REVIEW

The protein disorder cycle

Vladimir N. Uversky[1](http://orcid.org/0000-0002-4037-5857)

Received: 16 August 2021 / Accepted: 13 October 2021 / Published online: 3 November 2021 © International Union for Pure and Applied Biophysics (IUPAB) and Springer-Verlag GmbH Germany, part of Springer Nature 2021

Abstract

This mini-review represents a brief, disorder-centric consideration of the interplay between order and disorder in proteins. The goal here is to show that inside the cell, folding, non-folding, and misfolding of proteins are interlinked on multiple levels. This is evidenced by the highly heterogeneous spatio-temporal structural organization of a protein molecule, where one can fnd diferently (dis)ordered components that can undergo local or global order-to-disorder and disorder-to-order transitions needed for functionality. This is further illustrated by the fact that at particular moments of their life, most notably during their synthesis and degradation, all proteins are at least partially disordered. In addition to these intrinsic forms of disorder, proteins are constantly facing extrinsic disorder, which is intrinsic disorder in their functional partners. All this comprises the multileveled protein disorder cycle.

Keywords Intrinsically disordered protein region · Protein function · Nascent polypeptide chain · Protein degradation · Protein biosynthesis · Protein folding · Protein misfolding

The discovery of the intrinsic disorder phenomenon and successful penetration of this concept into protein science indicated a departure from the classical description of a protein function in terms of the "lock-and-key" model. Accumulated evidence indicates that protein intrinsic disorder is ubiquitous and inexorable, and the presence of structure-less but biologically active proteins dramatically expanded the universe of functional proteins in such a way that sometimes it may seem that the grinning faces of intrinsically disordered proteins (IDPs) and proteins with IDP regions (IDPRs) peek out from every corner of modern protein science. Altogether, there is no doubt now that the fate of a newly synthesized polypeptide chain is not limited to the functional folding and assembly or pathological misfolding, as was believed for a long time, but also includes a very important "non-folding" branch (see Fig. [1\)](#page-1-0). The choice between these pathways is determined by the peculiarities of the protein amino acid sequence and its environment (Uversky and Uversky [2014](#page-6-0)).

However, none of the indicated pathways (folding, nonfolding, or misfolding) represent a one-way road leading to

 \boxtimes Vladimir N. Uversky vuversky@usf.edu

a dead-end. Instead, inside cells, proteins are involved in a constant circulation between folded, non-folded, and misfolded forms, where already folded proteins can undergo at least partial unfolding and misfold or get (partially) unfolded for new functionality; where misfolded proteins can return back to their original folded or non-folded states; where IDPs/IDPRs can partially fold or misfold; and where such transitions can be repeated several times (Uversky [2003\)](#page-7-0). In other words, there is no folk-tale crossroad stone stating, "If you go left, you will lose your horse; if you go right, you will lose your life; if you go straight, you will live, but you will forget yourself," and the cellular fate of a protein is not predefned but can be rewritten (and multiple times at that). Let us examine this folding-non-folding-misfolding cycle by taking a closer look at the intrinsic disorder (non-folding) part.

The last two decades witnessed a triumph of the idea that protein functionality can be independent of the unique structure (Turoverov et al. [2010](#page-6-1); Dyson [2011](#page-5-0); Tompa [2011,](#page-6-2) [2012;](#page-6-3) Uversky [2013a](#page-7-1); van der Lee et al. [2014](#page-7-2)). IDPs and hybrid proteins with ordered domains and IDPRs are abundantly present not only in all proteomes analyzed so far (Uversky et al. [2000;](#page-7-3) Dunker et al. [2000](#page-4-0); Ward et al. [2004](#page-7-4); Schad et al. [2011](#page-6-4); Xue et al. [2012](#page-6-5); Pancsa and Tompa 2012; Peng et al. [2013a](#page-6-6), [2015](#page-6-7)) but even within the preeminent source of protein structural knowledge, the protein data bank (PDB), where the vast majority of proteins with resolved

¹ Department of Molecular Medicine and Byrd Alzheimer's Research Institute, Morsani College of Medicine, University of South Florida, Tampa, FL 33612, USA

Fig. 1 Schematic representation of the fate of a polypeptide chain in the cell. In addition to the folding-misfolding-nonfolding crossroad stone that seems to defne the cellular fate of a protein based on the peculiarities of its amino acid sequence, the model shows that this cellular fate of a protein is not pre-defned but can be rewritten, thereby generating a perpetual folding-non-foldingmisfolding cycle

X-ray structures have unobserved regions of varied length with missing electron density that frequently correspond to IDPRs (Le Gall et al. [2007](#page-6-8); Monzon et al. [2020](#page-6-9)). Furthermore, in PDB, many protein–protein and protein-nucleic acid complexes with stable structures are in fact formed by IDPs/IDPRs as a result of the disorder-to-order transition (Gunasekaran et al. [2004;](#page-5-1) Oldfeld et al. [2008;](#page-6-10) Wu et al. [2015](#page-7-6); Zhou et al. [2020](#page-7-7)).

The levels of IDPs/IDPRs in proteomes are correlated with the evolutionary complexity of the organisms, where the more advanced species have higher IDP/IDPR counts (Dunker et al. [2000](#page-4-0), [2015;](#page-5-2) Ward et al. [2004;](#page-7-4) Xue et al. [2012](#page-7-5); Peng et al. [2013a,](#page-6-6) [2015\)](#page-6-7). The functional range of these proteins is remarkably broad and complements functions of ordered proteins and domains (Wright and Dyson [1999](#page-7-8); Dunker et al. [2001,](#page-4-1) [2002a,](#page-5-3) [b](#page-5-4), [2005,](#page-5-5) [2008a,](#page-5-6) [b](#page-5-7); Dunker and Obradovic [2001;](#page-4-2) Dyson and Wright [2002,](#page-5-8) [2005;](#page-5-9) Uversky [2002a,](#page-6-11) [b](#page-6-12); Uversky et al. [2005](#page-7-9); Cortese et al. [2008](#page-4-3); Dunker and Uversky [2008;](#page-5-10) Oldfeld et al. [2008;](#page-6-10) Uversky and Dunker [2010](#page-7-10); van der Lee et al. [2014](#page-7-2); Oldfeld and Dunker [2014\)](#page-6-13).

Structurally, IDPs/IDPRs are characterized by remarkable spatio-temporal heterogeneity and can range from completely structure-less, coil-like conformational ensembles to compact (but still highly dynamic) molten globular ensembles, to proteins with a hybrid structure containing both ordered and disordered regions (Uversky and Dunker [2010](#page-7-10); Uversky [2013a](#page-7-1), [d,](#page-7-11) [e](#page-7-12); Dunker et al. [2013](#page-5-11)). Thus, intrinsic disorder has multiple faces afecting diferent levels of protein structural organization, where either the whole protein or its various regions can be (dis)ordered to a diferent degree. As a result, while looking at such a heterogeneous structure, one can identify fragments with diferent structural complexity and folding complicity and admire a highly dynamic and interchanging structural mosaic containing foldons (i.e., independently foldable protein units), inducible foldons (which are IDPRs capable of at least partial folding promoted by their interactions with binding partners), morphing inducible foldons (IDPRs with the potential to fold diferently due to binding to diferent partners), semi-foldons (regions that are always in a semi-folded state), and nonfoldons (IDPRs that never fold) (Uversky [2013e](#page-7-12), [2016a](#page-7-13), [b,](#page-7-14) [c,](#page-7-15) [2019a](#page-7-16), [b](#page-7-17)). Such exceptional spatio-temporal heterogeneity of IDPs/IDPRs is translated into their multifunctionality, as diferently (dis)ordered parts of a protein molecule might have diferent functions (Uversky [2015,](#page-7-18) [2016a](#page-7-13)).

IDPs/IDPRs behave as highly frustrated systems with no single thermodynamically stable state. This is refected in their free energy landscapes, which are relatively fat, do not have deep energy minima seen in the free energy landscapes of ordered globular proteins, and instead represent a hilly plateau with multiple shallow local minima corresponding to the allowed conformations separated by low hills indicating forbidden conformations (Uversky et al. [2008](#page-7-19); Turoverov et al. [2010;](#page-6-1) Fisher and Stultz [2011\)](#page-5-12). Such a fattened energy landscape is extremely sensitive to diferent environmental changes and can be modifed in a number of diferent ways. Depending on the peculiarities of the environment, some energy minima can be deepened, while some energy barriers can increase. This explains the conformational plasticity of IDPs/IDPRs, their extreme sensitivity to changes in the environment, the ability to specifcally interact with many partners of diferent nature, and to fold diferently as a result of these interactions (Uversky [2013e](#page-7-12)).

One should also remember that there is no actual boundary between order and disorder. Instead, structures of proteins can be presented as a continuous spectrum of diferently structured/disordered conformations that extends from the fully ordered to completely structure-less species, with everything in between (Uversky [2013e](#page-7-12); DeForte and Uversky [2016\)](#page-4-4). Such complex, highly heterogeneous spatiotemporal organization of the protein structural space forms the foundation of the structure–function continuum concept (Uversky [2016a,](#page-7-13) [b](#page-7-14), [2019a,](#page-7-16) [b](#page-7-17)), where instead of the classical "one gene-one protein-one structure-one function" model, any protein is considered as a highly dynamic conformational ensemble with a broad spectrum of structural features, and as such the structural heterogeneity and conformational plasticity of IDPs defne their remarkable multifunctionality and binding promiscuity (Uversky [2016b](#page-6-12), [2019b](#page-7-14); Fonin et al. [2019](#page-5-13)).

Therefore, multifunctionality of proteins is based on the constant order–disorder-order and disorder-order–disorder cycles. This is refected in the crucial dependence of the functionality of many ordered proteins on the existence of "unfoldons," i.e., ordered regions that must undergo an order-to-disorder transition to make the protein active (Uversky [2013e\)](#page-7-12), or, more generally, in a dormant (transient, conditional, or cryptic) disorder phenomenon (Jakob et al. [2014](#page-5-14); Creamer [2013](#page-4-5); Bardwell and Jakob [2012](#page-4-6)). On the other hand, the functionality of IDPs/IDPRs depends on the presence of disorder-based binding sites, called molecular recognition features (MoRFs), which are interactionprone disordered regions that can fold at binding to specifc partners (Cheng et al. [2007;](#page-4-7) Vacic et al. [2007](#page-7-20); Mohan et al. [2006;](#page-6-14) Oldfeld et al. [2005](#page-6-15)). The presence of such MoRFs (inducible foldons) or morphing MoRFs defnes the exceptional binding plasticity and promiscuity of IDPs/IDPRs (Uversky [2011b,](#page-7-21) [2013c\)](#page-7-22), where a single IDPR can bind to multiple partners gaining very diferent structures in the bound state (Oldfeld et al. [2008;](#page-6-10) Hsu et al. [2012,](#page-5-15) [2013](#page-5-16); Alterovitz et al. [2020](#page-4-8)), and explains the abundance of intrinsic disorder among hub proteins and their binding partners in various protein–protein interaction networks (Dunker et al. [2005](#page-5-5); Patil and Nakamura [2006](#page-6-16); Haynes et al. [2006;](#page-5-17) Ekman et al. [2006](#page-5-18); Dosztanyi et al. [2006;](#page-4-9) Singh et al. [2006](#page-6-17)).

Furthermore, the lack of stable structure defines the exceptional sensitivity of IDPs/IDPRs (which represent the edge of chaos systems) to even small environmental changes that can trigger very diferent structural and functional outputs, thereby serving as illustrative examples of the butterfy efect (Uversky [2013e,](#page-7-12) [2019a,](#page-7-16) [b](#page-7-17)). Finally, since the formation of the pathological aggregates, including amyloid fbrils, is critically dependent on the presence of partially folded aggregation-prone species, protein misfolding, by default, relies on the order-(partial) disorder or disorder- (partial) disorder transitions. For example, it was emphasized that amyloidogenic conformations must be relatively unfolded (but not completely unfolded or coil-like), as the inherent fexibility of such pre-molten globular intermediates defnes their capability to undergo conformational rearrangements necessary to form amyloid fbrils (Uversky and Fink [2004\)](#page-7-23). It is likely that similar conformational constrains are applicable to proteins involved in the formation of functional amyloids (Deshmukh et al. [2018](#page-4-10); Christensen et al. [2019;](#page-4-11) Jain and Chapman [2019](#page-5-19); Rubel et al. [2020;](#page-6-18) Sergeeva and Galkin [2020;](#page-6-19) Daskalov et al. [2021](#page-4-12); Levkovich et al. [2021](#page-6-20)).

Although the aforementioned examples serve as illustrations of the folding-non-folding-misfolding circle, it seems that any given protein constantly faces disorder throughout its entire life within the cell, and this disorder can be of intrinsic or extrinsic nature (i.e., intrinsic disorder in functional partners) (Uversky [2013b](#page-7-24)). Even proteins with the most ordered structures are synthesized as linear polymers that need to undergo disorder-to-order transitions to gain their functional structure. A newly synthesized nascent polypeptide chain passes through the 100-Å long, 10–28-Å-wide ribosomal exit tunnel (Gabashvili et al. [2001\)](#page-5-20), which, at the exit site, contains a ring of 7 ribosomal proteins (L4, L17, L22, L23, L24, L29, and L32) and which regulates translation via interaction of L22 with specifc nascent chains (Wilson and Nierhaus [2005\)](#page-7-25). Figure [2](#page-4-13) shows that all these proteins contain signifcant levels of disorder, as evidenced by their per-residue intrinsic disorder profles and by their unusual structural shapes, which (with the exception of L22) are not consistent with simple globular structure, suggesting that, similar to the majority of other ribosomal proteins, these proteins fold at binding (Peng et al. [2013b](#page-6-21)) and might preserve signifcant levels of disorder even in the bound state, forming so-called fuzzy complexes (Uversky [2011b](#page-7-21); Tompa and Fuxreiter [2008;](#page-6-22) Fuxreiter and Tompa [2012](#page-5-21); Sharma et al. [2015;](#page-6-23) Miskei et al. [2017;](#page-6-24) Fuxreiter [2020\)](#page-5-22).

These observations indicate that the cradle of a nascent polypeptide chain, being enriched in IDPs/IDPRs, is soft and fufy. Even chaperones and nanny-proteins, which guard a newly synthesized polypeptide chain before it properly folds and matures, are IDPs or hybrid proteins containing ordered domains and functional IDPRs (Tompa

◆ Fig.2 Order and disorder in ribosomal proteins L4, L17, L22, L23, L24, L29, and L32 forming the ring at exit site of the ribosomal exit tunnel. The left panels represent intrinsic disorder profles generated by these proteins by a set of commonly used disorder predictors, whereas the corresponding structures of these proteins modeled by AlphaFold (Jumper et al. [2021](#page-5-31)) are shown on the right side. Disorder profles were assembled by the DiSpi web crawler that aggregates the results from PONDR® VLXT (Romero et al. [2001\)](#page-6-32), PONDR® VL3 (Peng et al. [2006](#page-6-33)), PONDR® VLS2B (Peng et al. [2005](#page-6-34)), PONDR® FIT (Xue et al. [2010\)](#page-7-29), IUPred2 (Short) and IUPred2 (Long) (Dosztányi et al. [2005;](#page-4-14) Meszaros et al. [2018](#page-6-35)). The outputs of the evaluation of the per-residue disorder propensity (PRDP) by these tools are represented as real numbers between 1 (ideal prediction of disorder) and 0 (ideal prediction of order). Thresholds of 0.15 ≤ PRDP < 0.5 and PRDP ≥ 0.5 were used to identify flexible and disordered residues and regions. AlphaFold-predicted structures are colored based on the per-residue confdence scores (pLDDT) that range between 0 and 100: blue, cyan, yellow, and orange colors correspond to regions with very high (pLDDT>90), high (90>pLDDT>70), low $(70 > pLDDT > 50)$, and very low (pLDDT < 50) per-residue confidence scores. Some regions with low pLDDT may be unstructured in isolation

and Csermely [2004;](#page-6-25) Kovacs et al. [2009](#page-5-23); Tompa and Kovacs [2010;](#page-6-26) Uversky [2011a](#page-7-26); Tsvetkov et al. [2009](#page-6-27); Kovacs and Tompa [2012\)](#page-5-24). In its mature form, a protein is placed within the aforesaid perpetual folding-non-folding-misfolding circle, and even on its deathbed it faces disorder. In fact, both proteolytic digestion and proteasomal degradation are dependent on intrinsic disorder. Proteolysis of disordered substrates by numerous proteases is extremely fast (Dunker et al. [2001](#page-4-1); Iakoucheva et al. [2001;](#page-5-25) Fontana et al. [1986](#page-5-26), [1997a,](#page-5-27) [b,](#page-5-28) [1993](#page-5-29)), and ATP-dependent, active unfolding of protein substrates (likely by mechanical pulling of the polypeptide chain into their channel) represents a crucial functional step of proteasomes and their prokaryotic and archaeal analogues (Weber-Ban et al. [1999](#page-7-27); Kim et al. [2000](#page-5-30); Van Melderen et al. [1996](#page-7-28); Navon and Goldberg [2001;](#page-6-28) Lee et al. [2001](#page-6-29); Prakash and Matouschek [2004](#page-6-30); Prakash et al. [2004\)](#page-6-31).

In conclusion, intrinsic disorder represents a crucial part of the functional and dysfunctional life of any given protein, and no single protein can avoid some form of disorder (intrinsic, induced, or extrinsic) during its lifetime. In fact, some proteins are always at least partially disordered. Others, being mostly ordered, possess transient disorder and have to undergo at least partial unfolding to become functional. The functions of many proteins depend on extrinsic disorder, as these proteins are controlled, regulated, and activated via utilization of the functional intrinsic disorder of their partners. Furthermore, the birth and death of all proteins are inevitably associated with disorder, since protein biosynthesis involves generation of a linear polypeptide chain and proteasomal degradation relies on the induced unfolding of a degradation target.

Funding None

Declarations

Conflict of interest The author declares no competing interests.

Ethical approval Not applicable

References

- Alterovitz WL, Faraggi E, Oldfeld CJ, Meng J, Xue B, Huang F, Romero P, Kloczkowski A, Uversky VN, Dunker AK (2020) Many-to-one binding by intrinsically disordered protein regions. Pac Symp Biocomput 25:159–170. [https://doi.org/10.1142/97898](https://doi.org/10.1142/9789811215636_0015) [11215636_0015](https://doi.org/10.1142/9789811215636_0015)
- Bardwell JC, Jakob U (2012) Conditional disorder in chaperone action. Trends Biochem Sci 37(12):517–525. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.tibs.2012.08.006) [tibs.2012.08.006](https://doi.org/10.1016/j.tibs.2012.08.006)
- Cheng Y, Oldfeld CJ, Meng J, Romero P, Uversky VN, Dunker AK (2007) Mining alpha-helix-forming molecular recognition features with cross species sequence alignments. Biochemistry 46(47):13468–13477. <https://doi.org/10.1021/bi7012273>
- Christensen LFB, Schafer N, Wolf-Perez A, Madsen DJ, Otzen DE (2019) Bacterial amyloids: biogenesis and biomaterials. Adv Exp Med Biol 1174:113–159. [https://doi.org/10.1007/](https://doi.org/10.1007/978-981-13-9791-2_4) [978-981-13-9791-2_4](https://doi.org/10.1007/978-981-13-9791-2_4)
- Cortese MS, Uversky VN, Dunker AK (2008) Intrinsic disorder in scaffold proteins: getting more from less. Prog Biophys Mol Biol 98(1):85–106. [https://doi.org/10.1016/j.pbiomolbio.2008.](https://doi.org/10.1016/j.pbiomolbio.2008.05.007) [05.007](https://doi.org/10.1016/j.pbiomolbio.2008.05.007)
- Creamer T (2013) Transient disorder: calcineurin as an example. Intrin Disordered Proteins 1(1):e26412.<https://doi.org/10.4161/idp.26412>
- Daskalov A, El Mammeri N, Lends A, Shenoy J, Lamon G, Fichou Y, Saad A, Martinez D, Morvan E, Berbon M, Grelard A, Kaufmann B, Ferber M, Bardiaux B, Habenstein B, Saupe SJ, Loquet A (2021) Structures of pathological and functional amyloids and prions, a solid-state NMR perspective. Front Mol Neurosci 14:670513. <https://doi.org/10.3389/fnmol.2021.670513>
- DeForte S, Uversky VN (2016) Order, disorder, and everything in between. Molecules 21(8):1090. [https://doi.org/10.3390/molec](https://doi.org/10.3390/molecules21081090) [ules21081090](https://doi.org/10.3390/molecules21081090)
- Deshmukh M, Evans ML, Chapman MR (2018) Amyloid by design: intrinsic regulation of microbial amyloid assembly. J Mol Biol 430(20):3631–3641.<https://doi.org/10.1016/j.jmb.2018.07.007>
- Dosztányi Z, Csizmok V, Tompa P, Simon I (2005) IUPred: web server for the prediction of intrinsically unstructured regions of proteins based on estimated energy content. Bioinformatics 21(16):3433– 3434.<https://doi.org/10.1093/bioinformatics/bti541>
- Dosztanyi Z, Chen J, Dunker AK, Simon I, Tompa P (2006) Disorder and sequence repeats in hub proteins and their implications for network evolution. J Proteome Res 5(11):2985–2995. [https://doi.](https://doi.org/10.1021/pr060171o) [org/10.1021/pr060171o](https://doi.org/10.1021/pr060171o)
- Dunker AK, Obradovic Z (2001) The protein trinity–linking function and disorder. Nat Biotechnol 19(9):805–806. [https://doi.org/10.](https://doi.org/10.1038/nbt0901-805) [1038/nbt0901-805](https://doi.org/10.1038/nbt0901-805)
- Dunker AK, Obradovic Z, Romero P, Garner EC, Brown CJ (2000) Intrinsic protein disorder in complete genomes. Genome Inform Ser Workshop Genome Inform 11:161–171. [https://doi.org/10.](https://doi.org/10.11234/gi1990.11.161) [11234/gi1990.11.161](https://doi.org/10.11234/gi1990.11.161)
- Dunker AK, Lawson JD, Brown CJ, Williams RM, Romero P, Oh JS, Oldfeld CJ, Campen AM, Ratlif CM, Hipps KW, Ausio J,

Nissen MS, Reeves R, Kang C, Kissinger CR, Bailey RW, Griswold MD, Chiu W, Garner EC, Obradovic Z (2001) Intrinsically disordered protein. J Mol Graph Model 19(1):26–59. [https://doi.](https://doi.org/10.1016/s1093-3263(00)00138-8) [org/10.1016/s1093-3263\(00\)00138-8](https://doi.org/10.1016/s1093-3263(00)00138-8)

- Dunker AK, Brown CJ, Lawson JD, Iakoucheva LM, Obradovic Z (2002a) Intrinsic disorder and protein function. Biochemistry 41(21):6573–6582. <https://doi.org/10.1021/bi012159+>
- Dunker AK, Brown CJ, Obradovic Z (2002b) Identifcation and functions of usefully disordered proteins. Adv Protein Chem 62:25– 49. [https://doi.org/10.1016/s0065-3233\(02\)62004-2](https://doi.org/10.1016/s0065-3233(02)62004-2)
- Dunker AK, Cortese MS, Romero P, Iakoucheva LM, Uversky VN (2005) Flexible nets: the roles of intrinsic disorder in protein interaction networks. FEBS J 272(20):5129–5148. [https://doi.](https://doi.org/10.1111/j.1742-4658.2005.04948.x) [org/10.1111/j.1742-4658.2005.04948.x](https://doi.org/10.1111/j.1742-4658.2005.04948.x)
- Dunker AK, Babu MM, Barbar E, Blackledge M, Bondos SE, Dosztányi Z, Dyson HJ, Forman-Kay J, Fuxreiter M, Gsponer J, Han K-H, Jones DT, Longhi S, Metallo SJ, Nishikawa K, Nussinov R, Obradovic Z, Pappu R, Rost B, Selenko P, Subramaniam V, Sussman JL, Tompa P, Uversky VN (2013) What's in a name? Why these proteins are intrinsically disordered. Intrinsically Disord Proteins 1(1):e24157. <https://doi.org/10.4161/idp.24157>
- Dunker AK, Bondos SE, Huang F, Oldfeld CJ (2015) Intrinsically disordered proteins and multicellular organisms. Semin Cell Dev Biol 37:44–55.<https://doi.org/10.1016/j.semcdb.2014.09.025>
- Dunker AK, Oldfeld CJ, Meng J, Romero P, Yang JY, Chen JW, Vacic V, Obradovic Z, Uversky VN (2008a) The unfoldomics decade: an update on intrinsically disordered proteins. BMC Genomics 9(Suppl 2):S1. <https://doi.org/10.1186/1471-2164-9-S2-S1>
- Dunker AK, Silman I, Uversky VN, Sussman JL (2008b) Function and structure of inherently disordered proteins. Curr Opin Struct Biol 18(6):756–764. <https://doi.org/10.1016/j.sbi.2008.10.002>
- Dunker AK, Uversky VN (2008) Signal transduction via unstructured protein conduits. Nat Chem Biol 4(4):229–230. [https://doi.org/](https://doi.org/10.1038/nchembio0408-229) [10.1038/nchembio0408-229](https://doi.org/10.1038/nchembio0408-229)
- Dyson HJ (2011) Expanding the proteome: disordered and alternatively folded proteins. Q Rev Biophys 44(4):467–518. [https://doi.org/](https://doi.org/10.1017/S0033583511000060) [10.1017/S0033583511000060](https://doi.org/10.1017/S0033583511000060)
- Dyson HJ, Wright PE (2002) Coupling of folding and binding for unstructured proteins. Curr Opin Struct Biol 12(1):54–60. [https://](https://doi.org/10.1016/s0959-440x(02)00289-0) [doi.org/10.1016/s0959-440x\(02\)00289-0](https://doi.org/10.1016/s0959-440x(02)00289-0)
- Dyson HJ, Wright PE (2005) Intrinsically unstructured proteins and their functions. Nat Rev Mol Cell Biol 6(3):197–208. [https://doi.](https://doi.org/10.1038/Nrm1589) [org/10.1038/Nrm1589](https://doi.org/10.1038/Nrm1589)
- Ekman D, Light S, Bjorklund AK, Elofsson A (2006) What properties characterize the hub proteins of the protein-protein interaction network of Saccharomyces cerevisiae? Genome Biol 7(6):R45. <https://doi.org/10.1186/gb-2006-7-6-r45>
- Fisher CK, Stultz CM (2011) Constructing ensembles for intrinsically disordered proteins. Curr Opin Struct Biol 21(3):426–431. <https://doi.org/10.1016/j.sbi.2011.04.001>
- Fonin AV, Darling AL, Kuznetsova IM, Turoverov KK, Uversky VN (2019) Multi-functionality of proteins involved in GPCR and G protein signaling: making sense of structure-function continuum with intrinsic disorder-based proteoforms. Cell Mol Life Sci. <https://doi.org/10.1007/s00018-019-03276-1>
- Fontana A, Fassina G, Vita C, Dalzoppo D, Zamai M, Zambonin M (1986) Correlation between sites of limited proteolysis and segmental mobility in thermolysin. Biochemistry 25(8):1847–1851. <https://doi.org/10.1021/bi00356a001>
- Fontana A, Polverino de Laureto P, De Filippis V, Scaramella E, Zambonin M (1997a) Probing the partly folded states of proteins by limited proteolysis. Fold Des 2(2):R17-26. [https://doi.org/10.](https://doi.org/10.1016/S1359-0278(97)00010-2) [1016/S1359-0278\(97\)00010-2](https://doi.org/10.1016/S1359-0278(97)00010-2)
- Fontana A, Polverinode Laureto P, De Phillips V (1993) Molecular aspects of proteolysis of globular proteins. In: van den Tweel W, Harder A, Buitelear M (eds) Protein stability and stabilization.

Elsevier Science, Amsterdam, pp 101–110. [https://doi.org/10.](https://doi.org/10.1016/B978-0-444-89372-7.50017-8) [1016/B978-0-444-89372-7.50017-8](https://doi.org/10.1016/B978-0-444-89372-7.50017-8)

- Fontana A, Zambonin M, Polverino de Laureto P, De Filippis V, Clementi A, Scaramella E (1997b) Probing the conformational state of apomyoglobin by limited proteolysis. J Mol Biol 266(2):223– 230. <https://doi.org/10.1006/jmbi.1996.0787>
- Fuxreiter M (2020) Classifying the Binding Modes of Disordered Proteins. Int J Mol Sci 21(22):8615. [https://doi.org/10.3390/ijms2](https://doi.org/10.3390/ijms21228615) [1228615](https://doi.org/10.3390/ijms21228615)
- Fuxreiter M, Tompa P (2012) Fuzzy complexes: a more stochastic view of protein function. Adv Exp Med Biol 725:1–14. [https://doi.org/](https://doi.org/10.1007/978-1-4614-0659-4_1) [10.1007/978-1-4614-0659-4_1](https://doi.org/10.1007/978-1-4614-0659-4_1)
- Gabashvili IS, Gregory ST, Valle M, Grassucci R, Worbs M, Wahl MC, Dahlberg AE, Frank J (2001) The polypeptide tunnel system in the ribosome and its gating in erythromycin resistance mutants of L4 and L22. Mol Cell 8(1):181–188. [https://doi.org/10.1016/](https://doi.org/10.1016/s1097-2765(01)00293-3) [s1097-2765\(01\)00293-3](https://doi.org/10.1016/s1097-2765(01)00293-3)
- Gunasekaran K, Tsai CJ, Nussinov R (2004) Analysis of ordered and disordered protein complexes reveals structural features discriminating between stable and unstable monomers. J Mol Biol 341(5):1327–1341. <https://doi.org/10.1016/j.jmb.2004.07.002>
- Haynes C, Oldfeld CJ, Ji F, Klitgord N, Cusick ME, Radivojac P, Uversky VN, Vidal M, Iakoucheva LM (2006) Intrinsic disorder is a common feature of hub proteins from four eukaryotic interactomes. PLoS Comput Biol 2(8):e100. [https://doi.org/10.1371/](https://doi.org/10.1371/journal.pcbi.0020100) [journal.pcbi.0020100](https://doi.org/10.1371/journal.pcbi.0020100)
- Hsu WL, Oldfeld C, Meng J, Huang F, Xue B, Uversky VN, Romero P, Dunker AK (2012) Intrinsic protein disorder and protein-protein interactions. Pac Symp Biocomput:116–127. [https://doi.org/10.](https://doi.org/10.1142/9789814366496_0012) [1142/9789814366496_0012](https://doi.org/10.1142/9789814366496_0012)
- Hsu WL, Oldfeld CJ, Xue B, Meng J, Huang F, Romero P, Uversky VN, Dunker AK (2013) Exploring the binding diversity of intrinsically disordered proteins involved in one-to-many binding. Protein Sci 22(3):258–273. <https://doi.org/10.1002/pro.2207>
- Iakoucheva LM, Kimzey AL, Masselon CD, Bruce JE, Garner EC, Brown CJ, Dunker AK, Smith RD, Ackerman EJ (2001) Identifcation of intrinsic order and disorder in the DNA repair protein XPA. Protein Sci 10(3):560–571. [https://doi.org/10.1110/](https://doi.org/10.1110/ps.29401) [ps.29401](https://doi.org/10.1110/ps.29401)
- Jain N, Chapman MR (2019) Bacterial functional amyloids: Order from disorder. Biochim Biophys Acta Proteins Proteom 1867(10):954– 960. <https://doi.org/10.1016/j.bbapap.2019.05.010>
- Jakob U, Kriwacki RW, Uversky VN (2014) Conditionally and transiently disordered proteins: Awakening cryptic disorder to regulate protein function. Chem Rev 114(13):6779–6805. [https://doi.](https://doi.org/10.1021/cr400459c) [org/10.1021/cr400459c](https://doi.org/10.1021/cr400459c)
- Jumper J, Evans R, Pritzel A, Green T, Figurnov M, Ronneberger O, Tunyasuvunakool K, Bates R, Zidek A, Potapenko A, Bridgland A, Meyer C, Kohl SAA, Ballard AJ, Cowie A, Romera-Paredes B, Nikolov S, Jain R, Adler J, Back T, Petersen S, Reiman D, Clancy E, Zielinski M, Steinegger M, Pacholska M, Berghammer T, Bodenstein S, Silver D, Vinyals O, Senior AW, Kavukcuoglu K, Kohli P, Hassabis D (2021) Highly accurate protein structure prediction with AlphaFold. Nature. [https://doi.org/10.1038/](https://doi.org/10.1038/s41586-021-03819-2) [s41586-021-03819-2](https://doi.org/10.1038/s41586-021-03819-2)
- Kim YI, Burton RE, Burton BM, Sauer RT, Baker TA (2000) Dynamics of substrate denaturation and translocation by the ClpXP degradation machine. Mol Cell 5(4):639–648. [https://doi.org/](https://doi.org/10.1016/s1097-2765(00)80243-9) [10.1016/s1097-2765\(00\)80243-9](https://doi.org/10.1016/s1097-2765(00)80243-9)
- Kovacs D, Rakacs M, Agoston B, Lenkey K, Semrad K, Schroeder R, Tompa P (2009) Janus chaperones: assistance of both RNA- and protein-folding by ribosomal proteins. FEBS Lett 583(1):88–92. <https://doi.org/10.1016/j.febslet.2008.11.049>
- Kovacs D, Tompa P (2012) Diverse functional manifestations of intrinsic structural disorder in molecular chaperones. Biochem Soc Trans 40(5):963–968.<https://doi.org/10.1042/BST20120108>
- Le Gall T, Romero PR, Cortese MS, Uversky VN, Dunker AK (2007) Intrinsic disorder in the Protein Data Bank. J Biomol Struct Dyn 24(4):325–342. [https://doi.org/10.1080/07391102.2007.10507](https://doi.org/10.1080/07391102.2007.10507123) [123](https://doi.org/10.1080/07391102.2007.10507123)
- Lee C, Schwartz MP, Prakash S, Iwakura M, Matouschek A (2001) ATP-dependent proteases degrade their substrates by processively unraveling them from the degradation signal. Mol Cell 7(3):627–637. [https://doi.org/10.1016/s1097-2765\(01\)](https://doi.org/10.1016/s1097-2765(01)00209-x) [00209-x](https://doi.org/10.1016/s1097-2765(01)00209-x)
- Levkovich SA, Gazit E, Laor Bar-Yosef D (2021) Two Decades of Studying Functional Amyloids in Microorganisms. Trends Microbiol 29(3):251–265. [https://doi.org/10.1016/j.tim.2020.](https://doi.org/10.1016/j.tim.2020.09.005) [09.005](https://doi.org/10.1016/j.tim.2020.09.005)
- Meszaros B, Erdos G, Dosztanyi Z (2018) IUPred2A: context-dependent prediction of protein disorder as a function of redox state and protein binding. Nucleic Acids Res 46(W1):W329–W337. <https://doi.org/10.1093/nar/gky384>
- Miskei M, Gregus A, Sharma R, Duro N, Zsolyomi F, Fuxreiter M (2017) Fuzziness enables context dependence of protein interactions. FEBS Lett 591(17):2682–2695. [https://doi.org/10.1002/](https://doi.org/10.1002/1873-3468.12762) [1873-3468.12762](https://doi.org/10.1002/1873-3468.12762)
- Mohan A, Oldfeld CJ, Radivojac P, Vacic V, Cortese MS, Dunker AK, Uversky VN (2006) Analysis of molecular recognition features (MoRFs). J Mol Biol 362(5):1043–1059. [https://doi.org/10.](https://doi.org/10.1016/j.jmb.2006.07.087) [1016/j.jmb.2006.07.087](https://doi.org/10.1016/j.jmb.2006.07.087)
- Monzon AM, Necci M, Quaglia F, Walsh I, Zanotti G, Piovesan D, Tosatto SCE (2020) Experimentally Determined Long Intrinsically Disordered Protein Regions Are Now Abundant in the Protein Data Bank. Int J Mol Sci 21(12):4496. [https://doi.org/](https://doi.org/10.3390/ijms21124496) [10.3390/ijms21124496](https://doi.org/10.3390/ijms21124496)
- Navon A, Goldberg AL (2001) Proteins are unfolded on the surface of the ATPase ring before transport into the proteasome. Mol Cell 8(6):1339–1349. [https://doi.org/10.1016/s1097-2765\(01\)](https://doi.org/10.1016/s1097-2765(01)00407-5) [00407-5](https://doi.org/10.1016/s1097-2765(01)00407-5)
- Oldfeld CJ, Cheng Y, Cortese MS, Romero P, Uversky VN, Dunker AK (2005) Coupled folding and binding with alphahelix-forming molecular recognition elements. Biochemistry 44(37):12454–12470. <https://doi.org/10.1021/bi050736e>
- Oldfeld CJ, Dunker AK (2014) Intrinsically disordered proteins and intrinsically disordered protein regions. Annu Rev Biochem 83:553–584. [https://doi.org/10.1146/annurev-bioch](https://doi.org/10.1146/annurev-biochem-072711-164947) [em-072711-164947](https://doi.org/10.1146/annurev-biochem-072711-164947)
- Oldfeld CJ, Meng J, Yang JY, Yang MQ, Uversky VN, Dunker AK (2008) Flexible nets: disorder and induced ft in the associations of p53 and 14–3-3 with their partners. BMC Genomics 9(Suppl 1):S1.<https://doi.org/10.1186/1471-2164-9-S1-S1>
- Pancsa R, Tompa P (2012) Structural disorder in eukaryotes. PLoS ONE 7(4):e34687. [https://doi.org/10.1371/journal.pone.00346](https://doi.org/10.1371/journal.pone.0034687) [87](https://doi.org/10.1371/journal.pone.0034687)
- Patil A, Nakamura H (2006) Disordered domains and high surface charge confer hubs with the ability to interact with multiple proteins in interaction networks. FEBS Lett 580(8):2041–2045. <https://doi.org/10.1016/j.febslet.2006.03.003>
- Peng K, Radivojac P, Vucetic S, Dunker AK, Obradovic Z (2006) Length-dependent prediction of protein intrinsic disorder. BMC Bioinformatics 7:208.<https://doi.org/10.1186/1471-2105-7-208>
- Peng K, Vucetic S, Radivojac P, Brown CJ, Dunker AK, Obradovic Z (2005) Optimizing long intrinsic disorder predictors with protein evolutionary information. J Bioinform Comput Biol 3(1):35–60
- Peng Z, Mizianty MJ, Kurgan L (2013a) Genome-scale prediction of proteins with long intrinsically disordered regions. Proteins. <https://doi.org/10.1002/prot.24348>
- Peng Z, Oldfeld CJ, Xue B, Mizianty MJ, Dunker AK, Kurgan L, Uversky VN (2013b) A creature with hundred of waggly tails: Intrinsically disordered proteins in ribosome. Cell Mol Life Sci in press
- Peng Z, Yan J, Fan X, Mizianty MJ, Xue B, Wang K, Hu G, Uversky VN, Kurgan L (2015) Exceptionally abundant exceptions: comprehensive characterization of intrinsic disorder in all domains of life. Cell Mol Life Sci 72(1):137–151. [https://doi.org/10.1007/](https://doi.org/10.1007/s00018-014-1661-9) [s00018-014-1661-9](https://doi.org/10.1007/s00018-014-1661-9)
- Prakash S, Matouschek A (2004) Protein unfolding in the cell. Trends Biochem Sci 29(11):593–600. [https://doi.org/10.1016/j.tibs.](https://doi.org/10.1016/j.tibs.2004.09.011) [2004.09.011](https://doi.org/10.1016/j.tibs.2004.09.011)
- Prakash S, Tian L, Ratlif KS, Lehotzky RE, Matouschek A (2004) An unstructured initiation site is required for efficient proteasome-mediated degradation. Nat Struct Mol Biol 11(9):830–837. <https://doi.org/10.1038/nsmb814>
- Romero P, Obradovic Z, Li X, Garner EC, Brown CJ, Dunker AK (2001) Sequence complexity of disordered protein. Proteins 42(1):38–48
- Rubel MS, Fedotov SA, Grizel AV, Sopova JV, Malikova OA, Chernoff YO, Rubel AA (2020) Functional Mammalian Amyloids and Amyloid-Like Proteins. Life (Basel) 10(9):156. [https://doi.org/](https://doi.org/10.3390/life10090156) [10.3390/life10090156](https://doi.org/10.3390/life10090156)
- Schad E, Tompa P, Hegyi H (2011) The relationship between proteome size, structural disorder and organism complexity. Genome Biol 12(12):R120. <https://doi.org/10.1186/gb-2011-12-12-r120>
- Sergeeva AV, Galkin AP (2020) Functional amyloids of eukaryotes: criteria, classifcation, and biological signifcance. Curr Genet 66(5):849–866.<https://doi.org/10.1007/s00294-020-01079-7>
- Sharma R, Raduly Z, Miskei M, Fuxreiter M (2015) Fuzzy complexes: Specifc binding without complete folding. FEBS Lett 589(19 Pt A):2533–2542. [https://doi.org/10.1016/j.febslet.](https://doi.org/10.1016/j.febslet.2015.07.022) [2015.07.022](https://doi.org/10.1016/j.febslet.2015.07.022)
- Singh GP, Ganapathi M, Sandhu KS, Dash D (2006) Intrinsic unstructuredness and abundance of PEST motifs in eukaryotic proteomes. Proteins 62(2):309–315. [https://doi.org/10.1002/prot.](https://doi.org/10.1002/prot.20746) [20746](https://doi.org/10.1002/prot.20746)
- Tompa P (2011) Unstructural biology coming of age. Curr Opin Struct Biol 21(3):419–425. <https://doi.org/10.1016/j.sbi.2011.03.012>
- Tompa P (2012) Intrinsically disordered proteins: a 10-year recap. Trends Biochem Sci 37(12):509–516. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.tibs.2012.08.004) [tibs.2012.08.004](https://doi.org/10.1016/j.tibs.2012.08.004)
- Tompa P, Csermely P (2004) The role of structural disorder in the function of RNA and protein chaperones. FASEB J 18(11):1169– 1175. [https://doi.org/10.1096/f.04-1584rev](https://doi.org/10.1096/fj.04-1584rev)
- Tompa P, Fuxreiter M (2008) Fuzzy complexes: polymorphism and structural disorder in protein-protein interactions. Trends Biochem Sci 33(1):2–8.<https://doi.org/10.1016/j.tibs.2007.10.003>
- Tompa P, Kovacs D (2010) Intrinsically disordered chaperones in plants and animals. Biochem Cell Biol 88(2):167–174. [https://](https://doi.org/10.1139/o09-163) doi.org/10.1139/o09-163
- Tsvetkov P, Reuven N, Shaul Y (2009) The nanny model for IDPs. Nat Chem Biol 5(11):778–781. [https://doi.org/10.1038/nchem](https://doi.org/10.1038/nchembio.233) [bio.233](https://doi.org/10.1038/nchembio.233)
- Turoverov KK, Kuznetsova IM, Uversky VN (2010) The protein kingdom extended: ordered and intrinsically disordered proteins, their folding, supramolecular complex formation, and aggregation. Prog Biophys Mol Biol 102(2–3):73–84. [https://doi.org/10.](https://doi.org/10.1016/j.pbiomolbio.2010.01.003) [1016/j.pbiomolbio.2010.01.003](https://doi.org/10.1016/j.pbiomolbio.2010.01.003)
- Uversky AV, Uversky VN (2014) Amino acid code for protein folding, misfolding, and non-folding. In: Farkas E, Ryadnov M (eds) Amino acids, peptides, and proteins, vol 39. RCS Publishing, London, pp 192–236. [https://doi.org/10.1039/9781849739](https://doi.org/10.1039/9781849739962-00192) [962-00192](https://doi.org/10.1039/9781849739962-00192)
- Uversky VN (2002a) Natively unfolded proteins: a point where biology waits for physics. Protein Sci 11(4):739–756. [https://doi.org/10.](https://doi.org/10.1110/ps.4210102) [1110/ps.4210102](https://doi.org/10.1110/ps.4210102)
- Uversky VN (2002b) What does it mean to be natively unfolded? Eur J Biochem 269(1):2–12. [https://doi.org/10.1046/j.0014-2956.](https://doi.org/10.1046/j.0014-2956.2001.02649.x) [2001.02649.x](https://doi.org/10.1046/j.0014-2956.2001.02649.x)
- Uversky VN (2003) Protein folding revisited. A polypeptide chain at the folding-misfolding-nonfolding cross-roads: which way to go? Cell Mol Life Sci 60(9):1852–1871. [https://doi.org/10.1007/](https://doi.org/10.1007/s00018-003-3096-6) [s00018-003-3096-6](https://doi.org/10.1007/s00018-003-3096-6)
- Uversky VN (2011a) Flexible nets of malleable guardians: intrinsically disordered chaperones in neurodegenerative diseases. Chem Rev 111(2):1134–1166. <https://doi.org/10.1021/cr100186d>
- Uversky VN (2011b) Multitude of binding modes attainable by intrinsically disordered proteins: a portrait gallery of disorder-based complexes. Chem Soc Rev 40(3):1623–1634. [https://doi.org/10.](https://doi.org/10.1039/c0cs00057d) [1039/c0cs00057d](https://doi.org/10.1039/c0cs00057d)
- Uversky VN (2013a) A decade and a half of protein intrinsic disorder: biology still waits for physics. Protein Sci 22(6):693–724. [https://](https://doi.org/10.1002/pro.2261) doi.org/10.1002/pro.2261
- Uversky VN (2013b) Disorder in the lifetime of a protein. Intrinsically Disord Proteins 1(1):e26782.<https://doi.org/10.4161/idp.26782>
- Uversky VN (2013c) Intrinsic disorder-based protein interactions and their modulators. Curr Pharm Des 19(23):4191–4213. [https://doi.](https://doi.org/10.2174/1381612811319230005) [org/10.2174/1381612811319230005](https://doi.org/10.2174/1381612811319230005)
- Uversky VN (2013d) Under-folded proteins: Conformational ensembles and their roles in protein folding, function and pathogenesis. Biopolymers.<https://doi.org/10.1002/bip.22298>
- Uversky VN (2013e) Unusual biophysics of intrinsically disordered proteins. Biochim Biophys Acta 1834(5):932–951. [https://doi.](https://doi.org/10.1016/j.bbapap.2012.12.008) [org/10.1016/j.bbapap.2012.12.008](https://doi.org/10.1016/j.bbapap.2012.12.008)
- Uversky VN (2015) Functional roles of transiently and intrinsically disordered regions within proteins. FEBS J 282(7):1182–1189. <https://doi.org/10.1111/febs.13202>
- Uversky VN (2016a) Dancing Protein Clouds: The Strange Biology and Chaotic Physics of Intrinsically Disordered Proteins. J Biol Chem 291(13):6681–6688. [https://doi.org/10.1074/jbc.R115.](https://doi.org/10.1074/jbc.R115.685859) [685859](https://doi.org/10.1074/jbc.R115.685859)
- Uversky VN (2016b) p53 proteoforms and intrinsic disorder: an illustration of the protein structure-function continuum concept. Int J Mol Sci 17(11):1874. [https://doi.org/10.3390/ijms1](https://doi.org/10.3390/ijms17111874) [7111874](https://doi.org/10.3390/ijms17111874)
- Uversky VN (2016c) Paradoxes and wonders of intrinsic disorder: complexity of simplicity. Intrinsically Disord Proteins 4(1):e1135015. <https://doi.org/10.1080/21690707.2015.1135015>
- Uversky VN (2019a) Intrinsically disordered proteins and their "mysterious" (meta)physics. Front Phys 7:Article 10 (18 pages). [https://](https://doi.org/10.3389/fphy.2019.00010) doi.org/10.3389/fphy.2019.00010
- Uversky VN (2019b) Protein intrinsic disorder and structure-function continuum. Prog Mol Biol Transl Sci 166:1–17. [https://doi.org/](https://doi.org/10.1016/bs.pmbts.2019.05.003) [10.1016/bs.pmbts.2019.05.003](https://doi.org/10.1016/bs.pmbts.2019.05.003)
- Uversky VN, Dunker AK (2010) Understanding protein non-folding. BBA-Proteins Proteom 1804(6):1231–1264. [https://doi.org/10.](https://doi.org/10.1016/j.bbapap.2010.01.017) [1016/j.bbapap.2010.01.017](https://doi.org/10.1016/j.bbapap.2010.01.017)
- Uversky VN, Fink AL (2004) Conformational constraints for amyloid fbrillation: the importance of being unfolded. Biochim Biophys Acta 1698(2):131–153. [https://doi.org/10.1016/j.bbapap.2003.](https://doi.org/10.1016/j.bbapap.2003.12.008) [12.008](https://doi.org/10.1016/j.bbapap.2003.12.008)
- Uversky VN, Gillespie JR, Fink AL (2000) Why are "natively unfolded" proteins unstructured under physiologic conditions? Proteins 41(3):415–427. [https://doi.org/10.1002/1097-](https://doi.org/10.1002/1097-0134(20001115)41:3%3c415::AID-PROT130%3e3.0.CO;2-7) [0134\(20001115\)41:3%3c415::AID-PROT130%3e3.0.CO;2-7](https://doi.org/10.1002/1097-0134(20001115)41:3%3c415::AID-PROT130%3e3.0.CO;2-7)
- Uversky VN, Oldfeld CJ, Dunker AK (2005) Showing your ID: intrinsic disorder as an ID for recognition, regulation and cell signaling. J Mol Recognit 18(5):343–384. [https://doi.org/10.1002/](https://doi.org/10.1002/jmr.747) [jmr.747](https://doi.org/10.1002/jmr.747)
- Uversky VN, Oldfeld CJ, Dunker AK (2008) Intrinsically disordered proteins in human diseases: introducing the D2 concept. Annu Rev Biophys 37:215–246. [https://doi.org/10.1146/annurev.bioph](https://doi.org/10.1146/annurev.biophys.37.032807.125924) [ys.37.032807.125924](https://doi.org/10.1146/annurev.biophys.37.032807.125924)
- Vacic V, Oldfeld CJ, Mohan A, Radivojac P, Cortese MS, Uversky VN, Dunker AK (2007) Characterization of molecular recognition features, MoRFs, and their binding partners. J Proteome Res 6(6):2351–2366.<https://doi.org/10.1021/pr0701411>
- van der Lee R, Buljan M, Lang B, Weatheritt RJ, Daughdrill GW, Dunker AK, Fuxreiter M, Gough J, Gsponer J, Jones DT, Kim PM, Kriwacki RW, Oldfeld CJ, Pappu RV, Tompa P, Uversky VN, Wright PE, Babu MM (2014) Classifcation of intrinsically disordered regions and proteins. Chem Rev 114(13):6589–6631. <https://doi.org/10.1021/cr400525m>
- Van Melderen L, Thi MH, Lecchi P, Gottesman S, Couturier M, Maurizi MR (1996) ATP-dependent degradation of CcdA by Lon protease. Efects of secondary structure and heterologous subunit interactions. J Biol Chem 271(44):27730–27738. [https://doi.org/](https://doi.org/10.1074/jbc.271.44.27730) [10.1074/jbc.271.44.27730](https://doi.org/10.1074/jbc.271.44.27730)
- Ward JJ, Sodhi JS, McGuffin LJ, Buxton BF, Jones DT (2004) Prediction and functional analysis of native disorder in proteins from the three kingdoms of life. J Mol Biol 337(3):635–645. [https://](https://doi.org/10.1016/j.jmb.2004.02.002) doi.org/10.1016/j.jmb.2004.02.002
- Weber-Ban EU, Reid BG, Miranker AD, Horwich AL (1999) Global unfolding of a substrate protein by the Hsp100 chaperone ClpA. Nature 401(6748):90–93. <https://doi.org/10.1038/43481>
- Wilson DN, Nierhaus KH (2005) Ribosomal proteins in the spotlight. Crit Rev Biochem Mol Biol 40(5):243–267. [https://doi.org/10.](https://doi.org/10.1080/10409230500256523) [1080/10409230500256523](https://doi.org/10.1080/10409230500256523)
- Wright PE, Dyson HJ (1999) Intrinsically unstructured proteins: reassessing the protein structure-function paradigm. J Mol Biol 293(2):321–331.<https://doi.org/10.1006/jmbi.1999.3110>
- Wu Z, Hu G, Yang J, Peng Z, Uversky VN, Kurgan L (2015) In various protein complexes, disordered protomers have large per-residue surface areas and area of protein-. DNA- and RNA-binding interfaces. FEBS Lett 589(19 Pt A):2561–2569. [https://doi.org/10.](https://doi.org/10.1016/j.febslet.2015.08.014) [1016/j.febslet.2015.08.014](https://doi.org/10.1016/j.febslet.2015.08.014)
- Xue B, Dunbrack RL, Williams RW, Dunker AK, Uversky VN (2010) PONDR-FIT: a meta-predictor of intrinsically disordered amino acids. Biochim Biophys Acta 1804(4):996–1010. [https://doi.org/](https://doi.org/10.1016/j.bbapap.2010.01.011) [10.1016/j.bbapap.2010.01.011](https://doi.org/10.1016/j.bbapap.2010.01.011)
- Xue B, Dunker AK, Uversky VN (2012) Orderly order in protein intrinsic disorder distribution: disorder in 3500 proteomes from viruses and the three domains of life. J Biomol Struct Dyn 30(2):137–149. <https://doi.org/10.1080/07391102.2012.675145>
- Zhou J, Oldfeld CJ, Yan W, Shen B, Dunker AK (2020) Identifcation of Intrinsic Disorder in Complexes from the Protein Data Bank. ACS Omega 5(29):17883–17891. [https://doi.org/10.1021/acsom](https://doi.org/10.1021/acsomega.9b03927) [ega.9b03927](https://doi.org/10.1021/acsomega.9b03927)

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.