



# Application of Response Surface Methodology to Supercritical CO<sub>2</sub> Extraction: Case Study on Coextraction of Carotenoids and Oil from Rosehip Shells and Seeds

Edgar Uquiche<sup>1</sup> · Julián Arias<sup>2</sup> · José Manuel del Valle<sup>3</sup>

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## Abstract

Response surface methodology (RSM) is an experimental strategy widely used as a research tool in investigation. We reviewed 89 papers that used RSM to study the extraction of oils or minor lipids, using supercritical (SC) CO<sub>2</sub>, and observed that most of these studies have not contributed to an understanding of the extraction phenomenon, by neglecting prior knowledge on mass transfer or equilibrium relationships. We used the extraction of carotenoids from rosehip shells and oil from the seeds, as a case study to illustrate an improved strategy to apply RSM to oil-aided SC-CO<sub>2</sub> extraction of high-molecular-weight nonpolar solutes, such as carotenoids. We selected the temperature and density to characterize the effect of solvent conditions, the specific CO<sub>2</sub> consumption to characterize the interaction of solvent time and solvent power, and the percentage of seeds in the composite substrate to characterize the cosolvent effect of the oil. A rotatable central composite design was applied sequentially in three blocks, where the third block allowed incorporating quadratic coefficients to adequately describe the non-linear behavior of the responses.

**Keywords** Carotenoids · Extrusion · Design of experiments · *Rosa aff. rubiginosa* · *Rosa canina* L. · Supercritical extraction · Vegetable oil cosolvent

## Introduction

Reputed food engineering journals should generally reject conventional articles on the use of response surface methodology (RSM) to analyze the supercritical (SC) carbon dioxide (CO<sub>2</sub>) extraction of vegetable substrates or microalgae. Indeed, using RSM provides authors with optimized conditions for the selected range of operating variables, but fails to provide an understanding of the underlying mechanisms and lacks sufficient depth in engineering science. Sharif et al. [1] published a highly quoted review article on the

application of RSM to SC-CO<sub>2</sub> extraction processes that we will use as a reference point to back up the claim that heads this paragraph.

To the best of the knowledge of the authors, del Valle and Aguilera [2] were the first to apply RSM to the analysis of SC-CO<sub>2</sub> extraction processes, but this was due more to limitations of the experimental device than a consequence of careful planning. The authors performed SC-CO<sub>2</sub> extraction of mushroom oleoresin in a 1-gallon batch reactor that did not dispose of a high-pressure pump and had the CO<sub>2</sub> density ( $\rho$ ) and system temperature ( $T$ ) as the independent variables defining the conditions in the reactor. CO<sub>2</sub> density was controlled by adjusting the amount of liquid CO<sub>2</sub> loaded in the extraction vessel that was fed from a high-pressure cylinder equipped with a dip tube while gaseous CO<sub>2</sub> was continuously removed from the upper end of the reactor through a tubing that projected a measured length into the upper portion of the reactor. The supercritical conditions in the reactor were achieved by heating, and attainment of the desired  $\rho$ - $T$  conditions was confirmed by measuring the temperature and pressure ( $P$ ) in the reactor during extraction and using PVT tables for CO<sub>2</sub> to interpolate the corresponding values

✉ José Manuel del Valle  
delvalle@ing.puc.cl

<sup>1</sup> Departamento de Ingeniería Química, Center of Food Biotechnology and Bioseparations, Universidad de La Frontera, BIOREN, Temuco, Chile

<sup>2</sup> Escuela de Ingeniería Química, Universidad Industrial de Santander, Bucaramanga, Colombia

<sup>3</sup> Departamento de Ingeniería Química y Bioprocesos, Pontificia Universidad Católica de Chile, Avda. Vicuña Mackenna 4860, Macul, Santiago, Chile

of  $\rho$ . Dried *Suillus luteus* L. mushrooms used as substrate were ground, size-classified ( $-35 + 100$  mesh), moistened to  $130 \text{ g kg}^{-1}$  water/substrate, and uniaxially compressed in a 53-mm ID cylindrical mold to produce disks of 24-mm height in an Instron Universal Testing Machine operating with a flat plunger at 6 mm/s. The third experimental variable was the specific volume of the substrate, which was adjusted by varying the load of conditioned powder loaded to the compression cell. The compressed substrate disks were placed between flat plates and attached to the upper lid of the reaction vessel above the level of the liquid  $\text{CO}_2$  in all experiments. Following completion of extraction, after 6 h, the loaded supercritical phase was expanded to a second 1-gallon high-pressure reaction vessel filled with gaseous  $\text{CO}_2$  at room temperature that was slowly depressurized. Because of  $\text{CO}_2$  expansion, the solubilized mushroom oleoresin came out of the solution and precipitated out both in the extraction reactor and separation reactor. Thus, the oleoresin yield (the response variable) experiments were not determined based on the recovered extract in the baskets placed at the bottom of the two reactors, but on the assessment of the weight and moisture losses of substrate disks. del Valle and Aguilera [2] used a three-factor fractional factorial design of experiments (DoE) consisting of 18 runs at 16 different experimental conditions.

About 15 years after this first study, the corresponding author was back in Pontificia Universidad Católica (UC) de Chile as an assistant professor, when the *Laboratorio de Extracción de Materiales Biologicos* (LEMAB) procured a Process Development Unit (PDU) from Thar Technologies (Pittsburgh, PA) to carry out research on SC- $\text{CO}_2$  extraction processes from food materials. The first published work in this second stage was on the SC- $\text{CO}_2$  extraction of rosehip seed oil using RSM [3], which will be reviewed in “[Application of RSM on the Supercritical  \$\text{CO}\_2\$  Extraction of Oilseeds](#)” section. At that time, about 20 years ago, the WoS database reported only five other works on the application of RSM or DoEs to SC- $\text{CO}_2$  extraction processes in the Food Science Technology field.

Since 2000, the application of RSM and DoE in the field of food engineering, particularly in the context of SC- $\text{CO}_2$  extraction of vegetable substrates and microalgae, has gained significant attention. While RSM enables the determination of optimized conditions for a given range of operating variables, a debate persists regarding its limitations in providing a comprehensive understanding of the underlying mechanisms and engineering principles. In light of this, the primary objective of this review is to comprehensively discuss the utilization of RSM and DoE in SC- $\text{CO}_2$  extraction, specifically focusing on the extraction of vegetable oils and associated minor lipids from seeds and related substrates.

To achieve this goal, the review is structured as follows. “[RSM and its Application in Food Engineering](#)” section

provides a brief overview of RSM and DoE applications in the field of food engineering. “[Application of RSM on the Supercritical  \$\text{CO}\_2\$  Extraction of Oilseeds](#)” section describes and analyzes selected studies that have employed RSM for the SC- $\text{CO}_2$  extraction of oils and minor lipids from seeds and related substrates, highlighting both their advantages and limitations, as well as the importance of appropriate selection of dependent and independent variables, along with suitable experimental design strategies. “[Effect of Process Variables on the Supercritical  \$\text{CO}\_2\$  Extraction of Oil in Seeds](#)” section discusses the key variables that influence the supercritical extraction of vegetable oils from seeds. Furthermore, “[Supercritical  \$\text{CO}\_2\$  Extraction of Oils and Carotenoids in Rosehip Seeds and Shells](#)” section presents a state-of-the-art compilation, highlighting the opportunity to enhance the extraction of oils and carotenoids from rosehip seeds by including shells as part of the substrate. The subsequent section, “[Case Study](#)” section, serves as a case study, presenting an experimental work and statistical analysis of results for the coextraction of carotenoids and oil from rosehip shells and seeds. By addressing these topics, this review aims to provide a critical and up-to-date perspective on the application of RSM and DoE in SC- $\text{CO}_2$ , while also serving as a valuable reference and practical resource for researchers and professionals seeking to optimize the extraction processes of oils and minor lipids from seeds and related substrates. Finally, “[Conclusions](#)” section summarizes the main conclusion of this manuscript.

## Literature Search

As indicated above, this section will expand the “[Introduction](#)” section, thus is just a justification of this review article from the standpoint of the experience of one of the authors, to briefly describe the application of RSM and DoE in the food engineering field, to discuss and describe 89 works in literature on the application of these methodologies to the SC- $\text{CO}_2$  extraction of oil and minor lipids from seeds and related substrates, and to present and discuss the main variables affecting the SC- $\text{CO}_2$  extraction of oil in seeds, in general, as well as the extraction of oil and carotenoids in rosehip seeds and shells, specifically. As part of “[Supercritical  \$\text{CO}\_2\$  Extraction of Oils and Carotenoids in Rosehip Seeds and Shells](#)” section, we will justify the case study we devised for an improved analysis of RSM and DoE in the food engineering field that readers should consider in planning their experimental studies in the future.

## RSM and its Application in Food Engineering

DoE and RSM are widely used research methods that use statistical and model-fitting tools to predict one or more

dependent variables (responses) as a function of several independent variables (factors). These methods should be used mainly in those cases where an analytical description of the phenomenon under study is not yet available, and it is presumed that the change in the factors produces relatively mild alterations in the response(s) within an experimental region. The resulting models are categorized as empirical, or result-driven models, due to their lack of underlying theory and their limited ability to predict responses outside of the range of data that was used to fit the model.

DoEs were adapted from agricultural research to quality control in industry in the 1920s as reviewed by Bisgaard [4]. More recent authoritative reviews on the subject were published by Tanco et al. [5] and by de Oliveira et al. [6] that focus more specifically on the use of central composite designs (CCDs), of the type that will be used in our study, and data analysis using RSM. The reader interested on DoEs and RSM may consult the books or monographs of Haaland [7], Box et al. [8], Box and Draper [9], Gacula et al. [10], Myers et al. [11], Montgomery [12], Jones and Montgomery [13], and NIST/SEMATECH [14]. Response surface methodology has a major advantage over the one-factor-at-a-time approach in that it allows the evaluation of the effect of multiple factors (or independent variables) and their interactions on the response(s) [or dependent variable(s)] with a reduced number of trials. Interaction between independent variables is manifested when one factor does not produce the same effect on the response at different levels of another factor.

RSM studies allow (1) understanding the topography of the response surface (local maximum, local minimum, ridge lines); and (2) finding the region where the optimal response occurs. The goal is to move rapidly and efficiently along a path to get to a maximum or a minimum response so that the response is optimized. The methodology involves three steps [11]: (1) experimental design in which the independent variables and their experimental levels are set using well-established statistical experimental designs such as CCD; (2) response surface modeling through regression analysis; and (3) process optimization using the response surface models.

An important aspect of RSM is the DoE [9]. The objective of DoEs is the selection of experimental conditions where the response should be evaluated. Most of the criteria for optimal DoEs are associated with the mathematical model of the process. Generally, these mathematical models are polynomials with an unknown structure, so that the corresponding experiments are designed only for every particular problem. The choice of the DoEs can have a large influence on the accuracy of the approximation and the cost of constructing the response surface.

Table 1 summarizes the number of entries on DoE or RSM in the database of WoS under the category of Food Science Technology. About one-third of the contributions relate to extraction, and most of the 2240 entries on this

**Table 1** An overview of topics covered by articles indexed to the Web of Science in the “Food Science Technology” category when “Response Surface Methodology” OR “Design of Experiments” was queried in September 2021

Topic in Food Science	Number of articles
Extraction	2,240
Formulation	726
Fermentation	646
Reaction	621
Storage	614
Sensory analysis	498
Drying	454
Separation	435
Extrusion	308
Packaging	200
Thermal processing	162
Total articles 6,486	

subject relate in turn to emerging technologies such as those on subcritical or hot-pressurized water (29% of total), ultrasound-assisted (24%), microwave-assisted (16%), or supercritical CO<sub>2</sub> extraction (9%) processes. Extraction is followed by formulation, fermentation, reaction, and storage with about 10% each. Then, the subjects’ sensory analysis, drying, and separation account for 7–8% each. Finally, extrusion, packaging, and thermal processing account for less than 5% each. Consequent to our observations in Table 1, Yolmeh and Jafari [15], on reviewing the applications of RSM in the food industry, highlight extraction, drying, and production of microbial enzymes and some other metabolites (all supported by extensive tabulations of practical examples), besides formulation processes, microencapsulation, and controlled-release processes, optimization of enzymatic hydrolysis and clarification, and blanching (a thermal process).

In “Application of RSM on the Supercritical CO<sub>2</sub> Extraction of Oilseeds” section, we analyze critically the application of DoE and RSM to the SC-CO<sub>2</sub> extraction of oil from seeds. This subject has been implicitly analyzed in reviews focussing on experimental designs on SCFE [1], the supercritical fluid extraction (SCFE) of vegetable matrices [16], or SCFE of seed oils [17].

### Application of RSM on the Supercritical CO<sub>2</sub> Extraction of Oilseeds

To ensure a comprehensive and focused review, we employed a specific search equation on the Web of Science (Date 08/06/2021). The search equation used was “supercritical CO<sub>2</sub> extraction “ (searched in all fields) combined with “Response Surface Methodology” (searched in

all fields), “oil” (searched in all fields), “seed” (searched in all fields), “Proceeding Paper” or “Book Chapters” or “Review Article” (document types excluded) and “Food Science Technology” or “Engineering Chemical” (Web of Science categories). This search equation allowed us to narrow down our selection criteria and identify relevant articles that addressed the specific aspects of our study. By combining these keywords and filters, we aimed to retrieve articles that focused on supercritical CO<sub>2</sub> extraction, utilizing response surface methodology, and targeting applications in food science, technology, and chemical engineering with a specific emphasis on seed-related research. Through this rigorous search strategy, we obtained a collection of 89 scientific articles that met our predefined criteria. Appendix A in Supplementary Material lists all plant-derived substrates extracted in the selected works. Overall, 90 substrates, mostly seeds (84%), followed by byproducts (11%), and whole fruits (5%), were studied. The most popular was flaxseed (*Linum usitatissimum*), which appeared in six contributions.

From the chosen articles, we extracted valuable insights into the authors’ preferences concerning the factors and observables they prioritize when utilizing response surface methodology. Additionally, we captured information about their experimental strategies, considering the diverse options available for organizing experiments within the framework of response surface methodology. Table 2 summarizes the most studied factors or independent variables of the selected works. Most designs have three independent variables (60%)

followed by four (21%), five (12%), and two (7%). Regardless of the number of factors, all the RSM studies reported in Table 2 used pressure ( $P$ ) as an independent variable, and all but two also used temperature ( $T$ ).  $P$  and  $T$  are by far the thermodynamic variables preferred by researchers for exploring the region where the solvent or solvent mixture is a supercritical fluid. The convenience of replacing one of these thermodynamic variables by another, like density, that might be more suitable will be discussed later. Thirty-nine studies considered the CO<sub>2</sub> flow rate ( $Q$ ) and 36 the dynamic extraction time ( $t_d$ ) as independent variables, while 11 considered both. With 11 occurrences, the fifth most used factor was particle size ( $d_p$ ), a variable related to the inner resistance to mass transfer. Seventeen percent of the works considered the modifier concentration ( $MC$ ) as a factor, in that the modifier is generally added to increase the yield of some minor polar and/or high-molecular-weight substrate component. Finally, the specific CO<sub>2</sub> consumption ( $q$ ) and static extraction time ( $t_e$ ) were the least considered factors with 7 and 4 occurrences.

Six works of the 88 that we analyzed in our study are not listed in Table 2. These works are those that used one or more factors with only one occurrence. Jiang and Niu [58], for example, in addition to some of the factors mentioned in Table 2, included the substrate moisture in their preliminary trials; Wang et al. [99] used the amount of substrate loaded in the extractor ( $M_s$ ); and Bernardo-Gil et al. [100] conveniently used superficial CO<sub>2</sub> velocity ( $U$ ), an intensive variable

**Table 2** Literature search on the extraction of vegetable oils using supercritical CO<sub>2</sub>

Number of factors	Count*	Factors or independent variables								References
		$P$	$T$	$t_d$	$Q$	$d_p$	$MC$	$q$	$t_e$	
5	2	✓	✓	✓	✓	✓				[18, 19]
	2	✓	✓	✓			✓		✓	[20, 21]
	6	✓	✓		✓	✓	✓			[22–27]
4	9	✓	✓	✓	✓					[28–36]
	3	✓	✓	✓						[37–39]
	1	✓	✓	✓						[40]
	5	✓	✓			✓				[41–45]
	17	✓	✓	✓						[3, 46–61]
3	16	✓	✓		✓					[62–77]
	2	✓	✓			✓				[78, 79]
	4	✓	✓				✓			[80–83]
	7	✓	✓					✓		[84–90]
	1	✓		✓			✓			[91]
	1	✓		✓					✓	[92]
2	6	✓	✓							[93–98]
Total	82	82	80	36	38	18	13	7	4	

Most studied factors and their concurrence in studies that employ response surface methodology. ( $P$ ) pressure, ( $T$ ) temperature, ( $t_d$ ) dynamic extraction time, ( $Q$ ) CO<sub>2</sub> flow rate, ( $d_p$ ) particle size, ( $MC$ ) modifier concentration, ( $q$ ) specific CO<sub>2</sub> consumption, and ( $t_e$ ) static time

\*Count of studies with the same factors

that does not depend on the geometry or size of the extraction vessel. These less frequently studied variables encompass phenomena associated with substrate composition, bed density, and flow effects from a scalability perspective. Although these phenomena hold significance for scalability based on theoretical models, which highlight the importance of assessing the impact of their variations on model performance, the authors have demonstrated less interest in investigating or comprehending their effects. This disparity could be attributed to the predominant focus, observed in most papers, on exploring new sources of vegetable oils, rather than optimizing the extraction process for promising well-studied vegetable oils through a comprehensive understanding of the underlying transfer phenomena. Based on our experience, we have observed a limited number of published works on RSM employing factors that could contribute novel experimental information to the understanding of phenomena that are not yet sufficiently well described for the SC-CO<sub>2</sub> extraction of vegetable oils. Mixing speed and ultrasonic power [101], substrate pretreatment [102], and dispersion of the substrate in an inert matrix [103] may contribute to the understanding of SC-CO<sub>2</sub> extraction of oil from seeds but are rarely studied in RSM and DoE. Appendix A in Supplementary Material also details the selected responses: 89% of the works evaluated oil yield as a response, 34% evaluated the yield of a minor compound in the oil, 12% evaluated percent recovery, and 11% evaluated antioxidant activity. A response variable that was found with few occurrences (three times) in the review, but that may be very useful from the standpoint of describing the rate of the extraction process, was the slope of the first part of the cumulative extraction curve. The idea was used by Bernardo-Gil et al. [100], and then by Comin et al. [102]. Finally, Bojanić et al. [63] fitted semi-empirical models to extraction curves to obtain the initial slope values from best-fit parameters values. (This will be illustrated in “Effect of Process Variables on the Supercritical CO<sub>2</sub> Extraction of Oil in Seeds” section for the model of Naik et al. [104]).

Appendix B in Supplementary Material shows experimental designs used by researchers in their RSM applications. Among them, the central composite design (CCD) (35 occurrences) and the Box–Behnken design (BBD) (29 occurrences) stand out, both with characteristics of rotatability (or near-rotatability) appropriate for the application of RSM [11]. Other less popular experimental designs include the factorial (10 occurrences), face-centered (4), factorial multilevel (1), and fractional factorial (1) ones. More than half of these consulted manuscripts (52) studied 3 experimental factors, 20 studied 4, 11 studied 5, and only 6 papers studied 2 experimental factors. The advantage of the CCD lies in the sequential nature of its implementation, which allows experiments to be added to the factorial design (FD) as notions of lack of fit are observed, i.e., it is possible to

divide the design into blocks that do not affect the estimates of the coefficients in the second-order model (orthogonal blocking) [14]. On the other hand, the advantage of the BBD design is that they require fewer treatment combinations than CCDs in cases involving 3 or 4 factors. However, BBDs have a limited capacity for orthogonal blocking compared to CCDs, and also contain corner regions of poor predictive capability in corners [14].

Appendix B in Supplementary Material groups the works in Appendix A into two categories according to their implementation: those that used optimization designs or factorial designs, and those that screened factors prior to optimization experiments. The prescreening approach is convenient when studying many factors because it allows determining the significance of the effect of the factors on the response(s) before considering factor optimization. Examples of applications of the screening plus optimization approach include the one-factor-at-a-time (OFT) plus BBD with six occurrences, OFT + CCD with four, and fractional factorial design + CCD and factorial design + OFT with one each. This sequential approach may allow reducing the number of experiments without losing statistically relevant information. However, despite its advantages, four times less (6) occurrences of the prescreening approach were carried out than the approach of performing complete designs (25 occurrences) when studying the effect of 4 or 5 factors. Solving a 5-factor BBD or CCD involves performing 46 or 52 experiments, respectively. When performing prescreening experiments prior to optimization experiments, this number of experiments could be considerably reduced. Such could have been the case in the study conducted by Suryawanshi and Mohanty [23] who solved a five-factor BBD ( $T$ ,  $P$ ,  $d_p$ ,  $Q$ , and  $MC$ ) for the SC-CO<sub>2</sub> extraction of *Argemone mexicana* where they found that  $T$  and  $Q$  did not have a significant effect on extraction yield. If the authors had employed the prescreening approach using, for example, a fractional factorial design of resolution III, i.e., a  $2^{5-2}$  FFD, they would have found the lack of significance of  $T$  and  $Q$  with only 8 experiments that correspond to the factorial part of a three-factor CCD. A CCD of these characteristics would have required 12 additional experimental data points corresponding to the 6-star points and 6 center points. This two-stage approach requires less than half of the time and consumables needed to achieve the same goals.

The vast majority of the analyzed works estimated the optimal extraction conditions from a model limited to the studied experimental region. As detailed in Appendix A in Supplementary Material, 55% of these optimal conditions coincide with a corner or an edge of the experimental region. Thus, if the purpose of the RSM model is to identify an optimal condition, approximately half of the studies failed because they did not properly define the experimental region so that it actually enclosed the optimal. For the remaining



45% of the cases, the predicted optimum is within the experimental region, but only 45% of these predictions were confirmed by the authors by performing one or more experiments and comparing predicted and experimental values.

As also noted in Appendix A in Supplementary Material, in the majority of studies, first-order or second-order models were fitted following recommendations to implement RSM. However, we also found studies that used artificial neural networks (ANNs) in addition to second-order models. According to Ahangari et al. [17], researchers who used ANN found better predictions of optimal conditions, such is the case for the works of Chouaibi et al. [22], Amani et al. [62], Azmir et al. [69], and Zahedi and Azarpour [97]. From a practical standpoint, when there are resource limitations, it may be convenient to use ANN to find the best extraction conditions. However, the opacity of the internal mechanisms of ANNs precludes their contribution to explanations of the effects of the factors on the response(s). When the network provides a solution, it does not lead to explaining why and how the different factors contributed to the answer. This moves us away from understanding the phenomenon, which is usually sought when considering adopting DoEs, regarding building or confirming hypotheses through experiments.

From our review, only Sodeifian et al. [30], in addition to first-order and second-order models, tested the third-order model. Apparently, the authors did so without a leading reason and without adding more experiments to their design, suggesting an overfitting of the experimental data. The third-order model can improve the goodness of fit criteria of the model as in the study by Sodeifian et al. [30], but at the cost of an increased unreliability of predictions of responses outside conditions of training data.

Finally, we observed that it is not common to find studies that include validation tests for the models. Only 16% of the studies used cross-validation strategies such as leave one out (LOO) where a single data point a time is excluded prior to model fitting, and a comparison is made between the prediction of this “limited model” and the actual “excluded” experimental observation. The validation and the  $R^2$ -predictive or  $Q^2$  statistics inform about the accuracy of predictions of the response surface models by estimating the effect on predictions of the elimination of experimental data points used in model training. Incorporating validation tests enhances confidence in the optimization process by providing a more accurate evaluation of the model’s predictive capabilities and ensuring reliable outcomes.

The predictive power of a fitted response surface model at various points within the experimental region depends on the design being used. A second-order model can be constructed efficiently using CCDs [12]. CCDs are factorial ( $2^n$ ) designs augmented by additional center and axial experimental data points that allow estimation of the tuning parameters of a second-order model. It may be desirable to

have designs that predict uniformly at all constant distances from their center points. The variances and covariances of the best-fit coefficients of the model for such a rotatable design remain unchanged [10]. Because the reason for using response surface designs is generally to locate an unknown optimal, it makes sense to use a rotatable design that provides equal precision in the estimation of the shape of the surface in all directions [10]. A CCD becomes rotatable by choosing an appropriate axial distance to the center of any start point ( $\alpha$  times the coordinate of a factorial point in the same direction). The  $\alpha$ -value to achieve conversion of a design to a rotatable type depends upon the number of independent variables ( $n$ ,  $\alpha = 2^{n/4}$ ) [12].

The CCD can be applied sequentially. To form a CCD, we can start with a  $2^n$  factorial design [10] to fit a first-order model. If the ANOVA for the first-order model shows a significant lack of fit, axial points can be added to allow the inclusion of quadratic terms in the model, as well as supplemental center points to complete a CCD [11]. The number of center points and the axial distance  $\alpha$  are the two parameters we have to fix in a CCD. The center data points provide information about the experimental variance and curvature of the surface, whereas the axial data points allow estimating the coefficients of the second-order terms. A second-order model can describe surfaces that take many shapes. Therefore, the response surface model can represent a maximum, minimum, ridge, or saddle point [11].

When a second-order model is assembled sequentially from a first-order model, it is convenient considering the formation of blocks to reduce the variability that arises from “uncontrollable nuisance factors” [12]. The CCD can be easily carried out using an orthogonal blocking arrangement that provides block-independent coefficients of the response surface model [10]. In the absence of orthogonal experimental blocks, the matrix  $X'X$  will have some non-zero off-diagonal elements following least squares estimation. Consequently, it is impossible to calculate the sum of squares due to quadratic effects without first estimating the linear effects in the model. This impedes the coupling of experimental blocks to perform calculations. In addition to orthogonal, the added experimental blocks must be rotatable. However, because it is not always possible to obtain both rotatability and orthogonality, a value of  $\alpha$  is defined that warrants exact orthogonality and near-rotatability (as much as possible in practice). In practice, depending on the definition of  $\alpha$ , it is easier to guarantee exact orthogonality and get as close as possible to rotatability. For instance, if  $n = 4$ , if we have 16 cubic points and 8 axial points, together with 6 replicates of the center point, we can use an  $\alpha$ -value of 2 to satisfy rotatability and orthogonal blocking [10]. If the number of factors is large enough, the factorial part can be divided into two or more blocks. The axial component must form a single block. Myers et al. [11] reported a table

(blocking arrangements for orthogonal and near-rotatable composite design) of blocking arrangements for composite designs exhibiting rotatability and orthogonality [10].

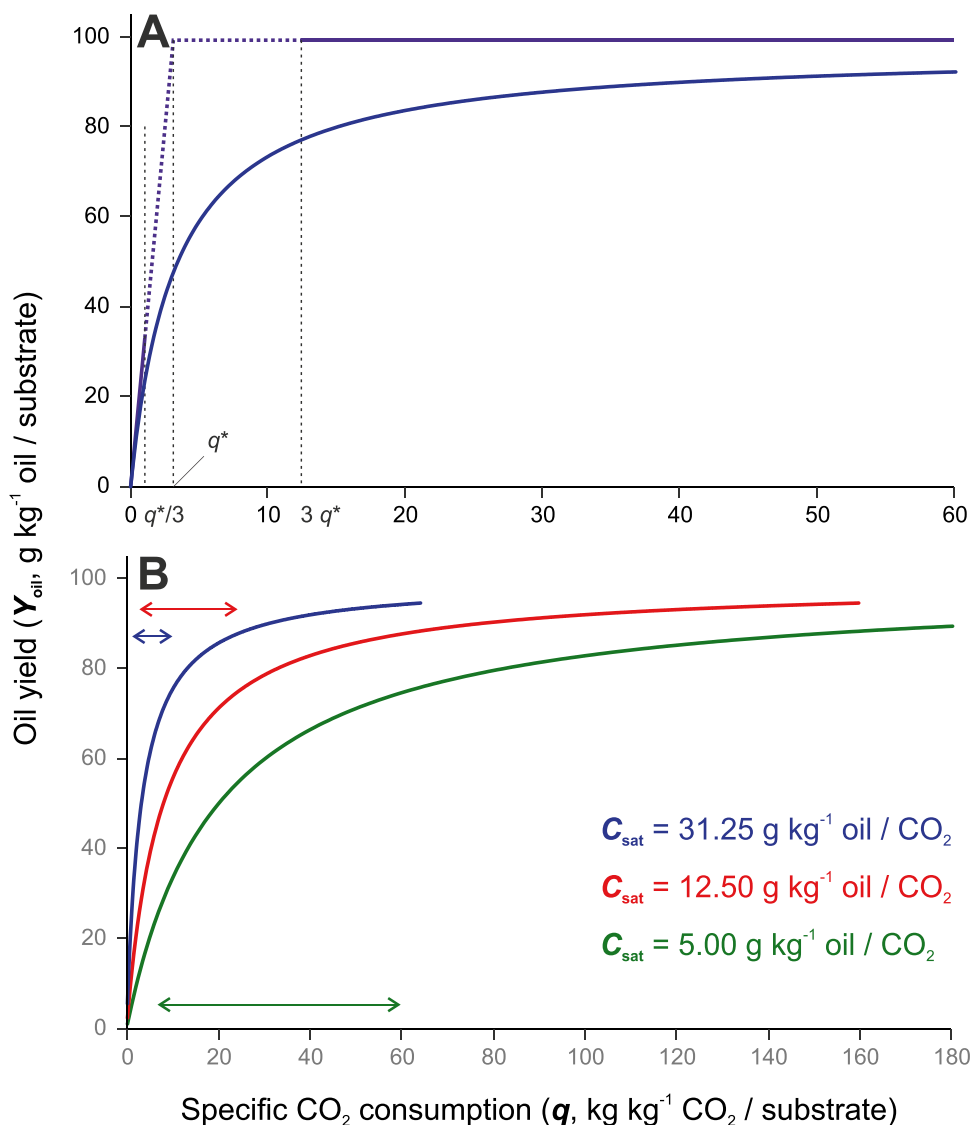
### Effect of Process Variables on the Supercritical CO<sub>2</sub> Extraction of Oil in Seeds

Ahangari et al. [17] recently reviewed current trends in the SC-CO<sub>2</sub> extraction of seed oils, including the effect of several parameters on process rate and yield. The SC-CO<sub>2</sub> extraction process can be visually analyzed by constructing a plot for the extraction yield of oil, or a minor lipid that is coextracted with the oil versus the extraction time or specific CO<sub>2</sub> consumption, keeping constant several independent variables such as the extraction vessel and vessel load, the substrate pretreatment and particle size, the CO<sub>2</sub> conditions (extraction temperature and pressure), and the

CO<sub>2</sub> flow rate passing through the extraction vessel. The extraction experiment is extended for a given extraction time up to fully removing the oil from the seed.

A cumulative extraction curve of oil extraction yield ( $Y$ ) versus specific CO<sub>2</sub> consumption ( $q$ ) is fairly uneventful [105] (Fig. 1). The curve has an initial straight portion by the origin, whose slope is related to the operational solubility of the oil in SC-CO<sub>2</sub> at the selected extraction conditions ( $C_{op}$ ). Following the initial straight portion, the cumulative extraction curve slopes down to reach a horizontal asymptote corresponding to the oil content of the substrate ( $Y_{\infty}$ ). The transition between the two regions depends on the mass transfer mechanisms and mass transfer parameters, which depend in turn on the physical properties and superficial velocity ( $U$ ) of the CO<sub>2</sub>, and the pretreatment and particle size and shape of the substrate. For a given substrate, changing extraction conditions affects mainly the operational solubility, but not the total extractable oil, as noted in Fig. 1B.

**Fig. 1** Cumulative extracción curve of vegetable oil from a pretreated seed. **A** Actual curve, initial asymptote defining the operational solubility of the oil in supercritical CO<sub>2</sub> at process conditions, and final, horizontal asymptote defining the extractable oil content of the pretreated seed. The two asymptotes intercept for a specific CO<sub>2</sub> consumption of  $q^*$ . **B** Cumulative extracción curve as a function of specific CO<sub>2</sub> consumption ( $q$ ) and operational solubility ( $C_{sat}$ , g kg<sup>-1</sup> oil/CO<sub>2</sub>) of the oil in supercritical CO<sub>2</sub>: high (31.25), intermediate (12.50) and low (5.0)



del Valle et al. [106] noted that the operational solubility in the SC-CO<sub>2</sub> extraction of oilseeds coincided with the actual thermodynamic solubility of vegetable oils (triacylglycerols of unbranched fatty acids) in SC-CO<sub>2</sub> at process conditions ( $C_{\text{sat}}$ ), and did not change depending on the substrate. This is so because the thermodynamic solubility depends more on the carbon chain length of the substituent fatty acids (generally 18, in most vegetable oils) than on their number of unsaturations.

Figure 1 is built using the empirical equation of Naik et al. [104] that is written in Eq. (1) as a function of  $C_{\text{sat}}$  and  $Y_{\infty}$ :

$$Y = \frac{Y_{\infty} q}{\frac{Y_{\infty}}{C_{\text{sat}}} + q} \left( = \frac{Y_{\infty} q}{q^* + q} \right) \quad (1)$$

The intersection between the two straight asymptotes ( $Y = C_{\text{sat}} q$ , initially; and  $Y = Y_{\infty}$ , at the end) occurs for a specific CO<sub>2</sub> consumption ( $q^*$ ) that is given by Eq. (2):

$$q^* = \frac{Y_{\infty}}{C_{\text{sat}}} \quad (2)$$

The percent discrepancy of  $Y$  given by Eq. (1) and the horizontal asymptote ( $Y = Y_{\infty}$ , for  $q > q^*$ ) for  $n$ -times  $q^*$  can be computed using Eq. (3a):

$$\% \text{ Error } (n q^*) = 100 \left( \frac{1}{Y_{\infty}} \right) \left[ Y_{\infty} - \frac{Y_{\infty}(n q^*)}{q^* + (n q^*)} \right] \left( = \frac{100}{1 + n} \right), \quad (3a)$$

whereas the percent discrepancy of  $Y$  given by Eq. (1) and the straight line by the origin ( $Y = C_{\text{sat}} q$ , for  $q < q^*$ ) for the  $n$ -th fraction of  $q^*$  can be computed using Eq. (3b):

$$\% \text{ Error } \left( \frac{q^*}{n} \right) = 100 \left( \frac{1}{C_{\text{sat}} q^* / n} \right) \left[ C_{\text{sat}} \left( \frac{Y_{\infty} / C_{\text{sat}}}{n} \right) - \frac{Y_{\infty} (q^* / n)}{q^* + (q^* / n)} \right] \left( = \frac{100}{1 + n} \right), \quad (3b)$$

where  $C_{\text{sat}} q^* = Y_{\infty}$  in Eq. (3b). Equation (3a) and Eq. (3b) indicate that for  $n = 99$ , the percent errors for  $q^*/99$  and  $99 q^*$  are both equal to  $100/(99 + 1)$  or 1%. Correspondingly, the percent errors for  $q^*/3$  and  $3 q^*$  equal both  $100/4 = 25\%$ . The percent error reaches a maximal value equal to 50% [ $100/(1 + 1)$ ] for  $q^*$  ( $n = 1$ ). Because the segmented straight lines in Fig. 1A are limited for  $q^*/3$  and  $3 q^*$ , the segmented straight segments in Fig. 1A limit those specific CO<sub>2</sub> consumption levels where the assumption that the solvent capacity of the CO<sub>2</sub> stream is fully employed overpredicts the actual yield predicted by Eq. (1) by a factor that increases from 25 to 50% between  $q^*/3$  and  $q^*$ , and a factor that decreases from 50 to 25% between  $q^*$  and  $3 q^*$ .

Ahangari et al. [17] list, among the parameters critically affecting the extraction of seed oils in a packed bed using neat CO<sub>2</sub>, the temperature ( $T$ ) and pressure ( $P$ ) conditions in the extraction vessel, the particle size of the substrate ( $d_p$ ), and the CO<sub>2</sub> flow rate ( $Q$ ) during the dynamic extraction time. Because  $Q$  is an extensive variable, it is inadequate to describe the SC-CO<sub>2</sub> extraction process (its effect depends on the size of the system). More appropriate intensive variables related to  $Q$  include the ratio of CO<sub>2</sub> mass flow to substrate loaded in the extraction vessel ( $Q/M_s$ , or the weight of CO<sub>2</sub> per unit weight of the substrate and per unit time, that is inversely proportional to the residence time of CO<sub>2</sub> in the extraction vessel) and the superficial CO<sub>2</sub> velocity ( $U$ , or the velocity the CO<sub>2</sub> stream would reach within the extraction vessel if it were empty, which will be defined in “Variations on Carotenoid Content in the Oil with Process Conditions” section). “Implication of Results” section will describe the effect of the system conditions ( $T$ ,  $P$ ) and extraction conditions (substrate pretreatment,  $d_p$ ,  $U$ ) on the shape of the cumulative extraction curve (Fig. 1). Figure 1 clearly indicates that, for any given substrate and extraction conditions ( $T$ ,  $P$ ,  $d_p$ ,  $U$ ), oil extraction yield increases as the specific CO<sub>2</sub> consumption ( $q$ ) increases, where  $q$  increases proportionally to the dynamic extraction time ( $t_d$ ,  $q = Q t/M_s$ ).

Besides the temperature and pressure conditions of the CO<sub>2</sub>, the substrate and its pretreatment affect largely the rate and yield of the SC-CO<sub>2</sub> extraction process. There are two main strategies to fracture the main inner barrier to mass transfer in vegetable substrates, namely the intact walls of the target-solute-containing cells. They consist of the rupture of surface cells by impact, abrasion, or cut mechanisms, and the fracture of inner cells by high-shear mechanisms. The effectiveness of the fracture of superficial cells using impact mills, abrasive mills, or cutters benefits from a reduction in particle size that increases the surface-to-volume ratio of ground particles or the ratio of ruptured surface cells to intact inner cells in the pretreated vegetable substrate [17, 107]. High-shear pretreatments for vegetable substrates include flaking, prepressing, extrusion, and pelletization [108] that do not resort to a reduction in particle size to increase the fraction of ruptured cells. A high-shear pretreatment ruptures cell walls and frees solutes from inner cells, on one hand, and restructures the substrate into a high-density matrix with interconnected pores, on the other. Not requiring a significant reduction in particle size to be effective to free the oil and minor lipids from the oil-containing cells effectively constitutes a significant advantage of high-shear pretreatments as compared to particle-size-reduction pretreatments, because fines are difficult to handle in packed beds as required in batch SC-CO<sub>2</sub> extraction processes for solid substrates. Indeed, fines have a tendency to aggregate and cause solvent channeling in the packed bed extraction vessel so as to diminish extraction efficiency [109–111], and to clog filters used to



separate suspended particles from the extract-loaded solvent phase thus delaying industrial processing [112]. An added advantage of the high-shear pretreatments as compared to particle-size-reduction pretreatments is that they allow an increase in bulk density of the substrate in the packed bed, so that the extraction vessel load can increase. Densification of the substrate increases the volumetric productivity of the process (or the weight of extract per unit volume of the extraction vessel and per unit time) even if such densification restricts inner mass transfer during extraction [113, 114]. In an extruder, moistened particles of ground material are forced into a spiraling and contracting channel by a screw that turns into a heated barrel. Under the influence of lubricant water and high temperature, plasticized material emerges from the extruder through the constricting opening placed at the end of the barrel (die) and water vapor flashes to an extent dependent on the moisture content and final temperature of the material [115]. Gardner [116] reported an increase in bulk density of hop cones from 150–250 kg/m<sup>3</sup> in ground, hammer-milled hops to 400 kg/m<sup>3</sup> in extruded hops, as well as an increase in  $\alpha$ -acid recovery from 75–80% from ground material to 95% from the extruded material, which they claimed was due to improved, shear-induced cell wall rupture of hop cone cells. Uquiche et al. [114] densified Jalapeño pepper using an extruder prior to SC-CO<sub>2</sub> extraction of capsaicin-rich oleoresins. Other examples of the use of extrusion as a pretreatment include the SC-CO<sub>2</sub> extraction of dried red pepper flakes [117, 118], *Nannochloropsis gaditana* microalgae [119], and marigold flower petals [120].

## Supercritical CO<sub>2</sub> Extraction of Oils and Carotenoids in Rosehip Seeds and Shells

Winther et al. [121] reviewed the chemical makeup and biological activity of rosehips. The pseudofruits are constituted of a red flesh pulp or hypanthium enclosing several thin-membrane-surrounded seeds or individual achenes (the actual botanical fruits). Following drying, the two parts of the rosehip pseudofruit give rise to shells and seeds, respectively. Rosehip seeds are rich in essential polyunsaturated fatty acids such as linoleic and  $\alpha$ -linolenic, whereas the rosehip shells are rich in flavonoids and carotenoids. Particularly, carotenoids are important minor lipid compounds because of their function as pro-vitamin A, antioxidant, and coloring agent. However, the solubility of carotenoids in SC-CO<sub>2</sub> is limited, which decreases their extraction yield from plant substrates.

Dąbrowska et al. [122] recently reviewed the extraction processes for rosehip seeds, with supercritical fluid extraction processes being prominently featured. Table 3 summarizes the conditions of the SC-CO<sub>2</sub> extraction of oil from seeds. The oil of different rosehip species has been extracted using SC-CO<sub>2</sub>, including *Rosa aff. rubiginosa* in the cases of del Valle et al. [3, 108, 126], Eggers et al. [111], and Mabe et al. [123], and *Rosa canina* L. in the cases of Szentmihályi et al. [125], Machmudah et al. [127, 131], Salgin et al. [128], Jahongir et al. [129], Illés et al. [130], and Tozzi et al. [132]. SC-CO<sub>2</sub> extraction has been carried out both at laboratory scale [3, 108, 126, 128, 131, 132] as well as pilot scale [3,

**Table 3** Works on supercritical CO<sub>2</sub> extraction of oil from rosehip (*Rosa aff. rubiginosa* or *Rosa canina* L) seeds and shells available in the open literature

Substrate	$d_p$	$T/P$	$Q/M_s$	$t_d$	Reference
Flaked seeds	–	40–60/30–50	0.21	90–270	[3]
Flaked seeds	–	40/30	0.084	270	[3]
Flaked seeds	0.53	40/30	0.60	120	[108]
Ground seeds	0.96	40–80/30–70	0.14–0.48	85–265	[111]
Ground seeds	–	15/52	0.025	330	[123]
Ground seeds	0.42–1.0	40–70/10–69	0.15–0.46	30	[124]
Ground seeds	< 0.36	35/25	–	80	[125]
Ground seeds	0.65	40–50 or 30–40	0.46	60–90	[126]
Ground seeds	0.56–2.1	40–80/15–45	–	180	[127]
Ground seeds	0.36–0.50	40–60/20–40	–	150	[128]
Ground seeds	0.27–1.4	50/30	–	120	[129]
Roller milled seeds	0.79	40/30 or 50/40	0.15–0.92	60–90	[126]
Roller milled seeds	1.0	40/30 or 50/40	0.46	60–90	[126]
Roller milled seeds	0.79	40/30 or 50/40	0.25	150	[126]
Ground fruit	< 0.36	35/25	–	–	[130]
Ground shells	0.67	40–80/15–45	–	150	[131]
Ground shells	–	70/30	1.0	–	[132]

Operating conditions and arrangement of the substrate studied in the experiments. Particle size ( $d_p$ , mm), temperature ( $T$ , °C), pressure ( $P$ , MPa), specific rate of CO<sub>2</sub> consumption [ $Q/M_s$ , (g CO<sub>2</sub>) (g substrate<sup>-1</sup>) min<sup>-1</sup>], and dynamic extraction time ( $t_d$ , min)

111, 126, 129]. Table 3 also includes a contribution on the extraction of oils from whole fruits, and a couple of contributions on the SC-CO<sub>2</sub> extraction of rosehip shell carotenoids, including the one Tozzi et al. [132], but in this work, ethanol was added as a cosolvent to increase their solubility so as to reduce CO<sub>2</sub> requirements.

There have been several works showing a positive effect of vegetable oil as a continuous cosolvent for the SC-CO<sub>2</sub> extraction of lycopene from tomato products [133–135], β-carotene from carrots [136], lutein from marigold flower petals [137, 138], and astaxanthin from shrimp shells [139]. Besides using vegetable oil as a continuous cosolvent, which can be added using a cosolvent pump, it is also possible to carry out extraction in a mixed substrate-packed bed containing a vegetable oil source, in addition to the carotenoid source, as done by adding hazelnut to help extract tomato lycopene [140], avocado for extracting either tomato lycopene [141] or red pepper capsanthin [142], or roasted coffee for extracting brown seaweed fucoxanthin [143]. It is clearly advantageous to use a single substrate to coextract lipids and carotenoids, as done by Machmudah et al. [144] to extract tomato flesh byproducts using tomato seeds as the cosubstrate, or by Ndayishimiye and Chun [145] from *Yuzu ichandrin* (a citrus fruit) peels using *Y. ichandrin* seeds as the cosubstrate. Consequently, we proposed using rosehip seeds as a cosubstrate of rosehip shells to help extract shell carotenoids using SC-CO<sub>2</sub>. Our proposal has manifested itself in industrial practice with two products offered by *Flavex Naturextrakte*, a large German food ingredients company. *Flavex Naturextrakte* offers both an oil for use in cosmetics [146] and an extract with the carotenoids from the fruit shells, which give it a reddish color [147].

## Case Study

In this section, we will study the coextraction of oil from the seeds and carotenoids from the shells of rosehips as a case study for the application of RSM and DoE to analyze the SC-CO<sub>2</sub> extraction of oil and minor lipids. The study uses as independent variables the extraction temperature, CO<sub>2</sub> density at the extraction conditions, specific CO<sub>2</sub> consumption during the dynamic extraction time, and seed-to-shell ratio in the substrate, and as dependent variables the oil extraction yield and carotenoid content of the oil. Substrate mixtures were manufactured by extrusion which allows an effective destruction of inner barriers to mass transfer, while avoiding the presence of fines. The vessel load, particle size of the substrate, and superficial CO<sub>2</sub> velocity were all kept constant in the unreported experiments of a student at Universidad de la Frontera (UFro). Appendix C in Supplementary Material presents the materials and methods of this unreported experimental work.

Table 4 shows the experimental results of oil yield and carotenoid content in the oil when extracting extruded rosehip seeds and shells using SC-CO<sub>2</sub> as a function of the extraction temperature, CO<sub>2</sub> density, specific CO<sub>2</sub> consumption, and content of seeds in the pretreated substrate. Oil yield ranged from about 50.0 to 80.5 g kg<sup>-1</sup> oil/substrate, depending on the substrate and the extraction conditions. To make results independent of the substrate, we computed the percent oil recovery, which was based on the fraction of seeds in the composite substrate (with the remainder being rosehip shells), and the experimentally measured oil contents of seeds and shells (Appendix C in Supplementary Material). The oil content of the extruded samples ranged from 50.5 g kg<sup>-1</sup> oil/substrate when using 240 g kg<sup>-1</sup> seed/substrate, to 99.1 g kg<sup>-1</sup> oil/substrate when using only seeds. The percent oil recovery should have been limited to 100% but, because of experimental errors was above that limit for up to 10–15% in a few selected opportunities when applying a high specific solvent consumption of CO<sub>2</sub> having a high solvent power (high temperature and density), or a few of those traits. The lowest percent oil recoveries were limited to 65–70% when applying a limited specific solvent consumption of CO<sub>2</sub> having a low solvent power (low temperature and density), or a few of those traits. The carotenoid content in the oil changed even more dramatically between 0.10 g kg<sup>-1</sup> carotenoids/oil when using rosehip seeds as the substrate to 5.5 g kg<sup>-1</sup> carotenoids/oil when using a substrate containing 24% of seeds and 76% of shells. The yield of carotenoids was computed as the product of the two main responses which, the same as the carotenoid content of the oil, ranged widely from 7.9 to 320 mg kg<sup>-1</sup> carotenoids/substrate. To make these results independent of the substrate, we computed the percent carotenoid recovery, which was also based on the fraction of seeds in the composite substrate, and the experimentally measured carotenoid contents of seeds and shells (Appendix C in Supplementary Material). The carotenoid content of the extruded samples increased from 73.4 mg kg<sup>-1</sup> oil/substrate when using only seeds to 185.3 mg kg<sup>-1</sup> carotenoids/substrate when using 240 g kg<sup>-1</sup> seed/substrate. The percent carotenoid recovery should have been limited to 100% but, because of compound experimental errors of both the yield of oil and the carotenoid content in the oil, it was above that limit six times for up to 70%. The recovery of carotenoids varied widely from 10% in the single experiment using the substrate containing only seeds to 170% in the single experiment using the substrate containing 24% of seeds and 76% of shells.

The remainder of this section will present the statistical analyses and graphical representations of the RSs separately for oil yield (“Variations on Oil Yield with Process Conditions” section) and carotenoid content in the oil (“Variations on Carotenoid Content in the Oil with Process Conditions” section) for the complete experimental CCD, which be followed by a discussion with the implications of our results.

**Table 4** Results of the application of the rotatable central composite design (CCD) to the supercritical CO<sub>2</sub> extraction of mixtures of seeds and shells of rosehip

<i>T</i>	$\rho$	<i>q</i>	<i>S</i>	<i>Y</i> <sub>oil</sub>	<i>C</i> <sub>car</sub>	<i>Y</i> <sub>car</sub>	<i>R</i> <sub>oil</sub>	<i>R</i> <sub>car</sub>
45	860	12	81	60.8	0.46	28.0	69.9	27.6
65	860	12	43	51.8	3.00	155.3	82.7	98.7
45	940	12	43	56.4	3.66	206.5	90.1	131.2
65	940	12	81	79.4	0.58	46.2	91.3	45.6
45	860	24	43	62.2	2.29	142.5	99.3	90.6
65	860	24	81	71.5	0.72	51.2	82.2	50.5
45	940	24	81	58.5	0.86	50.3	67.3	49.7
65	940	24	43	67.8	3.29	223.2	108.3	141.9
55	900	18	62	63.9	1.44	91.9	85.4	71.0
55	900	18	62	61.8	1.41	86.9	82.6	67.2
45	860	12	43	50.0	2.00	99.8	79.8	63.5
65	860	12	81	63.5	0.83	52.6	73.1	51.9
45	940	12	81	65.9	0.74	48.9	75.8	48.2
65	940	12	43	62.7	3.65	228.4	100.0	145.2
45	860	24	81	73.2	0.66	48.5	84.2	47.9
65	860	24	43	69.9	2.58	180.5	111.6	114.8
45	940	24	43	68.0	3.37	228.9	108.6	145.5
65	940	24	81	65.4	0.64	41.8	75.2	41.2
55	900	18	62	63.5	1.37	87.1	84.8	67.3
55	900	18	62	62.3	1.39	86.5	83.3	66.9
35	900	18	62	55.0	1.25	68.4	73.5	52.9
75	900	18	62	67.2	1.74	117.1	89.9	90.5
55	820	18	62	54.7	1.58	86.2	73.2	66.7
55	980	18	62	61.5	1.97	121.0	82.2	93.5
55	900	6	62	53.3	1.37	72.8	71.3	56.3
55	900	30	62	71.4	1.51	107.5	95.5	83.1
55	900	18	24	58.0	5.45	315.9	114.9	170.5
55	900	18	100	80.5	0.10	7.9	81.2	10.7
55	900	18	62	62.4	1.44	89.6	83.4	69.3
55	900	18	62	62.4	1.40	87.1	83.4	67.4

Factors correspond to temperature (*T*, °C), density ( $\rho$ , kg/m<sup>3</sup>), specific CO<sub>2</sub> consumption (*q*, kg kg<sup>-1</sup> CO<sub>2</sub>/substrate), and fraction of seeds in the composite substrate (*S*, g kg<sup>-1</sup> seeds/substrate). Response variables are the oil extraction yield (*Y*<sub>oil</sub>, g kg<sup>-1</sup> oil/substrate), the carotenoid content in the oil (*C*<sub>car</sub>, g kg<sup>-1</sup> carotenoids/oil), the carotenoids extraction yield (*Y*<sub>car</sub>, g kg<sup>-1</sup> carotenoid/substrate), the oil recovery (*R*<sub>oil</sub>, %), and the carotenoids recovery (*R*<sub>car</sub>, %)

Appendix D in Supplementary Material statistically demonstrates that the surfaces for oil yield and carotenoid content in the oil for Blocks 1 and 2, respectively, exhibit considerable curvature and therefore cannot be represented using a first-order model (not significant, statistically), which constitutes the first stage in the proposed methodology, before completing the experiments (Block 3) and analyzing the response surface for each response.

**Variations on Oil Yield with Process Conditions**

Clearly, the results in Appendix D of Supplementary Material indicate that a linear model is not adequate to describe the response for oil yield. There are curved regions that the

linear model does not explain, so that a second-order model, such as the one presented in Eq. (4) should be applied:

$$Y = \sum_{i=0}^4 \sum_{j=i}^4 \beta_{ij} X_i X_j \tag{4}$$

where the same as before,  $\beta_{00}$  is the intercept coefficient;  $\beta_{0i}$  is the main effect for the normalized factor “*i*” (0, 1, 2, 3, or 4);  $\beta_{ii}$  is the quadratic effect for the normalized factor “*i*”;  $\beta_{ij}$  is the interaction for the normalized effects of factors “*i*” and “*j*” (1, 2, 3, or 4, different from “*i*”); *X<sub>i</sub>* and *X<sub>j</sub>* are the levels of the coded independent variables *i* and *j*, respectively (*X*<sub>0</sub> = 1); and *Y* is a response variable, that in this section corresponds to oil yield (*Y*<sub>oil</sub>). The coded independent variables in Eq. (4) are defined as follows (Eqs. 5a–d):

**Table 5** Statistical descriptors of the second-order model performance (Block. Analysis of variance for the fitted regression coefficients using the oil extraction yield ( $Y_{oil}$ , g kg<sup>-1</sup> oil/substrate) as response

Regression coefficients	$Y_{oil}$	
	Estimate	<i>p</i> -value
Model ( <i>F</i> -value)	15.7	< 0.0001
$\beta_1$	2.555	0.002
$\beta_2$	1.449	0.053
$\beta_3$	3.426	< 0.0001
$\beta_4$	3.936	< 0.0001
$\beta_4^2$	1.979	0.006
$\beta_{23}$	-3.463	< 0.001
$\beta_{34}$	-3.007	0.002
Lack of fit	14.44	0.024
$R^2$	0.85	
Adjusted- $R^2$	0.79	
Signal-to-noise ratio	15.0	
C. V.	5.4	

$$X_1 = \frac{T - 55}{10}, \quad (5a)$$

$$X_2 = \frac{\rho - 900}{40}, \quad (5b)$$

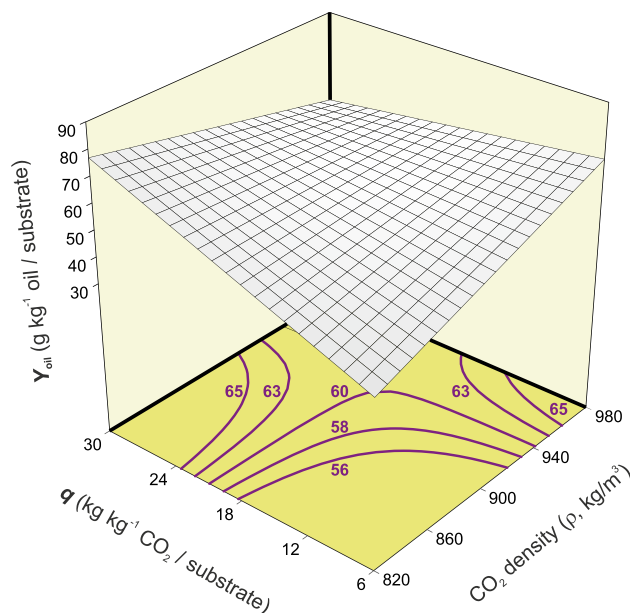
$$X_3 = \frac{q - 18}{6}, \text{ and} \quad (5c)$$

$$X_4 = \frac{S - 620}{190}, \quad (5d)$$

where  $T$  is the extraction temperature (°C),  $\rho$  (kg/m<sup>3</sup>) is the density of CO<sub>2</sub> at the process conditions (defined by the extraction temperature and pressure),  $q$  (kg kg<sup>-1</sup> CO<sub>2</sub>/substrate) is the specific solvent consumption, and  $S$  (g kg<sup>-1</sup> seeds/substrate) is the fraction of seeds in the composite substrate.

Table 5 summarizes the statistical indicators obtained from the ANOVA when the selected second-order model, Eq. (4), was applied to the results for  $Y_{oil}$  in Table 4. The second-order model was considered adequate, because of the insignificant lack of fit ( $p \leq 0.05$ ) relative to the pure error, the high signal-to-noise ratio (> 14), and the high coefficient of determination ( $R^2 = 0.85$ ). The  $R^2$  value indicates that the second-order model explains 84.6% of the variability in the values of  $Y_{oil}$ . Appendix D in Supplementary Material complements the statistical indicators in Table 5, showing that the residuals of the model [or the discrepancies between experimental values in Table 4 and those predicted by Eq. (4)] behave normally.

To present our results graphically, we proceeded to eliminate nonsignificant coefficients ( $p > 0.05$ ) in Eq. (4).



**Fig. 2** Surface plot of oil extraction yield ( $Y_{oil}$ ) with supercritical CO<sub>2</sub> as a function of the specific CO<sub>2</sub> consumption ( $q$ ) and CO<sub>2</sub> density ( $\rho$ ) at  $T = 45^\circ\text{C}$  and  $S = 62\%$ . Numbers on contour map correspond to oil yield ( $Y_{oil}$ , g kg<sup>-1</sup> oil/substrate)

However, as reported in Table 5, we kept the linear coefficient of dimensionless density ( $X_2$ ), which was not significant at the 5% level ( $p = 0.053$ ), to keep the hierarchy of the response surface model. Only three second-order coefficients that affected oil yield were kept in the model, namely the interaction between the coded density ( $X_2$ ) and the coded specific CO<sub>2</sub> consumption ( $X_3$ ), the interaction

**Table 6** Statistical descriptors of the second-order model performance. Analysis of variance for the fitted regression coefficients using the carotenoid content in the oil ( $C_{car}$ , g kg<sup>-1</sup> carotenoids/oil) as response

Regression coefficients	$C_{car}$	
	Estimate	<i>p</i> -value
Model ( <i>F</i> -value)	157.6	< 0.0001
$\beta_1$	0.933	0.028
$\beta_2$	2.095	< 0.0001
$\beta_4$	-12.101	< 0.0001
$\beta_2^2$	0.833	0.032
$\beta_4^2$	3.337	< 0.0001
$\beta_{12}$	-1.367	0.010
$\beta_{24}$	-2.459	< 0.0001
Lack of fit	88.7	0.002
$R^2$	0.98	
Adjusted- $R^2$	0.98	
Signal-to-noise ratio	43	
C. V.	11	

between the coded specific CO<sub>2</sub> consumption ( $X_3$ ) and the coded fraction of seeds in the substrate ( $X_4$ ), and the square of the fraction of seeds in the substrate ( $X_4^2$ ). Equation (6) reports the second-order model for oil extraction yield that can be derived for the significant coefficient values informed in Table 5:

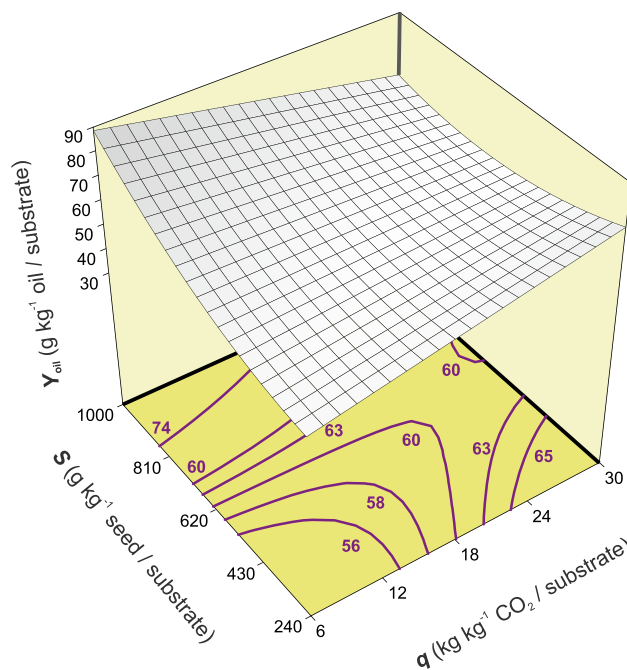
$$\begin{aligned}
 Y_{oil} = & 61.91 + 2.555 \left( \frac{T - 55}{10} \right) + 1.449 \left( \frac{\rho - 900}{40} \right) \\
 & + 3.426 \left( \frac{q - 18}{6} \right) + 3.936 \left( \frac{S - 620}{190} \right) + 1.979 \left( \frac{S - 620}{190} \right)^2 \\
 & - 3.463 \left( \frac{\rho - 900}{40} \right) \left( \frac{q - 18}{6} \right) - 3.007 \left( \frac{q - 18}{6} \right) \left( \frac{S - 620}{190} \right)
 \end{aligned}
 \tag{6}$$

RSM offers useful tools for understanding the behavior of a response as a function of several factors. However, when there are three or more independent variables, as in this case, it is difficult to visualize the behavior of the response. The nature of the response surface may be determined from the stationary point and the sign and magnitude of the eigenvalues in the so-called canonical analysis [12]. The eigenvalues of Eq. (6) are  $\lambda_1 = +1.1508$ ,  $\lambda_2 = -2.6540$ ,  $\lambda_3 = -0.3396$ , and  $\lambda_4 = +2.8469$ . Eigenvalues can be interpreted considering them in pairs: two positive values signal the presence of minimal value in the direction defined by the corresponding axis; two positive values signal the presence of a maximal value; and opposite signs eigenvalues signal that the surface portrays saddle points [10].

Statistical indicator  $Q^2$  for  $Y_{oil}$  was 0.62 which suggests a low accuracy of predictions and overfitting of the regression model since it depends considerably on the experimental data. This result suggests that the fitted model will have difficulties in predicting new data that does not match the calibration experiments.

Figure 2 shows the interaction between  $\rho$  and  $q$  for the extraction at 45 °C of a substrate containing 62% of seeds. The surface represents a saddle, which is common to find in cases of a relatively flat experimental region that does not include a true maximum or minimum [6]. What we observe are basically situations in which the maximum oil yield can be reached virtually with the exception of those cases in which little CO<sub>2</sub> is used, for small  $q$ , or its solvent power is low, for a low value of  $\rho$ , on the front and to the left when oil yield falls below the expected 75 g kg<sup>-1</sup> oil/substrate.

Figure 3 shows the interaction between  $q$  and  $S$  for  $T=45$  °C and  $\rho=940$  kg/m<sup>3</sup>, which also represents a saddle. The same as in the case of Fig. 3, we observed a plateau and a fall to the front and left when the oil content in the substrate decreases when  $S$  (the fraction of seeds in the substrate) or when  $q$  (the quantity of CO<sub>2</sub> used for extraction) decreases.



**Fig. 3** Surface plot of oil extraction yield ( $Y_{oil}$ ) with supercritical CO<sub>2</sub> as a function of the fraction of seeds in the composite substrate ( $S$ ) and specific CO<sub>2</sub> consumption ( $q$ ) at  $T=45^\circ\text{C}$  and  $\rho=940$  kg/m<sup>3</sup>. Numbers on contour map correspond to oil extraction yield ( $Y_{oil}$ , g kg<sup>-1</sup> oil/substrate)

### Variations on Carotenoid Content in the Oil with Process Conditions

Equation (4) was also used to represent the effect of the experimental factors on the carotenoid content in the oil, given that Appendix D of Supplementary Material indicated that a linear model was not adequate to describe this response. Table 6 summarizes the statistical indicators obtained from the ANOVA when Eq. (4) was applied to the results for  $C_{car}$  in Table 4. The same as in the case of oil yield, the second-order model was considered adequate, because of the insignificant lack of fit ( $p \leq 0.01$ ) relative to the pure error, the high signal-to-noise ratio ( $> 40$ ), and the high coefficient of determination ( $R^2 = 0.98$ ). This last coefficient indicates that the second-order model explains 98.2% of the variability in  $C_{car}$ . Appendix D in Supplementary Material complements the information provided by the statistical indicators in Table 6 showing that the residuals of Eq. (4) for carotenoid content in the oil behave normally.

As reported in Table 6, we proceeded to eliminate non-significant coefficients ( $p > 0.05$ ) in Eq. (4) to present our results graphically. We noted that the coded specific consumption of CO<sub>2</sub> ( $X_3$ ) does not affect the carotenoid content in the oil, which in practice indicates that the carotenoids and the oil are extracted in fixed proportions throughout extraction. Furthermore, only two of the remaining second-order

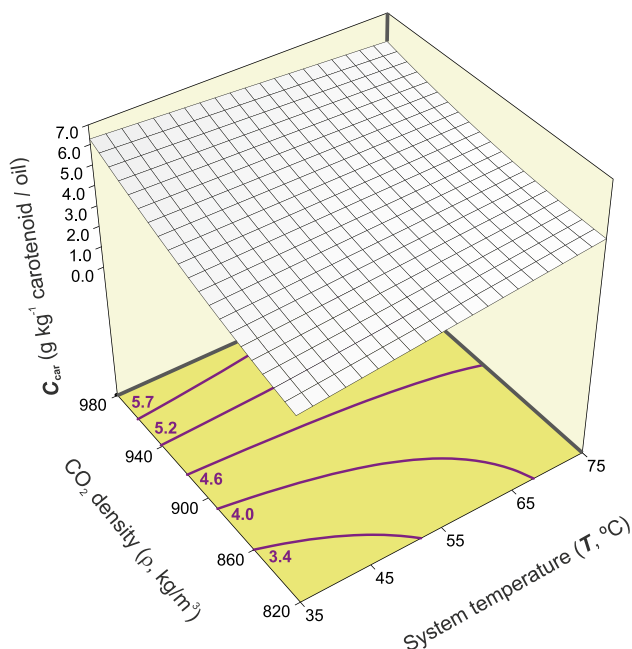


coefficients do not affect the carotenoid content in the oil, which are the square of the temperature ( $X_1^2$ ), and the interaction between the temperature and the fraction of seeds in the substrate ( $X_1X_4$ ). Equation (7) reports the second-order model for the carotenoid content in the oil based on the significant coefficient values informed in Table 6:

$$C_{car} = 14.233 + 0.933 \left( \frac{T-55}{10} \right) + 2.095 \left( \frac{\rho-900}{40} \right) - 12.101 \left( \frac{S-620}{190} \right) + 0.833 \left( \frac{\rho-900}{40} \right)^2 + 3.337 \left( \frac{S-620}{190} \right)^2 - 1.367 \left( \frac{T-55}{10} \right) \left( \frac{\rho-900}{40} \right) - 2.459 \left( \frac{\rho-900}{40} \right) \left( \frac{S-620}{190} \right) \quad (7)$$

The eigenvalues of Eq. (7) are  $\lambda_1 = +3.8955$ ,  $\lambda_2 = -0.6452$ ,  $\lambda_3 = +0.050$  (nearly null value), and  $\lambda_4 = +1.0779$ . Statistical indicator  $Q^2$  for  $C_{car}$  was 0.93. Thus, the accuracy of predictions of the polynomial model is better for  $C_{car}$  than for  $Y_{oil}$  ( $Q^2=0.62$ ), which indicates that the independent variables and their chosen combinations better describe and predict carotenoid content in the oil than oil yield.

Figure 4 shows the interaction between temperature and CO<sub>2</sub> density when extracting a substrate containing

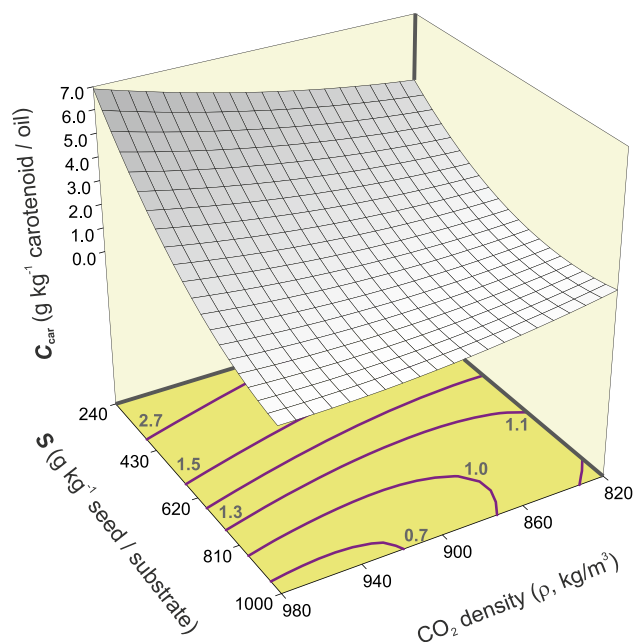


**Fig. 4** Surface plot of carotenoid content ( $C_{car}$ ) with supercritical CO<sub>2</sub> as a function of the CO<sub>2</sub> density ( $\rho$ ) and temperature ( $T$ ) at  $q=18 \text{ kg kg}^{-1} \text{ CO}_2/\text{substrate}$  and  $S=30\%$ . Numbers on contour map correspond to using the carotenoid content in the oil ( $C_{car}$ ,  $\text{g kg}^{-1}$  carotenoids/oil)

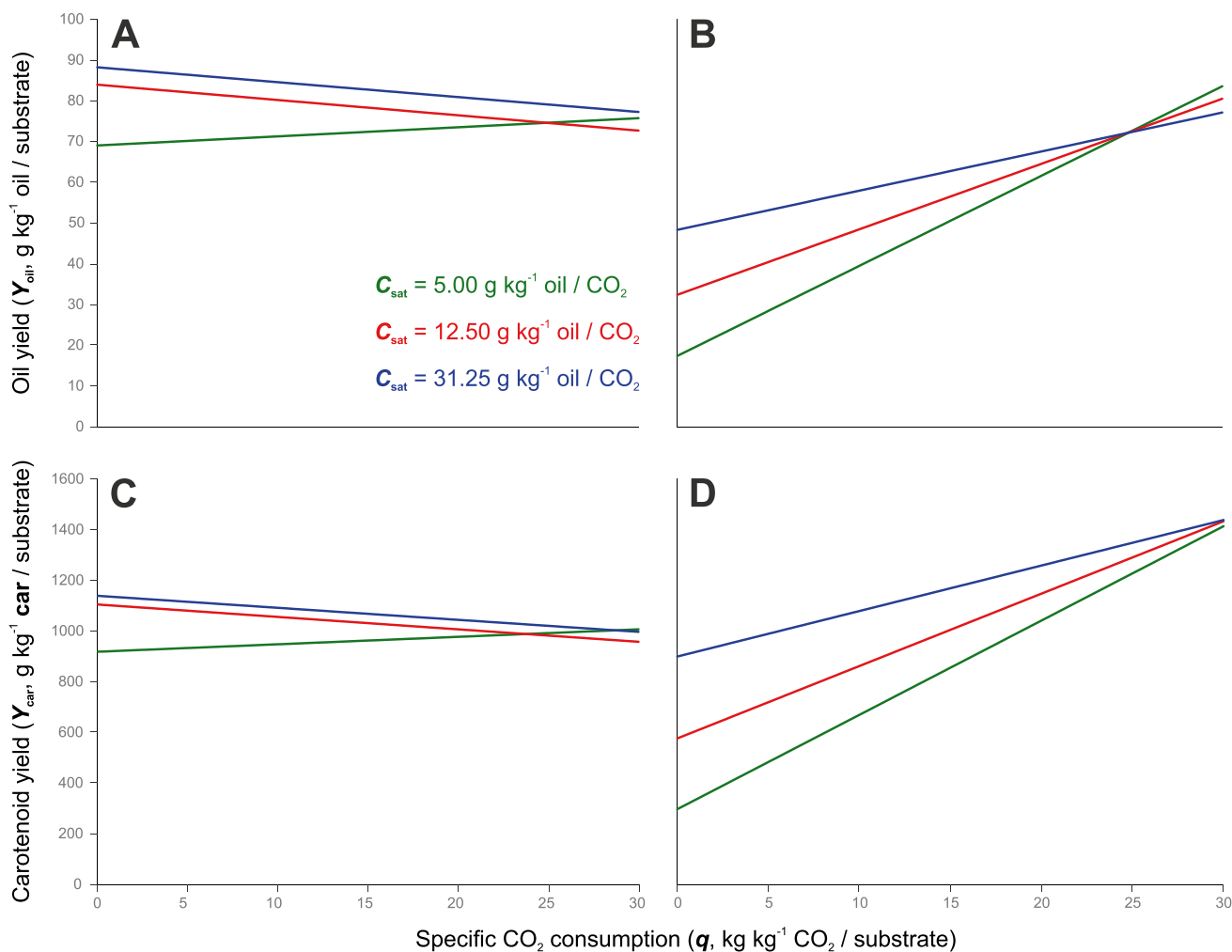
43% seeds using  $18 \text{ kg kg}^{-1} \text{ CO}_2/\text{substrate}$ .  $C_{car}$  changed slightly with increasing  $T$  and  $\rho$ , resulting in a fairly flat response surface graph. Indeed, when the CO<sub>2</sub> density increased from 820 to 980  $\text{kg/m}^3$ ,  $C_{car}$  increased slightly, perhaps because the extraction is more defined by the solute content in the substrate than by its solubility in CO<sub>2</sub> at the selected extraction conditions.

Figure 5 shows the interaction between  $\rho$  and  $S$  when extracting the raw material using  $18 \text{ kg kg}^{-1} \text{ CO}_2/\text{substrate}$  at  $75 \text{ }^\circ\text{C}$ . There is a great dependence on the carotenoid content in the oil when using CO<sub>2</sub> with high solvent power (for  $\rho=980 \text{ kg/m}^3$ ), the case where the carotenoid content in the substrate determines its content in the oil. We think this may be due to the large cosolvent effect of the oil on the solubilization of carotenoids. When using CO<sub>2</sub> with low solvent power (for  $\rho=820 \text{ kg/m}^3$ ), on the other hand, there is a smaller effect of the composition of the raw material on the content of carotenoids in the extracted oil.

Figure 6 presents results in the variations in oil yield (Fig. 6A, B), and carotenoid yield (Fig. 6C, D), for rosehip seeds (Fig. 6A, C), and rosehip fruits containing approximately 25.1% of seeds and the rest of shells (Fig. 6B, D). The carotenoid yield was estimated as the product of the oil yield by the carotenoid content in the oil, which were the two responses modeled in this work. The curves represent changes in the extraction conditions that increased the oil solubility in CO<sub>2</sub> from 5  $\text{g/kg}$  at  $43.6 \text{ }^\circ\text{C}$  and 23.8



**Fig. 5** Surface plot of carotenoid content ( $C_{car}$ ) with supercritical CO<sub>2</sub> as a function of the fraction of seeds in the composite substrate ( $S$ ) and CO<sub>2</sub> density ( $\rho$ ) at  $T=75 \text{ }^\circ\text{C}$  and  $q=18 \text{ kg kg}^{-1} \text{ CO}_2/\text{substrate}$ . Numbers on contour map correspond to using the carotenoid content in the oil ( $C_{car}$ ,  $\text{g kg}^{-1}$  carotenoids/oil)



**Fig. 6** Prediction of the extraction yield as a function of specific CO<sub>2</sub> consumption ( $q$ ) and saturation concentration of the oil in CO<sub>2</sub> ( $C_{sat}$ ) using the response surface model for **A** oil from seeds, **B** oil from fruits, **C** carotenoids from seeds, **D** carotenoids from fruits

MPa ( $\rho = 854.3$  kg/m<sup>3</sup>), 12.5 g/kg at 53.9 °C and 36.9 MPa ( $\rho = 895.8$  kg/m<sup>3</sup>), or 31.25 g/kg at 65.1 °C and 55.7 MPa ( $\rho = 940.2$  kg/m<sup>3</sup>).

**Implication of Results**

Figure 6 shows that the response surface is not capable of predicting extraction curves of oil yield versus specific CO<sub>2</sub> consumption, in the sense that they do not start at the

origin, and that they do not approach a horizontal asymptote for high values of  $q$  as suggested by Fig. 1. However, their trends approximate expected curves between  $q^*/3$  and  $3q^*$  in that it is not the thermodynamic solubility of the oil for low values of  $q$ , nor the oil content in the substrate which determines the variations in oil yield ( $Y_{oil}$ ) with the specific CO<sub>2</sub> consumption ( $q$ ). In the case of rosehip seeds (for  $S = 1000$  g kg<sup>-1</sup> seeds/substrate), it is apparent that it is the oil content in the seeds which determines the yield of

**Table 7** Effect of temperature ( $T$ ) and their interaction with pressure ( $T \times P$ ) on oil extraction yield from oilseeds reported in literature as a function of different pressure and temperature ranges

Substrate	$T$ (°C)	$P$ (MPa)	$T$ effect	$T \times P$ effect	Reference
<i>Camellia sinensis</i> seeds	40–60	17.5–22.5	Negative	Positive	[48]
<i>Glycine max</i> seeds	40–50	10–30	Negative	Not significant	[60]
<i>Brassica napus</i> seeds	40–60	20–30	Negative	Not significant	[55]
<i>Cyperus esculentus</i> L. nuts	40–80	20–40	Not significant	Positive	[56]
<i>Salvia hispanica</i> L. seeds	40–80	25–45	Not significant	Positive	[59]

oil, with the trend decreasing as the solvent power of the SC-CO<sub>2</sub> decreases (Fig. 6A). On the other hand, in the case of the complete fruit (for  $S = 251 \text{ g kg}^{-1}$  seeds/substrate), a trend closer to that expected is seen, which indicates that the solubility of the extract is lower than the solubility of the oil; as the solvent power of CO<sub>2</sub> increases, oil extraction is closer to being determined by the oil content in the substrate, at least until reaching a specific CO<sub>2</sub> consumption of  $25 \text{ kg kg}^{-1}$  CO<sub>2</sub>/substrate (Fig. 6B). The trend is more pronounced in the case of carotenoid extraction, in which there is a more marked effect of the solvent power of CO<sub>2</sub> (Fig. 6D).

RSM claims that it can provide an approximate picture of the behavior of a system with a limited number of experiments. However, this is not the case for the extraction of vegetable oil from an optimally sheared substrate, where there have been notable advances in science and technology aspects to allow a more accurate picture of the phenomena with an even more limited experimental effort. Indeed, del Valle [148] claimed that a single experiment was required to determine a best-fit pretreatment-dependent microstructural factor to describe the extraction of prepressed, flaked, pelletized, or extruded seeds that were ground and size-classified using the shrinking core model. According to del Valle [148] the extraction curve predicted by the shrinking core model will depend on four groups of factors. The first group corresponds to the substrate and depends on its treatment, which will define, in turn, the microstructural factor ( $F_M$ ) and the particle diameter ( $d_p$ ). The second group refers to the target solute, which in the case of a vegetable oil can be homologated to a triolein molecule. The third group relates to the extraction conditions that can be characterized by the process temperature ( $T$ ) and pressure ( $P$ ).  $T$  and  $P$ , in turn, and under the assumption that the limited amount of dissolved oil does not affect the physical properties of CO<sub>2</sub>, determine the density ( $\rho$ ) and viscosity ( $\mu$ ), which can be estimated using the NIST database [149]. The last physical property, the diffusivity of the oil in CO<sub>2</sub> under the extraction conditions ( $D_{12}$ ), can be estimated as a function of  $\mu$  and the molecular weight of triolein (a triglyceride usually employed to represent typical vegetable oils) using the equation of Funazukuri et al. [150]. The last group refers to the characteristics of the extraction process that depend on the CO<sub>2</sub> flow rate ( $Q$ ), and the volume ( $V_E$ ) and aspect ratio ( $L/D$ ) of the cylindrical extraction vessel which, in turn, define its high ( $L$ ) and internal diameter ( $D$ ). The superficial CO<sub>2</sub> velocity in the extraction vessel ( $U$ ) can be estimated by dividing  $Q$  by  $\rho$  and the cross-sectional area of the extractor ( $\pi D^2/4$ ). Then, the dimensionless Schmidt ( $Sc$ ) number can be determined, which depends on the physical properties of the system under study ( $\rho$ ,  $\mu$ ,  $D_{12}$ ). On the other hand, the dimensionless Reynolds ( $Re$ ) number, which characterizes the flow regime in the packed bed, depends on the physical properties of CO<sub>2</sub> ( $\rho$ ,  $\mu$ ),  $U$ , and

$d_p$ .  $Re$  and  $Sc$  allow the determination of the dimensionless Sherwood ( $Sh$ ) number using a specific correlation such as the one of King and Catchpole [151] for mass transfer in packed beds operating with supercritical fluids. Finally,  $\rho$  and  $T$  allow estimating the solubility of the oil in CO<sub>2</sub> ( $C_{\text{sat}}$ ) under the extraction conditions using the equation of del Valle et al. [106];  $F_M$  and  $D_{12}$  allow estimating the effective diffusivity of the oil in the pretreated substrate ( $D_e$ ) [108], which characterizes the internal mass transfer in the solid substrate; and  $Sh$  makes it possible to estimate the film coefficient ( $k_f$ ) that characterizes mass transfer by convection in the packed bed.

Something similar occurs in the case of seeds subjected to size reduction by impact, abrasion, or cutting mechanisms that cause rupture of surface cells. Extraction of these type of substrates can be represented by the intact-and-broken cell model [151–154] whose predictions depends on the fraction of broken cells, an oil partition coefficient between intact and broken cells, and mass transfer coefficients for the intact tissue (characterized by inner mass transfer coefficient  $k_s$ ), and from broken superficial cells to the surrounding SC-CO<sub>2</sub> phase (characterized by  $k_f$ ). Reverchon and Marrone [107] suggested using microscopy to characterize the parenchymatous cells and related the fraction of broken cells to the ratio  $\alpha$  of the volume of a superficial layer half the size of a typical parenchymatous cell ( $\delta$ ), and the volume of the whole particle, containing intact inner cells. For a spherical particle, the relationship is given by Eq. (8) [155]:

$$\alpha \left( = \frac{V_{\text{libre}}}{V} \right) = 3 \left( \frac{\delta}{d_p} \right) - 3 \left( \frac{\delta}{d_p} \right)^2 + \left( \frac{\delta}{d_p} \right)^3 \quad (8)$$

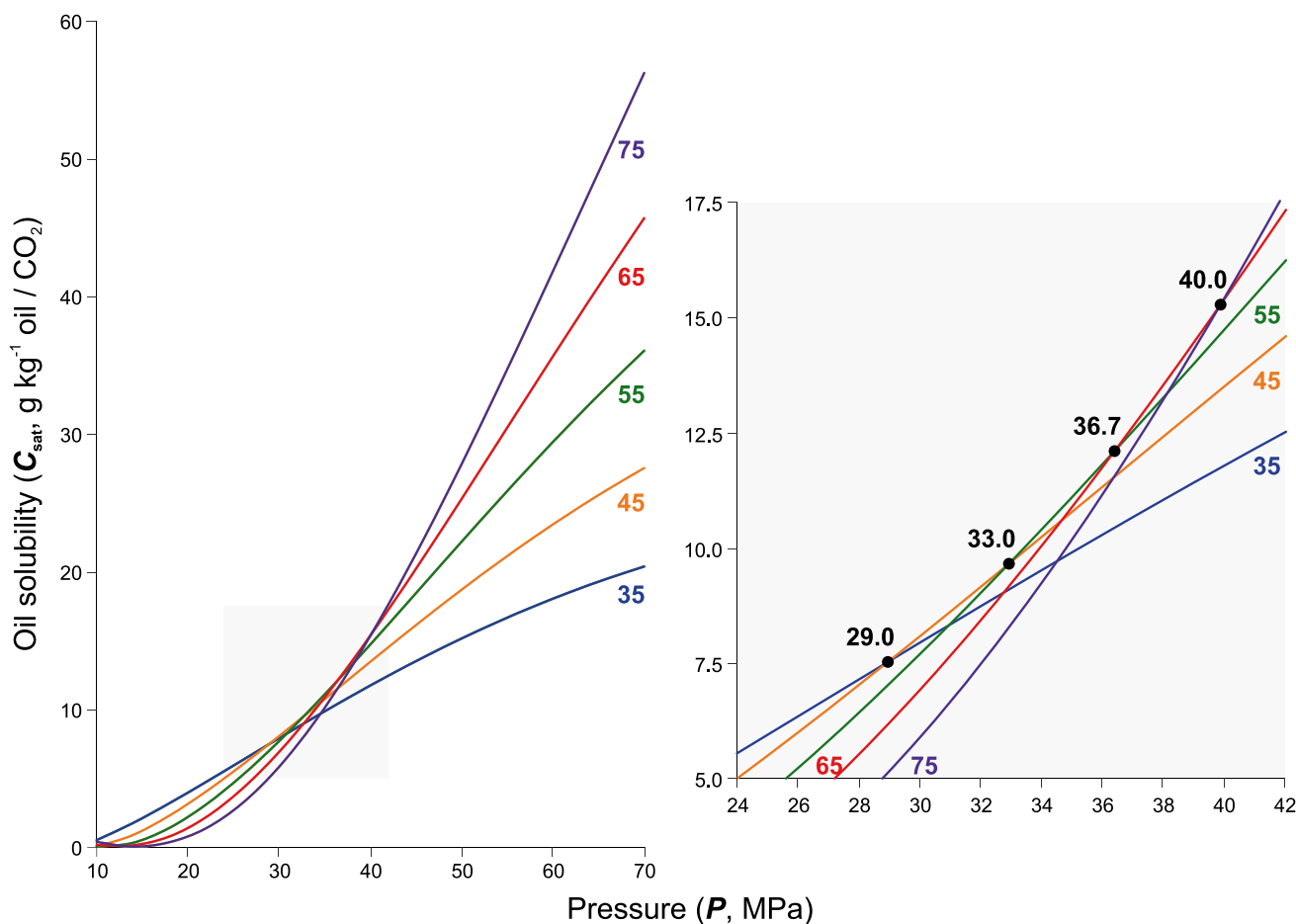
Recently, Arias et al. [156] estimated  $k_s$  as a function of a pretreatment-dependent microstructural factor ( $F_M$ ) and  $k_f$  using a dimensionless correlation for packed beds operating with supercritical fluids, and best-fitted the values of substrate-dependent  $\alpha$  and  $F_M$ , and extraction-condition-dependent  $K$  (partition coefficient) for the extraction curves for the *Lippia origanoides*' oleoresin using ethanol-modified SC-CO<sub>2</sub>. Urrego et al. [157] measured of equilibrium partition of rapeseed oil between SC-CO<sub>2</sub> and prepressed rapeseed.

There is some support to the claim that the use of vegetable oil as cosolvent improves the solubility of carotenoids in CO<sub>2</sub>, as done by Araus et al. [158] for the solubility of  $\beta$ -carotene in triolein-modified SC-CO<sub>2</sub>, and by Araus et al. [159] for the solubility of capsanthin in triolein-modified SC-CO<sub>2</sub>, both in the form of an enhancement factor, or an indication of how many times larger is the solubility of the carotenoid in triolein-modified CO<sub>2</sub> than in pure CO<sub>2</sub> at the same system temperature and pressure, and where the solubility of triolein in SC-CO<sub>2</sub> is not modified by the presence of small amounts of the dissolved carotenoids.

Clearly, oil solubility in SC-CO<sub>2</sub> under the extraction conditions determines the extraction curves for the oil and carotenoids in Fig. 6. Figure 7 shows the variations in oil solubility in SC-CO<sub>2</sub> as a function of temperature (different isotherms) and pressure (abscissa). For low pressures, the solubility of the oil decreases as temperature increases, due to the reduction in the density and the solvent power of CO<sub>2</sub> [160]. On the other hand, at high pressures, for which CO<sub>2</sub> is less compressible, the solubility of the oil increases with temperature, due to the increase in the vapor pressure and volatility of the oil. Therefore, there is an intermediate pressure region, at the so-called crossover pressure, which is highlighted in the expanded grey area in Fig. 7, in which the effects of the solvent power of CO<sub>2</sub> and the volatility of the oil are compensated, and the solubility of the oil in the CO<sub>2</sub> does not change with temperature [161]. In the case of vegetable oils, the crossover pressure increases with the temperature between 29 MPa for the crossover between 35 and 45 °C, and 40 MPa for the crossover between 65 and 75 °C (Fig. 7).

As a result of these changes in the oil solubility in CO<sub>2</sub> with temperature, a decrease in oil yield with increasing temperature was expected in studies at low pressures (below 29 MPa) as can be observed in the first three lines of Table 7. An increase in oil yield would be also expected with increasing temperature in studies at high pressures (above 40 MPa), but no such studies were found in the literature. Finally, at intermediate pressures, including the crossover pressure range between 29 and 40 MPa, mixed results would be expected, with an inconclusive effect of temperature on oil extraction yield, and a great statistical significance of the interactions between pressure and temperature, as highlighted in the last two lines of Table 7.

The supercritical of Chen et al. [86] predicts a crossover pressure of 31.4 to 35.2 MPa for a temperature interval from ca. 40 to 60 °C, but in this case, the crossover decreased instead of increased as the temperature increased. Chen et al. [86] used the system temperature and pressure, and the specific CO<sub>2</sub> consumption as the factors, and the percent oil extraction yield as the response, and Appendix E in



**Fig. 7** Crossover effect. **A** Solubility of vegetable oils in supercritical CO<sub>2</sub> as a function of temperature (different isotherms) and pressure (abscissa axis); **B** variations in oil solubility in SC-CO<sub>2</sub> in the experimental range of the extraction conditions studied

Supplementary Material details how this information was used to estimate the crossover pressure, which is a condition for which the derivative of an estimate of oil solubility as a function of temperature keeping pressure constant is null.

Unlike the complex effects of system temperature and pressure on solute solubility, which includes the crossover phenomena that modify the functional dependency of the solubility on system temperature, the equation proposed by Chrastil [162], on which Fig. 7 is based, predicts positive effects of both system temperature and CO<sub>2</sub> density on solute solubility. Indeed, Chrastil's equation predicts that solute solubility in SC-CO<sub>2</sub> depends on three factors, which are the solubility at an arbitrary reference condition; a correction factor for the CO<sub>2</sub> density, which depends on a factor  $k$  that defines the number of CO<sub>2</sub> molecules that form a solvate-complex with a single molecule of the solute; and a correction factor for the absolute temperature, which depends on the heat of solvation  $\Delta H$  of the solute, that includes the thermal effects to first vaporize the solute and to secondly dissolve it in CO<sub>2</sub> phase [106, 163].

Considering the positive effects of system temperature and CO<sub>2</sub> density on solute solubility predicted by the equation of Chrastil [162], on one hand, and the positive effect of solute solubility on the CO<sub>2</sub> extraction curves (Fig. 1B), it is strange that there are such a limited number of studies that use  $T$  and  $\rho$  instead of  $T$  and  $P$  as independent variables. An exception is the work of del Valle and Aguilera [2] mentioned in the “Introduction” section, but some additional contributions can be also mentioned, including those on the SC-CO<sub>2</sub> extraction of polyphenols from an inert matrix [164] or grape seeds [165], and on the SC-CO<sub>2</sub> desorption of lemon essential oil from silica gel [166]. We have also used  $T$  and  $\rho$  as independent variables to describe the SC-CO<sub>2</sub> extraction of oils and antioxidant carotenoids from pelletized *Nannochloropsis gaditana* microalgae using RSM [119, 167].

## Conclusions

Researchers in the areas of process engineering and food engineering should exercise caution when reviewing the state of the art when attempting to understand, describe, or analyze a process of interest. Its main objective should be to find and use models supported by fundamental principles, which allow safe scaling projections. In the event that the study is so novel that the effects of the factors on the response(s) are unknown, then they should be concerned with carefully selecting the independent variables, and the experimental space where they will study them, before embarking on doing the DoE and the subsequent RSM.

We suggest that future contributions to the supercritical extraction of solid matrices favor an approach based

on the construction of extraction curves (e.g., cumulative extraction yield versus specific solvent consumption). This information will allow the application of models to obtain mass transfer (e.g., diffusion coefficient) and equilibrium (e.g., operational solubility) parameters, relevant information for the scaling of the process, and for the understanding of the phenomenon. extraction. The response surface methodology only has one use when investigating the effect of an additional variable. For example, it would apply when investigating the effect of the type of pretreatment. Researchers should avoid using variables such as temperature since there are analytical expressions that describe the effect of these variables.

This study demonstrates the advantage of using  $T$ ,  $\rho$ , and  $q$  instead of the very commonly used  $T$ ,  $P$ , and  $t_d$  as independent variables in DoE and RSM studies on the SC-CO<sub>2</sub> extraction from oilseeds. Based on the review and analysis of the literature, this work claims that response surface designs have been used excessively, and consequently, most of the articles that have used these designs have not provided new information to understand the extractive phenomenon, particularly in the extraction of vegetable oils with SC-CO<sub>2</sub>. For example, in the carotenoid extraction case study, the response surface was not able to predict extraction curves descriptive of the extraction process. Furthermore, in this case study, we propose the use of  $\rho$  and  $T$  as independent variables, instead of  $P$  and  $T$ , because they bypass confounding effects of solute solubility in CO<sub>2</sub>, and the specific consumption of CO<sub>2</sub> is a more appropriate independent variable than time. Finally, the purpose of the case study was to exemplify the application of a sequential strategy of the RSM from a first-order model to a second-order one. This sequential experimentation strategy favored the determination of the effects of coextraction of oil and carotenoids, where the extraction of carotenoids from rosehip peels was favored when the proportion of rosehip seeds as cosubstrate increased.

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1007/s12393-023-09357-z>.

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on the Supercritical CO<sub>2</sub> Extraction of Oil in Seeds”, “Supercritical CO<sub>2</sub> Extraction of Oils and Carotenoids in Rosehip Seeds and Shells”, “Implication of Results”, and “Conclusions” sections), making illustrations (all figures), final editing, communicating with journal officials.

**Availability of Data and Material** Data is available on request and can be found in the supplementary material.

**Code Availability** Not applicable.

## Declarations

**Conflict of Interest** The authors declare no competing interests.

## References

- Sharif KM, Rahman MM, Azmir J, Mohamed A, Jahurul MHA, Sahena F, Zaidul ISM (2014) Experimental design of supercritical fluid extraction – a review. *J Food Eng* 124:105–116. <https://doi.org/10.1016/j.jfoodeng.2013.10.003>
- del Valle JM, Aguilera JM (1989) Effects of substrate densification and CO<sub>2</sub> conditions on supercritical extraction of mushroom oleoresins. *J Food Sci* 54:135–141. <https://doi.org/10.1111/j.1365-2621.1989.tb08586.x>
- del Valle JM, Bello S, Thiel J, Allen A, Chordia L (2000) Comparison of conventional and supercritical CO<sub>2</sub>-extracted rosehip oil. *Braz J Chem Eng* 17:335–348. <https://doi.org/10.1590/S0104-66322000000300010>
- Bisgaard S (1992) Industrial use of statistically designed experiments: case study references and some historical anecdotes. *Qual Eng* 4:547–562. <https://doi.org/10.1080/08982119208918936>
- Tanco M, Viles E, Ilzarbe L, Alvarez MJ (2009) Implementation of design of experiments projects in industry. *Appl Stoch Models Bus Ind* 25:478–505. <https://doi.org/10.1002/asmb.779>
- de Oliveira LG, de Paiva AP, Balestrassi PP, Ferreira JR, da Costa SC, da Silva Campos PH (2019) Response surface methodology for advanced manufacturing technology optimization: theoretical fundamentals, practical guidelines, and survey literature review. *Int J Adv Manuf Technol* 104:1785–1837. <https://doi.org/10.1007/s00170-019-03809-9>
- Haaland PD (1989) *Experimental design in biotechnology*, 1st edn. CRC Press, Florida. ISBN 978-1-003-06596-8
- Box GE, Hunter JS, Hunter WG (2005) *Statistics for experimenters: design, innovation, and Discovery*, 2nd edn. John Wiley & Sons, New Jersey. ISBN 978-0-471-71813-0
- Box GE, Draper NR (2007) *Response surfaces, mixtures, and ridge analyses*, 2nd edn. John Wiley & Sons, New Jersey. ISBN 978-0-470-05357-7
- Gacula MC Jr, Singh J, Bi J, Altan S (2009) *Statistical methods in food and consumer research*, 2nd edn. Academic Press, San Diego. ISBN 978-0-123-73716-8
- Myers RH, Montgomery DC, Anderson-Cook CM (2016) *Response surface methodology: process and product optimization using designed experiments*, 4th edn. John Wiley & Sons, New Jersey. ISBN 978-1-118-91601-8
- Montgomery DC (2019) *Design and analysis of experiments*, 10th edn. John Wiley & Sons, New Jersey. ISBN 978-1-119-49244-3
- Jones B, Montgomery DC (2019) *Design of experiments: a modern approach*, 1st edn. John Wiley & Sons, New Jersey. ISBN 978-1-119-61119-6
- e-Handbook of Statistical Methods (2022) NIST/SEMATECH, Gaithersburg. <http://www.itl.nist.gov/div898/handbook/>. <https://doi.org/10.18434/M32189>. Accessed 8 Oct 2023
- Yolmeh M, Jafari SM (2017) Applications of response surface methodology in the food industry processes. *Food Bioprocess Technol* 10:413–433. <https://doi.org/10.1007/s11947-016-1855-2>
- de Melo MMR, Silvestre AJD, Silva CM (2014) Supercritical fluid extraction of vegetable matrices: applications, trends and future perspectives of a convincing green technology. *J Supercrit Fluids* 92:115–176. <https://doi.org/10.1016/j.supflu.2014.04.007>
- Ahangari H, King JW, Ehsani A, Yousefi M (2021) Supercritical fluid extraction of seed oils – a short review of current trends. *Trends Food Sci Technol* 111:249–260. <https://doi.org/10.1016/j.tifs.2021.02.066>
- Peng WL, Mohd-Nasir H, Setapar SHM, Ahmad A, Lokhat D (2020) Optimization of process variables using response surface methodology for tocopherol extraction from Roselle seed oil by supercritical carbon dioxide. *Ind Crops Prod* 143:111886. <https://doi.org/10.1016/j.indcrop.2019.111886>
- Zhang S, Zu YG, Fu YJ, Luo M, Li WL, Efferth T (2010) Supercritical carbon dioxide extraction of seed oil from yellow horn (*Xanthoceras sorbifolia* Bunge.) and its antioxidant activity. *Biores Technol* 101:2537–2544. <https://doi.org/10.1016/j.biortech.2009.11.082>
- Ara KM, Jowkarderis M, Raofie F (2015) Optimization of supercritical fluid extraction of essential oils and fatty acids from flixweed (*Descurainia sophia* L.) seed using response surface methodology and central composite design. *J Food Sci Technol* 52(7):4450–4458. <https://doi.org/10.1007/S13197-014-1353-3>
- Ara KM, Karami M, Raofie F (2014) Application of response surface methodology for the optimization of supercritical carbon dioxide extraction and ultrasound-assisted extraction of *Capparis spinosa* seed oil. *J Supercrit Fluids* 85:173–182. <https://doi.org/10.1016/j.supflu.2013.10.016>
- Chouaibi M, Rigane K, Ferrari G (2020) Extraction of Citrullus colocynthis L. seed oil by supercritical carbon dioxide process using response surface methodology (RSM) and artificial neural network (ANN) approaches. *Ind Crops Prod* 158:113002. <https://doi.org/10.1016/j.indcrop.2020.113002>
- Suryawanshi B, Mohanty B (2018) Modeling and optimization of process parameters for supercritical CO<sub>2</sub> extraction of *Argemone mexicana* (L.) seed oil. *Chem Eng Commun* 206:1087–1106. <https://doi.org/10.1080/00986445.2018.1547712>
- Devi V, Khanam S (2019) Development of generalized and simplified models for supercritical fluid extraction: case study of papaya (*Carica papaya*) seed oil. *Chem Eng Res Des* 150:341–358. <https://doi.org/10.1016/j.cherd.2019.08.006>
- Devi V, Khanam S (2019) Comparative study of different extraction processes for hemp (*Cannabis sativa*) seed oil considering physical, chemical and industrial-scale economic aspects. *J Clean Prod* 207:645–657. <https://doi.org/10.1016/j.jclepro.2018.10.036>
- Suryawanshi B, Mohanty B (2018) Modeling and optimization: supercritical CO<sub>2</sub> extraction of *Pongamia pinnata* (L.) seed oil. *J Environ Chem Eng* 6:2660–2673. <https://doi.org/10.1016/j.jece.2018.04.014>
- Rai A, Mohanty B, Bhargava R (2015) Modeling and response surface analysis of supercritical extraction of watermelon seed oil using carbon dioxide. *Sep Purif Technol* 141:354–365. <https://doi.org/10.1016/j.seppur.2014.12.016>
- Daraee A, Ghoreishi SM, Hedayati A (2019) Supercritical CO<sub>2</sub> extraction of chlorogenic acid from sunflower (*Helianthus annuus*) seed kernels: modeling and optimization by response surface methodology. *J Supercrit Fluids* 144:19–27. <https://doi.org/10.1016/j.supflu.2018.10.001>
- Zhang Z, Liu Y, Che L (2018) Optimization of supercritical carbon dioxide extraction of *Eucommia ulmoides* seed oil and quality evaluation of the oil. *J Oleo Sci* 67:255–263. <https://doi.org/10.5650/jos.ess17153>

30. Sodeifian G, Ghorbandoost S, Sajadian SA, Saadati Ardestani N (2016) Extraction of oil from *Pistacia khinjuk* using supercritical carbon dioxide: experimental and modeling. *J Supercrit Fluids* 110:265–274. <https://doi.org/10.1016/j.supflu.2015.12.004>
31. Maran JP, Priya B (2015) Supercritical fluid extraction of oil from muskmelon (*Cucumis melo*) seeds. *J Taiwan Inst Chem Eng* 47:71–78. <https://doi.org/10.1016/j.jtice.2014.10.007>
32. Ni Q, Gao Q, Yu W, Liu X, Xu G, Zhang Y (2015) Supercritical carbon dioxide extraction of oils from two *Torreya grandis* varieties seeds and their physicochemical and antioxidant properties. *LWT Food Sci Technol* 60:1226–1234. <https://doi.org/10.1016/j.lwt.2014.09.007>
33. Kraujalis P, Venskutonis PR (2013) Optimisation of supercritical carbon dioxide extraction of amaranth seeds by response surface methodology and characterization of extracts isolated from different plant cultivars. *J Supercrit Fluids* 73:80–86. <https://doi.org/10.1016/j.supflu.2012.11.009>
34. Zhao S, Zhang D (2013) A parametric study of supercritical carbon dioxide extraction of oil from *Moringa oleifera* seeds using a response surface methodology. *Sep Purif Technol* 113:9–17. <https://doi.org/10.1016/j.seppur.2013.03.041>
35. Zhang QA, Fan XH, Zhang ZQ, Zhang BS, Zhang ZQ, Jia XY (2009) Optimization of SC-CO<sub>2</sub> extraction of oil from almond pretreated with autoclaving. *LWT Food Sci Technol* 42:1530–1537. <https://doi.org/10.1016/j.lwt.2009.05.007>
36. Xu X, Gao Y, Liu G, Wang Q, Zhao J (2008) Optimization of supercritical carbon dioxide extraction of sea buckthorn (*Hippophaë thamnoides* L.) oil using response surface methodology. *LWT Food Sci Technol* 41:1223–1231. <https://doi.org/10.1016/j.lwt.2007.08.002>
37. Liu Z, Liu B, Kang H, Yue H, Chen C, Jiang L, Shao Y (2019) Subcritical fluid extraction of *Lycium ruthenicum* seeds oil and its antioxidant activity. *Int J Food Sci Technol* 54:161–169. <https://doi.org/10.1111/ijfs.13920>
38. Sodeifian G, Sajadian SA, Saadati-Ardestani N (2017) Supercritical fluid extraction of omega-3 from *Dracocephalum kotschy* seed oil: process optimization and oil properties. *J Supercrit Fluids* 119:139–149. <https://doi.org/10.1016/j.supflu.2016.08.019>
39. Yu J, Wang J, Liu C, Liu Z, Wang Q (2012) Application of response surface methodology to optimise supercritical carbon dioxide extraction of oil from rapeseed (*Brassica napus* L.). *Int J Food Sci Technol* 47:1115–1121. <https://doi.org/10.1111/j.1365-2621.2012.02948.x>
40. Ghoreishi SM, Bataghva E (2011) Supercritical extraction of evening primrose oil: experimental optimization via response surface methodology. *AIChE J* 57:3378–3384. <https://doi.org/10.1002/aic.12532>
41. Çelik HT, Gürü M (2015) Extraction of oil and silybin compounds from milk thistle seeds using supercritical carbon dioxide. *J Supercrit Fluids* 100:105–109. <https://doi.org/10.1016/j.supflu.2015.02.025>
42. Özkal SG (2009) Response surface analysis and modeling of flaxseed oil yield in supercritical carbon dioxide. *J Am Oil Chem Soc* 86:1129–1135. <https://doi.org/10.1007/s11746-009-1448-6>
43. Özkal SG, Yener ME, Bayindirli L (2005) Response surfaces of apricot kernel oil yield in supercritical carbon dioxide. *LWT Food Sci Technol* 38:611–616. <https://doi.org/10.1016/j.lwt.2004.08.003>
44. de Lucas A, Rincón J, Gracia I (2003) Influence of operation variables on quality parameters of olive husk oil extracted with CO<sub>2</sub>: three-step sequential extraction. *J Am Oil Chem Soc* 80:181–188. <https://doi.org/10.1007/s11746-003-0674-4>
45. de Lucas A, Rincón J, Gracia I (2002) Influence of operating variables on yield and quality parameters of olive husk oil extracted with supercritical carbon dioxide. *J Am Oil Chem Soc* 79:237–243. <https://doi.org/10.1007/s11746-002-0467-9>
46. Durante M, Ferramosca A, Treppiccione L, Di Giacomo M, Zara V, Montefusco A, Piro G, Mita G, Bergamo P, Lenucci MS (2020) Application of response surface methodology (RSM) for the optimization of supercritical CO<sub>2</sub> extraction of oil from patè olive cake: yield, content of bioactive molecules and biological effects in vivo. *Food Chem* 332:127405. <https://doi.org/10.1016/j.foodchem.2020.127405>
47. Luan Z, Jie, Li, P. pei, Li, D., Meng, X. ping, & Sun, J. (2020) Optimization of supercritical-CO<sub>2</sub> extraction of *Iris lactea* seed oil: component analysis and antioxidant activity of the oil. *Ind Crops Prod* 152:112553. <https://doi.org/10.1016/j.indcrop.2020.112553>
48. Muangrat R, Jirarattanarangsri W (2020) Physicochemical properties and antioxidant activity of oil extracted from Assam tea seeds (*Camellia sinensis* var. *assamica*) by supercritical CO<sub>2</sub> extraction. *J Food Process Preserv* 44:e14364. <https://doi.org/10.1111/jfpp.14364>
49. Jelani NAA, Azlan A, Khoo HE, Razman MR (2019) Fatty acid profile and antioxidant properties of oils extracted from dabai pulp using supercritical carbon dioxide extraction. *Int Food Res J* 26:1587–1598
50. Mo L, Mengxi W, Dongya C, Lingyan K, Mei X, Hong L (2019) Extraction and analysis of flaxseed oil by supercritical CO<sub>2</sub> method optimized by RSM and GC-MS. *Curr Top Nutraceutical Res* 17:11–16
51. Kulkarni NG, Kar JR, Singhal RS (2017) Extraction of flaxseed oil: a comparative study of three-phase partitioning and supercritical carbon dioxide using response surface methodology. *Food Bioprocess Technol* 10:940–948. <https://doi.org/10.1007/s11947-017-1877-4>
52. Hossain MS, Norulaini NN, Naim AA, Zulkhairi AM, Bennama MM, Omar AM (2016) Utilization of the supercritical carbon dioxide extraction technology for the production of deoiled palm kernel cake. *J CO<sub>2</sub> Utiliz* 16:121–129. <https://doi.org/10.1016/j.jcou.2016.06.010>
53. Wang L, Wang X, Wang P, Xiao Y, Liu Q (2016) Optimization of supercritical carbon dioxide extraction, physicochemical and cytotoxicity properties of *Gynostemma pentaphyllum* seed oil: a potential source of conjugated linolenic acids. *Sep Purif Technol* 159:147–156. <https://doi.org/10.1016/j.seppur.2016.01.007>
54. Belayneh HD, Wehling RL, Cahoon E, Ciftci ON (2015) Extraction of omega-3-rich oil from *Camelina sativa* seed using supercritical carbon dioxide. *J Supercrit Fluids* 104:153–159. <https://doi.org/10.1016/j.supflu.2015.06.002>
55. Cvjetko M, Jokić S, Lepojević Ž, Vidović S, Marić B, Radojčić Redovniković I (2012) Optimization of the supercritical CO<sub>2</sub> extraction of oil from rapeseed using response surface methodology. *Food Technol Biotechnol* 50:208–215
56. Lasekan O, Abdulkarim SM (2012) Extraction of oil from tiger nut (*Cyperus esculentus* L.) with supercritical carbon dioxide (SC-CO<sub>2</sub>). *LWT Food Sci Technol* 47:287–292. <https://doi.org/10.1016/j.lwt.2012.01.021>
57. Yang R, Wang H, Jing N, Ding C, Suo Y, You J (2012) Trigonella foenum-graecum L. seed oil obtained by supercritical CO<sub>2</sub> extraction. *J Am Oil Chem Soc* 89(12):2269–2278. <https://doi.org/10.1007/s11746-012-2123-x>
58. Jiang ST, Niu L (2011) Optimization and evaluation of wheat germ oil extracted by supercritical CO<sub>2</sub>. *Grasas Aceites* 62:181–189. <https://doi.org/10.3989/gya.078710>
59. Ixtaina VY, Vega A, Nolasco SM, Tomás MC, Gimeno M, Bárzana E, Tecante A (2010) Supercritical carbon dioxide extraction of oil from Mexican chia seed (*Salvia hispanica* L.): characterization and process optimization. *J Supercrit Fluids* 55:192–199. <https://doi.org/10.1016/j.supflu.2010.06.003>

60. Jokić S, Zeković Z, Vidović S, Sudar R, Nemet I, Bilić M, Velić D (2010) Supercritical CO<sub>2</sub> extraction of soybean oil: process optimisation and triacylglycerol composition. *Int J Food Sci Technol* 45:1939–1946. <https://doi.org/10.1111/j.1365-2621.2010.02358.x>
61. Bhattacharjee P, Singhal RS, Tiwari SR (2007) Supercritical carbon dioxide extraction of cottonseed oil. *J Food Eng* 79:892–898. <https://doi.org/10.1016/j.jfoodeng.2006.03.009>
62. Amani M, Ardestani NS, Honarvar B (2021) Experimental optimization and modeling of supercritical fluid extraction of oil from *Pinus gerardiana*. *Chem Eng Technol* 44:578–588. <https://doi.org/10.1002/ceat.202000347>
63. Bojanić N, Teslić N, Rakić D, Brdar M, Fišteš A, Zeković Z, Bodroža-Solarov M, Pavlić B (2019) Extraction kinetics modeling of wheat germ oil supercritical fluid extraction. *J Food Process Preserv* 43:e14098. <https://doi.org/10.1111/jfpp.14098>
64. Satyannarayana S, Anjaneyulu B, Neeharika TSVR, Rani KNP, Chakrabarti PP (2018) Process optimization for the supercritical carbon dioxide (SC-CO<sub>2</sub>) extraction of wheat germ oil with respect to yield, and phosphorous and tocol contents using a Box Behnken design. *Grasas Aceites* 69:e259–e259. <https://doi.org/10.3989/gya.0102181>
65. Yamamoto N, Murakami K, Kimthet C, Wahyudiono W, Onwona-Agyeman S, Kanda H, Goto M (2018) Lipids from *Vitellaria paradoxa* Gaertn seeds by supercritical CO<sub>2</sub>: extraction and optimization of parameters by response surface methodology. *Engineering Journal-Thailand* 22:31–44. <https://doi.org/10.4186/ej.2018.22.5.31>
66. Rombaut N, Savoie R, Hecke EV, Thomasset B (2017) Supercritical CO<sub>2</sub> extraction of linseed: optimization by experimental design with regards to oil yield and composition. *Eur J Lipid Sci Technol* 119:1600078. <https://doi.org/10.1002/ejlt.201600078>
67. Aladić K, Vidović S, Vladić J, Balić D, Jukić H, Jokić S (2016) Effect of supercritical CO<sub>2</sub> extraction process parameters on oil yield and pigment content from by-product hemp cake. *Int J Food Sci Technol* 51:885–893. <https://doi.org/10.1111/ijfs.13041>
68. Danlami JM, Zaini MAA, Arsad A, Yunus MAC (2015) A parametric investigation of castor oil (*Ricinus communis* L) extraction using supercritical carbon dioxide via response surface optimization. *J Taiwan Inst Chem Eng* 53:32–39. <https://doi.org/10.1016/j.jtice.2015.02.033>
69. Azmir J, Zaidul ISM, Sharif KM, Uddin MS, Jahurul MHA, Jinap S, Hajeb P, Mohamed A (2014) Supercritical carbon dioxide extraction of highly unsaturated oil from *Phaleria macrocarpa* seed. *Food Res Int* 65:394–400. <https://doi.org/10.1016/j.foodres.2014.06.049>
70. Akanda MdJH, Sarker MdZI, Norulaini N, Ferdosh S, Rahman MM, Omar AKM (2013) Optimization of supercritical carbon dioxide extraction parameters of cocoa butter analogy fat from mango seed kernel oil using response surface methodology. *J Food Sci Technol* 52(1):319–326. <https://doi.org/10.1007/s13197-013-0979-x>
71. Zhang JP, Hou XL, Yu T, Li Y, Dong HY (2012) Response surface optimization of *Nigella glandulifera* Freyn seed oil yield by supercritical carbon dioxide extraction. *J Integr Agric* 11:151–158. [https://doi.org/10.1016/s1671-2927\(12\)60793-7](https://doi.org/10.1016/s1671-2927(12)60793-7)
72. He W, Gao Y, Yuan F, Bao Y, Liu F, Dong J (2010) Optimization of supercritical carbon dioxide extraction of Gardenia fruit oil and the analysis of functional components. *J Am Oil Chem Soc* 87:1071–1079. <https://doi.org/10.1007/s11746-010-1592-z>
73. Nyam KL, Tan CP, Karim R, Lai OM, Long K, Man YBC (2010) Extraction of tocopherol-enriched oils from Kalahari melon and roselle seeds by supercritical fluid extraction (SFE-CO<sub>2</sub>). *Food Chem* 119:1278–1283. <https://doi.org/10.1016/j.foodchem.2009.08.007>
74. Nyam KL, Tan CP, Lai OM, Long K, Che Man YB (2010) Optimization of supercritical fluid extraction of phytosterol from roselle seeds with a central composite design model. *Food Bioprod Process* 88:239–246. <https://doi.org/10.1016/j.fbp.2009.11.002>
75. Liu G, Xu X, Hao Q, Gao Y (2009) Supercritical CO<sub>2</sub> extraction optimization of pomegranate (*Punica granatum* L.) seed oil using response surface methodology. *LWT Food Sci Technol* 42:1491–1495. <https://doi.org/10.1016/j.lwt.2009.04.011>
76. Jiao SS, Li D, Huang ZG, Zhang ZS, Bhandari B, Chen XD, Mao ZH (2008) Optimization of supercritical carbon dioxide extraction of flaxseed oil using response surface methodology. *Int J Food Eng* 4(4). <https://doi.org/10.2202/1556-3758.1409>
77. Özkal SG, Yener ME, Salgın U, Mehmetoğlu Ü (2004) Response surfaces of hazelnut oil yield in supercritical carbon dioxide. *Eur Food Res Technol* 220(1):74–78. <https://doi.org/10.1007/s00217-004-1013-3>
78. Da Porto C, Voinovich D, Decorti D, Natolino A (2012) Response surface optimization of hemp seed (*Cannabis sativa* L.) oil yield and oxidation stability by supercritical carbon dioxide extraction. *J Supercrit Fluids* 68:45–51. <https://doi.org/10.1016/j.supflu.2012.04.008>
79. Louaer M, Zermane A, Larkeche O, Meniai AH (2019) Experimental study and optimization of the extraction of Algerian date stones oil (*Phoenix dactylifera* L.) using supercritical carbon dioxide. *J Food Process Eng* 42:e13049. <https://doi.org/10.1111/jfpe.13049>
80. Li G, Song C, You J, Sun Z, Xia L, Suo Y (2011) Optimisation of red pepper seed oil extraction using supercritical CO<sub>2</sub> and analysis of the composition by reversed-phase HPLC-FLD-MS/MS. *Int J Food Sci Technol* 46:44–51. <https://doi.org/10.1111/j.1365-2621.2010.02441.x>
81. Xia L, You J, Li G, Sun Z, Suo Y (2011) Compositional and antioxidant activity analysis of *Zanthoxylum bungeanum* seed oil obtained by supercritical CO<sub>2</sub> fluid extraction. *J Am Oil Chem Soc* 88:23–32. <https://doi.org/10.1007/s11746-010-1644-4>
82. Li G, Sun Z, Xia L, Shi J, Liu Y, Suo Y, You J (2010) Supercritical CO<sub>2</sub> oil extraction from Chinese star anise seed and simultaneous compositional analysis using HPLC by fluorescence detection and online atmospheric CI-MS identification. *J Sci Food Agric* 90:1905–1913. <https://doi.org/10.1002/jsfa.4031>
83. Palazoglu TK, Balaban MO (1998) Supercritical CO<sub>2</sub> extraction of lipids from roasted pistachio nuts. *Trans ASAE* 41:679–684. <https://doi.org/10.13031/2013.17193>
84. Bogdanovic A, Tadic V, Ristic M, Petrovic S, Skala D (2016) Optimization of supercritical CO<sub>2</sub> extraction of Fenugreek seed (*Trigonella foenum-graecum* L.) and calculating of extracts solubility. *J Supercrit Fluids* 117:297–307. <https://doi.org/10.1016/j.supflu.2016.07.010>
85. Chen C-R, Cheng Y-J, Shieh C-J, Hsiang D, Chang C-MJ (2013) Oil production from de-shelled *Aquilaria crassna* seeds using supercritical carbon dioxide extraction. *J Am Oil Chem Soc* 90:9–16. <https://doi.org/10.1007/s11746-012-2147-2>
86. Chen CR, Cheng YJ, Ching YC, Hsiang D, Chang CMJ (2012) Green production of energetic *Jatropha* oil from de-shelled *Jatropha curcas* L. seeds using supercritical carbon dioxide extraction. *J Supercrit Fluids* 66:137–143. <https://doi.org/10.1016/j.supflu.2012.01.010>
87. Cheng YJ, Shieh CJ, Wang YC, Lai SM, Chang CMJ (2012) Supercritical carbon dioxide extraction of omega-3 oil compounds from *Ficus awkeotsang* Makino achenes. *Sep Purif Technol* 98:62–68. <https://doi.org/10.1016/j.seppur.2012.06.030>
88. Chen WH, Chen CH, Chang CMJ, Liao BC, Hsiang D (2010) Supercritical carbon dioxide extraction of triglycerides from *Aquilaria crassna* seeds. *Sep Purif Technol* 73:135–141. <https://doi.org/10.1016/j.seppur.2010.03.016>
89. Chen WH, Chen CH, Chang CMJ, Chiu YH, Hsiang D (2009) Supercritical carbon dioxide extraction of triglycerides from



- Jatropha curcas* L. seeds. *J Supercrit Fluids* 51:174–180. <https://doi.org/10.1016/j.supflu.2009.08.010>
90. Nik Norulaini NA, Setianto WB, Zaidul ISM, Nawi AH, Azizi CYM, Omar AKM (2009) Effects of supercritical carbon dioxide extraction parameters on virgin coconut oil yield and medium-chain triglyceride content. *Food Chem* 116:193–197. <https://doi.org/10.1016/j.foodchem.2009.02.030>
  91. Ivanov DS, Čolović RR, Lević JD, Sredanović SA (2012) Optimization of supercritical fluid extraction of linseed oil using RSM. *Eur J Lipid Sci Technol* 114:807–815. <https://doi.org/10.1002/ejlt.201100347>
  92. Bimakr M, Rahman RA, Ganjloo A, Taip FS, Adzahan NM, Sarker MZI (2015) Characterization of valuable compounds from Winter Melon (*Benincasa hispida* (Thunb.) Cogn.) seeds using supercritical carbon dioxide extraction combined with pressure swing technique. *Food Bioprocess Technol* 9:396–406. <https://doi.org/10.1007/s11947-015-1636-3>
  93. Bilgiç-Keleş S, Şahin-Yeşilçubuk N, Barla-Demirköz A, Karakaş M (2019) Response surface optimization and modelling for supercritical carbon dioxide extraction of *Echium vulgare* seed oil. *J Supercrit Fluids* 143:365–369. <https://doi.org/10.1016/j.supflu.2018.09.008>
  94. de Oliveira NA, Mazzali MR, Fukumasu H, Gonçalves CB, de Oliveira AL (2019) Composition and physical properties of babassu seed (*Orbignya phalerata*) oil obtained by supercritical CO<sub>2</sub> extraction. *J Supercrit Fluids* 150:21–29. <https://doi.org/10.1016/j.supflu.2019.04.009>
  95. Gustinelli G, Eliasson L, Svelander C, Alminger M, Ahrné L (2018) Supercritical CO<sub>2</sub> extraction of bilberry (*Vaccinium myrtillus* L.) seed oil: fatty acid composition and antioxidant activity. *J Supercrit Fluids* 135:91–97. <https://doi.org/10.1016/j.supflu.2018.01.002>
  96. Jokić S, Bijuk M, Aladić K, Bilić M, Molnar M (2016) Optimisation of supercritical CO<sub>2</sub> extraction of grape seed oil using response surface methodology. *Int J Food Sci Technol* 51:403–410. <https://doi.org/10.1111/ijfs.12986>
  97. Zahedi G, Azarpour A (2011) Optimization of supercritical carbon dioxide extraction of *Passiflora* seed oil. *J Supercrit Fluids* 58:40–48. <https://doi.org/10.1016/j.supflu.2011.04.013>
  98. Began G, Manohar B, Udaya Sankar K, Appu Rao AG (2000) Response surfaces for solubility of crude soy lecithin lipid in supercritical carbon dioxide. *Eur Food Res Technol* 210(3):209–212. <https://doi.org/10.1007/pl00005513>
  99. Wang W, Han S, Jiao Z, Cheng J, Song J (2019) Antioxidant activity and total polyphenols content of camellia oil extracted by optimized supercritical carbon dioxide. *J Am Oil Chem Soc* 96:1275–1289. <https://doi.org/10.1002/aocs.12285>
  100. Bernardo-Gil MG, Grenha J, Santos J, Cardoso P (2002) Supercritical fluid extraction and characterization of oil from hazelnut. *Eur J Lipid Sci Technol* 104:402–409. [https://doi.org/10.1002/1438-9312\(200207\)104:7<402:aid-ejlt402>3.0.co;2-n](https://doi.org/10.1002/1438-9312(200207)104:7<402:aid-ejlt402>3.0.co;2-n)
  101. Soares JF, Prá VD, Barrales FM, dos Santos P, Kuhn RC, Rezende CA, Martínez J, Mazutti MA (2018) Extraction of rice bran oil using supercritical CO<sub>2</sub> combined with ultrasound. *Braz J Chem Eng* 35:785–794. <https://doi.org/10.1590/0104-6632.20180352s20160447>
  102. Comin LM, Temelli F, Saldaña MA (2010) Supercritical CO<sub>2</sub> extraction of flax lignans. *J Am Oil Chem Soc* 88(5):707–715. <https://doi.org/10.1007/s11746-010-1704-9>
  103. Sheibani A, Ghaziaskar HS (2008) Pressurized fluid extraction of pistachio oil using a modified supercritical fluid extractor and factorial design for optimization. *LWT Food Sci Technol* 41:1472–1477. <https://doi.org/10.1016/j.lwt.2007.09.002>
  104. Naik SN, Lentz H, Maheshwari RC (1989) Extraction of perfumes and flavours from plant materials with liquid carbon dioxide under liquid–vapor equilibrium conditions. *Fluid Phase Equilib* 49:115–126. [https://doi.org/10.1016/0378-3812\(89\)80009-3](https://doi.org/10.1016/0378-3812(89)80009-3)
  105. Brunner G (1994) Gas extraction: an introduction to fundamentals of supercritical fluids and the application to separation processes. Springer, New York. [https://doi.org/10.1007/978-3-662-07380-3\\_1](https://doi.org/10.1007/978-3-662-07380-3_1)
  106. del Valle JM, de la Fuente JC, Uquiche E (2012) A refined equation for predicting the solubility of vegetable oils in high-pressure CO<sub>2</sub>. *J Supercrit Fluids* 67:60–70. <https://doi.org/10.1016/j.supflu.2012.02.004>
  107. Reverchon E, Marrone C (2001) Modeling and simulation of the supercritical CO<sub>2</sub> extraction of vegetable oils. *J Supercrit Fluids* 19:161–175. [https://doi.org/10.1016/s0896-8446\(00\)00093-0](https://doi.org/10.1016/s0896-8446(00)00093-0)
  108. del Valle JM, Germain JC, Uquiche E, Zetzl C, Brunner G (2006) Microstructural effects on internal mass transfer of lipids in prepressed and flaked vegetable substrates. *J Supercrit Fluids* 37:178–190. <https://doi.org/10.1016/j.supflu.2005.09.002>
  109. Reyes FA, Muñoz LA, Hansen A, del Valle JM (2015) Water relationships in *Haematococcus pluvialis* and their effect in high-pressure agglomeration for supercritical CO<sub>2</sub> extraction. *J Food Eng* 162:18–24. <https://doi.org/10.1016/j.jfoodeng.2015.03.038>
  110. del Valle JM, Glatzel V, Martínez JL (2012) Supercritical CO<sub>2</sub> extraction of allicin from garlic flakes: screening and kinetic studies. *Food Res Int* 45:216–224. <https://doi.org/10.1016/j.foodres.2011.10.021>
  111. Eggers R, Ambrogi A, von Schnitzler J (2000) Special features of SCF solid extraction of natural products: deoiling of wheat gluten and extraction of rose hip oil. *Braz J Chem Eng* 17:329–334. <https://doi.org/10.1590/S0104-6632200000300009>
  112. Fernández-Trujillo JP (2007) Extracción convencional de oleoresina de pimentón dulce y picante I. Generalidades, composición, proceso e innovaciones y aplicaciones. *Grasas Aceites* 58:252–263
  113. Sielfeld C, del Valle JM, Sastre F (2019) Effect of pelletization on supercritical CO<sub>2</sub> extraction of rosemary antioxidants. *J Supercrit Fluids* 147:162–171. <https://doi.org/10.1016/j.supflu.2016.04.010>
  114. Uquiche E, del Valle JM, Ihl M (2005) Microstructure-extractability relationships in the extraction of prepelletized Jalapeño peppers with supercritical carbon dioxide. *J Food Sci* 70:E379–E386. <https://doi.org/10.1111/j.1365-2621.2005.tb11442.x>
  115. Muthukumarappan K, Swamy GJ (2020). Extrusion processing of foods. In M.S. Rahman (Ed), *Handbook of Food Preservation*, 3rd edition., pp. 647–658). CRC Press, Boca Raton, FL. ISBN 978-0-429-09148-3
  116. NIST Standard Reference Database 23: Reference Fluid Thermodynamic and Transport Properties-REFPROP, Version 10.0 (2018) National institute of standards and technology, standard reference data program, Gaithersburg. <https://www.nist.gov/srd/refprop>. Accessed 8 Oct 2023.
  117. Uquiche E, Millao S, del Valle JM (2022) Extrusion affects supercritical CO<sub>2</sub> extraction of red pepper (*Capsicum annuum* L.) oleoresin. *J Food Eng* 316:110829. <https://doi.org/10.1016/j.jfoodeng.2021.110829>
  118. Arous-Sarmiento KA (2013) Identification of the effect of entrainers (triolein or ethanol) on the carotenoids extraction from red paprika (*Capsicum annuum* L.) using supercritical carbon dioxide (SC-CO<sub>2</sub>). Pontificia Universidad Católica de Chile, Santiago, Chile
  119. Millao S, Uquiche E (2016) Extraction of oil and carotenoids from pelletized microalgae using supercritical carbon dioxide. *J Supercrit Fluids* 116:223–231. <https://doi.org/10.1016/j.supflu.2016.05.049>
  120. Arous K, Temelli F, del Valle JM, de la Fuente JC, Robert P (2011) Supercritical extraction of petals and pellets of marigold flowers using ethanol-modified CO<sub>2</sub>. In XI Conference on Engineering and Food (ICEF 11), Athens, Greece

121. Winther K, Sophie Vinther Hansen A, Campbell-Tofte J (2016) Bio-active ingredients of rose hips (*Rosa canina* L) with special reference to antioxidative and anti-inflammatory properties: in vitro studies. *Botanics* 6:11–23. <https://doi.org/10.2147/btat.s91385>
122. Dąbrowska M, Maciejczyk E, Kalembe D (2019) Rose hip seed oil: methods of extraction and chemical composition. *Eur J Lipid Sci Technol* 121:1800440. <https://doi.org/10.1002/ejlt.201800440>
123. Mabe GD, Foco GM, Brignole EA, Bottini SB (1999) Extraction of rosa mosqueta (*Rosa aff. rubiginosa*) oil with dense fluids. *Acta Horticulturae*. International Society for Horticultural Science (ISHS), Leuven, Belgium, pp 37–44
124. Reverchon E, Kazianus A, Marrone C (2000) Supercritical CO<sub>2</sub> extraction of hiprose seed oil: experiments and mathematical modelling. *Chem Eng Sci* 55:2195–2201. [https://doi.org/10.1016/S0009-2509\(99\)00519-9](https://doi.org/10.1016/S0009-2509(99)00519-9)
125. Szentmihályi K, Vinkler P, Lakatos B, Illés V, Then M (2002) Rose hip (*Rosa canina* L.) oil obtained from waste hip seeds by different extraction methods. *Bioresour Technol* 82:195–201. [https://doi.org/10.1016/S0960-8524\(01\)00161-4](https://doi.org/10.1016/S0960-8524(01)00161-4)
126. del Valle JM, Rivera O, Mattea M, Ruetsch L, Daghero J, Flores A (2004) Supercritical CO<sub>2</sub> processing of pretreated rosehip seeds: effect of process scale on oil extraction kinetics. *J Supercrit Fluids* 31:159–174. <https://doi.org/10.1016/j.supflu.2003.11.005>
127. Machmudah S, Kawahito Y, Sasaki M, Goto M (2007) Supercritical CO<sub>2</sub> extraction of rosehip seed oil: fatty acids composition and process optimization. *J Supercrit Fluids* 41:421–428. <https://doi.org/10.1016/j.supflu.2006.12.011>
128. Salgın U, Salgın S, Ekici DD, Uludağ G (2016) Oil recovery in rosehip seeds from food plant waste products using supercritical CO<sub>2</sub> extraction. *J Supercrit Fluids* 118:194–202. <https://doi.org/10.1016/j.supflu.2016.08.011>
129. Jahongir H, Miansong Z, Amankeldi I, Yu Z, Changheng L (2019) The influence of particle size on supercritical extraction of dog rose (*Rosa canina*) seed oil. *J King Saud Univ Eng Sci* 31:140–143. <https://doi.org/10.1016/j.jksues.2018.04.004>
130. Illés V, Szalai O, Then M, Daood H, Perneczki S (1997) Extraction of hiprose fruit by supercritical CO<sub>2</sub> and propane. *J Supercrit Fluids* 10:209–218. [https://doi.org/10.1016/S0896-8446\(97\)00018-1](https://doi.org/10.1016/S0896-8446(97)00018-1)
131. Machmudah S, Kawahito Y, Sasaki M, Goto M (2008) Process optimization and extraction rate analysis of carotenoids extraction from rosehip fruit using supercritical CO<sub>2</sub>. *J Supercrit Fluids* 44:308–314. <https://doi.org/10.1016/j.supflu.2007.09.032>
132. TTozzi R, Mulinacci N, Storlikken K, Pasquali I, Vincieri FF, Bettini R (2008) Supercritical extraction of carotenoids from *Rosa canina* L. hips and their formulation with  $\beta$ -cyclodextrin. *AAPS PharmSciTech* 9:693. <https://doi.org/10.1208/s12249-008-9100-6>
133. Watanabe Y, Honda M, Higashiura T, Fukaya S, Machmudah S, Wahyudiono K, H., & Goto, M. (2018) Rapid and selective concentration of lycopene z-isomers from tomato pulp by supercritical CO<sub>2</sub> with co-solvents. *Solvent Extr Res Dev, Jpn* 25:47–57
134. Shi J, Yi C, Xue SJ, Jiang Y, Ma Y, Li D (2009) Effects of modifiers on the profile of lycopene extracted from tomato skins by supercritical CO<sub>2</sub>. *J Food Eng* 93:431–436. <https://doi.org/10.1016/j.jfoodeng.2009.02.008>
135. Vasapollo G, Longo L, Rescio L, Ciurlia L (2004) Innovative supercritical CO<sub>2</sub> extraction of lycopene from tomato in the presence of vegetable oil as co-solvent. *J Supercrit Fluids* 29:87–96. [https://doi.org/10.1016/S0896-8446\(03\)00039-1](https://doi.org/10.1016/S0896-8446(03)00039-1)
136. Sun M, Temelli F (2006) Supercritical carbon dioxide extraction of carotenoids from carrot using canola oil as a continuous co-solvent. *J Supercrit Fluids* 37(3):397–408. <https://doi.org/10.1016/j.supflu.2006.01.008>
137. Palumpitag W, Prasitchoke P, Goto M, Shotipruk A (2011) Supercritical carbon dioxide extraction of marigold lutein fatty acid esters: effects of cosolvents and saponification conditions. *Sep Sci Technol* 46:605–610. <https://doi.org/10.1080/01496395.2010.533739>
138. Ma Q, Xu X, Gao Y, Wang Q, Zhao J (2008) Optimisation of supercritical carbon dioxide extraction of lutein esters from marigold (*Tagetes erect* L.) with soybean oil as a co-solvent. *Int J Food Sci Technol* 43:1763–1769. <https://doi.org/10.1111/j.1365-2621.2007.01694.x>
139. Razi Parjikolaei B, Cardoso LC, Fernández-Ponce MT, Mantell Serrano C, Frette XC, Christensen KV (2015) Northern shrimp (*Pandalus borealis*) processing waste: effect of supercritical fluid extraction technique on carotenoid extract concentration. *Chem Eng Trans* 43:1045–1050
140. Ciurlia L, Blevé M, Rescio L (2009) Supercritical carbon dioxide co-extraction of tomatoes (*Lycopersicon esculentum* L.) and hazelnuts (*Corylus avellana* L.): a new procedure in obtaining a source of natural lycopene. *J Supercrit Fluids* 49:338–344. <https://doi.org/10.1016/j.supflu.2009.03.003>
141. Barros HDFQ, Grimaldi R, Cabral FA (2017) Lycopene-rich avocado oil obtained by simultaneous supercritical extraction from avocado pulp and tomato pomace. *J Supercrit Fluids* 120:1–6. <https://doi.org/10.1016/j.supflu.2016.09.021>
142. Barros HDFQ, Coutinho JP, Grimaldi R, Godoy HT, Cabral FA (2016) Simultaneous extraction of edible oil from avocado and capsanthin from red bell pepper using supercritical carbon dioxide as solvent. *J Supercrit Fluids* 107:315–320. <https://doi.org/10.1016/j.supflu.2015.09.025>
143. Getachew AT, Saravana PS, Cho YJ, Woo HC, Chun BS (2018) Concurrent extraction of oil from roasted coffee (*Coffea arabica*) and fucoxanthin from brown seaweed (*Saccharina japonica*) using supercritical carbon dioxide. *J CO<sub>2</sub> Utiliz* 25:137–146. <https://doi.org/10.1016/j.jcou.2018.03.018>
144. Machmudah S, Winardi S, Sasaki M, Goto M, Kusumoto N, Hayakawa K (2012) Lycopene extraction from tomato peel by-product containing tomato seed using supercritical carbon dioxide. *J Food Eng* 108:290–296. <https://doi.org/10.1016/j.jfoodeng.2011.08.012>
145. Ndayishimiye J, Chun BS (2017) Optimization of carotenoids and antioxidant activity of oils obtained from a co-extraction of citrus (*Yuzu ichandrin*) by-products using supercritical carbon dioxide. *Biomass Bioenerg* 106:1–7. <https://doi.org/10.1016/j.biombioe.2017.08.014>
146. (2022) Flavex Naturextrakte: General Specification Rosehip CO<sub>2</sub>-to extract (organic) DE-ÖKO- 013, Type No. 046.005. [https://www.flavex.com/en/produkt/046\\_005/](https://www.flavex.com/en/produkt/046_005/)
147. Flavex Naturextrakte: General Specification Rosehip Seed CO<sub>2</sub>-to extract (organic) DE- ÖKO-013, Type No. 046.004. [https://www.flavex.com/en/produkt/046\\_004/](https://www.flavex.com/en/produkt/046_004/)
148. del Valle JM (2015) Extraction of natural compounds using supercritical CO<sub>2</sub>: going from the laboratory to the industrial application. *J Supercrit Fluids* 96:180–199. <https://doi.org/10.1016/j.supflu.2014.10.001>
149. Gardner DS (1982) Industrial scale hop extraction with liquid carbon dioxide. *Chem Ind* 19:402–405
150. Funazukuri T, Toriumi M, Yui K, Kong CY, Kagei S (2009) Correlation for binary diffusion coefficients of lipids in supercritical carbon dioxide. In 9th International Symposium on Supercritical Fluids, Arcachon, France
151. King MB, Catchpole O (1993) Physico-chemical data required for the near-critical fluid extraction process. In: King MJ, Bott TR (eds) Extraction of natural products using near-critical solvents. Blackie Academic & Professional, Glasgow, UK, pp 184–231
152. Sovová H (2012) Steps of supercritical fluid extraction of natural products and their characteristic times. *J Supercrit Fluids* 66:73–79. <https://doi.org/10.1016/j.supflu.2011.11.004>



153. Sovová H (2005) Mathematical model for supercritical fluid extraction of natural products and extraction curve evaluation. *J Supercrit Fluids* 33:35–52. <https://doi.org/10.1016/j.supflu.2004.03.005>
154. Sovová H (1994) Rate of the vegetable oil extraction with supercritical CO<sub>2</sub>—I. Modelling of extraction curves. *Chem Eng Sci* 49:409–414. [https://doi.org/10.1016/0009-2509\(94\)87012-8](https://doi.org/10.1016/0009-2509(94)87012-8)
155. del Valle JM, Jiménez M, de la Fuente JC (2003) Extraction kinetics of pre-pelletized Jalapeño peppers with supercritical CO<sub>2</sub>. *J Supercrit Fluids* 25:33–44. [https://doi.org/10.1016/s0896-8446\(02\)00090-6](https://doi.org/10.1016/s0896-8446(02)00090-6)
156. Arias J, Martínez J, Stashenko E, del Valle JM, Núñez GA (2022) Supercritical CO<sub>2</sub> extraction of pinocembrin from *Lippia origanoides* distillation residues. 2. Mathematical modeling of mass transfer kinetics as a function of substrate pretreatment. *J Supercrit Fluids* 180:105458. <https://doi.org/10.1016/j.supflu.2021.105458>
157. Urrego FA, Núñez GA, Donaire YD, del Valle JM (2015) Equilibrium partition of rapeseed oil between supercritical CO<sub>2</sub> and prepressed rapeseed. *J Supercrit Fluids* 102:80–91. <https://doi.org/10.1016/j.supflu.2015.04.004>
158. Arous KA, Canales RI, del Valle JM, de la Fuente JC (2011) Solubility of  $\beta$ -carotene in ethanol- and triolein-modified CO<sub>2</sub>. *J Chem Thermodyn* 43:1991–2001. <https://doi.org/10.1016/j.jct.2011.07.013>
159. Arous KA, del Valle JM, Robert PS, de la Fuente JC (2012) Effect of triolein addition on the solubility of capsanthin in supercritical carbon dioxide. *J Chem Thermodyn* 51:190–194. <https://doi.org/10.1016/j.jct.2012.02.030>
160. Mansoori GA, Schulz K, Martinelli E (1988) Bioseparation using supercritical fluid extraction/retrograde condensation. *Bio/Technology* 6:393–396
161. Foster NR, Gurdial GS, Yun JSL, Liong KK, Tilly KD, Ting SST, Singh H, Lee JH (1991) Significance of the crossover pressure in solid-supercritical fluid phase equilibria. *Ind Eng Chem Res* 30:1955–1964. <https://doi.org/10.1021/ie00056a044>
162. Chrastil J (1982) Solubility of solids and liquids in supercritical gases. *J Phys Chem* 86:3016–3021. <https://doi.org/10.1021/j100212a041>
163. Valenzuela LM, Reveco-Chilla AG, del Valle JM (2014) Modeling solubility in supercritical carbon dioxide using quantitative structure–property relationships. *J Supercrit Fluids* 94:113–122. <https://doi.org/10.1016/j.supflu.2014.06.022>
164. Palma M, Taylor LT (1999) Statistical design for optimization of extraction of polyphenols from an inert matrix using carbon dioxide-based fluids. *Anal Chim Acta* 391:321–329. [https://doi.org/10.1016/s0003-2670\(99\)00210-x](https://doi.org/10.1016/s0003-2670(99)00210-x)
165. Palma M, Taylor LT (1999) Extraction of polyphenolic compounds from grape seeds with near critical carbon dioxide. *J Chromatogr A* 849:117–124. [https://doi.org/10.1016/s0021-9673\(99\)00569-5](https://doi.org/10.1016/s0021-9673(99)00569-5)
166. Coelho SRM, Araújo JMA, Scheuermann ESS (2000) Remoção de limoneno do óleo essencial de limão siciliano adsorvido em sílica gel pelo CO<sub>2</sub> supercrítico. *Arquivos de Ciências da Saúde da UNIPAR* 4(247–250):63
167. Millao S, Uquiche E (2016) Antioxidant activity of supercritical extracts from *Nannochloropsis gaditana*: correlation with its content of carotenoids and tocopherols. *J Supercrit Fluids* 111:143–150. <https://doi.org/10.1016/j.supflu.2016.02.002>

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