




Impact of postharvest exogenous γ -aminobutyric acid treatment on cucumber fruit in response to chilling tolerance

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Abstract Low-temperature storage is generally used to extend postharvest lifetime and to inhibit decay of cucumber fruit, but it also enhances the intensity of chilling injury. The capability of γ -aminobutyric acid to enhance antioxidant enzyme activities and reduce chilling injury was studied in cucumber (*Cucumis sativus* L.) fruit stored at 1 °C for 5 weeks. The purpose of this study was to define if the GABA-induced modification in antioxidant system and phospholipase activity is linked to the reduced chilling injury in cold-stored cucumber fruit. Alleviation of chilling injury by GABA treatment was related to increased content of proline, endogenous GABA and enhanced activities of CAT and SOD, together with reduced activities of PLC, PLD and LOX. We suggest that PLC, LOX and PLD are associated with chilling injury initiation by involvement in a signaling pathway and membrane deterioration. Therefore the results obtained in this study suggest GABA's potential for postharvest applications for reducing chilling injury symptom in cucumber fruit.

Keywords GABA · Cucumber fruit · Antioxidant enzyme · LOX · PLC · PLD

Abbreviations

AOS	Active oxygen species
APX	Ascorbate peroxidase (EC 1.11.1.11)
CAT	Catalase (EC 1.11.1.6)
CI	Chilling injury
EL	Electrolyte leakage
GABA	γ -Aminobutyric acid
LOX	Lipoxygenases (EC 1.13.11)
MDA	Malonyl-dialdehyde
PLC	Phospholipase C (EC 3.1.4.11)
SOD	Superoxide dismutase (EC 1.15.1.1)

Introduction

Horticultural plants are used for food, either as edible products, or for culinary ingredients, and for medicinal use. They are genetically very diverse group and play a major role in modern society and economy. They are considered an important component of traditional food, and are also central to healthy diets of urban population (Benjak et al. 2005; Celik et al. 2007; Ercisli et al. 2010; Rop et al. 2014; Canan et al. 2016; Hricova et al. 2016; Zorenc et al. 2016).

Chilling injury (CI) is a physiological disturbance that restricts the storage of chilling-sensitive cucumber fruit at low temperatures (Lurie and Crisosto 2005). Two main assumptions, which are not pairwise exclusive, have been forwarded to clarify the injury effect of low temperature in this fruit. The first assumption proposes that chilling results in cell membrane stabilizations, which induces inhibition of membrane-bound enzymes or carrier activities (Wolfe 2006; Zhang et al. 2010). This stability is dependent on the fatty acid composition of the membranes. Some evidence in favor of the membrane rigidification hypothesis has been

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found (Nishida and Murata 1996; Promyou et al. 2008), but other reports found no obvious relationship between membrane fatty acid compound and CI sensitivity (Prasad 2001; Wongsheree et al. 2009; Sirikesorn et al. 2013). The second assumption refers to generation of active oxygen species (AOS). AOS consist of superoxide, hydroxyl radicals and hydrogen peroxide (H₂O₂). AOS rapidly react with different molecules, including proteins and DNA, and cause membrane lipid peroxidation (Rice-Evans et al. 1997). This leads to cellular damage or cell death (Blokhina et al. 2003). In presence of transition metal ions (such as Zn, Cu and Fe) hydrogen peroxide may react with superoxide radical (O₂⁻) to hydroxyl radicals (OH). Among the primary produced radicals, the hydroxyl radicals are the most reactive and hence the most noxious (Apel and Hirt 2004).

GABA (γ-Aminobutyric acid) is a non-protein amino acid, conserved from bacteria through yeasts to vertebrates. GABA was detected in plants almost 60 years ago (Malekzadeh et al. 2012). As an intrinsic signal molecule, GABA is very useful in adjustment of stress reactions (Kinnersley and Turano 2000; Malekzadeh et al. 2014).

GABA synthesis system regulates H⁺ in cytosol, functioning as a pH-stat (Sawaki et al. 2009). GABA can mitigate oxidative damage induced by aluminium and proton stresses on barley seedlings (Song et al. 2010). Shi et al. (2010) reported that GABA regulates expression of genes involved in generation of H₂O₂ in *Caragana intermedia* roots under salinity. Exogenous GABA mitigates CI in tomato and wheat plants under chilling stress (Malekzadeh et al. 2012, 2014). A deeper understanding of GABA in decreasing CI needs further investigation.

CI affects cell membrane integrity (Rui et al. 2010). Lipid peroxidation is known to be responsible for damage to cell membrane integrity which can be measured by the malonyl-dialdehyde (MDA) generation (Wise and Naylor 1987). The peroxidation of membrane fatty acids generates MDA and its level is utilized as a marker of oxidative stress. A rise in MDA production indicates the occurrence of lipid peroxidation, which results in damage to the cell membrane (Hodges et al. 1999). MDA and electrolyte leakage (EL) content are popular physiological markers of membrane lipid peroxidation and loss of membrane semi-permeability. These markers are used to indirectly calculate cell membrane integrity (Malekzadeh et al. 2014).

This study aimed to evaluate whether GABA-affected changes in the antioxidant system and MDA, proline and EL content are linked to the enhanced tolerance to CI in cold-stored cucumber fruit.

Materials and methods

Plant material and treatments

Hydroponic cluster cucumber (*Cucumis sativus* L.) was obtained from a commercial greenhouse in Tehran, Iran. Fruits were harvested and immediately transported to the laboratory. The fruit surface was disinfected in 0.1% sodium hypochlorite solution for 2 min, thoroughly washed in tap water and air-dried at 25 °C prior to use. Based on previous studies (Kinnersley and Turano 2000; Deewatthanawong et al. 2010a, b; Shang et al. 2011; Malekzadeh et al. 2012, 2014), the first group was immersed in 5 mM solution for 10 min, whereas the second group of fruit was soaked in sterile deionized water for 10 min and served as controls. All fruits were then air-dried for approximately 30 min and stored at 1 °C and 80–90% relative humidity (RH). Samples were collected from fruit after 3 or 5 weeks of storage at 1 °C for measurements of levels of CI, EL, H₂O₂, MDA, GABA and proline content, and the activities of SOD, CAT, APX, LOX, PLC and PLD. Each experiment was replicated three times.

Untreated and treated fruits were sampled on day 0, after 3 and 5 weeks of cold storage. Equatorial slices of sampled fruit were diced, frozen in liquid nitrogen and stored at –80° C for enzyme analysis. For CI evaluation, fruit of each treatment was sampled weekly from cold storage and held at 20 °C for 3 days.

CI index

The degree of CI was visually assessed on the mesocarp surface, following a double cut parallel to the axial diameter (Ding et al. 2002). The extent of flesh browning was divided into 4 classes: 0 = no browning; 1 = browning covering < 25% of the fruit surface; 2 = browning covering < 50%, but > 25% of surface; 3 = browning covering > 50%. The average extent of cold damage was expressed as a CI index calculated using the following formula:

$$CI\ index\ (\%) = \frac{\sum[(CI\ level)(Number\ of\ fruit\ at\ CI\ level)]}{Total\ number\ of\ fruit\ in\ the\ treatment} \times 100$$

Determination of MDA and proline contents

MDA content was measured by the thiobarbituric acid method described by Chen et al. (2008). The MDA concentration was calculated according to the following formula:

$$\text{MDA } (\mu\text{mol g}^{-1} \text{FW}) = 6.45 \times (\text{A532} - \text{A600}) 0.56 \times \text{A450}$$

where A532, A600 and A450 represent the absorbance of the mixture at 532, 600, and 450 nm, respectively.

Proline content was measured using the acid-ninhydrin method described by Shan et al. (2007). Proline content was expressed as μg proline g^{-1} FW.

Electrolyte leakage

The rate of electrolyte leakage was measured according to the method described by Chen et al. (2008) with modification. Cylinders of cucumber mesocarp tissue were excised with a 10-mm diameter stainless steel cork borer. Two pieces of 4 mm thickness were cut from each cylinder. After being rinsed three times (2–3 min) with deionized water, 10 pieces were put into 50 mL of deionized water and shaken at 100 cycles per min for 30 min. Conductivity was measured using a Conductance Bridge (DDS-11A, Yamei Electron Instrument Factory, Hangzhou, China). Total conductivity was obtained after keeping the flasks boiling for 10 min, and electrolyte leakage was expressed as percentage of total conductivity.

Enzyme assays

The activity of Superoxide dismutase was determined using the slightly modified method of Xu et al. (2008). Fresh samples (0.5 g) were rapidly extracted in a pre-chilled mortar on an ice bath with 5 mL of ice-cold phosphate buffer (100 mM, pH 7.8) containing 1 mM EDTA and 5% (w/v) PVP. After centrifugation at $10,000 \times g$ for 30 min at 4 °C, the supernatant was used for SOD analysis. One hundred μL of the enzyme extract was mixed with 2.5 mL of 100 mM phosphate buffer (pH 7.8), 75 μL of 55 mM methionine, 300 μL of 0.75 mM nitroblue tetrazolium (NBT) and 60 μL of 0.1 mM riboflavin in a test tube. The test tubes containing the reaction solution were irradiated under 2 fluorescent light tubes ($40 \mu\text{mol m}^{-2} \text{s}^{-1}$) for 10 min. The absorbance measured at 560 nm with a UV/visible spectrophotometer (HACH, USA). Blanks and controls were run in the same manner, but without illumination and enzyme, respectively. One unit of SOD activity was determined by the method of Rao et al. (1996). The reaction medium contained 50 mmol L^{-1} sodium phosphate buffer (pH 7.8), 14 mmol L^{-1} methionine, 3 $\mu\text{mol L}^{-1}$ EDTA, 1 $\mu\text{mol L}^{-1}$ nitroblue tetrazolium (NBT), 60 $\mu\text{mol L}^{-1}$ riboflavin and 0.1 mL of SOD extract. The formation of blue formazan was monitored by recording the absorbance at 560 nm. One unit of SOD activity was defined as the amount of enzyme that caused 50% inhibition of NBT.

CAT activity was assayed according to the method of Chance and Maehly (1955). This involved monitoring the

disappearance of H_2O_2 by recording the decrease in absorbance at 240 nm of a reaction mixture containing 50 mmol L^{-1} sodium phosphate buffer (pH 7), 12.5 mmol L^{-1} H_2O_2 and 20 μL of CAT extract. One unit of CAT activity was defined as the amount of enzyme that decomposed 1 $\mu\text{mol H}_2\text{O}_2 \text{ min}^{-1}$ at 30 °C. Ascorbate peroxidase measurement was adapted from the method of Vicente et al. (2006). 1 g frozen tissue was ground with 5 mL of 50 mM sodium phosphate buffer (pH 7.0), containing 0.1 mM EDTA, 1 mM ascorbic acid and 1% polyvinylpyrrolidone (PVP). The homogenate was centrifuged at $10,000g$ for 20 min at 4 °C and the supernatant was used to determine APX activity. One unit of APX activity is defined as the amount of enzyme that oxidized 1 μmol ascorbate per minute at 30 °C.

LOX activity was assayed using the method of Todd et al. (1990). The standard assay mixture contained 200 μL of Tween 20 and 40 μL of linoleic acid (Aldrich, Milwaukee, WI, USA) in 40 mL of 0.1 M sodium phosphate buffer (pH 7). To 1 mL of standard assay mixture in a cuvette, 0.2 mL of LOX extract was added. One unit of LOX activity was defined as the amount of enzyme that caused an increase in absorption at 234 nm of 0.01 min^{-1} at 25 °C with linoleic acid as substrate.

PLC and PLD activities were determined by the procedures of Kurioka and Matsuda (1976) and Gupta and Wold (1980) respectively using D-nitrophenylphosphorylcholine (NPPC; Aldrich) as substrate. The reaction mixture for PLC consisted of 1 mL Tris-HCl buffer (0.25 mol L^{-1} , pH 7.2 containing 20 mmol L^{-1} NPPC and 600 g L^{-1} D-sorbitol), and 0.3 mL of enzyme extract. For PLD, 0.9 mL Ca-acetate (50 mmol L^{-1} , pH 5.6) containing 27.4 mM NPPC was mixed with 0.1 mL (0.4 U) of acid phosphatase (Aldrich) dissolved in 50 mM Ca-acetate (pH 5.6) along with 0.3 mL of enzyme extract. In both cases, after incubation for 60 min at 37 °C, 0.1 mL of 50 mM NaOH was added and the D-nitrophenol content was determined at 400 nm. One unit of PLC or PLD activity was defined as the amount of enzyme that catalyzed the formation of 1 nmol D-nitrophenol h^{-1} .

H_2O_2 analysis

For H_2O_2 determination, 2 g of frozen tissue was homogenized with 5 mL of chilled pure acetone and centrifuged at $10,000 \times g$ for 20 min at 4 °C. The supernatant was collected for H_2O_2 analysis by a method based on titanium oxidation (Patterson et al. 1984). The acetone extract (supernatant) was mixed with hydrochloric acid containing 200 mL L^{-1} TiCl_4 and 17 mol L^{-1} ammonia solution. The precipitate was washed with acetone and dissolved in 2 M sulfuric acid for spectrophotometric measurement at 410 nm. H_2O_2 concentration was determined from a standard graph (from 5 to 1 mM

H₂O₂, constructed by direct addition of H₂O₂ to the titanium solution) and expressed as nmol g⁻¹ FW (Cao et al. 2009).

GABA determination

The GABA concentration in fruit pulp was estimated by the method of Zhang and Bown (1997). GABA was determined on the basis of the increase in A₃₄₀ after 30 min following supplier recommendations for commercially available of GABase (Aldrich), a spectrophotometric-coupled enzyme assay system for GABA. The resulting values were compared with a standard curve constructed using known amounts of GABA and expressed as µg GABA/g fresh weight (FW).

Statistical analysis

Five cucumber fruits from each treatment were taken at each sampling. All the assays were carried out in triplicate. Data analyses were performed using SPSS software version 16. Results were analyzed using one-way analysis of variance (ANOVA) followed by Tukey's multiple comparison test. Correlation between various parameters was also investigated. Significance was determined at $p < 0.05$ and the results were expressed as mean values and standard error (SE) of the means.

Results

According to ANOVA results, the interaction effects of week and GABA treatments were significant for all the traits (Table 1).

Effect of GABA treatment on CI in cucumber fruit

Index of CI in cucumber fruit increased during storage period, this increasing was delayed when treated with

exogenous GABA application. Chilling injury symptoms after 10 days storage at 1 °C were visible in cucumber fruit. Significant difference(s) in the incidence of CI between control fruit and GABA-treated fruit were observed (Fig. 1). Exogenous GABA treatment reduced CI symptoms and maintained quality of fruit (Fig. 2).

Levels of MDA and Electrolyte leakage in cucumber fruit

MDA and electrolyte leakage content in cucumber fruit increased during storage (Fig. 2). Levels of MDA and EL in control fruit were higher than that of GABA-treated fruit during storage. GABA-treated fruit had 23% less EL than control fruit after 3 weeks of storage. In line with this decreased EL content, the amount of MDA in GABA-treated fruit was 34% less than control fruit after 5 weeks of storage. Exogenous GABA treatment reduced MDA content and EL in cucumber fruit during storage.

Effect of GABA treatment on proline content in cucumber fruit

Proline content increases in both control as well as GABA treated fruits (Fig. 2). The proline content in the GABA-treated fruit was significantly ($p < 0.05$) higher than in control fruit. Also, the proline content in GABA-treated fruit was 168.7% higher than that in control after storage for 5 weeks (Fig. 2).

Effect of GABA treatment on SOD, APX and CAT activities and H₂O₂ content in cucumber fruit

As shown in Fig. 3, activity of ROS-scavenging system in cucumber significantly affected by postharvest GABA treatment during storage at 1 °C.

H₂O₂ content and SOD activity increased during storage, while APX and CAT activities decreased. GABA

Table 1 Analysis of variance (ANOVA) of the effect of week and GABA on some growth, enzymes, biochemical and mineral nutrient parameters cucumber fruit

Source	df	CI	EL	Prolin	MDA	H ₂ O ₂	CAT
Week	2	200.1**	3.5**	87.6**	35.8**	126.8**	136.2**
GABA	1	33.4**	5.7**	58.8**	18.33	40.9**	89.2**
Week* GABA	2	9.2**	2.3**	14.0**	3.0**	16.3**	17.9**
CV %		78.94	35.96	42.6	40.71	54.58	33.68
	df	SOD	APX	PLC	PLD	LOX	GABA content
Week	2	954.5**	214.1**	152.1**	561.8**	47.2**	309.7**
GABA	1	63.8**	1.9**	43.4**	89.3**	46.0**	210.3**
Week* GABA	2	10.1**	1.3*	12.8*	27.9**	16.3**	62.7**
CV %		53.90	31.53	40.38	28.23	30.11	41.87

*, ** Effects significant at probability of 0.05 and 0.01 respectively; ns non-significant ($p > 0.05$)



Fig. 1 Appearances and transects of cucumber fruits stored at 1 ± 0.5 °C for 5 weeks. Fruits were treated with 5 mM GABA for 10 min and then stored. The fruit in the control was stored without GABA treatment. Appearances (a) and transects (b)

treatment resulted in increased SOD and CAT activities, inhibited the increase in H_2O_2 levels and had no significant impact on APX activity when compared to control cucumber fruit (Fig. 3).

The results indicate that, SOD activity in the cucumber fruit increased with storage period and reached the peak after 5 weeks of storage at 1 °C.

SOD activity in GABA-treated cucumber fruit was 19% higher than in the control cucumber fruit at 5 weeks of storage period.

Exogenous GABA treatment maintained higher CAT activity compared with the control fruits over the whole storage period.

Although CAT decreased with storage time, the activity in GABA-treated cucumber fruit was 63% higher than control at 5 weeks of storage.

APX activity significantly decreased during the storage period, reduced to half of the 0 day after 5 weeks of storage with no significant difference between GABA-treated and control fruit.

Levels of H_2O_2 in cucumber fruit enhanced gradually during storage. However, the rise in H_2O_2 level was significantly slower with GABA treatment. As could be seen in Fig. 3, the content of H_2O_2 in GABA-treated fruit was

26 and 37% lower than that of control fruit at week 3 and 5, respectively.

Effect of GABA treatment on Lipoxygenase, PLC and PLD activities in cucumber fruit

LOX, PLC and PLD activities increased during storage (Fig. 4). Like SOD, these activities were also much lower in GABA treatment than control. LOX, PLD and PLC activities in exogenous GABA-treated fruit were 38, 20 and 28% lower than that of control fruit at week 5 of storage, respectively.

Discussion

GABA levels increase rapidly in response to environmental stress in plants, which in turn, strengthens the defense system of plants. Increase in GABA content was also observed due to low oxygen and high CO_2 storage in strawberry (Deewatthanawong et al. 2010a) and tomato (Deewatthanawong et al. 2010b). GABA concentrations in soybean leaves increased 20–40 folds within 5 min of cold-treatment (Wallace et al. 1984). Chilling injury symptoms

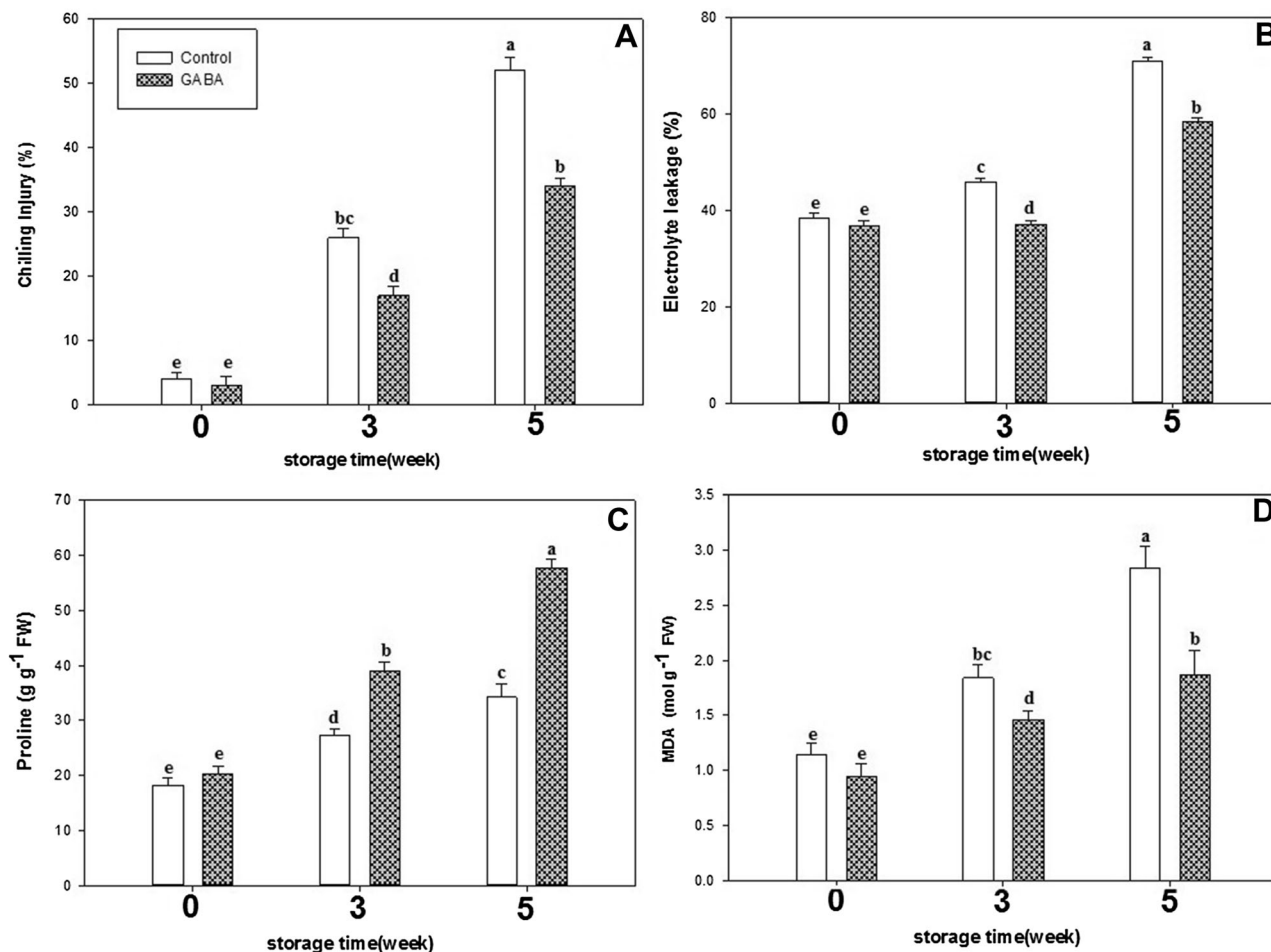


Fig. 2 The effects of exogenous GABA on CI (a), electrolyte leakage (b), proline (c) and MDA (d) content in cucumber fruit during storage period. Vertical bars represent standard error of the means. Different letters indicate statistically different means at $p < 0.05$

of wheat and tomato seedlings during 15 days of storage at 2 °C, were alleviated by exogenous GABA treatments (Malekzadeh et al. 2012, 2014). In this study, treatment of 5 mM exogenous GABA alleviated symptoms of chilling in cucumber fruit stored at 1 °C for 5 weeks and delayed CI percent increase (Fig. 2).

A significant increase in EL in cucumber tissue exposed to low temperatures is a good marker of cold stress. The results shown that the amount of electrolyte leakage significantly increased in cucumber fruit after 3 weeks (Fig. 2). Accompanying this rapid rise in EL of control fruit, the increase was delayed in GABA-treated cucumbers. Environmental stresses induce reactive oxygen species (ROS) generation that causes cell oxidative damage and lipid peroxidation, so that MDA (a final product of lipid peroxidation) can be used as an assay of cell oxidative damage (Xu et al. 2009). In this study, the change in level of MDA and electrolyte leakage shared similar patterns. The accumulation of MDA under chilling stress resulted in lipid peroxidation (Fig. 2). These results revealed that

chilling condition might mediate catabolic reactions targeting cell membranes.

To limit damage to cells by reactive oxygen species, plant cells employ the reactive oxygen species (ROS) scavenger system. In a previous study, Chongchatuporn et al. (2013), observed that overall chilling injury induced enhancement in the activities of APX, SOD and CAT and the content of MDA, H₂O₂ and EL in mango fruit.

Our finding also revealed that, level of MDA content was increased and SOD activity declined in fruit. Further, the results show that GABA treatment significantly delayed the MDA increase in cucumber fruits (Fig. 1), together with significant increase in the activity of SOD (Fig. 2). Therefore, the results suggest that the peripherally administered GABA can scavenge ROS and protect the tissue against active carbonyl harm.

Apel and Hirt (2004) reported that, the generation of O₂⁻ catalyzed by several enzymes including LOX and NADPH oxidase. The application of GABA resulted in lower activity of LOX (Ding et al. 2007; Hatamnia et al. 2016).

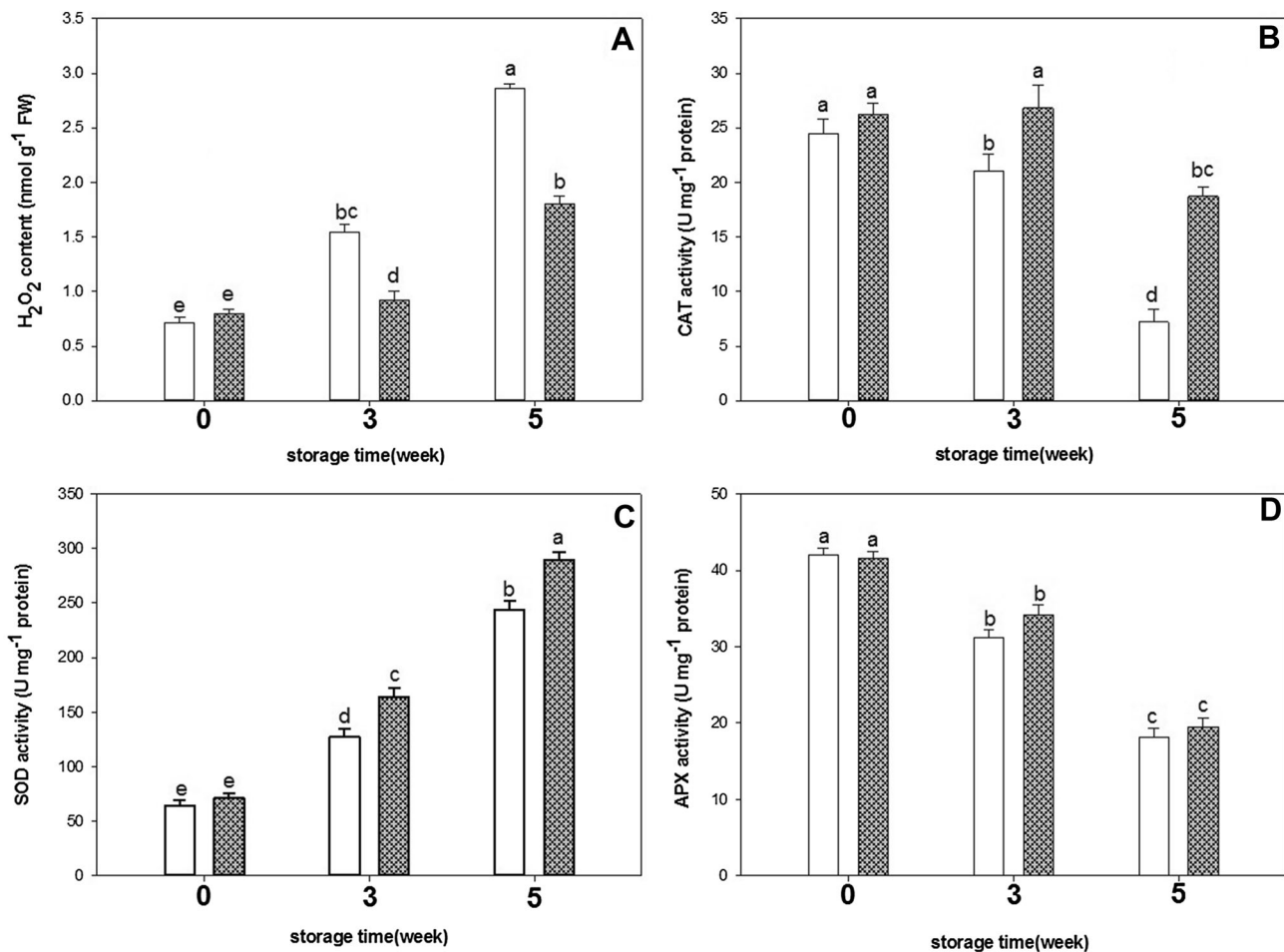


Fig. 3 The effects of exogenous GABA on H₂O₂ content (a), CAT (b) SOD (c) and APX (d) activities in cucumber fruit during storage period. Vertical bars represent standard error of the means. Different letters indicate statistically different means at $p < 0.05$

Our results also show that, the activity of SOD, the O₂⁻ scavenging enzyme, did increase by GABA treatment. SOD has been demonstrated as a target of ROS (Qin et al. 2009). The effect of GABA treatment on H₂O₂ may attribute to the GABA-induced higher activity of SOD.

Membrane damage during stress condition is initiated by a lipolytic cascade, with PLD and LOX being critical to the phospholipid catabolism (Bargmann et al. 2009). PLD-mediated hydrolysis increased under environmental stresses such as chilling and salt stresses. The higher activities of PLD and LOX were shown to be associated with their gene expression (Zhao et al. 2010; Malekzadeh 2015) and increases of these two enzyme activities were accompanied by the induction of CI (Mao et al. 2007; Rui et al. 2010). LOX plays an important role in peroxidative damage in membrane lipids in plants. Lower LOX activity was shown to be related to chilling tolerance of plants as membrane fluidity and lipid unsaturation was enhanced by LOX (Lee et al. 2005).

PLD, PLC and LOX activities induced by chilling might stimulate corresponding physiological reactions related to deterioration and senescence during the storage period.

In this study also, development of chilling injury was accompanied by an increase in PLC, PLD and LOX activities, which reached the maximum at week 5 at 1 °C. Activities of PLC, PLD and LOX increased during storage, but to a lesser extent in GABA-treated cucumbers suggesting that chilling resistance was enhanced by exogenous GABA application probably due to inhibition of these enzymes.

In summary, results of this investigation showed that GABA treatment can improve the chilling-tolerance of cucumber fruit by improving antioxidant enzyme activities, and decreasing the level of ROS, thus protecting membranes from chilling stress. Together with other studies (Trobacher et al. 2013; Yu et al. 2014), it is suggested that the exogenous GABA treatment can prove to be a promising and safe postharvest technology for increasing

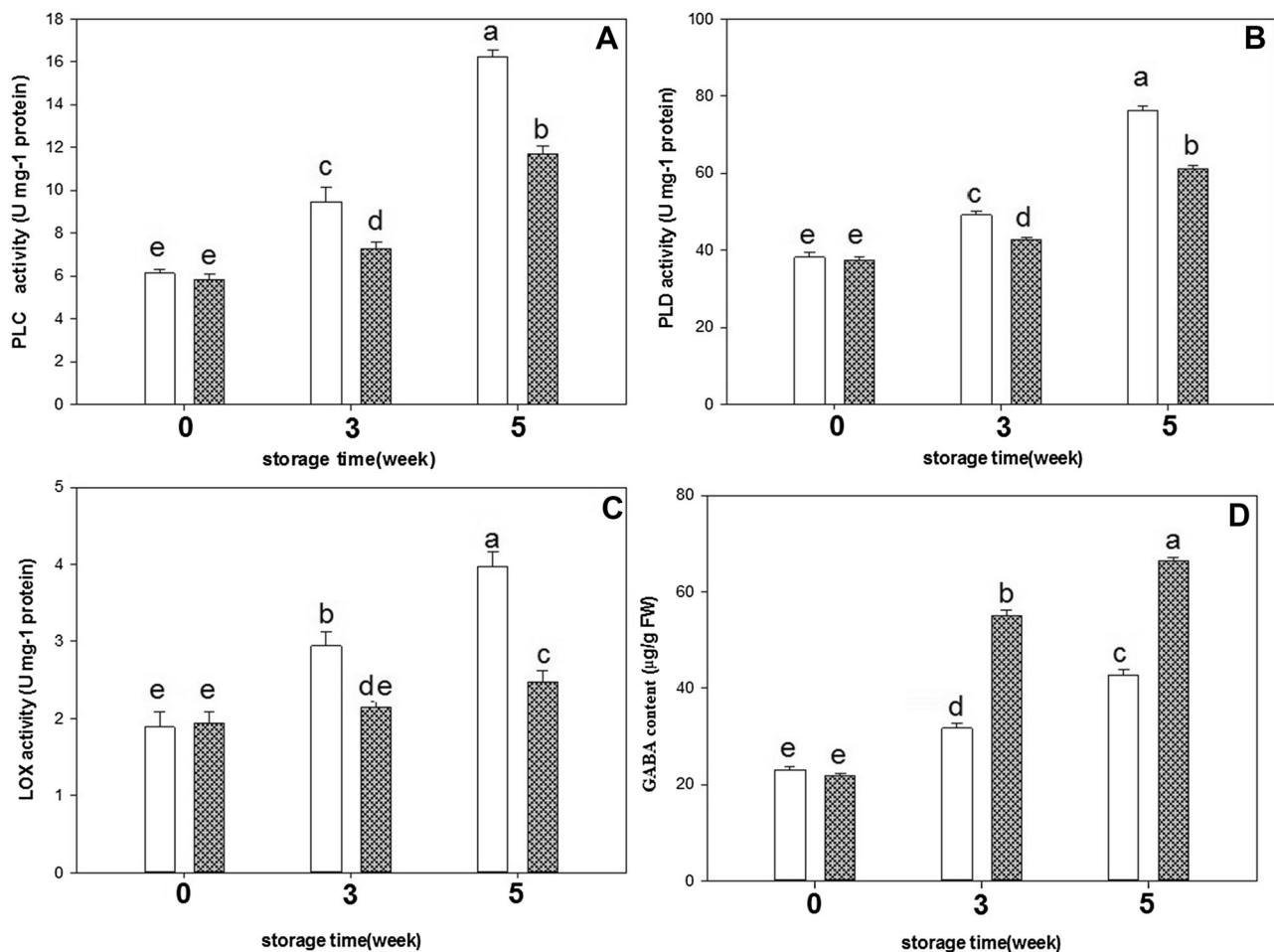


Fig. 4 The effects of exogenous GABA on PLC (a), PLD (b), LOX (c) activities and endogenous GABA content (d) in cucumber fruit during storage period. Vertical bars represent standard error of the means. Different letters indicate statistically different means at $p < 0.05$

the shelf life of harvested cucumber fruit and thereby maintaining its quality. Further studies are needed to elucidate the role of exogenous GABA in chilling tolerance at the molecular level.

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