

Imaging atherosclerotic burden and inflammation: Insights into the spectrum of atherosclerotic disease in HIV

Ahmed Tawakol, MD,^a and Steven K. Grinspoon, MD^b

^a Cardiology Division, Department of Medicine, Massachusetts General Hospital and Harvard Medical School, Boston, MA

^b Program in Nutritional Metabolism, Department of Medicine, Massachusetts General Hospital and Harvard Medical School, Boston, MA

Received Nov 25, 2014; accepted Nov 25, 2014
doi:10.1007/s12350-014-0044-9

See related article, pp. 372–380

HIV disease, left inadequately treated, leads to an inexorable decline in immune function resulting in AIDS and ultimately in death from opportunistic infections. However, advances in treatment of HIV have resulted in a dramatic reduction in AIDS-related mortality.¹ With the widespread use of highly active antiretroviral therapy (HAART), HIV infection has been transformed to a chronic disease associated with long-term viral suppression but at the cost of an increase in atherothrombotic diseases.^{2,3} Hence currently there is substantial interest in developing a better understanding of the mechanisms underlying the increased risk of atherothrombosis in HIV disease.

While HIV infection is characterized by a decline of peripheral blood CD4+ T cells, a paradoxical chronic immune activation of T cells and monocytes is routinely seen in virally suppressed HIV disease.⁴⁻⁶ Multiple studies in HIV-infected individuals show that this persistently heightened state of immune activation likely contributes to the increased incidence of atherosclerosis,^{2,7,8} itself a chronic inflammatory condition. Numerous potential causes may contribute to this chronic immune activation, including increased microbial translocation from the gut due to CD4+ T

lymphocytes depletion,⁹ persistent low-level viral replication,¹⁰ increased levels of lipopolysaccharide (LPS), sCD14,¹¹ HIV gene products, and CMV-specific T-cell activation¹² among other mechanisms. In this regard, an increasing number of studies have begun to assess the relationship between atherosclerosis and immune activation in HIV-infected individuals.

Recent studies by our group and others have shown substantial increases in subclinical coronary plaque.^{13,14} Using CT angiography, we have observed increases in non-calcified plaque volume¹⁵ and vulnerable plaque features in association with sCD163 an immune activation marker, indicative of monocyte activation.¹⁶ In a large study of over 700 HIV-infected patients and control subjects in the MACS cohort patients, Post et al similarly demonstrated increased non-calcified plaque.¹⁴ In a follow-up analysis, sCD163 was shown to increase in association with overall plaque indices, again using CTA.¹⁷ These studies support the notion, across a broad range of patients with HIV infection, (including elite controllers who have well controlled viremia without ART),¹⁸ that immune activation is increased in HIV, and is associated with increased coronary atherosclerotic indices.

Additionally, numerous studies have shown that HIV infection is associated with an increase in extra-coronary atherosclerosis, such as can be demonstrated by measurement of carotid intima-media thickness (cIMT). Hsue et al demonstrated that cIMT is greater in HIV patients than in age-matched control subjects and progresses much more rapidly than in individuals without HIV infection.¹⁹ The severity of carotid atherosclerosis seen in treated HIV infection (as IMT) correlates with markers of immune activation.²⁰ Others have shown that the rate of progression of cIMT in HIV is predicted by markers of immune activation and

Reprint requests: Ahmed Tawakol, MD, Cardiology Division, Department of Medicine, Massachusetts General Hospital and Harvard Medical School, Boston, MA; atawakol@partners.org

J Nucl Cardiol 2015;22:381–4.
1071-3581/\$34.00

Copyright © 2014 American Society of Nuclear Cardiology.

microbial translocation, respectively.⁸ Moreover, the abnormally advanced atherosclerotic disease is manifest even in children with HIV compared to age-matched controls.²¹ A link between HIV disease, premature and progressive atherosclerosis, and immune activation is becoming increasingly well established.

There is, not surprisingly, substantial interest in studying atherosclerotic inflammation in the context of HIV. Imaging of arterial inflammation is now routinely performed using fluorodeoxyglucose positron emission tomography (FDG-PET). FDG-PET imaging has been used successfully in clinical oncology and for detection of occult infection. Tissue accumulation of FDG is reflective of the rate of tissue glycolysis, which is higher in areas containing metabolically active tumor cells and in inflamed tissues. A large body of basic cellular physiology data demonstrates that activated macrophages have an unusually high metabolic rate^{22,23} especially after classical/innate activation²⁴ hence avidly accumulate FDG.²⁵ While one pre-clinical study suggested that hypoxia is a more substantial stimulus of macrophage glycolysis than inflammatory stimuli,²⁶ that study stands in opposition to the much broader literature; further, hypoxia itself is a potent stimulus of macrophage pro-inflammatory activation.²⁷ Moreover, FDG accumulation within the artery wall has repeatedly been found to correlate with atherosclerotic plaque macrophage density of.²⁸⁻³⁰ Further, several studies have demonstrated that increased plaque inflammation (measured using FDG-PET) predicts an increased rate of atherosclerotic plaque progression.^{31,32} In addition, higher arterial FDG uptake is associated with a substantially increased risk for *subsequent* stroke and MI.³³⁻³⁵ Taken together, these studies provide justification for using arterial FDG-PET/CT imaging to provide a meaningful measure of arterial inflammation in human clinical studies.

More recently, Yarasheski et al and our group separately employed FDG-PET to assess arterial inflammation in HIV.^{7,36} These two studies demonstrated increased arterial FDG-PET uptake in HIV-infected individuals, both in the carotid and descending aorta. Furthermore, we observed that the degree of arterial inflammation in HIV correlates with markers of monocyte activation,⁷ and in a separate study, we observed that extra-coronary arterial inflammation correlates with the presence of high-risk coronary plaques.³⁷ Hsue et al recently extended those findings, and observed that the tissue measures on FDG accumulation relate to markers of viral persistence in HIV.³⁸ Hence a substantial literature has emerged in HIV-positive individuals, which links immune activation to arterial inflammation.

It is in the context of these prior studies that new data from Knudsen et al, in this issue of *Journal of Nuclear Cardiology*, should be considered. Knudsen

et al performed FDG-PET in HIV-infected patients and controls and found no significant differences in arterial inflammation between the HIV-positive individuals and the HIV-negative control subjects (who were matched for some, but not all major CVD risk factors). At first blush, these data seem to contrast with those of Yarasheski et al and Subramanian et al. However, a closer look at the data reveals some important differences in study design and potentially of the patient population that may have contributed to the results and which may provide important insights.

First, the Knudsen et al study showed no evidence of increased subclinical atherosclerosis (by cIMT), in the HIV-positive cohort compared to the HIV-negative controls. This finding runs counter to the findings from a sizeable number of studies, some of which suggest that HIV is associated with a premature aging phenotype, with advanced subclinical atherosclerosis by on average 15 years.³⁹ Of further note, the HIV-positive group in the Knudsen study did not manifest an increase in sCD163 or other immune activation markers, which also runs counter to a large body of data.^{15,17}

It follows from these observations, that the HIV-positive population that was studied by Knudsen et al was somewhat atypical in that the group lacked evidence of either increased immune activation or pre-clinical atherosclerosis. Accordingly, it comes with less surprise that this particular population had no significant increase in arterial inflammation either. Indeed, in the study by Subramanian et al, higher levels of arterial inflammation were seen primarily in individuals with higher levels of immune activation (i.e., sSD138 levels). Thus perhaps the findings of the current study are appropriate for the specific population studied. Perhaps the lesson of the Knudsen study, viewed in the context of existing studies, is that among HIV-positive individuals without pre-clinical atherosclerotic disease or heightened immune activation, there is no increase in arterial inflammation. However, among HIV patients with subclinical atherosclerotic disease and increased immune activation (a group which represents a large proportion of the HIV population), arterial inflammation is increased.

The findings of Knudsen et al highlight the need for further study. Their study, taken together with the previous studies of arterial inflammation in HIV, suggests that a spectrum of arterial inflammation exists in HIV: from higher-to-lower arterial inflammation in individuals with or without immune activation/ pre-clinical atherosclerosis, respectively. Accordingly, a question of proximal importance is whether the spectrum of immune activation and atherosclerotic inflammation in HIV-positive individuals provides any disease insights. For example, can imaging measures of arterial inflammation predict the risk of atherothrombosis in an HIV-positive

population, (much as it can be used in non-infected populations). Furthermore, it remains to be seen whether measures of arterial inflammation could be used to study novel therapies in HIV-positive populations. Thus far, several multicenter randomized controlled trials have utilized serial FDG-PET imaging to evaluate the effect of drugs on atherosclerotic inflammation (in non-infected populations). Data from those studies suggest that treatment-related changes in arterial inflammation seem to be predictive of clinical efficacy. Accordingly, imaging treatment effects using arterial FDG-PET/CT imaging might provide needed insights regarding potential therapeutic efficacy of new therapies. With that stated, studies are now underway to evaluate whether anti-inflammatory strategies (such as low-dose methotrexate) can reduce peripheral markers of inflammation and arterial inflammation in HIV. Concurrently, other atherosclerosis imaging tools such as coronary CT angiography are also being used to study the impact of statin therapy on atherosclerosis progression in HIV. Directionally positive findings in those imaging surrogate studies could justify larger clinical outcome trials to define the therapeutic efficacy of those treatments.

The study of Knudsen et al provides an additional patch to a quilt of data emerging on atherosclerosis in HIV, and suggests that at least some individuals with HIV might have low levels of immune activation, have little evidence of pre-clinical atherosclerosis, or atherosclerotic inflammation, and might thus be spared of the advanced atherosclerotic disease that is otherwise reported in the majority of HIV-infected populations studied. More studies are needed to better understand the mechanisms underlying the increased risk of atherothrombosis in HIV. Ultimately, studies evaluating atherosclerosis in HIV might pave the way toward a new era, where infected individuals can be spared of both AIDS as well as the atherothrombotic complications associated with HIV infection.

Disclosures

Dr. Grinspoon has consulted with AstraZeneca, Navidea, NovoNordisk, and Theratechnologies, and received grant support from Gilead, Amgen, and Theratechnologies, unrelated to this manuscript. Dr Tawakol has consulted with Actelion, Amgen, Cerenis, Genetech, and Takeda and received grant support from Genetech and Takeda, unrelated to this manuscript.

References

1. May MT, Sterne JA, Costagliola D, Sabin CA, Phillips AN, Justice AC, et al. HIV treatment response and prognosis in Europe and

- North America in the first decade of highly active antiretroviral therapy: A collaborative analysis. *Lancet* 2006;368:451-8.
2. Triant VA, Lee H, Hadigan C, Grinspoon SK. Increased acute myocardial infarction rates and cardiovascular risk factors among patients with human immunodeficiency virus disease. *J Clin Endocrinol Metab* 2007;92:2506-12.
3. Freiberg MS, Chang CC, Kuller LH, Skanderson M, Lowy E, Kraemer KL, et al. HIV infection and the risk of acute myocardial infarction. *JAMA Intern Med* 2013;173:614-22.
4. Deeks SG, Walker BD. The immune response to AIDS virus infection: Good, bad, or both? *J Clin Invest* 2004;113:808-10. doi: [10.1172/JCI200421318](https://doi.org/10.1172/JCI200421318).
5. Melnick JL, Adam E, DeBaakey ME. Possible role of cytomegalovirus in atherogenesis. *JAMA* 1990;263:2204-7.
6. Hsue PY, Hunt PW, Sinclair E, Bredt B, Franklin A, Killian M, et al. Increased carotid intima-media thickness in HIV patients is associated with increased cytomegalovirus-specific T-cell responses. *AIDS* 2006;20:2275-83.
7. Subramanian S, Tawakol A, Burdo TH, Abbara S, Wei J, Vijayakumar J, et al. Arterial inflammation in patients with HIV. *JAMA* 2012;308:379-86.
8. Kelesidis T, Kendall MA, Yang OO, Hodis HN, Currier JS. Biomarkers of microbial translocation and macrophage activation: Association with progression of subclinical atherosclerosis in HIV-1 infection. *J Infect Dis* 2012;206:1558-67.
9. Estes JD, Harris LD, Klatt NR, Tabb B, Pittaluga S, Paiardini M, et al. Damaged intestinal epithelial integrity linked to microbial translocation in pathogenic simian immunodeficiency virus infections. *PLoS Pathog* 2010;6:e1001052.
10. Yukl SA, Shergill AK, McQuaid K, Gianella S, Lampiris H, Hare CB, et al. Effect of raltegravir-containing intensification on HIV burden and T-cell activation in multiple gut sites of HIV-positive adults on suppressive antiretroviral therapy. *AIDS* 2010;24:2451-60.
11. Marchetti G, Cozzi-Lepri A, Merlini E, Bellistri GM, Castagna A, Galli M, et al. Microbial translocation predicts disease progression of HIV-infected antiretroviral-naive patients with high CD4+ cell count. *AIDS* 2011;25:1385-94.
12. Sacre K, Hunt PW, Hsue PY, Maidji E, Martin JN, Deeks SG, et al. A role for cytomegalovirus-specific CD4+CX3CR1+ T cells and cytomegalovirus-induced T-cell immunopathology in HIV-associated atherosclerosis. *AIDS* 2012;26:805-14.
13. Lo J, Abbara S, Shturman L, Soni A, Wei J, Rocha-Filho JA, et al. Increased prevalence of subclinical coronary atherosclerosis detected by coronary computed tomography angiography in HIV-infected men. *AIDS* 2010;24:243-53.
14. Post WS, Budoff M, Kingsley L, Palella FJ Jr, Witt MD, Li X, et al. Associations between HIV infection and subclinical coronary atherosclerosis. *Ann Intern Med* 2014;160:458-67.
15. Burdo TH, Lo J, Abbara S, Wei J, DeLelys ME, Preffer F, et al. Soluble CD163, a novel marker of activated macrophages, is elevated and associated with noncalcified coronary plaque in HIV-infected patients. *J Infect Dis* 2011;204:1227-36.
16. Zanni MV, Abbara S, Lo J, Wai B, Hark D, Marmarelis E, et al. Increased coronary atherosclerotic plaque vulnerability by coronary computed tomography angiography in HIV-infected men. *AIDS* 2013;27:1263-72.
17. McKibben RA, Margolick JB, Grinspoon S, Li X, Palella FJ, Jr, Kingsley LA, et al. Elevated levels of monocyte activation markers are associated with subclinical atherosclerosis in men with and those without HIV infection. *J Infect Dis* 2014. doi: [10.1093/infdis/jiu594](https://doi.org/10.1093/infdis/jiu594).
18. Pereyra F, Lo J, Triant VA, Wei J, Buzon MJ, Fitch KV, et al. Increased coronary atherosclerosis and immune activation in HIV-1 elite controllers. *AIDS* 2012;26:2409-12.

19. Hsue PY, Lo JC, Franklin A, Bolger AF, Martin JN, Deeks SG, et al. Progression of atherosclerosis as assessed by carotid intima-media thickness in patients with HIV infection. *Circulation* 2004;109:1603-8.
20. Alonso-Villaverde C, Coll B, Parra S, Montero M, Calvo N, Tous M, et al. Atherosclerosis in patients infected with HIV is influenced by a mutant monocyte chemoattractant protein-1 allele. *Circulation* 2004;110:2204-9.
21. Charakida M, Donald AE, Green H, Storry C, Clapson M, Caslake M, et al. Early structural and functional changes of the vasculature in HIV-infected children: Impact of disease and antiretroviral therapy. *Circulation* 2005;112:103-9.
22. Garede A, Henderson SO, Moncada S. Activated macrophages utilize glycolytic ATP to maintain mitochondrial membrane potential and prevent apoptotic cell death. *Cell Death Differ* 2010;17:1540-50.
23. Cramer T, Yamanishi Y, Clausen BE, Forster I, Pawlinski R, Mackman N, et al. HIF-1 α is essential for myeloid cell-mediated inflammation. *Cell* 2003;112:645-57.
24. Rodriguez-Prados JC, Traves PG, Cuenca J, Rico D, Aragones J, Martin-Sanz P, et al. Substrate fate in activated macrophages: A comparison between innate, classic, and alternative activation. *J Immunol* 2010;185:605-14.
25. Kubota R, Kubota K, Yamada S, Tada M, Ido T, Tamahashi N. Microautoradiographic study for the differentiation of intratumoral macrophages, granulation tissues and cancer cells by the dynamics of fluorine-18-fluorodeoxyglucose uptake. *J Nucl Med* 1994;35:104-12.
26. Folco EJ, Sheikine Y, Rocha VZ, Christen T, Shvartz E, Sukhova GK, et al. Hypoxia but not inflammation augments glucose uptake in human macrophages: Implications for imaging atherosclerosis with 18fluorine-labeled 2-deoxy-D-glucose positron emission tomography. *J Am Coll Cardiol* 2011;58:603-14.
27. Eltzschig HK, Carmeliet P. Hypoxia and inflammation. *N Engl J Med* 2011;364:656-65.
28. Tawakol A, Migrino RQ, Bashian GG, Bedri S, Vermylen D, Cury RC, et al. In vivo 18F-fluorodeoxyglucose positron emission tomography imaging provides a noninvasive measure of carotid plaque inflammation in patients. *J Am Coll Cardiol* 2006;48:1818-24.
29. Graebe M, Pedersen SF, Borgwardt L, Hojgaard L, Sillesen H, Kjaer A. Molecular pathology in vulnerable carotid plaques: Correlation with [18]-fluorodeoxyglucose positron emission tomography (FDG-PET). *Eur J Vasc Endovasc Surg* 2009;37:714-21.
30. Pedersen SF, Graebe M, Fisker Hag AM, Hojgaard L, Sillesen H, Kjaer A. Gene expression and 18FDG uptake in atherosclerotic carotid plaques. *Nucl Med Commun* 2010;31:423-9.
31. Fayad ZA, Mani V, Woodward M, Kallend D, Abt M, Burgess T, et al. Safety and efficacy of dalcetrapib on atherosclerotic disease using novel non-invasive multimodality imaging (dal-PLAQUE): A randomised clinical trial. *Lancet* 2011;377:1547-59.
32. Abdelbaky A, Corsini E, Figueroa AL, Fontanez S, Subramanian S, Ferencik M, et al. Focal arterial inflammation precedes subsequent calcification in the same location: A longitudinal FDG-PET/CT study. *Circ Cardiovasc Imaging* 2013;6:747-54.
33. Rominger A, Saam T, Wolpers S, Cyran CC, Schmidt M, Foerster S, et al. 18F-FDG PET/CT identifies patients at risk for future vascular events in an otherwise asymptomatic cohort with neoplastic disease. *J Nucl Med* 2009;50:1611-20.
34. Paulmier B, Duet M, Khayat R, Pierquet-Ghazzar N, Laissy JP, Maunoury C, et al. Arterial wall uptake of fluorodeoxyglucose on PET imaging in stable cancer disease patients indicates higher risk for cardiovascular events. *J Nucl Cardiol* 2008;15:209-17.
35. Figueroa AL, Abdelbaky A, Truong QA, Corsini E, Macnabb MH, Lavender ZR, et al. Measurement of arterial activity on routine FDG PET/CT images improves prediction of risk of future CV events. *JACC Cardiovasc Imaging* 2013;6:1250-9.
36. Yarasheski KE, Laciny E, Overton ET, Reeds DN, Harrod M, Baldwin S, et al. 18FDG PET-CT imaging detects arterial inflammation and early atherosclerosis in HIV-infected adults with cardiovascular disease risk factors. *J Inflamm (Lond)* 2012;9:26.
37. Tawakol A, Lo J, Zanni MV, Marmarelis E, Ihenachor EJ, MacNabb M, et al. Increased arterial inflammation relates to high-risk coronary plaque morphology in HIV-infected patients. *J Acquir Immune Defic Syndr* 2014;66:164-71.
38. Hsue PY, MacNabb M, Kaplan R, Nitta E, Deeks S, Martin J, et al. Arterial inflammation in HIV as measured by FDG-PET/CT is associated with splenic activity conference on retroviruses and opportunistic infections, Boston, 2014.
39. Guaraldi G, Zona S, Alexopoulos N, Orlando G, Carli F, Ligabue G, et al. Coronary aging in HIV-infected patients. *Clin Infect Dis* 2009;49:1756-62.