index among groups, with Group *D* showing the highest values. The study emphasized international ships as primary introducers of NIS. Unique fndings included diferences in biofouling based on ship shape, antifouling paint conditions, and speed, highlighting the need for tailored management strategies based on navigation type and attachment area. This study urges further research to explore diferences in attachment areas and emphasizes the importance of obtaining more information about ships for efective management.

Keywords Biofouling · Hull fouling · Invasive alien species (IAS) · Macroinvertebrate · Non-indigenous species (NIS)

1 Introduction

The rapid growth of maritime transportation since the industrial revolution has increased the movement of marine organisms via ships, both in the ballast and when attached to the hull (Suk [2018\)](#page-9-0). Several studies have suggested that biofouling introduces and spreads more organisms than those introduced through ballast water (Eldredge and Carlton [2002](#page-9-1); Farrapeira et al. [2011](#page-9-2)).

In 2004, the International Maritime Organization (IMO) adopted the International Convention for the Control and Management of Ships' Ballast Water and Sediments in response to the challenges posed by the movement of organisms via ships. For organisms adhering to hulls, in 2011, the IMO introduced a guideline for the control and management of ship biofouling aimed at minimizing the transfer of invasive aquatic species. However, unlike the preventive measures targeting ballast water, in which the recommendation took the form of a resolution, the guideline is not legally enforceable (Suk [2018\)](#page-9-0). In several countries, national laws regulating the management of hull-attached organisms have been enacted (Hyun et al. [2018](#page-9-3); Park et al. [2022;](#page-9-4) Shin and Park [2020](#page-9-5)), but there are no clear guidelines in South Korea.

Biofouling on ships has increased, despite advances in antifouling paints (Lewis [2001\)](#page-9-6). Biofouling of the hull causes a variety of economic and environmental problems, including increasing the cost of ship repair and maintenance (Schultz et al. [2011](#page-9-7)), and exerting drag,

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which reduces energy efficiency and, in turn, increases greenhouse gas emissions (Liu et al. [2023](#page-9-8)). In particular, biofouling of the hull is an important vector for the introduction of non-indigenous species (NIS). International ships are a major route for the introduction, spread, and settlement of NIS (Davidson et al. [2009](#page-8-0)), with negative efects on native species and human health (Georgiades et al. [2021](#page-9-9); Shine et al. [2000](#page-9-10)). Information on biofouling species is needed to determine the impacts of NIS on marine ecosystems, but studies are very limited.

Ships have large underwater surface areas; this favors the growth of marine organisms (Liu et al. [2023\)](#page-9-8), whose composition and density may differ depending on the attachment area. For example, niche areas are less afected by external factors than exposed areas and, due to their complex structure, are less likely to be coated with antifouling paint, thus providing a safe sheltering place for organisms (Davidson et al. [2009;](#page-8-0) Moser et al. [2017;](#page-9-11) Otani [2006](#page-9-12); Ulman et al. [2019](#page-9-13)). Bulbous bows also increase biofouling due to hydrodynamic forces (Alamsyah et al. [2018\)](#page-8-1). For such reasons, macroinvertebrates that attach to the hull may have regional specifcity.

Several recent studies have reported that international ships are the main vector for introducing NIS (Çinar et al. [2014](#page-8-2); Pinochet et al. [2023\)](#page-9-14). However, while barnacles have been surveyed as an evaluation method for managing biofouling in South Korea (Park et al. [2022](#page-9-4); Shin and Park [2020\)](#page-9-5), only biofouling of international ships has been investigated (Park et al. [2022\)](#page-9-4). No accurate species information has been obtained, and the number of studies is limited (Lee et al. [2010;](#page-9-15) Park et al. [2022](#page-9-4)). This study aimed to investigate and compare the species distribution patterns of attached macroinvertebrates on six ships with diferent navigation types (domestic–international and international) and attachment areas in 2021 and 2022.

2 Materials and Methods

Six ships (*A*–*F*) were assessed to investigate the management of attached macroinvertebrates. Ship type, size, construction date, main routes, average speed, type of antifouling paint used, and cleaning status were determined. Information on ship management was obtained directly from ship owners and inspectors. Additional operational information was obtained from TradLinx [\(www.tradlinx.com](http://www.tradlinx.com)), the Integrated Port Management Information System (new.portmis.go.kr), a public database of ship operations (www.data.go.kr), and My Ship Tracking [\(www.myshiptracking.com](http://www.myshiptracking.com)).

Scuba divers collected quadrat samples $(15 \times 15 \text{ cm})$ from eight attachment areas (bottom, bow, hull, rudder, screw, shaft, stern, and thruster)on six ships (three each in 2021 and 2022) (Fig. [1](#page-1-0)a), and qualitative samples were collected from eight attachment areas (bottom, bow, hull, rudder, screw, shaft, stern, and thruster) where high concentrations of macroinvertebrates were randomly sampled (Fig. [1](#page-1-0)b). For qualitative analyses, sections within the eight areas with high concentrations of macroinvertebrates were randomly sampled. The samples were transported to the laboratory, where species identifcation was performed by sieving the organisms through a 1-mm mesh sieve, after which the densities and wet weights of the organisms were recorded. Once analyzed, the samples were fxed in 70% ethanol.

The number and wet weight (*g*) of individuals were measured, and the organisms were identified at the species level. Macroinvertebrate number and wet weight (*g*) were calculated per m^2 , and the species number, density

Fig. 1 Sample methods for attached macroinvertebrates: **a** Quantitative sampling; **b** Qualitative sampling

(individuals/ $m²$), and biomass (g/m²) were analyzed. The Shannon–Weiner diversity index $H(\log_e)$ (Colwell [2005\)](#page-8-3) was calculated for the density data. Analysis of variance (ANOVA) was used to determine differences in density based on navigation type and attachment area (SigmaPlot 11; Systat Software Inc., San Jose, CA, USA). The results of cluster analysis of biofouling community structure were analyzed using the Bray–Curtis similarity index (Chapman [1998\)](#page-8-4) on fourth-root-transformed species density data. A similarity profile permutation (SIMPROF) test was used to identify differences among species groups, and a similarity percentage (SIMPER) analysis was used to determine which species were the main drivers of the similarity and dissimilarity values of the groups. Permutational analysis of variance (PER-MANOVA) was used to analyze differences in macroinvertebrate species similarity among attachment areas and navigation types. Community analyses were performed using PRIMER 7 (Systat Software Inc.).

3 Results

3.1 Ship Information

The six investigated ships were designated as *A*–*F* based on the date of the survey. Specific information on the ships is presented in Table [1](#page-3-0). Three of the ships (*A*–*C*) were research ships that were operated both domestically and internationally. The other three ships (*D*–*F*) were full container ships that were operated only internationally. The container ships were larger than the research ships and were operated at higher speeds. The antifouling paint was oldest on ship *B* (February 2003), while the hulls of ships *D* and *F* had not been cleaned. Information on hull cleaning was not available for ship *C* (Table [1](#page-3-0)).

3.2 Species Composition

Forty species belonging to seven phyla were identified in the qualitative and quantitative surveys (Table [2\)](#page-4-0). Based on navigation type, 21 species belonging to seven phyla appeared on domestic–international ships, and 22 species belonging to three phyla appeared on international ships. Among all species, 17 were attached, 10 of which were NIS; 7 attached NIS were on domestic–international ships, while 5 were on international ships (Table [2\)](#page-4-0).

Ten dominant species were high ranking based on average density (> 2% average density); *Balanus trigonus* (Thecostraca) predominated (45.39%) (Table [3\)](#page-5-0), followed by *Amphibalanus amphitrite* (Thecostraca) (9.22%), *Bugula neritina* (Gymnolaemata) (7.56%),

Magallana gigas (Bivalvia) (6.18%), *Amphibalanus reticulatus* (Thecostraca) (4.66%), *Fistulobalanus kondakovi* (Thecostraca) (3.91%), *Pseudopotamilla occelata* (Polychaeta), *Actinia equina* (Anthozoa), *Megabalanus rosa* (Thecostraca), and *Monocorophium acherusicum* (Amphipoda) (Table [3\)](#page-5-0). *A. amphitrite* had the highest frequency of appearance, at 81.82%, followed by *B. trigonus* (36.36%) and *M. acherusicum* (36.36%) (Table [3\)](#page-5-0).

3.3 Distribution Patterns According to Navigation Type and Attachment Area

Cluster and non-metric multidimensional scaling analyses (nMDS) of species density revealed four groups (PERMANOVA, df = 3, *F* = 5.919, *p* = 0.001; Fig. [2\)](#page-5-1). Groups *A* and *B* were domestic–international and differed significantly from groups *C* and *D*, which were international (PERMANOVA, $df = 1$, $F = 4.036$, $p = 0.005$). Group *A* was attached at the stern of domestic–international ships; based on SIMPER analysis, the average similarity was 50.63%, and the species that contributed to the community was *B. trigonus*. Group B was attached at the bow and midships of domestic–international ships; the average similarity was 54.95%, and the species that contributed to the community were *B. neritina*, *P. occelata*, *A. equina*, and *A. amphitrite*. Group *C* was attached to the midships and stern of international ships; the average similarity was 47.27%, and the species that contributed to the community was *A. amphitrite*. Group *D* was attached at the bow of international ships; the average similarity was 34.97%, and the species that contributed to the community were *M. rosa*, *A. amphitrite*, and *Ericthonius pugnax* (Table [4\)](#page-6-0).

There were significant differences among groups in number of species (one-way ANOVA, $df = 3$, $p = 0.009$), species density (one-way ANOVA, $df = 3$, $p = 0.005$), species richness (one-way ANOVA, $df = 3$, $p = 0.01$), and diversity index (H) (One-way ANOVA, $df = 3$, $p = 0.05$) (Fig. [3](#page-6-1)), all four of which were highest in group *D*. Based on the SIMPER analysis, the average dissimilarity between groups ranged from 76.65% (*D* and *C*) to 100% (*A* and *C*) (Table [5](#page-7-0)). In addition to the previously mentioned species, *Hydroides elegans* and *Koinostylochus* sp. also contributed to the dissimilarity between communities (Table [5](#page-7-0)).

The effect of navigation type was significantly dependent on the attachment area (two-way ANOVA, $df = 2$, $F = 10.18$, $p=0.02$; Table [6](#page-7-1)). On domestic–international vessels, there was a significant difference between midships and stern $(p=0.004)$ and bow and stern $(p=0.03)$; on international vessels, there was a signifcant diference between bow and midships ($p = 0.002$; Table [6\)](#page-7-1).

Phylum	Species name	NIS classification	NIS origin (first record areas)	Navigation type
Annelida	Pseudopotamila occelata	Indigenous species		$D+I$
Annelida	Nereis sp.	Indigenous species		$D+I$
Annelida	Lepidonutus sp.	Indigenous species		$D+I$
Annelida	Hydroides elegans	NIS	Indo-Pacific (Cinar 2012)	$D+I$
Arthropoda	Unidentified sp.	Indigenous species		I
Arthropoda	Stenothoe valida	Indigenous species		$D+I$
Arthropoda	Protomedeia sp.	Indigenous species		Ι
Arthropoda	<i>Pareurystheus</i> sp.	Indigenous species		Ι
Arthropoda	Monocorophium insidiosum	Indigenous species		Ι
Arthropoda	Monocorophium acherusicum	Indigenous species		$\mathbf I$
Arthropoda	Melita hoshinoi	Indigenous species		$\mathbf I$
Arthropoda	Megabalanus rosa	NIS	Japan, Formosa (Carlton et al. 2011)	Ι
Arthropoda	Lepas anatifera	Indigenous species		I
Arthropoda	Jassa slatteryi	Indigenous species		I
Arthropoda	Gnorimosphaeroma ovatum	Indigenous species		$D+I$
Arthropoda	Gnorimosphaeroma naktongense	Indigenous species		I
Arthropoda	Fistulobalanus kondakovi	Indigenous species		\bf{I}
Arthropoda	Ericthonius pugnax	Indigenous species		\bf{I}
Arthropoda	Crassicorophium crassicorne	Indigenous species		I
Arthropoda	Conchoderma auritum	NIS	Cosmopolitan (Foster and Willan 2010)	$D+I, I$
Arthropoda	Ceradocus sp.	Indigenous species		$D+I$
Arthropoda	Caprella scaura	Indigenous species		$D+I$
Arthropoda	Caprella penantis	Indigenous species		$D+I$
Arthropoda	Caprella equilibra	Indigenous species		Ι
Arthropoda	Caprella californica	Indigenous species		I
Arthropoda	Balanus trigonus	Indigenous species		$D+I, I$
Arthropoda	Apocorophium acutum	Indigenous species		$D+I$
Arthropoda	Anoplodactylus crassus	Indigenous species		$D+I$
Arthropoda	Amphibalanus reticulatus	NIS	Indo-Pacific (Carlton et al. 2011)	I
Arthropoda	Amphibalanus eburneus	NIS	Northwest Atlantic (Carlton et al. 2011)	Ι
Arthropoda	Amphibalanus amphitrite	NIS	Indo-Pacific (Carlton et al. 2011)	$D+I, I$
Bryozoa	Bugula neritina	NIS	Pacific Coast (Robertson 1905)	$D+I$
Chordata	Ciona intestinalis	NIS	Europe (Park et al. 2018)	$D+I$
Cnidaria	Plumularia setacea	Indigenous species		\bf{I}
Cnidaria	Actinia equina	Indigenous species		$D+I$
Mollusca	Perna viridis	NIS	Indo-Pacific (Siddall 1980)	$D+I$
Mollusca	Mytilus galloprovincialis	NIS	Mediterranean Sea (Siddall 1980)	$D+I$
Mollusca	Magallana gigas	Indigenous species		$D+I$
Mollusca	Brachidontes mutabilis	Indigenous species		Ι
Platyhelminthes	Kinostylochus sp.	Indigenous species		$D+I$

Table 2 List of species detected on the seven studied ships: species name, non-indigenous species (NIS) classifcation, NIS origin (frst recorded area), and appearance of macroinvertebrate by navigation type

D domestic, *I* International

4 Discussion

Forty species were identifed in qualitative and quantitative surveys of eight attachment areas on six ships (Table [2](#page-4-0)). *B. trigonus* was the most predominant species (Table [3](#page-5-0)), while *A. amphitrite* had the highest frequency of appearance (Table [3\)](#page-5-0). *A*. *amphitrite*, *Amphibalanus eburneus*, *A. reticulatus*, *B. neritina*, *H. elegans*, *Ciona intestinalis*, *Conchoderma auritum*, *M. rosa*, *Mytilus galloprovincialis*, and *Perna viridis* were NIS identifed in this study (Carlton et al. [2011;](#page-8-5) Çinar [2012;](#page-8-6) Foster and Willan [2010;](#page-9-16) Lee et al. [2010;](#page-9-15) Park et al. [2018;](#page-9-17) Siddall **Table 3** Dominant species ranking based on the density of macroinvertebrates as a result of quantitative survey

Taxa	Species	Average density (individuals/m ²)	% of total density	Frequency $(\%)$
ACr	Balanus trigonus	1454.55	45.39	36.36
ACr	Amphibalanus amphitrite	295.62	9.22	81.82
BGy	Bugula neritina	242.42	7.56	27.27
MBi	Magallana gigas	197.98	6.18	18.18
ACr	Amphibalanus reticulatus	149.49	4.66	9.09
ACr	Fistulobalanus kondakovi	125.25	3.91	9.09
APo	Pseudopotamila occelata	123.23	3.85	27.27
CAn	Actinia equina	84.85	2.65	27.27
ACr	Megabalanus rosa	80.81	2.52	18.18
ACr	Monocorophium acherusicum	72.73	2.27	36.36

ACr Crustacea, *BGy* Gymnolaemata, *MBi* Bivalvia, *APo* Polychaeta, *CAn* Anthozoa

Fig. 2 Cluster analysis and non-metric multidimensional scaling (nMDS) using Bray–Curtis similarities, based on the fourth-root transformed abundance data

[1980](#page-9-19)). In previous studies, only juvenile *P*. *viridis* were reported domestically (Lee et al. [2010](#page-9-15)) However, in this study, adults were also discovered. Thus, we anticipated that *P*. *viridis* would establish itself in Korea in the future. Additionally, *C*. *intestinalis* is a known biofouling organism and a designated NIS, and it is managed as a disturbance organism by the Ministry of Oceans and Fisheries (Park et al. [2018](#page-9-17)). *C*. *intestinalis* may decrease ecosystem diversity.

The species composition and distribution patterns differed between the navigation types (Table [2\)](#page-4-0). Reported that international ships are the main factor of NIS, and the domestic–international navigation type acts as a secondary factor in spreading NIS in the country (Otani [2006](#page-9-12)). In this study, *A*. *reticulatus* and *M*. *rosa* appeared only on the international navigation type of ship (Table [1\)](#page-3-0). This species was NIS with a low habitat density on domestic–international (Kim et al. [2019\)](#page-9-20). Among the NIS, *A*. *amphitrite*, *A*. *eburneus*, *C*. *intestinalis*, and *M*. *galloprovincialis* are completely settled in country (Kim et al. [2019;](#page-9-20) Lee et al. [2010](#page-9-15); Lee and Shin [2014](#page-9-21)), and *Hydroides eleganss* is a representative invasive species (Bagaveeva and Zvyagintsev [2000\)](#page-8-7). In this study, *A*. *amphitrite* and *A*. *eburneus* were dense on international hulls, but *H. eleganss* was dense on the domestic and international sterns, and *C*. *intestinalis*, *H. eleganss* and *M*. *galloprovincialis* were dense on the mid and bow (Fig. [4\)](#page-8-8). Previous studies concluded that ship speed is one of the most important determinants of the survival of biofouling organisms (Coutts et al. [2010](#page-8-9); Davidson et al. [2009](#page-8-0)). The average speed of the domestic–international ships examined in this study was 12 kn, which is less than half that of international ships (29.7 kn) (Table [1\)](#page-3-0). Species density was higher on the slower domestic–international ships (Fig. [2](#page-5-1)). International ships were the main contributor to NIS, and domestic–international vessels will spread such species into domestic waters, where they have not yet been introduced.

In this study, no clear trend emerged when comparing density, cleaning information, and operating period on each ship (Figure [5\)](#page-8-10). However, the number and density of species on the examined attachment areas difered based on the navigation type (Figure [3](#page-6-1)). While previous studies showed diferences in exposed and niche areas due to external environmental factors (Moser et al. [2017](#page-9-11)), this study found no such diference. However, a unique fnding of this study was the diference in biofouling between bows, midships, and sterns depending on the navigation type (Figure [4\)](#page-8-8). This $Table 4$

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Fig. 3 The macroinvertebrate communities according to navigation type and groups: **a** Number of species (*S*); **b** Density (*N*); **c** Richness (*d*); **d** Diversity (H´)

diference can be attributed to several factors, such as the condition of the antifouling paint, the shape of the ship, and ship speed. Cargo ships, in particular, have a unique bow shape and may have a high rate of antifouling paint removal (Coutts and Taylor [2004\)](#page-8-11), which can increased biofouling due to external factors, such as shear and pressure forces (Alamsyah et al. [2018\)](#page-8-1). The international ships investigated in this study were also cargo ships (Table [1\)](#page-3-0), and the occurrence of high biofouling in Group *D* (bow) was infuenced by these factors. Group *B*, which showed high biofouling in midships and bows, appeared to have high biofouling even on the fat and wide midships due to low-speed operation (Coutts et al. [2010;](#page-8-9) Davidson et al. [2009](#page-8-0)). These results suggest that diferent management approaches are needed for each attached area depending on the navigation type.

Maritime transportation has brought signifcant benefts to humanity, but it has also caused many problems. Korea is surrounded by the sea on three sides, and its many ports, together with its oceanographic characteristics, provide a perfect environment for the settlement of NIS (Kim et al. [2020;](#page-9-22) Ubagan et al. [2021](#page-9-23)). However, only a few NIS have been surveyed to date, and their current status is unknown (Lee et al. [2010](#page-9-15)). The results of this study indicate that different management strategies are required depending on the

Table 5 SIMPER analysis of main characterizing species at each group (*A*–*D*; Av. Abund, average abundance; Av. Diss, average dissimilarity; Contrib%, contribution%; Cum%, cumulative%; 5<Contrib%)

	Species		Av. Abund Av. Abund Av. Diss Diss/SD Contrib%				$Cum.\%$
Groups A and B average dissimilarity = 92.09	Balanus trigonus	9.09	0.86	16.41	2.06	17.82	17.82
	Bugula neritina	$\boldsymbol{0}$	5.46	10.58	3.75	11.49	29.31
	Pseudopotamila occelata	$\mathbf{0}$	4.61	8.98	2.83	9.76	39.07
	Amphibalanus amphitrite	$\mathbf{0}$	4.22	8.35	2.62	9.07	48.14
	Actinia equina	$\mathbf{0}$	4.15	8.17	2.82	8.87	57.01
	Magallana gigas	3.4	0.86	6.21	0.99	6.74	63.75
Groups A and C average	Balanus trigonus	9.09	$\boldsymbol{0}$	38.94	2.24	38.94	38.94
$dissimilarity = 100.00$	Amphibalanus amphitrite	$\boldsymbol{0}$	4.32	17.7	4	17.7	56.64
	Magallana gigas	3.4	$\boldsymbol{0}$	12.11	0.91	12.11	68.75
	Monocorophium acherusicum	$\mathbf{0}$	1.97	6.92	0.91	6.92	75.67
Groups A and D average dissimilarity = 93.28	Balanus trigonus	9.09	2.1	12.61	1.79	13.52	13.52
	Amphibalanus amphitrite	$\boldsymbol{0}$	5.62	9.6	6.19	10.29	23.82
	Megabalanus rosa	$\boldsymbol{0}$	5.01	8.66	14.13	9.29	33.1
	Ericthonius pugnax	$\mathbf{0}$	4.36	7.76	2.33	8.31	41.42
	Monocorophium acherusicum	$\boldsymbol{0}$	3.36	5.88	4.96	6.3	47.72
	Fistulobalanus kondakovi	$\mathbf{0}$	3.05	5.71	0.86	6.12	53.84
	Magallana gigas	3.4	$\boldsymbol{0}$	5.56	0.86	5.96	59.8
	Amphibalanus reticulatus	$\boldsymbol{0}$	3.18	5.1	0.86	5.47	65.26
Groups B and D average dissimilarity = 86.70	Bugula neritina	5.46	$\overline{0}$	6.66	5.96	7.68	7.68
	Megabalanus rosa	$\mathbf{0}$	5.01	6.09	6.45	7.02	14.7
	Pseudopotamila occelata	4.61	$\overline{0}$	5.64	3.63	6.5	21.2
	Ericthonius pugnax	$\boldsymbol{0}$	4.36	5.41	2.4	6.24	27.44
	Actinia equina	4.15	$\boldsymbol{0}$	5.11	3.93	5.89	33.33
Groups B and C average dissimilarity = 83.54	Bugula neritina	5.46	$\boldsymbol{0}$	11.38	3.44	13.63	13.63
	Pseudopotamila occelata	4.61	$\boldsymbol{0}$	9.67	2.71	11.57	25.2
	Actinia equina	4.15	$\mathbf{0}$	8.8	2.68	10.53	35.73
	Hydroides elegans	2.5	$\boldsymbol{0}$	4.29	1.34	5.14	40.87
	Kinostylochus sp.	2.5	$\boldsymbol{0}$	4.29	1.34	5.14	46.01
Groups D and C average dissimilarity = 76.65	Megabalanus rosa	5.01	$\boldsymbol{0}$	9.2	10.38	12	12
	Ericthonius pugnax	4.36	$\boldsymbol{0}$	8.25	2.44	10.77	22.77
	Fistulobalanus kondakovi	3.05	$\boldsymbol{0}$	6.09	0.93	7.95	30.72
	Amphibalanus reticulatus	3.18	$\boldsymbol{0}$	5.39	0.93	7.03	37.74
	Monocorophium acherusicum 3.36		1.97	3.92	1.25	5.12	42.86

Table 6 Results of statistical analysis of navigation and attachment area efects Shannon–Wiener diversity

(*p*-values: *,<0.05)

Fig. 4 The density of domestically settled non-indigenous species (NIS) by group

Fig. 5 The density, operating date, and cleaning information (*C*/*Y*=Cleaning/Yes, *C*/*N*=Cleaning/No and *C*/Un=Cleaning/ Unknown) of each ship

type of ship and the location of the operation, as there were diferences in species number and density between the navigation types and attachment areas. Future research should investigate the diferences between attachment areas with by increasing sampling frequency. Close communication with ship owners and shipping companies is required to obtain more ship information.

5 Conclusion

This study investigated the diversity and distribution patterns of biofouling macroinvertebrates on diferent types of ships. We identifed 40 species, among which *B. trigonus* was the most dominant in terms of density. Several NIS were identifed, including *A. amphitrite*, *A. eburneus*, and *M. rosa*, and *P. viridis* could potentially settle in Korean waters

in the future. Signifcant diferences in species composition and density were found between navigation types, indicating that international ships play a major role in introducing NIS, while domestic–international ships contribute to their spread; ship speed was determined to be a crucial factor, with slower speeds associated with higher species densities. The variation in species diversity between navigation types and attachment areas underscores the need for tailored management strategies.

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Data availability The raw data that support the fndings of this study are available from the corresponding authorupon reasonable request to corresponding author and with permission from Korea Institue of Ocean Science andTechnology (KIOST).

References

- Alamsyah MA, Hakim ML, Utama I (2018) Study of shear and pressure flow on the variation of ship hull shapes as one of the biofouling growth factors. In: Proceedings of the 3rd international conference on marine technology. SCITEPRESS-Science and Technology Publications
- Bagaveeva EV, Zvyagintsev AYu (2000) The Introduction of Polychaetes Hydroides elegans (Haswell), Polydora limicola Annenkova, and Pseudopotamilla occelata Moore to the Northwestern Part of the East Sea. Ocean Polar Res 22(1):25-36
- Carlton JT, Newman WA, Pitombo FB (2011) Barnacle invasions: Introduced, cryptogenic, and range expanding cirripedia of North and South America. In: Galil BS, Clark PF, Carlton JT (eds), In the wrong place-Alien marine crustaceans: Distribution, biology and impacts. Springer Dordrecht, Berlin, pp 159–213. [https://doi.](https://doi.org/10.1007/978-94-007-0591-3_5) [org/10.1007/978-94-007-0591-3_5](https://doi.org/10.1007/978-94-007-0591-3_5)
- Chapman M (1998) Relationships between spatial patterns of benthic assemblages in a mangrove forest using diferent levels of taxonomic resolution. Mar Ecol-Prog Ser 162:71–78
- Çinar ME (2012) Alien polychaete species worldwide: current status and their impacts. J Mar Biol Assoc UK 93(5):1257–1278. [https://](https://doi.org/10.1017/s0025315412001646) doi.org/10.1017/s0025315412001646
- Çinar ME, Arianoutsou M, Zenetos A, Golani D (2014) Impacts of invasive alien marine species on ecosystem services and biodiversity: a pan-European review. Aquat Inv 9(4):391–423
- Colwell RK (2005) EstimateS: Statistical estimation of species richness and shared species from samples. Version 7.5. User's Guide and application. <http://purl.oclc.org/estimates>
- Coutts AD, Taylor MD (2004) A preliminary investigation of biosecurity risks associated with biofouling on merchant vessels in New Zealand. New Zeal J Mar Fresh 38(2):215–229
- Coutts AD, Piola RF, Taylor MD, Hewitt CL, Gardner JP (2010) The efect of vessel speed on the survivorship of biofouling organisms at diferent hull locations. Biofouling 26(5):539–553
- Davidson IC, Brown CW, Sytsma MD, Ruiz GM (2009) The role of containerships as transfer mechanisms of marine biofouling species. J Bioad Biof Res 25:645–655. [https://doi.org/10.1080/08927](https://doi.org/10.1080/08927010903046268) [010903046268](https://doi.org/10.1080/08927010903046268)
- Eldredge LG, Carlton JT (2002) Hawaiian marine bioinvasions: a preliminary assessment. Pac Sci 56(2):211–212
- Farrapeira CMR, de Oliveira Tenório D, do Amaral FD (2011) Vessel biofouling as an inadvertent vector of benthic invertebrates occurring in Brazil. Mar Pollut Bull 62(4):832–839
- Foster BA, Willan RC (2010) Foreign barnacles transported to New Zealand on an oil platform. New Zeal J Mar Fresh 13(1):143–149. <https://doi.org/10.1080/00288330.1979.9515788>
- Georgiades E, Scianni C, Davidson I, Tamburri MN, First MR, Ruiz G, Ellard K, Deveney M, Kluza D (2021) The role of vessel biofouling in the translocation of marine pathogens: management considerations and challenges. Front Mar Sci 8:660125
- Hyun B, Jang P-G, Shin K, Kang J-H, Jang M-C (2018). Ship's Hull Fouling Management and In-Water Cleaning Techniques. J Korean Soc Mar Environ Saf 24:785-795[.https://doi.org/10.7837/](https://doi.org/10.7837/kosomes.2018.24.6.785) [kosomes.2018.24.6.785](https://doi.org/10.7837/kosomes.2018.24.6.785)
- Kim H, Lee S-K, Kim W, Min BUM (2019) Report on the current status of the distribution of invasive barnacles in marine national park areas of Korea. J Nat Park Res 10(2):249–257
- Kim HK, Chan BK, Lee S-K, Kim W (2020) Biogeography of intertidal and subtidal native and invasive barnacles in Korea in relation to oceanographic current ecoregions and global climatic changes. J Mar Biol Assoc UK 100(7):1079–1091
- Lee T, Shin S (2014) Morphological and molecular identifcation of an introduced alien sea squirt (Tunicata: Ascidiacea) in Korea. P Biol Soc Wash 127(1):284–297
- Lee J-S, Lee YS, Min D-K (2010) Introduced molluscan species to Korea. Korean J Malacol 26(1):45–49
- Lewis J (2001) Hull fouling as a vector for the translocation of marine organisms. phase 1 study-hull fouling research. Water Res Ser Rep 14:1–129
- Liu S, Kee YH, Shang B, Papanikolaou A (2023) Assessment of the economic, environmental and safety impact of biofouling on a ship's hull and propeller. Ocean Eng 285:115481
- Moser CS, Wier TP, First MR, Grant JF, Riley SC, Robbins-Wamsley SH, Tamburri MN, Ruiz GM, Miller AW, Drake LA (2017) Quantifying the extent of niche areas in the global feet of commercial ships: the potential for "super-hot spots" of biofouling. Biol Inv 19(6):1745–1759
- Otani M (2006) Important vectors for marine organisms unintentionally introduced to Japanese waters. In: Koike F (ed) Assessment and control of biological invasion risks. Japan and the World Conservation Union (IUCN), Gland, pp 92–103
- Park J-U, Hong J, Kim DG, Yoon TJ, Shin S (2018) Prediction of the suitable habitats of marine invasive species, ciona robusta based on RCP scenarios. Korean J Environ Biol 36(4):687–693
- Park J, Hoe C, Kim H, Cho Y (2022) Study on the biofouling management of international ships entering South Korea. J Korean Soc Mar Environ Safe 28(1):10–18

Pinochet J, Brante A, Daguin-Thiébaut C, Tellier F, Viard F (2023) Investigating the risk of non-indigenous species introduction through ship hulls in Chile. Manag Biol Invasions 14(1):156–177

Robertson A (1905) Non-incrusting chilostomatous Bryozoa of the west coast of North America (Vol. 2). The University Press, p 322

Schultz MP, Bendick J, Holm E, Hertel W (2011) Economic impact of biofouling on a naval surface ship. Biofouling 27(1):87–98

- Shin Y-H, Park H-S (2020) A case study on the management of biofouling for protection of the marine ecosystem. J Navi Port Res 44(3):151–157
- Shine C, Williams N, Gündling L (2000) A guide to designing legal and institutional frameworks on alien invasive species. IUCN, Gland, p 153
- Siddall SE (1980) A clarifcation of the *Genus Perna* (Mytilidae). Bull Mar Sci 30(4):858–870
- Suk J (2018) A study on the regulatory framework related to ship's biofouling. Korea Inst Mar Law 30(1):139–173
- Ubagan MD, Lee YS, Lee T, Hong J, Kim IH, Shin S (2021) Settlement and recruitment potential of four invasive and one indigenous barnacles in South Korea and their future. Sustainability 13(2):634
- Ulman A, Ferrario J, Forcada A, Seebens H, Arvanitidis C, Occhipinti-Ambrogi A, Marchini A (2019) Alien species spreading via biofouling on recreational vessels in the Mediterranean Sea. J Appl Ecol 56(12):2620–2629

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