



Combination therapy involving HSP90 inhibitors for combating cancer: an overview of clinical and preclinical progress

Yajun Liu¹ · Chenyao Li² · Hongwei Liu^{3,4} · Shutao Tan⁵

Received: 20 October 2023 / Accepted: 26 March 2024 / Published online: 17 April 2024
© The Pharmaceutical Society of Korea 2024

Abstract

The molecular chaperone heat shock protein 90 (HSP90) regulates multiple crucial signalling pathways in cancer by driving the maturation of key signalling components, thereby playing a crucial role in tumorigenesis and drug resistance in cancer. Inhibition of HSP90 results in metastable conformational collapse of its client proteins and their proteasomal degradation. Considerable efforts have been devoted to the development of small-molecule inhibitors targeting HSP90, and more than 20 inhibitors have been evaluated in clinical trials for cancer therapy. However, owing to disadvantages such as organ toxicity and drug resistance, only one HSP90 inhibitor has been approved for use in clinical settings. In recent years, HSP90 inhibitors used in combination with other anti-cancer therapies have shown remarkable potential in the treatment of cancer. HSP90 inhibitors work synergistically with various anti-cancer therapies, including chemotherapy, targeted therapy, radiation therapy and immunotherapy. HSP90 inhibitors can improve the pharmacological effects of the above-mentioned therapies and reduce treatment resistance. This review provides an overview of the use of combination therapy with HSP90 inhibitors and other anti-cancer therapies in clinical and preclinical studies reported in the past decade and summarises design strategies and prospects for these combination therapies. Altogether, this review provides a theoretical basis for further research and application of these combination therapies in the treatment of cancer.

Keywords HSP90 inhibitors · Cancer therapy · Drug combination · Synergistic effect · Drug resistance

Introduction

Heat shock proteins (HSPs) are a family of evolutionarily conserved molecular chaperons. They are responsible for maintaining protein homeostasis and play an essential role in preventing misfolding and aggregation of proteins, stabilising the conformation of intracellular proteins, assisting the transmembrane transport of proteins, mediating the transport of proteins to target organelles, driving the assembly and degradation of oligomeric structures and degrading irreversibly damaged proteins in cells (Lang et al. 2021).

HSP90 is one of the most active molecular chaperones in cells. The HSP90 family has four members that are localised in different organelles. Inducible HSP90 α and constitutive HSP90 β are located in the cytosol, whereas GRP94 and TRAP1 are located in the endoplasmic reticulum (ER) and mitochondria, respectively. HSP90 contains three conserved regions as follows: (1) the N-terminal domain (NTD) containing an ATP-binding pocket known as the ‘Bergerat fold’ and a subdomain that binds co-chaperones, such as p50 and cdc37 (Smith et al. 2015); (2) the C-terminal domain (CTD)

✉ Yajun Liu
yjliu85@dlut.edu.cn

✉ Hongwei Liu
hwliu1425@163.com

✉ Shutao Tan
tanst@sj-hospital.org

¹ School of Pharmaceutical Engineering, Shenyang Pharmaceutical University, Wenhua Road 103, Shenyang 110016, China

² School of Life and Pharmaceutical Sciences, Dalian University of Technology, Dagong Road 2, Panjin 124221, China

³ Department of Head and Neck Surgery, Liaoning Cancer Hospital and Institute, Shenyang 110042, China

⁴ Affiliated Cancer Hospital of Dalian University of Technology, Shenyang 110042, China

⁵ Department of Urology, Shengjing Hospital of China Medical University, Sanhao Street 36, Shenyang 110004, China

containing subdomains that form HSP90 dimers and sites for interaction with other co-chaperones (Donnelly and Blagg 2008); (3) the middle domain (MD), which is a stretch of highly charged amino acid sequences connecting the NTD and CTD (Ali et al. 2006).

Although HSP90 inhibitors hold great promise in the treatment of cancer, clinical trials have not demonstrated satisfactory results. Monotherapy with HSP90 inhibitors leads to complications such as organ toxicity and acquired drug resistance (Erlichman 2009). In recent years, combination therapy involving HSP90 inhibitors has emerged as an important strategy for treating cancer (Ren et al. 2022). In this review, we summarised the typical examples of the combined use of HSP90 inhibitors and other anti-cancer therapies in both clinical trials and preclinical experiments reported in the past decade. In addition, we discussed the rationale for using HSP90 inhibitors to increase the efficacy of other anti-cancer therapies and overcome drug resistance. These anti-cancer therapies include traditional chemotherapy, targeted therapy, radiotherapy and immunotherapy.

Role of HSP90 in cancer

HSP90 has received substantial attention from scholars owing to its role in the prevention of misfolding and degradation of several mutated and overexpressed oncoproteins. Consequently, it serves as a critical factor for oncogene addiction and cancer cell survival (Whitesell and Lindquist 2005; Hoter et al. 2018). HSP90 is responsible for stabilising and activating more than 400 client proteins that are involved in signal transduction, hormone response and transcriptional regulation in cells (Zhao et al. 2005; Schopf et al. 2017). The updated list of HSP90 clients is available on the Picard lab website (<https://www.picard.ch/downloads/Hsp90interactors.pdf>). The client proteins of HSP90 mediate tumour cell proliferation and survival as well as tumour angiogenesis, invasion and metastasis and are involved in the evolution of all hallmarks of cancer (Miyata et al. 2013). Cancer cells are vulnerable to proteotoxic stress, partly because the accumulation of mutant molecules may lead to cell death (Bagatell and Whitesell 2004). Under proteotoxic stress conditions, the transcription factor heat shock factor 1 (HSF-1) dissociates from HSP90 and initiates the transcription and expression of HSPs. Cancer cells require higher expression levels of HSP90 to govern protein homeostasis. HSP90 is upregulated in various cancers. Plasma HSP90 has been identified as a biomarker for the diagnosis of hepatocellular carcinoma and assessment of treatment efficacy (Wei et al. 2020). In addition, HSP90 has been associated with the aggressiveness or metastatic potential of cancer cells and regulates the immune response in cancer (Snigireva et al. 2014; Graner 2016).

Inhibition of HSP90 leads to the degradation of cancer-related client proteins via the ubiquitin–proteasome system, consequently disturbing multiple signalling pathways, and induces the apoptosis of cancer cells. The discovery of HSP90 inhibitors can be traced back to the identification of geldanamycin, a natural antibiotic, which competitively binds to the ATP-binding pocket in the NTD of HSP90 (Whitesell et al. 1994). When HSP90 encounters geldanamycin, the client protein dissociates from the HSP90 complex, and the chaperone cycle halts. Tanespimycin (17-AAG), a geldanamycin semi-synthetic derivative, was the first HSP90 inhibitor to enter clinical trials in 1999. In the past two decades, significant attempts have been made to identify HSP90 inhibitors with good pharmacodynamic and pharmacokinetic properties and acceptable safety profiles. To date, clinical trials have evaluated the therapeutic role of more than 20 HSP90 inhibitors in cancer. The discovery of HSP90 inhibitors has been well summarised and discussed in some excellent reviews (Li et al. 2020, 2021; Xiao and Liu 2020; Serwetnyk and Blagg 2021; Yu et al. 2022). The chemical structures of representative HSP90 inhibitors are shown in Fig. 1. HSP90 inhibitors used as monotherapy in clinical trials are summarised in Table 1. The data were retrieved from *clinicaltrials.gov*, and only trials that have been completed are presented.

Combination therapy involving HSP90 inhibitors in cancer treatment

In the past several years, numerous studies have attempted to evaluate the potential of combination therapy with HSP90 inhibitors and other anti-cancer therapies. A total of 28 clinical trials on 8 HSP90 inhibitors combined with other therapies are summarised in Table 2. Targeted small-molecule drugs represent the major class of drugs that are combined with HSP90 inhibitors. Other drugs include traditional chemotherapeutics, biological therapeutics and hormones.

HSP90 inhibitors can synergistically act with other anti-cancer drugs in different ways to enhance treatment outcomes. First, HSP90 inhibitors can degrade anti-cancer biological targets given that they are clients of HSP90. Second, HSP90 inhibitors block the cooperative signalling pathways and accelerate the apoptosis of cancer cells. Third, HSP90 inhibitors delay DNA repair, which may improve the therapeutic efficacy of DNA-targeting anti-cancer drugs. Fourth, HSP90 inhibitors can reverse immunosuppression in cancer cells and enhance immunotherapies. HSP90 inhibitors have been widely used to overcome drug resistance (Jhaveri and Modi 2012). Effective cancer therapy is often hampered by drug resistance during treatment (Vasan et al. 2019). Overexpression or mutation of biological targets is a major cause of resistance to conventional or targeted anti-cancer

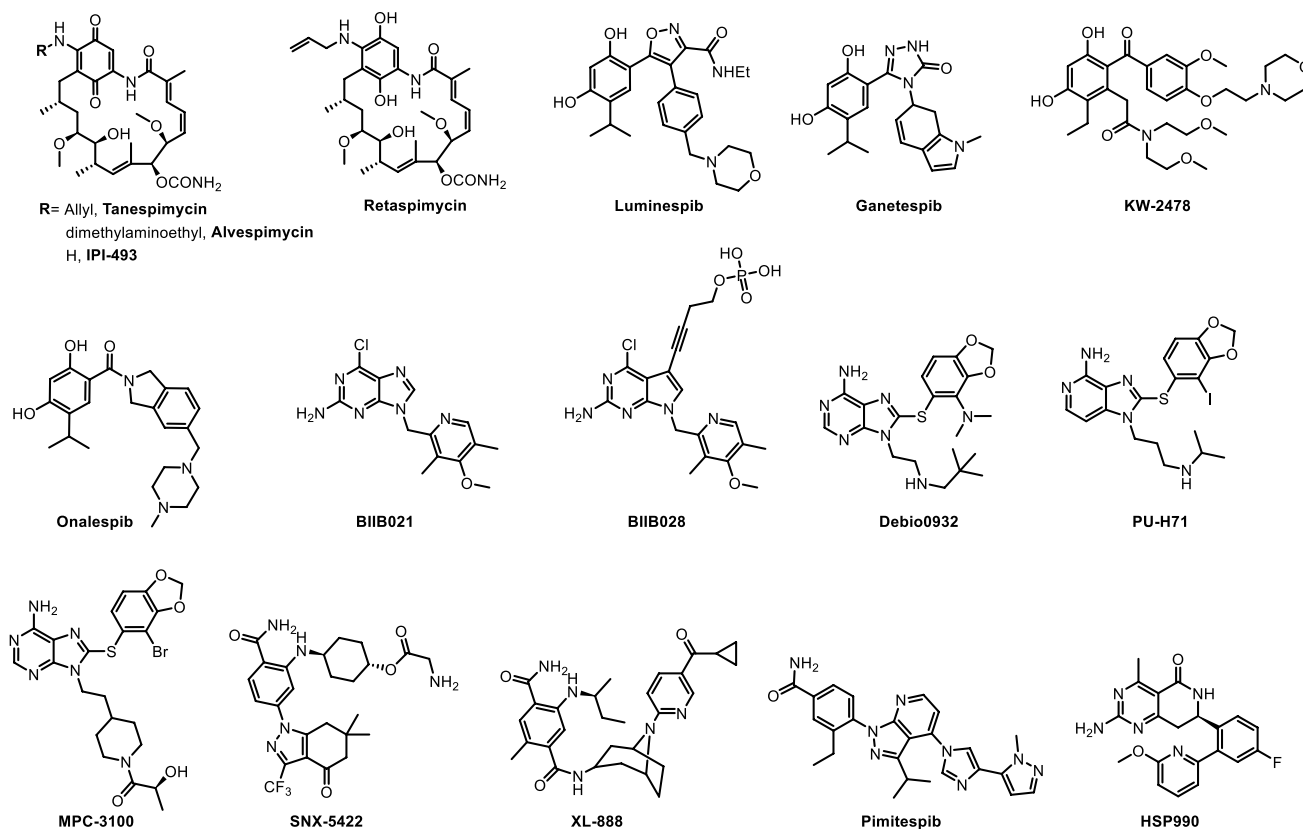


Fig. 1 Representative HSP90 inhibitors

drugs in many human cancers. Because many biological targets are clients of HSP90, inhibition of HSP90 leads to their degradation irrespective of whether they are mutated. The client proteins of HSP90 involved in drug resistance are well summarised in a review by Vergnaud-Gauduchon (Mathieu et al. 2019). Activation of bypass signalling often impairs the action of anti-cancer drugs, especially kinase inhibitors. HSP90 inhibitors can disrupt compensatory signalling pathways and restore the therapeutic potential of anti-cancer drugs. In addition, some drug resistance-related proteins in cancer cells have been identified as clients of HSP90; therefore, their depletion by HSP90 inhibitors is considered an effective strategy for overcoming the corresponding resistance.

Combination with HSP90 inhibitors in clinical trials

Pimitespib (TAS-116)

Pimitespib is an orally active HSP90 inhibitor that competitively binds to the ATP-binding site in the NTD of HSP90 (Uno et al. 2019). Pimitespib exhibits a stronger specific

binding affinity for HSP90 α and HSP90 β than for the highly homologous HSP90 family members GRP94 and TRAP1. In 2022, pimitespib received its first approval in Japan for the treatment of gastrointestinal stromal tumour (GIST). Pimitespib exhibits strong anti-cancer activity in xenograft models without causing evident ocular toxicity in rats (Ohkubo et al. 2015). A phase I study revealed that pimitespib had an acceptable safety profile with potent anti-tumour activity in patients with advanced solid tumours, including those with heavily pre-treated GIST (Shimomura et al. 2019). In a phase II study, pimitespib exerted significant therapeutic effects against advanced GIST that was refractory to standard treatment with imatinib, sunitinib and regorafenib (Doi et al. 2019). The median progression-free survival (PFS) was 4.4 months, and the 12-week progression-free rate was 73.4%. Treatment-related adverse events were observed in all patients; however, they can be resolved via dose modification. In phase III trials of pimitespib, patients with previously treated GIST had significantly prolonged PFS when compared with patients treated with a placebo (Honma et al. 2021; Kurokawa et al. 2022; Sawaki et al. 2022).

Imatinib is an oral tyrosine receptor kinase inhibitor targeting PDGFR, c-KIT, BCR/ABL and v-ABL (Cohen et al. 2021). It has been approved for the treatment of Ph-positive

Table 1 Monotherapy with HSP90 inhibitors in the clinical trials

Entry	HSP90 inhibitors	Conditions	Phase	Registration no
1	Ganetespi	Leukemia	I	NCT00964873
2	Ganetespi	Solid tumors	I	NCT00687934
3	Ganetespi	Hematologic malignancies	I	NCT00858572
4	Ganetespi	Colon cancer Rectal cancer	I	NCT01111838
5	Ganetespi	Hepatocellular carcinoma	I	NCT01665937
6	Ganetespi	Non small cell lung cancer	II	NCT01031225
7	Ganetespi	Ocular melanoma	II	NCT01200238
8	Ganetespi	HER2 ⁺ or triple negative breast cancer	II	NCT01677455
9	Ganetespi	Small cell lung cancer	II	NCT01173523
10	Ganetespi	Metastatic hormone-resistant prostate cancer	II	NCT01270880
11	Ganetespi	Advanced esophagogastric cancer	II	NCT01167114
12	Luminespi	Gastrointestinal stromal tumor	II	NCT01404650
13	Luminespi	Non small cell lung cancer	II	NCT01752400 NCT01124864
14	Luminespi	Advanced solid tumors	I	NCT01132625
15	Luminespi	Breast cancer, hematologic neoplasms	I/II	NCT00526045
16	Onalespi	Refractory solid tumors	I	NCT01246102
17	Onalespi	Metastatic solid tumors	I	NCT00878423
18	SNX-5422	Refractory solid tumor malignancies	I	NCT00506805 NCT01611623
19	SNX-5422	Hematologic neoplasms	I	NCT00595686
20	SNX-5422	Neuroendocrine tumors	I	NCT02063958
21	SNX-5422	Resistant lung adenocarcinoma	I	NCT01851096
22	SNX-5422	Solid tumor cancers and lymphomas	I	NCT00644072
23	SNX-5422		I	NCT00647764
24	SNX-5422	Refractory hematological malignancies	I	NCT01635712
25	BIIB021	Advanced solid tumors	I	NCT00345189
26	MPC-3100	Refractory or relapsed cancer	I	NCT00920205
27	Pimitepi	Advanced solid tumors	I	NCT02965885
28	HSP990	Advanced solid malignancies	I	NCT00879905
29	BIIB028	Advanced solid tumors	I	NCT00725933
30	Debio 0932(CUDC-305)	Solid tumors or lymphoma	I	NCT01168752
31	Alvespimycin	Advanced solid tumor or lymphoma	I	NCT00088868
32	PU-H71	Advanced malignancies	I	NCT01393509
33	Retaspimycin	Lung cancer	I/II	NCT00431015

chronic myeloid leukaemia (CML) and GIST. Mutations in *KIT* or *PDGFRA* block the clinical activity of imatinib against GIST. Both *KIT* and *PDGFRA* are clients of HSP90 and can be downregulated by HSP90 inhibitors. Therefore, the combination of an HSP90 inhibitor and imatinib is considered an effective strategy for overcoming imatinib resistance in GIST. Saito et al. reported that pimitespi inhibited the growth of both imatinib-resistant GIST cell lines and EGFR-mutated lung cancer cell lines (Saito et al. 2020). Recently, a phase I study has been initiated to investigate the effects of pimitespi combined with imatinib in 78 patients with advanced GIST refractory to imatinib (NCT05245968).

Nivolumab is a human monoclonal antibody that targets programmed cell death-1 ligand 1 (PD-L1) and

blocks its interaction with programmed cell death-1 (PD-1) (Sharma and Allison 2015). The Food and Drug Administration (FDA) has approved nivolumab for the treatment of metastatic non-small cell carcinoma, metastatic melanoma, relapsed Hodgkin's lymphoma, metastatic head and neck cancer (HNC) and advanced renal cell carcinoma. However, nivolumab does not benefit patients with microsatellite-stable (MSS) or mismatch repair (MMR)-proficient colorectal cancer (CRC) (Hirano et al. 2021). Regulatory T cells are reported to induce resistance to anti-PD1/PD-L1 inhibitors by inhibiting effective anti-tumour immunity (Togashi et al. 2019). Pimitepi can degrade the signal transducer and activator of transcription 5 (STAT-5) and reduce the regulatory

Table 2 Combination therapies with HSP90 inhibitors in clinical trials

Entry	Combination therapy	Conditions	Phase	Registration no
1	Luminespib /alpelisib	Advanced or metastatic gastric cancer	I	NCT01613950
2	Luminespib /capecitabine	Metastatic or unresectable solid tumor malignancy	I	NCT01226732
3	Luminespib /pemetrexed disodium	Recurrent NSCLC; squamous cell lung cancer; Stage IV non-small cell lung cancer	I	NCT01784640
4	Luminespib /cetuximab	KRAS wild-type metastatic colorectal cancer	I	NCT01294826
5	Luminespib /erlotinib hydrochloride	Adenocarcinoma of the lung; non-small cell lung cancer	I/II	NCT01259089
6	Luminespib /bortezomib/dexamethasone	Relapsed or refractory multiple myeloma	I/II	NCT00708292
7	Luminespib /trastuzumab	Advanced HER2-positive breast cancer	I/II	NCT01271920
8	Luminespib /bortezomib/dexamethasone	Relapsed or refractory multiple myeloma	I/II	NCT00708292
9	Ganetespib /carboplatin/paclitaxel/radiation	Esophageal cancer	I	NCT02389751
10	Ganetespib /docetaxel	Solid tumor malignancies	I	NCT01183364
11	Ganetespib /paclitaxel/trastuzumab/pertuzumab	Metastatic HER2 positive breast cancer	I	NCT02060253
12	Ganetespib /pemetrexed/cisplatin (carboplatin)	Pleural mesothelioma	I	NCT01590160
13	Ganetespib /plerixafor/quizartinib	Acute myeloid leukaemia; high risk myelodysplastic syndrome	I/II	NCT01236144
14	Ganetespib /paclitaxel	Platinum-resistant epithelial ovarian cancer	I/II	NCT02012192
15	Ganetespib /paclitaxel	Refractory sarcomas and MPNSTs	I/II	NCT02008877
16	Ganetespib /standard chemotherapy	Breast cancer	II	NCT01042379
17	Ganetespib /docetaxel	Non-small cell lung cancer	II/III	NCT01348126
18	Onalespib /olaparib	Advanced solid tumor	I	NCT02898207
19	Onalespib /abiraterone acetate/prednisone (prednisolone)	Castration-resistant prostate cancer	I/II	NCT01685268
20	Onalespib /imatinib	Gastrointestinal stromal tumor	II	NCT01294202
21	Onalespib /crizotinib	Non-small cell lung cancer	I/II	NCT01712217
22	Onalespib /AT7519	Advanced solid tumor	I	NCT02503709
23	Pimitiespib /imatinib/sunitinib	Gastrointestinal stromal tumor	I	NCT05245968
24	Pimitiespib /nivolumab	Colorectal cancer; solid tumor	II	EPOC1704
25	XL888 /vemurafenib	Advanced BRAF ^{V600E} -mutant melanoma	I	NCT01657591
26	SNX-5422 /carboplatin/paclitaxel	Advanced non-small cell lung cancer and small cell lung cancer	I	NCT01892046
27	KW-2478 /bortezomib	Relapsed/refractory multiple myeloma	I/II	NCT01063907
28	Tanespimycin /bortezomib	Multiple myeloma in first relapse (BMS TIME-1)	III	NCT00546780

T cells to enhance the anti-tumour activity of anti-PD1/PD-L1 inhibitors. Therefore, the combination of pimitiespib and anti-PD1/PD-L1 inhibitors is considered an effective strategy for the treatment of cancer. The maximum tolerated dose and recommended phase II dose of the combination of pimitiespib and nivolumab were evaluated in a dose-discovery and -expansion phase Ib trial (Kawazoe et al. 2021). A total of 44 patients with CRC and other solid tumours were enrolled in this trial. Dose-limiting toxicity was not observed at any dose level. The trial revealed that the combination of 160 mg pimitiespib plus nivolumab had a manageable safety profile and anti-tumour activity, especially in MSS CRC, with an objective response rate (ORR) of 16%.

Onalespib (AT13387)

Onalespib, a resorcinol-derived HSP90 inhibitor, has a strong affinity for HSP90 α (Woodhead et al. 2010). A first-in-human phase I dose-escalation study revealed that onalespib had an acceptable safety profile and exhibited linear pharmacokinetic properties and preliminary anti-tumour activity (Shapiro et al. 2015). HSP70 induction was observed as proof of target engagement. Notably, the unfavourable hepatotoxicity of the first-generation ansamycin class of HSP90 inhibitors that hampered their clinical development has not been observed in studies on onalespib. Moreover, dose-limiting toxicity has not been observed with a once-weekly regimen. In another phase I clinical

trial, no responses were observed in 31 patients with various advanced solid tumours (Do et al. 2015). However, 8 patients had stable disease for > 2 cycles, and 1 patient with hepatocellular carcinoma remained under observation for six cycles.

AT7519, a pan-inhibitor of cyclin-dependent kinases (CDKs), exhibits anti-tumour activity in patients with advanced solid tumours, chronic lymphocytic leukaemia and mantle cell lymphoma (Wyatt et al. 2008). Do et al. reported that AT7519 inhibited the compensatory expression of HSP70 induced by onalespib (Do et al. 2020). They conducted a phase I clinical trial of onalespib combined with AT7519 in patients with advanced solid tumours. Plasma analysis revealed an increase in HSP70 expression after treatment with onalespib alone and a decrease in HSP70 expression after treatment with the combination of onalespib and AT7519. Therefore, AT7519 may overcome the acquired drug resistance to HSP90 inhibitors caused by heat shock response, providing a rationale for combining AT7519 with HSP90 inhibitors. Partial responses were observed in a palate adenocarcinoma and Sertoli–Leydig tumor; a colorectal and an endometrial cancer patient remained on study for 10 cycles with stable disease as the best response. Most adverse effects accounted for grades 1 and 2 (92.3%), followed by grade 3 (7%) and grade 4 (0.7%). In addition, no clinically relevant pharmacokinetic interactions were observed for either drug. In a later work, the El-Deiry group discovered that co-inhibition of CDK and HSP90 resulted in the destabilisation of hypoxia-inducible factor-1 α (HIF-1 α) and showed a synergistic effect on reducing the viability of cancer cells (Zhao et al. 2021).

Olaparib is a poly(ADP-ribose) polymerase (PARP) inhibitor that has been approved for the treatment of patients with advanced ovarian cancer and human epidermal growth factor receptor 2 (HER2)-negative breast cancer with deleterious germline BRCA mutation (gBRCAm), who have been previously treated with chemotherapy (Paik 2021). Olaparib has shown remarkable anti-tumour activity in homologous recombination (HR)-deficient cancers but not in HR-proficient cancers (Parsels et al. 2021). Choi et al. (2014) identified the HSP90 inhibitor tanespimycin as a suppressor of HR based on a connectivity map. They found that it enhanced sensitivity to olaparib and carboplatin in HR-proficient ovarian cancer cells, suggesting a rational combination of HSP90 inhibitors and olaparib. Konstantinopoulos et al. conducted a phase I clinical trial of onalespib combined with olaparib in patients with advanced solid tumours (Konstantinopoulos et al. 2022) Although no objective responses were observed, disease stabilisation for \geq 24 weeks was observed in 7/22 (32%) patients, including those with BRCA-mutated ovarian cancer and acquired resistance to PARP inhibitors and those with tumours harbouring alterations in the retinoblastoma pathway. Dose-limiting toxicity was not observed in

patients treated with 300 mg olaparib/40 mg onalespib and 200 mg olaparib/80 mg onalespib. Pharmacokinetic analysis revealed that co-administration of olaparib and onalespib did not affect the steady-state pharmacokinetic behaviour of either agent.

Abiraterone is an inhibitor of 17 α -hydroxylase/C17, 20-lyase (CYP17) that has been approved for the treatment of castration-resistant prostate cancer (CRPC) in combination with prednisone. Resistance to abiraterone is attributed to the increased expression of androgen receptor (AR), the expression of alternatively spliced AR and somatic point mutations (Rice et al. 2019). Therefore, alternative strategies that can effectively target both persistent full-length AR (AR-FL) and AR splice variants should be developed for the treatment of CRPC. Onalespib can effectively deplete both AR-FL and AR-V7 protein, thereby inhibiting the growth of 22Rv1 (CRPC) cells expressing AR-V7 and VCaP cells overexpressing AR-FL (Ferraldeschi et al. 2016). These results indicate that the depletion of AR-FL and AR-V7 by HSP90 inhibitors can benefit CRPC. Therefore, future studies should investigate the efficacy of the combination of HSP90 inhibitors and abiraterone in the clinical treatment of advanced CRPC. However, in the phase I/II clinical trial, the combination of onalespib and abiraterone with either prednisone or prednisolone was not found to benefit patients with CRPC (Slovin et al. 2019). No patient showed an objective or prostate-specific antigen response in either regimen. Moreover, circulating tumour cell counts declined transiently and increased rapidly after treatment, suggesting that HSP90 inhibition was not effective in blocking the AR signalling pathway and inducing the apoptosis of cancer cells.

Like other HSP90 inhibitors, onalespib has been demonstrated to be effective in overcoming imatinib resistance in GIST. In a dose-escalation phase I study, the combination of onalespib and imatinib was well tolerated in patients with TKI-resistant metastatic GIST (Wagner et al. 2016). In particular, 9 (35%) patients, including 2 patients with KIT mutations, had stable disease, which was considered the best response. Disease control was achieved at 4 months in 5 patients (19%), and the median PFS period was 112 days. In addition, 1 patient with PDGFRA-mutant GIST had a partial response for more than 376 days. The highest dose of onalespib in combination with imatinib in patients with normal baseline renal function was 220 mg/m² once weekly for 3 weeks of every 4-week cycle. Approximately 81% of patients reported > 1 onalespib-related gastrointestinal disorder.

Small-molecule BRAF inhibitors, such as vemurafenib and dabrafenib, lead to rapid and effective responses in patients with metastatic melanoma; however, resistance resulting in therapeutic escape is common (Savoia et al. 2020). Relapse is often caused by the recovery of signalling in the MAPK and/or PI3K/AKT pathway (Shi et al.

2014). Since many key drivers in both signalling pathways are known as the clients of HSP90, inhibition of HSP90 is considered a promising strategy for overcoming resistance to BRAF inhibitors. Both N-terminal and C-terminal inhibitors of HSP90 have been shown in preclinical investigations to reverse the resistance to BRAF and BRAF-MEK inhibitors and delay the onset of BRAF inhibitor resistance (Paraiso et al. 2012; Smyth et al. 2014; Sanchez et al. 2021; Sasame et al. 2022). The safety and efficacy of onalespib, in combination with dabrafenib and trametinib (a MEK inhibitor), were examined in a recent Phase I study by the Mooradian group in BRAF^{V600E/K}-mutant solid tumors (Mooradian et al. 2023). In this study, twenty-two patients with metastatic, BRAF^{V600E}-mutant solid tumors were enrolled. With an overall response rate of 9.5%, a disease control rate of 47.6%, and a median overall survival of 5.1 months, two patients had a partial response and eight had stable disease. Dose-limiting toxicities were myelosuppression and fatigue that occurred in two patients at high dose levels, respectively.

Ganetespib (STA-9090)

Ganetespib is an injectable second-generation small-molecule inhibitor of HSP90. It exhibits a favourable safety profile and does not cause ocular and liver toxicities associated with the use of first-generation ansamycin-type HSP90 inhibitors and some second-generation HSP90 inhibitors (Proia and Bates 2014). In a Phase II study, ganetespib administered once weekly was well tolerated in patients with heavily pre-treated advanced GIST, with no evidence of severe liver, ocular, cardiac or renal toxicity (Demetri et al. 2011). In other phase II clinical trials, ganetespib showed controllable toxicity in previously treated patients with advanced oesophageal gastric cancer and patients with refractory metastatic CRC (Kwak et al. 2013; Cercek et al. 2014).

Pemetrexed, an antifolate drug, inhibits several key enzymes, including dihydrofolate reductase (DHFR), thymidylate synthetase (TS) and glycinamide ribonucleotide formyltransferase (GARFT) in the folate metabolic pathway (Rossi et al. 2018). It significantly affects DNA synthesis in cancer cells and inhibits cancer growth. Combination chemotherapy with pemetrexed and cisplatin is a standard treatment strategy for pleural mesothelioma (PM); however, the median overall survival (OS) is only approximately 12 months. Oncogenic kinases such as AXL and MET, which are involved in the progression of PM, are clients of HSP90. In addition, TS is also a client of HSP90 and has been associated with pemetrexed activity and antifolate resistance (Abu Lila et al. 2016). Therefore, the combined use of pemetrexed and HSP90 inhibitors may lead to synergistic anti-tumour effects and help to overcome drug resistance. In phase I clinical trial, combination therapy

with ganetespib, pemetrexed and cisplatin/carboplatin was well-tolerated in patients with PM, with evident anti-tumour activity, particularly at the recommended dose of 200 mg/m² (Fennell et al. 2018). The partial tumour response rate was 61%. Among 27 patients, 7 patients had tumour burden reduction of > 50%, and 1 patient remained progression-free even after 37 months.

Taxanes, such as paclitaxel (Taxol) and its derivative docetaxel, are widely used chemotherapeutic drugs. They inhibit tubulin depolymerisation and are widely used in the treatment of various solid tumours, including breast, prostate, gastric, head and neck, ovarian and pancreatic cancers and non-small cell lung cancer (NSCLC) (Mosca et al. 2021). Nguyen et al. reported that tanespimycin led to a 5–22-fold increase in the cytotoxicity of paclitaxel in the lung cancer cell line H358 (Nguyen et al. 2001). The combination of tanespimycin and paclitaxel effectively suppressed tumour growth in vivo and significantly prolonged the survival of mice bearing H358 xenografts. Tanespimycin downregulated the expression of mutant EGFR and p53, and reduced microvasculature, thus increasing the sensitivity of tumours to taxanes. Ray-Coquard et al. completed the first clinical trial potentially targeting stabilized mutant gain-of-function p53 protein via the mechanism of depletion by HSP90 inhibition (Ray-Coquard et al. 2019). In the phase I/II trial in women with high-grade platinum-resistant epithelial ovarian cancer, administration of ganetespib combined with paclitaxel was found to be safe. Of the 10 enrolled patients, 2 patients achieved a partial response with an ORR of 20% and 4 patients had stable disease (disease control rate of 60%). In addition, no ocular or liver toxicity was observed in the 10 patients. Lang et al. evaluated ganetespib in combination with standard chemotherapy in patients with high-risk early-stage breast cancer (Lang et al. 2022). A total of 93 patients with HER2-negative breast cancer were administered ganetespib every 3 weeks with weekly paclitaxel over 12 weeks, followed by doxorubicin. In the overall study population, ganetespib exhibited limited clinical efficacy in hormone receptor-positive early-stage breast cancer. The final estimated pathological complete response rates were 26% versus 18% in patients with HER2-negative cancer, 38% versus 22% in patients with HR-negative/HER2-negative cancer and 15% versus 14% in HR-positive/HER2-negative cancer in the ganetespib versus the control group, respectively. None of the 18 tested biomarkers (including HSP90, GR/efflux, proliferation, DNA repair and immune biomarkers) was significantly associated with pathological complete response in HER2-negative breast cancer. This suggests HSP90 inhibitors may serve as potential anti-cancer agents in other clinical settings such as HER2-positive disease or in combination with anti-PD1 neoadjuvant chemotherapy in triple-negative breast cancer. In a phase II clinical study, Ramalingam et al. evaluated the combination of ganetespib

and docetaxel for the second-line treatment of 381 patients with advanced NSCLC (Ramalingam et al. 2015). Although no efficacy was observed in non-adenocarcinoma patients, PFS (4.5 months) and OS (10.2 months) were significantly prolonged in advanced lung adenocarcinoma patients after > 6 months of diagnosis of advanced disease. In addition, the combination therapy exhibited an acceptable safety profile, as ganetespib did not increase adverse events compared with docetaxel alone. The subsequent phase III trial included 677 patients with stage IIIB or IV adenocarcinoma; 335 were randomly assigned to ganetespib and docetaxel, while 337 received docetaxel alone (Pillai et al. 2020). However, this combination regimen did not improve the survival for salvage therapy of patients. The most common grade 3 or 4 adverse event in both arms was neutropenia.

Sirolimus (Rapamune), a macrocyclic antibiotic with immunosuppressive activity, was approved in 1999 for use in patients undergoing organ transplantation. Recently, it has received substantial attention owing to its potential as an anti-cancer drug. As an inhibitor of the mammalian target of rapamycin (mTOR), sirolimus prevents the propagation of IL-2-mediated cell proliferation signalling through the PI3K/AKT/mTOR pathway (Namba et al. 2006). De Raedt et al. (2011) reported that malignant peripheral nerve sheath tumours (MPNST) are hypersensitive to ER stress-inducing agents (De Raedt et al. 2011). Considering that HSP90 inhibitors can enhance ER stress in cancer cells, they tested the combination of IPI-504, an HSP90 inhibitor, and sirolimus in aggressive mouse models. The two drugs exerted synergistic effects by promoting unresolvable ER stress, resulting in catastrophic damage to ER and mitochondria. Based on this result, a phase I/II clinical trial was conducted to examine the efficacy of ganetespib combined with sirolimus in 20 patients with refractory sarcoma and MPNST (Kim et al. 2020a, b). Ganetespib was intravenously administered, and sirolimus was orally taken. The most common adverse reactions were diarrhoea, elevated liver transaminase levels and fatigue, which were not attributed to dose-limiting toxicity. Although the combination was well tolerated and toxicity was manageable, no responses were observed.

XL888

XL888, an oral HSP90 inhibitor with a good kinase selectivity profile, can significantly inhibit the activity of HSP90 and exert minimal inhibitory effects against 29 other diverse kinases (Bussenius et al. 2012). Preclinical studies have revealed that XL888 is a good candidate for the treatment of melanoma, advanced pancreatic cancer/CRC and liver cancer (Haarberg et al. 2013). A phase I study evaluating the safety and pharmacokinetic properties of XL888 in adult patients with solid tumours was initiated in 2015 but terminated 3 years later. At present, several clinical trials are

actively investigating the combination of XL888 and other anti-cancer drugs.

Eroglu et al. initiated a clinical study to evaluate the combination of vemurafenib and XL888 in 21 patients with advanced BRAF^{V600E}-mutant melanoma (Eroglu et al. 2018). Objective responses were observed in 15 of 20 evaluated patients, with 3 patients demonstrating a complete response and 12 patients demonstrating a partial response. The median PFS and OS were 9.2 months and 34.6 months, respectively. The side effects were tolerated, and the most common grade-3/4 toxicity was skin toxicity.

SNX-5422

SNX-5422 is an oral prodrug of SNX-2112, a highly selective inhibitor targeting HSP90 (Huang et al. 2009). SNX-5422 has been investigated in clinical trials in patients with various cancers, including those with HER2-positive breast cancer, neuroendocrine tumour, resistant lung adenocarcinoma and refractory haematological malignancies. In phase I clinical trials, the preliminary clinical activity of SNX-5422 has been validated in patients with prostate cancer, HER2-positive breast cancer, transformed lymphoma and multiple myeloma (Rajan et al. 2011; Reddy et al. 2013; Infante et al. 2014). However, ocular toxicity has been observed in patients and animal models treated with SNX-5422, preventing further investigation of SNX-5422 as a monotherapy for cancer.

Friedman et al. reported that SNX-5422/-2112 exhibited synergistic activity with carboplatin and paclitaxel in HSP90-overexpressing head and neck squamous cancer cell lines (Friedman et al. 2013). Gutierrez et al. conducted a phase I clinical trial to evaluate the therapeutic efficacy of the combination of SNX-5422, carboplatin and paclitaxel followed by SNX-5422 maintenance therapy in 23 patients with advanced NSCLC (n = 20) and SCLC (n = 3) (Gutierrez et al. 2021). The combination therapy was well tolerated and exerted synergistic effects. A total of 6 patients with NSCLC had a partial response (33%), 10 patients had stable disease (56%) and 2 patients had primary progressive disease (11%). The most common treatment-related grade-3/4 adverse events were gastrointestinal reactions including diarrhoea and nausea. More importantly, ocular toxicity was not observed.

KW-2478

KW-2478 is an HSP90 inhibitor with high binding affinity and potent anti-tumour activity. In a phase I clinical trial on patients with B-cell malignancies, 24 of the 25 (96%) evaluated patients had stable disease, with 5 patients being free of disease progression for ≥ 6 months (Yong et al. 2016). In

addition, KW-2478 was well tolerated, with no evidence of DLT or unexpected toxicity.

Bortezomib is an FDA-approved proteasome inhibitor used for the treatment of multiple myeloma and mantle cell lymphoma (Tan et al. 2019). Inhibition of the proteasome with bortezomib leads to the accumulation of immunoglobulin-derived defective ribosomal products, known as misfolded/unfolded protein response (UPR), which causes apoptosis owing to extensive ER stress (Meister et al. 2007). Because HSP90 inhibitors can induce the accumulation of misfolded/unfolded proteins, they are supposed to enhance the anti-myeloma activity of bortezomib. Several preclinical studies have reported the synergistic anti-cancer activity of HSP90 inhibitors and bortezomib in multiple myeloma (Wright 2010). In a phase I/II clinical trial, the combination of KW-2478 and bortezomib was well tolerated in patients with relapsed/refractory multiple myeloma, with no evidence of overlapping toxicity (Cavenagh et al. 2017). The overall response rate was 39.2% and the clinical benefit rate was 51.9% in the evaluated population (n = 79). The median PFS and duration of response were 6.8 months and 5.6 months, respectively. The anti-myeloma activity of the combination therapy appeared relatively modest; however, the good tolerability of this combination may support further exploration of dosing schedules and other combinations.

Combination of HSP90 inhibitors in preclinical studies

Chemotherapeutics

Chemotherapeutic drugs usually kill cancer cells by targeting their DNA synthesis and replication, representing an essential strategy for the treatment of cancer. However, chemotherapy response is often limited by new or acquired drug resistance. Because many DNA repair-related proteins serve as HSP90 clients, HSP90 inhibitors can disrupt double-strand break repair and sensitise tumours to chemotherapeutic drugs (Pennisi et al. 2016).

Cisplatin

Platinum-based anti-cancer agents induce both intra- and inter-strand crosslinks in DNA through covalent interaction with the nucleophilic N-7 atom of the purine base (Sikov 2015). These crosslinks inhibit DNA synthesis by preventing RNA polymerase II from passing the site of the DNA adduct. The combination with HSP90 inhibitors has been demonstrated as an effective strategy for enhancing the therapeutic potential of cisplatin. Preclinical studies have demonstrated that the combination of cisplatin and HSP90 inhibitors induces DNA damage response and decreases cell

viability in diffuse large B-cell lymphoma (DLBCL), ovarian cancer and HNC.

Although many patients with DLBCL achieve long-term remission after first-line treatment with platinum-based drugs, nearly 40% of these patients develop refractory disease or relapse after the initial remission (Vaidya and Witzig 2014). Tanespimycin, a first-generation HSP90 inhibitor, has been reported to enhance the anti-neoplastic effects of cisplatin in seven DLBCL cell lines (Schmidt et al. 2022). These synergistic effects were mediated by induced DNA damage, leading to increased levels of apoptosis. Cell lines with the lowest sensitivity to cisplatin treatment (e.g. RIVA, OCI-Ly7 and DB) exhibited the strongest response to the drug combination, suggesting that patients with cisplatin-resistant tumours can benefit from inhibition of HSP90.

Zhang et al. reported that exposure to three HSP90 inhibitors, including geldanamycin, tanespimycin and alvespimycin, in combination with cisplatin resulted in synergistic cytotoxic and pro-apoptotic effects on the human ovarian cancer cell line SKOV3 (Zhang et al. 2015). This combination led to a > 2-fold decrease in the resistance of SKOV3 cells to cisplatin. The mRNA and protein expression levels of various drug resistance-related genes were more dramatically altered by combination therapy than by monotherapy with HSP90 inhibitors or cisplatin. Moita et al. found that HSP90 inhibitors luminespib and HSP990 considerably improved the cisplatin potency in ovarian cancer cell lines (A2780, CaOV3, OVCAR3 and cisplatin-resistant subclones) via apoptosis induction (Rodrigues Moita et al. 2020). Triple combinations of histone deacetylase (HDAC) inhibitor, HSP90 inhibitor and cisplatin, however, did not outperform dual combinations.

The Spiegelberg group demonstrated that the HSP90 inhibitor onalespib enhanced the efficacy of cisplatin and reversed cisplatin resistance in ovarian and head and neck cancer cells (Mortensen et al. 2020). This combination delayed DNA repair as evidenced by an increase in the number of double-strand breaks in cells co-treated with cisplatin and onalespib. In addition, the Dobbstein group reported the synergistic effects of cisplatin and onalespib in both *in vitro* and *in vivo* models of cisplatin-resistant pancreatic ductal adenocarcinoma (Ewers et al. 2021). These synergistic effects were attributed to the HSP90 inhibitor-induced degradation of factors involved in the Fanconi anaemia pathway, the increased DNA damage and chromosome fragmentation and the enhanced accumulation of DNA-bound platinum.

Mouse double minute X (MDMX) binds to the N-terminal region of p53 or heterodimerises with its homologue MDM2 by interacting with its C-terminal domain to augment p53 degradation (Marine et al. 2006). Tanespimycin can destabilise MDMX and inhibit the interaction between MDMX and p53, resulting in the activation of the p53

signalling pathway and inhibition of cancer cell survival pathways, such as PI3K/AKT (Vaseva et al. 2011). Roh et al. demonstrated that tanespimycin and cisplatin exerted synergistic anti-tumour effects on HNC cells (Roh et al. 2013). Tanespimycin effectively activated and stabilised p53, inhibited the interaction between p53 and MDMX and induced the apoptosis of HNC cells.

5-Fluorouracil

5-FU, a nucleotide analogue, inhibits TS and blocks the synthesis of 2'-deoxythymidine-5'-monophosphate (dTMP) (Chon et al. 2017). Although 5-FU is widely used in the treatment of various cancers, its clinical application is limited owing to drug resistance. The El-Rayes group hypothesised that HSP90 inhibition can sensitise CRC cells to 5-FU through downregulation of TS and disruption of proliferation-related signalling pathways (Nagaraju et al. 2014). Ganetespi decreased the RNA and protein expression levels of cyclin D1 and pRb and induced p21 expression, leading to G₀/G₁ cell cycle arrest in the CRC cell lines HCT-116 and HT-29. This effect was associated with the downregulation of the transcription factor E2F1 and its target gene TS. In addition, ganetespi inhibited the PI3K/AKT and ERK signalling pathways in CRC cells, and the combination of ganetespi, oxaliplatin and 5-FU exhibited synergistic anti-tumour activity in both CRC cells and animal models.

Trabectedin

Trabectedin, a minor groove DNA double helix inhibitor, is used as second line therapy for myxoid liposarcoma (MLS). According to the Aman group's work, HSP90 inhibitors 17-DMAG, luminespi and STA-9090 showed synergistic effect with trabectedin in vitro (Vannas et al. 2022). In an MLS patient-derived xenograft model, 17-DMAG inhibited the tumor growth, but surprisingly, luminespi caused an increase in tumor growth.

Methotrexate

Numerous diseases, including cancer, autoimmune diseases, and rheumatoid arthritis have been successfully treated with methotrexate. The Hussain and Lila group discovered that ganetespi acted synergistically with methotrexate against A549 cells (Subaiea et al. 2023). They demonstrated that the combination of ganetespi and methotrexate inhibited the migration and invasion of A549 cells via obstruction of the NF- κ B signalling pathway. Additionally, their work showed this combination synergistically augmented the ROS production, and induced caspase activation and nuclear condensation and fragmentation in A549 cells.

Targeted anti-cancer agents

Kinase inhibitors

Epidermal growth factor receptor (EGFR), a member of the HER family, is a transmembrane receptor tyrosine kinase. EGFR alternations, including amplification of the EGFR gene and point mutations within the kinase-coding domain of EGFR, have been detected in many cancers, including lung, colorectal, ovarian and breast cancers and glioblastoma (Sabbah et al. 2020). The first- and second-generation EGFR tyrosine kinase inhibitors (TKIs) gefitinib, lapatinib, erlotinib, icotinib, afatinib and dacomitinib have advantages over various platinum-based chemotherapeutic drugs for the treatment of patients with advanced NSCLC with EGFR mutation. However, most patients treated with EGFR TKIs inevitably develop acquired resistance through multiple mechanisms after a median period of 10–14 months (Sequist et al. 2011). EGFR mutation in exon 20 (T790M) and c-MET amplification are responsible for acquired resistance to EGFR TKIs (Chong and Jänne 2013; He et al. 2021). The T790M mutation accounts for approximately 50% of these resistance cases. In addition, c-MET amplification can transactivate ERBB3 (HER3), resulting in PI3K activation independent of EGFR kinase activity. Combining HSP90 inhibitors with EGFR inhibitors is a promising therapeutic strategy for EGFR inhibitor-resistant NSCLC because HSP90 inhibitors can deplete various receptor tyrosine kinases, including EGFR, and inhibit the entire EGFR–PI3K–AKT–mTOR–p70S6K–S6 signalling axis irrespective of the presence of the T790M mutation (Shimamura et al. 2008). Codony-Servat et al. demonstrated that osimertinib and HSP90 inhibitors, ganetespi and luminespi, synergistically inhibited the growth of NSCLC cell lines and osimertinib-resistant cell lines, indicating that combination therapy is a promising strategy for overcoming osimertinib resistance (Codony-Servat et al. 2019). Luminespi decreased the protein expression of membrane receptors, such as EGFR, STAT3, MET, YAP and AKT, and reduced the phosphorylation of several membrane receptors activated by osimertinib. The Ye and Xu group found that ganetespi was able to reverse the development of lapatinib resistance in HER2-positive breast cancer cells (Ye et al. 2021). Ganetespi and lapatinib synergistically decreased the expression of HER2/3, EGFR, AKT and ERK, and enhanced the induction of apoptosis and G1 arrest. Additionally, this combination led to the depletion of STAT3, which is probably involved in the lapatinib resistance of HER2-positive breast cancer cells. In both SKBR3 and SKBR3-L xenografts, ganetespi augmented the inhibition of tumor growth. The Park group reported that HSP90 was involved in the lapatinib resistance mechanisms, and the combination of 17-DMAG and lapatinib showed synergistical antitumour

effect in the ER-positive, HER2-overexpressing breast cancer cell line LR-BT474, and the xenograft model as well (Lee et al. 2020).

Sorafenib is a multikinase inhibitor that is considered the standard first-line option for treating advanced hepatocellular carcinoma. However, prolonged treatment with sorafenib leads to the resistance, which are featured by the reduction of microvessel density and intratumoral hypoxia (Tang et al. 2020). This hypoxia is mediated by a client of HSP90, hypoxia-inducible factors (HIFs). The Saber group reported that ganetespib augmented sorafenib efficacy via necroptosis induction in hepatocellular carcinoma (Saber et al. 2023a, b). It is interesting that the treatment with sorafenib, ganetespib, or their combination did not result in a significant change in the levels of HSP90 in HepG2 cells subjected to hypoxia. Ganetespib reduced the expression of LAMP2 and increased the expression of MLKL, suggesting that ganetespib treatment enhanced the process of necroptosis when combined with sorafenib. They further found that ganetespib and STAT3 inhibitor nifuroxazid exerted a synergistic effect in hepatocellular carcinoma (Saber et al. 2023a, b). Ganetespib decreased the levels of the HSP90 client proteins HIF-1 α and pSTAT3, resulting in the suppression of the STAT3/HIF-1 α complex transcriptional activity. This combination therapy exhibited promising survival prolongation in mice models.

Burkitt lymphoma (BL) is a germinal centre B cell-derived malignancy with a survival rate of <20% in patients with recurrent disease. Giulino-Roth et al. used PU-H71 as a probe and identified multiple proteins in the PI3K/AKT/mTOR signalling pathway as HSP90 clients in BL (Giulino-Roth et al. 2017). PU-H71 and the PI3K/mTOR inhibitor dactolisib (BEZ235) exerted synergistic effects in both in vitro and in vivo models of lymphoma. In another work, the Choi group reported a strong synergistic effect between 17-DMAG and dactolisib in cisplatin-resistant human bladder cancer cells (Kim et al. 2020a, b).

To identify effective pancreatic ductal adenocarcinoma (PDAC) therapies, the Leach group performed a large-scale, unbiased screen of 16 single-agent and 41 two-drug targeted therapy combinations, and found the synergy between HSP90 inhibition and MEK inhibition in an orthotopic mouse model (Grbovic-Huezo et al. 2020). The MEK1/2 inhibitor trametinib and the broad-spectrum receptor tyrosine kinase (RTK) inhibitor sunitinib significantly increased the effectiveness of PU-H71 without significant body weight loss over the 2-week treatment period. The combination of PU-H71 and trametinib led to an 80% increase in the survival of PDAC-bearing mice. Mechanistic study revealed HSP90 inhibition by PU-H71 can overcome the compensatory activation of resistance pathways, including components of the PI3K/AKT/mTOR signalling axis, which was induced by single-agent MEK

inhibition. Later research in Wiener's group revealed that this combination also acted synergistically in colorectal cancer organoid (Soós et al. 2023). The Takahashi group recently reported that the combination of HSP90 inhibitor pimitespid and sunitinib exerted synergistic effect on imatinib-resistant gastrointestinal stromal tumors (Teranishi et al. 2023). This combination effectively downregulated KIT signalling and angiogenic signalling pathways in GIST cells, and decreased the tumour microvessel density in the xenograft models.

Ibrutinib, the first small-molecule drug targeting Bruton's tyrosine kinase (BTK), has been approved for the treatment of various blood cancers and chronic graft-versus-host disease (cGVHD). Although it has demonstrated excellent activity in the clinical treatment of patients with chronic lymphocytic leukaemia (CLL), it does not eliminate the disease or induce durable responses without continuous therapy (Byrd et al. 2013). The TC481S mutation in BTK has been identified as the primary mechanism underlying resistance to ibrutinib in patients with CLL. In addition, survival-related signalling pathways such as the PI3K-AKT, classical and alternative NF- κ B and receptor tyrosine kinase-like orphan receptor 1 (ROR1)-mediated pathways may contribute to primary resistance to ibrutinib (Yu et al. 2017). Given that inhibition of HSP90 affects multiple signalling pathways involved in the development of ibrutinib resistance, the Yu group performed iTRAQ-based quantitative proteomic analysis and identified 130 potential client proteins of HSP90 (Liu et al. 2020). Specifically, ROR1 was identified as a novel client protein of HSP90. Inhibition of HSP90 induced ROR1 degradation and hence attenuated the tumour escape ability of ibrutinib. In addition, ibrutinib inhibited HSP90 inhibitor-induced degradation of ibrutinib-binding proteins including BTK, BLK, LCK and LYN but not that of proteins that do not bind to ibrutinib, such as ROR1. The HSP90 inhibitor alvespimycin induced ROR1 degradation and worked synergistically with ibrutinib in vitro and in vivo. Further analysis revealed that inhibition of HSP90 by alvespimycin suppressed the BCR signalling pathway in CLL cells. Furthermore, the Hertlein group demonstrated that the HSP90 inhibitor SNX-5422 was effective in depleting both wild-type BTK and BTK-C481S (Chen et al. 2021). Compared with vehicle treatment or monotherapy, the combination of SNX-5422 and ibrutinib resulted in a significant survival benefit in the E μ -TCL1 mouse model of CLL. Later work in the Tu and Liu group reported that the HSP90 inhibitor ganetespib improved the sensitivity of mantle cell lymphoma to ibrutinib (Lu et al. 2022). Ganetespib enhanced the cell cycle arrest and apoptosis in vitro, and significantly increased inhibition of tumor growth mediated by ibrutinib in the mouse models.

Proteostasis inhibitors

Cancer cells are particularly reliant on the proteostasis network, however targeting single component in this network might result in compensation. The Gestwicki group investigated the binary combinations of four proteostasis inhibitors, including 17-DMAG, JG-98 (HSP70 inhibitor), bortezomib (proteasome inhibitor) and CB-5083 (p97 inhibitor) in prostate cancer cell line 22Rv1 cells (Shkedi et al. 2021). The combinations of Hsp90-p97 and Hsp90-Hsp70 were modestly synergistic, while the combination of p97 and proteasome inhibitor was the most strongly synergistic. The synergistic effects of the combinations with Hsp90 and Hsp70 inhibitors may be partly attributable to the induced degradation of both AR-FL and AR splice variants, interrupting the AR signalling required for growth of 22Rv1 cells.

Anti-androgen drugs

Anti-androgens are a class of drugs that bind to AR and block the effects of androgen to inhibit the growth of cancer cells (Student et al. 2020). They are commonly used to treat metastatic castration-sensitive prostate cancer (mCSPC) and mCRPC. Because AR is a client protein of HSP90, HSP90 inhibitors may enhance the therapeutic efficacy of anti-androgen drugs in prostate cancer. Recently, the Stoyanova group tested 22 chalcones for their inhibitory effects on tumour growth across the National Cancer Institute's 60 human cancer cell lines and found that SU086 was highly effective in inhibiting the growth of two prostate cancer cell lines, DU145 and PC3, and its activity was independent of the AR status (Rice et al. 2022). HSP90 was identified as the target of SU086 based on cellular thermal shift assay. SU086 can bind to HSP90 α and HSP90 β isoforms and, unlike classical HSP90 inhibitors, reduce HSP90 protein levels in prostate cancer cells. Further evaluation revealed that SU086 exerted excellent inhibitory effects on tumour growth in vivo both as a single drug and in combination with anti-androgen therapy. SU086 worked synergistically with the second-generation anti-androgen drugs enzalutamide and abiraterone in C4-2 CRPC cells. In C4-2 xenografts, SU086 combined with enzalutamide or abiraterone halted tumour growth without causing measurable toxicity during 21 days of treatment.

Fatty acid oxidation inhibitors

The Butler group conducted a proteomic analysis of 30 clinical prostate tumors and found that HSP90 inhibitor luminespib significantly increased the abundance of proteins involved in oxidative phosphorylation and fatty acid metabolism (Nassar et al. 2020). Therefore, the inhibition of fatty acid oxidation would sensitize prostate cancer cells

to HSP90 inhibition. The combination of luminespib and perhexiline significantly reduced the cell viability of prostate cancer cells by inducing cell arrest and apoptosis. It is notable that perhexiline attenuated the heat shock response of luminespib.

Glucose-6-phosphate dehydrogenase (G6PD) inhibitor

Dehydroepiandrosterone (DHEA) is endogenously secreted as a precursor for the synthesis of various hormones. DHEA is a non-competitive inhibitor of G6PD. The Saeed group found that PU-H71 and DHEA exerted a synergistic effect on promoting apoptosis in triple-negative breast cancer cell line MDA-MB-231 (Soudan et al. 2020). The cells treated with PU-H71 and DHEA combination showed a highly significant increase in ROS levels in comparison to either drug alone. It is interesting that the combination of PU-H71 and DHEA resulted in a significant reduction in HSP90 expression, suggesting a better therapeutic outcome in comparison to the single drug, PU-H71.

Monoclonal antibodies

Trastuzumab, a humanized monoclonal antibody targeting HER2, has been approved for the treatment of early and advanced HER2-positive breast cancer and advanced HER2-positive gastric cancer (Swain et al. 2023). Mechanisms underlying the development of resistance to trastuzumab include the activation of PI3K signalling, presence of a truncated form of HER2 lacking an antibody-binding epitope and enhancement of signal through HER3 or other receptor tyrosine kinases such as IGF-IR (Bai et al. 2018; Derakhshani et al. 2020). Because many signalling proteins involved in the development of resistance are known client proteins of HSP90 (Huang et al. 2010), the combined use of HSP90 inhibitors and trastuzumab is a rational strategy for overcoming resistance to trastuzumab. Wainberg et al. examined the synergistic effects of the HSP90 inhibitor luminespib and trastuzumab in both breast and gastric cancers. This combination therapy induced more potent HER2 degradation and G₂-phase cell cycle arrest and increased cell apoptosis (Wainberg et al. 2013). In addition, it exhibited stronger anti-tumour efficacy than either drug alone in a xenograft model of trastuzumab-resistant gastric cancer. Recent studies also revealed that the combination of trastuzumab and HSP90 inhibitors such as geldanamycin and 17-AAG significantly increased the cytotoxicity compared to trastuzumab alone in HER2 overexpressing cancer cell lines (Skeie et al. 2020; McCombs et al. 2021).

Glembatumumab vedotin is an antibody drug conjugate targeting transmembrane glycoprotein NMB (GPNMB) (Rose et al. 2017). Glembatumumab vedotin exhibits variable efficacy against GPNMB-positive metastatic TNBC as

a single agent. Biondini et al. reported that suppression of HSP90 increases GPNMB expression and cell-surface localization, sensitizing breast cancer cells to glembatumumab vedotin (Biondini et al. 2022). HSP90 inhibition resulted in lysosomal dispersion towards the cell periphery and fusion with the plasma membrane, which delivered GPNMB to the surface of breast cancer cells.

Radiation therapy

Radiation therapy is an essential cancer treatment strategy that involves the use of radiation to kill cancer cells through DNA damage. However, patients with cancer often develop resistance to radiation therapy (Kabakov et al. 2010; Kim et al. 2015). Although the resistance mechanisms are multifaceted, numerous studies have revealed that multiple HSP90 clients are involved in these mechanisms.

HIF-1 α is responsible for inducing radiation resistance through the hypoxia signalling pathway (Albadari et al. 2019). Given that HIF-1 α is a client protein of HSP90, the Multhoff group investigated the effects of HSP90 inhibitors on radiation therapy (Schilling et al. 2012). The HSP90 inhibitors luminespib and tanespimycin significantly enhanced the radiosensitivity of H1339 lung cancer cells under normoxic and hypoxic conditions. These synergistic effects were found to be independent of HIF-1 α , as HSP90 inhibitors surprisingly increased HIF-1 α levels in H1339 cells. On the contrary, the El-Rayes group demonstrated that ganetespib significantly reduced HIF-1 α levels in PDAC cells (Nagaraju et al. 2019). Compared with either ganetespib or radiation plus 5-FU treatment, the combination of ganetespib, radiotherapy and 5-FU resulted in more potent inhibition of tumour growth in HPAC tumour xenograft models. The combination treatment was well tolerated, with no measurable toxicity or significant reduction in the body weight of animals.

The Krawczyk group found that ganetespib potentiated cytotoxicity as well as radio- and chemosensitizing effects in cervix cancer cell lines (Vriend et al. 2017). HSP90 inhibition enhanced the induction of DNA damage by hyperthermia, and reduced thermotolerance. Later, this group demonstrated that this combination treatment was effective in 10 different cell lines (Scutigliani et al. 2022). Ganetespib can reduce the thermal dose and temperature required for sensitisation. In a subcutaneous murine model of cervix cancer, ganetespib enhanced the effects of radio-thermotherapy in four out of six mice. The Chen group demonstrated that the synergistic effect between ganetespib and hyperthermia therapy was attributable to the HSP90 inhibition-induced degradation of DNA-PKcs protein and suppression of *PRKDC* transcription (Liu et al. 2021).

The Harrington group revealed that luminespib induced radiosensitivity in athymic mice bearing tumour xenografts

established using human HNC cells (Zaidi et al. 2012). The combination of radiotherapy and luminespib significantly delayed tumour growth and increased surrogate end-point survival. Luminespib depleted HSP90 clients, including AKT, ErbB2 and cRAF, in HeLa and HN3 cells and delayed Rad51 foci formation and subsequent resolution of radiation-induced double-strand DNA break repair.

In 2020, the Spiegelberg group reported that onalespib and radiotherapy exerted synergistic antitumour effect in radiosensitive HCT116 and radioresistant A431 cells (Spiegelberg et al. 2020). The Nyati group revealed that onalespib sensitised radiotherapy even at a subcytotoxic concentration (Mehta et al. 2020). A low-dose onalespib reduced the expression of DNA repair proteins, without affecting the majority of HSP90 clients. They showed that a combination of onalespib and radiotherapy produced significant tumor growth inhibition in HNSCC and pancreatic xenografts. In a recent study, the Mitchell group reported that onalespib increased radiosensitivity in both in vitro and in vivo models of HNC (Naz et al. 2021). The combined use of onalespib and ionising radiation (IR) resulted in G₂/M-phase cell cycle arrest; inhibited DNA repair and altered tumour metabolism, including nucleotide metabolism, glycolysis and tricarboxylic acid cycle. Compared with IR alone, the combination therapy synergistically delayed tumour growth in HNC xenograft models. HSP70 expression was elevated and CDK4 expression was reduced in onalespib-treated tumours, suggesting the target engagement of HSP90. Onalespib significantly reduced the expression of HIF-1 and other IR-induced marker proteins involved in vasculogenesis, such as SDF-1, CD11b⁺ and CD45, indicating that the combination of onalespib and radiotherapy can reduce tumour recurrence after radiation therapy.

The Tran group reported that ganetespib induced radiosensitivity in liver cancer cells (Chettiar et al. 2016). The combination of ganetespib and radiotherapy induced supra-additive radiosensitivity in all tested liver cancer cell lines, including Hep3B, HepG2 and HUH7, at low nanomolar concentrations with enhancement ratios of 1.33–1.78. In addition, the combination therapy delayed tumour growth in HepG2 tumour xenografts. Furthermore, the Lin group reported that ganetespib inhibited DNA damage repair induced by radiation and intensified G₂-M-phase arrest in four types of NSCLC cells (Wang et al. 2016). Ganetespib significantly enhanced the therapeutic efficacy of radiation in H460 xenografts; however, synergistic effects were not observed in A549 xenografts. These findings suggest that radiosensitivity mediated by HSP90 inhibitors is tumour-specific.

In 2021, the Zhao and Lu group reported that HSP90 inhibitor XL888 enhanced cell apoptosis via downregulating STAT3 after insufficient radiofrequency ablation in hepatocellular carcinoma (Sun et al. 2021). XL888 increased the

heat-induced cell apoptosis of hepatocellular carcinoma cells by increasing the levels of cleaved caspase 3 and PARP. Further investigation revealed that combination of XL888 and heat treatment significantly reduced the expression level of STAT3 and its phosphorylated level, leading to the reduced expression levels of AKT, phosphorylated AKT, Bcl-xL, Bcl-2 as well as Mcl-1, and the increased expression level of BAX.

The Xue group and the Lauber group demonstrated that HSP90 inhibitors NW457, NXD30001 and onalespib, showed synergistic effects with radiotherapy in glioblastoma cells (Chen et al. 2020; Orth et al. 2021; Xu et al. 2022). HSP90 inhibitors reduced the expression of DNA damage repair regulators induced by radiotherapy. These combinations inhibited the tumor growth and prolonged the survival in animal models. Notably, Xu et al. claimed that onalespib sensitized glioblastoma cells to the combination of radiation and temozolomide by depleting the homologous recombination pathway. A recent work in the Lauber group revealed that NW457 augmented the efficacy of ionizing irradiation in CRC cells (Ernst et al. 2020). It is interesting that the combination of NW457 and radiotherapy induced the release of damage-associated molecular patterns, which enforced the differentiation of a monocyte-derived antigen presenting cell phenotype and triggered the priming of allogeneic T cell responses.

The Erin group demonstrated that PU-H71 enhanced the radiosensitivity of metastasized breast cancer cells 4T1 (Kale et al. 2020). PU-H71 decreased a variety of client proteins and increased the cytotoxicity of radiotherapy. However, the treatment of PU-H71 induced the expression of angiogenic and inflammatory factors such as MIP-2, SDF-1, and VEGF, suggesting an inflammatory response. In another work by Nowosielska's group, the combination of HSP90 inhibitor NVP-AUY922 and the whole-body low dose radiotherapy suppressed clonogenic potential in Lewis lung cancer cells and reduced tumorigenesis in mice (Nowosielska et al. 2021). However, this combination was inferior to these of radiotherapy and blockade of the immune checkpoints such as CTLA-4 and PD-1.

Immunotherapy

In addition to chemotherapy, radiotherapy and surgery, immunotherapy is an effective treatment strategy for cancer. Immunotherapy can be divided into five categories as follows: vaccine-based, cytokine-based, checkpoint blockade-based, adoptive cell transfer-based and small-molecule-based immunotherapies (Zhou et al. 2021). HSP90 is involved in immune modulation in cancer and plays an essential role in antigen presentation, effector immune cell function and regulation of inflammatory processes (Zininga et al. 2018). Therefore, the combination of HSP90 inhibitors

and immunotherapy represents a promising strategy for the treatment of cancer (Graner 2021).

The Storkus group demonstrated that the HSP90 inhibitor alvespimycin functioned as an adjuvant to erythropoietin-producing hepatoma (EPH) receptor A2 (EphA2)-specific active vaccination or adoptive transfer of EphA2-specific CD₈⁺ T cells (Rao et al. 2012). Alvespimycin reconditioned the tumour microenvironment and improved the recruitment of therapeutic T cells. Treatment with alvespimycin led to degradation of the HSP90 client protein EphA2 and subsequently increased recognition of tumour cells by type-1 anti-EphA2 CD₈⁺ T cells.

Given that intrinsic signalling pathways related to tumours interfere with processes essential for an effective anti-tumour immune response, the Hwu group identified small molecules with the potential to improve responses to immunotherapy by assessing their effects on autologous-T-cell-driven elimination of primary melanoma cells (Mbofung et al. 2017). The HSP90 inhibitor ganetespib was identified from 850 bioactive compounds. It enhanced the elimination of patient-derived human melanoma cells by their autologous T cells in vitro and potentiated responses to anti-CTLA4 and anti-PD1 therapies in vivo. Mechanistically, inhibition of HSP90 by ganetespib upregulated the interferon response genes IFIT1, IFIT2 and IFIT3 in tumours. Considering the intrinsic toxicities of pan-HSP90 inhibitors, the Lu group investigated the efficacy of an HSP90 β -selective inhibitor NDNB1182 in affecting the immune checkpoint blockade immunotherapy (Rahmy et al. 2022). NDNB1182 effectively downregulated CDK4 and increased the expression of endogenous retroviral elements and interferon-stimulated genes. Moreover, NDNB1182 enhances immune checkpoint blockade therapy with superior tolerability to ganetespib in murine tumors.

A recent study by the Lairson group revealed that HSP90 inhibitors reduced the mRNA and protein expression levels of immune checkpoint molecules, namely, PD-L1 and PD-L2, in target cells and potentially improved the efficacy of immunotherapy (Zavareh et al. 2021). Phenotype-based high-throughput screening revealed that HSP90 inhibitors affected the expression of PD-L1 and PD-L2 at clinically relevant concentrations in vitro and in vivo. Compared with monotherapy, combination therapy with anti-PD-L1 antibody and ganetespib exhibited better anti-tumour activity in syngeneic C57BL/6 mouse models of colon adenocarcinoma established using MC-38 cells.

Peptide receptor radionuclide therapy is a highly targeted and effective form of radiotherapy with minimal side effects for treating neuroendocrine tumors with an abundance of somatostatin receptors. ¹⁷⁷Lu-DOTATATE was approved for therapy of gastroenteropancreatic neuroendocrine cancer in Europe. The Nestor group found that onalespib was a radiosensitizer to potentiate the therapeutic outcomes of ¹⁷⁷Lu-DOTATATE in neuroendocrine tumor xenografts (Lundsten

et al. 2020). Later, they further developed an anti-carcinoembryonic antigen humanized antibody ^{177}Lu -DOTA-M5A and investigated the therapeutic effect of ^{177}Lu -DOTA-M5A and/or onalespib in CRC (Mohajershojai et al. 2022). Onalespib synergistically enhanced the therapeutic effects of ^{177}Lu -DOTA-M5A in three-dimensional colorectal spheroid models. This combination exerted cooperative effects on the cellular tumor suppressive pathways RTK and MAPK and therefore increased apoptosis.

Cytokine-induced killer (CIK) cell is a novel approach for adoptive cell-based immunotherapy of cancer. The Schmidt-Wolf group demonstrated that HSP90 inhibitors showed synergistic effect with CIK cells in Burkitt's lymphoma (BL) cells (Ge et al. 2023). HSP90 inhibitors, 17-DMAG and ganetespib, worked synergistically with CIK cells against BL cell lines, BL-41 and Raji. This combination significantly increased the expression of Fas and induced the caspase 3/7-dependent apoptosis in BL cells.

Conclusion

Recent clinical and preclinical studies have revealed that the combination of HSP90 inhibitors and other anti-cancer drugs is a viable method to re-introduce HSP90 inhibitors to anti-cancer therapy. Because many oncoproteins and drug resistance-related proteins are clients of HSP90, HSP90 inhibitors can cooperate well with various clinical anti-cancer therapies, including chemotherapy, targeted therapy, radiation therapy and immunotherapy, to improve the treatment efficacy and overcome drug resistance. Pioneering examples discussed in this review have broadened the prospects for the rational and effective use of HSP90 inhibitors in cancer treatment.

Since the initial discovery of HSP90 as a promising therapeutic target for anti-cancer therapy, tremendous progress has been made in the identification of numerous HSP90 inhibitors. Because HSP90 is required for stabilising many oncoproteins, HSP90 inhibitors often have strong and broad-spectrum anti-cancer activities. Although HSP90 inhibitors have demonstrated effectiveness in both *in vitro* and *in vivo* models, clinical trials investigating their use as monotherapy have reported unsatisfactory results, which may be attributed to organ toxicity and heat shock response induced by the inhibitors. Therefore, developing robust strategies for the successful application of HSP90 inhibitors in anti-cancer therapy is necessary.

Combination therapy represents an important therapeutic strategy for improving treatment efficacy and safety. It involves the combined use of two or more therapies with different but synergistic pharmacological mechanisms of action. Theoretically, HSP90 inhibitors can act synergistically with any anti-cancer therapy given that the mechanism

of action relies on the inhibition of the client proteins of HSP90. Inhibition of HSP90 results in the degradation of many drug resistance-related proteins; therefore, combination therapy involving HSP90 inhibitors may represent an effective strategy for overcoming drug resistance in advanced and refractory cancers. Unlike monotherapy, combination therapy allows the use of lower doses of each drug, which can help to reduce the intrinsic toxicity of HSP90 inhibitors. Based on the above-mentioned reasons, the combined use of HSP90 inhibitors and other anti-cancer therapies is considered an alternative to monotherapy for cancer treatment. In the past decade, many clinical and preclinical studies have evaluated different combinations of HSP90 inhibitors and other anti-cancer therapies. Some combinations exhibited positive therapeutic effects in patients with cancer, demonstrating their promising therapeutic potential, whereas some combinations demonstrated no therapeutic benefit in clinical trials. Therefore, the rational design of combination therapies involving HSP90 inhibitors should be improved, and the efficacy, toxicity, dosage and administration of these combination regimens should be continuously investigated in clinical trials.

Given that numerous proteins have been identified as clients of HSP90, inhibition of HSP90 may lead to complicated changes in signalling in cancer cells. The biological function of HSP90 warrants further investigation. Establishing a comprehensive map of HSP90 client proteins is important for rationalising combination strategies involving HSP90 inhibitors. Given that many drugs targeting HSP90 clients have not yet been evaluated in combination with HSP90 inhibitors, expanding combination therapies to clinical trials with different therapeutic targets and indications may help to realise the complete potential of HSP90 inhibitors in combination therapy.

In combination therapies reported to date, HSP90 inhibitors have been used as 'adjuvant' supporting other anti-cancer therapies to achieve the goals of enhancing efficacy or overcoming resistance. On the contrary, other anti-cancer therapies have been rarely used to improve the efficacy and safety of HSP90 inhibitors. Therefore, drugs that can negate the side effects of HSP90 inhibitors, such as organ toxicity and heat shock response, should be developed for promoting the use of HSP90 inhibitors as a 'striker' in cancer treatment. For example, the CDK inhibitor AT7519 can inhibit the compensatory expression of HSP70 induced by onalespib, suggesting a novel method to overcome the heat shock response.

HSP90 inhibitors used in existing combination therapies predominantly target the N-terminus of HSP90. Over the past several years, substantial progress has been made in the development of C-terminal and isoform-specific inhibitors of HSP90, which have a better safety profile and (or) do not induce heat shock response. These novel HSP90 inhibitors

may open broader avenues for the development of effective anti-cancer therapies. More combination therapies involving HSP90 inhibitors should be developed and investigated in clinical and preclinical trials to improve cancer treatment in the future.

Acknowledgements This work is funded by the National Key Research and Development Program [Grant No. 2017YFC1702006], the Natural Science Foundation of Liaoning Province of China [Grant No. 2020-MS-105], and the Fundamental Research Funds for the Central Universities [Grant No. DUT22YG111].

Author contributions Yajun Liu and Chenyao Li prepared the original manuscript; Yajun Liu and Shutao Tan revised the manuscript. Yajun Liu, Shutao Tan and Hongwei Liu conceived the manuscript and polished writing.

Data availability Not applicable.

Declarations

Conflict of interest The authors declare no conflicts of interest.

References

- Abu Lila AS, Kato C, Fukushima M, Huang CL, Wada H, Ishida T (2016) Downregulation of thymidylate synthase by RNAi molecules enhances the antitumor effect of pemetrexed in an orthotopic malignant mesothelioma xenograft mouse model. *Int J Oncol* 48(4):1399–1407. <https://doi.org/10.3892/ijo.2016.3367>
- Albadari N, Deng S, Li W (2019) The transcriptional factors HIF-1 and HIF-2 and their novel inhibitors in cancer therapy. *Expert Opin Drug Discov* 14(7):667–682. <https://doi.org/10.1080/17460441.2019.1613370>
- Ali MMU, Roe SM, Vaughan CK, Meyer P, Panaretou B, Piper PW, Prodromou C, Pearl LH (2006) Crystal structure of an Hsp90-nucleotide-p23/Sba1 closed chaperone complex. *Nature* 440(7087):1013–1017. <https://doi.org/10.1038/nature04716>
- Bagatell R, Whitesell L (2004) Altered Hsp90 function in cancer: a unique therapeutic opportunity. *Mol Cancer Ther* 3(8):1021–1030. <https://doi.org/10.1097/01.cmr.0000138826.11538.5e>
- Bai XP, Ni J, Beretov J, Graham P, Li Y (2018) Cancer stem cell in breast cancer therapeutic resistance. *Cancer Treat Rev* 69:152–163. <https://doi.org/10.1016/j.ctrv.2018.07.004>
- Biondini M, Kiepas A, El-Houjeiri L, Annis MG, Hsu BE, Fortier AM, Morin G, Martina JA, Sirois I, Aguilar-Mahecha A, Gruosso T, McQuirk S, Rose AAN, Tokat UM, Johnson RM, Sahin O, Bareke E, St-Pierre J, Park M, Basik M, Majewski J, Puertollano R, Pause A, Huang S, Keler T, Siegel PM (2022) HSP90 inhibitors induce GPNMB cell-surface expression by modulating lysosomal positioning and sensitize breast cancer cells to glematumumab vedotin. *Oncogene* 41(12):1701–1717. <https://doi.org/10.1038/s41388-022-02206-z>
- Busenius J, Blazey CM, Aay N, Anand NK, Arcalas A, Baik T, Bowles OJ, Buhr CA, Costanzo S, Curtis JK, DeFina SC, Dubenko L, Heuer TS, Huang P, Jaeger C, Joshi A, Kennedy AR, Kim AI, Lara K, Lee J, Li J, Loughheed JC, Ma S, Malek S, Manalo JC, Martini JF, McGrath G, Nicoll M, Nuss JM, Pack M, Peto CJ, Tsang TH, Wang L, Womble SW, Yakes M, Zhang W, Rice KD (2012) Discovery of XL888: a novel tropane-derived small molecule inhibitor of HSP90. *Bioorg Med Chem Lett* 22(17):5396–5404. <https://doi.org/10.1016/j.bmcl.2012.07.052>
- Byrd JC, O'Brien S, James DF (2013) Ibrutinib in relapsed chronic lymphocytic leukemia. *New Engl J Med* 369(13):1278–1279. <https://doi.org/10.1056/NEJMc1309710>
- Cavenagh J, Oakervee H, Baetiong-Caguioa P, Davies F, Gharibo M, Rabin N, Kurman M, Novak B, Shirraishi N, Nakashima D, Akinaga S, Yong K (2017) A phase I/II study of KW-2478, an Hsp90 inhibitor, in combination with bortezomib in patients with relapsed/refractory multiple myeloma. *Br J Cancer* 117(9):1295–1302. <https://doi.org/10.1038/bjc.2017.302>
- Cercek A, Shia J, Gollub M, Chou JF, Capanu M, Raasch P, Reidy-Lagunes D, Proia DA, Vakiani E, Solit DB, Saltz LB (2014) Ganetespib, a novel Hsp90 inhibitor in patients with KRAS mutated and wild type, refractory metastatic colorectal cancer. *Clin Colorectal Cancer* 13(4):207–212. <https://doi.org/10.1016/j.clcc.2014.09.001>
- Chen H, Gong Y, Ma Y, Thompson RC, Wang J, Cheng Z, Xue L (2020) A brain-penetrating Hsp90 inhibitor NXD30001 inhibits glioblastoma as a monotherapy or in combination with radiation. *Front Pharmacol* 11:974. <https://doi.org/10.3389/fphar.2020.00974>
- Chen TML, Harrington B, Truxall J, Wasmuth R, Prouty A, Sloan S, Lehman AM, Sampath D, Orlemans E, Baiocchi RA, Alinari L, Byrd JC, Woyach JA, Hertlein E (2021) Preclinical evaluation of the Hsp90 inhibitor SNX-5422 in ibrutinib resistant CLL. *J Hematol Oncol* 14(1):36. <https://doi.org/10.1186/S13045-021-01039-9>
- Chettiar ST, Malek R, Annadanam A, Nugent KM, Kato Y, Wang HL, Cades JA, Taparra K, Belcaid Z, Ballew M, Manmiller S, Proia D, Lim M, Anders RA, Herman JM, Tran PT (2016) Ganetespib radiosensitization for liver cancer therapy. *Cancer Biol Ther* 17(4):457–466. <https://doi.org/10.1080/15384047.2016.1156258>
- Choi YE, Battelli C, Watson J, Liu J, Curtis J, Morse AN, Matulonis UA, Chowdhury D, Konstantinopoulos PA (2014) Sublethal concentrations of 17-AAG suppress homologous recombination DNA repair and enhance sensitivity to carboplatin and olaparib in HR proficient ovarian cancer cells. *Oncotarget* 5(9):2678–2687. <https://doi.org/10.18632/oncotarget.1929>
- Chon J, Stover PJ, Field MS (2017) Targeting nuclear thymidylate biosynthesis. *Mol Aspects Med* 53:48–56. <https://doi.org/10.1016/j.mam.2016.11.005>
- Chong CR, Jänne PA (2013) The quest to overcome resistance to EGFR-targeted therapies in cancer. *Nat Med* 19(11):1389–1400. <https://doi.org/10.1038/nm.3388>
- Codony-Servat J, Viteri S, Codony-Servat C, Ito M, Bracht JWP, Berenguer J, Chaib I, Molina-Vila MA, Karachaliou N, Rosell R (2019) Hsp90 inhibitors enhance the antitumoral effect of osimertinib in parental and osimertinib-resistant non-small cell lung cancer cell lines. *Transl Lung Cancer Res* 8(4):340–351. <https://doi.org/10.21037/tlcr.2019.08.22>
- Cohen P, Cross D, Jänne PA (2021) Kinase drug discovery 20 years after imatinib: progress and future directions. *Nat Rev Drug Discov* 20(7):551–569. <https://doi.org/10.1038/s41573-021-00195-4>
- De Raedt T, Walton Z, Yecies JL, Li DA, Chen YM, Malone CF, Maertens O, Jeong SM, Bronson RT, Lebleu V, Kalluri R, Normant E, Haigis MC, Manning BD, Wong KK, Macleod KF, Cichowski K (2011) Exploiting cancer cell vulnerabilities to develop a combination therapy for Ras-driven tumors. *Cancer Cell* 20(3):400–413. <https://doi.org/10.1016/j.ccr.2011.08.014>
- Demetri GD, Heinrich MC, Chmielowski B, Morgan JA, George S, Bradley R, Blackman RK, Teofilovici F, Fletcher JA, Tap WD, von Mehren M (2011) An open-label phase II study of the Hsp90 inhibitor ganetespib (STA-9090) in patients (pts) with metastatic and/or unresectable GIST. *J Clin Oncol* 29(15):10011. https://doi.org/10.1200/jco.2011.29.15_suppl.10011
- Derakhshani A, Rezaei Z, Safarpour H, Sabri M, Mir A, Sanati MA, Vahidian F, Gholamiyan Moghadam A, Aghadokht A,

- Hajiasgharzadeh K, Baradaran B (2020) Overcoming trastuzumab resistance in HER2-positive breast cancer using combination therapy. *J Cell Physiol* 235(4):3142–3156. <https://doi.org/10.1002/jcp.29216>
- Do K, Speranza G, Chang LC, Polley EC, Bishop R, Zhu WM, Trepel JB, Lee S, Lee MJ, Kinders RJ, Phillips L, Collins J, Lyons J, Jeong W, Antony R, Chen AP, Neckers L, Doroshow JH, Kummar S (2015) Phase I study of the heat shock protein 90 (Hsp90) inhibitor onalespib (AT13387) administered on a daily for 2 consecutive days per week dosing schedule in patients with advanced solid tumors. *Investig New Drug* 33(4):921–930. <https://doi.org/10.1007/s10637-015-0255-1>
- Do KT, Coyne GO, Hays JL, Supko JG, Liu SPV, Beebe K, Neckers L, Trepel JB, Lee MJ, Smyth T, Gannon C, Hedglin J, Muzikansky A, Campos S, Lyons J, Ivy P, Doroshow JH, Chen ALP, Shapiro GI (2020) Phase I study of the HSP90 inhibitor onalespib in combination with AT7519, a pan-CDK inhibitor, in patients with advanced solid tumors. *Cancer Chemother Pharmacol* 86(6):815–827. <https://doi.org/10.1007/s00280-020-04176-z>
- Doi T, Kurokawa Y, Sawaki A, Komatsu Y, Ozaka M, Takahashi T, Naito Y, Ohkubo S, Nishida T (2019) Efficacy and safety of TAS-116, an oral inhibitor of heat shock protein 90, in patients with metastatic or unresectable gastrointestinal stromal tumour refractory to imatinib, sunitinib and regorafenib: a phase II, single-arm trial. *Eur J Cancer* 121:29–39. <https://doi.org/10.1016/j.ejca.2019.08.009>
- Donnelly A, Blagg BSJ (2008) Novobiocin and additional inhibitors of the Hsp90 C-terminal nucleotide-binding pocket. *Curr Med Chem* 15(26):2702–2717. <https://doi.org/10.2174/092986708786242895>
- Erllichman C (2009) Tanespimycin: the opportunities and challenges of targeting heat shock protein 90. *Expert Opin Investig Drugs* 18(6):861–868. <https://doi.org/10.1517/13543780902953699>
- Ernst A, Hennel R, Krombach J, Kapfhammer H, Brix N, Zuchtriegel G, Uhl B, Reichel CA, Frey B, Gaipf US, Winssinger N, Shirasawa S, Sasazuki T, Sperandio M, Belka C, Lauber K (2020) Priming of anti-tumor immune mechanisms by radiotherapy is augmented by inhibition of heat shock protein 90. *Front Oncol* 10:1668. <https://doi.org/10.3389/fonc.2020.01668>
- Eroglu Z, Chen YA, Gibney GT, Weber JS, Kudchadkar RR, Khushalani NI, Markowitz J, Brohl AS, Tetteh LF, Ramadan H, Arnone G, Li JN, Zhao XH, Sharma R, Darville LNF, Fang B, Smalley I, Messina JL, Koomen JM, Sondak VK, Smalley KSM (2018) Combined BRAF and HSP90 inhibition in patients with unresectable BRAF(V600E)-mutant melanoma. *Clin Cancer Res* 24(22):5516–5524. <https://doi.org/10.1158/1078-0432.CCR-18-0565>
- Ewers KM, Patil S, Kopp W, Thomale J, Quilitz T, Magerhans A, Wang X, Hessmann E, Dobbelsstein M (2021) HSP90 inhibition synergizes with cisplatin to eliminate basal-like pancreatic ductal adenocarcinoma cells. *Cancers* 13(24):6163. <https://doi.org/10.3390/Cancers13246163>
- Fennell D, Danson S, Forster M, Talbot D, Woll P, Child J, Ngai Y, Farrelly L, Hackshaw A, Sharkey A, Busacca S, Hastings R, Barnes D, Nicolson M, Taylor P, Ahmed S, Wheeler G (2018) Phase I study of HSP90 inhibitor ganetespib with pemetrexed and cisplatin/carboplatin chemotherapy for pleural mesothelioma. *J Thorac Oncol* 13(10):S397. <https://doi.org/10.1016/j.jtho.2018.08.415>
- Ferraldeschi R, Welti J, Powers MV, Yuan W, Smyth T, Seed G, Riisnaes R, Hedayat S, Wang H, Crespo M, Rodrigues DN, Figueiredo I, Miranda S, Carreira S, Lyons JF, Sharp S, Plymate SR, Attard G, Wallis N, Workman P, de Bono JS (2016) Second-generation HSP90 inhibitor onalespib blocks mRNA splicing of androgen receptor variant 7 in prostate cancer cells. *Cancer Res* 76(9):2731–2742. <https://doi.org/10.1158/0008-5472.CAN-15-2186>
- Friedman JA, Wise SC, Hu M, Gouveia C, Vander Broek R, Freudl-sperger C, Kannabiran VR, Arun P, Mitchell JB, Chen Z, Van Waes C (2013) HSP90 inhibitor SNX5422/2112 targets the dys-regulated signal and transcription factor network and malignant phenotype of head and neck squamous cell carcinoma. *Transl Oncol* 6(4):429–441. <https://doi.org/10.1593/tlo.13292>
- Ge F, Wang Y, Sharma A, Yang Y, Liu H, Essler M, Jaehde U, Schmidt-Wolf IGH (2023) Cytokine-induced killer cells in combination with heat shock protein 90 inhibitors functioning via the Fas/FasL axis provides rationale for a potential clinical benefit in Burkitt's lymphoma. *Int J Mol Sci* 24(15):12476. <https://doi.org/10.3390/ijms241512476>
- Giulino-Roth L, van Besien HJ, Dalton T, Totonchy JE, Rodina A, Taldone T, Bolaender A, Erdjument-Bromage H, Sadek J, Chadburn A, Barth MJ, Dela Cruz FS, Rainey A, Kung AL, Chiosis G, Cesarman E (2017) Inhibition of Hsp90 suppresses PI3K/AKT/mTOR signaling and has antitumor activity in Burkitt lymphoma. *Mol Cancer Ther* 16(9):1779–1790. <https://doi.org/10.1158/1535-7163.MCT-16-0848>
- Graner MW (2016) HSP90 and immune modulation in cancer. *Adv Cancer Res* 129:191–224. <https://doi.org/10.1016/bs.acr.2015.10.001>
- Graner MW (2021) Making HSP90 inhibitors great again? Unite for better cancer immunotherapy. *Cell Chem Biol* 28(2):118–120. <https://doi.org/10.1016/j.chembiol.2021.02.002>
- Grbovic-Huezo O, Pitter KL, Lecomte N, Saglimbeni J, Askan G, Holm M, Melchor JP, Chandwani R, Joshi S, Haglund C, Iacobuzio-Donahue CA, Chiosis G, Tammela T, Leach SD (2020) Unbiased in vivo preclinical evaluation of anticancer drugs identifies effective therapy for the treatment of pancreatic adenocarcinoma. *Proc Natl Acad Sci USA* 117(48):30670–30678. <https://doi.org/10.1073/pnas.1920240117>
- Gutierrez M, Guo RB, Giaccone G, Liu SV, Hao ZL, Hilton C, Hinson JM, Kris MG, Orlemans EO, Drilon A (2021) Phase I multicenter study of the HSP90 inhibitor SNX-5422 plus carboplatin and paclitaxel in patients with lung cancers. *Lung Cancer* 162:23–28. <https://doi.org/10.1016/j.lungcan.2021.10.001>
- Haarberg HE, Paraiso KHT, Wood E, Rebecca VW, Sondak VK, Koomen JM, Smalley KSM (2013) Inhibition of Wee1, AKT, and CDK4 underlies the efficacy of the HSP90 inhibitor XL888 in an in vivo model of NRAS-mutant melanoma. *Mol Cancer Ther* 12(6):901–912. <https://doi.org/10.1158/1535-7163.MCT-12-1003>
- He J, Huang Z, Han L, Gong Y, Xie C (2021) Mechanisms and management of 3rd-generation EGFR-TKI resistance in advanced non-small cell lung cancer (review). *Int J Oncol* 59(5):90. <https://doi.org/10.3892/ijo.2021.5270>
- Hirano H, Takashima A, Hamaguchi T, Shida D, Kanemitsu Y (2021) Current status and perspectives of immune checkpoint inhibitors for colorectal cancer. *Jpn J Clin Oncol* 51(1):10–19. <https://doi.org/10.1093/jjco/hyaa200>
- Honma Y, Kurokawa Y, Sawaki A, Naito Y, Iwagami S, Baba H, Komatsu Y, Nishida T, Doi T (2021) Randomized, double-blind, placebo (PL)-controlled, phase III trial of pimitespib (TAS-116), an oral inhibitor of heat shock protein 90 (HSP90), in patients (pts) with advanced gastrointestinal stromal tumor (GIST) refractory to imatinib (IM), sunitinib (SU) and regorafenib (REG). *J Clin Oncol* 39(15):11524. https://doi.org/10.1200/Jco.2021.39.15_Suppl.11524
- Hoter A, El-Sabban ME, Naim HY (2018) The HSP90 family: structure, regulation, function, and implications in health and disease. *Int J Mol Sci* 19(9):2560. <https://doi.org/10.3390/ijms19092560>
- Huang KH, Veal JM, Fadden RP, Rice JW, Eaves J, Strachan JP, Barabasz AF, Foley BE, Barta TE, Ma W, Silinski MA, Hu M,

- Partridge JM, Scott A, DuBois LG, Freed T, Steed PM, Ommen AJ, Smith ED, Hughes PF, Woodward AR, Hanson GJ, McCall WS, Markworth CJ, Hinkley L, Jenks M, Geng LF, Lewis M, Otto J, Pronk B, Verleysen K, Hall SE (2009) Discovery of novel 2-aminobenzamide inhibitors of heat shock protein 90 as potent, selective and orally active antitumor agents. *J Med Chem* 52(14):4288–4305. <https://doi.org/10.1021/jm900230j>
- Huang XP, Gao LZ, Wang SL, McManaman JL, Thor AD, Yang XH, Esteva FJ, Liu BL (2010) Heterotrimerization of the growth factor receptors erbB2, erbB3, and insulin-like growth factor-I receptor in breast cancer cells resistant to herceptin. *Cancer Res* 70(3):1204–1214. <https://doi.org/10.1158/0008-5472.CAN-09-3321>
- Infante JR, Weiss GJ, Jones S, Tibes R, Bauer TM, Bendell JC, Hinson JM Jr, Von Hoff DD, Burris HA 3rd, Orlemans EO, Ramanaathan RK (2014) Phase I dose-escalation studies of SNX-5422, an orally bioavailable heat shock protein 90 inhibitor, in patients with refractory solid tumours. *Eur J Cancer* 50(17):2897–2904. <https://doi.org/10.1016/j.ejca.2014.07.017>
- Jhaveri K, Modi S (2012) HSP90 inhibitors for cancer therapy and overcoming drug resistance. *Adv Pharmacol* 65:471–517. <https://doi.org/10.1016/b978-0-12-397927-8.00015-4>
- Kabakov AE, Kudryavtsev VA, Gabai VL (2010) Hsp90 inhibitors as promising agents for radiotherapy. *J Mol Med* 88(3):241–247. <https://doi.org/10.1007/s00109-009-0562-0>
- Kale Ş, Korcum AF, Dündar E, Erin N (2020) HSP90 inhibitor PU-H71 increases radiosensitivity of breast cancer cells metastasized to visceral organs and alters the levels of inflammatory mediators. *Naunyn-Schmiedeberg's Arch Pharmacol* 393(2):253–262. <https://doi.org/10.1007/s00210-019-01725-z>
- Kawazoe A, Itahashi K, Yamamoto N, Kotani D, Kuboki Y, Taniguchi H, Harano K, Naito Y, Suzuki M, Fukutani M, Higuchi T, Ikeno T, Wakabayashi M, Sato A, Koyama S, Nishikawa H, Shitara K (2021) TAS-116 (pimitepsib), an oral HSP90 inhibitor, in combination with nivolumab in patients with colorectal cancer and other solid tumors: an open-label, dose-finding, and expansion phase Ib trial (EPOC1704). *Clin Cancer Res* 27(24):6709–6715. <https://doi.org/10.1158/1078-0432.CCR-21-1929>
- Kim BM, Hong Y, Lee S, Liu P, Lim JH, Lee YH, Lee TH, Chang KT, Hong Y (2015) Therapeutic implications for overcoming radiation resistance in cancer therapy. *Int J Mol Sci* 16(11):26880–26913. <https://doi.org/10.3390/ijms161125991>
- Kim A, Lu Y, Okuno SH, Reinke D, Maertens O, Perentesis J, Basu M, Wolters PL, De Raedt T, Chawla S, Chugh R, Van Tine BA, O'Sullivan G, Chen A, Cichowski K, Widemann BC (2020a) Targeting refractory sarcomas and malignant peripheral nerve sheath tumors in a phase I/II study of sirolimus in combination with ganetespib (SARC023). *Sarcoma* 2020:5784876. <https://doi.org/10.1155/2020/5784876>
- Kim HJ, Gong MK, Yoon CY, Kang J, Yun M, Cho NH, Rha SY, Choi YD (2020b) Synergistic antitumor effects of combined treatment with HSP90 inhibitor and PI3K/mTOR dual inhibitor in cisplatin-resistant human bladder cancer cells. *Yonsei Med J* 61(7):587–596. <https://doi.org/10.3349/ymj.2020.61.7.587>
- Konstantinopoulos PA, Cheng SC, Supko JG, Polak M, Wahner-Hendrickson AE, Ivy SP, Bowes B, Sawyer H, Basada P, Hayes M, Curtis J, Horowitz N, Wright AA, Campos SM, Ivanova EV, Pawletz CP, Palakurthi S, Liu JF, D'Andrea AD, Gokhale PC, Chowdhury D, Matulonis UA, Shapiro GI (2022) Combined PARP and HSP90 inhibition: preclinical and phase I evaluation in patients with advanced solid tumours. *Br J Cancer* 126(7):1027–1036. <https://doi.org/10.1038/s41416-021-01664-8>
- Kurokawa Y, Honma Y, Sawaki A, Naito Y, Iwagami S, Komatsu Y, Takahashi T, Nishida T, Doi T (2022) Pimitepsib in patients with advanced gastrointestinal stromal tumor (CHAPTER-GIST-301): a randomized, double-blind, placebo-controlled phase III trial. *Ann Oncol* 33(9):959–967. <https://doi.org/10.1016/j.annonc.2022.05.518>
- Kwak EL, Goyal L, Abrams TA, Carpenter A, Wolpin BM, Wadlow RC, Allen JN, Heist RS, McCleary NJ, Chan JA, Goessling W, Schrag D, Evans C, Ng K, Enzinger PC, Ryan DP (2013) A phase II clinical trial of ganetespib (STA-9090) in previously treated patients with advanced esophagogastric cancers. *J Clin Oncol* 31(15):4090. https://doi.org/10.1200/jco.2013.31.15_suppl.4090
- Lang BJ, Guerrero ME, Prince TL, Okusha Y, Bonorino C, Calderwood SK (2021) The functions and regulation of heat shock proteins; key orchestrators of proteostasis and the heat shock response. *Arch Toxicol* 95(6):1943–1970. <https://doi.org/10.1007/s00204-021-03070-8>
- Lang JE, Forero-Torres A, Yee D, Yau C, Wolf D, Park J, Parker BA, Chien AJ, Wallace AM, Murthy R, Albain KS, Ellis ED, Beckwith H, Haley BB, Elias AD, Boughey JC, Yung RL, Isaacs C, Clark AS, Han HS, Nanda R, Khan QJ, Edmiston KK, Stringer-Reasor E, Price E, Joe B, Liu MC, Brown-Swigart L, Petricoin EF, Wulfkühle JD, Buxton M, Clennell JL, Sanil A, Berry S, Asare SM, Wilson A, Hirst GL, Singhrao R, Asare AL, Matthews JB, Melisko M, Perlmutter J, Rugo HS, Symmans WF, van't Veer LJ, Hylton NM, DeMichele AM, Berry DA, Esserman LJ (2022) Safety and efficacy of HSP90 inhibitor ganetespib for neoadjuvant treatment of stage II/III breast cancer. *npj Breast Cancer* 8(1):128. <https://doi.org/10.1038/s41523-022-00493-z>
- Lee HJ, Shin S, Kang J, Han KC, Kim YH, Bae JW, Park KH (2020) HSP90 inhibitor, 17-DMAG, alone and in combination with lapatinib attenuates acquired lapatinib-resistance in ER-positive, HER2-overexpressing breast cancer cell line. *Cancers (Basel)* 12(9):2630. <https://doi.org/10.3390/cancers12092630>
- Li L, Wang L, You QD, Xu XL (2020) Heat shock protein 90 inhibitors: an update on achievements, challenges, and future directions. *J Med Chem* 63(5):1798–1822. <https://doi.org/10.1021/acs.jmedchem.9b00940>
- Li L, Chen NN, You QD, Xu XL (2021) An updated patent review of anticancer Hsp90 inhibitors (2013-present). *Expert Opin Ther Pat* 31(1):67–80. <https://doi.org/10.1080/13543776.2021.1829595>
- Liu ZJ, Liu J, Zhang TM, Shi MX, Chen XF, Chen Y, Yu J (2020) Destabilization of ROR1 enhances activity of Ibrutinib against chronic lymphocytic leukemia in vivo. *Pharmacol Res* 151:104512. <https://doi.org/10.1016/j.phrs.2019.104512>
- Liu L, Deng Y, Zheng Z, Deng Z, Zhang J, Li J, Liang M, Zhou X, Tan W, Yang H, Neckers LM, Zou F, Chen X (2021) Hsp90 inhibitor STA9090 sensitizes hepatocellular carcinoma to hyperthermia-induced DNA damage by suppressing DNA-PKcs protein stability and mRNA transcription. *Mol Cancer Ther* 20(10):1880–1892. <https://doi.org/10.1158/1535-7163.mct-21-0215>
- Lu Z, Wang Z, Tu Z, Liu H (2022) HSP90 inhibitor ganetespib enhances the sensitivity of mantle cell lymphoma to Bruton's tyrosine kinase inhibitor ibrutinib. *Front Pharmacol* 13:864194. <https://doi.org/10.3389/fphar.2022.864194>
- Lundsten S, Spiegelberg D, Raval NR, Nestor M (2020) The radiosensitizer Onalespib increases complete remission in (177)Lu-DOTATATE-treated mice bearing neuroendocrine tumor xenografts. *Eur J Nucl Med Mol Imaging* 47(4):980–990. <https://doi.org/10.1007/s00259-019-04673-1>
- Marine JC, Francoz S, Maetens M, Wahl G, Toledo F, Lozano G (2006) Keeping p53 in check: essential and synergistic functions of Mdm2 and Mdm4. *Cell Death Differ* 13(6):927–934. <https://doi.org/10.1038/sj.cdd.4401912>
- Mathieu C, Messaoudi S, Fattal E, Vergnaud-Gauchon J (2019) Cancer drug resistance: rationale for drug delivery systems and targeted inhibition of HSP90 family proteins. *Cancer Drug Resist (Alhambra, Calif)* 2(3):381–398. <https://doi.org/10.20517/cdr.2019.26>

- Mbofung RM, McKenzie JA, Malu S, Zhang M, Peng WY, Liu CW, Kuitatse I, Tieu T, Williams L, Devi S, Ashkin E, Xu CY, Huang L, Zhang MY, Talukder AH, Tripathi SC, Khong H, Satani N, Muller FL, Roszik J, Heffernan T, Allison JP, Lizee G, Hanash SM, Proia D, Amaria R, Davis RE, Hwu P (2017) HSP90 inhibition enhances cancer immunotherapy by upregulating interferon response genes. *Nat Commun* 8(1):451. <https://doi.org/10.1038/s41467-017-00449-z>
- McCombs JR, Chang HP, Shah DK, Owen SC (2021) Antibody-drug conjugate and free geldanamycin combination therapy enhances anti-cancer efficacy. *Int J Pharm* 610:121272. <https://doi.org/10.1016/j.ijpharm.2021.121272>
- Mehta RK, Pal S, Kondapi K, Sitto M, Dewar C, Devasia T, Schipper MJ, Thomas DG, Basur V, Pai MP, Morishima Y, Osawa Y, Pratt WB, Lawrence TS, Nyati MK (2020) Low-dose Hsp90 inhibitor selectively radiosensitizes HNSCC and pancreatic xenografts. *Clin Cancer Res* 26(19):5246–5257. <https://doi.org/10.1158/1078-0432.ccr-19-3102>
- Meister S, Schubert U, Neubert K, Herrmann K, Burger R, Gramatzki M, Hahn S, Schreiber S, Wilhelm S, Herrmann M, Jack HM, Voll RE (2007) Extensive immunoglobulin production sensitizes myeloma cells for proteasome inhibition. *Cancer Res* 67(4):1783–1792. <https://doi.org/10.1158/0008-5472.CAN-06-2258>
- Miyata Y, Nakamoto H, Neckers L (2013) The therapeutic target Hsp90 and cancer hallmarks. *Curr Pharm Des* 19(3):347–365. <https://doi.org/10.2174/138161213804143725>
- Mohajershaj T, Jha P, Boström A, Frejd FY, Yazaki PJ, Nestor M (2022) In vitro characterization of (177)Lu-DOTA-M5A anti-carcinoembryonic antigen humanized antibody and HSP90 inhibition for potentiated radioimmunotherapy of colorectal cancer. *Front Oncol* 12:849338. <https://doi.org/10.3389/fonc.2022.849338>
- Mooradian MJ, Cleary JM, Giobbie-Hurder A, Darville LNF, Parikh A, Buchbinder EI, Cohen JV, Lawrence DP, Shapiro GI, Keer H, Chen HX, Ivy SP, Smalley KSM, Koomen JM, Sullivan RJ (2023) Dose-escalation trial of combination dabrafenib, trametinib, and AT13387 in patients with BRAF-mutant solid tumors. *Cancer* 129(12):1904–1918. <https://doi.org/10.1002/ncr.34730>
- Mortensen ACL, Mohajershaj T, Hariri M, Pettersson M, Spiegelberg D (2020) Overcoming limitations of cisplatin therapy by additional treatment with the HSP90 inhibitor onalespib. *Front Oncol* 10:532285. <https://doi.org/10.3389/Fonc.2020.532285>
- Mosca L, Ilari A, Fazi F, Assaraf YG, Colotti G (2021) Taxanes in cancer treatment: activity, chemoresistance and its overcoming. *Drug Resist Updat* 54:100742. <https://doi.org/10.1016/j.drug.2020.100742>
- Nagaraju GP, Alese OB, Landry J, Diaz R, El-Rayes BF (2014) HSP90 inhibition downregulates thymidylate synthase and sensitizes colorectal cancer cell lines to the effect of 5FU-based chemotherapy. *Oncotarget* 5(20):9980–9991. <https://doi.org/10.18632/oncotarget.2484>
- Nagaraju GP, Zakka KM, Landry JC, Shaib WL, Lesinski GB, El-Rayes BF (2019) Inhibition of HSP90 overcomes resistance to chemotherapy and radiotherapy in pancreatic cancer. *Int J Cancer* 145(6):1529–1537. <https://doi.org/10.1002/ijc.32227>
- Namba R, Young LJ, Abbey CK, Kim L, Damonte P, Borowsky AD, Qi JY, Tepper CG, MacLeod CL, Cardiff RD, Gregg JP (2006) Rapamycin inhibits growth of premalignant and malignant mammary lesions in a mouse model of ductal carcinoma in situ. *Clin Cancer Res* 12(8):2613–2621. <https://doi.org/10.1158/1078-0432.CCR-05-2170>
- Nassar ZD, Mah CY, Centenera MM, Irani S, Sadowski MC, Scott JS, Nguyen EV, Nagarajan SR, Moldovan M, Lynn DJ, Daly RJ, Hoy AJ, Butler LM (2020) Fatty acid oxidation is an adaptive survival pathway induced in prostate tumors by HSP90 inhibition. *Mol Cancer Res* 18(10):1500–1511. <https://doi.org/10.1158/1541-7786.mcr-20-0570>
- Naz S, Leiker AJ, Choudhuri R, Preston O, Sowers AL, Gohain S, Gamson J, Mathias A, Van Waes C, Cook JA, Mitchell JB (2021) Pharmacological inhibition of HSP90 radiosensitizes head and neck squamous cell carcinoma xenograft by inhibition of DNA damage repair, nucleotide metabolism, and radiation-induced tumor vasculogenesis. *Int J Radiat Oncol* 110(5):1295–1305. <https://doi.org/10.1016/j.ijrobp.2021.03.048>
- Nguyen DM, Lorang D, Chen GA, Stewart JH, Tabibi E, Schrupp DS (2001) Enhancement of paclitaxel-mediated cytotoxicity in lung cancer cells by 17-allylamino geldanamycin: in vitro and in vivo analysis. *Ann Thorac Surg* 72(2):371–378. [https://doi.org/10.1016/S0003-4975\(01\)02787-4](https://doi.org/10.1016/S0003-4975(01)02787-4)
- Nowosielska EM, Cheda A, Pocięgiel M, Cheda L, Szymański P, Wiedlocha A (2021) Effects of a unique combination of the whole-body low dose radiotherapy with inactivation of two immune checkpoints and/or a heat shock protein on the transplantable lung cancer in mice. *Int J Mol Sci* 22(12):6309. <https://doi.org/10.3390/ijms22126309>
- Ohkubo S, Kodama Y, Muraoka H, Hitotsumachi H, Yoshimura C, Kitade M, Hashimoto A, Ito K, Gomori A, Takahashi K, Shibata Y, Kanoh A, Yonekura K (2015) TAS-116, a highly selective inhibitor of heat shock protein 90 α and β , demonstrates potent antitumor activity and minimal ocular toxicity in preclinical models. *Mol Cancer Ther* 14(1):14–22. <https://doi.org/10.1158/1535-7163.mct-14-0219>
- Orth M, Albrecht V, Seidl K, Kinzel L, Unger K, Hess J, Kreutzer L, Sun N, Stegen B, Nieto A, Maas J, Winssinger N, Friedl AA, Walch AK, Belka C, Zitzelsberger H, Niyazi M, Lauber K (2021) Inhibition of HSP90 as a strategy to radiosensitize glioblastoma: targeting the DNA damage response and beyond. *Front Oncol* 11:612354. <https://doi.org/10.3389/fonc.2021.612354>
- Paik J (2021) Olaparib: a review as first-line maintenance therapy in advanced ovarian cancer. *Target Oncol* 16(6):847–856. <https://doi.org/10.1007/s11523-021-00842-1>
- Paraiso KHT, Haarberg HE, Wood E, Rebecca VW, Chen YA, Xiang Y, Ribas A, Lo RS, Weber JS, Sondak VK, John JK, Sarnaik AA, Koomen JM, Smalley KSM (2012) The HSP90 inhibitor XL888 overcomes BRAF inhibitor resistance mediated through diverse mechanisms. *Clin Cancer Res* 18(9):2502–2514. <https://doi.org/10.1158/1078-0432.CCR-11-2612>
- Parsels LA, Engelke CG, Parsels J, Flanagan SA, Zhang Q, Tanska D, Wahl DR, Canman CE, Lawrence TS, Morgan MA (2021) Combinatorial efficacy of olaparib with radiation and ATR inhibitor requires PARP1 protein in homologous recombination-proficient pancreatic cancer. *Mol Cancer Ther* 20(2):263–273. <https://doi.org/10.1158/1535-7163.MCT-20-0365>
- Pennisi R, Ascenzi P, di Masi A (2016) Hsp90: a new player in DNA repair? *Biomolecules* 5(4):2589–2618. <https://doi.org/10.3390/biom5042589>
- Pillai RN, Fennell DA, Kovcin V, Ciuleanu TE, Ramlau R, Kowalski D, Schenker M, Yalcin I, Teofilovici F, Vukovic VM, Ramalingam SS (2020) Randomized phase III study of ganetespib, a heat shock protein 90 inhibitor, with docetaxel versus docetaxel in advanced non-small-cell lung cancer (GALAXY-2). *J Clin Oncol* 38(6):613–622. <https://doi.org/10.1200/jco.19.00816>
- Proia DA, Bates RC (2014) Ganetespib and HSP90: translating preclinical hypotheses into clinical promise. *Cancer Res* 74(5):1294–1300. <https://doi.org/10.1158/0008-5472.CAN-13-3263>
- Rahmy S, Mishra SJ, Murphy S, Blass BSJ, Lu X (2022) Hsp90 β inhibition upregulates interferon response and enhances immune checkpoint blockade therapy in murine tumors. *Front Immunol* 13:1005045. <https://doi.org/10.3389/fimmu.2022.1005045>
- Rajan A, Kelly RJ, Trepel JB, Kim YS, Alarcon SV, Kummar S, Gutierrez M, Crandon S, Zein WM, Jain L, Mannargudi B, Figg

- WD, Houk BE, Shnaidman M, Brega N, Giaccone G (2011) A phase I study of PF-04929113 (SNX-5422), an orally bioavailable heat shock protein 90 inhibitor, in patients with refractory solid tumor malignancies and lymphomas. *Clin Cancer Res* 17(21):6831–6839. <https://doi.org/10.1158/1078-0432.CCR-11-0821>
- Ramalingam S, Goss G, Rosell R, Schmid-Bindert G, Zaric B, Andric Z, Bondarenko I, Komov D, Ceric T, Khuri F, Samarzija M, Felip E, Ciuleanu T, Hirsh V, Wehler T, Spicer J, Salgia R, Shapiro G, Sheldon E, Teofilovici F, Vukovic V, Fennell D (2015) A randomized phase II study of ganetespib, a heat shock protein 90 inhibitor, in combination with docetaxel in second-line therapy of advanced non-small cell lung cancer (GALAXY-1). *Ann Oncol* 26(8):1741–1748. <https://doi.org/10.1093/annonc/mdv220>
- Rao A, Taylor JL, Chi-Sabins N, Kawabe M, Gooding WE, Storkus WJ (2012) Combination therapy with HSP90 inhibitor 17-DMAG reconditions the tumor microenvironment to improve recruitment of therapeutic T cells. *Cancer Res* 72(13):3196–3206. <https://doi.org/10.1158/0008-5472.CAN-12-0538>
- Ray-Coquard I, Braicu I, Berger R, Mahner S, Sehouli J, Pujade-Lauraine E, Cassier PA, Moll UM, Ulmer H, Leunen K, Zeimet AG, Marth C, Vergote I, Concin N, Ins GCP (2019) Part I of GANNET53: a European multicenter phase I/II trial of the Hsp90 inhibitor ganetespib combined with weekly paclitaxel in women with high-grade, platinum-resistant epithelial ovarian cancer—a study of the GANNET53 Consortium. *Front Oncol* 9:832. <https://doi.org/10.3389/Fonc.2019.00832>
- Reddy N, Voorhees PM, Houk BE, Brega N, Hinson JM, Jillela A (2013) Phase I trial of the HSP90 inhibitor PF-04929113 (SNX5422) in adult patients with recurrent, refractory hematologic malignancies. *Clin Lymphoma Myeloma Leuk* 13(4):385–391. <https://doi.org/10.1016/j.clml.2013.03.010>
- Ren X, Li T, Zhang W, Yang X (2022) Targeting heat-shock protein 90 in cancer: an update on combination therapy. *Cells* 11(16):2556. <https://doi.org/10.3390/cells11162556>
- Rice MA, Malhotra SV, Stoyanova T (2019) Second-generation antiandrogens: from discovery to standard of care in castration resistant prostate cancer. *Front Oncol* 9:801. <https://doi.org/10.3389/Fonc.2019.00801>
- Rice MA, Kumar V, Tailor D, Garcia-Marques FJ, Hsu EC, Liu SQ, Bermudez A, Kanchustambham V, Shankar V, Inde Z, Alabi BR, Muruganantham A, Shen M, Pandrala M, Nolley R, Aslan M, Ghoochani A, Agarwal A, Backup M, Kumar M, Going CC, Peehl DM, Dixon SJ, Zare RN, Brooks JD, Pitteri SJ, Malhotra SV, Stoyanova T (2022) SU086, an inhibitor of HSP90, impairs glycolysis and represents a treatment strategy for advanced prostate cancer. *Cell Rep Med* 3(2):100502. <https://doi.org/10.1016/J.Xcrm.2021.100502>
- Rodrigues Moita AJ, Bandolik JJ, Hansen FK, Kurz T, Hamacher A, Kassack MU (2020) Priming with HDAC inhibitors sensitizes ovarian cancer cells to treatment with cisplatin and HSP90 inhibitors. *Int J Mol Sci* 21(21):8300. <https://doi.org/10.3390/ijms21218300>
- Roh JL, Kim EH, Park HB, Park JY (2013) The Hsp90 inhibitor 17-(allylamino)-17-demethoxy geldanamycin increases cisplatin antitumor activity by inducing p53-mediated apoptosis in head and neck cancer. *Cell Death Dis* 4:e956. <https://doi.org/10.1038/cddis.2013.488>
- Rose AAN, Biondini M, Curiel R, Siegel PM (2017) Targeting GPNMB with glembatumumab vedotin: current developments and future opportunities for the treatment of cancer. *Pharmacol Ther* 179:127–141. <https://doi.org/10.1016/j.pharmthera.2017.05.010>
- Rossi G, Alama A, Genova C, Rijavec E, Tagliamento M, Biello F, Coco S, Dal Bello MG, Boccardo S, Grossi F (2018) The evolving role of pemetrexed disodium for the treatment of non-small cell lung cancer. *Expert Opin Pharmacother* 19(17):1969–1976. <https://doi.org/10.1080/14656566.2018.1536746>
- Sabbah DA, Hajjo R, Sweidan K (2020) Review on epidermal growth factor receptor (EGFR) structure, signaling pathways, interactions, and recent updates of EGFR inhibitors. *Curr Top Med Chem* 20(10):815–834. <https://doi.org/10.2174/1568026620666200303123102>
- Saber S, El-Fattah EEA, Abdelhamid AM, Mourad AAE, Hamouda MAM, Elrabat A, Zakaria S, Haleem AA, Mohamed SZ, Elgharabawy RM, Morsy NE, El Adle KN, Mohammed OA, El-Bahouty WB, Mostafa SA, Abdelhady R, Galal O, ElSaid ZH, Yahya G, Shata A, Youssef ME (2023a) Innovative challenge for the inhibition of hepatocellular carcinoma progression by combined targeting of HSP90 and STAT3/HIF-1 α signaling. *Biomed Pharmacother* 158:114196. <https://doi.org/10.1016/j.biopha.2022.114196>
- Saber S, Hasan AM, Mohammed OA, Saleh LA, Hashish AA, Alamri MMS, Al-Ameer AY, Alfaifi J, Senbel A, Aboregela AM, Khalid TBA, Abdel-Reheim MA, Cavalu S (2023b) Ganetespib (STA-9090) augments sorafenib efficacy via necroptosis induction in hepatocellular carcinoma: implications from preclinical data for a novel therapeutic approach. *Biomed Pharmacother* 164:114918. <https://doi.org/10.1016/j.biopha.2023.114918>
- Saito Y, Takahashi T, Obata Y, Nishida T, Ohkubo S, Nakagawa F, Serada S, Fujimoto M, Ohkawara T, Nishigaki T, Sugase T, Koh M, Ishida T, Tanaka K, Miyazaki Y, Makino T, Kurokawa Y, Nakajima K, Yamasaki M, Hirota S, Naka T, Mori M, Doki Y (2020) TAS-116 inhibits oncogenic KIT signalling on the Golgi in both imatinib-naive and imatinib-resistant gastrointestinal stromal tumours. *Br J Cancer* 122(5):658–667. <https://doi.org/10.1038/s41416-019-0688-y>
- Sanchez JN, Subramanian C, Chanda M, Shangguan G, Zhang N, Wang T, Timmermann BN, Blagg BJS, Cohen MS (2021) A novel C-terminal Hsp90 inhibitor KU758 synergizes efficacy in combination with BRAF or MEK inhibitors and targets drug-resistant pathways in BRAF-mutant melanomas. *Melanoma Res* 31(3):197–207. <https://doi.org/10.1097/cmr.0000000000000734>
- Sasame J, Ikegaya N, Kawazu M, Natsumeda M, Hayashi T, Isoda M, Satomi K, Tomiyama A, Oshima A, Honma H, Miyake Y, Takabayashi K, Nakamura T, Ueno T, Matsushita Y, Iwashita H, Kanemaru Y, Murata H, Ryo A, Terashima K, Yamanaka S, Fujii Y, Mano H, Komori T, Ichimura K, Cahill DP, Wakimoto H, Yamamoto T, Tateishi K (2022) HSP90 inhibition overcomes resistance to molecular targeted therapy in BRAFV600E-mutant high-grade glioma. *Clin Cancer Res* 28(11):2425–2439. <https://doi.org/10.1158/1078-0432.ccr-21-3622>
- Savoia P, Zavattaro E, Cremona O (2020) Clinical implications of acquired BRAF inhibitors resistance in melanoma. *Int J Mol Sci* 21(24):9730. <https://doi.org/10.3390/ijms21249730>
- Sawaki A, Kurokawa Y, Honma Y, Naito Y, Iwagami S, Baba H, Komatsu Y, Nishida T, Doi T (2022) A phase III trial of pimipespib (TAS-116) in patients with advanced gastrointestinal stromal tumor: CHAPTER-GIST-301. *Ann Oncol* 33:S467–S467. <https://doi.org/10.1016/j.annonc.2022.05.071>
- Schilling D, Bayer C, Li W, Molls M, Vaupel P, Multhoff G (2012) Radiosensitization of normoxic and hypoxic H1339 lung tumor cells by heat shock protein 90 inhibition is independent of hypoxia inducible factor-1 alpha. *PLoS ONE* 7(2):e31110. <https://doi.org/10.1371/journal.pone.0031110>
- Schmidt L, Issa II, Haraldsdottir H, Hald JL, Schmitz A, Due H, Dybkaer K (2022) Hsp90 inhibition sensitizes DLBCL cells to cisplatin. *Cancer Chemother Pharmacol* 89(4):431–440. <https://doi.org/10.1007/s00280-022-04407-5>
- Schopf FH, Biebl MM, Buchner J (2017) The HSP90 chaperone machinery. *Nat Rev Mol Cell Biol* 18(6):345–360. <https://doi.org/10.1038/nrm.2017.20>

- Scutigliani EM, Liang Y, IJff M, Rodermond H, Mei X, Korver MP, Orié VS, Hoebe RA, Picavet DI, Oei A, Kanaar R, Krawczyk PM (2022) Evaluation of the heat shock protein 90 inhibitor ganetespiib as a sensitizer to hyperthermia-based cancer treatments. *Cancers (Basel)* 14(21):5250. <https://doi.org/10.3390/cancers14215250>
- Sequist LV, Waltman BA, Dias-Santagata D, Digumarthy S, Turke AB, Fidias P, Bergethon K, Shaw AT, Gettinger S, Cosper AK, Akhavanfard S, Heist RS, Temel J, Christensen JG, Wain JC, Lynch TJ, Vernovsky K, Mark EJ, Lanuti M, Iafrate AJ, Mino-Kenudson M, Engelman JA (2011) Genotypic and histological evolution of lung cancers acquiring resistance to EGFR inhibitors. *Sci Transl Med* 3(75):75ra26. <https://doi.org/10.1126/scitranslmed.3002003>
- Serwetyk MA, Blagg BSJ (2021) The disruption of protein-protein interactions with co-chaperones and client substrates as a strategy towards Hsp90 inhibition. *Acta Pharm Sin B* 11(6):1446–1468. <https://doi.org/10.1016/j.apsb.2020.11.015>
- Shapiro GI, Kwak E, Dezube BJ, Yule M, Ayrton J, Lyons J, Mahadevan D (2015) First-in-human phase I dose escalation study of a second-generation non-ansamycin HSP90 inhibitor, AT13387, in patients with advanced solid tumors. *Clin Cancer Res* 21(1):87–97. <https://doi.org/10.1158/1078-0432.CCR-14-0979>
- Sharma P, Allison JP (2015) The future of immune checkpoint therapy. *Science* 348(6230):56–61. <https://doi.org/10.1126/science.aaa8172>
- Shi H, Hugo W, Kong X, Hong A, Koya RC, Moriceau G, Chodon T, Guo R, Johnson DB, Dahlman KB, Kelley MC, Kefford RF, Chmielowski B, Glaspy JA, Sosman JA, van Baren N, Long GV, Ribas A, Lo RS (2014) Acquired resistance and clonal evolution in melanoma during BRAF inhibitor therapy. *Cancer Discov* 4(1):80–93. <https://doi.org/10.1158/2159-8290.cd-13-0642>
- Shimamura T, Li D, Ji H, Haringsma HJ, Liniker E, Borgman CL, Lowell AM, Minami Y, McNamara K, Perera SA, Zaghlul S, Thomas RK, Greulich H, Kobayashi S, Chirieac LR, Padera RF, Kubo S, Takahashi M, Tenen DG, Meyerson M, Wong KK, Shapiro GI (2008) Hsp90 inhibition suppresses mutant EGFR-T790M signaling and overcomes kinase inhibitor resistance. *Cancer Res* 68(14):5827–5838. <https://doi.org/10.1158/0008-5472.CAN-07-5428>
- Shimomura A, Yamamoto N, Kondol S, Fujiwara Y, Suzuki S, Yanagitani N, Horiike A, Kitazono S, Ohyanagi F, Doi T, Kuboki Y, Kawazoe A, Shitara K, Ohno I, Banerji U, Sundar R, Ohkubo S, Calleja EM, Nishio M (2019) First-in-human phase I study of an oral HSP90 inhibitor, TAS-116, in patients with advanced solid tumors. *Mol Cancer Ther* 18(3):531–540. <https://doi.org/10.1158/1535-7163.MCT-18-0831>
- Shkedi A, Adkisson M, Schroeder A, Eckalbar WL, Kuo SY, Neckers L, Gestwicki JE (2021) Inhibitor combinations reveal wiring of the proteostasis network in prostate cancer cells. *J Med Chem* 64(19):14809–14821. <https://doi.org/10.1021/acs.jmedchem.1c01342>
- Sikov WM (2015) Assessing the role of platinum agents in aggressive breast cancers. *Curr Oncol Rep* 17(2):3. <https://doi.org/10.1007/S11912-014-0428-7>
- Skeie M, Nikolaysen F, Chitano Y, Stang E (2020) Hsp90 inhibition and co-incubation with pertuzumab induce internalization and degradation of trastuzumab: implications for use of T-DM1. *J Cell Mol Med* 24(17):10258–10262. <https://doi.org/10.1111/jcmm.15643>
- Slovins S, Hussain S, Saad F, Garcia J, Picus J, Ferraldeschi R, Crespo M, Flohr P, Riisnaes R, Lin CC, Keer H, Oganessian A, Workman P, de Bono J (2019) Pharmacodynamic and clinical results from a phase I/II study of the HSP90 inhibitor onalespiib in combination with abiraterone acetate in prostate cancer. *Clin Cancer Res* 25(15):4624–4633. <https://doi.org/10.1158/1078-0432.CCR-18-3212>
- Smith JR, de Billy E, Hobbs S, Powers M, Prodromou C, Pearl L, Clarke PA, Worknan P (2015) Restricting direct interaction of CDC37 with HSP90 does not compromise chaperoning of client proteins. *Oncogene* 34(1):15–26. <https://doi.org/10.1038/onc.2013.519>
- Smyth T, Paraiso KHT, Hearn K, Rodriguez-Lopez AM, Munck JM, Haarberg HE, Sondak VK, Thompson NT, Azab M, Lyons JF, Smalley KSM, Wallis NG (2014) Inhibition of HSP90 by AT13387 delays the emergence of resistance to BRAF inhibitors and overcomes resistance to dual BRAF and MEK inhibition in melanoma models. *Mol Cancer Ther* 13(12):2793–2804. <https://doi.org/10.1158/1535-7163.MCT-14-0452>
- Snigireva AV, Vrublevskaia VV, Skarga YY, Evdokimovskaya YV, Morenkov OS (2014) Effect of heat shock protein 90 (Hsp90) on migration and invasion of human cancer cells in vitro. *Bull Exp Biol Med* 157(4):476–478. <https://doi.org/10.1007/s10517-014-2595-9>
- Soós A, Kelemen A, Orosz A, Szvicsek Z, Tölgyes T, Dede K, Bursics A, Wiener Z (2023) High CD142 level marks tumor-promoting fibroblasts with targeting potential in colorectal cancer. *Int J Mol Sci* 24(14):11585. <https://doi.org/10.3390/ijms241411585>
- Soudan H, Saeed H, Eldemellawy M, Shalaby M, Hassan M, Elkewedi M, El-Nikhely N (2020) Heat shock protein 90 α inhibitor, PU-H71 in combination with DHEA promoting apoptosis in triple-negative breast cancer cell line MDA-MB-231. *Acta Biochim Pol* 67(4):561–570. https://doi.org/10.18388/abp.2020_5418
- Spiegelberg D, Abramenkova A, Mortensen ACL, Lundsten S, Nestor M, Stenerlöw B (2020) The HSP90 inhibitor Onalespiib exerts synergistic anti-cancer effects when combined with radiotherapy: an in vitro and in vivo approach. *Sci Rep* 10(1):5923. <https://doi.org/10.1038/s41598-020-62293-4>
- Student S, Hejmo T, Poterala-Hejmo A, Lesniak A, Buldak R (2020) Anti-androgen hormonal therapy for cancer and other diseases. *Eur J Pharmacol* 866:172783. <https://doi.org/10.1016/J.ejphar.2019.172783>
- Subaiea G, Rizvi SMD, Yadav HKS, Al Hagbani T, Abdallah MH, Khafagy ES, Gangadharappa HV, Hussain T, Abu Lila AS (2023) Ganetespiib with methotrexate acts synergistically to impede NF- κ B/p65 signaling in human lung cancer A549 cells. *Pharmaceuticals* 16(2):230. <https://doi.org/10.3390/ph16020230>
- Sun C, Bai M, Ke W, Wang X, Zhao X, Lu Z (2021) The HSP90 inhibitor, XL888, enhanced cell apoptosis via downregulating STAT3 after insufficient radiofrequency ablation in hepatocellular carcinoma. *Life Sci* 282:119762. <https://doi.org/10.1016/j.lfs.2021.119762>
- Swain SM, Shastry M, Hamilton E (2023) Targeting HER2-positive breast cancer: advances and future directions. *Nat Rev Drug Discov* 22(2):101–126. <https://doi.org/10.1038/s41573-022-00579-0>
- Tan CRC, Abdul-Majeed S, Cael B, Barta SK (2019) Clinical pharmacokinetics and pharmacodynamics of bortezomib. *Clin Pharmacokinet* 58(2):157–168. <https://doi.org/10.1007/s40262-018-0679-9>
- Tang W, Chen Z, Zhang W, Cheng Y, Zhang B, Wu F, Wang Q, Wang S, Rong D, Reiter FP, De Toni EN, Wang X (2020) The mechanisms of sorafenib resistance in hepatocellular carcinoma: theoretical basis and therapeutic aspects. *Signal Transduct Target Ther* 5(1):87. <https://doi.org/10.1038/s41392-020-0187-x>
- Teranishi R, Takahashi T, Obata Y, Nishida T, Ohkubo S, Kazuno H, Saito Y, Serada S, Fujimoto M, Kurokawa Y, Saito T, Yamamoto K, Yamashita K, Tanaka K, Makino T, Nakajima K, Hirota S, Naka T, Eguchi H, Doki Y (2023) Combination of pimatespiib (TAS-116) with sunitinib is an effective therapy for imatinib-resistant gastrointestinal stromal tumors. *Int J Cancer* 152(12):2580–2593. <https://doi.org/10.1002/ijc.34461>

- Togashi Y, Shitara K, Nishikawa H (2019) Regulatory T cells in cancer immunosuppression—implications for anticancer therapy. *Nat Rev Clin Oncol* 16(6):356–371. <https://doi.org/10.1038/s41571-019-0175-7>
- Uno T, Kawai Y, Yamashita S, Oshiumi H, Yoshimura C, Mizutani T, Suzuki T, Chong KT, Shigeno K, Ohkubo M, Kodama Y, Muraoka H, Funabashi K, Takahashi K, Ohkubo S, Kitade M (2019) Discovery of 3-ethyl-4-(3-isopropyl-4-(4-(1-methyl-1H-pyrazol-4-yl)-1H-imidazol-1-yl)-1H-pyrazolo[3,4-b]pyridin-1-yl)benzamide (TAS-116) as a potent, selective, and orally available HSP90 inhibitor. *J Med Chem* 62(2):531–551. <https://doi.org/10.1021/acs.jmedchem.8b01085>
- Vaidya R, Witzig TE (2014) Prognostic factors for diffuse large B-cell lymphoma in the R(X)CHOP era. *Ann Oncol* 25(11):2124–2133. <https://doi.org/10.1093/annonc/mdu109>
- Vannas C, Andersson L, Dolatabadi S, Ranji P, Lindén M, Jonasson E, Ståhlberg A, Fagman H, Åman P (2022) Different HSP90 inhibitors exert divergent effect on myxoid liposarcoma in vitro and in vivo. *Biomedicines* 10(3):624. <https://doi.org/10.3390/biomedicines10030624>
- Vasan N, Baselga J, Hyman DM (2019) A view on drug resistance in cancer. *Nature* 575(7782):299–309. <https://doi.org/10.1038/s41586-019-1730-1>
- Vaseva AV, Yallowitz AR, Marchenko ND, Xu S, Moll UM (2011) Blockade of Hsp90 by 17AAG antagonizes MDMX and synergizes with Nutlin to induce p53-mediated apoptosis in solid tumors. *Cell Death Dis* 2(5):e156. <https://doi.org/10.1038/cddis.2011.39>
- Vriend LEM, van den Tempel N, Oei AL, L'Acosta M, Pieterse FJ, Franken NAP, Kanaar R, Krawczyk PM (2017) Boosting the effects of hyperthermia-based anticancer treatments by HSP90 inhibition. *Oncotarget* 8(57):97490–97503. <https://doi.org/10.18632/oncotarget.22142>
- Wagner AJ, Agulnik M, Heinrich MC, Mahadevan D, Riedel RF, von Mehren M, Trent J, Demetri GD, Corless CL, Yule M, Lyons JF, Oganessian A, Keer H (2016) Dose-escalation study of a second-generation non-ansamycin HSP90 inhibitor, onalespib (AT13387), in combination with imatinib in patients with metastatic gastrointestinal stromal tumour. *Eur J Cancer* 61:94–101. <https://doi.org/10.1016/j.ejca.2016.03.076>
- Wainberg ZA, Anghel A, Rogers AM, Desai AJ, Kalous O, Conklin D, Ayala R, O'Brien NA, Quadt C, Akimov M, Slamon DJ, Finn RS (2013) Inhibition of HSP90 with AUY922 induces synergy in HER2-amplified trastuzumab-resistant breast and gastric cancer. *Mol Cancer Ther* 12(4):509–519. <https://doi.org/10.1158/1535-7163.MCT-12-0507>
- Wang YF, Liu H, Diao LX, Potter A, Zhang JH, Qiao YW, Wang J, Proia DA, Tailor R, Komaki R, Lin SH (2016) Hsp90 inhibitor ganetespib sensitizes non-small cell lung cancer to radiation but has variable effects with chemoradiation. *Clin Cancer Res* 22(23):5876–5886. <https://doi.org/10.1158/1078-0432.CCR-15-2190>
- Wei W, Liu M, Ning S, Wei J, Zhong J, Li J, Cai Z, Zhang L (2020) Diagnostic value of plasma HSP90 α levels for detection of hepatocellular carcinoma. *BMC Cancer* 20(1):6. <https://doi.org/10.1186/s12885-019-6489-0>
- Whitesell L, Lindquist SL (2005) HSP90 and the chaperoning of cancer. *Nat Rev Cancer* 5(10):761–772. <https://doi.org/10.1038/nrc1716>
- Whitesell L, Mimnaugh EG, De Costa B, Myers CE, Neckers LM (1994) Inhibition of heat shock protein HSP90-pp60v-src heteroprotein complex formation by benzoquinone ansamycins: essential role for stress proteins in oncogenic transformation. *Proc Natl Acad Sci USA* 91(18):8324–8328. <https://doi.org/10.1073/pnas.91.18.8324>
- Woodhead AJ, Angove H, Carr MG, Chessari G, Congreve M, Coyle JE, Cosme J, Graham B, Day PJ, Downham R, Fazal L, Feltell R, Figueroa E, Frederickson M, Lewis J, McMenamin R, Murray CW, O'Brien MA, Parra L, Patel S, Phillips T, Rees DC, Rich S, Smith DM, Trewartha G, Vinkovic M, Williams B, Woolford AJA (2010) Discovery of (2,4-dihydroxy-5-isopropylphenyl)-[5-(4-methylpiperazin-1-ylmethyl)-1,3-dihydroisindol-2-yl]methanone (AT13387), a novel inhibitor of the molecular chaperone Hsp90 by fragment based drug design. *J Med Chem* 53(16):5956–5969. <https://doi.org/10.1021/jm100060b>
- Wright JJ (2010) Combination therapy of bortezomib with novel targeted agents: an emerging treatment strategy. *Clin Cancer Res* 16(16):4094–4104. <https://doi.org/10.1158/1078-0432.CCR-09-2882>
- Wyatt PG, Woodhead AJ, Berdini V, Boulstridge JA, Carr MG, Cross DM, Davis DJ, Devine LA, Early TR, Feltell RE, Lewis EJ, McMenamin RL, Navarro EF, O'Brien MA, O'Reilly M, Reule M, Saxty G, Seavers LC, Smith DM, Squires MS, Trewartha G, Walker MT, Woolford AJ (2008) Identification of N-(4-piperidinyl)-4-(2,6-dichlorobenzoylamino)-1H-pyrazole-3-carboxamide (AT7519), a novel cyclin dependent kinase inhibitor using fragment-based X-ray crystallography and structure based drug design. *J Med Chem* 51(16):4986–4999. <https://doi.org/10.1021/jm800382h>
- Xiao Y, Liu YJ (2020) Recent advances in the discovery of novel HSP90 inhibitors: an update from 2014. *Curr Drug Targets* 21(3):302–317. <https://doi.org/10.2174/1389450120666190829162544>
- Xu J, Wu PJ, Lai TH, Sharma P, Canella A, Welker AM, Beattie CE, Elder JB, Easley M, Lonser R, Jacob NK, Pietrzak M, Timmers CM, Lang F, Sampath D, Puduvali VK (2022) Disruption of DNA repair and survival pathways through heat shock protein inhibition by onalespib to sensitize malignant gliomas to chemoradiation therapy. *Clin Cancer Res* 28(9):1979–1990. <https://doi.org/10.1158/1078-0432.ccr-20-0468>
- Ye M, Huang W, Liu R, Kong Y, Liu Y, Chen X, Xu J (2021) Synergistic activity of the HSP90 inhibitor ganetespib with lapatinib reverses acquired lapatinib resistance in HER2-positive breast cancer cells. *Front Pharmacol* 12:651516. <https://doi.org/10.3389/fphar.2021.651516>
- Yong K, Cavet J, Johnson P, Morgan G, Williams C, Nakashima D, Akinaga S, Oakervee H, Cavenagh J (2016) Phase I study of KW-2478, a novel Hsp90 inhibitor, in patients with B-cell malignancies. *Br J Cancer* 114(1):7–13. <https://doi.org/10.1038/bjc.2015.422>
- Yu J, Chen L, Cui B, Wu C, Choi MY, Chen Y, Zhang L, Rassenti LZ, Widhopf GF, Kipps TJ (2017) Cirtuzumab inhibits Wnt5a-induced Rac1 activation in chronic lymphocytic leukemia treated with ibrutinib. *Leukemia* 31(6):1333–1339. <https://doi.org/10.1038/leu.2016.368>
- Yu J, Zhang C, Song C (2022) Pan- and isoform-specific inhibition of Hsp90: design strategy and recent advances. *Eur J Med Chem* 238:114516. <https://doi.org/10.1016/J.Ejmech.2022.114516>
- Zaidi S, McLaughlin M, Bhide SA, Eccles SA, Workman P, Nutting CM, Huddart RA, Harrington KJ (2012) The HSP90 inhibitor NVP-AUY922 radiosensitizes by abrogation of homologous recombination resulting in mitotic entry with unresolved DNA damage. *PLoS ONE* 7(4):e35436. <https://doi.org/10.1371/journal.pone.0035436>
- Zavareh RB, Spangenberg SH, Woods A, Martinez-Pena F, Lairson LL (2021) HSP90 inhibition enhances cancer immunotherapy by modulating the surface expression of multiple immune checkpoint proteins. *Cell Chem Biol* 28(2):158. <https://doi.org/10.1016/j.chembiol.2020.10.005>

- Zhang ZM, Xie Z, Sun GY, Yang PF, Li J, Yang HF, Xiao S, Liu Y, Qiu HB, Qin LJ, Zhang C, Zhang FH, Shan BE (2015) Reversing drug resistance of cisplatin by HSP90 inhibitors in human ovarian cancer cells. *Int J Clin Exp Med* 8(5):6687–6701
- Zhao R, Davey M, Hsu YC, Kaplanek P, Tong A, Parsons AB, Krogan N, Cagney G, Mai D, Greenblatt J, Boone C, Emili A, Houry WA (2005) Navigating the chaperone network: an integrative map of physical and genetic interactions mediated by the HSP90 chaperone. *Cell* 120(5):715–727. <https://doi.org/10.1016/j.cell.2004.12.024>
- Zhao S, Zhou L, Dicker DT, Lev A, Zhang S, Ross E, El-Deiry WS (2021) Anti-cancer efficacy including Rb-deficient tumors and VHL-independent HIF1 α proteasomal destabilization by dual targeting of CDK1 or CDK4/6 and HSP90. *Sci Rep* 11(1):20871. <https://doi.org/10.1038/s41598-021-00150-8>
- Zhou BQ, Liu JX, Lin MA, Zhu JY, Chen WR (2021) Recent advances in immunotherapy, immunoadjuvant, and nanomaterial-based combination immunotherapy. *Coordin Chem Rev* 442:214009. <https://doi.org/10.1016/j.Ccr.2021.214009>
- Zininga T, Ramatsui L, Shonhai A (2018) Heat shock proteins as immunomodulators. *Molecules* 23(11):2846. <https://doi.org/10.3390/molecules23112846>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.