



# Photobiology of lipofuscin granules in the retinal pigment epithelium cells of the eye: norm, pathology, age

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## Abstract

Lipofuscin granules (LGs) are accumulated in the retinal pigment epithelium (RPE) cells. The progressive LG accumulation can somehow lead to pathology and accelerate the aging process. The review examines composition, spectral properties and photoactivity of LGs isolated from the human cadaver eyes. By use of atomic force microscopy and near-field microscopy, we have revealed the fluorescent heterogeneity of LGs. We have discovered the generation of reactive oxygen species by LGs, and found that LGs and melanolipofuscin granules are capable of photoinduced oxidation of lipids. It was shown that A2E, as the main fluorophore (bisretinoid) of LGs, is much less active as an oxidation photosensitizer than other fluorophores (bisretinoids) of LGs. Photooxidized products of bisretinoids pose a much greater danger to the cell than non-oxidized one. Our studies of the fluorescent properties of LGs and their fluorophores (bisretinoids) showed for the first time that their spectral characteristics change (shift to the short-wavelength region) in pathology and after exposure to ionizing radiation. By recording the fluorescence spectra and fluorescence decay kinetics of oxidized products of LG fluorophores, it is possible to improve the methods of early diagnosis of degenerative diseases. Lipofuscin (“aging pigment”) is not an inert “slag”. The photoactivity of LGs can pose a significant danger to the RPE cells. Fluorescence characteristics of LGs are a tool to detect early stages of degeneration in the retina and RPE.

**Keywords** Retinal pigment epithelium · Lipofuscin granules · Bisretinoids · Photosensitization · Autofluorescence · AMD

## Introduction

Lipofuscin granules, or age pigment, are accumulated with aging in retinal pigment epithelium (RPE) cells of human eye and remain there to the end of life, occupying up to 20% of cell cytoplasmic volume (Feeney–Burns et al. 1984; Jung et al. 2077; Yin 1996). The mechanism of LG formation is related to the physiological functions of the retina and RPE.

The retina is the light-sensitive tissue of the eye (Fig. 1a). It consists of several layers of neurons interconnected by synapses (Fig. 1b). The primary light-sensing cells in the retina are the photoreceptor cells, rods and cones. The RPE is the pigmented single-cell layer just outside the retina, firmly attached to the underlying choroid and is in close contact

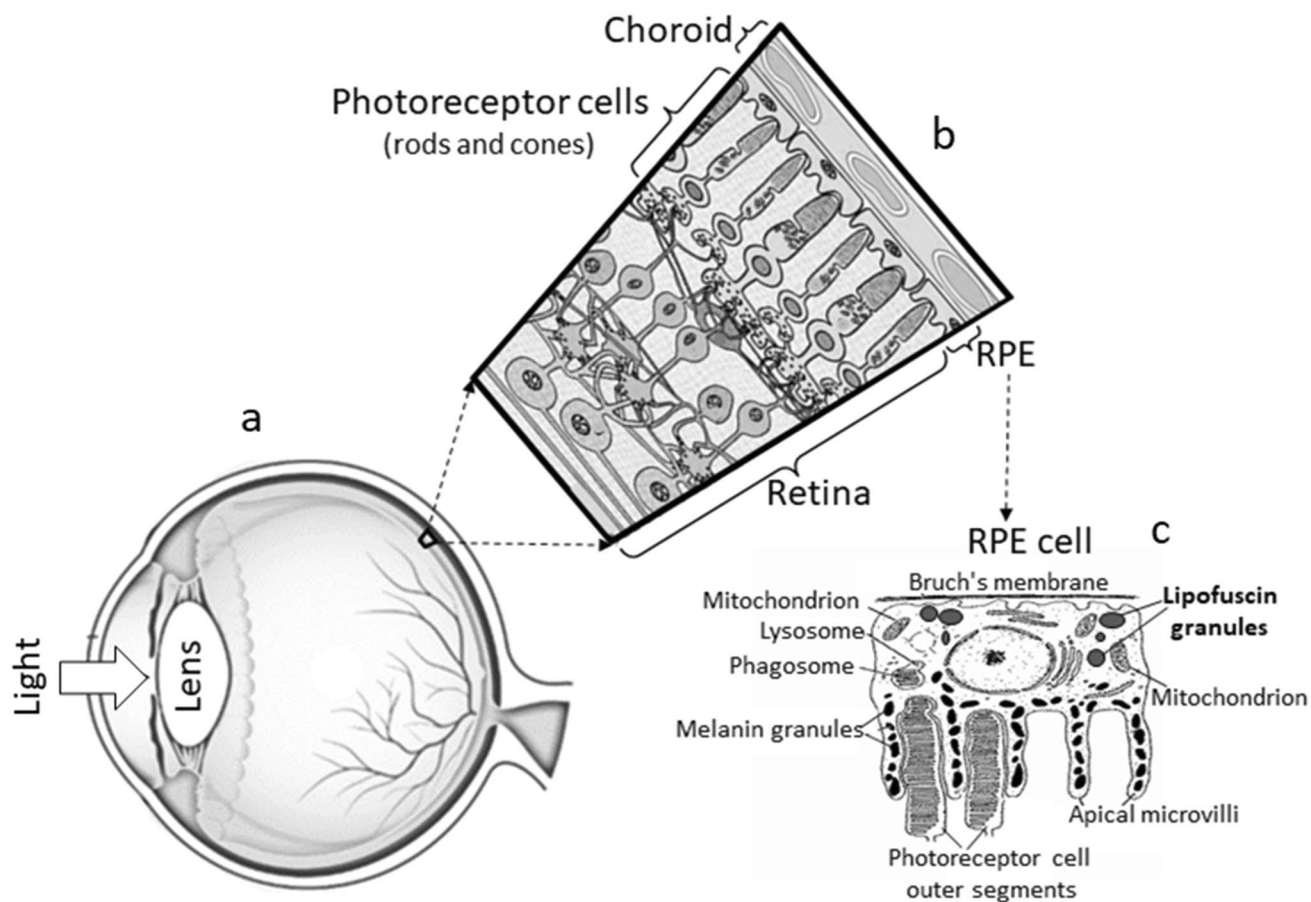
with the photoreceptor cells (Fig. 1b, c). The RPE has several crucial functions for vision, namely, scattered light absorption, epithelial transport, spatial ion buffering, visual cycle, phagocytosis of photoreceptor outer segment membranes, secretion and immune modulation (Strauss 2005).

With exposure to light during rhodopsin photolysis, retinoid side products can be produced in photoreceptor cells. Biogenesis of these products occurs when two molecules of *all-trans* retinal condense with one molecule of phosphatidylethanolamine (Fig. 2) in the photoreceptor membrane (Wolf 2003). Evolution has developed a powerful mechanism that prevents the accumulation of retinoid side products in terminally differentiated photoreceptor cells (Young 1967). Throughout life, the debris of the photoreceptor outer segment apical part, are phagocytized and digested by the RPE cells, while new photoreceptor discs with rhodopsin molecules are synthesized by the photoreceptor inner segments (Kennedy et al. 1995). However, the lysosomal enzyme system of the RPE cell is not effective in degrading of the photoreceptor outer segment debris, because the latter are supposed to contain modified

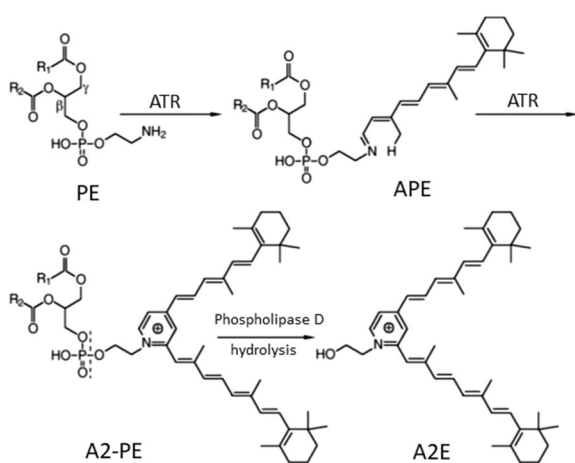
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**Fig. 1** Scheme of ocular exposure to light (a), the retina and retinal pigment epithelium (RPE) (b), and RPE cell containing LGs (c). Figure was modified from Yakovleva et al. (2022b)



**Fig. 2** Scheme of *N*-retinyl-*N*-retinylideneethanolamine (A2E) formation. ATR — *all-trans* retinal; PE — phosphatidylethanolamine; APE — *N*-retinylideneethanolamine; A2PE — *N*-retinyl-*N*-retinylideneethanolamine

retinoid side products of rhodopsin photolysis as well as modified lipids and proteins. In other words, the lysosomal enzyme system of the RPE cell cannot recognize such modified molecules and do not digest them (Feeney 1973). As a consequence, lipofuscin granules (LGs), containing retinoid derivatives, are formed in the RPE cells (Fig. 1c).

LGs have been long believed to be just a cell metabolism by-product. It turned, however, that they are photochemically active. As we have shown in the early 1990s, LGs can generate oxygen reactive species (ROS) upon photoexcitation with visible light, which account for their phototoxicity (Boulton et al. 1993). The main photo-inducible generators of ROS in LGs are retinoid side products. The features of these compounds, such as photosensitizing, have been studied in detail (Boulton et al. 1993; Sparrow et al. 2000; Rozanowska et al. 1995, 1998, 2005; Avallé et al. 2005). There is the correlation between LG accumulation in the RPE cells and development of degenerative retinal diseases, including such a severe and widespread retinopathy as age-related macular degeneration AMD (Holz et al. 2004; Katz 2002; Sparrow and Boulton 2005).

LGs are heterogeneous, composed of mixtures of proteins and lipids, including more than 21 different fluorescent compounds (Warburton et al. 2005; Bazan et al. 1990; Eldred and Katz 1988; Ng et al. 2008; Sparrow et al. 2009). The structure and fluorescence properties of LGs were analyzed by atomic force microscopy (AFM) and near-field microscopy (Yakovleva et al. 2016; Petrukhin et al. 2005; Warburton et al. 2005; Clancy et al. 2000). Figure 3a shows an image of aggregated LGs. Single LGs were found to be approximately 0.7–1.0  $\mu\text{m}$  in diameter, with a fairly uniform density. Figure 3b shows the fluorescence emission spectra of aggregated LGs, as determined by near-field microscopy, at the two points marked on Fig. 3a. The LG fluorescence emission spectrum 1 at point 1 (Fig. 3a) is broad and has several emission maxima, ranging approximately from 570 to 625 nm, that correlates with the fluorescence properties of LGs in suspension (Feldman et al. 2015; Boulton et al. 1990; Haralampus-Grynaviski et al. 2003).

Major sources of LG fluorescence are retinoid side products (Kennedy et al. 1995; Sparrow et al. 2012). They are bisretinoids (BisRets) and their oxidation and degradation derivatives (BisRets-OX) (Lamb and Simon 2004; Sakai et al. 1996; Sparrow et al. 2008, 2012; Wang et al. 2006a, b; Wu et al. 2010; Kim et al. 2007; Feldman et al. 2015). *N*-retinyl-*N*-retinylideneethanolamine (A2E) (Fig. 2) is the most widely studied BisRet (Lamb and Simon 2004; Sakai et al. 1996; Sparrow et al. 2008).

LGs exhibit distinct fluorescence in the visible region. Based on its measurement, fundus autofluorescence (AF) imaging is a modern noninvasive diagnostic method for revealing age-related base changes and degenerative retinal and RPE pathology. AF allows assessment of the condition, integrity and viability of the photoreceptor/RPE complex (Schmitz-Valckenberg et al. 2007).

This review considers the phototoxic properties of BisRets in LGs, as well as the cytotoxic properties of the

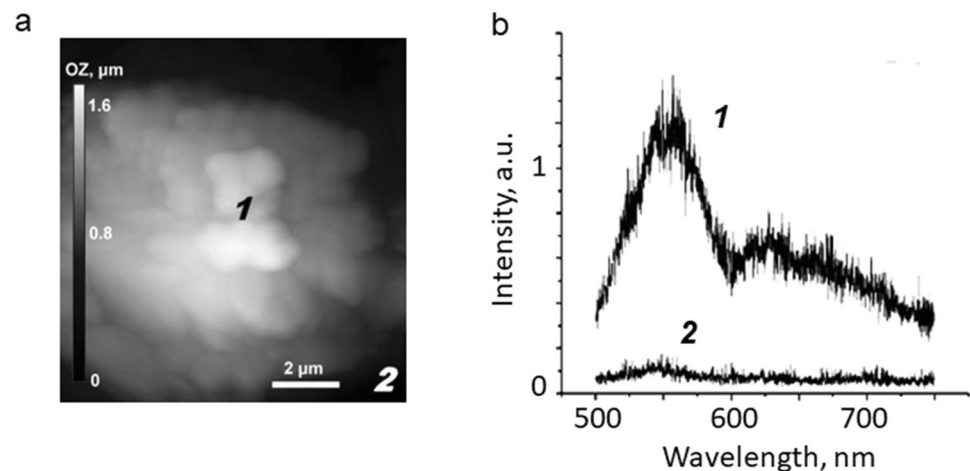
BisRets-OX. The review also examines the fluorescent properties of LGs in health and disease for the use of this knowledge in expanding the capabilities of the fundus AF method for diagnosing degenerative diseases of the retina and RPE.

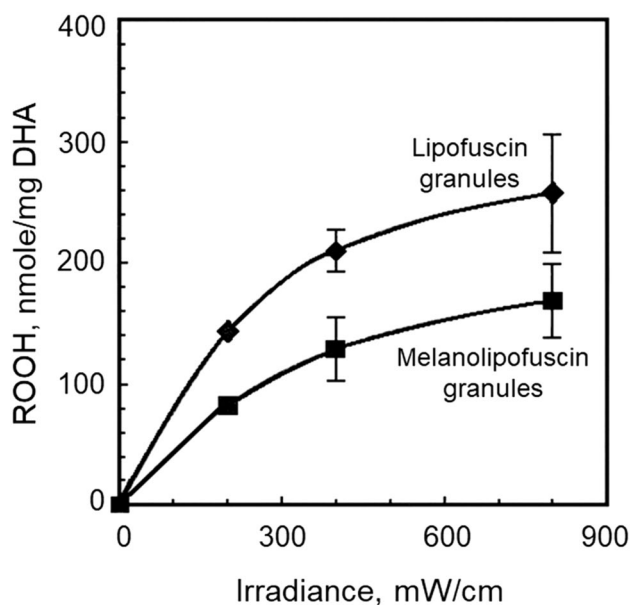
## Phototoxic properties of LGs in the RPE cell

It is well known that LGs in the RPE cells under the influence of visible light stimulate the oxidation of both lipids (Yin 1996; Wassel et al. 1999; Dontsov et al. 1999; Dontsov et al. 2005; Dontsov et al. 2012; Nowak 2013) and proteins (Wassell et al., 1999; Rozanowska et al., 2002; Wiktor et al., 2018). This is due to the ability of LGs to produce ROS under the action of visible light (Boulton et al. 1993; Rozanowska et al. 1995, 1998, 2002). LGs most effectively photogenerate ROS in the blue-green region of the spectrum (400–520 nm) (Boulton et al. 1993, 2004; Dontsov et al. 2012). Figure 4 demonstrates data on oxidation of 4,7,10,13,16,19-docosahexaenoic acid (DHA) induced by blue-green light irradiation and sensitized with LG and melanolipofuscin granules from human RPE cells. Both types of granules containing lipofuscin stimulated DHA peroxidation, but LGs were more active in this respect.

The ROS generation by LGs in the RPE under the action of blue light can explain the “blue light hazard” phenomenon for the retina (Rozanowska et al. 1995; Boulton et al. 2004). It has been shown, for example, that in the RPE cells loaded with LGs in vitro and irradiated with blue-green light, there is a significant increase in lipid and protein oxidation, accompanied by such morphological changes as loss of lysosome integrity (Brunk et al. 1995; Nilsson et al. 2003; Sparrow and Boulton 2005; Shamsi and Boulton 2001), damage to the cell membrane and increased vacuolization of the cytoplasm compared to the RPE cells irradiated with long-wavelength (> 550 nm) light (Boulton 2001). LGs in

**Fig. 3** Atomic force microscopy (AFM) image of aggregated LGs on a cover slip (a) and near-field microscopy analysis of the fluorescence emission spectra (b). The excitation wavelength was 420 nm. Emission spectrum 1 was recorded from point 1 (a) and corresponded to the aggregated LGs, whereas emission spectrum 2 was recorded from point 2 (a) and corresponded to the part of the slide glass without LGs. OZ represents the height ( $\mu\text{m}$ ) of the sample (Yakovleva et al. 2016)

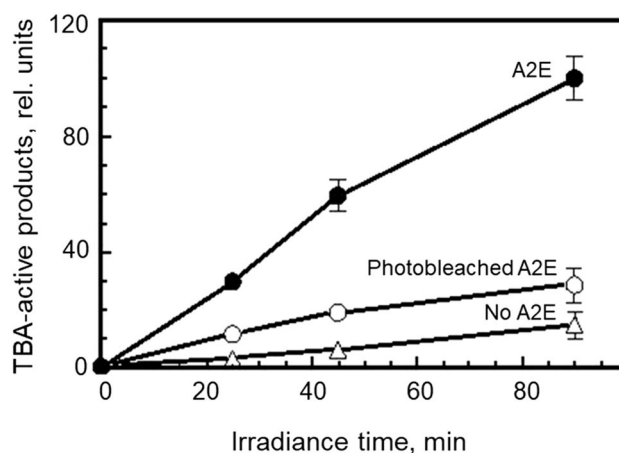




**Fig. 4** DHA (4,7,10,13,16,19-docosahexaenoic acid) peroxidation produced by photoactivation of lipofuscin and melanolipofuscin RPE pigment granules. The argon ion laser was used to generate blue-green light (488.1 nm and 514.5 nm; granule concentration was  $1.5 \times 10^7$  granules/mL). Figure was modified from Dontsov et al. (1999)

the RPE are also capable of photogeneration of singlet oxygen, the efficiency of which decreases monotonically with increasing wavelength of excitation light (Rożanowska, et al. 1998, 2004; Avasle et al. 2005).

The phototoxicity of LGs is associated with the presence in them of bisretinoid fluorophores that absorb light in the blue region of the spectrum. One of the main LG bisretinoids, A2E, has been shown to be localized not only in LGs, but also in other RPE cell compartments. Although the main amount of A2E is found mainly in lysosomes (Eldred and Lasky 1993; Sparrow et al. 1999; Holz et al. 1999; Schutt et al. 2007), but to a lesser extent, A2E accumulates in the mitochondria, Golgi apparatus, and cytoplasmic membrane (Schutt et al. 2007). The amount of A2E in the RPE cells in vivo can reach 800 pmol/eye (Parish et al. 1998). This level of BisRets in the RPE cells can be obtained in vitro by incubating cells with 15–30  $\mu\text{M}$  A2E (Sparrow et al. 1999; Roberts et al. 2002; Lakkaraju et al. 2007). The RPE cells from donors older than 50 years have significantly higher concentration of A2E (2–3 times more) compared to young donors (Bhosale et al. 2009). Due to chemical structure, A2E can exhibit dark toxicity, acting as an amphiphilic detergent capable of destroying phospholipid membranes (Eldred and Lasky 1993; De and Sakmar 2002; Lakkaraju et al. 2007; Sokolov et al. 2007; Dontsov et al. 2012) and induce cell apoptosis in the dark (Suter et al. 2000). A2E exposed to blue light sensitizes lipid peroxidation (Dontsov



**Fig. 5** Photoinduced peroxidation of bovine photoreceptor outer segments (POS) with non-irradiated and pre-irradiated bisretinoid A2E. TBA (thiobarbituric acid) — active products. The concentration of A2E was 100  $\mu\text{M}$ . The samples were irradiated with visible light (390–700 nm) with an energy of 100 mW/cm<sup>2</sup>. Photobleached A2E was prepared by irradiating the original bisretinoid with an LED source (wavelength 450 nm) for 60 min. Concentration of POS was 0.2 mg of rhodopsin per 1 mL. Figure was modified from Dontsov et al. (2012)

et al. 2005, 2012, 2016), causes destruction of lysosomal membranes (Schutt et al. 2000a, b), and inhibits mitochondrial cytochrome oxidase (Suter et al. 2000). The mechanism of the phototoxic action of A2E is associated with its ability to photoproduce ROS. It is known that A2E can photogenerate superoxide radicals (Pawlak et al. 2002, 2003; Gaillard et al. 2004; Broniec et al. 2005) and singlet oxygen (Ragauskaitė et al. 2001; Cantrell et al. 2001; Pawlak et al. 2003). However, the efficiency of these processes is not very high compared to other RPE chromophores (all-*trans* retinal, protoporphyrins) (Bynoe et al. 1998; Pawlak et al. 2003; Maeda et al. 2009; Wielgus et al. 2010). Moreover, A2E is a much less potent sensitizer than LGs in the RPE cells (Rożanowska et al. 1998; Pawlak et al. 2002; Boulton et al. 2004; Dontsov et al. 2005). Apparently, the higher photosensitizing activity of LGs is associated with the presence of other photoreactive substances in them.

A2E in the presence of oxygen is easily oxidized by irradiation with the formation of numerous oxidation products (see below). Photobleached A2E loses its ability to enhance photoinduced lipid peroxidation (Fig. 5).

Figure 5 demonstrates that non-irradiated A2E is a much more effective sensitizer of photooxidation of the outer segments of photoreceptor cells than photobleached A2E. As is known, in the RPE cells, in addition to lipofuscin-containing granules, there are melanin-containing organelles — melanosomes. If the former exhibit photosensitizing properties, enhancing the production of ROS when exposed to blue light, then melanosomes, on the contrary, serve to protect

the RPE cells from the damaging effects of light and ROS (Ostrovsky et al. 1987; Wang et al. 2006a, b; Ostrovsky and Dontsov 2019). Age-related changes occurring in the RPE cells contribute to the enhancement of photooxidative processes induced by LGs. This is mainly due to an increase in the number of lipofuscin-containing granules in the RPE cell, as well as a significant drop in the concentration of melanin (Feeney–Burns et al. 1984, 1990; Sarna et al. 2003; Yacout et al. 2019). This leads to weakening of the protective effect of melanin, which includes screening of photosensitive cell structures from excess light (Ostrovsky et al. 2018), antioxidant and antiradical protection (Ostrovsky et al. 1987; Wang et al. 2006a, b; Ostrovsky and Dontsov 2019), as well as the binding of BisRets into inactive complexes (Dontsov et al. 2013; Sakina et al. 2013). A decrease in the effectiveness of antioxidant protection of melanin in the RPE cells with age is apparently associated with its photooxidative destruction in melanolipofuscin granules under the action of superoxide radicals and the formation of degradation products that do not have antioxidant activity and, on the contrary, exhibit prooxidant properties (Zareba et al. 2006; Dontsov et al. 2017; Mahendra et al. 2020; Olchawa et al. 2021).

### Cytotoxic properties of LGs in the RPE cell

In the presence of oxygen, LG BisRets themselves can be photo-oxidized to form various products, consisting primarily of epoxides, peroxides, aldehydes, and ketones, which are potentially cytotoxic (Wu et al. 2010; Ben-Shabat et al. 2002; Feldman et al. 2015; Sparrow et al. 2012; Yakovleva et al. 2006; Dontsov et al. 2009; Yoon et al. 2012). The cytotoxic properties of BisRets-OX in LGs have not been fully investigated. The role of these compounds in pathological processes of the RPE remains controversial. Some studies have suggested that highly reactive cytotoxic carbonyl compounds, aldehydes and ketones, are formed during photooxidation of BisRets in LGs (Schütt et al. 2000; Sparrow et al. 2000; Wang et al. 2006a, b). By contrast, other studies (Murdaugh et al. 2010, 2011) suggest that BisRet-OX interacts with itself or with A2E, forming products with a higher molecular weight inside LGs. Most of these compounds are also hydrophobic and remain inside LGs, resulting in the concomitant diminution of its reactivity *in vivo*. Clarifying the roles of BisRets-OX is important to delineate the mechanisms of pathological ocular diseases, especially AMD.

Our prior findings have demonstrated that LG BisRet-OX content is higher in AMD eyes than in normal eyes, which was indicated by changes in the characteristics of LG fluorescence spectra and in the parameters of fluorescence decay kinetic curves (Feldman et al. 2018). Specifically, the fluorescence excitation at 488 nm of samples

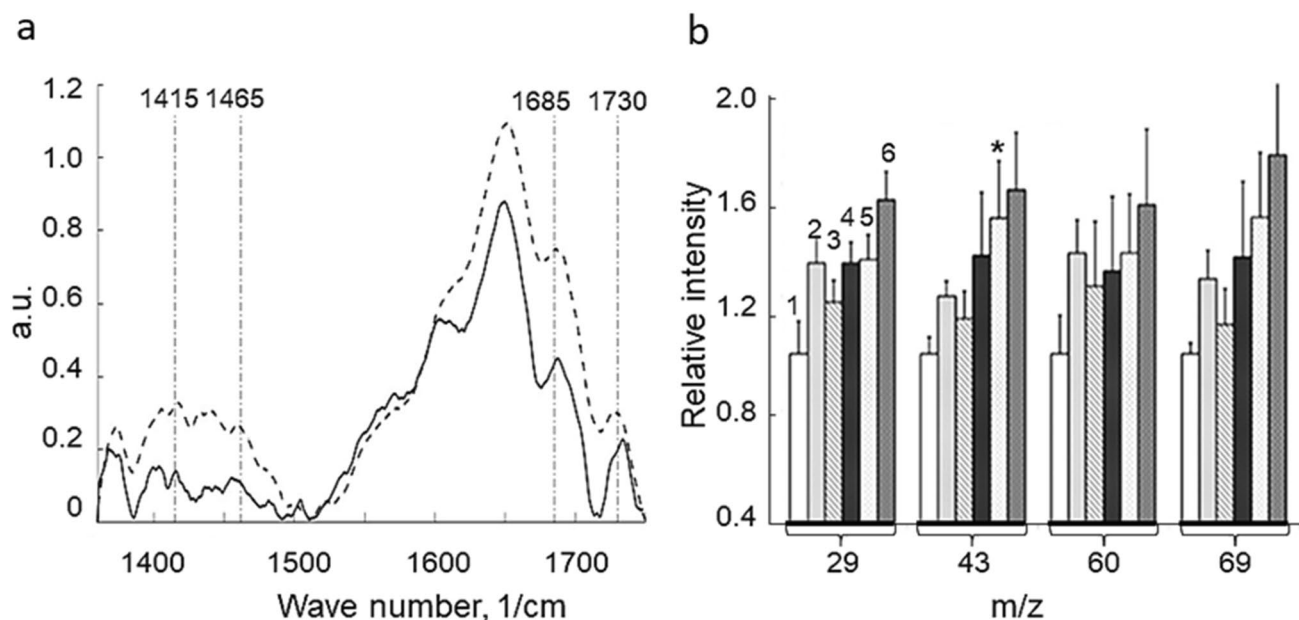
from eyes with AMD increases the fluorescence intensity of the band at 556 nm, and the contribution of BisRets-OX to total fluorescence increases. However, the pathophysiological, or protective, properties of these products remain controversial, as prior studies have suggested conflicting roles (Murdaugh et al. 2010, 2011), and as BisRets-OX could also potentially become a neutral product eventually. Investigating the potential release of BisRets-OX from LGs into the RPE cell cytoplasm and the assessment of their toxicity to cellular structures is thus fundamental to understanding the pathogenesis of retinal diseases.

The chemical characteristics of BisRets-OX in LGs, which were obtained from healthy donor eyes, have been studied (Yakovleva et al. 2022a). Raman spectroscopy and Time-of-Flight secondary ion mass spectrometry (ToF–SIMS) analysis identified the presence of free-state aldehydes and ketones within LGs (Fig. 6). It has been shown, that these substances are formed as a result of Bis-Ret photooxidation and can accumulate in LGs. This is consistent with prior findings (Wu et al. 2010; Wang et al. 2006a, b).

Together, fluorescence spectroscopy, high-performance liquid chromatography, and mass spectrometry revealed that BisRets-OX have both hydrophilic and amphiphilic properties, allowing their diffusion through LG membrane into the RPE cell cytoplasm (Yakovleva et al. 2022a). These products contain cytotoxic carbonyls, which are thiobarbituric acid (TBA)-active products (Fig. 7).

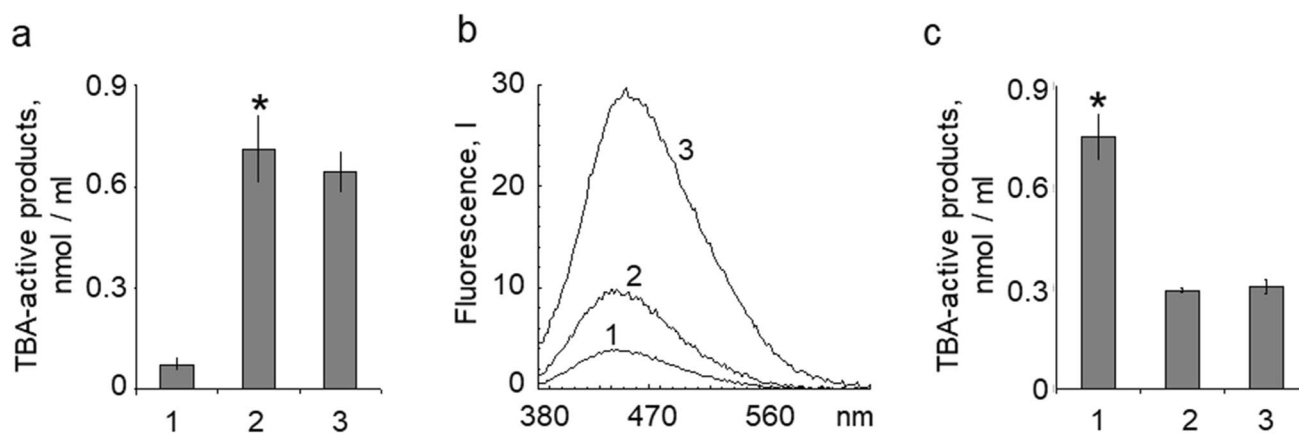
There are a number of works (Schutt et al. 2003; Ye et al. 2016; Hyttinen et al. 2018; Rózanowska and Rózanowski 2022) where it was shown that the source of TBA-active products in LGs is lipid peroxidation end-products *i.e.*, highly reactive electrophilic aldehydes like malondialdehyde (MDA) and 4-hydroxynonenal (HNE). Some recent studies have shown that LG BisRets are involved in the development of photoinduced glycative stress, not only in RPE tissue, but also in adjacent tissues, in particular, in Bruch's membrane (Zhou et al. 2015; Thao et al. 2014). It was suggested that the development of glycative stress in the RPE cells is largely associated with the photooxidative destruction of LG BisRets, leading to the formation of water-soluble reactive carbonyls which are extremely cytotoxic molecules (Schleicher et al. 2001). They are assumed to be the main precursors of advanced glycation end products (AGEs) formation (Rowan et al. 2018; Lin et al. 2016) and can be formed by direct oxidative decay of BisRets (Kim et al. 2021). In our work (Dontsov et al. 2022), we have shown that water-soluble carbonyl compounds formed during A2E photooxidation cause modification of serum albumin and hemoglobin. The antiglycation agent, aminoguanidine, has inhibited the process of protein modification. It is assumed that these carbonyl products can initiate the inflammatory processes in the retina and RPE.

It should be noted that BisRets-OX can be formed not only during BisRet photo-oxidation, but also during the



**Fig. 6** **a** Averaged Raman spectra for LG suspensions before (solid line) and after (dotted line) visible light irradiation for 100 min. Three independent experiments were conducted. In each experiment, 25 spectra were obtained ( $p < 0.05$ ). **(b)** ToF-SIMS analysis of LG suspensions before irradiation (1) and after visible light irradiation for 2 (2), 10 (3), 40 (4), 100 (5), and 160 (6) min. On the abscissa axis, the

numbers correspond to the mass of positive fragment ions, containing carbonyl groups (29: CHO+; 43: C<sub>2</sub>H<sub>3</sub>O+; 60: C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>+; 69: C<sub>4</sub>H<sub>5</sub>O+). On the ordinate axis, the relative intensities of the corresponding positive fragment ions are plotted as relative units. Data are presented as means  $\pm$  SD from nine independent experiments. \*  $p < 0.01$  (Yakovleva et al. 2022a)



**Fig. 7** **a** Concentrations of TBA-active products in samples of the supernatants from the original LG suspension (1), LG suspension irradiated with visible light for 60 min (2), or LG suspension oxidized by superoxide radicals (3). **(b)** Fluorescence spectra of the supernatants (the spectrum numbers correspond to the sample numbers in

panel A). The fluorescence excitation wavelength was 365 nm. **(c)** Supernatant content of TBA-active products from LG suspension irradiated with visible light for 90 min (1), and in the aqueous (2) and chloroform (3) fractions. Data are presented as means  $\pm$  SD from three independent experiments. \*  $p < 0.05$  (Yakovleva et al. 2022a)

oxidative destruction of BisRets in the dark via non-lipo-fuscin ROS (Yakovleva et al. 2022a, b). Thus, a significant change in the fluorescent properties of retinoids in the retina and RPE from mouse eye exposed to ionizing radiation (IR) was detected (Yakovleva et al. 2022b). Such changes occur when retinoids are oxidized, suggesting that IR induce oxidation and degradation of retinoids, similar to photo-oxidation

of BisRets in the human RPE (Feldman et al. 2015, 2018). However, in the case of IR, in the absence of light, the source of ROS is different. For example, these can be ROS-generated by mitochondria, caused by IR, or water radiolysis (Kobashigawa et al. 2011; Azzam et al. 2012; Belli and Indovina 2020). Thus, LGs can have a damaging effect on the RPE cell through the BisRets-OX formation. Therefore,

BisRets-OX are a likely aggravating factor in the progression of various senile eye pathologies.

The RPE cells are constantly exposed to (photo)oxidative stress. This is facilitated by high oxygen consumption and prolonged exposure to light (Beatty et al. 2000). An important factor contributing to the increase in oxidative stress in the RPE cells is the progressive age-related accumulation of LGs (Feeney–Burns et al. 1984; Wing et al. 1978; Delori et al. 2001) and a decrease in the content of melanosomes (Schmidt and Peisch 1986; Sarna et al. 2003; Dontsov et al. 2017; Yacout et al. 2019). These processes lead to the appearance of damaged and modified cellular proteins, lipids, and DNA (Kohen and Nyska 2002). Oxidative stress and inflammation are of great importance in the development of degenerative processes in the RPE cells. So, it is believed that oxidative stress plays a central role in the development of AMD (Beatty et al. 2000; Datta et al. 2017; Abokyi et al. 2020; Ruan et al. 2021). It has been shown that in the RPE cells from donor eyes with AMD, compared with normal eyes, there are increased levels of TBA-active products, protein carbonyls (Totan et al. 2009), a high content of carboxyethylpyrrole in Bruch's membrane (Crabb et al. 2002; Lu et al. 2009), as well as oxidative damage and dysfunction of mitochondria (Terluk et al. 2015; Blasiak et al. 2013; Golestaneh et al. 2018). Also, in AMD, the accumulation of damaged proteins and disruption of the autophagy process are noted (Mitter et al. 2014). The modified and damaged proteins formed as a result of oxidative stress, which are incapable of repair by heat shock proteins, are directed to the proteasome for purification. However, if the activity of the proteasome is reduced (Zhang et al. 2008; Fernandes et al. 2008), proteins aggregate and can be degraded by autophagy (Fig. 8).

Autophagy is also used by the RPE cells to digest damaged mitochondria (mitophagy). A decrease in the activity of lysosomal enzymes as a result of oxidative stress (Brunk et al. 1995; Nilsson et al. 2003; Sparrow and Boulton 2005) leads to inhibition of the autophagy process. ROS also destroy the integration relationship with proteasomes and autophagy, which ultimately leads to increased accumulation of toxic aggregates, development of chronic inflammation, activation of the complement system, formation of extracellular drusen, and death of the RPE cells (Kinnunen et al. 2012; Ferrington et al. 2016; Moreno-García et al. 2018).

### Fluorescence characteristics of LGs as a tool to detect early stages of degeneration in the retina and RPE

LGs exhibit distinct fluorescence in the visible region. BisRets and BisRets-OX are major sources of LG fluorescence. Fundus autofluorescence (FAF) imaging is a noninvasive, prospective diagnostic method based on the

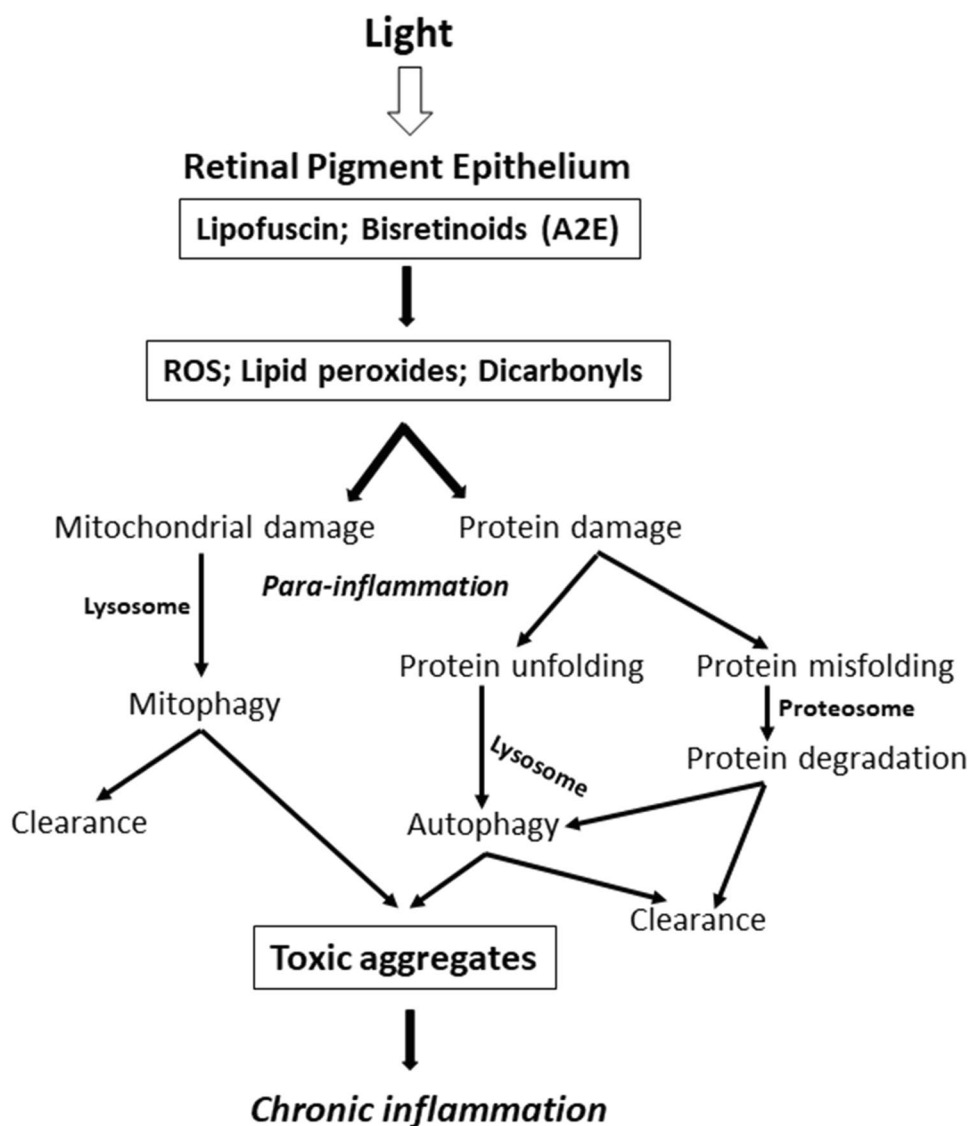
detection of LG fluorescence in the RPE cells (Von Ruckmann et al. 1997; Holz et al. 2007; Sparrow et al. 2010a, b). FAF is excited by a wavelength of 488 nm, yielding monochromatic images in the long-wavelength region, starting at 500 nm, resulting from the fluorescence of BisRets and BisRets-OX. FAF can detect early phenotypic changes in RPE, occurring prior to the progression of disease. Analysis of FAF patterns can provide detailed qualitative information that allows the detection of areas of pathology, thereby differentiating among different types of ocular disease. At present, however, it is not possible to quantify the detected changes. The degree of disease progression must be assessed subjectively by an expert, who compares patterns with those of normal eyes.

Efforts are underway to expand the capabilities of this diagnostic method, based on knowledge about the spectral characteristics of LG fluorophores. Increased blue-green autofluorescence of the Bruch's membrane, relative to the yellow-orange autofluorescence of RPE-associated lipofuscin, is associated with AMD (Marmorstein et al. 2002). In addition, with the use of fluorescence lifetime imaging ophthalmoscopy (FLIO) *in vivo*, healthy eyes were shown to exhibit different patterns than those of AMD eyes (Sauer et al. 2018a, b, c; Schweitzer et al. 2009). Specific patterns were also detected for retinitis pigmentosa (Andersen et al. 2018; Dysli et al. 2018), Stargardt disease (Dysli et al. 2016), macular telangiectasia type 2 (MacTel) (Sauer et al. 2018a, b, c), and other diseases (Sauer et al. 2018a, b, c). Changes from normal fluorescence parameters have also been observed in the eyes of patients with diabetes (Schweitzer et al. 2015) and Alzheimer's disease (Jentsch et al. 2014; Sadda et al. 2019). However, it should be noted that none of these publications have explained the nature or underlying mechanism of these differences. Nevertheless, these findings, especially in AMD, were confirmed by experimental research, which has demonstrated that the quantitative and qualitative properties of LG fluorophores change during pathologic development (Feldman et al. 2015, 2018; Wu et al. 2010).

The main drawback of these experimental studies is the inability to analyze LG fluorophore composition *in vivo*, as more than 20 fluorophores, bisretinoids, and their derivatives have been identified to date. Nevertheless, one of the main objectives of *in vivo* FAF and FLIO methods is to determine differences in the fluorescence characteristics of the fundus between individuals with ocular pathology and those with healthy eyes.

Recently, we have shown that the content of BisRets-OX increases with the development of AMD (Feldman et al. 2018). Because, the fluorescence can show the accumulation of BisRets-OX in LGs, therefore BisRets-OX may be an indicator of AMD progression. Changes in the quantitative and qualitative composition of fluorophores and their

**Fig. 8** Schematic illustration of LG role in the development of photooxidative stress in the RPE cell



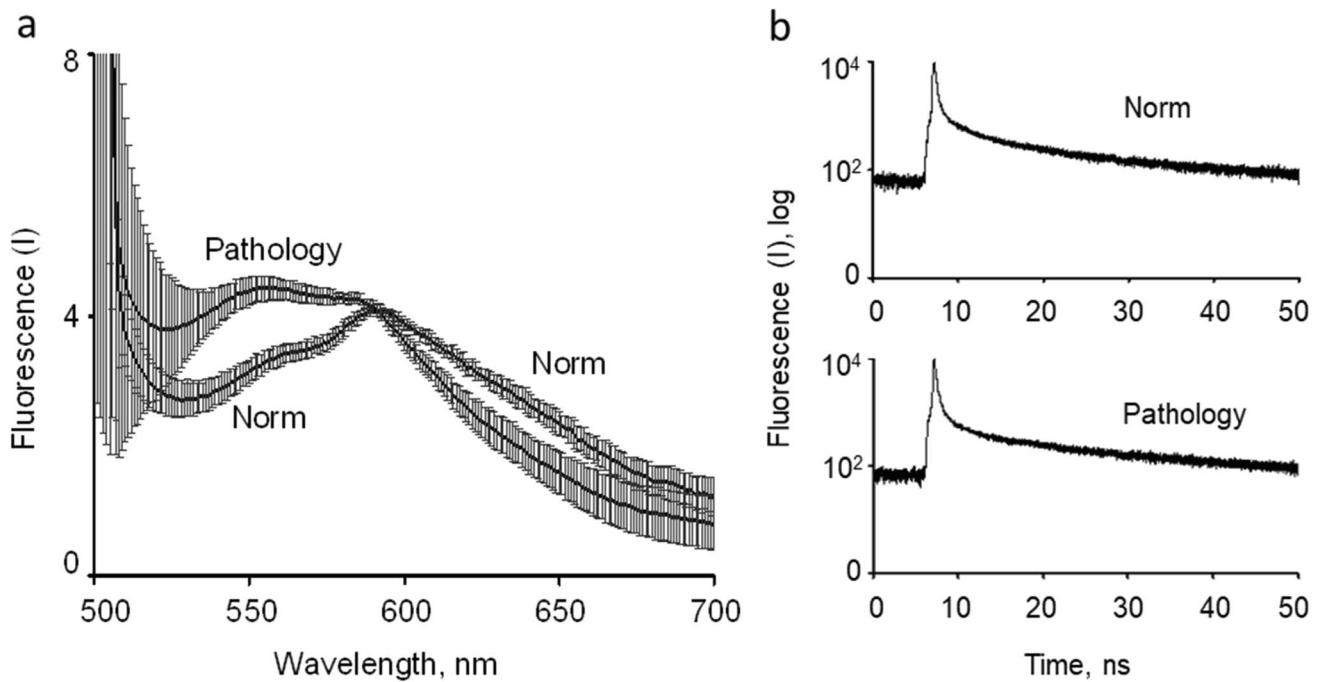
spectral characteristics are determined only by the presence or absence of pathological changes in the RPE, but are independent of age. These patterns have characteristic and reproducible features that can be used as diagnostic indicators of visual pathology (Fig. 9a).

Moreover, the fluorescence lifetimes were measured by picosecond-resolved time correlated single photon counting technique (Feldman et al. 2018; Yakovleva et al. 2020) (Fig. 9b). It was shown that BisRets-OX exhibited a longer fluorescence lifetime (average value approximately 6 ns) and a shorter wavelength maximum (530–580 nm). Further, these products significantly contributed (more than 30%) to total fluorescence compared to the other fluorophores in LGs. Thus, the contribution of BisRets-OX to autofluorescence decay kinetics is an important characteristic for fluorescence lifetime imaging microscopy data analysis (Feldman et al. 2018; Yakovleva et al. 2020).

Based on the data obtained, we can deduce that the specific pattern observed in AMD eyes in vivo using FLIO could be due to accumulation of BisRets-OX in LGs (Sauer et al. 2018a, b, c; Schweitzer et al. 2009). Thus, we suggest this as an additional approach to expand the diagnostic capabilities of the FLIO method. Instead of using the averaged fluorescence lifetimes for different eye tissues with fluorescence excitation at 468 nm (Schweitzer et al. 2007), we propose to focus only on LG fluorophores from the RPE with fluorescence excitation at 488 nm (similar to the FAF method) in order to determine the contribution of BisRets-OX to total fluorescence or determine the averaged fluorescence lifetime, because the higher BisRet-OX content in the RPE from AMD eyes increased the average fluorescence lifetime (Feldman et al. 2018; Yakovleva et al. 2020).

Earlier, we have shown that LG BisRets can be oxidized not only in the presence of light, but also by ROS of





**Fig. 9** **a** Comparative statistical analysis of spectral characteristics of the RPE cell suspensions from cadaver eyes without (Norm) and with (Pathology) signs of AMD. Fluorescence spectra were averaged for 19 normal eyes (from donors aged 27–74 years), and for 12 AMD eyes (from donors aged 59–88 years). The excitation wavelength was 488 nm, with emission spectra normalized at 592 nm. **(b)** Fluorescence decay kinetic curves (normalized) of LG fluorophores

of the RPE cells from human cadaver eyes. The samples of the RPE cell suspensions were from normal eyes (Norm) from a 74-year-old donor, and the other from eyes with AMD (Pathology) from a 75-year-old donor. Fluorescence was excited at 485 nm (pulse duration, 30 ps), and kinetic curves were recorded at 540 nm. Figure was modified from Feldman et al. (2018)

a non-lipofuscin nature (Yakovleva et al. 2022a, b). In other words, the spectral properties of LG fluorophores can be markers of oxidative stress which can initiate degenerative processes in the retina and RPE. Thus, our findings contribute to progress in the creation of rapid testing of the oxidative stress development in living organisms, and contribute to development of a predictive criterion for increased AMD risk in future.

Thus, there is a possibility to improve the FAF and FLIO techniques to obtain additional information from the total fluorescence patterns. Quantitative determination of increases in BisRets-OX in LGs may be used to establish quantitative diagnostic criteria for degenerative processes in the retina and RPE even in the absence of visible manifestation of the disease.

## Conclusion

Lipofuscin was discovered by Virchow R. as early as 1847, and the term “lipofuscin” itself has been used since 1912. Earlier, lipofuscin was traditionally referred to as intracellular and inert “slag” as a marker of aging. It is known that the progressive accumulation of lipofuscin can somehow lead

to pathology and accelerate the aging process. However, the mechanism of pathogenetic action of lipofuscin remained unknown until recently.

The nature of the lipofuscin accumulation in cells is associated with the destruction of cell organelles that have not been utilized by lysosomes. In the case of the RPE, these are mainly nondigested fragments of phagocytosed outer segments of photoreceptors. The accumulation of LGs is explained by the absence of enzymes in the cell that can degrade it. So far, there are no effective ways to both slow down the LG accumulation in the cell and remove it from the cell. Despite some encouraging experimental results on the removal of LGs from RPE cells, they have not reached clinical use.

Significant interest in the pathogenetic role of lipofuscin arose in the early nineties after we discovered the photoactivity of lipofuscin granules isolated from human cadaver eyes (Ostrovsky et al. 1992; Boulton et al. 1993). It turned out that LGs are not an inert “slag”, but extracellular structures capable of ROS generating under the action of visible light.

Approximately, at the same time and later, active development and improvement of optical methods for recording fundus autofluorescence began. It was about short-wavelength

autofluorescence as a non-invasive diagnostic method for detecting LGs, a by-product of the visual cycle, which accumulates in the RPE cells with age or disease.

Over the past decades, we have conducted research both in the direction of the action of light on LGs, and in the direction of changes in the spectral characteristics of the LGs fluorescence in pathology and under the action of ionizing radiation. The results of these studies significantly expanded our understanding of both the photoactivity and fluorescence of LGs and the fluorophores (bisretinoids) contained in them. The main results of these studies are as follows.

First, our studies of the structure and fluorescence properties of LGs by AFM and near-field microscopy revealed the fluorescent heterogeneity of LGs. This means that twenty or more LG fluorophores are unevenly distributed in the granule. It would of course be interesting to compare this distribution in non-photooxidized (normal) and photooxidized (pathological) LGs.

Secondly, our study of the phototoxicity of LGs which are photo-inducible generators of ROS, showed the following. LGs and melanolipofuscin granules of human RPE cells are capable of photoinduced oxidation of lipids. At the same time, LGs are more active than melanolipofuscin granules. Another rather unexpected result is that A2E, as the main fluorophore (BisRet) of LGs capable of photogeneration of ROS, was much less active as an oxidation photosensitizer than other fluorophores (BisRets) of LGs. In this regard, it would be important to establish which BisRet or group of BisRet fluorophores represent the greatest danger in terms of phototoxicity. Fundamentally important for understanding the mechanisms of LG phototoxicity is the fact that BisRets-OX pose a much greater danger to the cell than, non-oxidized products. Moreover, the BisRets-OX formed in the dark as a result of BisRet oxidative degradation also have significant toxicity for the cell.

Thirdly, our studies of the fluorescent properties of LGs and their fluorophores (BisRets and BisRets-OX) showed for the first time that their spectral characteristics change, namely, they shift to the short-wavelength region, in pathology (AMD) and after exposure to ionizing radiation (gamma-rays and protons).

It is important to emphasize the promise of recording not only changes in the spectra, but also the decay kinetics of FAF for the early diagnosis of degenerative diseases of the retina. These changes are associated precisely with the formation and accumulation of LG BisRets-OX. In other words, by recording the fluorescence decay kinetics of oxidized products of LG BisRets-OX, it is possible to significantly improve the methods of early diagnosis of degenerative diseases, primarily AMD.

Thus, there is no need to talk about any inertness of lipofuscin (“aging pigment”) as an inert “slag”. The photoactivity of lipofuscin can pose a significant danger to

the RPE cells. Based on this, it is necessary to observe the well-known light hygiene measures for the senile and diseased eye (sunglasses, colored intraocular lenses, etc.). The search for pharmacological agents that prevent the toxicity of LG fluorophores (BisRets) and their oxidation products is extremely relevant. It would also be very important to find a drug capable of delaying the formation of LGs into the cells or removing (destroying) LGs from the cell.

**Author contribution** All authors had the idea for the article and performed the literature search and data analysis; T.B. Feldman, A.E. Dontsov and M.A. Ostrovsky drafted the work. All authors read and approved the final manuscript.

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## Declarations

**Conflict of interest** The authors declare no competing interests.

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