#### RESEARCH



# Wheat debranning: effects on mycotoxins, phenolic content, and antioxidant activity

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#### Abstract

The debranning process, at an industrial scale, was applied to grains of two wheat cultivars to determine its effect on *Fusarium* mycotoxin content and antioxidant activity. Grain samples from the BRS Marcante and BRS Reporte wheat cultivars, naturally contaminated by *Fusarium*, were used in the study. The dry wheat samples were processed on the polisher once or twice and evaluated by hardness index, chemical composition (moisture, protein, and ash), deoxynivalenol (DON) and zearalenone (ZON) levels, phenolic content, and antioxidant activity. In the BRS Marcante cultivar, the debranning process only slightly reduced the DON and ZON contents in whole-wheat flours compared with the previous cleaning treatment (no-debranned). In the BRS Reporte cultivar, the DON concentration decreased by 36% at a debranning ratio of 5%, obtained by polishing, compared with prior cleaning treatment (no-debranned). In addition, the polishing reduced the ZON level by 56% compared with the cleaned wheat. The debranning process did not reduce the antioxidant capacity. Therefore, debranning is a suitable technology to obtain safer and healthier food by minimizing the mycotoxin content and retaining antioxidant capacity.

Keywords Debranning · Deoxynivalenol · Zearalenone · Whole-wheat flour · Antioxidant activity

# Introduction

Wheat is one of the essential staple grains in the world, and it is one of the most important food sources for humans (Cheli et al. 2017). *Fusarium* head blight (FHB) is a disease caused by the *Fusarium graminearum* species complex in

#### Highlights

- The debranning process significantly reduced the DON contamination of the BRS Reporte wheat cultivar.
- The debranning process reduced the level of ZON in the BRS Reporte cultivar.
- The antioxidant capacity was preserved after the debranning process.

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Southern Brazil. The pathogen produces deoxynivalenol (DON) and zearalenone (ZON) mycotoxins (Del Ponte et al. 2015). DON disrupts normal cell function and is related to deleterious effects on human health, such as anorexia, weight loss, malnutrition, dysfunction endocrine, and immunological changes (Pestka 2010; Terzi et al. 2014). ZON causes hyperestrogenic syndrome and reproductive dysfunction (Iqbal et al. 2014). To prevent mycotoxicosis, the maximum limits of mycotoxins are regulated in many countries. International mycotoxins regulation differs among countries, with the acceptable level of DON ranging from 500 to 2000 ppb for foods intended for human consumption (Poroșnicu et al. 2023). În Brazil, the maximum DON limit allowed for unprocessed wheat is 2000 ppb, 1250 ppb for milled wheat, and 1000 ppb for flour (ANVISA 2021). The maximum limit for ZON is 400 ppb for wheat in grains, 200 ppb for milled wheat, and 100 ppb for wheat flour and other derivatives (ANVISA 2021).

According to Cheli et al. (2013), the highest concentration of toxins was found in the outer part of the kernel. The high DON concentration in the bran is of concern because it is used for animal feed, and the bran is also used for direct human consumption due to their health benefits. The milling process removes the bran fraction layers, but it includes key ingredients, rich in bioactive compounds, for cerealbased foods with high antioxidant and nutritional potential (Ciccoritti et al. 2017).

Debranning is based on the sequential removal of the outer grain layers by abrasion and friction, until the desired level, which will depend on the raw materials and the final products (Dexter and Wood 1996). Debranning represents an alternative for using outer layers of kernels rich in natural bioactive compounds (i.e., natural antioxidants and fiber). These compounds potentially benefit human health and are removed during traditional milling (Delcour et al. 2012). Therefore, studies of the effects of debranning technology are important to define the distributions of healthy compounds that can then be preserved during milling and of contaminants (toxic trace elements) to prevent them from entering the food chain (Ficco et al. 2020).

There are many reports of debranning in wheat (Dexter and Wood 1996; De Brier et al. 2015; Martini et al. 2015; Giordano and Blandino 2018). However, there is limited information about applying the debranning process at an industrial scale and its impact on mycotoxin content. Zhao et al. (2019) reported the effect of light debranning in wheat decreased the deoxynivalenol (DON) level by 15.89% at a debranning ratio, percentage of removed bran from the wheat kernel, of 1.2% in the production line. Cheli et al. (2013) reported that the effect of debranning and the efficiency of mycotoxin removal are incredibly variable; the DON reduction ranged from 15 to 78%.

This work aimed to evaluate the effect of the industrial debranning process on *Fusarium* mycotoxins and antioxidant activity in whole-wheat flour from wheat grains harvested from two commercial wheat fields naturally infected by *Fusarium* head blight.

# **Material and methods**

#### Wheat samples

Grain samples from the 2019 crop season of wheat cultivars BRS Marcante and BRS Reponte were obtained from commercial fields. The geographical origin of the wheat fields was Passo Fundo, located in the Rio Grande do Sul State (28° 13' 30.2" S 52° 24' 31.9" W). Wheat samples were likely to be naturally contaminated by *Fusarium gramine-arum*, based on former studies (Del Ponte et al. 2015).

From September to November of 2019, the average temperature was 16.3, 19.3, and 21.2 °C, and there were 11, 13, and 12 days of rainfall, with total values of 55.8, 337.0, and 115.5 mm, respectively (INMET 2022). The humid and hot weather during the flowering and grain filling time was favorable to the FHB epidemic and consequent accumulation of mycotoxins in wheat.

The two wheat cultivars present different rheological properties. The cultivar BRS Marcante has high gluten strength ( $W = > 220 \times 10$ -4 J) and stability (13–20 min); BRS Reponte is characterized by intermediate gluten strength ( $W = 160 \times 10$ -4 J) and stability (6 min). Both cultivars are moderately resistant to *Fusarium* head blight (FHB). Moderately resistant cultivars show reduced symptoms compared to susceptible cultivars when exposed to the *Fusarium* disease, these categories are used based on their reaction to disease infection. The wheat samples were kept at 8 °C in the cold chamber before the analyses.

#### **Cleaning process**

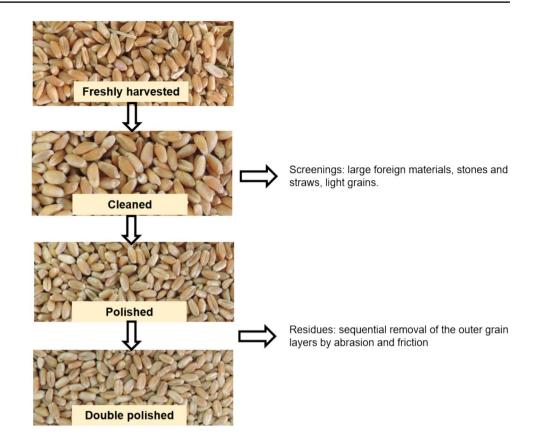
Air-sieve separators (Kepler Webber, model: LC 160–1, Panambi/RS – Brazil) were used to primarily clean impurities from the samples, which differ from the wheat grain with geometric parameters and aerodynamic properties. The sieves' adjustments were as follows: 1st 6 mm round; 2nd 1.75 mm oblong; 3rd 5 mm round; and 4th 2 mm oblong. For BRS Marcante and BRS Reporte cultivars, the relative amount of discarded grain in the cleaning process was 11.6% and 8.9%, respectively.

### Industrial debranning process

The wheat grains were debranned using a commercial polisher (Model Peeling Idugel Mod. PI). The polisher has a capacity of debranning of 5 tons per hour of wheat. The weight of the samples entering the polisher was approximately 500 kg. The dry wheat samples were processed through the polisher once (polished treatment) or two times (double polished treatment) (Fig. 1). In all the debranning treatments, the sampling procedure was to collect a sample each 5 min during the debranning process, thus resulting in four repetitions with 2 kg each. The samples and the residues were weighed before and after the debranning procedure. The discarded residues for the BRS Marcante were 1% in the polished and 2% in the double-polished treatment. Furthermore, for BRS Reponte, discarded residues were 8% in both treatments, polished and double-polished. The moisture content was recorded in each treatment (Portable Grain Moisture equipment AL-102 ECO-Agrologic). After each debranning passage, the equipment was thoroughly cleaned with a dust vacuum and compressed air to minimize cross-contamination.

#### Milling procedures and grain hardness index

The grain samples after each treatment were milled to obtain whole-wheat flour fraction. The samples were milled in a Fig. 1 Schematic treatments are freshly harvested, cleaned, and industrial debranning processes



Laboratory Mill 3100 (Perten Instruments, Huddinge, Sweden). The wheat fractions were grounded to pass through a 0.8-mm screen.

The wheat grain hardness index (GHI) was determined using NIR (NIR instrument FOSS XDS; RCA, Hoganas, Sweden) for the whole-wheat flour. The reference method used for the hardness calibration development was a singlekernel characterization system (SKCS), method 55–31.01 (AACC International 2010).

# Moisture, protein, and ash contents

Moisture and protein contents were determined in 24 samples by 44–15.02 and 39–10.01 methods, respectively (AACC International 2010).

Ash content was determined according to the ICC Standard method 104/1 (ICC 1990) using a muffle furnace at 900 °C for 2.5 h. Protein and ash results were expressed as g/100 g.

# Deoxynivalenol and zearalenone analysis

The wheat samples were grounded in ultra-centrifugal mill model ZM200 (Retsch) with a 1-mm sieve, weighed into a 50 mL Falcon-type tube, and 24 mL of extraction solvent containing methanol: ultrapure water (70:30, v/v) was

added for mycotoxin extraction (Mallmann et al. 2021). After agitation in a vortex mixer for 20 min at 2500 rpm, the content of the tube was centrifuged at  $2465 \times g$  for 10 min (5804R, Eppendorf) and diluted in a vial-type flask.

DON and ZON contents were determined, in 24 samples, by ultra-high-performance liquid chromatographytandem with triple quadrupole mass spectrometry (UHPLC-MS/MS) before and after the industrial debranning process. Briefly, the chromatographic separation was performed at 25 °C on a Gemini C18-column,  $150 \times 4.6$  mm i.d. with 5 µm particle size, equipped with a  $C184 \times 3$  mm i.d. security guard cartridge (Phenomenex, Torrance, CA, USA) (Oliveira et al. 2017). A binary gradient at a flow rate of 0.8 mL/min was performed with solvent A (water) and solvent B (acetonitrile), both modified with 0.5% (v/v) ammonium acetate (Sulyok et al. 2006). The analytical method was developed in-house and validated by the references ISO/IEC 17025: 2017 and Malachová et al. (2014). The accuracy of the technique used to measure DON and ZON was evaluated by recovery experiments (purified analytes in blank wheat) and a certified reference material (CRM), as detailed in Mallmann et al. (2017). The limits of quantification and recovery for DON and ZON were 200 µg.kg-1/80% and 20 µg.kg-1/85%, respectively.

#### Phenolic content and antioxidant activity

In the different treatments carried out on wheat, the free and bound phenolics were extracted according to the methods presented by Adom and Liu (2002) and Sosulski et al. (1982), respectively. Briefly, for the analysis of total phenolic compounds, approximately 2 g of the sample was subjected to extraction using 70% acetone, going through a homogenization and centrifugation processes, with a subsequent stage of concentration and redissolution of the extract (Alves et al. 2016). For the analysis of complexed phenolic compounds, the residue from the extraction of free phenolic compounds was used, which was subjected to basification, heat treatment, and acidification, with subsequent concentration and resuspension in 70% acetone (Alves et al. 2016). The extracts of the free and complexed phenolic compounds were used to measure the total phenolic content (TPC) and antioxidant activity by 2,2-diphenyl-1-picryl-hydrazyl (DPPH) and 2,2 azino bis (3-ethylbenzene thiazoline-6sulfonic acid) (ABTS) radicals (López-Perea et al. 2019).

#### **Statistical analysis**

A software R was used for statistical analysis and figure plotting (R Development Core Team 2018). The packages ggplot2 and ggpubr were used in this study. Analytical determinations conducted in triplicates for each cultivar were submitted to a one-way analysis of variance (ANOVA). A pairwise comparison of mean effects was applied for each cultivar, and those with  $p \le 0.05$  were considered significant.

For phenolic content and antioxidant activity, a comparison of the means was ascertained by Tukey's test to a 5% significance level using an analysis of the variance (ANOVA).

# **Results and discussion**

## Grain hardness index and chemical composition

For the parameter grain hardness index (GHI), the samples differed significantly ( $p \le 0.05$ ) after the debranning processes (polished and double-polished) in both cultivars (Table 1).

In our study, the moisture content ranged from 11.6 to 12.1% and was not significantly affected by debranning (Table 1). The ash content was significantly reduced with the progressive removal of the wheat grains' external layers by debranning (double-polished) in both cultivars (Table 1). Similarly, Ficco et al. (2020) reported that the ash levels for all wheat genotypes decreased progressively with increasing debranning time and after milling.

The protein level was not significantly affected by the debranning process, except for the cultivar BRS Marcante, which demonstrated a significant increase in protein level in the double-polished treatment (Table 1). De Brier et al. (2015) did not report any significant differences in the protein content after the debranning process in wheat.

#### **Deoxynivalenol content**

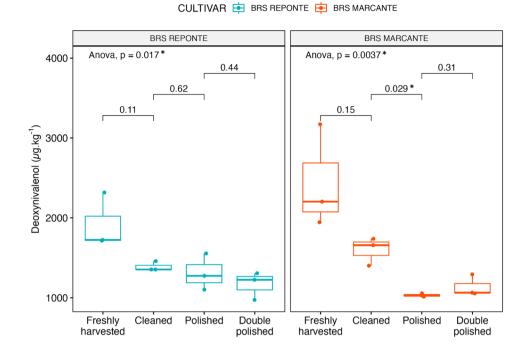
The DON contents of the whole-wheat flours from BRS Marcante and BRS Reporte cultivars are reported in Fig. 2.

In freshly harvested grains, obtained from wheat cultivars, DON was detected at higher levels (Fig. 2). After the cleaning process that eliminated light and shriveled grains (physical removal), DON contamination decreased by 28% and 34% for BRS Marcante and BRS Reporte cultivars, respectively, but did not differ significantly (Fig. 2). These

Table 1 Wheat sample Cultivars Treatment Grain hardness Moisture (%) Ash (%) Protein (%) characterization before and index after debranning and milling processes. Each value represents **BRS** Marcante Freshly harvested 77.4 11.6 1.77 11.8 the mean of three replicate test Cleaned 77.9 11.6 1.74 11.8 portions analyzed Polished 82.3\* 11.8 1.70 11.8 Double-polished 80.6\* 11.7 1.66\* 12.0\* **BRS** Reponte Freshly harvested 72.0 11.8 1.66 13.2 Cleaned 71.9 11.7 1.60 13.2 Polished 73.8\* 12.1 1.61 13.2 Double-polished 72.9\* 11.9 1.51\* 13.4

\*Significantly different by test f ( $p \le 0.05$ )

**Fig. 2** Effect of the debranning process on deoxynivalenol (DON) levels in whole-wheat flours obtained from the BRS Marcante and BRS Reponte cultivars. \**t*.test *p*-values for pairwise group mean comparison and overall *F*-test *p*-value (n = 24)



findings might account for the heterogeneity of mycotoxin distribution in wheat grains and the time of the *Fusarium* infection (Cheli et al. 2013).

In the ANOVA analyses between every two subsequent treatments (pair comparison), the debranning process reduced the DON content significantly by 36% (p=0.029) in the BRS Reporte cultivar when compared with the cleaning treatment (Fig. 2). The treatment polished was equivalent to the debranning level of 5%, with the removal of part of the aleurone of the wheat grains. Similarly, Ciccoritti et al. (2017) reported that debranning is an exciting technology that can provide debranned kernels, which can be used directly to produce improved whole products.

In the BRS Marcante cultivar, the debranning only slightly reduced the DON content in the treatments polished and double-polished, which were equivalent to the debranning levels of 1 and 2%, respectively, when compared with the cleaned treatment (Fig. 2). These differences among wheat cultivars might be due to variations in grain hardness and thickness of the branny layer (Katyal et al. 2019). Besides, the initial level of DON contamination in the BRS Reporte cultivar (2440.0  $\mu$ g.kg<sup>-1</sup>) was higher than in the BRS Marcante cultivar (1918.4 µg.kg<sup>-1</sup>). Zhao et al. (2019) reported that the DON concentration in wheat decreased by 15.9% at a debranning ratio of 1.2% in the production line. Moreover, for the laboratory experiment, the maximum DON removal for grain and flour was 23.4% and 21.9%, respectively (Zhao et al. 2019). Finally, Ríos et al. (2009) showed that the debranning process in two wheat samples naturally contaminated with Fusarium was more efficient than milling alone in reducing the DON content, regardless of its initial levels.

The cleaning and debranning process can be associated during postharvest to alleviate DON content in wheat byproducts. The cleaning process is useful in reducing DON content in wheat by eliminating the light and shriveled grains, especially in the crop seasons characterized by days with excessive moisture associated with warmer temperatures at the heading stage of wheat crop development (Tibola et al. 2016). Diversely, light debranning before milling is more useful when *Fusarium* infections happen later in the crop season, corresponding to the grain filling stage when the infected grains are similar to sound ones and the mycotoxins are located more externally in the wheat grains.

The progressive debranning of wheat grains did not reduce mycotoxin contamination. The treatments polished and double-polished did not present differences in the whole-wheat flours obtained from BRS Marcante and BRS Reponte cultivars. Our results for the DON content were similar to those reported by Sovrani et al. (2012). These authors stated that DON contamination decreased from the external to the internal layers: 64% of total contamination of kernel was found in the 0-5% and 5-10% fractions. The entire debranning with 25% tissue removal resulted only in a slightly higher DON loss of approximately 67% (Sovrani et al. 2012). Furthermore, Tibola et al. (2019) reported that the DON content decreased by 25%, 31%, and 31% in the debranning times of 15, 30, and 60 s, respectively, compared to the no-debranned samples. The distribution profile of mycotoxins in the ground wheat fractions is variable. It depends on mycotoxin, chemical traits, genotypes, fungal contamination levels, and grinding methodology (Cheli et al. 2010, 2013).

Wheat sorting, cleaning, debranning, and milling influence mycotoxin distribution in wheat products entering the food chain (Cheli et al. 2017). In our study, the DON level obtained in the residues from debranning was 12,743  $\mu$ g.kg<sup>-1</sup> and 18,931  $\mu$ g.kg<sup>-1</sup>, for BRS Marcante and BRS Reporte cultivars, respectively. Similarly, the DON content in the residues from double-polished was 18,575  $\mu$ g.kg<sup>-1</sup> and 21,508  $\mu$ g.kg<sup>-1</sup>, for BRS Marcante and BRS Reporte cultivars, respectively.

In Brazil, the DON upper limits established for wholewheat flour in 2021 were 1250  $\mu$ g.kg<sup>-1</sup> (ANVISA 2021). Considering this level, the wheat samples from the cultivars BRS Marcante and BRS Reporte reduced the mycotoxin contamination to levels near the permitted limit after cleaning and debranning processes.

#### Zearalenone content

Cleaning and debranning processes significantly reduced the ZON contamination in the whole-wheat flours from the BRS Reporte cultivar compared with freshly harvested (Fig. 3). The BRS Reporte cultivar showed a ZON level statistically higher than the cultivar BRS Marcante in all treatments (Fig. 3).

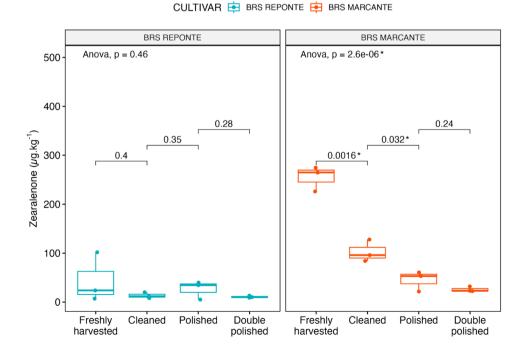
For the cultivar BRS Marcante, the effect of debranning in reducing ZON content was not statistically significant (Fig. 3). Otherwise, the debranning process reduced the level of ZON by 56% when compared with the cleaned sample for the BRS Reporte cultivar (102.6  $\mu$ g.kg<sup>-1</sup>) (Fig. 3). Another reduction in ZON levels (43%) was observed by double-polished, but it was not statistically significant (Fig. 3). Considering both these debranning treatments, the discarded residue was 8%. The ZON level obtained in the residues from debranning was 346 and 2494  $\mu$ g.kg<sup>-1</sup>, for BRS Marcante and BRS Reporte cultivars, respectively.

The second pass on the polisher equipment had no significant effect on the ZON contamination in both cultivars (Fig. 3). In the treatment double-polished, ZON content found in the residue was 498 and 1721  $\mu$ g.kg<sup>-1</sup>, for BRS Marcante and BRS Reporte cultivars, respectively.

#### Phenolic compounds and antioxidant activity

BRS Reporte and BRS Marcante cultivars, the debranning process reduced the total phenolic content in both free and bound extracts (Table 2). For BRS Reporte and BRS Marcante cultivars, the double-polished treatment showed a 14% and 7.6% reduction in free phenolics, respectively, concerning cleaned wheat. In our study, the free and bound phenolic contents were similar. Otherwise, Zhang et al. (2018) concluded that the contribution of bound phenolic acids to the total phenolic content and antioxidant activity was significantly higher than that of free phenolic acids. The BRS Marcante cultivar presented higher levels of total phenolic

**Fig. 3** Effect of the debranning process on zearalenone (ZON) levels in whole-wheat flours obtained from the BRS Marcante and BRS Reponte cultivars. \**t*.test *p*-values for pairwise group means comparison and overall *F*-test *p*-value (n = 24)



Cultivar	Treatment	Total phenolic compounds	compounds	Antioxidant activity	vity			
		Free	Bound		DPPH assay		ABTS assay	
		(mg GAE. kg <sup>-1</sup> d.w)	(mg GAE.kg <sup>-1</sup> d.w)	Free (µmol TE.g <sup>-1</sup> )	Bound (µmol TE.g <sup>-1</sup> )	- Free (μmol TE.g <sup>-1</sup> )		Bound (µmol TE.g <sup>-1</sup> )
BRS Reponte	Freshly harvested $48.6\pm0.9^{d}$	$48.6 \pm 0.9^{d}$	$47.3 \pm 0.5^{d}$	$28.5 \pm 0.3^{a}$	$38.4 \pm 8.9^{a}$	$33.9 \pm 0.1^{a}$		$30.9 \pm 0.2$ bc
	Cleaned	$45.7 \pm 0.9^{d}$	$46.5 \pm 0.6^{d}$	$29.0 \pm 0.2^{a}$	$44.6 \pm 0.3^{a}$	$34.5 \pm 0.4^{a}$		$31.3\pm0.6$ <sup>abc</sup>
	Polished	$42.2 \pm 0.3^{e}$	$45.4 \pm 1.0^{de}$	$29.3 \pm 0.4^{a}$	$45.4\pm0.7^{a}$	$35.4 \pm 0.7^{a}$		$32.5\pm0.2^{a}$
	Double-polished	$39.3 \pm 1.0^{\rm e}$	$42.9 \pm 0.2^{\circ}$	$29.2 \pm 0.0^{a}$	$44.6 \pm 0.4^{a}$	$35.2 \pm 0.3^{a}$		$32.5\pm0.3^{a}$
BRS Marcante	Freshly harvested $70.2 \pm 0.7^{a}$	$70.2 \pm 0.7^{a}$	$70.6 \pm 1.0^{a}$	$28.5 \pm 0.6^{a}$	$44.8\pm0.1^{\rm a}$	$35.1 \pm 0.6^{a}$		$30.1\pm0.3$ °
	Cleaned	$68.1 \pm 0.3^{\mathrm{ab}}$	$69.0 \pm 1.7^{ab}$	$28.8 \pm 0.6^{a}$	$44.5 \pm 0.9^{a}$	$35.0 \pm 0.8^{a}$		$30.1\pm1.0^{\circ}$
	Polished	$65.2 \pm 1.5^{bc}$	$67.0 \pm 1.2^{\mathrm{bc}}$	$29.1 \pm 0.6^{a}$	$44.8\pm0.7^{a}$	$34.8 \pm 0.6^{a}$		$31.2 \pm 0.5$ <sup>abc</sup>
	Double-polished $62.9 \pm 1.2^{\circ}$	$62.9 \pm 1.2^{\circ}$	$65.1 \pm 1.1^{\circ}$	$29.2 \pm 0.4^{a}$	$44.8 \pm 0.8^{a}$	$35.4 \pm 0.6^{a}$		$31.8 \pm 0.4$ <sup>ab</sup>

The results for antioxidant activity referring to the DPPH test for the BRS Reporte and BRS Marcante cultivars indicated that debranning treatments did not influence the free and bound phenolic contents compared to the cleaned sample (Table 2). A similar pattern was observed in the ABTS assay, where the total antioxidant capacity did not reduce in the debranned whole-wheat flour when compared with previous treatments (Table 2).

The bound phenolics were more abundant than the free form, following Adom and Liu (2002), who reported that most phenolic compounds in grains exist in bound form (75% in oats and wheat). Bound phenolics, such as phenolic acids and flavonoids, display strong bioactivities, including anticancer, anti-inflammation, and cardiovascular disease ameliorating properties (Shahidi and Yeo 2016).

There were no significant differences in free and bound phenolics among the cultivars. According to Giordano et al. (2017) regardless of the cultivar considered, the bran fraction resulted in a total antioxidant activity 10 and three-fold higher than the refined flour and wholegrain flour, respectively.

The future direction of the research should investigate in more detail the degree of the debranning process and the percentage of grain tissue removal to preserve healthy compounds and reduce mycotoxin content. Moreover, more samples from different cultivars, with contrasting levels of mycotoxin content, should be included to improve the evaluation of the debranning process and identify appropriated wheat fractions to make superior end-use wheat products.

# Conclusion

*GAE* gallic acid equivalent, *TE* Trolox equivalent

The debranning process has the potential to be an effective post-harvest strategy to minimize mycotoxin contamination in wheat products. The debranning process was effective in reducing DON contamination in the BRS Reporte cultivar (initial level of 2440.0  $\mu$ g.kg<sup>-1</sup>), contributing to obtaining safer whole-wheat products. Furthermore, for BRS Reporte and BRS Marcante wheat cultivars, the debranning process did not reduce the antioxidant activity. Our results are relevant for the proper definition, optimization, and scale-up of in-line debranning at an industrial scale to reduce mycotoxin contamination and produce safer and healthier wheat-based food products.

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Author contribution Dr. CST was responsible for conceiving the idea, carrying out the work, and writing the MS, while Dr. LE obtained the samples for the experiment, Dr. JMCF carried out the statistical

**Table 2** Free and bound phenolic contents and antioxidant activity by the DPPH and ABTS methods of the whole-wheat flour subjected to the debranning process. Each value represents the

analysis and corrected the manuscript, Dr. DS conducted the industrial debranning experiment, and Drs. ERZ and ARGD carried out antioxidant experiments. All the authors revised the manuscript.

**Data availability** No datasets were generated or analysed during the current study.

# Declarations

Competing interests The authors declare no competing interests.

# References

- AACC International. American Association of Cereal Chemists International (2010) Approved Methods of Analysis, 11th Ed. St. Paul, MN. Method 39–10.01. Near-infrared reflectance method for protein determination in small grains. Method 44–15.02. Moisture air-oven method. Method 55–31.01: Single-kernel characterization system for wheat kernel texture
- Adom KK, Liu RH (2002) Antioxidant activity of grains. J Agric Food Chem 50:6182–6187. https://doi.org/10.1021/jf0205099
- Alves GH, Ferreira CD, Vivian PG, Monks JL, Elias MC, Vanier NL, de Oliveira M (2016) The revisited levels of free and bound phenolics in rice: effects of the extraction procedure. Food Chem 208:116–123. https://doi.org/10.1016/j.foodchem.2016.03.107
- ANVISA. National Health Surveillance Agency (2021) Normative Instruction n° 88, of March 26, 2021. Establishes the maximum tolerated limits (LMT) of contaminants in food. *Diário Oficial da União*, Ed. 61, Section 1, p. 226. Brasília, DF, 2021
- Cheli F, Campagnoli A, Ventura V, Brera C, Berdini C, Palmaccio E, Dell'Orto V (2010) Effects of industrial processing on the distributions of deoxynivalenol, cadmium and lead in durum wheat milling fractions. LWT – Food Sci Technol 43:1050–1057. https:// doi.org/10.1016/j.lwt.2010.01.024
- Cheli F, Pinotti L, Rossi L, Dell'Orto V (2013) Effect of milling procedures on mycotoxin distribution in wheat fractions: a review. LWT – Food Sci Technol 54:307–314. https://doi.org/10.1016/j.lwt.2013.05.040
- Cheli F, Pinotti L, Novacco M, Ottoboni M, Tretola M, Dell'Orto V (2017) Mycotoxins in wheat and mitigation measures. In: Wanyera, R, Owuoche, J. (ed.). Wheat improvement, management, and utilization. IntechOpen, London. https://doi.org/10.5772/67240. Available from: https://www.intechopen.com/books/wheat-impro vement-management-and-utilization/mycotoxins-in-wheat-andmitigation-measures. Accessed on 17 Apr 2020
- Ciccoritti R, Taddei F, Nicoletti I, Gazza L, Corradini D, D'Egidio MG, Martini D (2017) Use of bran fractions and debranned kernels for the development of pasta with high nutritional and healthy potential. Food Chem 225:77–86. https://doi.org/10.1016/j.foodc hem.2017.01.005
- De Brier N, Hemdane S, Dornez E, Gomand SV, Delcour JA, Courtin CM (2015) Structure, chemical composition, and enzymatic activities of pearlings and bran obtained from pearled wheat (*Triticum aestivum* L.) by roller milling. J Cereal Sci 62:66–72. https://doi.org/10.1016/j.jcs.2014.12.009
- Del Ponte EM, Spolti P, Ward TJ, Gomes LB, Nicolli CP, Kuhnem PR, Silva CN, Tessmann DJ (2015) Regional and field-specific factors affect the composition of Fusarium head blight pathogens in subtropical no-till wheat agroecosystem of Brazil. Phytopathology 105(2):246–254
- Delcour JA, Rouau X, Courtin CM, Poutanen K, Ranieri R (2012) Technologies for enhanced exploitation of the health-promoting potential of cereals. Trends Food Sci Technol 25:78–86. https:// doi.org/10.1016/j.tifs.2012.01.007

- Dexter JE, Wood PJ (1996) Recent applications of debranning of wheat before milling. Trends Food Sci Technol 7:35–41. https://doi.org/ 10.1016/0924-2244(96)81326-4
- Ficco DBM, Borrelli GM, Miedico O, Giovanniello V, Tarallo M, Pompa C, De Vita P, Chiaravalle AE (2020) Effects of grain debranning on bioactive compounds, antioxidant capacity and essential and toxic trace elements in purple durum wheat. LWT – Food Sci Technol 118:108734. https://doi.org/10.1016/j.lwt. 2019.108734
- Giordano D, Blandino M (2018) Arsenic, lead and cadmium distribution in the pearled fractions of different winter wheat cultivars (*Triticum aestivum* L.). J Cereal Sci 80:94–101
- Giordano D, Locatelli M, Travaglia F, Bordiga M, Reyneri A, Coïsson JD, Blandino M (2017) Bioactive compound and antioxidant activity distribution in roller-milled and pearled fractions of conventional and pigmented wheat varieties. Food Chem 233:483–491. https:// doi.org/10.1016/j.foodchem.2017.04.065
- ICC. International Association for Cereal Science and Technology (1990) Standard n° 104/1: *determination of ash in cereals and cereal products*. Vienna, Austria, Approved 1960, Revised 1990
- INMET Instituto Nacional De Meteorologia Do Brasil (2022) Banco de dados Meteorológicos (BDMEP). Available in: https://bdmep. inmet.gov.br/#. Access in: 20 Aug 2022
- Iqbal SZ, Asi MR, Jinap S, Rashid U (2014) Detection of aflatoxins and zearalenone contamination in wheat-derived products. Food Control 35:223–226. https://doi.org/10.1016/j.foodcont.2013.06.048
- Katyal M, Singh N, Virdi AS, Kaur A, Chopra N, Agarwal S, Roy JK (2019) Effect of debranning on grains and meal characteristics of different Indian and exotic wheat varieties. Food Res Int 123:327– 339. https://doi.org/10.1016/j.foodres.2019.04.036
- López-Perea P, Guzmán-Ortiz FA, Román-Gutiérrez AD, Castro-Rosas J, Gómez-Aldapa CA, Rodríguez-Marín ML, Falfán-Cortés RN, González-Olivares LG, Torruco-Uco JG (2019) Bioactive compounds and antioxidant activity of wheat bran and barley husk in the extracts with different polarity. Int J Food Prop 22(1):646–658. https://doi.org/10.1080/10942912.2019.1600543
- Malachová A, Sulyok M, Beltrán E, Berthiller F, Krska R (2014) Optimization and validation of a quantitative liquid chromatographytandem mass spectrometric method covering 295 bacterial and fungal metabolites including all regulated mycotoxins in four model food matrices. J Chromatogr A 1362:145–156
- Mallmann CA, Dilkin P, Mallmann AO, Oliveira MS, Adaniya ZNC, Tonini C (2017) Prevalence and levels of deoxynivalenol and zearalenone in commercial barley and wheat grain produced in Southern Brazil: an eight-year (2008 to 2015) summary. Tropical Plant Pathology 42(3):146–152
- Mallmann CA, Simões CT, Vidal JK, da Silva CR, de Lima Schlösser LM, de Almeida CA (2021) Occurrence and concentration of mycotoxins in maize dried distillers' grains produced in Brazil. World Mycotoxin J 14(3):259–268. https://doi.org/10.3920/WMJ2020.2669
- Martini D, D'Egidio MG, Nicoletti I, Corradini D, Taddei F (2015) Effects of durum wheat debranning on total antioxidant capacity and content and profile of phenolic acids. J Funct Foods 17:83–92. https://doi.org/10.1016/j.jff.2015.04.054
- Oliveira MS, Rocha A, Sulyok M, Krska R, Mallmann CA (2017) Natural mycotoxin contamination of maize (Zea mays L.) in the South region of Brazil. Food Control 73:127–132
- Pestka JJ (2010) Deoxynivalenol: mechanisms of action, human exposure, and toxicological relevance. Arch Toxicol 84:663–679. https://doi.org/10.1007/s00204-010-0579-8
- Poroșnicu I, Ailincăi LI, Neculai-Văleanu AS, Ariton MA, Mareş M (2023) Legislative aspects regarding the control of mycotoxins. scientific papers animal science and biotechnologies 56(2):153–153
- R Development Core Team (2018) E: The R project for statistical computing, version 3.5.0. Vienna: R Foundation for Statistical Computing. http://www.R-project.org

- Ríos G, Pinson-Gadais L, Abecassis J, Zakhia-Rozis N, Lullien-Pellerin V (2009) Assessment of dehulling efficiency to reduce deoxynivalenol and *Fusarium* levels in durum wheat grains. J Cereal Sci 49:387–392. https://doi.org/10.1016/j.jcs.2009.01.003
- Shahidi F, Yeo JD (2016) Insoluble-bound phenolics in food. Molecules 21(9):1216. https://doi.org/10.3390/molecules21091216
- Sosulski F, Krygier K, Hogge L (1982) Free, esterified, and insolublebound phenolic acids. 3. Composition of phenolic acids in cereal and potato flour. J Agric Food Chem 30:337–340. https://doi.org/ 10.1021/jf00110a030
- Sovrani V, Blandino M, Scarpino V et al (2012) Bioactive compound content, antioxidant activity, deoxynivalenol, and heavy metal contamination of pearled wheat fractions. Food Chem 135:39–46. https://doi.org/10.1016/j.foodchem.2012.04.045
- Sulyok M, Berthiller F, Krska R, Schuhmacher R (2006) Development and validation of a liquid chromatography/tandem mass spectrometric method for the determination of 39 mycotoxins in wheat and maize. Rapid Communications in Mass Spectrometry: an International Journal Devoted to the Rapid Dissemination of upto-the-Minute Research in Mass Spectrometry 20(18):2649–2659
- Terzi V, Tumino G, Stanca AM, Morcia C (2014) Reducing the incidence of cereal head infection and mycotoxins in small grain cereal species. J Cereal Sci 59(3):284–293
- Tibola CS, Fernandes JMC, Guarienti EM (2016) Effect of cleaning, sorting, and milling processes in wheat mycotoxin content.

Food Control 60:174–179. https://doi.org/10.1016/j.foodcont. 2015.07.031

- Tibola CS, Guarienti EM, Dias ARG, Nicolau M, Devos RJB, Teixeira DD (2019) Effect of debranning process on deoxynivalenol content in whole-wheat flours. Cereal Chem 96:717–724. https://doi. org/10.1002/cche.10168
- Zhang J, Ding Y, Dong H, Hou H, Zhang X (2018) Distribution of phenolic acids and antioxidant activities of different bran fractions from three pigmented wheat varieties. J Chem 1–9. https://doi.org/ 10.1155/2018/6459243
- Zhao J, Liu X, Wang F (2019) DON reduction of wheat grain without compromising the lab-scale milling properties of flour. Grain Oil Sci Technol 2(3):62–66. https://doi.org/10.1016/j.gaost.2019.09. 001

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