

Trehalose accumulation enhances drought tolerance by modulating photosynthesis and ROS‑antioxidant balance in drought sensitive and tolerant rice cultivars

Akhil Mohanan1 · Anjali Kodigudla1 · Dhana Ramya Raman1 · Kavya Bakka² · Dinakar Challabathula[1](http://orcid.org/0000-0002-7158-5902)

Received: 28 July 2023 / Revised: 30 November 2023 / Accepted: 11 December 2023 / Published online: 28 December 2023 © Prof. H.S. Srivastava Foundation for Science and Society 2023

Abstract

Trehalose being an integral part for plant growth, development and abiotic stress tolerance is accumulated in minute amounts in angiosperms with few exceptions from resurrection plants. In the current study, two rice cultivars difering in drought tolerance were used to analyse the role of trehalose in modulating photosynthesis and ROS-antioxidant balance leading to improvement in drought tolerance. Accumulation of trehalose in leaves of Vaisakh (drought-tolerant) and Aiswarya (droughtsensitive) rice cultivars was observed by spraying 50 mM trehalose and 100 μ M validamycin A (trehalase inhibitor) followed by vacuum infltration. Compared to stress sensitive Aiswarya cultivar, higher trehalose levels were observed in leaves of Vaisakh not only under control conditions but also under drought conditions corresponding with increased root length. The increase in leaf trehalose by treatment with trehalose or validamycin A corresponded well with a decrease in electrolyte leakage in sensitive and tolerant plants. Decreased ROS levels were refected as increase in antioxidant enzyme activity and their gene expression in leaves of both the cultivars treated with trehalose or Validamycin A under control and drought conditions signifying the importance of trehalose in modulating the ROS-antioxidant balance for cellular protection. Further, higher chlorophyll, higher photosynthetic activity and modulation in other gas exchange parameters upon treatment with trehalose or validamycin A strongly suggested the benefcial role of trehalose for stress tolerance. Trehalose accumulation helped the tolerant cultivar adjust towards drought by maintaining higher water status and alleviating the ROS toxicity by efective activation and increment in antioxidant enzyme activity along with enhanced photosynthesis.

Keywords Antioxidant enzymes · Drought stress · Gene expression · Photosynthesis · ROS · Trehalose

Introduction

Rice (Oryza sativa) is the widely cultivated crop considered as staple food of not only India but also in diferent parts of Asia, Latin America and Africa. Fluctuations in the environmental conditions pose a threat of stress to rice plants afecting their growth and productivity. Decades of breeding practices and research has led to development of drought tolerant rice varieties with varied stress tolerance

limit (Reyes [2023\)](#page-13-0). Among the varied mechanisms, accumulation of compatible solutes and sugars are considered efective as they play a crucial role in cellular protection and abiotic stress tolerance (Blum [2017;](#page-12-0) Huang et al. [2020](#page-12-1)). In many instances, some of the unusual sugars such as rafnose, stachyose, verbascose, octulose etc. were also observed in several of the plant species whose functions are not yet clearly understood (Peters et al. [2007;](#page-13-1) Dinakar and Bartels [2013;](#page-12-2) Egert et al. [2015](#page-12-3); Sengupta et al. [2015](#page-13-2)). Drought induced accumulation of trehalose although is not common in many plants, is observed in several desiccation tolerant plants such as *Selaginella lepidophylla* and *Myrothamnus fabellifolia* (Drennan et al. [1993](#page-12-4); Iturriaga et al. [2009](#page-12-5)). The trehalose accumulating transgenic plants, with altered trehalose biosynthesis or degradation showed tolerance towards multiple stresses suggesting its importance for abiotic stress tolerance (Li et al. [2011](#page-13-3); Lin et al. [2019](#page-13-4); Kosar et al. [2019;](#page-12-6) Hassan et al. [2022\)](#page-12-7). Modulation in

 \boxtimes Dinakar Challabathula dinakarc@cutn.ac.in

¹ Department of Life Sciences, School of Life Sciences, Central University of Tamil Nadu, Thiruvarur, Tamil Nadu 610 005, India

Department of Microbiology, School of Life Sciences, Central University of Tamil Nadu, Thiruvarur, Tamil Nadu 610005, India

physiological, biochemical, molecular and photosynthetic functions upon exogenous application of trehalose is also observed in several plants including *Arabidopsis thaliana*, *Triticum aestivum* and *Zea mays* (Yang et al. [2014;](#page-14-0) Kosar et al. [2019](#page-12-6); Hassan et al. [2022](#page-12-7); Zhang et al. [2022\)](#page-14-1).

Drought stress reduces leaf water potential, turgor pressure and affects various physiological and biochemical functions including photosynthesis, chlorophyll synthesis and carbohydrate metabolism signifcantly afecting the growth and cellular homeostasis ultimately leading to drastic decrease in crop productivity and causing cell death (Biswas et al. [2016;](#page-12-8) Bashir et al. [2021](#page-12-9); Kaur et al. [2021](#page-12-10); Challabathula et al. [2022\)](#page-12-11). The declining water status is critical as most of rice cultivated areas are rain fed and the cultivars developed mostly seek good irrigation indicating the importance of water for productivity of rice plants. The prolonged drought increases the cellular ROS levels leading to oxidative damage. Although ROS production and signalling is important in eliciting the antioxidant defence responses for withstanding stress and protecting the cellular integrity, sensitive cultivars fail to employ the mechanisms that could efficiently regulate ROS accumulation thereby resulting in increased oxidative damage. The accumulation of ROS such as singlet oxygen $({}^{1}O_2)$, superoxide radical (O_2^-) , hydrogen peroxide (H_2O_2) and the hydroxyl radical (HO•) constitutes unrestricted oxidation of cellular components leading to oxidative damage and cell death. Prolonged drought makes the sensitive cultivars vulnerable to ROS induced damages, predominantly the lipid peroxidation of membranes and oxidation of amino acid side chains and protein backbone (Asada [2006;](#page-12-12) Das and Roy-Choudhury [2014](#page-12-13); Mittler [2017](#page-13-5)). Higher H_2O_2 accumulation was observed in the sensitive rice cultivar, Aiswarya during drought than the tolerant cultivar Vaisakh suggesting the efective ROS management in tolerant cultivar (Challabathula et al. [2022\)](#page-12-11). In another study, although, the salt tolerant cultivars showed higher generation of H_2O_2 and higher activity of ROS generating enzyme NADPH oxidase, it was compensated by a higher activity of both enzymatic and non-enzymatic antioxidant system (Kaur et al. [2016](#page-12-14)). Compared to drought sensitive plants, the drought tolerant plants are endowed with several mechanisms for decreasing the ROS induced oxidative damage, maintenance of the cellular water status and limitation in membrane damage during stress (Sharma and Dubey [2005](#page-13-6); Das and Roychoudhury [2014;](#page-12-13) Mishra and Panda [2017](#page-13-7); Irato and Santovito [2021](#page-12-15); Challabathula et al. [2022](#page-12-11)).

Plants sense the changes in water deficit and send the signals from root to leaves by accumulating abscisic acid (ABA) (Biswas et al. [2016\)](#page-12-8). The ABA signalling is necessary to control and initiate the stomatal closure preventing water loss by transpiration. An ABA independent regulatory system also exists in inducing the stomatal closure which later gets the assistance from ABA accumulation (Tombesi et al. [2015](#page-13-8)). The stomatal closure, as a drawback, results in the limitation of gas exchange thereby decreasing the photosynthesis. Since the entry of $CO₂$ is restricted due to stomatal closure, the photosynthesis majorly depends on internal $CO₂$ availability. The inadequate availability of $CO₂$ to chloroplasts results in decreased Ribulose-1-5-bisphosphate carboxylase oxygenase (RubisCO) activity thereby afecting photosynthesis. Drought also is known to afect the chloroplast, by damaging its ultrastructure, damaging the thylakoids and degrading chlorophyll molecules (Wang et al. [2022](#page-13-9)). Free radical generation and lipid peroxidation also are drought associated changes that occur in chloroplasts. In stress tolerant plants, photosynthesis is maintained to a certain extent by early sensing and preparing for an upcoming stress. Maintenance of higher intracellular $CO₂$ levels, higher photosynthesis rate and enhanced antioxidant defence system are signifcant features of drought tolerant plants (Ullah et al. [2021\)](#page-13-10). The accumulation of osmolytes is one of the important strategies adopted by tolerant plants. Stress induced expression of genes involved in osmolyte biosynthesis, water channel proteins and protective enzymes are also known to be involved in protection from drought stress induced oxidative damage (Ullah et al. [2021](#page-13-10)).

Trehalose accumulation in plants leads to stress tolerance. The accumulation of trehalose in the dehydrated tissues of resurrection plants are signifcant and are often discussed for the association of this sugar with to stress tolerance. The desiccation/dehydration induced changes in cells causes membrane damage and disintegration of macromolecules. Trehalose acts as an osmoprotectant molecule stabilizing the cell membranes and proteins by forming a glassy matrix around them. The modulation of oxidative stress response, induction of stress responsive genes, improved photosynthesis is observed in plants accumulating trehalose (Luo et al. [2021](#page-13-11); Zhang et al. [2022](#page-14-1); Mohanan et al. [2023\)](#page-13-12). Rice plants exposed to drought stress showed increased accumulation of trehalose. Further, the transgenic rice plants accumulating trehalose showed improved drought tolerance (Mostofa et al. [2015a](#page-13-13); Kosar et al. [2019](#page-12-6); Sadak et al. [2019;](#page-13-14) Hassan et al. [2022](#page-12-7)). The accumulation of trehalose either through exogenous application or by generation of transgenic plants harbouring genes for trehalose metabolism has led to enhanced drought tolerance with better photosynthesis (Garg et al. [2002](#page-12-16); Karim et al. [2007](#page-12-17); Van Houtte et al. [2013\)](#page-13-15). The exogenous application of trehalose leads to the accumulation of trehalose along with induction of stress responsive dehydrin genes in *Arabidopsis thaliana* plants (Mohanan et al. [2023](#page-13-12)). Another approach for the accumulation of trehalose in leaves can be achieved through the application of trehalase inhibitor validamycin A. Validamycin A is an aminoglycoside antibiotic with strong potential to inhibit trehalases in plants, insects and fungi. Further due to its fungicidal and antibacterial activity, it is used to control sheath-blight disease in

rice and soil borne pathogens (Ishikawa et al. [2005](#page-12-18); Yang et al. [2023\)](#page-14-2). In *A. thaliana*, validamycin A increases the trehalose accumulation by effectively inhibiting the trehalase enzyme activity (Müller et al. [2001](#page-13-16); Vogel et al. [2001](#page-13-17)). Further, in Tobacco plants, 100 µM validamycin inhibited more than 99% of trehalase activity indicating its efectiveness in increasing the trehalose accumulation in plants (Goddijn et al. [1997](#page-12-19)).

In our previous study, the diferential modulation of photosynthesis, ROS and antioxidants were evaluated in drought stress sensitive and tolerant rice cultivars (Aiswarya and Vaisakh) along with salt stress tolerant cultivar Vyttila upon inhibition of cytochrome oxidase (COX) and alternative oxidase (AOX) pathways of mitochondrial oxidative electron transport during drought and salinity stress (Challabathula et al. [2022](#page-12-11)). The current study is focused on exploring the importance of trehalose for drought stress tolerance using the stress tolerant Vaisakh and sensitive Aiswarya plants. The modulation in photosynthesis and ROS-antioxidant balance in drought sensitive and tolerant rice cultivars upon treatment with trehalose or validamycin during drought stress conditions was analysed to shed light on the importance of trehalose for photosynthesis and ROS-antioxidant homeostasis during drought stress.

Materials and methods

Plant growth and treatment

Seeds of drought sensitive rice cultivar Aiswarya were obtained from the Regional Agricultural Research Station, Pattambi, Kerala and the seeds of drought tolerant rice cultivar Vaisakh were obtained from Rice Research Station, Vyttila, Kerala, India. The seeds were surface sterilized with Sodium hypochlorite (4% v/v) followed by thoroughly washing them with sterile distilled water. After cold-stratifcation at 4 °C for three days, the seeds were directly sown in circular pots (7 cm height and 5 and 6 cm diameter at bottom and top respectively) containing sterile, dry, potting mixture consisting of red soil, farmyard manure and soilrite (1:1:1 ratio). The pots flled with 75 gm of the potting mixture were wetted with 100 ml of water. The pots were transferred to growth chambers maintained at 25 $\mathrm{°C/}$ 22 $\mathrm{°C}$ the day/ night temperature, and 8/16 hrs light (110 µmol m⁻² sec⁻¹ PPFD illumination) /dark photoperiods. While control plants were regularly irrigated with 50 ml of water, drought stress was imposed by discontinuing irrigation of pots which were initially maintaining at a constant soil moisture of >90%. The trehalose (50 mM) and validamycin A (100 μ M) treatment was done by foliar spraying and vacuum infltration (20 KPa pressure for 2 min) of 10 ml of solution prior to imposing drought stress treatment following the method described by

Challabathula et al. [\(2022\)](#page-12-11). Drought stress treatments were terminated after ten days and the leaf samples were collected for further analysis.

Measurement of relative water content (RWC) and root length

Leaf RWC was measured according to Kuroki et al. [\(2019](#page-12-20)). Even-sized leaves (3 replicates for each treatment) from the control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment were cut from base and the fresh weight (FW) was measured. The leaves were immersed in distilled water for 24 h and the turgid weight (TW) was measured. The dry weight (DW) was measured after drying the leaves for 24 h at 80 $^{\circ}$ C in a hot air oven. The leaf RWC was calculated using the following formula:

$$
RWC(\%) = \frac{(FW - DW)}{(TW - DW)} \times 100
$$

Roots from the control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment were removed from the soil and cleaned with fresh water prior to measuring the root length. Root length (in cms) of at least 10 plants from each treatment were measured using a standard measuring scale starting from the base to the tip of the longest root.

Trehalose estimation

Trehalose estimation from the leaves of control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment were carried out as per the method described by Mostofa et al. [\(2015a\)](#page-13-13). The leaves, 50 mg from each treatment were cut, ground with 1 ml of hot 80% ethanol followed by centrifugation at 10,000 rpm for 20 min at room temperature. The supernatants collected were dried at 80 °C to remove the ethanol traces and were re-suspended in 5 ml of distilled water. The 300 µl of 0.2 N sulphuric acid was added to 200 µl of the suspended solution and were boiled at 100 °C for 10 min followed by chilling on ice for 5 min. To this mixture, 300 µl of 0.6 N sodium hydroxide was added and was boiled for 10 min and were cooled on ice for 5 min. To this, 4 ml of anthrone reagent was followed by boiling for 10 min and chilling for 5 min respectively. The absorbance of the solutions was recorded at 630 nm using a UV - visible spectrophotometer. The trehalose contents were calculated using a standard curve and the graphs were plotted.

Measurement of leaf electrical conductivity (EC) and histochemical staining for ROS accumulation in leaves

For the assessment of membrane damages in leaves, leaf electrical conductivity was measured following the methodology described by Challabathula et al. [\(2022](#page-12-11)). Even sized leaves from control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment were detached and were cut into smaller pieces and dispersed and vortexed in 10 ml of ultrapure distilled water. Initial conductivity (EC1) of the solution was measured after 2 h of incubation using a conductivity meter 304, conductivity cell type CD-10 cell $K1 \pm 10\%$ (Systronics India Ltd, India) set within a range of 2 mS at 25° C. The final conductivity (EC2) was measured after boiling the samples at 100 o C for 10 min. The relative leaf EC was calculated as per Muchate et al. ([2019](#page-13-18)) using the following formula,

$$
EC(\%) = \frac{EC_1}{EC_2} \times 100
$$

The histochemical staining for ROS such as H_2O_2 and O_2 ⁻ in the leaves of control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment was performed according to Taj and Challa-bathula ([2021\)](#page-13-19). Histochemical staining of leaves for H_2O_2 was performed with 3, 3'-diaminobenzidine (DAB) and O_2 ⁻ was performed using nitro blue tetrazolium (NBT). The leaves that were cut from the base were immersed in either 1 mg ml⁻¹, NBT or 1 mg ml⁻¹ DAB in 10 mM potassium phosphate buffer pH 7.8 followed by vacuum infiltration for 2 min at 20 KPa for three times. The samples were kept under darkness for overnight followed by illumination under continuous light (300 µmol m^{-2} s⁻¹) for 8 h. The pigments of the leaves were removed by destaining the leaves with methanol, acetic acid, and glycerol in a 3:2:1 ratio and fxed using fxative solution. After complete destaining, the Images were taken at 5 x magnifcation using a stereo zoom microscope, Nikon SMZ800N attached with Nikon DS-Fi3 camera (Nikon, Japan).

In‑gel antioxidant activity staining

In-gel antioxidant enzyme activities were performed by specifc staining procedures by following the methodology described by Analin et al. [\(2020\)](#page-12-21). The proteins were extracted from 500 mg of leaf samples from control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment by grinding in liquid nitrogen and thoroughly dissolving in the extraction bufer consisting of 1mM PMSF, a protease inhibitor, in 50mM sodium phosphate buffer pH 7.0 followed by centrifugation at 12,000 rpm for 5 min at 4 °C. The supernatant was used for protein quantifcation using Lowry's method (Lowry et al. [1951\)](#page-13-20). Native PAGE was done on gel consisting of 10% resolving gel (10% acrylamide-bisacrylamide, 375mM Tris-HCl pH 8.8, 10% glycerol, 0.05% Ammonium Persulphate; APS and 0.05% tetramethylethylenediamine; TEMED) and stacking gel containing 4% acrylamide-bisacrylamide, 126mM Tris-HCl pH 6.8, 10% glycerol, 0.05% APS and 0.05% TEMED. From the extracted total protein, 100 µg protein was loaded onto the gels and were run in the running bufer (tris - glycine for SOD and CAT, and tris-glycineascorbate for APX) for overnight at 4 °C at 30 V. Prior to the protein loading, the gels were pre-run in their respective running buffers. The gels were stained for the visualization of enzyme bands. For SOD, the gel was initially soaked in 2.45 mM NBT and incubated in dark for 20 min. The solution was removed and the gel was incubated in a mixture containing 28 mM TEMED and 2.4 µM ribofavin in 50 mM potassium phosphate pH 7.8 buffer and kept under illumination. Transparent bands were seen in violet stained background after 30 min of incubation. For CAT activity, the respective gel was washed with distilled water thrice, 15 min each time. The gel was then incubated in 20 mM hydrogen peroxide for 10 min in dark. After discarding the solution, the gel was washed with distilled water thrice, 5 min each time. Then, 1% ferric chloride and 1% potassium ferricyanide were prepared separately and poured onto the gel. After 10 to 15 min, transparent bands in green stained background were observed (Woodbury et al. [1971\)](#page-14-3). The gel run with tris- glycine -ascorbate was used to stain for APX. After electrophoresis, the gel was initially washed thrice with 2 mM ascorbic acid in 50 mM sodium phosphate bufer pH 7.0, 10 min each time followed by incubation in a solution containing 4 mM ascorbic acid and 20 mM hydrogen peroxide in 50 mM sodium phosphate bufer pH 7.0 for 20 min. Then, a solution of 28 mM TEMED and 2.4 mM NBT in 50 mM potassium phosphate bufer pH 7.8 was prepared and added to the gel. The gel was incubated for 20 min until transparent bands were visible in dark blue background (Lee and Lee [2000\)](#page-13-21). The gels were scanned using the scanner, CanoScan LiDE120 (Canon, USA).

RNA extraction and gene expression

The RNA extraction was performed following the method described by Valenzuela-Avendaño et al. ([2005\)](#page-13-22). The leaves of control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment were ground in liquid nitrogen using the extraction bufer (38% bufer saturated phenol, 0.8 M guanidine thiocyanate, 0.4 M ammonium thiocyanate, 0.1 M sodium acetate and 5% glycerol). The leaf extracts incubated in room temperature for 10 min were centrifuged for 10 min at 9200 rpm and 300 µl of chloroform-isoamyl alcohol was added to the supernatant, vortexed and centrifuged at 9200 rpm for 10 min at 4 °C. In a fresh tube, the aqueous phase on the upper side was taken and 375 µl each of isopropanol and 0.8 M sodium citrate/1 M sodium chloride were added and incubated at room temperature for 10 min followed by centrifugation at 10,100 rpm for 10 min at 4 °C. The pellet was washed using ice cold 70% ethanol followed by centrifugation at 10,100 rpm for 10 min at 4 °C. The RNA pellets were dissolved in RNase free water and quantifed using Genova nano spectrophotometer (Jenway, UK). Using iScript cDNA synthesis kit, the cDNA was synthesized using 1 µg of RNA (Bio-rad, USA) which is used as template in RT-PCR reaction along with gene specifc primers for monitoring the gene expression levels. The list of primers and the sequences are provided in the Supplementary Table 1. Actin was used as a house keeping gene. The PCR program consisted of initial denaturation and denaturation both set at 95 $\mathrm{^{\circ}C}$ for 1 min followed by annealing for 1 min set at 60 °C. The extension was set at 72

^oC for 2 min. A final extension was done at 72 °C for 2 min. C for 2 min. A final extension was done at 72° C for 2 min with 28 cycles. The amplifed products along with 1 kb DNA ladder were separated on 2% agarose gel, and the bands were visualized using gel documentation system (Fusion Solo S, Vilber Lour mat, France).

Chlorophyll estimation

Chlorophyll estimation was done according to Arnon ([1949](#page-12-22)). From the leaves of control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment, 100 mg leaves were ground with 80% acetone at 4 ^oC under dark. The extracts were centrifuged and the supernatants were used for chlorophyll estimation spectrophotometrically by measuring the absorbance of chlorophyll a, chlorophyll b, at 663, and 645 nm, respectively. Arnon [\(1949\)](#page-12-22) equations were used to calculate the chlorophyll contents.

Chlorophyll b(mg g⁻¹FW) = (22.9 x(Ab645) – 4.68(Ab663))x $\frac{V}{100}$ xWand Chlorophyll a(mg g⁻¹FW) = (12.7 x(Ab663) – 2.69(Ab645))x $\frac{V}{100}xW$, Total chlorophyll $(mg g^{-1}FW) = Chlorophyll a + chlorophyll b$

where V is fnal volume of chlorophyll extracted in 80% acetone; and W, fresh weight of the leaf used.

Measurements of leaf gas exchange parameters

The leaf gas exchange parameters including $CO₂$ assimilation rate, stomatal conductance (g_s) , intercellular CO_2 concentration and the ratio of intercellular to ambient $CO₂$ concentration (Ci/Ca) were measured in the leaves of control and drought stress treated rice cultivars with and without trehalose and validamycin A treatment using a portable photosynthesis system (LI-6400 XT; LI-COR Inc., Lincoln, Neb. USA) following Yao et al. [\(2017](#page-14-4)) and Analin et al. ([2020](#page-12-21)). Four leaves of the rosette were used for covering the 2 cm^2 chamber area of the gas analyser. A PPFD of 1500 μ mol photons m⁻² s⁻¹, CO₂ concentration at 400 μ mol mol⁻¹ and temperature 25 ± 2 °C were set for the measurements.

Statistical analysis

All the experiments were conducted in at least three replicates and data represents an average of three biological replicates. The normality of the data was performed using Shapiro–Wilk test. The statistical analysis was done using Multiple comparison by means of Two-way ANOVA, the Tukey's method. The statistically signifcant diferent groups were represented with different alphabets over the bars showing significant differences among them. $(p = 0.001)$ with an overall significance level = 0.05. Error bars represent means \pm SE ($n=3$) obtained from three biological replicates. The calculation of statistics and plotting of graphs were done using GraphPad Prism version 8.0.0 for Windows (GraphPad Software, San Diego, California USA).

Results

Trehalose accumulation in rice cultivars alters the drought associated physiological changes

As already reported by Challabathula et al. ([2022](#page-12-11)), the drought tolerant cultivar Vaisakh showed superior response than the sensitive cultivar Aiswarya which was reflected in its phenotype, leaf relative water content levels and leaf electrical conductivity (EC) during drought stress (Fig. [1\)](#page-5-0). Upon drought, while the sensitive cultivar maintained RWC < 20% which resulted in drying with increased EC (82%, Fig. [1a](#page-5-0) and b), the tolerant cultivar maintained higher RWC and comparatively lower EC than the sensitive cultivar (Fig. [1](#page-5-0)c). Interestingly, the leaves of the tolerant plant showed higher trehalose (3.54 µg g^{-1} FW) levels in its leaves than the sensitive cultivar (1.6 µg g-1 FW) under control conditions hinting towards the protective role of this sugar (Fig. [1d](#page-5-0)). Further, the trehalose accumulation in leaves of both sensitive and tolerant plants was although found to be enhanced by treatment with either 50 mM trehalose or 100 μ M validamycin A, higher trehalose accumulation was observed in stress tolerant cultivar Vaisakh (Fig. [1](#page-5-0)d). The accumulation of trehalose helped both the cultivars to maintain a higher

Fig. 1 Trehalose accumulation and associated changes in rice cultivars Vaisakh and Aiswarya during drought. The plants were subjected to drought stress for 10 days and trehalose and validamycin A treatments were given as described in Materials and methods. The treatments are represented as; control plants, C; control+50 mM trehalose, TC; control+100 µM validamycin A, VC; drought stressed, D; drought+50 mM trehalose, TD; and drought+100 µM validamycin A, VD. "dh" indicates dehydration/drought stress treatment. Unstressed and drought stressed Vaisakh (grey bars) and Aiswarya

RWC and a lower EC indicating the importance of trehalose in drought tolerance.

Trehalose accumulation alters root length in rice cultivars during drought stress

Trehalose accumulating seedlings of drought stress tolerant Vaisakh and drought stress sensitive Aiswarya cultivars showed diferential root growth patterns. While signifcant increase in root growth was observed in Vaisakh cultivar upon trehalose accumulation, the root growth was marginally increased in Aiswarya cultivar (Fig. [2a](#page-6-0), b and c). Although, drought stress has led to decrease in root growth in Vaisakh plants, accumulation of trehalose

(black bars) were represented with solid and pattern flled bars respectively. **a** The phenotypic changes in Vaisakh and Aiswarya. **b** The relative water content, **c** electrical conductivity and **d** trehalose accumulation in Vaisakh and Aiswarya cultivars is plotted. Two-way ANOVA using Tukey's multiple comparisons method was used to assess the statistical signifcance. Diferent alphabets indicate statistically significant differences among each group. $P = < 0.001$ with overall significance level = 0.05. Error bars represent means \pm SE (n = 3)

due to trehalose/validamycin A supplementation has led to increased root growth.

Trehalose accumulation diferentially modulates oxidative stress responses in drought sensitive and tolerant rice cultivars

The oxidative stress responses of the trehalose accumulating drought sensitive and drought tolerant rice cultivars were assessed by analysing the ROS accumulation in leaves by histochemical staining and associated antioxidant enzyme activities of SOD, CAT and APX by activity staining. Drought stress has resulted in increased ROS, (H_2O_2) in leaves of sensitive cultivar which was aggravated with

Fig. 2 Root growth in drought sensitive and drought tolerant rice cultivars treated with trehalose and validamycin A prior to imposing drought stress conditions. The validamycin A and trehalose treatments were done as described in Materials and methods and the plants were subjected to drought stress for 10 days. Root morphology of (**a**) Vaisakh, (**b)** Aiswarya and (**c)** root length in rice cultivars is shown. The treatments are represented as; control plants, C; control + 50 mM trehalose, TC; control + 100 μ M validamycin A, VC; drought stressed, D; drought+50 mM trehalose, TD; and

drought+100 µM validamycin A, VD. "Dehy" indicates dehydration/ drought stress treatment. Unstressed and drought stressed Vaisakh (grey bars) and Aiswarya (black bars) were represented solid and pattern flled bars respectively. Two-way ANOVA using Tukey's multiple comparisons method was used to assess the statistical signifcance was represented with diferent alphabets representing statistically significant differences among each group. $P = < 0.001$ with overall significance level=0.05. Error bars represent means \pm SE (n=10). Scale bar represents 1 cm

Fig. 3 Histochemical staining for ROS and analysis of antioxidant enzyme activity in rice cultivars difering in drought tolerance upon treatment with trehalose and validamycin A prior to imposing drought stress conditions. The localization of (a) H₂O₂ and superoxide in Vaisakh (left plate) and Aiswarya (right plate) were done by staining with diaminobenzidine (DAB) and nitroblue tetrazo-

lium (NBT) respectively. (**b**) The in-gel antioxidant enzyme activity of SOD, CAT and APX. The treatments are represented as; control plants, C; drought stressed, D; control+50 mM trehalose, T; drought + 50 mM trehalose, TD; control + 100 μ M validamycin A, V; and drought $+100 \mu M$ validamycin A, VD.

validamycin A and trehalose treatment (Fig. [3a](#page-6-1)). Although, mild increase in ROS levels were observed in drought tolerant cultivar, the H_2O_2 levels were lower than those observed

in leaves of stress sensitive cultivar Aiswarya. Further, trehalose accumulation resulted in decline in H_2O_2 accumulation during drought stress in stress tolerant cultivar. Compared to leaves of Vaisakh, higher levels of superoxide radicals were observed during drought stress conditions in the leaves of Aiswarya cultivar. However, the changes were marginal in the presence of validamycin A and trehalose. The results with ROS were in agreement with in-gel antioxidant enzyme activity wherein SOD activity was higher in both the cultivars upon trehalose and validamycin treatment during drought stress. Although, the induction in SOD and APX activities by drought were observed in both the cultivars, the induction of antioxidant enzyme activities during drought and with the treatment of trehalose and validamycin A were comparatively higher in tolerant cultivar than in sensitive cultivar (Fig. [3b](#page-6-1)). The induction in the CAT activity was observed by the treatment with trehalose and Validamycin A in leaves of Vaisakh plants, however, the CAT activity is not observed during drought stress. Compared to Vaisakh plants, the leaves of Aiswarya plants showed higher CAT activity during drought stress. Further, while the gene expression levels of APX increased with trehalose and validamycin A treatment in both Vaisakh and Aiswarya (Fig. [4\)](#page-8-0), higher expression levels were observed in the Vaisakh cultivar (Fig. [4](#page-8-0)a and b). Increased expression of APX and Cu/Zn SOD in leaves of Vaisakh plants during drought upon treatment with validamycin A was observed. The expression of CAT gene was only observed in trehalose and validamyin A treated leaves of Vaisakh plants (Fig. [4](#page-8-0)a and b). The results indicated modulation of ROS accumulation and associated antioxidant activities in trehalose accumulating plants during stressed and unstressed conditions.

Chlorophyll accumulation in leaves of Vaisakh and Aiswarya cultivars upon treatment with trehalose and validamycin A under drought stress conditions.

Increased chlorophyll content was observed in the leaves of Vaisakh and Aiswarya cultivars upon treatment with trehalose and validamycin A (Fig. [5](#page-8-1)). However, the chlorophyll content was signifcantly higher in leaves of Vaisakh than Aiswarya. Among chlorophyll a and chlorophyll b contents, while the chlorophyll a content showed significant increase in trehalose or validamycin a treated leaves of Vaisakh plants, the chlorophyll b content was decreased (Fig. [5](#page-8-1)b). Contrary to this, the decline in chlorophyll a content and an increase in chlorophyll b content was observed after trehalose and validamycin treatment in Aiswarya cultivar (Fig. [5a](#page-8-1) and b). During drought, comparatively higher chlorophyll a and chlorophyll b levels were observed in the leaves of Vaisakh plants than Aiswarya plants upon supplementation with trehalose and validamycin A (Fig. [5a](#page-8-1), b).

The photosynthetic responses towards trehalose accumulation in rice cultivars during drought stress

The $CO₂$ assimilation rates were comparatively higher $(5.129 \text{ µmol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$, Fig. [6a](#page-9-0)) in the leaves of Vaisakh than Aiswarya (2.361 µmol CO_2 m⁻² s⁻¹, Fig. [6a](#page-9-0)). Further, the rates were signifcantly increased when the leaves of Vaisakh plants were treated with 50 mM trehalose or 100 μ M validamycin A. Similar observations were noticed for other gas exchange parameters such as stomatal conductance (gs, Fig. [6](#page-9-0)b) internal $CO₂$ (Ci, Fig. [6c](#page-9-0)) and Ci/Ca ratio (Fig. 6d). In stress sensitive cultivar Aiswarya, trehalose treatment has led to 3-fold increase in $CO₂$ assimilation and 2-fold increase when treated with validamycin A (Fig. [6](#page-9-0)a). The other gas exchange parameters also increased with trehalose treatment, but decreased with validamycin A treatment. A drastic decrease in $CO₂$ assimilation rates and gs were observed in both the cultivars when exposed to drought. While the leaves of Vaisakh maintained comparatively higher $CO₂$ assimilation rates (0.516 µmol CO_2 m⁻² s⁻¹), the leaves of Aiswarya showed a very low CO_2 assimilation rate (0.033 µmol CO_2) $m^{-2} s^{-1}$) with higher gs (0.023 mol H₂O m⁻² s⁻¹, Fig. [6](#page-9-0)a and b). Trehalose accumulation through 50 mM trehalose and 100 µM validamycin, improved the gas exchange parameters where Vaisakh showed higher photosynthetic CO2 assimilation rates.

Correlation of physiological parameters with trehalose accumulation in leaves of Vaisakh and Aiswarya plants

While the trehalose accumulation in leaves of Vaisakh plants under unstressed condition showed a positive correlation with physiological parameters like RWC $(r=0.95)$, total chlorophyll content (0.94) and the gas exchange parameters like $CO₂$ assimilation (0.65), gs (0.98), ci (0.94) and Ci/ Ca ratio (0.93), the EL was negatively correlated (-0.42). This was consistent with the antioxidant gene expression of Cu/ Zn SOD (0.99), APX (1) and CAT (0.49). Although the trehalose accumulating leaves of drought stressed Vaisakh plants showed negative correlation with root length, EL, gs, Ci and Ci/Ca, the positive correlation with RWC (0.99), CO_2 assimilation (0.86) and antioxidant gene expression was observed. In contrast to this, the drought sensitive Aiswarya cultivar with trehalose treatment showed positive correlation for photosynthesis and APX gene expression under control conditions and RWC, chlorophyll content, photosynthetic $CO₂$ assimilation and APX gene expression under drought stress conditions. These results indicated a trehalose assisted improvement of physiological, biochemical characteristics along with improvement

Fig. 4 The expression analysis of genes encoding antioxidant enzymes *Cu/Zn SOD*, *CAT*, and *APX* in the trehalose accumulating leaves of drought-tolerant Vaisakh and drought-sensitive Aiswarya during drought stress conditions. Actin was used as a reference gene for constitutive expression. The treatments are represented as; control plants, C; drought stressed, D; control+50 mM trehalose, T; drought + 50 mM trehalose, TD; control + 100 μ M validamycin A, V, and drought $+100 \mu M$ validamycin A, VD. "ne" denotes no expression and "Dehy" indicates dehydration/drought stress treatment. A densitogram was made based on the relative expression of gene of interest with actin. (**a**) The gel picture is with the relative expression of (**b**) *APX*, (*Cu / Zn SOD* and CAT expression in Vaisakh (grey bars) and Aiswarya (black bars). Unstressed and drought stressed plants are shown as solid and pattern flled bars respectively. Twoway ANOVA using Tukey's multiple comparisons method was used to assess the statistical signifcance was represented with diferent alphabets representing statistically signifcant diferences among each group. $P = < 0.001$ with overall significance level = 0.05. Error bars represent means \pm SE (n=3)

Fig. 5 Chlorophyll content in leaves of trehalose accumulating drought-tolerant Vaisakh and drought-sensitive Aiswarya subjected to drought stress. The treatments are represented as; control plants, C; control+50 mM trehalose, TC; control+100 µM validamycin A, VC; drought stressed, D; drought + 50 mM trehalose, TD; drought + 100 µM validamycin A, VD. "dh" indicates dehydration/drought stress treatment. Unstressed and drought stressed plants are shown as solid and pattern flled bars respectively. (**a**) Chlorophyll a (**b**) Chlorophyll b and (**c**) total chlorophyll in Vaisakh (grey bars) and Aiswarya (black bars). Two-way ANOVA using Tukey's multiple comparisons method was used to assess the statistical signifcance. Diferent alphabets above the bars represent statistically signifcant diferences among each group $P = < 0.001$ with overall significance level = 0.05. Error bars represent means \pm SE (n=3)

in antioxidant gene expression and gas exchange parameters helping the tolerant cultivar to manage dehydration.

Discussion

Drought tolerance is a complex phenomenon involving mechanisms for efficient sensing of dehydration along with recruitment of mechanisms for defending and minimizing

Fig. 6 Photosynthetic gas exchange parameters analysed in trehalose accumulating drought-tolerant Vaisakh and drought-sensitive Aiswarya plants subjected to drought stress. The treatments are represented as; control plants, C; control+50 mM trehalose, TC; control + 100 μ M validamycin A, VC; drought stressed, D; drought + 50 mM trehalose, TD; drought + 100 μM validamycin A, VD. "dh" indicates dehydration/drought stress treatment. (a) CO₂ assimilation rates (**b**) stomatal conductance (**c**) Ci (**d**) Ci/ Ca in Vaisakh (grey bars) and Aiswarya (black bars) cultivars are plotted. Two-way ANOVA using Tukey's multiple comparisons method was used to assess the statistical signifcance. Diferent alphabets represent statistically signifcant differences among each group $P = 0.001$ with overall significance level = 0.05. Error bars represent means \pm SE (n=3)

the stress associated damage. Rice cultivars difering in the sensitivity and tolerance to drought serve as valuable resource for identifying the mechanisms of stress tolerance. Analysing the intrinsic diferences in between drought sensitive and tolerant cultivars and identifying the mechanisms employed by the tolerant plants is important to unravel the trivial components for stress tolerance behaviour (Bartels and Dinakar [2013\)](#page-12-2). Among the two rice cultivars used in the current study, Vaisakh is a drought tolerant and Aiswarya is drought sensitive (Fig. [1](#page-5-0)a, Challabathula et al. [2022\)](#page-12-11). Previously, we used these cultivars along with salt tolerant rice cultivar Vyttila to analyse the importance of cytochrome oxidase and alternative oxidase pathways of mitochondrial oxidative electron transport chain in modulating photosynthesis, ROS and antioxidants during salinity and drought stress (Dinakar et al., 2022). In the current study, the importance of trehalose in modulating the photosynthesis and ROS-antioxidant balance during drought stress in Vaisakh and Aiswarya rice cultivars is analysed. Since the drought tolerant Vaisakh plants were able to rehydrate after 10 days of drought stress as indicated in our previous study (Challabathula et al. [2022\)](#page-12-11), the drought stress was imposed for 10 days duration for both sensitive and tolerant plants. In the current and previous study with Aiswarya and Vaisakh cultivars, the Relative water content of leaves was considered as the primary indicator of stress wherein the tolerant cultivar maintained around 40% RWC which supported its tolerance and revival upon rehydration. Thus 10 days of dehydration stress was imposed.

Trehalose is known to maintain the oxidative balance and protect cells from stress mediated damage. Although trehalose accumulation was once thought to be restricted to only lower order plants and certain angiosperm resurrection plants, the identifcation of trehalose biosynthesis genes from many angiosperm plants suggested an active biosynthesis pathway. However, it is intriguing that irrespective of the presence of trehalose biosynthesis genes in plants, the accumulation in the tissues is very scarce or in untraceable amounts. The leaves of *A. thaliana* plants treated with 50 mM trehalose and 100 μ M Validamycin resulted in the accumulation of trehalose in the range of 1.5- 2 μ g g⁻¹ FW and has shown improved ROS- antioxidant balance and induction of stress responsive dehydrin genes under unstressed conditions in *A. thaliana* plants (Mohanan et al. [2023](#page-13-12)). Hence, similar concentrations were used in the current study to study the role of trehalose during drought stress. The presence of trehalose observed in the leaves both Vaisakh and Aiswarya cultivars suggests the operation of active trehalose metabolic pathway in rice cultivars (Fig. [1](#page-5-0)d). Further, compared to the stress sensitive cultivar, the signifcantly higher levels of trehalose observed in leaves of Vaisakh plants signifes the existence of diferences in the accumulation of trehalose in between stress tolerant and sensitive cultivar. Furthermore, the drought induced increase in trehalose levels in stress tolerant Vaisakh plants indicates the active role of trehalose under stress conditions. The trehalose content in stress sensitive Aiswarya plants is not increased during drought coinciding with its stress sensitive nature (Fig. [1d](#page-5-0)). The increase in leaf trehalose content upon treatment of the leaves with trehalose or validamycin A suggests active uptake of trehalose from external supplementation (Fig. 1d). Further, the higher accumulation of trehalose in validamycin A treated leaves than trehalose treated leaves suggests higher activity of trehalase in the plants (Goddijn et al. [1997;](#page-12-19) Müller et al. [2001\)](#page-13-16). Trehalase prevents the trehalose accumulation by breaking trehalose into two glucose molecules and validamycin A, due to structural similarity with trehalose shows higher affinity towards trehalase and inhibits its activity. In Arabidopsis, the trehalose accumulation gradually increased in their tissues with decreasing trehalase activity upon treatment of leaves with validamycin A (Müller et al. [2001\)](#page-13-16). The increase in trehalose accumulation post trehalose and validamycin A treatment helped the rice cultivars to show positive response during drought stress. The drought along with validamycin A and trehalose promoted higher trehalose accumulation in Vaisakh plants than Aiswarya. Like the drought tolerant Vaisakh, the lower trehalose accumulation still improved stress responses as inferred from the water status and the lower membrane damages in Aiswarya (Fig. [1](#page-5-0)b, c and d).

Trehalose biosynthesis is a two-step process where trehalose is formed from glucose-6-phosphate and UDP-glucose through an intermediate sugar, trehalose-6-phosphate. Two enzymes, trehalose-6-phosphate synthase (TPS) and trehalose-6-phosphate phosphatase (TPP) are involved in the formation of trehalose-6-phosphate from the precursors and for conversion of trehalose-6-phosphate to trehalose (Iordachescu and Imai, 2008). Heterologous expression of yeast trehalose biosynthesis genes or overexpression of plant trehalose metabolism genes provided greater protection to plants from stress mediated injuries along with improvement of photosynthesis. Further, the overexpression of rice TPP led to signifcant increase in yield and improvement in photosynthesis during unstressed and mild drought stressed conditions in maize plants (Nuccio et al. [2015](#page-13-23)). While the overexpression of AtTPPF in *A. thaliana* resulted in increased trehalose accumulation with higher survival rate, improved recovery after drought stress and reduced ROS accumulation, mutant plants lacking TPPF showed higher H_2O_2 accumulation in its leaves (Lin et al. [2019](#page-13-4)). In agreement with this, the trehalose accumulated leaves of tolerant rice cultivar showed lower H_2O_2 accumulation than that was observed during drought in untreated plants (Fig. [3](#page-6-1)a). Contrastingly, the trehalose accumulation triggered the H_2O_2 accumulation in stress sensitive plants. This may be due to the trehalose mediated ROS accumulation and associated signalling

(Fig. [3a](#page-6-1)). Similar to our observation, treatment of tobacco leaves with exogenously supplied trehalose triggered ROS accumulation (Shi et al. [2019\)](#page-13-24). Trehalose induced increase in antioxidant activities are also observed in plants during abiotic stress conditions. In *A. thaliana*, the SOD, APX and POD activities increased with trehalose treatment during salt stress. Several reports suggested that the increased ROS accumulation followed by an increase in antioxidant activities and other positive stress responses lead to improved stress tolerance (Garg and Manchanda, 2009). The presence of multiple SOD isoforms in leaves of rice plants and their activation under stress conditions was reported earlier. Similarly, increased APX activity along with the appearance of new isoforms in stress tolerant rice plants under stress conditions was also reported (Challabathula et al. [2022](#page-12-11)). Sugars and their metabolic enzymes could possibly interact with ROS and its signalling pathways which directly or indirectly enhances the drought stress tolerance by modulating ROS production (Couée et al. [2006](#page-12-23); Bolouri-Moghaddam et al. [2010\)](#page-12-24). Exogenous application of sucrose and trehalose resulted in increased expression of antioxidant enzymes during drought (Kaur et al. [2021](#page-12-10)). In *Triticum aestivum* L., supplementation of trehalose resulted in increased O_2 - and H_2O_2 scavenging by activation of SOD and CAT (Luo et al. [2008](#page-13-25)). The rice cultivars, Vaisakh and Aiswarya used in the current study showed diferences in their responses towards drought when they are accumulating trehalose. While the tolerant cultivar was equipped with better antioxidant system with increased activity by trehalose and stress, this was consistent even when the plants are unstressed. The higher activity of SOD and APX, might be the reason for these plants to have lower ROS during drought (Fig. [3b](#page-6-1)). Although similar response was observed in stress sensitive cultivar Aiswarya, increased ROS accumulation was observed during drought. These variations were also supported by the antioxidant gene expression where Vaisakh showed a higher Cu/Zn SOD and APX expression during trehalose treatment (Fig. [4](#page-8-0)a and b). This confrms the existence of diferential modulation in oxidative balance during drought in drought tolerant and sensitive rice cultivars. The tolerant cultivar accumulated higher levels of trehalose than sensitive cultivar under nonstress conditions indicating its capability of early sensing of stress and adjusting their defence strategies even before they experience the actual stress.

Photosynthesis is an important phenomenon that is known to be severely afected by drought stress. Diferences exist in the regulation of photosynthesis during drought in sensitive and tolerant cultivars. Compared to stress sensitive Aiswarya cultivar, the stress tolerant cultivar Vaisakh maintained a higher carbon assimilation and better gas exchange parameters at 1500 µmol photons m^{-2} s⁻¹ PPFD (Taj and Challabathula [2021;](#page-13-19) Challabathula et al. [2022\)](#page-12-11). Upon experiencing drought, plants minimize the gas exchange and transpiration

process by forcing the stomata to close. The stomatal closure decreases the photosynthesis due to unavailability of $CO₂$ and activates ABA as a mobile signal (Meyer and Genty [1998](#page-13-26); Flexas et al. 2004). The decline in $CO₂$ assimilation rates observed in both the cultivars during drought corresponds to lower stomatal conductance and lower internal CO2 levels (Challabathula et al. [2022;](#page-12-11) Fig. [6](#page-9-0)a-d). However, the induction in $CO₂$ assimilation rates upon treatment with trehalose and validamycin A in both the cultivars indicates the importance of trehalose for photosynthetic activity (Fig. [6a](#page-9-0)). The association between $CO₂$ assimilation rates and stomatal conductance reveals that the induction of $CO₂$ assimilation rates were higher in trehalose and validamycin A treated leaves of tolerant cultivar Vaisakh than stress sensitive Aiswarya cultivar (Fig. [6](#page-9-0)). Although the stomatal conductance was relatively higher in trehalose treated leaves of Aiswarya cultivar, the photosynthetic $CO₂$ assimilation rates were higher in Vaisakh cultivar. Lower stomatal density along with smaller stomatal size in drought tolerant cultivars exerts infuence on stomatal conductance. Direct association in between the increase in the intracellular trehalose levels to increase in photosynthetic carbon assimilation rates are observed in both the cultivars suggesting the signifcant role played by trehalose in increasing the photosynthesis (Fig. [6a](#page-9-0)). However, since stress tolerant cultivar accumulated more amounts of trehalose, the $CO₂$ assimilation rates were also higher. Increased Fv/Fm ratio was observed in maize and wheat plants treated with diferent concentrations of trehalose during heat stress indicating increased photosynthetic performance (Zhang et al. [2022](#page-14-1)). Similar responses were also observed in leaves of *Ocimum basilicum* L., (Zulfqar et al. [2021](#page-14-5)). It was also reported that the exogenous trehalose application could protect the photosystem II in winter wheat by promoting the cyclic electron flow during drought stress (Luo et al. [2021](#page-13-11)). Reports also suggested trehalose interacting with the sugar signalling pathways leading to the enhancement in photosynthesis (Oszvald et al. [2018\)](#page-13-27). The drought stress increased the Ci value and Ci/Ca ratio in both cultivars which was expected as a result of the mesophyll conductance (Fig. [6c](#page-9-0) and d). The trehalose accumulation comparatively reduced these parameters in drought stressed tolerant cultivar, indicating a lower $CO₂$ release caused due to the stress induced photorespiration. It is also noted that the trehalose mediates the increase in photosynthesis during drought in tolerant cultivar which might be due to comparatively higher chlorophyll pigments.

Trehalose accumulation signifcantly increased the chlorophyll content in unstressed plants and protected them from drought stress induced damages. Although both Vaisakh and Aiswarya cultivars have shown increased total chlorophyll content upon trehalose or validamycin A treatment, the content was higher in Vaisakh cultivar (Fig. [5](#page-8-1)). Trehalose help the plants to maintain their photosynthesis by protecting

the chloroplast ultrastructure, regulation of stomatal closure through ABA signalling, maintenance of water status, improvement in photosynthetic pigment concentration and improving the gas exchange parameters in unstressed and stressed conditions (Van Houtte et al. [2013](#page-13-15); Wang et al. 2020; Luo et al. [2021](#page-13-11)). Vaisakh plants being stress tolerant were able to maintain a lower water loss, better oxidative balance, chlorophyll content, and improved photosynthesis and minimized drought associated damages that helped them to extend their survival to prolonged drought conditions. Drought sensitive Aiswarya, on the other hand, is susceptible to drought, showed higher water loss leading to membrane damage, increased ROS which are not compensated by timely detection and employing of sufficient antioxidant machinery leading to an altered metabolism leaving lower photosynthetic performance (Challabathula et al. [2022\)](#page-12-11). Trehalose accumulation helps these cultivars in diferent ways to withstand drought, where the tolerant cultivar is supported by an improved root growth and lower oxidative damages. The trehalose accumulation counteracts the drought induced ROS accumulation with timely signalling and increasing the antioxidant enzyme activity along with a superior chlorophyll stability and gas exchange parameters. In sensitive cultivar, the trehalose accumulation may have role in ROS signalling which results in antioxidant defence but up to a certain extent of water loss.

Conclusions

The active trehalose metabolic pathway is operational in both drought sensitive and drought tolerant rice cultivars with diferential accumulation of trehalose. Compared to the stress sensitive cultivar, signifcantly higher levels of trehalose observed in leaves of drought stress tolerant Vaisakh plants emphasize the importance of trehalose for stress tolerance. Further, trehalose or validamycin A treatment led to increase in trehalose content in the leaves promoting drought tolerance. Compared to stress sensitive Aiswarya plants, drought stress and the treatment of leaves with validamycin A or trehalose has led to higher trehalose accumulation in Vaisakh plants. Trehalose accumulation helped tolerant cultivar, Vaisakh, to adjust towards drought by maintaining a higher water status and decrease the ROS by efective activation of antioxidant enzyme activity and antioxidant gene expression along with enhanced photosynthesis. Further, the multifaceted roles of trehalose in signaling, activation of stress responsive genes/proteins, growth and development of plants during stress conditions needs to be clearly deciphered.

Supplementary Information The online version contains supplementary material available at<https://doi.org/10.1007/s12298-023-01404-7>. **Acknowledgements** AM acknowledges the UGC fellowship, Rajiv Gandhi National Fellowship 2017-18 for supporting the research work through the Fellowship. DC acknowledges the SERB funded projects (NO/SB/EMEQ-299/2014) and (CRG/2021/005916) for facilities. The authors acknowledge the Central Instrumentation Facility of Department of Life Sciences for equipment and other common facilities.

Author contributions AM and DC designed research and experiments. AM, AK and DR performed the experiments, interpretation of data and manuscript writing was done by DC, BK and AM. Every author contributed to the article and approved the submitted version of manuscript.

Declarations

Conflict of interest The authors declare that they have no conficts of interest. We declare that we do not have any commercial or associative interest that represents a confict of interest in connection with the work submitted.

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