



# Is tuberous sclerosis complex-associated autism a preventable and treatable disorder?

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Received: 5 May 2023 / Accepted: 10 September 2023 / Published online: 25 October 2023  
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## Abstract

**Background** Tuberous sclerosis complex (TSC) is a genetic disorder caused by inactivating mutations in the *TSC1* and *TSC2* genes, causing overactivation of the mechanistic (previously referred to as mammalian) target of rapamycin (mTOR) signaling pathway in fetal life. The mTOR pathway plays a crucial role in several brain processes leading to TSC-related epilepsy, intellectual disability, and autism spectrum disorder (ASD). Pre-natal or early post-natal diagnosis of TSC is now possible in a growing number of pre-symptomatic infants.

**Data sources** We searched PubMed for peer-reviewed publications published between January 2010 and April 2023 with the terms “tuberous sclerosis”, “autism”, or “autism spectrum disorder,” animal models”, “preclinical studies”, “neurobiology”, and “treatment”.

**Results** Prospective studies have highlighted that developmental trajectories in TSC infants who were later diagnosed with ASD already show motor, visual and social communication skills in the first year of life delays. Reliable genetic, cellular, electroencephalography and magnetic resonance imaging biomarkers can identify pre-symptomatic TSC infants at high risk for having autism and epilepsy.

**Conclusions** Preventing epilepsy or improving therapy for seizures associated with prompt and tailored treatment strategies for autism in a sensitive developmental time window could have the potential to mitigate autistic symptoms in infants with TSC.

**Keywords** Animal model · Autism spectrum disorders · Developmental and epileptic encephalopathy · Mechanisms · mTOR · Risk · Tuberous sclerosis complex

## Introduction

Autism spectrum disorder (ASD) is a heterogeneous neurodevelopmental disorder characterized by deficits in social communication and interactions and restrictive and repetitive patterns of behavior or interests [1]. The prevalence of ASD has progressively grown over the last few years, with the last estimation among children aged 8 years in an American monitoring network being 1 in 44 [2]. ASD is a lifelong condition, and although there is a lack of definite cure for the core symptoms of ASD, some behaviors can be modified by early intervention strategies and parent-mediated communication-focused treatment [3–5].

Influenced by a combination of genetic and environmental factors, ASD is likely polygenic, with many genes having a minor effect on the overall clinical presentation [6]. Progress has been made in understanding genetic underpinnings, and whole-genome sequencing is becoming the first-line test for

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children with ASD [7]. Sequencing technology has improved the capability of identifying possible ASD risk genes, such as synaptic activity-related genes, as well as genes related to molecular regulatory systems, transcription, chromatin modeling, or mechanistic target of rapamycin (mTOR) pathways [8], and shed light on the relative impact of specific biological processes among different autism phenotypes [9].

The discovery of different molecular mechanisms underlying the genetics of autism is now an active area of research [10, 11]. However, the disruption of different neurodevelopmental pathways associated with a relatively high number of genes makes it difficult to disentangle the exact mechanisms involved in ASD [12]. Only a few ASD-related diseases have monogenic causes [13, 14].

Tuberous sclerosis complex (TSC) is a multisystem genetic disorder characterized by age-related development of benign tumors across several organs, including the brain, skin, kidneys, heart, and eyes, and is caused by inactivating mutations in either *TSC1* or *TSC2* genes [15, 16]. The genetic variant results in an overactivation of the mTOR signaling pathway, which plays a crucial role in several brain development processes. The underlying brain pathology of TSC, including cortical tubers and dysplastic cortex and white matter connectivity abnormalities, predispose patients to multiple overlapping comorbidities, including seizures, intellectual disability and a spectrum of behavioral abnormalities with a high incidence of ASD [16, 17].

Overall, more than 2000 pathogenic *TSC1/2* variants have been described, making the genotype–phenotype correlation challenging. Identification of a pathogenic variant in *TSC1/2* is sufficient for the diagnosis of TSC, regardless of clinical findings [18]. However, about 10% of TSC individuals meeting clinical diagnostic criteria have no mutation identified by current genetic testing [19]. Pre-natal diagnosis or early post-natal diagnosis of infants with TSC by the detection of major TSC criteria, such as cardiac rhabdomyoma, subependymal nodules, tubers, or cortical dysplasia, enables major effective clinical surveillance in pre-symptomatic patients not only for seizures but also for autism [20, 21]. This review will discuss new predictive biomarkers for TSC infants with ASD symptoms, the preventive value of early identification of developmental abnormalities, and novel mechanism-based treatment options for TSC-associated ASD.

## Biomarkers for autism spectrum disorder and epilepsy

Epidemiological studies of TSC revealed a surprisingly high incidence of ASD ranging between 17% and 63% [10]. In a single cohort of 103 patients evaluated by a single neuropsychologist, the prevalence of ASD was found in 40%

of children [22]. The TOSCA (Tuberous Sclerosis registry to increase disease Awareness) database provided a good understanding of the natural course of TSC-associated neuropsychiatric disorders [23], with about 50% of individuals having intellectual disability [16, 24].

In TSC children with autistic features, epilepsy often precedes the onset of ASD, raising the issue of the effects of the seizures themselves on the developing brain [25]. In a systematic review and meta-analysis to investigate the association with ASD, the link between seizures and ASD in children with TSC could not be inferred, but a strong association was found with seizure onset during infancy and specifically with infantile spasms [26]. Early onset and persistent seizures, as well as region-specific interictal epileptiform electroencephalography (EEG) activity, may prevent the development of inappropriate neural connectivity leading to autistic-like behaviors. Electrophysiological disturbances may interfere with activity-dependent mechanisms involved in synaptogenesis and dendritic arborization, and these disturbances may disrupt the proper establishment of social cognitive representation, interfering with the process that enables neuronal cells to develop specialization of social functions [27, 28]. In a large multicenter observational study evaluating biomarkers predictive of ASD, early seizure onset in these patients negatively impacted neurodevelopmental outcomes at 24 months of age [29]. Epilepsy, which often begins in the first years of life, is the most common (80%–90% of patients) of a large spectrum of neurological and psychiatric manifestations of patients with TSC, which also includes intellectual disability, behavioral abnormalities, and ASD [30, 31].

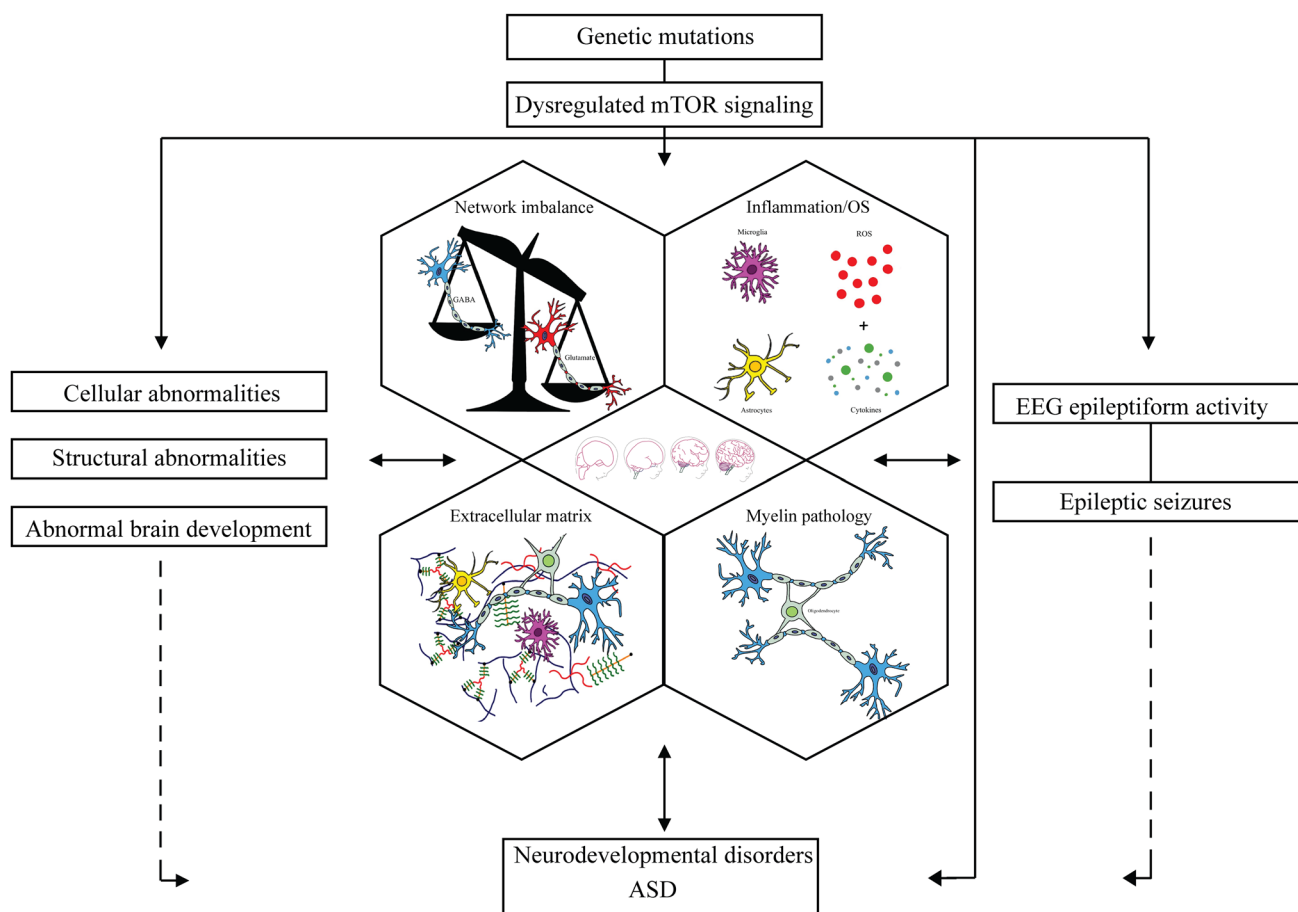
Abnormal behavioral manifestations simulating autistic-like symptoms were first described in 1932 in 29 individuals with impaired social contact, repetitive and stereotyped behaviors, and absence of normal speech [32]. However, it was only after the first description of autism that these behaviors were recognized as core characteristics of ASD [33]. In addition to the early description, the association between ASD and TSC has only recently been systematically investigated [31]. ASD features in TSC may vary from those identified in idiopathic ASD, because they present a higher score on the social communication domain and a lower score on the repetitive domain [34]. In addition, the male-to-female proportion in TSC is reportedly approximately equal [24].

Solid evidence now shows that a *TSC* gene variant could be sufficient to lead to some social deficits; however, the appearance of early onset and persistent seizures could have an additive effect in increasing the likelihood of autistic-like behaviors. This idea is strengthened by the findings that mutations in *TSC1* and *TSC2* are associated with both epilepsy and ASD [35]. In epilepsy, mTOR activation has been implicated in the development

of seizures [36, 37], while in ASD, mTOR dysregulation has been linked to altered neuronal connectivity and synaptic function [38–40]. Moreover, mTOR dysregulation has been reported in a clinical series of children with idiopathic autism; therefore, mTOR dysregulation may represent a direct pathway to both epilepsy and autism [41]. Interestingly, the variability of the neurological and behavioral phenotypes in TSC infants may be related to the degree of loss of function of *TSC1/2* genes, suggesting that mTOR dysregulation could play a direct role in determining susceptibility to both ASD and epilepsy [37, 42].

## Molecular and cellular mechanisms

Complex cellular and molecular events play a role in determining a high risk for both epilepsy and the neurodevelopmental disorders associated with TSC, including ASD and intellectual disability [37, 43] (Fig. 1). TSC and ASD share molecular and cellular mechanisms, including the dysregulation of signaling pathways such as the mTOR pathway, which plays a critical role in regulating cell growth, proliferation, differentiation, and spine dynamics. The downstream results of mTOR dysfunction include multiple cellular processes in the developing brain, including neuronal



**Fig. 1** Mechanisms underlying mTOR-related DEE in TSC and ASD development. Schematic representation of the intricate relationship between genetic mutations, dysregulated mTOR signaling, and downstream mechanisms leading to epilepsy and neurodevelopmental disorders such as ASD. Loss-of-function mutations in *TSC1* or *TSC2* lead to hyperactivation of the mTOR pathway, resulting in cellular and molecular consequences such as network imbalance, inflammation, oxidative stress, extracellular matrix dysfunction, and myelin pathology. As discussed in “Molecular and cellular mechanisms”, growing evidence highlights the interconnected nature of multiple pathological mechanisms in the context of TSC-associated ASD, emphasizing

the mutual relationships between inflammation/oxidative stress, ECM remodeling, myelin pathology, and network dysfunction. These consequences are in direct interplay with cellular and structural abnormalities in the brain, abnormal brain development, and epileptic seizures. The complex interplay among all these factors plays a crucial role in the development of neurodevelopmental disorders such as ASD. *mTOR* mechanistic target of rapamycin, *DEE* developmental and epileptic encephalopathy, *TSC* tuberous sclerosis complex, *ASD* autism spectrum disorder, *ECM* extracellular matrix, *OS* oxidative stress, *ROS* reactive oxygen species, *EEG* electroencephalography, *GABA* gamma-aminobutyric acid

cell morphology, number and shape of synapses, and white matter connectivity [38–40, 44].

One of the principal shared mechanisms that has been proposed between epilepsy and ASD is an abnormal balance between excitatory and inhibitory neurotransmission [45]. Multiple aspects play a role in this imbalance, starting with microtubule cytoskeletal network alterations and abnormal synaptic function [39, 46]. Czapski et al. reported abnormalities in the microtubule-associated protein Tau (MAP-Tau) and reduced levels of MAP1B and neurofilament light proteins, as well as nerve ending swelling in a TSC mouse model of ASD (*Tsc2*<sup>±</sup> mice) [39]. *Tsc2* haploinsufficiency also produces alterations in the expression of synaptic proteins (VAMP1/2 and phospho-synapsin-1), synaptic density, synaptic vesicles, and synaptic mitochondria [46]. Abnormalities in synaptic structure, including synaptic density and mitochondrial ultrastructure, have also been reported in different neuronal in vitro and in vivo models of TSC (*Tsc1/2*-deficient neurons) [47].

Several lines of evidence have also suggested that perturbation of mTOR signaling can lead to a developmental disruption of GABAergic interneurons and enhanced glutamatergic function, leading to abnormal synaptic homeostasis and explaining the concomitant presence of epilepsy and autism in infants with TSC [48–52]. In a recent study, organoids derived from patients with heterozygous *TSC2* mutations were examined, revealing excessive production of mid-gestational human interneurons. This finding further strengthens the association between mTOR dysregulation and the development of GABAergic interneurons [53]. GABAergic deficits are implicated in the dysfunctional neural circuits underlying both neurodevelopmental disorders and epileptogenesis [54–56]. A developmental disruption of GABAergic interneurons linked to abnormal tangential migration can also possibly explain the concomitant occurrence of epilepsy and autism [57–60]. Interestingly, additional studies using resected brain samples from pediatric epilepsy patients, including patients with TSC, have provided compelling evidence of a delay in the physiological maturation of GABAergic signaling (GABAergic immaturity) leading to alterations in excitatory/inhibitory (E/I) balance at the network level [37, 61].

In epilepsy, seizures are associated with an increase in pro-inflammatory cytokines [62, 63], while in ASD, immune dysregulation has been linked to altered neural connectivity, synaptic function, and behavior [64, 65]. Several genes associated with immune function have been implicated in ASD epilepsy, further highlighting the overlap between the two disorders [66]. The mTOR signaling pathway is known to be a central regulator of immune responses [67, 68]. Immunohistochemical and large-scale transcriptomic studies in brain tissue from TSC patients have shown enrichment of reactive

astrocytes and microglia associated with activation of both innate and adaptive immune responses and induction of different inflammatory pathways, which may contribute to epileptogenesis and associated comorbidities (Fig. 1) [69–74]. Interestingly, pre-natal activation of inflammatory pathways has been observed even in developing TSC brain lesions [72, 75]. Notably, functional studies have provided compelling evidence that neuroinflammatory processes and disrupted chloride homeostasis can synergistically contribute to increased brain excitability, shedding light on the intriguing connection between neuroinflammation and the abnormal balance of excitatory and inhibitory neurotransmission in TSC. Indeed, cytokines, such as interleukin-beta, have been recognized as neuromodulators that exert specific effects on GABAergic neurotransmission [76–78]. These findings highlight the interconnected nature of multiple pathological mechanisms in the context of TSC-associated ASD (Fig. 1). Furthermore, oxidative stress, a state of imbalance between the production of reactive oxygen species and the antioxidant defense system, has been implicated in the pathogenesis of various neurological disorders, including epilepsy and ASD [79, 80]. Recent studies have provided evidence of early and sustained oxidative stress in the TSC brain. Interestingly, the extent of oxidative stress is suggested to predict the neuroinflammatory state of the brain [72, 73, 81]. Thus, inflammation and oxidative stress may act synergistically to support the development of a dysfunction underlying both epilepsy and ASD (Fig. 1).

The extracellular matrix (ECM) is a complex network of molecules that surrounds cells and plays a crucial role in regulating cell behavior, including cell adhesion, migration, and signaling [82, 83]. Growing evidence suggests that dysregulation of the ECM may contribute to the pathogenesis of both epilepsy and ASD. For example, studies have shown alterations in the expression and distribution of ECM molecules in the brains of individuals with epilepsy and ASD, suggesting that ECM remodeling may be involved in the development and progression of these disorders [84, 85]. In particular, neuropathological and molecular studies provided evidence of ECM remodeling and dysfunctional cell adhesion molecules in brain tissue from TSC patients [69, 70, 86] (Fig. 1). Higher expression of matrix metalloproteinases (MMPs) has been shown in TSC tubers compared to normal neocortical brain tissue and perituberal cortex [87]. Reactive astrocytes represent an important source of MMPs, and their release may critically contribute to ECM remodeling and pathological synaptic plasticity [84, 88]. Dysregulation of MMP may contribute to the degradation of perineuronal nets around fast-spiking inhibitory interneurons, which is associated with network dysfunction [89, 90]. These observations further emphasize the mutual relationships between neuroinflammation, ECM remodeling, and network dysfunction (Fig. 1).

Finally, myelin, the fatty substance that surrounds and insulates axons in the nervous system, is critical for proper neuronal communication and signaling. Abnormalities in myelin structure and function have been implicated in the pathogenesis of ASD and epilepsy [91]. Growing evidence from neuropathological and in vitro studies strongly suggests that myelin reduction is an intrinsic component of the pathological mechanisms linked to mechanistic target of rapamycin complex 1 (mTORC1) deregulation during brain development [92]. Interestingly, myelin pathology extends beyond white matter, also impacting gray matter myelination. There is compelling evidence that a decrease in the number of oligodendrocytes is associated with lower cognitive functioning, and lower levels of myelin-associated oligodendrocyte basic protein have been linked to the presence of ASD [93]. Furthermore, myelin pathology in TSC has been shown to be associated with the presence of inflammatory cells (T-lymphocytes) [93]. Additionally, recent studies propose that disrupted communication between oligodendrocytes and inhibitory interneurons could contribute to the formation of the pathological network, warranting further investigation in relation to TSC-associated epilepsy and ASD [94, 95]. Taken together, these findings suggest a complex interrelationship between inflammation, myelin pathology, and network dysfunction, highlighting their role in the development of associated neuropsychiatric comorbidities (Fig. 1).

In ASD, myelin dysfunction may lead to impaired communication between neurons and contribute to the social and communication deficits characteristic of the disorder [96]. In epilepsy, myelin abnormalities may contribute to the development and propagation of seizures by disrupting the balance between excitatory and inhibitory signaling in the brain [97]. TSC can be considered a model of genetically determined developmental and epileptic encephalopathy (DEE), a group of severe epilepsies that are characterized by seizures that are often drug resistant and encephalopathy [98]. DEEs represent a heterogeneous group of disorders characterized by early onset, often difficult-to-treat epileptic seizures and EEG abnormalities associated with developmental impairment, which is related to the underlying genetic defects and may possibly worsen as a consequence of epileptiform activity [98–100].

### Pre-clinical studies evaluating the efficacy of mTOR inhibitors in epilepsy and autism

A variety of animal models of TSC, including mice, rats, zebrafish, and non-human primates, each have unique advantages and limitations [37, 101]. These models have been used to investigate the molecular mechanisms underlying

TSC, test the efficacy of potential therapies, and explore the natural history of the disease. Studies involving the co-occurrence of epilepsy and ASD in TSC have predominantly been conducted using rodents [37, 101]. Several animal models have been generated to identify the molecular and cellular mechanisms underlying the development of epilepsy and ASD. These animal models have been utilized not only to investigate the pathophysiology of TSC and ASD but also to test the efficacy of potential treatments (summarized in Table 1).

There is a significant role for the overactivation of the mTOR signaling pathway in linking the pathogenesis of these disorders [35, 37, 111, 112]. The inhibition of this pathway can be achieved by pharmacological agents such as rapamycin and its analogs, known as mTOR inhibitors, which target mTORC1 and lead to a reduction in the hyperactivity of the pathway [37, 43, 101]. The use of these mTOR inhibitors has shown promise in preclinical studies for the treatment of epilepsy, autism, and related neuropsychiatric disorders and could prevent susceptibility to autistic-like behaviors and epilepsy [16, 37].

Rapamycin, a selective inhibitor of the mTOR complex, was observed to reverse impaired social interactions in a mouse model of TSC (*Tsc2*<sup>±</sup> mouse model). Additionally, treatment with rapamycin immediately before and after seizures reversed the increased glutamatergic neurotransmission and attenuated later epilepsy and autistic-like behaviors, indicating that mTOR overactivation is involved in both epileptogenesis and altered social behaviors in a rodent model of acute hypoxia-induced neonatal seizures. Further studies have shown that rapamycin was able to not only reduce seizure susceptibility but also attenuate autistic-like behaviors in rodent models of TSC-ASD (Table 1).

Sato et al. [104] reported that rapamycin treatment alleviated impaired social interaction in both *Tsc1*<sup>±</sup> and *Tsc2*<sup>±</sup> mice. In the *Tsc2*-hGFAP mouse model, which is characterized by cortical abnormalities, seizures, and cognitive deficits resulting from *Tsc2* removal in embryonic neural progenitor cells, the effects of rapamycin treatment on neurodevelopmental defects were found to vary depending on pre-natal and/or post-natal administration [105].

In a rat model with *Tsc2* haploinsufficiency and induced developmental status epilepticus, social deficits were observed. Treatment with the mTOR inhibitor everolimus successfully reversed these deficits, bringing the behavior of the experimental group to control levels [106]. Early epileptic activity had an additional effect on the ASD-like phenotype in an animal model with *Tsc2* haploinsufficiency. However, the ASD-like social impairment induced by *Tsc2*<sup>±</sup> responded well to everolimus therapy, while the behavioral consequences of prolonged epileptic activity were not ameliorated by the drug [108]. In a *TSC2* conditional knockout mouse model, sirolimus was

**Table 1** Summary of the main preclinical studies evaluating the efficacy of the mTOR inhibitors on epilepsy and behavioral deficits

Authors	Mouse model/mechanism	Drug	Results
Ehninger et al., 2008 [102]	<i>Tsc2</i> <sup>±</sup> mice	Rapamycin	Rapamycin rescued not only the synaptic plasticity, but also the behavioral deficits
Talos et al., 2012 [103]	Rodent model of acute hypoxia-induced neonatal seizures (early life seizures, mTORC1 activity)	Rapamycin	Reversion of early increase in glutamatergic neurotransmission and seizure susceptibility and attenuation of later life epilepsy and autistic-like behavior
Sato et al., 2012 [104]	<i>Tsc1</i> <sup>+/-</sup> and <i>Tsc2</i> <sup>+/-</sup> mice	Rapamycin	Reversion of impairment of reciprocal social interaction
Way et al., 2012 [105]	<i>Tsc2</i> -hGFAP mouse model		The differential effects of pre-natal and/or post-natal rapamycin on neurodevelopmental defects and cognition
Tsai et al., 2012 [106]	<i>Tsc1</i> <sup>fllox/fllox</sup> mice <sup>a</sup>	Rapamycin	Reduction of seizure susceptibility and attenuation of autistic-like behaviors
Schneider et al., 2017 [107]	<i>Tsc2</i> <sup>±</sup> rats	Everolimus	Reversion of social deficit behaviors in the <i>Tsc2</i> haploinsufficiency plus DSE
Petrasek et al., 2021 [108]	<i>Tsc2</i> <sup>±</sup> rats	Everolimus	Persistent improvement of the social deficit induced by <i>Tsc2</i> <sup>±</sup> , no response to deficits related to DSE
Koike-Kumagai et al., 2022 [109]	<i>TSC2</i> conditional knockout mice (Mitf-cre, <i>Tsc2</i> KO; <i>Tsc2</i> cKO)	Rapamycin	Reduction of seizures and neuropsychiatric symptoms via changes of microglial polarity, except for ASD
Kashii et al., 2023 [110]	<i>TSC1</i> and <i>TSC2</i> double heterozygous mutant ( <i>TscD</i> <sup>±</sup> ) mice	Rapamycin	Impairments in social behaviors were rescued by rapamycin treatment

DSE developmental status epilepticus, mTOR mechanistic target of rapamycin, mTORC1 mechanistic target of rapamycin complex 1, TSC tuberous sclerosis complex, ASD autism spectrum disorder, KO knockout, cKO conditional knockout. <sup>a</sup>*Tsc1* deleted in cerebellar Purkinje cell [L7Cre; *Tsc1*<sup>fllox/+</sup> (het) or L7Cre; *Tsc1*<sup>fllox/fllox</sup> (mutant)]

observed to change the polarity of microglia via inhibition of mTORC1 and reduce multiple neuropsychiatric symptoms in TSC mice, except for ASD [109]. Finally, Kashii et al. investigated social behaviors in *Tsc1* and *Tsc2* double heterozygous mutant mice by the social interaction test and three-chambered sociability tests [110]; impairments in social behaviors were rescued by rapamycin treatment. Interestingly, the gene expression changes compared with wild-type mice were enriched in the inflammation-related canonical pathway.

Collectively, these findings suggest that mTOR pathway dysregulation plays a significant role in the interaction between epilepsy and autism and highlight the potential of mTOR inhibitors as a treatment option for these disorders. Pharmacological targeting of this pathway with mTOR inhibitors such as rapamycin has shown promise in both preventing susceptibility to the development of autistic-like behaviors and epilepsy and reversing these symptoms in animal models. While there are limitations to the use of animal models, the knowledge gained from these studies is critical to the development of new treatments and a better understanding of the underlying mechanisms of these disorders.

## Factors for an increased risk of autism spectrum disorder

Potential risk factors for the co-occurrence of ASD in TSC infants include genetic mutations, structural brain abnormalities, and epileptic activity [24]. In the multicenter Tuberous Sclerosis Complex Autism Center of Excellence Research Network study group, having a *TSC2* pathogenic variant was associated with significantly lower Mullen Scales of Early Learning scores at 24 months, independent of seizures [113]. Recent evidence has shown that *TSC2* genetic variants may cause a high risk of developing ASD in comparison to *TSC1* gene variants [114].

The detection of fetal brain lesions on magnetic resonance imaging is associated with subsequent poor neurodevelopmental outcomes [115]. Structural brain abnormalities, including tuber burden, cyst-like tubers, and abnormal connectivity, have been observed in children with ASD-associated autism [22, 116]. Abnormal fusiform gyrus connectivity captures the risk of developing ASD as early as 12 months of age [117]. A recent study provided

evidence of a strong association between tuber involvement of the right fusiform face area and ASD diagnosis [118]. Moreover, imaging studies examining brain–behavior relations have shown early abnormalities in white matter in infants with TSC who later develop ASD [91, 119, 120]. White matter diffusion tensor imaging abnormalities have been detected in children with ASD in comparison with children without ASD, suggesting microstructural changes in myelination and axonal integrity [121]. Furthermore, cerebellar volume loss, perhaps reflecting Purkinje cell degeneration, may predict neurodevelopmental severity in patients with *TSC2* mutations [122].

In an early developmental stage during the critical period of synaptogenesis, early onset refractory seizures may contribute to autistic-like behaviors. Persistent seizure activity may disrupt connectivity in cortical areas that are crucial for the normal development of emotional and cognitive functions, including the cingulum [123] and the temporal areas [22]. Furthermore, the presence of multifocal epileptiform abnormalities on EEG and relative alpha band power could enhance the detection of risk for ASD, since it was significantly elevated in an ASD group of TSC infants at 24 months of age [124]. Sleep dysfunction is strictly connected with ASD in infants with TSC and is dependent on mTOR overactivation [125]. Sleep disturbances and perturbation or disruption of the sleep–wake cycle could be correlated with ASD symptoms [126]. In a multicenter prospective observational study, language variables by 12 months predicted an ASD diagnosis at 36 months [127]. Serum-based biomarkers

such as microRNAs have shown potential for early risk assessment of ASD [128]. Early predictors of impending ASD are summarized in Table 2. Identifying early indicators of impending autism and epilepsy can have a clear impact on prevention and early intervention efforts [130].

## Advances in early recognition of autism spectrum disorder

The Autism Diagnostic Observation Scale (ADOS) combined with expert clinical assessment is considered the gold standard for the diagnosis of ASD even before 24 months [131]. However, behavioral markers detected from the autism observation scale of infants were consistent with ADOS items at 24 months [132]. Recently, we gained a deeper understanding of earlier developmental manifestations of TSC that precede ASD or intellectual disability [29, 133]. Already at 6 months, infants with TSC that will be classified as having ASD at the age of 24 months tend to diverge from a typical neurodevelopmental trajectory and present a lower level of developmental skills when compared with infants classified with TSC and not ASD at 24 months [134]. Rigorous prospective studies of the developmental trajectories of high-risk infants are needed [135]. However, although TSC is diagnosed early during development, only a few clinical studies have evaluated the early trajectory deviation leading to developmental delay and ASD, monitoring infants with TSC prospectively at multiple time points. In

**Table 2** Clinical, electroencephalography, magnetic resonance imaging, and genetic and serum biomarkers of ASD

Author	Clinical biomarkers
Moavero et al., 2020 [129]	Early onset refractory seizures and infantile spasms
Zhang et al., 2020 [125]	Sleep dysfunctions
Schoenberg et al., 2020 [127]	Language variables
EEG biomarkers	
Cook et al., 2020 [124]	Early multifocal epileptiform abnormalities on EEG
Cook et al., 2020 [124]	Alpha band power EEG abnormalities during sleep
MRI biomarkers	
Hulshof et al., 2022 [115]	Fetal brain tumors
Ruppe et al., 2014 [116]	Tuber burden and abnormal connectivity
Peters et al., 2019 [121]	White matter DTI abnormalities
Numis et al., 2011 [22]	Cyst like tubers
Scherrer et al., 2020 [117]	Abnormal fusiform gyrus connectivity
Srivastava et al., 2018 [122]	Cerebellar volume loss
Genetic biomarkers	
Ogórek et al., 2020 [114]	<i>TSC2</i> variants
Farach et al., 2020 [113]	<i>TSC2</i> variants
Serum biomarkers	
Scheper et al., 2022 [128]	Serum-based biomarkers

ASD autism spectrum disorder, MRI magnetic resonance imaging, EEG electroencephalography, DTI diffusion tensor imaging, TSC tuberous sclerosis complex

the epileptogenesis in a genetic model of epilepsy–tuberous sclerosis complex (EPISTOP) trial, there was evidence that in TSC infants receiving a diagnosis of ASD at 2 years of age, some developmental abnormalities were already evident at 6 months of age, expanding to all developmental domains at 12 months [133].

## Clinical trials for tuberous sclerosis complex-associated autism spectrum disorder

### Targeting E/I balance with GABAergic drugs as possible secondary prevention of ASD

Among the mentioned factors that increase the risk for ASD in TSC, only seizures may be potentially preventable, and prompt detection and treatment of both interictal epileptiform discharges on EEG and clinical seizures could improve the course of the disorder, reducing the negative impact of early seizures on neurodevelopment [29].

Seizures are preceded by a latent period of epileptogenesis in which interictal epileptiform discharges may appear in the EEG [136]. Prompt control of EEG abnormalities and seizures plays a crucial role in preventing subsequent epileptic encephalopathy and in reducing the cognitive and behavioral consequences of epilepsy [124, 137]. Vigabatrin is known to be particularly beneficial in TSC-associated infantile spasms and may have disease-specific efficacy, particularly due to the GABAergic effect [138, 139]. Therefore, pre- and postepileptic windows for secondary prevention of ASD may exist. A shorter gap between seizure onset and the start of vigabatrin treatment could reduce the risk of epileptic encephalopathy and the severity of intellectual disability and ASD, minimizing the negative impact of early life seizures [140]. Pre-symptomatic treatment with vigabatrin given during the period of epileptogenesis and before any clinically detectable seizure may show even better efficacy. In an international randomized-controlled trial, preventive treatment with vigabatrin resulted in a reduced risk of developing infantile spasms and drug-resistant epilepsy in comparison with treatment initiated after seizure onset [141]. However, in the EPISTOP trial, autistic features at 24 months of age were detected irrespective of treatment strategy, although preventive treatment resulted in a better neuropsychological result than in other historical cohorts of TSC children [141].

Bumetanide has been reported to alter the synaptic excitation/inhibition balance by strengthening the action of GABA, thereby attenuating the severity of ASD in animal models [142]. Bumetanide treatment is a potential treatment to ameliorate the behavioral burden and quality of life associated with TSC [143]. Furthermore, bumetanide has

been reported to have effects on EEG, reflected in increased absolute and relative alpha power and functional E/I ratio and in decreased central frequency; these EEG findings were related to medium-to-high repetitive behavior scale-revised improvement [144].

### Targeting translated and epigenetic regulation with mTOR inhibitors

The involvement of mTOR signaling in determining the behavioral phenotype associated with TSC led to the hypothesis that mTOR inhibitors could also have benefits in ASD symptoms. The negative effect of mTOR overactivity on white matter microstructural integrity was improved by mTOR inhibition with everolimus, and rapid white matter maturation was observed during long-term treatment in younger patients [121]. Clinical evidence based on anecdotal cases showed contradictory results about the beneficial effects of mTOR inhibitors, including rapamycin and its derivative everolimus, on autistic symptoms in a few patients [145].

In a prospective double-blind randomized and placebo-controlled study to evaluate treatment efficacy with everolimus on neurocognition and behavior in children with TSC, no significant improvement in neurocognitive functions was observed in 47 individuals with TSC [146]. However, no definite conclusions can be achieved from this study considering that the drug was used in individuals older than 6 years of age and already with symptoms. What is still unclear is the right timing of everolimus. However, the safety of everolimus has been proven in young children under the age of 24 months [147]. After the promising results from preclinical studies, clinical trials with mTOR inhibitors are now underway, even if the right timing to start everolimus is still unclear. Since the dysregulation of the mTOR pathway is a possible pathological substrate for TSC-related seizures and developmental encephalopathy, different windows of opportunity for the prevention of the development of seizure and co-occurring cognitive and behavioral disorders could be possible [36]. mTOR signaling is already activated in fetal tubers in various brain cell populations, including dysmorphic neurons within the dysplastic cortex and giant cells within the subcortical white matter, providing a likely pathological substrate for early onset epilepsy [43, 75, 148]. These findings raise the possibility that modifying epileptogenesis in the immature brain by mTOR inhibition may also prevent other manifestations of network dysfunctions leading to autism and other neurobehavioral comorbidities.

In a single case report, a fetus diagnosed with TSC at the age of 19 weeks of gestation who presented cardiac and brain tumors on pre-natal ultrasound and underwent intrauterine treatment with everolimus presented regression and



subsequent stabilization of the cardiac and brain lesions. Additionally, the patient did not develop seizures or autism and presented good psychomotor development [149].

## Non-pharmacological treatments

Brain social skills develop through an active process of post-natal interaction with the environment. The efficacy of environmental enrichment with enhanced somatosensory, cognitive, visual, and motor stimulation in the critical developmental window of synaptogenesis has been reported to improve joint engagement, joint attention, and social interaction in toddlers at high risk of ASD [150–152].

Due to the greater plasticity of developmental neural systems, intensive and targeted parent-mediated behavioral interventions can speed up the developmental trajectories and ameliorate the disabling effects of ASD [152]. In TSC infants, new evidence shows that early environmental enrichment may restore functional and structural changes and improve impaired synaptic plasticity. In a pilot study on five children with TSC aged 1–3 years, early play-based, parent-mediated behavioral interventions were able to improve communications and social skills [151].

Clinical trials on early intervention in toddlers aged 12–16 months are in progress, yet standardized measures to capture the effect of clinical interventions are needed. The recent TSC consensus workshop for clinical recommendations emphasized the need to offer early intervention services focused on building social communication skills in children with ASD concerns (Specchio et al., submitted to EJPN). Early screening for sleep problems and personalized approaches with sleep interventions may improve not only sleep but also daytime behaviors in TSC children with ASD. Melatonin, a hormone involved in the endogenous synchronization of internal biological clocks, may facilitate falling asleep and maintaining sleep in children with ASD [153, 154].

Epilepsy surgery in TSC typically involves the resection or disconnection of the epileptogenic tubers or the placement of neuromodulation devices, such as vagus nerve stimulation or responsive neurostimulation systems. Multiple studies have demonstrated the effectiveness of epilepsy surgery in TSC patients with refractory seizures. A study by Jansen et al. [155] reported a significant reduction in seizure frequency and improved seizure control in 70% of TSC patients who underwent epilepsy surgery. Another study by Specchio et al. [156] showed that surgical interventions, including focal resections and hemispherectomies, resulted in a favorable seizure outcome in TSC patients, with over 60% achieving seizure freedom or a significant reduction in seizure frequency. The potential risks and benefits of surgery should be carefully considered, as TSC patients may have multiple tubers or cortical abnormalities, making the surgical approach challenging. Epilepsy surgery can be a valuable therapeutic option for individuals with TSC who have medically refractory seizures. It offers the potential for improved seizure control and enhanced quality of life.

## Future directions and conclusions

The emerging clinical neuroscience of TSC-associated ASD, possible prevention, and early intervention are shown in Table 3. Significant advances made in the neurobiology of TSC and animal models have clearly suggested that the neurodevelopmental manifestations of TSC, including ASD, may be a direct consequence of gene variants and cell signaling abnormalities. Therefore, mTOR signaling overactivity through both direct and indirect pathways plays a role in the disruption of GABAergic interneurons during early critical stages of neurodevelopment and confers a high susceptibility to autism in infants and young children with TSC.

**Table 3** The emerging clinical neuroscience of TSC-associated ASD and possible prevention and early intervention

TSC is a genetically defined model for the early emergence of ASD
Progress has been made in understanding the cell consequences of <i>TSC1/2</i> pathogenic variants and the consequent mTOR overactivation in different cell types at specific stages of brain development
Genetic, cellular, EEG, and MRI biomarkers are now available to identify pre-symptomatic TSC infants at risk for developing ASD and epilepsy
Progress in early detection of autistic symptoms has promoted earlier interventions which should begin soon after the diagnosis of TSC in a sensitive time window, to improve synaptic connectivity and reach more positive developmental outcomes
Mouse models are making it possible to examine the effect of the <i>TSC</i> gene variant, to better understand pathological pathways, moving toward a targeted therapy
Preventive trials with mTOR inhibitors should be now designed in pre-symptomatic infants to evaluate if this strategy could have disease modifying effects, preventing, or mitigating TSC-associated ASD symptomatology

TSC tuberous sclerosis complex, ASD autism spectrum disorder, mTOR mechanistic target of rapamycin, EEG electroencephalography, MRI magnetic resonance imaging

Several risk factors for both autism and epilepsy have been described, and progress in the early identification of prodromic signs has been made. These findings are changing our clinical practice and have provided new insights for behavioral and pharmacological therapeutic options. However, in this specific context, it is difficult to separate treatment from prevention, particularly from tertiary prevention. Especially, when dealing with chronic conditions, tertiary prevention aims to diminish the effects of a disease or disorder by reducing or minimizing disability while boosting the person's quality of life. In epidemiological terms, the aim of tertiary prevention is to lessen the impact of complications. An increase in the knowledge and identification of subclinical ASD endophenotypes might even allow interference with the progression toward a full-blown ASD developmental trajectory. From this perspective, future advancements might lead to the possibility of developing secondary prevention strategies, further highlighting the preventive value of early detection.

Even though animal models demonstrated promising results in improving or reversing behavioral symptoms, recent clinical findings showed that no therapeutic options targeting either GABAergic or mTOR inhibition with everolimus were able to determine clinically significant behavioral gains even in children with a concomitant decrease in seizure frequency after targeted therapy.

Elucidating the cellular and molecular basis of TSC-associated ASD has shown a close link between the encephalopathic process and the epileptogenesis leading to ASD or epilepsy. Predicting, preventing, and improving treatment of TSC-associated seizures, including pharmacological and surgical intervention, could reduce the burden of autistic symptoms in infants with TSC. Future studies should clarify whether early parent-mediated behavioral intervention in a developmental sensitive time window associated with early prescription of mTOR inhibitors has the potential to mitigate the severity of ASD symptoms.

**Acknowledgements** SN was supported by Next-Generation EU (NGEU) and funded by the Ministry of University and Research (MUR), National Recovery and Resilience Plan (NRRP), under project No. MNESYS (PE0000006)—a multiscale integrated approach to the study of the nervous system in health and disease (DN. 1553 11.10.2022).

**Author contributions** CP conceived the review. SM and AE contributed particularly to “[Molecular and cellular mechanisms](#)” and “[Pre-clinical studies evaluating the efficacy of mTOR inhibitors in epilepsy and autism](#)” and the preparation of Fig. 1. All authors drafted and prepared the manuscript. All authors read, revised, and approved the final manuscript.

**Funding** AE and SM were supported by Healthcare Research and Medical Sciences (ZonMw; No. 09120012010007).

**Data availability** Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

## Declarations

**Ethical approval** Not required.

**Conflict of interest** No financial or non-financial benefits have been received or will be received from any party related directly or indirectly to the subject of this article. The authors have no conflict of interest to declare.

## References

1. American Psychiatric Association. Diagnostic and statistical manual of mental disorders. 5th ed. Washington: American Psychiatric Association; 2013.
2. Maenner MJ, Shaw KA, Bakian AV, Bilder DA, Durkin MS, Esler A, et al. Prevalence and characteristics of autism spectrum disorder among children aged 8 years—autism and developmental disabilities monitoring network, 11 sites, United States, 2018. *MMWR Surveill Summ.* 2021;70:1–16.
3. Tarver J, Palmer M, Webb S, Scott S, Slonims V, Simonoff E, et al. Child and parent outcomes following parent interventions for child emotional and behavioral problems in autism spectrum disorders: a systematic review and meta-analysis. *Autism.* 2019;23:1630–44.
4. Wang SH, Zhang HT, Zou YY, Cheng SM, Zou XB, Chen KY. Efficacy and moderating factors of the Early Start Denver Model in Chinese toddlers with autism spectrum disorder: a longitudinal study. *World J Pediatr.* 2023;19:741–52.
5. Aaronson B, Estes A, Rogers SJ, Dawson G, Bernier R. The early start Denver model intervention and mu rhythm attenuation in autism spectrum disorders. *J Autism Dev Disord.* 2022;52:3304–13.
6. Cheroni C, Caporale N, Testa G. Autism spectrum disorder at the crossroad between genes and environment: contributions, convergences, and interactions in ASD developmental pathophysiology. *Mol Autism.* 2020;11:69.
7. Fernandez BA, Scherer SW. Syndromic autism spectrum disorders: moving from a clinically defined to a molecularly defined approach. *Dialogues Clin Neurosci.* 2017;19:353–71.
8. Lim HK, Yoon JH, Song M. Autism Spectrum disorder genes: disease-related networks and compensatory strategies. *Front Mol Neurosci.* 2022;15:922840.
9. Emberti Gialloreti L, Enea R, Di Micco V, Di Giovanni D, Curatolo P. Clustering analysis supports the detection of biological processes related to autism spectrum disorder. *Genes (Basel).* 2020;11:1476.
10. Specchio N, Di Micco V, Trivisano M, Ferretti A, Curatolo P. The epilepsy-autism spectrum disorder phenotype in the era of molecular genetics and precision therapy. *Epilepsia.* 2022;63:6–21.
11. Di Giovanni D, Enea R, Di Micco V, Benvenuto A, Curatolo P, Emberti GL. Using machine learning to explore shared genetic pathways and possible endophenotypes in autism spectrum disorder. *Genes (Basel).* 2023;14:313.
12. Emberti Gialloreti L, Curatolo P. Autism spectrum disorder: why do we know so little? *Front Neurol.* 2018;9:670.
13. Hulbert SW, Jiang YH. Monogenic mouse models of autism spectrum disorders: common mechanisms and missing links. *Neuroscience.* 2016;321:3–23.
14. Benvenuto A, Moavero R, Alessandrelli R, Manzi B, Curatolo P. Syndromic autism: causes and pathogenetic pathways. *World J Pediatr.* 2009;5:169–76.
15. Curatolo P, Bombardieri R, Jozwiak S. Tuberous sclerosis. *Lancet.* 2008;372:657–68.

16. Curatolo P, Specchio N, Aronica E. Advances in the genetics and neuropathology of tuberous sclerosis complex: edging closer to targeted therapy. *Lancet Neurol.* 2022;21:843–56.
17. de Vries PJ, Wilde L, de Vries MC, Moavero R, Pearson DA, Curatolo P. A clinical update on tuberous sclerosis complex-associated neuropsychiatric disorders (TAND). *Am J Med Genet C Semin Med Genet.* 2018;178:309–20.
18. Northrup H, Aronow ME, Bebin EM, Bissler J, Darling TN, de Vries PJ, et al. Updated international tuberous sclerosis complex diagnostic criteria and surveillance and management recommendations. *Pediatr Neurol.* 2021;123:50–66.
19. Canevini MP, Kotulska-Jozwiak K, Curatolo P, La Briola F, Peron A, Słowińska M, et al. Current concepts on epilepsy management in tuberous sclerosis complex. *Am J Med Genet C Semin Med Genet.* 2018;178:299–308.
20. Dragoumi P, O'Callaghan F, Zafeiriou DI. Diagnosis of tuberous sclerosis complex in the fetus. *Eur J Paediatr Neurol.* 2018;22:1027–34.
21. Davis PE, Filip-Dhima R, Sideridis G, Peters JM, Au KS, Northrup H, et al. Presentation and diagnosis of tuberous sclerosis complex in infants. *Pediatrics.* 2017;140:e20164040.
22. Numis AL, Major P, Montenegro MA, Muzykewicz DA, Pulsifer MB, Thiele EA. Identification of risk factors for autism spectrum disorders in tuberous sclerosis complex. *Neurology.* 2011;76:981–7.
23. Kingswood JC, D'Augères GB, Belousova E, Ferreira JC, Carter T, Castellana R, et al. Tuberous Sclerosis registry to increase disease Awareness (TOSCA)—baseline data on 2093 patients. *Orphanet J Rare Dis.* 2017;12:2.
24. Curatolo P, Moavero R, de Vries PJ. Neurological and neuropsychiatric aspects of tuberous sclerosis complex. *Lancet Neurol.* 2015;14:733–45.
25. Chu-Shore CJ, Major P, Camposano S, Muzykewicz D, Thiele EA. The natural history of epilepsy in tuberous sclerosis complex. *Epilepsia.* 2010;51:1236–41.
26. Mitchell RA, Mitchell M, Williams K. The autism spectrum disorder phenotype in children with tuberous sclerosis complex: a systematic review and meta-analysis. *Dev Med Child Neurol.* 2022;64:1214–29.
27. Ebert DH, Greenberg ME. Activity-dependent neuronal signaling and autism spectrum disorder. *Nature.* 2013;493:327–37.
28. Jiang CC, Lin LS, Long S, Ke XY, Fukunaga K, Lu YM, et al. Signalling pathways in autism spectrum disorder: mechanisms and therapeutic implications. *Signal Transduct Target Ther.* 2022;7:229.
29. Capal JK, Bernardino-Cuesta B, Horn PS, Murray D, Byars AW, Bing NM, et al. Influence of seizures on early development in tuberous sclerosis complex. *Epilepsy Behav.* 2017;70:245–52.
30. Nabbout R, Belousova E, Benedik MP, Carter T, Cottin V, Curatolo P, et al. Epilepsy in tuberous sclerosis complex: findings from the TOSCA study. *Epilepsia Open.* 2019;4:73–84.
31. Specchio N, Pietrafusa N, Trivisano M, Moavero R, De Palma L, Ferretti A, et al. Autism and epilepsy in patients with tuberous sclerosis complex. *Front Neurol.* 2020;11:639.
32. Critchley M, Earl C. Tuberous sclerosis and allied conditions. *Brain.* 1932;55:311–46.
33. Kanner L. Autistic disturbances of affective contact. *Nerv Child.* 1943;2:217–50.
34. Moss J, Howlin P. Autism spectrum disorders in genetic syndromes: implications for diagnosis, intervention and understanding the wider autism spectrum disorder population. *J Intellect Disabil Res.* 2009;53:852–73.
35. Waltereit R, Japs B, Schneider M, de Vries PJ, Bartsch D. Epilepsy and *Tsc2* haploinsufficiency lead to autistic-like social deficit behaviors in rats. *Behav Genet.* 2011;41:364–72.
36. Nguyen LH, Mahadeo T, Bordey A. mTOR hyperactivity levels influence the severity of epilepsy and associated neuropathology in an experimental model of tuberous sclerosis complex and focal cortical dysplasia. *J Neurosci.* 2019;39:2762–73.
37. Aronica E, Specchio N, Luinburg MJ, Curatolo P. Epileptogenesis in tuberous sclerosis complex-related developmental and epileptic encephalopathy. *Brain.* 2023;146:2694–710.
38. Chaudry S, Vasudevan N. mTOR-dependent spine dynamics in autism. *Front Mol Neurosci.* 2022;15:877609.
39. Czapski GA, Babiec L, Jęsko H, Gąssowska-Dobrowolska M, Cieślik M, Matuszewska M, et al. Synaptic alterations in a transgenic model of tuberous sclerosis complex: relevance to autism spectrum disorders. *Int J Mol Sci.* 2021;22:10058.
40. Pagani M, Barsotti N, Bertero A, Trakoshis S, Ulysse L, Locarno A, et al. mTOR-related synaptic pathology causes autism spectrum disorder-associated functional hyperconnectivity. *Nat Commun.* 2021;12:6084.
41. Rosina E, Battan B, Siracusano M, Di Criscio L, Hollis F, Pacini L, et al. Disruption of mTOR and MAPK pathways correlates with severity in idiopathic autism. *Transl Psychiatry.* 2019;9:50.
42. Mühlbner A, Bongaarts A, Sarnat HB, Scholl T, Aronica E. New insights into a spectrum of developmental malformations related to mTOR dysregulations: challenges and perspectives. *J Anat.* 2019;235:521–42.
43. Curatolo P, Moavero R, van Scheppingen J, Aronica E. mTOR dysregulation and tuberous sclerosis-related epilepsy. *Expert Rev Neurother.* 2018;18:185–201.
44. Bassetti D, Luhmann HJ, Kirischuk S. Effects of mutations in *TSC* genes on neurodevelopment and synaptic transmission. *Int J Mol Sci.* 2021;22:7273.
45. Bozzi Y, Provenzano G, Casarosa S. Neurobiological bases of autism-epilepsy comorbidity: a focus on excitation/inhibition imbalance. *Eur J Neurosci.* 2018;47:534–48.
46. Gąssowska-Dobrowolska M, Czapski GA, Cieślik M, Zajdel K, Frontczak-Baniewicz M, Babiec L, et al. Microtubule cytoskeletal network alterations in a transgenic model of tuberous sclerosis complex: relevance to autism spectrum disorders. *Int J Mol Sci.* 2023;24:7303.
47. Ebrahimi-Fakhari D, Saffari A, Wahlster L, Di Nardo A, Turner D, Lewis TL, et al. Impaired mitochondrial dynamics and mitophagy in neuronal models of tuberous sclerosis complex. *Cell Rep.* 2016;17:1053–70.
48. Fu C, Cawthon B, Clinkscales W, Bruce A, Winzenburger P, Ess KC. GABAergic interneuron development and function is modulated by the *Tsc1* gene. *Cereb Cortex.* 2012;22:2111–9.
49. Ka M, Smith AL, Kim WY. mTOR controls genesis and autophagy of GABAergic interneurons during brain development. *Autophagy.* 2017;13:1348–63.
50. Amegandjin CA, Choudhury M, Jadhav V, Carriço JN, Quintal A, Berryer M, et al. Sensitive period for rescuing parvalbumin interneurons connectivity and social behavior deficits caused by *TSC1* loss. *Nat Commun.* 2021;12:3653.
51. Yang J, Yang X, Tang K. Interneuron development and dysfunction. *FEBS J.* 2022;289:2318–36.
52. Iannone AF, De Marco García NV. The emergence of network activity patterns in the somatosensory cortex: an early window to autism spectrum disorders. *Neuroscience.* 2021;466:298–309.
53. Eichmüller OL, Corsini NS, Vértesy Á, Morassut I, Scholl T, Gruber VE, et al. Amplification of human interneuron progenitors promotes brain tumors and neurological defects. *Science.* 2022;375:eabf5546.
54. Katsarou AM, Moshé SL, Galanopoulou AS. Interneuronopathies and their role in early life epilepsies and neurodevelopmental disorders. *Epilepsia Open.* 2017;2:284–306.

55. Cherubini E, Di Cristo G, Avoli M. Dysregulation of GABAergic signaling in neurodevelopmental disorders: targeting cation-chloride co-transporters to re-establish a proper E/I balance. *Front Cell Neurosci.* 2021;15:813441.
56. Powell EM. Interneuron development and epilepsy: early genetic defects cause long-term consequences in seizures and susceptibility. *Epilepsy Curr.* 2013;13:172–6.
57. Milovanovic M, Grujicic R. Electroencephalography in assessment of autism spectrum disorders: a review. *Front Psychiatry.* 2021;12:686021.
58. Powell EM, Campbell DB, Stanwood GD, Davis C, Noebels JL, Levitt P. Genetic disruption of cortical interneuron development causes region- and GABA cell type-specific deficits, epilepsy, and behavioral dysfunction. *J Neurosci.* 2003;23:622–31.
59. Dufour BD, McBride E, Bartley T, Juarez P, Martínez-Cerdeño V. Distinct patterns of GABAergic interneuron pathology in autism are associated with intellectual impairment and stereotypic behaviors. *Autism.* 2023;27:1730–45.
60. Righes Marafija J, Vendramin Pasquetti M, Calcagnotto ME. GABAergic interneurons in epilepsy: more than a simple change in inhibition. *Epilepsy Behav.* 2021;121:106935.
61. Ruffolo G, Iyer A, Cifelli P, Roseti C, Mühlebner A, van Scheppingen J, et al. Functional aspects of early brain development are preserved in tuberous sclerosis complex (TSC) epileptogenic lesions. *Neurobiol Dis.* 2016;95:93–101.
62. Vezzani A, Aronica E, Mazarati A, Pittman QJ. Epilepsy and brain inflammation. *Exp Neurol.* 2013;244:11–21.
63. Vezzani A, Ravizza T, Bedner P, Aronica E, Steinhäuser C, Boison D. Astrocytes in the initiation and progression of epilepsy. *Nat Rev Neurol.* 2022;18:707–22.
64. Matta SM, Hill-Yardin EL, Crack PJ. The influence of neuroinflammation in autism spectrum disorder. *Brain Behav Immun.* 2019;79:75–90.
65. Robinson-Agramonte MLA, Noris García E, Fraga Guerra J, Vega Hurtado Y, Antonucci N, Semprún-Hernández N, et al. Immune dysregulation in autism spectrum disorder: what do we know about It? *Int J Mol Sci.* 2022;23:3033.
66. Zahra A, Wang Y, Wang Q, Wu J. Shared etiology in autism spectrum disorder and epilepsy with functional disability. *Behav Neurol.* 2022;2022:5893519.
67. Weichhart T, Hengstschläger M, Linke M. Regulation of innate immune cell function by mTOR. *Nat Rev Immunol.* 2015;15:599–614.
68. Jones RG, Pearce EJ. mTORing immunity: mTOR signaling in the development and function of tissue-resident immune cells. *Immunity.* 2017;46:730–42.
69. Boer K, Crino PB, Gorter JA, Nellist M, Jansen FE, Spliet WGM, et al. Gene expression analysis of tuberous sclerosis complex cortical tubers reveals increased expression of adhesion and inflammatory factors. *Brain Pathol.* 2010;20:704–19.
70. Mills JD, Iyer AM, van Scheppingen J, Bongaarts A, Anink JJ, Janssen B, et al. Coding and small non-coding transcriptional landscape of tuberous sclerosis complex cortical tubers: implications for pathophysiology and treatment. *Sci Rep.* 2017;7:8089.
71. Gruber VE, Luinburg MJ, Colleselli K, Endmayr V, Anink JJ, Zimmer TS, et al. Increased expression of complement components in tuberous sclerosis complex and focal cortical dysplasia type 2B brain lesions. *Epilepsia.* 2022;63:364–74.
72. Zimmer TS, Korotkov A, Zwakenberg S, Jansen FE, Zwartkruis FJT, Rensing NR, et al. Upregulation of the pathogenic transcription factor SPI1/PU.1 in tuberous sclerosis complex and focal cortical dysplasia by oxidative stress. *Brain Pathol.* 2021;31:e12949.
73. Arena A, Zimmer TS, van Scheppingen J, Korotkov A, Anink JJ, Mühlebner A, et al. Oxidative stress and inflammation in a spectrum of epileptogenic cortical malformations: molecular insights into their interdependence. *Brain Pathol.* 2019;29:351–65.
74. Fuso A, Iyer AM, van Scheppingen J, Maccarrone M, Scholl T, Hainfellner JA, et al. Promoter-specific hypomethylation correlates with IL-1 $\beta$  overexpression in tuberous sclerosis complex (TSC). *J Mol Neurosci.* 2016;59:464–70.
75. Prabowo AS, Anink JJ, Lammens M, Nellist M, van den Ouweland AMW, Adle-Biassette H, et al. Fetal brain lesions in tuberous sclerosis complex: TORC1 activation and inflammation. *Brain Pathol.* 2013;23:45–59.
76. Alfano V, Romagnolo A, Mills JD, Cifelli P, Gaeta A, Morano A, et al. Unexpected effect of IL-1 $\beta$  on the function of GABAA receptors in pediatric focal cortical dysplasia. *Brain Sci Brain Sci.* 2022;12:807.
77. Ruffolo G, Alfano V, Romagnolo A, Zimmer T, Mills JD, Cifelli P, et al. GABAA receptor function is enhanced by Interleukin-10 in human epileptogenic gangliogliomas and its effect is counteracted by Interleukin-1 $\beta$ . *Sci Rep.* 2022;12:17956.
78. Palma E, Ruffolo G, Cifelli P, Roseti C, van Vliet EA, Aronica E. Modulation of GABAA receptors in the treatment of epilepsy. *Curr Pharm Des.* 2017;23:5563–8.
79. Usui N, Kobayashi H, Shimada S. Neuroinflammation and oxidative stress in the pathogenesis of autism spectrum disorder. *Int J Mol Sci.* 2023;24:5487.
80. Terrone G, Balosso S, Pauletti A, Ravizza T, Vezzani A. Inflammation and reactive oxygen species as disease modifiers in epilepsy. *Neuropharmacology.* 2020;167:107742.
81. Zimmer TS, Ciriminna G, Arena A, Anink JJ, Korotkov A, Jansen FE, et al. Chronic activation of anti-oxidant pathways and iron accumulation in epileptogenic malformations. *Neuropathol Appl Neurobiol.* 2020;46:546–63.
82. Walma DAC, Yamada KM. The extracellular matrix in development. *Development.* 2020;147:dev175596.
83. Lu P, Takai K, Weaver VM, Werb Z. Extracellular matrix degradation and remodeling in development and disease. *Cold Spring Harb Perspect Biol.* 2011;3:a005058.
84. Dityatev A, Fellin T. Extracellular matrix in plasticity and epileptogenesis. *Neuron Glia Biol.* 2008;4:235–47.
85. Leifeld J, Förster E, Reiss G, Hamad MIK. Considering the role of extracellular matrix molecules, in particular reelin, in granule cell dispersion related to temporal lobe epilepsy. *Front Cell Dev Biol.* 2022;10:917575.
86. Korotkov A, Luinburg MJ, Romagnolo A, Zimmer TS, van Scheppingen J, Bongaarts A, et al. Down-regulation of the brain-specific cell-adhesion molecule contactin-3 in tuberous sclerosis complex during the early postnatal period. *J Neurodev Disord.* 2022;14:8.
87. Broekaart DWM, Scheppingen J, Anink JJ, Wierts L, Hof B, Jansen FE, et al. Increased matrix metalloproteinases expression in tuberous sclerosis complex: modulation by microRNA 146a and 147b in vitro. *Neuropathol Appl Neurobiol.* 2020;46:142–59.
88. Broekaart DW, Bertran A, Jia S, Korotkov A, Senkov O, Bongaarts A, et al. The matrix metalloproteinase inhibitor IPR-179 has antiseizure and antiepileptogenic effects. *J Clin Invest.* 2021;131:e138332.
89. Rogers SL, Rankin-Gee E, Risbud RM, Porter BE, Marsh ED. Normal development of the perineuronal net in humans; in patients with and without epilepsy. *Neuroscience.* 2018;384:350–60.
90. Chaunsali L, Tewari BP, Sontheimer H. Perineuronal net dynamics in the pathophysiology of epilepsy. *Epilepsy Curr.* 2021;21:273–81.
91. Prohl AK, Scherrer B, Tomas-Fernandez X, Davis PE, Filip-Dhima R, Prabhu SP, et al. Early white matter development is

- abnormal in tuberous sclerosis complex patients who develop autism spectrum disorder. *J Neurodev Disord.* 2019;11:36.
92. Gruber VE, Lang J, Endmayr V, Diehm R, Pimpel B, Glatter S, et al. Impaired myelin production due to an intrinsic failure of oligodendrocytes in mTORopathies. *Neuropathol Appl Neurobiol.* 2021;47:812–25.
  93. Mühlebner A, van Scheppingen J, de Neef A, Bongaarts A, Zimmer TS, Mills JD, et al. Myelin pathology beyond white matter in tuberous sclerosis complex (TSC) cortical tubers. *J Neuropathol Exp Neurol.* 2020;79:1054–64.
  94. Zonouzi M, Berger D, Jokhi V, Kedaigle A, Lichtman J, Arlotta P. Individual oligodendrocytes show bias for inhibitory axons in the neocortex. *Cell Rep.* 2019;27:2799–808.e3.
  95. Fang LP, Zhao N, Caudal LC, Chang HF, Zhao R, Lin CH, et al. Impaired bidirectional communication between interneurons and oligodendrocyte precursor cells affects social cognitive behavior. *Nat Commun.* 2022;13:1394.
  96. Galvez-Contreras AY, Zarate-Lopez D, Torres-Chavez AL, Gonzalez-Perez O. Role of oligodendrocytes and myelin in the pathophysiology of autism spectrum disorder. *Brain Sci.* 2020;10:951.
  97. Bromfield EB, Cavazos JE, Sirven JI. An introduction to epilepsy. London: Routledge; 2006.
  98. Specchio N, Curatolo P. Developmental and epileptic encephalopathies: what we do and do not know. *Brain.* 2021;144:32–43.
  99. Scheffer IE, Liao J. Deciphering the concepts behind “Epileptic encephalopathy” and “Developmental and epileptic encephalopathy”. *Eur J Paediatr Neurol.* 2020;24:11–4.
  100. Specchio N, Wirrell EC, Scheffer IE, Nabbout R, Riney K, Samia P, et al. International league against epilepsy classification and definition of epilepsy syndromes with onset in childhood: position paper by the ILAE task force on nosology and definitions. *Epilepsia.* 2022;63:1398–442.
  101. Moavero R, Mühlebner A, Luinenburg MJ, Craiu D, Aronica E, Curatolo P. Genetic pathogenesis of the epileptogenic lesions in tuberous sclerosis complex: therapeutic targeting of the mTOR pathway. *Epilepsy Behav.* 2022;131:107713.
  102. Ehninger D, Han S, Shilyansky C, Zhou Y, Li W, Kwiatkowski DJ, et al. Reversal of learning deficits in a *Tsc2<sup>+/-</sup>* mouse model of tuberous sclerosis. *Nat Med.* 2008;14:843–8.
  103. Talos DM, Sun H, Kosaras B, Joseph A, Folkert RD, Poduri A, et al. Altered inhibition in tuberous sclerosis and type IIb cortical dysplasia. *Ann Neurol.* 2012;71:539–51.
  104. Sato A, Kasai S, Kobayashi T, Takamatsu Y, Hino O, Ikeda K, et al. Rapamycin reverses impaired social interaction in mouse models of tuberous sclerosis complex. *Nat Commun.* 2012;3:1292.
  105. Way SW, Rozas NS, Wu HC, McKenna J, Reith RM, Hashmi SS, et al. The differential effects of prenatal and/or postnatal rapamycin on neurodevelopmental defects and cognition in a neuroglial mouse model of tuberous sclerosis complex. *Hum Mol Genet.* 2012;21:3226–36.
  106. Tsai PT, Hull C, Chu Y, Greene-Colozzi E, Sadowski AR, Leech JM, et al. Autistic-like behaviour and cerebellar dysfunction in Purkinje cell *Tsc1* mutant mice. *Nature.* 2012;488:647–51.
  107. Schneider M, de Vries PJ, Schöning K, Rößner V, Waltereit R. mTOR inhibitor reverses autistic-like social deficit behaviours in adult rats with both *Tsc2* haploinsufficiency and developmental status epilepticus. *Eur Arch Psychiatry Clin Neurosci.* 2017;267:455–63.
  108. Petrasko T, Vojtechova I, Klovrcza O, Tuckova K, Vejmla C, Rak J, et al. mTOR inhibitor improves autistic-like behaviors related to *Tsc2* haploinsufficiency but not following developmental status epilepticus. *J Neurodev Disord.* 2021;13:14.
  109. Koike-Kumagai M, Fujimoto M, Wataya-Kaneda M. Sirolimus relieves seizures and neuropsychiatric symptoms via changes of microglial polarity in tuberous sclerosis complex model mice. *Neuropharmacology.* 2022;218:109203.
  110. Kashii H, Kasai S, Sato A, Hagino Y, Nishito Y, Kobayashi T, et al. *Tsc2* mutation rather than *Tsc1* mutation dominantly causes a social deficit in a mouse model of tuberous sclerosis complex. *Hum Genomics.* 2023;17:4.
  111. McMahon JJ, Yu W, Yang J, Feng H, Helm M, McMahon E, et al. Seizure-dependent mTOR activation in 5-HT neurons promotes autism-like behaviors in mice. *Neurobiol Dis.* 2015;73:296–306.
  112. Ruffolo G, Gaeta A, Cannata B, Pinzaglia C, Aronica E, Morano A, et al. GABAergic neurotransmission in human tissues is modulated by cannabidiol. *Life (Basel).* 2022;12:2042.
  113. Farach LS, Richard MA, Lupo PJ, Sahin M, Krueger DA, Wu JY, et al. Epilepsy risk prediction model for patients with tuberous sclerosis complex. *Pediatr Neurol.* 2020;113:46–50.
  114. Ogórek B, Hamieh L, Hulshof HM, Lasseter K, Klonowska K, Kuijff H, et al. *TSC2* pathogenic variants are predictive of severe clinical manifestations in TSC infants: results of the EPISTOP study. *Genet Med.* 2020;22:1489–97.
  115. Hulshof HM, Kuijff HJ, Kotulska K, Curatolo P, Weschke B, Riney K, et al. Association of early MRI characteristics with subsequent epilepsy and neurodevelopmental outcomes in children with tuberous sclerosis complex. *Neurology.* 2022;98:e1216–25.
  116. Ruppe V, Dilsiz P, Reiss CS, Carlson C, Devinsky O, Zagzag D, et al. Developmental brain abnormalities in tuberous sclerosis complex: a comparative tissue analysis of cortical tubers and perituberal cortex. *Epilepsia.* 2014;55:539–50.
  117. Scherrer B, Prohl AK, Taquet M, Kapur K, Peters JM, Tomas-Fernandez X, et al. The connectivity fingerprint of the fusiform gyrus captures the risk of developing autism in infants with tuberous sclerosis complex. *Cereb Cortex.* 2020;30:2199–214.
  118. Cohen AL, Kroeck MR, Wall J, McManus P, Ovchinnikova A, Sahin M, et al. Tubers affecting the fusiform face area are associated with autism diagnosis. *Ann Neurol.* 2023;93:577–90.
  119. Sato A, Tominaga K, Iwatani Y, Kato Y, Wataya-Kaneda M, Makita K, et al. Abnormal white matter microstructure in the limbic system is associated with tuberous sclerosis complex-associated neuropsychiatric disorders. *Front Neurol.* 2022;13:782479.
  120. Vanes LD, Tye C, Tournier JD, Combes AJE, Shephard E, Liang H, et al. White matter disruptions related to inattention and autism spectrum symptoms in tuberous sclerosis complex. *NeuroImage Clin.* 2022;36:103163.
  121. Peters JM, Prohl A, Kapur K, Nath A, Scherrer B, Clancy S, et al. Longitudinal effects of everolimus on white matter diffusion in tuberous sclerosis complex. *Pediatr Neurol.* 2019;90:24–30.
  122. Srivastava S, Prohl AK, Scherrer B, Kapur K, Krueger DA, Warfield SK, et al. Cerebellar volume as an imaging marker of development in infants with tuberous sclerosis complex. *Neurology.* 2018;90:e1493–500.
  123. Moavero R, Napolitano A, Cusmai R, Vigevano F, Figà-Talamanca L, Calbi G, et al. White matter disruption is associated with persistent seizures in tuberous sclerosis complex. *Epilepsy Behav.* 2016;60:63–7.
  124. Cook IA, Wilson AC, Peters JM, Goyal MN, Bebin EM, Northrup H, et al. EEG spectral features in sleep of autism spectrum disorders in children with tuberous sclerosis complex. *J Autism Dev Disord.* 2020;50:916–23.
  125. Zhang B, Guo D, Han L, Rensing N, Satoh A, Wong M. Hypothalamic orexin and mechanistic target of rapamycin activation mediate sleep dysfunction in a mouse model of tuberous sclerosis complex. *Neurobiol Dis.* 2020;134:104615.
  126. Elkhatib Smidt SD, Ghorai A, Taylor SC, Gehring BN, Dow HC, Langer A, et al. The relationship between autism spectrum and sleep-wake traits. *Autism Res.* 2022;15:641–52.

127. Schoenberger A, Capal JK, Ondracek A, Horn PS, Murray D, Byars AW, et al. Language predictors of autism spectrum disorder in young children with tuberous sclerosis complex. *Epilepsy Behav.* 2020;103:106844.
128. Scheper M, Romagnolo A, Besharat ZM, Iyer AM, Moavero R, Hertzberg C, et al. MiRNAs and isomiRs: serum-based biomarkers for the development of intellectual disability and autism spectrum disorder in tuberous sclerosis complex. *Biomedicines.* 2022;10:1838.
129. Moavero R, Kotulska K, Lagae L, Benvenuto A, Emberti Gialloreti L, Weschke B, et al. Is autism driven by epilepsy in infants with tuberous sclerosis complex? *Ann Clin Transl Neurol.* 2020;7:1371–81.
130. Talbott MR, Miller MR. Future directions for infant identification and intervention for autism spectrum disorder from a transdiagnostic perspective. *J Clin Child Adolesc Psychol.* 2020;49:688–700.
131. Randall M, Egberts KJ, Samtani A, Scholten RJ, Hooft L, Livingstone N, et al. Diagnostic tests for autism spectrum disorder (ASD) in preschool children. *Cochrane Database Syst Rev.* 2018;2018:CD009044.
132. Capal JK, Horn PS, Murray DS, Byars AW, Bing NM, Kent B, et al. Utility of the autism observation scale for infants in early identification of autism in tuberous sclerosis complex. *Pediatr Neurol.* 2017;75:80–6.
133. Moavero R, Benvenuto A, Emberti Gialloreti L, Siracusano M, Kotulska K, Weschke B, et al. Early clinical predictors of autism spectrum disorder in infants with tuberous sclerosis complex: results from the EPISTOP Study. *J Clin Med.* 2019;8:788.
134. Jeste SS, Sahin M, Bolton P, Ploubidis GB, Humphrey A. Characterization of autism in young children with tuberous sclerosis complex. *J Child Neurol.* 2008;23:520–5.
135. Zachor DA, Curatolo P. Recommendations for early diagnosis and intervention in autism spectrum disorders: an Italian-Israeli consensus conference. *Eur J Paediatr Neurol.* 2014;18:107–18.
136. Wu JY, Goyal M, Peters JM, Krueger D, Sahin M, Northrup H, et al. Scalp EEG spikes predict impending epilepsy in TSC infants: a longitudinal observational study. *Epilepsia.* 2019;60:2428–36.
137. Nabbout R, Kuchenbuch M, Chiron C, Curatolo P. Pharmacotherapy for seizures in tuberous sclerosis complex. *CNS Drugs.* 2021;35:965–83.
138. Curatolo P, Verdecchia M, Bombardieri R. Vigabatrin for tuberous sclerosis complex. *Brain Dev.* 2001;23:649–53.
139. van der Poest CE, Jansen FE, Braun KPJ, Peters JM. Update on drug management of refractory epilepsy in tuberous sclerosis complex. *Pediatr Drugs.* 2020;22:73–84.
140. Bombardieri R, Pinci M, Moavero R, Cerminara C, Curatolo P. Early control of seizures improves long-term outcome in children with tuberous sclerosis complex. *Eur J Paediatr Neurol.* 2010;14:146–9.
141. Kotulska K, Kwiatkowski DJ, Curatolo P, Weschke B, Riney K, Jansen F, et al. Prevention of epilepsy in infants with tuberous sclerosis complex in the EPISTOP trial. *Ann Neurol.* 2021;89:304–14.
142. Zhang L, Huang CC, Dai Y, Luo Q, Ji Y, Wang K, et al. Correction: Symptom improvement in children with autism spectrum disorder following bumetanide administration is associated with decreased GABA/glutamate ratios. *Transl Psychiatry.* 2020;10:63.
143. van Andel DM, Sprengers JJ, Oranje B, Scheepers FE, Jansen FE, Bruining H. Effects of bumetanide on neurodevelopmental impairments in patients with tuberous sclerosis complex: an open-label pilot study. *Mol Autism.* 2020;11:30.
144. Juarez-Martinez EL, Sprengers JJ, Cristian G, Oranje B, van Andel DM, Avramiea AE, et al. Prediction of behavioral improvement through resting-state electroencephalography and clinical severity in a randomized controlled trial testing bumetanide in autism spectrum disorder. *Biol Psychiatry Cogn Neurosci Neuroimaging.* 2023;8:251–61.
145. Overwater IE, Rietman AB, Mous SE, Bindels-de Heus K, Rizopoulos D, ten Hoopen LW, et al. A randomized controlled trial with everolimus for IQ and autism in tuberous sclerosis complex. *Neurology.* 2019;93:e200–9.
146. Krueger DA, Sadhwani A, Byars AW, de Vries PJ, Franz DN, Whittemore VH, et al. Everolimus for treatment of tuberous sclerosis complex-associated neuropsychiatric disorders. *Ann Clin Transl Neurol.* 2017;4:877–87.
147. Krueger DA, Capal JK, Curatolo P, Devinsky O, Ess K, Tzadok M, et al. Short-term safety of mTOR inhibitors in infants and very young children with tuberous sclerosis complex (TSC): multicentre clinical experience. *Eur J Paediatr Neurol.* 2018;22:1066–73.
148. Gelot AB, Represa A. Progression of fetal brain lesions in tuberous sclerosis complex. *Front Neurosci.* 2020;14:899.
149. Cavalheiro S, da Costa MDS, Richtmann R. Everolimus as a possible prenatal treatment of in utero diagnosed subependymal lesions in tuberous sclerosis complex: a case report. *Childs Nerv Syst.* 2021;37:3897–9.
150. Nithianantharajah J, Hannan AJ. Enriched environments, experience-dependent plasticity and disorders of the nervous system. *Nat Rev Neurosci.* 2006;7:697–709.
151. McDonald NM, Hyde C, Choi AB, Gulsrud AC, Kasari C, Nelson CA, et al. Improving developmental abilities in infants with tuberous sclerosis complex: a pilot behavioral intervention study. *Infants Young Child.* 2020;33:108–18.
152. Kasari C. Update on behavioral interventions for autism and developmental disabilities. *Curr Opin Neurol.* 2015;28:124–9.
153. Bruni O, Cortesi F, Giannotti F, Curatolo P. Sleep disorders in tuberous sclerosis: a polysomnographic study. *Brain Dev.* 1995;17:52–6.
154. Bruni O, Alonso-Alconada D, Besag F, Biran V, Braam W, Cortese S, et al. Current role of melatonin in pediatric neurology: clinical recommendations. *Eur J Paediatr Neurol.* 2015;19:122–33.
155. Jansen FE, Van Huffelen AC, Algra A, Van Nieuwenhuizen O. Epilepsy surgery in tuberous sclerosis: a systematic review. *Epilepsia.* 2007;48:1477–84.
156. Specchio N, Pepi C, de Palma L, Moavero R, De Benedictis A, Marras CE, et al. Surgery for drug-resistant tuberous sclerosis complex-associated epilepsy: who, when, and what. *Epileptic Disord.* 2021;23:53–73.

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