**RESEARCH ARTICLE**





# **Efects of Foliar Potassium Supplementation on Yield and Nutrient Uptake of Plant Sugarcane**

NattawatRadasai<sup>1</sup> · Daojarus Ketrot<sup>1</sup> D · Saowanuch Tawornpruek<sup>1</sup> · Tawatchai Inboonchuay<sup>2</sup> · **Acharaporn Wongsuksri3**

Received: 18 March 2024 / Accepted: 25 June 2024 © The Author(s), under exclusive licence to Society for Sugar Research & Promotion 2024

#### **Abstract**

Sugarcane, a globally signifcant economic crop, depends on potassium (K) for critical processes such as photosynthesis and sugar translocation. This study explored the impacts of various foliar K supplements, including  $2.5\%$  w/v KCl, KNO<sub>3</sub>,  $K_2SO_4$ , and  $K_2SiO_3$ , alongside diluted molasses and vinasse (5 × dilution). The field experiment was conducted on sugarcane grown in soil with sufficient soil K levels, applying foliar solutions at 120 days at 2667 L/ha. The results indicated that combining soil chemical fertilizers with foliar  $K_2SiO_3$  and  $KNO_3$  resulted in the highest yields of 155.19 and 154.81 tons/ ha, respectively, signifcantly outperforming the foliar water combined with soil chemical fertilizers (132.81 tons/ha) and the control (no basal fertilizer with foliar water, at 130.67 tons/ha, *P*≤0.05). This enhancement is expected to result from the improvement in chlorophyll content and photosynthesis, enabled by timely K and nutrient acquisition, bypassing root transport. However, no signifcant diferences were noted among the foliar K forms. Foliar K application also afected nutrient concentrations and uptake, with molasses showing the highest nutrient absorption in stalks: N (322 kg/ha), K (215 kg/ha), S (80.9 kg/ha), and Si (23.2 kg/ha) ( $P \le 0.05$ ). These findings provide valuable insights and recommendations for utilizing foliar application of  $K_2SiO_3$  and  $KNO_3$  to improve plant sugarcane yield, as well as employing molasses foliar application to enhance nutrient uptake in sugarcane cultivated in soils with adequate K.

**Keywords** Foliar fertilizer · Potassium silicate · Potassium nitrate · Molasses · Plant sugarcane

# **Introduction**

Sugarcane (*Saccharum* spp.) is an essential agro-industrial crop cultivated in over 130 countries across tropical and subtropical regions, primarily for sugar, ethanol, electricity, and other derivatives production (de Souza Barbosa et al. [2020](#page-9-0)). As the leading primary crop, sugarcane represented 20 percent of global production in 2020, with 1.9 billion tonnes (FAO [2022](#page-9-1)). However, yields vary signifcantly, ranging

- Department of Soil Science, Faculty of Agriculture, Kasetsart University, Bangkok 10900, Thailand
- Department of Soil Science, Faculty of Agriculture at Kamphaeng Saen, Kasetsart University, Kamphaeng Saen Campus, Nakhon Pathom 73140, Thailand
- <sup>3</sup> Suphan Buri Field Crops Research Center, Department of Agriculture, U-Thong District, Suphan Buri 72160, Thailand

Published online: 09 July 2024

from 6.27 to 120.74 tons/ha across 91 countries in 2023 (FAOSTAT [2024](#page-9-2)), infuenced by factors such as the growing period length, crop type (plant or ratoon), management practices, and environmental conditions. This variation is in stark contrast to the theoretical yield potential of up to 381 tons/ha (Waclawovsky et al. [2010](#page-10-0)), indicating substantial room for production improvement. Enhancing sugarcane productivity hinges on selecting crop varieties, optimizing growth environments, and implementing efective management practices. Particularly, potassium (K) management is vital for improving yield and quality, given its signifcant roles in growth, photosynthesis, drought and cold resistance (Jaiswal et al. [2021](#page-9-3); Johnson et al. [2022](#page-9-4)), and sugar transport, impacting sugarcane juice quality and parameters like commercial cane sugar (CCS), brix, and overall sugar yield (Watanabe et al. [2016;](#page-10-1) Johnson et al. [2022\)](#page-9-4).

Potassium chloride (KCl) is the most widely used K fertilizer in crop nutrition, primarily due to its low cost and high K<sub>2</sub>O content, which reduces handling, storage, transport, and application expenses (Mikkelsen and Roberts [2021](#page-9-5)).

 $\boxtimes$  Daojarus Ketrot fagrdrk@ku.ac.th

However, studies show that  $K^+$  and  $Cl^-$  are abundant in sugarcane juice and negatively correlate with sucrose concentration. Cl<sup>−</sup> can adversely affect sugarcane quality if K<sup>+</sup> is supplied in excess (Watanabe et al. [2016;](#page-10-1) Watanabe et al. [2017](#page-10-2)). Other K fertilizers, like  $KNO_3$ ,  $K_2SO_4$ , and  $K_2SiO_3$ , provide essential nutrients including N, S, and Si. N is vital for growth and development (Zeng et al. [2020\)](#page-10-3), S enhances yield and sweetness (Abdelrahman et al. [2014\)](#page-9-6), and Si helps mitigate water stress, improving transpiration, photosynthesis, and nutrient use efficiency while reducing water deficit damage (Teixeira et al. [2022;](#page-9-7) Costa et al. [2023\)](#page-8-0). Molasses and vinasse, by-products of the sugar industry, are rich in nutrients, particularly K, and their application in sugarcane cultivation can improve productivity and quality, ofering a cost-efective nutrient source (Christofoletti [2013](#page-8-1); Pal-monari et al. [2020\)](#page-9-8). Jiang et al. [\(2012\)](#page-9-9) reported that continuous application of vinasse to sugarcane felds for 2–3 years resulted in enhanced soil physicochemical properties. Specifcally, they observed a decrease in soil bulk density and an increase in total porosity, capillary porosity, and soil K content. Similarly, Wu et al. [\(2023](#page-10-4)) found that the combined application of chemical fertilizer with moderate molasses (160 mg C/kg soil) can increase sugarcane biomass grown in pot experiment and facilitate the utilization of stable organic phosphorus (Po) by altering bacterial community and soil properties.

Foliar fertilization involves applying nutrients directly to plant leaves, allowing for efficient nutrient absorption, particularly during growth spurts or when additional nutrition is required. While the increased use of foliar fertilizers in sugarcane cultivation has been explored, most studies focus on comparing or combining foliar with soil applications. Notably, foliar application of K in maize, even with suffcient soil K levels, has been shown to improve yield and sweetness by promoting chlorophyll synthesis (Suwannarit and Sestapukdee [1989](#page-9-10)). This fnding has led to research on the effects of foliar K supplementation in sugarcane fields with adequate soil K, using various K fertilizer formulations to examine their impact on productivity and nutrient uptake. The research also considers the use of molasses and vinasse in foliar applications, aiming to provide a source of supplementary foliar K fertilizer.

# **Materials and Methods**

## **Study Area and Soil Analysis**

To investigate the effects of foliar K supplementation on sugarcane yield and nutrient uptake, a feld experiment was conducted at the Kasetsart University Kamphaeng Saen campus, located in Nakhon Pathom province, central Thailand (coordinates: 14°02′09.6″ N, 99°57′33.4″ E, at an elevation of 9 m). The study site's soil is part of the Kamphaeng Saen (Ks) Series, classifed as Typic Haplustalfs. A composite soil sample was collected from the topsoil (0–15 cm) before the experiment, air-dried, homogenized by grinding and sieving (0.5 mm and 2 mm), and analyzed using standard physicochemical methods (Soil Survey Staff [2014\)](#page-9-11). The analysis included soil texture, pH1:1, electrical conductivity at saturated soil paste extracts (ECe), organic matter (OM), cation exchange capacity (CEC), total N, available P, K, Ca, and Mg. For extractable S and Si in soil, assessment was conducted using  $Ca(H_2PO_4)_2$  and KCl extraction methods, respectively (Fox et al. [1964](#page-9-12); Crusciol et al. [2018\)](#page-8-2).

## **Analysis of Molasses and Vinasse, a By‑Product from Sugarcane‑Sugar Industry**

In this study, molasses and vinasse were analyzed for pH and EC using a pH meter and an electrical conductivity meter, respectively. Total solids (TS) and suspended solids (SS) were determined through dry evaporation and Buchner's funnel methods, with dissolved solids (DS) calculated as DS=TS−SS. The total concentrations of elements such as P, K, Ca, Mg, S, Cu, Fe, Mn, Zn, Ni, Si, Na, Al, Cr, As, and Cd in molasses and vinasse were analyzed via aqua regia digestion (3:1 v/v, HCl:  $HNO<sub>3</sub>$ ) (Chen and Ma [2001\)](#page-8-3), followed by quantifcation using an inductively coupled plasma-optical emission spectrometer (ICP-OES) (Optima 8300, PerkinElmer, Waltham, MA, USA). P and S concentrations were measured using a UV–Visible spectrophotometer (Lambda 265, PerkinElmer), while total N was determined through wet oxidation with the micro-Kjeldahl method (Soil Survey Staff [2014\)](#page-9-11).

#### **Field Experiment and Sugarcane Cultivation**

The feld experiment was set up as a randomized complete block design (RCBD) with four replications. The Khon Kaen 3 variety of *Saccharum* spp. hybrid, known for its drought tolerance, high yield, sugar content, stalk number, and favorable ratooning characteristics, was selected (Ponragdee et al. [2011](#page-9-13)). The experiment took place during the 2022/23 growing season, focusing on the plant cane. Eight treatments of foliar K applications were established: T1—Control (no basal fertilizer+foliar water); T2−BF+water (basal fertilizer+foliar water); T3−BF+ KCl (basal fertilizer+2.5% w/v foliar KCl); T4−BF+ KNO<sub>3</sub> (basal fertilizer + 2.5% w/v foliar KNO<sub>3</sub>); T5−BF+K<sub>2</sub>SO<sub>4</sub> (basal fertilizer + 2.5% w/v foliar K<sub>2</sub>SO<sub>4</sub>); T6−BF + K<sub>2</sub>SiO<sub>3</sub> (basal fertilizer + 2.5% w/v foliar K<sub>2</sub>SiO<sub>3</sub>); T7−BF+molasses (basal fertilizer+foliar diluted molasses at 5x); and T8−BF+vinasse (basal fertilizer+foliar diluted vinasse at 5×). Each experimental plot measured  $7.5 \times 10 \text{ m}^2$ , consisting of five rows of sugarcane with a 1.5 m inter-row spacing and 2.0 m between plots.

Foliar fertilizer solution	pH	$EC$ ( $dS/m$ )	$K$ (%, w/v)		
Water	7.18	0.03	<b>Not</b> detected		
$2.5\%$ w/v KCl	6.66	38.35	1.45		
$2.5\%$ w/v KNO <sub>3</sub>	9.02	26.50	1.00		
2.5%w/v $K_2SO_4$	2.82	28.48	1.04		
$2.5\%$ w/v K <sub>2</sub> SiO <sub>3</sub>	4.50	14.88	0.20		
Diluted molasses $(5 \times$ dilution)	4.88	17.00	0.97		
Diluted vinasse $(5 \times$ dilution)	4.50	17.21	0.53		

<span id="page-2-0"></span>**Table 1** pH and electrical conductivity of foliar fertilizer solution used in this study

Basal fertilizer application followed the Department of Agriculture Thailand's 2021 recommendations in accordance with the soil analysis values (OM, available P, and available K) (Department of Agriculture [2021](#page-9-14)): 75 kg N/ ha, 18.75 kg  $P_2O_5/ha$ , and 112.5 kg K<sub>2</sub>O/ha, applied 30 days post-planting for treatments T2–T8. However, N fertilizer was applied twice, at 30 and 90 days post-planting. Foliar K fertilizers were applied at 120 days post-planting, during the grand or vegetative growth phase (120–240 days) (Somard et al. [2021\)](#page-9-15), using a battery sprayer at 2667 L/ha (20 L/ plot). During this phase, sugarcane undergoes rapid growth in height and biomass developing a robust root system and a dense canopy of leaves. The solution was sprayed on the tops and stalks to ensure even coverage. Two control treatments (T1 and T2) involved foliar water application to assess the potential impact of water application. A leaf binder  $(C_9H_{19}C_6H_4(OCH_2CH_2)_9OH)$ , which enhancing absorption and reducing loss from rain wash-of, was added to all fertilizer solutions at a ratio of 1 mL–2 L. Table [1](#page-2-0) details the pH, EC, and K content of the foliar fertilizer solution, diluted molasses, and vinasse used in the experiment.

## **Sugarcane Growth Parameters and Productive Yields**

Sugarcane was harvested 12 months after planting from plots of 9  $m<sup>2</sup>$ , focusing on three central rows, each two meters long, to determine fresh weight and stalk count. Ten representative stalks were randomly selected from each plot to measure stalk length, diameter, and sugarcane juice quality. Juice quality assessments included Fiber (%) obtained by spinning and drying, Brix (%) measured with a refractometer, and Polarity (%) evaluated using a polarimeter. Subsequently, Thailand's commercial cane sugar (CCS), originally known as Pure Obtainable Cane Sugar or POCS, was calculated as follows (Kirasak et al. [2015;](#page-9-16) Kulasekara et al. [2024](#page-9-17)):

$$
CCS (\%) = \frac{3(\% \text{Polarity})}{2} \left( 1 - \frac{(\% \text{Fiber}) + 5}{100} \right) - \frac{\% \text{Brix}}{2} \left( 1 - \frac{(\% \text{Fiber}) + 3}{100} \right) \tag{1}
$$

Purity, sugar recovery, and sugar yield also were calculated as follows (Mehareb and Abazied [2017;](#page-9-18) Divakar et al. [2023](#page-9-19)):

$$
Purity (\%) = \left(\frac{\% \text{Polarity}}{\% \text{Brix}}\right) \times 100 \tag{2}
$$

Sugar recovery (%) =  $[\%$  Polarity-0.4(% Brix – % Polarity)  $\times$  0.73 (3)

Sugar yield (tons/ha) = 
$$
\frac{\text{Yield (tons/ha)} \times \% \text{Sugar recovery}}{100}
$$
 (4)

Furthermore, cane stalks and tops, including leaves and upper parts, were randomly collected from each plot and oven-dried at 70 °C to achieve a consistent dry weight. These samples were homogenized by grinding. The total N content in the cane stalk and tops was determined using the micro-Kjeldahl method for wet oxidation  $(H_2SO_4-NaSO_4-Se)$ , the resulting solution was then used to measure the total concentrations of P, K, and Si. To measure the total concentrations of S, an acid digestion mixture (3:1  $HNO<sub>3</sub>:HClO<sub>4</sub>$ ) was used. The concentrations of P and S were determined through UV–Visible spectrophotometry, whereas K and Si concentrations were analyzed using atomic absorption spectroscopy (AAS). These concentration data were then used to calculate the nutrient uptake of sugarcane per unit area.

#### **Statistical Analysis**

All data collected in this study, encompassing growth parameters, productive yields, nutrient concentrations, and nutrient uptakes, were analyzed using analysis of variance (ANOVA) with F-tests to statistically evaluate mean diferences. Post hoc multiple comparisons were conducted to identify signifcant diferences among treatments, using Duncan's test at the 0.05 probability level ( $P \le 0.05$ ).

# **Results and Discussion**

#### **General Soil Characteristics**

The soil properties (Table [2](#page-3-0)) show that the soil had a loamy texture, a natural pH of 6.96, and was non-saline, with an ECe of 1.24 dS/m. The soil had a low CEC of 12.49 cmol $\sim$ kg and a moderate OM content of 17.30 g/kg. The total N, available P, and K in the soil were measured at 0.74 g/ kg, 86.89 mg/kg, and 50.23 mg/kg, respectively. Additionally, the soil contained 3,407 mg/kg of available Ca and 168.71 mg/kg of Mg, while extractable S and Si were 39.25 mg/kg and 16.12 mg/kg, respectively.

<span id="page-3-0"></span>**Table 2** Physicochemical properties of the studied soil before sugarcane cultivation

<span id="page-3-1"></span>**Table 3** Chemical properties and composition of molasses and vinasse used in this study

Parameters	Unit	Value	
Sand	g/kg	332	
Silt	g/kg	255	
Clay	g/kg	413	
Soil texture		Loam	
pH (1:1 H <sub>2</sub> O)		6.96	
ECe	dS/m	1.24	
Cation exchange capacity; CEC	cmol./kg	12.49	
Organic matter; OM	g/kg	17.30	
Total N	g/kg	0.74	
Available P	mg/kg	86.89	
Available K	mg/kg	50.23	
Available Ca	mg/kg	3,407	
Available Mg	mg/kg	168.71	
Extractable S	mg/kg	39.25	
Extractable Si	mg/kg	16.12	

#### **Property of Molasses and Vinasse**

The molasses and vinasse used in this study were highly acidic, with pH values of 4.93 and 4.31, respectively, vinasse being the more acidic. This higher acidity in vinasse could be due to the fermentation process. Both substances had high EC values, 26.45 dS/m for molasses and 38.80 dS/m for vinasse. Molasses had higher levels of dissolved and total solids, as well as viscosity, than vinasse. Both were rich in plant nutrients like K, N, Ca, S, Mg, P, and various micronutrients, with molasses generally containing higher concentrations of these elements. No toxic elements, such as As, Cd, and Pb, were detected in either substance (Table [3\)](#page-3-1).

## **Efect of Foliar K Supplementation on Growth Parameters and Yields of Plant Sugarcane**

K is an essential nutrient crucial for the growth, yield, and sugar content of sugarcane crops. This study investigated the effects of foliar supplementation with various K sources, including KCl,  $KNO_3$ ,  $K_2SO_4$ ,  $K_2SiO_3$ , diluted molasses, and diluted vinasse, on sugarcane growth parameters. The results showed that no signifcant diferences were observed among diferent foliar supplementation treatments; however, foliar  $K_2SiO_3(T6)$  supplementation tended to increase cane length (337.49 cm) and diameter (28.61 mm) (Table [4](#page-4-0)). Similarly, foliar supplementation with diluted vinasse (T8) appeared to raise the number of millable cane (86,667 stalk/ha), although no signifcant diferences were observed among the treatments for these growth parameters. In contrast, foliar supplementation with  $K_2SiO_3$  (T6) and  $KNO_3$ (T4) signifcantly enhanced cane yield, producing 155.19



Remark \*measuring at 1:2 H<sub>2</sub>O, and *ND* Not detected

and 154.81 tons/ha, respectively ( $P \le 0.05$ ). These yields were signifcantly greater than those of the foliar water (T2) and control (T1) treatments, which were 132.81 and 130.67 tons/ha, respectively, as shown in Table [4](#page-4-0). Even though the changes in yield attributes were not statistically signifcant, the combined efect of slight improvements across several parameters could collectively impact the total cane yield signifcantly. This aligns with several studies that have shown that yield attributes, regardless of statistical signifcance, can still contribute to diferences in yield, or in some cases, may not affect yield at all (Raposo Junior et al. [2013;](#page-9-20) Desalegn et al. [2023;](#page-9-21) Kumar et al. [2023](#page-9-22)). Nevertheless, among the different forms of foliar K supplementation (T3-8), no signifcant diferences in cane yield were found. Furthermore, no signifcant diferences in sugar yield among the treatments were observed, possibly because the increase in cane yield was not substantial enough to signifcantly boost sugar yield. However, there was a noticeable trend towards increased sugar yield with foliar supplementation of  $KNO_3$ and  $K_2SiO_3$  (19.96 and 19.70 tons/ha, respectively), which correlates with the increase in cane yield (Table [4\)](#page-4-0).

K primarily acts as an enzyme activator in plant metabolism, photosynthesis, starch formation, and the translocation <span id="page-4-0"></span>**Table 4** Efects of foliar K supplementation on length, diameter, number of millable cane, cane yield, and sugar yield of sugarcane



of proteins and sugars in sugarcane (Cui and Tcherkez [2021\)](#page-8-4). For high yields and optimal juice quality, sugarcane requires K in quantities at least equal to or greater than those of N and P (Kwong  $2002$ ). Foliar supplementation with  $K_2SiO_3$  not only enhanced K availability but also supplied additional silicon (Si) to the sugarcane, tending to increased cane length and diameter, and consequently, higher crop weight, as shown in Table [4](#page-4-0). Si played a role in reducing carbon (C) concentration while enhancing N and P absorption, suggesting that Si benefts physiological processes, thereby increasing biomass production (Frazão et al. [2020\)](#page-9-24). Additionally, Si reduced transpiration rates and increased chlorophyll concentration in the leaves (Teixeira et al. [2022](#page-9-7); Costa et al. [2023\)](#page-8-0), potentially infuencing sugarcane yield in this study. The foliar application of K and N, particularly in the form of  $KNO<sub>3</sub>$ , at 120 days after planting supported the sugarcane during its tillering and grand growth phases, facilitating improved plant growth and development. Nutrient scarcity in the soil can hinder plant growth, but foliar nutrient application mitigates soil nutrient fxation and ensures timely nutrient availability to plants. Consistent with Bamrungrai et al. ([2022](#page-8-5)), it was found that combining

soil NPK application with foliar N and K applications during the dry season at 90 days after planting (DAP) and the rainy season at 210 DAP, especially under fooding stress conditions, positively afected yield components, ultimately enhancing sugarcane yield.

# **Efect of Foliar K Supplementation on Quality of Sugarcane Juice**

The impact of foliar K supplementation on sugarcane juice components was examined, revealing no signifcant changes in fiber content (12.80–14.30%), brix (21.11–21.51%), polarity (18.49–19.20%), purity (87.52–89.83%), CCS (13.78–14.53%), and sugar recovery (12.68–13.31%) in sugarcane grown in K-sufficient soils (Table  $5$ ). This observation is consistent with prior studies, indicating that while K fertilizer could increase cane yield, it did not alter the sucrose concentration (polarity) in the cane (Kwong [2002](#page-9-23)). However, concerning sugarcane yield (Table [4](#page-4-0)), foliar supplementation with  $K_2SiO_3$  (T6) and  $KNO_3$  (T4) achieved the highest sugarcane yield. Despite showing a tendency to reduce CCS, the diference was not statistically signifcant

<span id="page-4-1"></span>



(Table [5\)](#page-4-1). Furthermore, a noticeable trend towards improved juice quality was observed with foliar application of diluted molasses (T7) and vinasse (T8) compared to other treatments (Table [5](#page-4-1)). Bamrungrai et al. [\(2022\)](#page-8-5) also noted that the combined application of soil NPK and foliar N and K during both the dry and rainy seasons did not signifcantly afect the sugar quality components. Conversely, Ali et al. [\(1997\)](#page-8-6) reported that split applications of N fertilizer, both solid and foliar, could enhance the polarity and CCS of sugarcane. However, these efects were not observed in our study's N-containing treatments, including  $KNO<sub>3</sub> (T4)$ , diluted molasses (T7), and vinasse (T8).

## **Efect of Foliar K Supplementation on Nutrient Concentration of Sugarcane**

The fndings revealed that nutrient concentrations, including N, P, K, and S, were higher in the tops of the sugarcane than in the cane stalks, with foliar K supplementation significantly affecting these concentrations at  $P \leq 0.05$ (Table [6](#page-5-0)). Foliar application of diluted molasses (T7) signifcantly increased the total K concentration in both the tops and cane stalks, reaching 25.14 and 2.80 g/kg, respectively. In addition, foliar application with  $K_2SO_4$ (T5), diluted vinasse (T8), and  $KNO<sub>3</sub>$  (T4) raised the total K concentration in the cane stalks to 2.70, 2.56, and 2.43 g/kg, respectively, compared to 2.11 g/kg for foliar water (T2) and 1.33 g/kg for the control (T1). Moreover, foliar application of diluted vinasse (T8) signifcantly increased total N and Si concentrations in the cane stalks, reaching 4.99 and 0.34 g/kg, respectively. Likewise, foliar applications with  $K_2SiO_3(T6)$ , diluted molasses (T7), and  $K<sub>2</sub>SO<sub>4</sub>$  (T5) resulted in higher total Si concentrations in the cane stalks compared to 0.25 g/kg for foliar water (T2) and 0.24 g/kg for the control (T1) (Table  $6$ ). Foliar K supplementation with diferent K sources results in sugarcane obtaining other plant nutrients besides K, the availability

of which varies depending on the form of the fertilizer. Foliar applications of molasses and vinasse are advantageous over others due to their rich nutrient content. This is consistent with the fndings of Gaafar et al. ([2019](#page-9-25)), who reported that foliar application of molasses and vinasse increased the concentrations of N, P, and K in fruit of sweet pepper.

Conversely, foliar applications with  $KNO<sub>3</sub> (T4)$  and  $K_2SiO_3$  (T6) significantly reduced total P and N concentrations in the tops of the sugarcane, with values of 1.73 and 6.86 g/kg, respectively. Similarly, the application of diluted molasses and vinasse decreased total P and S concentrations in the cane stalks, with values of 0.53 and 0.96 g/kg, respectively, compared to foliar water (T2) and the control (T1). Additionally, foliar supplementation with  $K_2SiO_3$  (T6) notably reduced the total S concentration in the tops (1.88 g/kg) and the total K concentration in the cane stalks  $(1.89 \text{ g/kg})$ , akin to the effect of diluted molasses (T7), which also signifcantly reduced the total S concentration in the tops (1.90 g/kg). Furthermore, foliar K supplementation did not impact the total Si concentration in the tops of the sugarcane (Table  $6$ ). However, solely considering the concentration of plant nutrients in sugarcane may not be sufficient due to the dilution effect. Therefore, it is advisable to also consider data on the uptake of plant nutrients in sugarcane alongside (Jarrell and Beverly [1981\)](#page-9-26), as discussed in the next section.

# **Efect of Foliar K Supplementation on Nutrient Uptake of Sugarcane**

Nutrient accumulation in sugarcane primarily occurs in the cane stalks, aligning the plant's overall nutrient uptake with that of the stalks. Foliar K supplementation resulted in signifcant variations in nutrient uptake in sugarcane at *P*≤0.05, except for the leaves, as shown in Figs. [1](#page-6-0) and [2.](#page-7-0) Sugarcane responded positively to soil fertilization (T2-8),

<span id="page-5-0"></span>**Table 6** Efects of foliar K supplementation on nutrient concentration in the tops and stalks of sugarcane (dry weight basis)

Treatments	Element concentration in top $(g/kg)$				Element concentration in cane $(g/kg)$					
	Total N	<b>Total P</b>	Total K	Total S	Total Si	Total N	Total P	Total K	Total S	<b>Total Si</b>
T1-Control	8.55ab	1.94a	21.23c	2.07 <sub>bc</sub>	0.27	1.85d	0.74abc	1.33f	1.04 <sub>bcd</sub>	0.24e
T2-BF + Water	7.88abc	1.89a	22.11bc	2.25ab	0.27	3.78 <sub>bc</sub>	0.75abc	2.11d	1.08abc	$0.25$ de
T3-BF+KCl	8.53ab	1.93a	23.09abc	2.16bc	0.27	3.36c	0.65 <sub>bcd</sub>	2.23d	1.07abc	$0.26$ cd
$T4-BF+KNO3$	8.68a	1.73 <sub>b</sub>	22.68abc	1.99bc	0.27	3.77bc	0.81a	2.43c	1.14ab	$0.27$ cd
$T5-BF+K_2SO_4$	7.89abc	1.82ab	24.20ab	2.49a	0.26	3.48c	0.87a	2.70ab	1.16a	0.28c
$T6-BF+K_2SiO_3$	6.86d	1.85ab	23.86abc	1.88c	0.27	4.14b	$0.62$ cd	1.89e	$0.97$ cd	0.31 <sub>b</sub>
T7-BF+Molasses	7.27 cd	1.92a	25.14a	1.90c	0.27	4.28b	0.53d	2.80a	1.11ab	0.31 <sub>b</sub>
T8-BF + Vinasse	7.67bcd	1.86a	22.31bc	2.10 <sub>bc</sub>	0.28	4.99a	0.80ab	2.56bc	0.96d	0.34a
F-test	*	$\ast$	*	∗	ns	*	*	*	*	$\ast$
C.V(%)	9.61	5.61	10.37	11.70	11.88	11.88	16.68	6.73	8.93	0.00

<span id="page-6-0"></span>**Fig. 1** Efects of foliar K supplementation on N (**a**), P (**b**), and K (**c**) uptake in the tops, stalks, and whole plant of sugarcane. The bars on the columns represent mean values  $\pm$  standard deviation. Lowercase letters indicate signifcant diferences at  $P \leq 0.05$  between treatments







showing increased N uptake compared to the unfertilized control (T1) in both the cane stalks and the entire plant, as illustrated in Fig. [1](#page-6-0)a. Foliar application with diluted molasses (T7) led to the highest uptakes of N, K, S, and Si in the cane stalks, at 322, 215, 80.9, and 23.2 kg/ha, respectively, and in the entire plant, at 335, 259, 84.3, and 23.6 kg/ha, respectively, which were signifcantly higher than those from foliar water (T2) and the control (T1) (Figs. [1](#page-6-0) and [2\)](#page-7-0). Diluted vinasse application also enhanced N and Si uptake in both the cane stalks (287 and 20.0 kg/ha, respectively) and the entire plant (300 and 20.6 kg/ha, respectively), as indicated in Figs. [1](#page-6-0)a and [2](#page-7-0)b. Foliar application with  $K_2SiO_3$  (T6) increased Si uptake in the cane stalks (18.1 kg/ha) and the entire plant (18.6 kg/ha) (Fig. [2](#page-7-0)b). Moreover, foliar application with diluted vinasse (T8) raised K uptake in the entire plant to 198 kg/ha (Fig. [1](#page-6-0)c).





<span id="page-7-0"></span>**Fig. 2** Efects of foliar K supplementation on S (**a**) and Si (**b**) uptake in the tops, stalks, and whole plant of sugarcane. The bars on the columns represent±standard deviation. Lowercase letters indicate signifcant diferences at *P*≤0.05 between treatments

Molasses, rich in plant nutrients like  $N$  and  $K$  and containing S and Si (Table [3](#page-3-1)), facilitated the accumulation of these elements in the sugarcane (Figs. [1](#page-6-0) and [2](#page-7-0)). N promotes growth and canopy expansion, improving light absorption efficiency, essential for increased cane stalk production (Milford et al. [2000](#page-9-27); Shah et al. [2024](#page-9-28)), as observed with foliar supplementation with diluted vinasse (T8), molasses (T7), and  $KNO<sub>3</sub>$  (T4), all containing N. K, acting as an osmotic solute, is vital for sustaining cell turgor that drives N-stimulated growth, highlighting the need for adequate K to support the utilization of unassimilated N in the cane, thus aiding the transition of reducing sugars into sucrose during maturity (Kwong [2002\)](#page-9-23). Despite its lower K content,  $K_2SiO_3$  led to the highest yield (Tables [1](#page-2-0) and [4](#page-4-0)), correlating with the lower total K concentration in the cane stalks (Table  $6$ ), possibly due to a dilution effect. However, foliar K supplementation did not infuence P uptake in the sugarcane in this study (Fig. [1](#page-6-0)b), even though P was indirectly supplied through foliar application with diluted molasses and vinasse. This absence of effect could be due to the high P availability in the soil (Table [2\)](#page-3-0) and soil fertilization.

This study, conducted on sugarcane grown in soil with sufficient K levels, revealed that foliar K supplementation, particularly  $K_2SiO_3$  and  $KNO_3$ , significantly influenced cane yield and internal nutrient dynamics, including nutrient concentration and uptake, and exhibited a trend towards enhancing specifc growth parameters. Other factors, such as environmental conditions (temperature, moisture, soil drainage), may contribute to the lack of statistically signifcant changes in growth parameters (Zhao and Li [2015\)](#page-10-5).

## **Conclusion**

Foliar K supplementation with 2.5% w/v of  $K_2SiO_3$  and  $KNO<sub>3</sub>$  at 120 days post-planting significantly increased cane yields, surpassing those from foliar water and the unfertilized control. Despite the lack of statistical signifcance, foliar K supplementation has shown a trend towards enhancing certain yield attributes. The presence of Si from  $K_2SiO_3$  likely bolstered physiological processes, leading to increased biomass production. Moreover, the supplementation of K and N, especially through  $KNO_3$ , proved advantageous during critical growth stages, thereby fostering overall plant growth and development. Despite the observed yield improvements, it's worth noting that foliar K supplementation did not exert signifcant impacts on sugarcane juice quality. Furthermore, our study revealed substantial variations in nutrient concentration and uptake in plant sugarcane due to foliar K supplementation, with diluted molasses leading to the highest uptakes of N, K, S, and Si in cane stalks. Molasses, being rich in N and K as well as S and Si, promoted the accumulation of these nutrients in sugarcane. This underscores the potential of foliar application in preventing soil nutrient fxation and ensuring timely nutrient availability, enhancing the viability of this approach. In conclusion, our study highlights the efectiveness of foliar K supplementation in increasing cane yield and nutrient uptake in sugarcane. Further research into foliar K supplementation, particularly in ratoon sugarcane, focusing on concentration, rate, and duration, is essential to optimize sugarcane yield under various soil fertility conditions.

**Acknowledgements** This research is funded by the Graduate School Fellowship Program in agriculture and agro-industry from the Agricultural Research Development Agency (Public Organization) as of fscal year 2022. The authors are grateful to MITR PHOL GROUP for molasses and vinasse used in this study.

**Author contributions** All authors contributed to the study conception and design. Material preparation, and data collection were performed by Nattawat Radasai, Daojarus Ketrot, and Tawatchai Inboonchuay. Sample and data analysis were performed by Nattawat Radasai, Daojarus Ketrot, Saowanuch Tawornpruek and Acharaporn Wongsuksri. The frst draft of the manuscript was written by Nattawat Radasai, and Daojarus Ketrot. All authors commented on previous versions of the manuscript and read and approved the fnal manuscript.

#### **Declarations**

**Conflict of interest** The authors declare that they have no competing interests.

#### **References**

- <span id="page-8-6"></span>Ali, S.A., M.M.R.K. Afridi, and R.G. Singh. 1997. Comparative efficiency of soil and foliar applied nitrogen in sugarcane, quality parameters and leaf nitrogen content. *Indian Journal of Plant Physiology* 2 (1): 75–78.
- <span id="page-8-5"></span>Bamrungrai, J., A. Polthanee, B. Tubana, V. Tre-loges, and A. Promkhambut. 2022. Efects of soil and foliar applied fertilizers on growth, yield and sugar quality of two sugarcane cultivars under rainfed conditions. *Indian Journal of Agricultural Research.* 56 (5): 545–550. [https://doi.org/10.18805/IJARe.AF-688.](https://doi.org/10.18805/IJARe.AF-688)
- <span id="page-8-3"></span>Chen, M., and L.Q. Ma. 2001. Comparison of three aqua regia digestion methods for twenty Florida soils. *Soil Science Society of America Journal* 65 (2): 491–499.
- <span id="page-8-1"></span>Christofoletti, C.A.J.P., J.E. Escher, J.F.U. Marinho. Correia, and C.S. Fontanetti. 2013. Sugarcane vinasse: Environmental implications of its use. *Waste Management* 33 (12): 2752–2761. [https://doi.org/](https://doi.org/10.1016/j.wasman.2013.09.005) [10.1016/j.wasman.2013.09.005](https://doi.org/10.1016/j.wasman.2013.09.005).
- <span id="page-8-0"></span>Costa, M.G., M.R. de Prado, M.M. Sarah, L.F. Palaretti, C.M. de Piccolo, and J.P. Souza Júnior. 2023. New approaches to the efects of Si on sugarcane ratoon under irrigation in quartzipsamments, eutrophic red oxisol, and dystrophic red oxisol. *BMC Plant Biology* 23: 51. [https://doi.org/10.1186/s12870-023-04077-2.](https://doi.org/10.1186/s12870-023-04077-2)
- <span id="page-8-2"></span>Crusciol, C.A.C., D.P. de Arruda, A.M. Fernandes, J.A. Antonangelo, L.R.F. Alleoni, C.A.C. do Nascimento, O.B. Rossato, and J.M. McCray. 2018. Methods and extractants to evaluate silicon availability for sugarcane. *Scientifc Reports* 8: 916. [https://doi.org/10.](https://doi.org/10.1038/s41598-018-19240-1) [1038/s41598-018-19240-1.](https://doi.org/10.1038/s41598-018-19240-1)
- <span id="page-8-4"></span>Cui, J., and G. Tcherkez. 2021. Potassium dependency of enzymes in plant primary metabolism. *Plant Physiology and Biochemistry* 166: 522–530. <https://doi.org/10.1016/j.plaphy.2021.06.017>.
- <span id="page-9-0"></span>de Souza Barbosa, G.V., J.M. dos Santos, C.A. Diniz, D.E. Cursi, and H.P. Hofmann. 2020. Energy cane breeding. In *Sugarcane biorefnery, technology and perspectives*, ed. F. In Santos, S.C. Rabelo, M.D. Matos, and P. Eichler. Amsterdam: Academic Press Elsevier.
- <span id="page-9-14"></span>Department of Agriculture. 2021. The recommendation for fertilizer application based on soil analysis for economic crops. Department of Agriculture, Ministry of Agriculture and Cooperatives, Thailand. 102
- <span id="page-9-21"></span>Desalegn, B., E. Kebede, H. Legesse, and T. Fite. 2023. Sugarcane productivity and sugar yield improvement: Selecting variety, nitrogen fertilizer rate, and bioregulator as a frst-line treatment. *Heliyon* 9: e15520. [https://doi.org/10.1016/j.heliyon.2023.e15520.](https://doi.org/10.1016/j.heliyon.2023.e15520)
- <span id="page-9-19"></span>Divakar, S., R.K. Jha, D.N. Kamat, and A. Singh. 2023. Validation of candidate genebased EST-SSR markers for sugar yield in sugarcane. *Frontiers in Plant Science* 14: 273740. [https://doi.org/10.](https://doi.org/10.3389/fpls.2023.1273740) [3389/fpls.2023.1273740](https://doi.org/10.3389/fpls.2023.1273740).
- <span id="page-9-1"></span>FAO. 2022. World Food and Agriculture: Statistical Yearbook 2022. Food and agriculture organization (FAO) of the United Nations, Rome, Italy. [https://doi.org/10.4060/cc2211en.](https://doi.org/10.4060/cc2211en) Accessed 29 February 2024.
- <span id="page-9-2"></span>FAOSTAT. 2024. Statistical database. Food and agriculture organization (FAO) of the United Nations, Rome. Italy. Available at [https://](https://www.fao.org/faostat/en/#data/QCL) [www.fao.org/faostat/en/#data/QCL.](https://www.fao.org/faostat/en/#data/QCL) Accessed 1 March 2024.
- <span id="page-9-12"></span>Fox, R.L., R.A. Olson, and H.F. Rhoades. 1964. Evaluating the sulfur status of soils by plant and soil tests. *Soil Science Society of America Journal* 28: 243–246. [https://doi.org/10.2136/sssaj1964.](https://doi.org/10.2136/sssaj1964.03615995002800020034x) [03615995002800020034x](https://doi.org/10.2136/sssaj1964.03615995002800020034x).
- <span id="page-9-24"></span>Frazão, J.J., R. de Mello Prado, J.P. de Souza Júnior, and D.R. Rossatto. 2020. Silicon changes C:N: P stoichiometry of sugarcane and its consequences for photosynthesis, biomass partitioning and plant growth. *Scientifc Reports* 10: 12492. [https://doi.org/10.1038/](https://doi.org/10.1038/s41598-020-69310-6) [s41598-020-69310-6](https://doi.org/10.1038/s41598-020-69310-6).
- <span id="page-9-25"></span>Gaafar, M.S., N.M.M. EL-Shimi, and M.M. Helmy. 2019. Efect of foliar and soil application of some residuals of sugar cane products (molasses and vinasses) with mineral fertilizer levels on growth, yield and quality of sweet pepper. *Menoufa Journal of Plant Production* 4 (5): 353–373. [https://doi.org/10.21608/mjppf.](https://doi.org/10.21608/mjppf.2019.174948) [2019.174948](https://doi.org/10.21608/mjppf.2019.174948).
- <span id="page-9-6"></span>Hamid, A.M., and Y.M. Dagash. 2014. Effect of sulfur on sugarcane yield and quality at the heavyclay soil "Vertisols" of Sudan. *Universal Journal of Applied Science* 2 (3): 68–71. [https://doi.org/10.](https://doi.org/10.13189/ujas.2014.020303) [13189/ujas.2014.020303](https://doi.org/10.13189/ujas.2014.020303).
- <span id="page-9-3"></span>Jaiswal, V.P., S.K. Shukla, L. Sharma, I. Singh, A.D. Pathak, M. Nagargade, A. Ghosh, C. Gupta, A. Gaur, S.K. Awasthi, R. Tiwari, A. Srivastava, and E. Masto. 2021. Potassium infuencing physiological parameters, photosynthesis and sugarcane yield in subtropical India. *Sugar Tech* 23 (2): 343–359. [https://doi.org/10.1007/](https://doi.org/10.1007/s12355-020-00905-z) [s12355-020-00905-z.](https://doi.org/10.1007/s12355-020-00905-z)
- <span id="page-9-26"></span>Jarrell, W.M., and R.B. Beverly. 1981. The dilution efect in plant nutrition studies. *Advances in Agronomy* 34: 197–224. [https://doi.org/](https://doi.org/10.1016/S0065-2113(08)60887-1) [10.1016/S0065-2113\(08\)60887-1](https://doi.org/10.1016/S0065-2113(08)60887-1).
- <span id="page-9-9"></span>Jiang, Z.P., Y.R. Li, G.P. Wei, Q. Liao, T.M. Su, Y.C. Meng, H.Y. Zhang, and C.Y. Lu. 2012. Efect of long-term vinasse application on physico-chemical properties of sugarcane feld soils. *Sugar Tech* 14 (4): 412–417. [https://doi.org/10.1007/s12355-012-0174-9.](https://doi.org/10.1007/s12355-012-0174-9)
- <span id="page-9-4"></span>Johnson, R., K. Vishwakarma, Md.S. Hossen, V. Kumar, A.M. Shackira, J.T. Puthur, G. Abdi, M. Sarraf, and M. Hasanuzzaman. 2022. Potassium in plants: Growth regulation, signaling, and environmental stress tolerance. *Plant Physiology and Biochemistry* 172: 56–69. [https://doi.org/10.1016/j.plaphy.2022.](https://doi.org/10.1016/j.plaphy.2022.01.001) [01.001](https://doi.org/10.1016/j.plaphy.2022.01.001).
- <span id="page-9-16"></span>Kirasak, K., T. Sansayawichai, W. Ponragdee, and A. Thipyawat. 2015. Determination of commercial cane sugar (CCS) using near infrared spectroscopy. *Thai Agricultural Research Journal* 33 (2): 159–168.<https://doi.org/10.14456/thaidoa-agres.2015.8>.
- <span id="page-9-17"></span>Kulasekara, B.R., B.D.S.K. Ariyawansha, H.A.S. Weerasinghe, K.A.D. Kodithuwakku, and U.W.L.M. Kumarasiri. 2024. Response of soil, cane yield and cane quality parameters to zinc fertiliser under alfsols condition in Sri Lanka: A preliminary investigation. *Sugar Tech.* 26 (3): 915–919. [https://doi.org/10.1007/](https://doi.org/10.1007/s12355-023-01353-1) [s12355-023-01353-1](https://doi.org/10.1007/s12355-023-01353-1).
- <span id="page-9-22"></span>Kumar, N., L. Rana, A.K. Singh, B. Pramanick, A. Gaber, A.M. Alsuhaibani, M. Skalicky, and A. Hossain. 2023. Precise macronutrient application can improve cane yield and nutrient uptake in widely spaced plant-ratoon cycles in the Indo-Gangetic plains of India. *Frontiers in Sustainable Food Systems* 7: 1223881. [https://doi.org/](https://doi.org/10.3389/fsufs.2023.1223881) [10.3389/fsufs.2023.1223881](https://doi.org/10.3389/fsufs.2023.1223881).
- <span id="page-9-23"></span>Kwong, K.F.N.K. 2002. The effects of potassium on growth, development, yield and quality of sugarcane. In *Potassium for sustainable production. Proceedings of the International Symposium on role of potassium in nutrient management for sustainable crop production in India*, 3–5 December 2001, New Delhi, eds. Pasricha, N.S., and S.K. Bansal, 430–444. Basel, Switzerland: International Potash Institute.
- <span id="page-9-18"></span>Mehareb, E.M., and S.R. Abazied. 2017. Genetic variability of some promising sugarcane varieties (*Saccharum spp*) under harvesting ages for juice quality traits, cane and sugar yield. *Open Access Journal of Agricultural Research* 2 (2): 1–14. [https://doi.org/10.](https://doi.org/10.23880/oajar-16000127) [23880/oajar-16000127](https://doi.org/10.23880/oajar-16000127).
- <span id="page-9-5"></span>Mikkelsen, R.L., and T.L. Roberts. 2021. Inputs: potassium sources for agricultural systems. In *Improving potassium recommendations for agricultural crops*, ed. T.S. Murrell, R.L. Mikkelsen, G. Sulewski, R. Norton, and M.L. Thompson. Cham: Springer.
- <span id="page-9-27"></span>Milford, G.F.J., M.J. Armstrong, P.J. Jarvis, B.J. Houghton, D.M. Bellett-Travers, J. Jones, and R.A. Leigh. 2000. Efects of potassium fertilizer on the yield, quality and potassium oftake of sugar beet crops grown on soils of diferent potassium status. *Journal of Agricultural Science* 135: 1–10.
- <span id="page-9-8"></span>Palmonari, A., D. Cavallini, C.J. Snifen, L. Fernandes, P. Holder, L. Fagioli, I. Fusaro, G. Biagi, A. Formigoni, and L. Mammi. 2020. Short communication: Characterization of molasses chemical composition. *Journal of Dairy Science* 103 (7): 6244–6249. <https://doi.org/10.3168/jds.2019-17644>.
- <span id="page-9-13"></span>Ponragdee, W., T. Sansayawichi, P. Sarawat, T. Moulanon, P. Kapetch, and U. Leabwon. 2011. Khon Kaen 3: A sugarcane variety for the northeast. *Thai Agricultural Research Journal* 29 (3): 283–301.
- <span id="page-9-20"></span>Raposo Junior, J.L., J.A.G. Neto, and L.V.S. Sacramento. 2013. Evaluation of diferent foliar fertilizers on the crop production of sugarcane. *Journal of Plant Nutrition* 36 (3): 459–469. [https://doi.org/](https://doi.org/10.1080/01904167.2012.748066) [10.1080/01904167.2012.748066](https://doi.org/10.1080/01904167.2012.748066).
- <span id="page-9-28"></span>Shah, I.H., W. Jinhui, X. Li, M.K. Hameed, M.A. Manzoor, P. Li, Y. Zhang, Q. Niu, and L. Chang. 2024. Exploring the role of nitrogen and potassium in photosynthesis implications for sugar: Accumulation and translocation in horticultural crops. *Scientia Horticulturae* 327: 112832.<https://doi.org/10.1016/j.scienta.2023.112832>.
- <span id="page-9-11"></span>Soil Survey Staff. 2014. Kellogg soil Survey laboratory methods manual. Soil survey investigations report No.42, Version 5.0. Burt, R. and Soil Survey Staf, eds. U.S. Department of Agriculture, Natural resources conservation service.
- <span id="page-9-15"></span>Som-ard, J., C. Atzberger, E. Izquierdo-Verdiguier, F. Vuolo, and M. Immitzer. 2021. Remote sensing applications in sugarcane cultivation: A review. *Remote Sensing* 13: 4040. [https://doi.org/10.](https://doi.org/10.3390/rs13204040) [3390/rs13204040](https://doi.org/10.3390/rs13204040).
- <span id="page-9-10"></span>Suwanarit, A., and M. Sestapukdee. 1989. Stimulating effects of foliar K-fertilizer applied at the appropriate stage of development of maize: A new way to increase yield and improve quality. *Plant and Soil* 120: 111–124.
- <span id="page-9-7"></span>Teixeira, G.C.M., R.M. de Prado, A.M.S. Rocha, A.S.B. de Oliveira Filho, G.S. da Sousa Junior, and P.L. Gratão. 2022. Action of silicon on the activity of antioxidant enzymes and on physiological mechanisms mitigates water deficit in sugarcane and energy

cane plants. *Scientifc Reports* 12: 1–18. [https://doi.org/10.1038/](https://doi.org/10.1038/s41598-022-21680-9) [s41598-022-21680-9](https://doi.org/10.1038/s41598-022-21680-9).

- <span id="page-10-0"></span>Waclawovsky, A.J., P.M. Sato, C.G. Lembke, P.H. Moore, and G.M. Souza. 2010. Sugarcane for bioenergy production: An assessment of yield and regulation of sucrose content. *Plant Biotechnology Journal* 8: 263–276. [https://doi.org/10.1111/j.1467-7652.2009.](https://doi.org/10.1111/j.1467-7652.2009.00491.x) [00491.x.](https://doi.org/10.1111/j.1467-7652.2009.00491.x)
- <span id="page-10-2"></span>Watanabe, K., J. Tominaga, S. Yabuta, H. Takaragawa, R. Suwa, M. Ueno, and Y. Kawamitsu. 2017. Efects of diferent kinds of potassium and chloride salts on sugarcane quality and photosynthesis. *Sugar Tech* 19 (4): 378–385. [https://doi.org/10.1007/](https://doi.org/10.1007/s12355-016-0486-2) [s12355-016-0486-2.](https://doi.org/10.1007/s12355-016-0486-2)
- <span id="page-10-1"></span>Watnabe, K., M. Nakabaru, E. Taira, M. Ueno, and Y. Kawamitsu. 2016. Relationships between nutrient and sucrose concentration in sugarcane juice and use of juice analysis for nutrient diagnosis in Japan. *Plant Production Science* 19 (2): 215–222. [https://doi.](https://doi.org/10.1080/1343943X.2015.1128106) [org/10.1080/1343943X.2015.1128106.](https://doi.org/10.1080/1343943X.2015.1128106)
- <span id="page-10-4"></span>Wu, Q., W. Zhou, Y. Lu, S. Li, D. Shen, Q. Ling, D. Chen, and J. Ao. 2023. Combined chemical fertilizers with molasses increase soil stable organic phosphorus mineralization in sugarcane seedling

stage. *Sugar Tech* 25 (3): 552–561. [https://doi.org/10.1007/](https://doi.org/10.1007/s12355-022-01196-2) [s12355-022-01196-2](https://doi.org/10.1007/s12355-022-01196-2).

- <span id="page-10-3"></span>Zeng, X.-P., K. Zhu, J.-M. Lu, Y. Jiang, L.-T. Yang, Y.-X. Xing, and Y.-R. Li. 2020. Long-term efects of diferent nitrogen levels on growth, yield, and quality in sugarcane. *Agronomy* 10: 353. [https://](https://doi.org/10.3390/agronomy10030353) [doi.org/10.3390/agronomy10030353](https://doi.org/10.3390/agronomy10030353).
- <span id="page-10-5"></span>Zhao, D., and Y. Li. 2015. Climate change and sugarcane production: Potential impact and mitigation strategies. *International Journal of Agronomy* 2015: 547386.<https://doi.org/10.1155/2015/547386>.

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.