## **ORIGINAL ARTICLE**



# **Blood Cadmium and Abdominal Aortic Calcifcation in Population with Diferent Weight Statuses: a Population‑Based Study**

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#### **Abstract**

The aim of our study was to assess the efect of blood cadmium levels (B-Cd) on abdominal aortic calcifcation (AAC). We used the data from the 2013–2014 NHANES database. A total of 1530 participants were included in our study, with a mean AAC score of  $1.40 \pm 0.10$ , and a prevalence of severe AAC of 7.98%. Participants with higher B-Cd quartiles showed a higher prevalence of severe AAC. B-Cd was positively associated with higher AAC scores and increased risk of severe AAC. In the obese population, blood cadmium levels showed a positive association with the risk of severe AAC. There may be a positive correlation between B-Cd levels and AAC scores and risk of severe AAC, and this correlation is more pronounced in the obese population. Therefore, the cadmium load in AAC patients in the obese population should be considered in clinical work.

**Keywords** Blood cadmium · Abdominal aortic calcifcation · Underweight · Normal weight · Overweight · Obese

## **Introduction**

Vascular calcifcation (VC) is defned as a pathological process of abnormal deposition of calcium, phosphorus, calcium phosphate complexes, or other minerals in the vessel wall, which eventually leads to vascular sclerosis [\[1](#page-11-0)]. In addition to this, VC is also associated with abnormal deposition of lipoproteins in the intima, chronic infammation, and endoplasmic reticulum stress [[2\]](#page-11-1). VC is a common condition in patients with diabetes and chronic kidney disease (CKD) and the prevalence of VC in patients with CKD is more than 70% [\[3](#page-11-2), [4\]](#page-11-3). The development of VC may be related to the conversion of vascular smooth muscle cells (VSMCs) to an osteoblast-like phenotype due to mineral deposition [\[5](#page-11-4)]. VC is a signifcant predictor of the occurrence of cardiovascular

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events and death, and one study found that VC was signifcantly associated with the risk of cardiovascular disease, myocardial infraction, and heart failure in patients with CKD [[6\]](#page-11-5). The results of a randomized controlled trial indicated that patients treated with intravenous sodium thiosulfate did not develop new heart valve calcifcations and their iliac artery calcifcation was relieved to some extent, although sodium thiosulfate did not improve abdominal aortic calcifcation in uremia patients [\[7](#page-11-6)]. Another phase 2b randomized trial demonstrated that SNF472 (myo-inositol hexaphosphate) signifcantly reduced the progression of coronary artery calcifcation and aortic valve calcifcation in patients with end-stage renal disease treated with hemodialysis [\[8](#page-11-7)]. Although the above drugs have shown efective potential in the treatment of VC, large randomized controlled trials are still needed to confirm the therapeutic efficacy of sodium thiosulfate. Therefore, prevention and management of VC in clinical time are crucial.

Abdominal aortic calcification (AAC), which has received increasing attention in recent years, is commonly seen in patients with diabetes and chronic kidney disease [[9,](#page-11-8) [10\]](#page-11-9). The national prevalence of AAC in the US population is 28.8% [\[11](#page-11-10)]. The prevalence of AAC increases with age and can be as high as 96% in people aged 85 years and older [[12\]](#page-11-11). AAC can be used as a prognostic indicator for continuous ambulatory peritoneal dialysis [[13\]](#page-11-12). In addition, AAC is also a good predictor of the risk of cardiovascular events [[14\]](#page-11-13). A meta-analysis showed that patients with CKD with any AAC had a higher risk of cardiovascular events and fatal cardiovascular events [\[15\]](#page-11-14). In order to assess the severity of AAC, Kauppila et al. developed a novel scoring system, which was calculated according to lateral X-ray of lumbar region to score the severity of anterior and posterior aortic calcifcation of each lumbar segment [\[16](#page-11-15)]. The AAC score ranges from 0 to 24, with higher AAC scores associated with more severe calcifcation of the abdominal aorta. This score provides a simple and low-cost assessment for subclinical vascular disease. The AAC score has been widely used in the follow-up clinical research [[17](#page-12-0)].

Cadmium is a common toxic non-essential heavy metal element to human body, cadmium is mainly found in contaminated soil and water sources [[18](#page-12-1)]. Cadmium has a long half-life, the main causes of cadmium exposure include consumption of contaminated food, long-term work in cadmium-contaminated environment, which makes cadmium exposure a global environmental problem that seriouly afects people's health and brings socio-economic burden [\[19](#page-12-2), [20\]](#page-12-3). A study evaluating cadmium exposure on blood pressure in the Korean general population showed that blood cadmium levels were higher in hypertensive patients than in non-hypertensive patients, and that systolic and diastolic blood pressure and mean arterial blood pressure were positively correlated with blood cadmium levels in the participants [\[21](#page-12-4)]. Another prospective cohort study showed that the adjusted HRs in the highest quantile cadmium exposure level group were 1.73 (95% CI: 1.52–1.97) for allcause mortality, 1.72 (95% CI: 1.28–2.30) for cardiovascular disease (CVD) mortality compared with the lowest quantile of cadmium exposure level group [\[22\]](#page-12-5). Cadmium is also an independent risk factor for atherosclerosis [\[23](#page-12-6), [24](#page-12-7)].

The increase in overweight and obese populations is a major global public health problem that poses a considerable socioeconomic burden. Globally, the proportion of adults with a body mass index (BMI) of 25 kg/m<sup>2</sup> or greater increased from 28.8 to 36.9% between 1980 and 2013 [\[25](#page-12-8)]. As of 2015, there were 603.7 million obese adults worldwide [\[26\]](#page-12-9). Obesity is strongly associated with type II diabetes, cardiovascular disease, osteoarthritis, stroke, and other diseases [[27\]](#page-12-10). More than two-thirds of deaths associated with high BMI are due to cardiovascular disease [[26](#page-12-9)].

The relationship between diferent blood cadmium concentrations and AAC is not well understood, and the association between obesity and AAC is contradictory, with one study fnding a negative association between increasing BMI and AAC [[11\]](#page-11-10). We, therefore, use data from the National Health and Nutrition Examination Survey (NHANES) to assess the relationship between cadmium exposure and AAC in people of diferent weight status. We hypothesized that cadmium exposure is positively associated with AAC scores and the risk of severe AAC, and that this positive correlation is more obvious in overweight and obese people. This may

provide new insights into the management and intervention of AAC in clinical practice.

## **Materials Methods**

#### **Study Population**

The National Health and Nutrition Examination Survey (NHANES), conducted by the National Center for Health Statistics, is an ongoing program designed to assess the health and nutrition status of the US population. The crosssectional study employs a complex multi-stage probability sampling design that is continuously updated with survey data in a continuous 2-year replication cycle and has a relatively large representation of participants [[28\]](#page-12-11). The NHANES study protocol was approved by the National Center for Health Statistics Research Ethics Review Committee, and each participant provided written consent prior to pariticipation in the study. All NHANES data are available at<https://www.cdc.gov/nchs/nhanes/>.

We used data from the NHANES database from 2013 to 2014 because only these 2 years of the survey cycle contained data on AAC scores. We excluded participants younger than 40 years of age because participants younger than 40 years of age did not have DXA scans, so they lacked AAC score data. In addition, we also excluded participants who lacked data on blood cadmium and AAC scores. In our study, we initially inclued 10,175 participants, after excluding 6360 participants aged<40 years old, and participants lacking blood cadmium and AAC scores, we fnally included 1530 participants older than 40 years old with complete blood cadmium and AAC scores in our analysis (Fig. [1](#page-2-0)).

#### **Exposure and Outcome Defnitions**

Blood cadmium (B-Cd) was designed as exposure variable. The dilution of the blood in the sample preparation step prior to analysis is a simple dilution of 1 part sample  $+1$ part water $+48$  parts diluent. The effects of the chemicals in the diluent are to release metals bound to red blood cells making them available for ionization, reduce ionization suppression by the biological matrix, prevent clogging of the sample introduction system pathways by undissolved biological solids, and allow introduction of internal standards to be utilized in the analysis step. Tetramethylammonium hydroxide (TMAH, 0.4% v/v) and Triton X-100TM (0.05%) in the sample diluent solubilizes blood components. Whole blood specimens are processed, stored, and shipped to the National Center for Environmental Health, and Centers for Disease Control and Prevention for analysis. Liquid samples are introduced into the mass spectrometer through the inductively coupled plasma (ICP) ionization source.



AAC

Excluded incompleted data of  $N = 255$ 

<span id="page-2-0"></span>**Fig. 1** Flowchart of the sample selection from National Health and Nutrition Examination Survey (NHANES) 2013–2014

 $N = 1530$ 

Cadmium ions frst passed through a focal region, then the dynamic reaction cell (DRC, ELAN®DRC II, PerkinElmer Norwalk, CT), the quadrupole mass flter, and fnally were selectively counted in rapid sequence at the detector where the B-Cd level was accurately measured.When the blood cadmium measurement was below the lower limit of detection (LLOD), they were divided by the square root of 2 [\[29](#page-12-12)]. The variable code used for blood cadmium concentration was LBXBCD in NHANES.

We designed the AAC score and severe AAC as outcome variables. The AAC score was used to assess the severity of abdominal aortic calcifcation, where a higher AAC score was associated with a more severe degree of abdominal aortic calcifcation. Kauppila et al. invented a scoring system to quantify AAC, which was evaluated based on lumbar spine images obtained using dual-enegry X-ray absorptionmetry, by quantifying the diferent severity of calcifcation in each segment of the aortic wall in the anterior region of the lumbar spone L1-4, with a range of 0 to 6 from each segment, with a total AAC score ranging from 0 to 24 [\[16](#page-11-15)]. In 2013–2014, lateral DXA scans of the thoraco-lumbar spine were administered in the NHANES mobile examination center (MEC). The variable code used for blood cadmium concentration was DXAAC24 in NHANES. If the AAC score is greater than 6, it is defned as severe AAC, which represents severe abdominal aortic calcifcation [\[30](#page-12-13)].

#### **Covariates**

Continuous Variables in this study included age, BMI, fastglucose, hemoglobin A1c (HbA1c), serium creatinine, serium uric acid, serium phosphorus, serium calcium,

triglyceride, total chloesterol, systolic bolld pressure, diastolic blood pressure, and the estimated glomerular fltration rate (eGFR). eGFR was calculated according to the CKD Epidemiology Collaboration (CKD-EPI) creatinine equation. Categorical variables in this study included gender (male/female), race (Mexican American/Non-Hispanic Black/Non-Hispanic White/Other races), education level (<9th grade/9–11th grade/college graduate or above/high school graduate/some college or AA degree), marriage (married/not married), the ratio of family income to poverty (RIP), diabetes, hypertension, alcohol drinker, and smoker. Alcohol use and smoking status was obtained from selfreported information. Hypertension was defned based on a self-reported diagnosis of hypertension, diastolic blood pressure≥90 mmHg or systolic blood pressure≥140 mmHg, or the use of antihypertensive medications [[31](#page-12-14)]. Diabetes mellitus was defned base on a self-reported diagnosis of diabete mellitus, 2-h plasma glucose≥200 mg/dL in an oral glucose tolerance test, HbAlc ≥ 6.5%, use of oral hypoglycemic agents, or fasting glucose  $\geq$  126 mg/dL [\[32](#page-12-15)]. BMI was categorized as <18.5, 18.5–24.9, 25.0–29.9, and  $\geq$  30 kg/m2, which corresponded to underweight, normal weight, overweight, and obese groups among the participants. RIP was categorized as  $< 1$ , 1–5, and  $> 5$ .

#### **Statistical Analysis**

We followed CDC guidelines when performing all statistical analyses, considering NHANES sampling weights from the complex multistage cluster survey design [[33\]](#page-12-16). Categorical and continuous variables were described as percentages and means±standard error (S.E.), respectively. We assessed the diference among diferent B-Cd quartiles via a weighted student's *t* test for continuous variables and weighted chisquare test for categorical variables. We used multivariate linear regression to assess the relationship between B-Cd and AAC scores, while  $\beta$  (effect size) was used to present the results. We also tested the independent correlation between B-Cd and severe AAC using multivariate logistic regression, and the results were expressed as OR (odds ratio). In Model 1, no covariates were adjusted. Model 2 was adjusted for age, sex, and races. Model 3 was adjusted for age, sex and races, educational levels, marriage, RIP, BMI, Fastglucose, HbA1c, systolic blood pressure, diastolic blood pressure, diabetes, hypertension, total cholesterol, serum creatinine, serum uric acid, serum phosphorus, serum calcium, eGFR, alcohol use, triglyceride, and total 25-hydroxyvitamin D. Model 4 is a further adjustment of the smoking status based on Model 3. *P*<0.05 was considered statistically signifcant. For the analysis of the correlation between B-Cd and AAC in the population with diferent weight status, we removed the underweight population because of its small number, only 22 people, accounting for about 1% of the total included population, and we partially adjusted the adjustment variables in the analysis for the population with diferent weight status in order to ensure the balance between the data. In Model a, no covariates were adjusted. Model b was adjusted for age, sex, and races. Model c was adjusted for age, sex, races, education level, RIP, diabetes, hypertension, serum creatinine, serum uric acid, serum phosphorus, serum calcium, eGFR, alcohol use, and total 25-hydroxyvitamin D. Model d is a further adjustment of the smoking status based on model c. All analysis was performed using R version 4.2.1. (<http://www.R-project.org>, The R Foundation).

# **Results**

#### **Baseline Characteristics of Participants**

The baseline characteristics of the study population stratifed by B-Cd quartiles are shown in Table [1](#page-4-0). A total of 1530 participants aged 40 years or older were included, with a mean age of  $57.24 \pm 0.37$ , of whom 48.71% were men and 51.29% were women, with a mean blood Cd of  $0.47 \pm 0.02$ and a range of B-Cd quartiles of 0.07–0.21, 0.21–0.335, 0.335–0.63, and 0.63–7.23. Among the diferent quartiles of B-Cd, there were signifcant diferences in age, sex, race, BMI classifcation, education, systolic blood pressure, diastolic blood pressure, eGFR, serum uric acid, hypertension, diabetes, total 25-hydroxyvitamin D, triglycerides, serum calcium and serum phosphorus, poverty level, smoking, and AAC score. The prevalence of severe AAC increases with increasing BLL quartiles. Compared with the lowest quartile, participants in the highest B-Cd quartile were more likely to be female and Non-Hispanic Black, more likely to have hypertension, a history of alcohol consumption, smoking and poverty, as well as lower diastolic blood pressure, higher systolic blood pressure and triglyceride levels. The highest B-Cd quartile had a blood cadmium concentration of  $1.25 \pm 0.07$ ug/L (Fig. [2\)](#page-5-0). In the total population, the mean score for AAC was  $1.40 \pm 0.1$  and the prevalence of severe AAC was 7.98%. Participants in the highest quartile of B-Cd exhibited a higher risk of developing severe AAC compared to the lowest quartile (Q1: 4.05%; Q2: 9.00%; Q3: 10.04%; Q4: 10.37%) (Fig. [3\)](#page-5-1).

## **The Relationship Between the Blood Cadmium and AAC Score and Severe AAC**

The relationship between blood cadmium and AAC scores was observed in Table [2.](#page-6-0) The association between blood cadmium and AAC scores was not obvious in Model 1 and Model 2 (Model 1:  $β = 0.28$ , 95% confidence interval, CI:−0.18–0.73, *P*=0.21; Model2: β=0.36, 95% confdence interval,CI:−0.05–0.77, *P*=0.08). When age, sex and races, educational levels, marriage, RIP, BMI, Fastglucose, HbA1c, systolic blood pressure, diastolic blood pressure, diabetes, hypertension, total cholesterol, serum creatinine, serum uric acid, serum phosphorus, serum calcium, eGFR, alcohol use, triglyceride, and total 25-hydroxyvitamin D were adjusted in model3, we observed a positive association between blood cadmium levels and AAC socres, a 1-unit increase in log2-transformed blood cadmium levels was associated with a 0.3 unit higher AAC socre (Model 3: β=0.30, 95% confdence interval,CI:−0.02–0.61, *P*=0.04). When we further adjusted for smoking, blood cadmium and AAC score scores no longer had a positive correlation.

To further assess the correlation between blood cadmium concentrations and AAC scores, we converted blood cadmium concentrations from a continuous variable to a categorical variable (quartiles). In the unadjusted model, we observed an increase of 0.61 and 0.75 units in AAC scores for B-Cd Quartiles 2, 3, respectively, compared to their lowest quartiles. However, when adjusted for the corresponding covariates, the relationship between B-Cd and AAC scores was no longer signifcant.

The relationship between B-Cd and severe AAC has been revealed in Table [3](#page-6-1), We found that increased B-Cd was positively associated with a higher risk of severe AAC in Model 2 and Model 3 (Model 2:  $OR = 1.79$ ; 95% CI: 1.19–2.69; *P* = 0.01; Model 3: OR = 2.18; 95% CI: 1.34–3.55;  $P = 0.004$ ). In addition, we also observed that in the unadjusted model, the OR (reference to Quartile 1) was 2.34 (95% CI: 1.16–4.71; *P*=0.02) for Quartile 2, 2.64 for Quartile 3 (95% CI: 1.01–6.88; *P*=0.04), and 2.74 for Quartile 4 (95% CI: 1.05–7.14; *P*=0.04). In Model 3, the adjusted OR for Quartile 4 was 8.59 (95% CI: 2.35–31.37;  $P=0.003$ ) (reference to Quartile 1), suggesting that partipants in Quartile 4 had a signifcant 759% higher risk of severe AAC than those in the B-Cd Quartile 1. However, we found that a positive relationship between higher B-Cd and increased risk of severe AAC only existed in Quartile 2 (95% CI: 1.03–27.60; *P*=0.04) of blood cadmium levels for Model 4.

# **The Relationship Between the Blood Cadmium and AAC Score and Severe AAC in Population With Diferent Weight Statuses**

For the analysis of the correlation between B-Cd and AAC in the population with diferent weight status, we removed the underweight population because of its small number, and we partially adjusted the adjustment variables in the analysis for the population with diferent weight status in order to ensure the balance between the data. In the unadjusted model, we observed a positive correlation between blood cadmium levels and AAC scores in B-Cd Quartile 3 of the overweight population ( $β = 1.17, 95%$ 

<span id="page-4-0"></span>



*RIP*, ratio of family income to poverty; *BMI*, body mass index; *DM*, diabetes; *AAC*, abdominal aortic calcifcation

The bold entries represented statistical signifcance

<span id="page-5-0"></span>**Fig. 2** Distribution of blood cadmium concentrations at diferent blood cadmium concentration quartiles



confdence interval, CI:0.21–2.12, *P*=0.021). But in the fully adjusted model, this correlation no longer existed (Table [4,](#page-7-0) Fig. [4](#page-8-0)).

In the model adjusted for age, sex, and race, we observed a positive association between blood cadmium concentrations and the prevalence of severe AAC in overweight and obese populations (Overweight OR 2.49; 95% CI: 1.14–4.71; *P* = 0.027. Obesity OR 2.08; 95% CI:  $1.05-4.12$ ;  $P = 0.039$ ). In the overweight population, the risk of developing severe AAC increased 149% with each unit of blood cadmium concentration, while in the obese population, the risk of developing severe AAC

<span id="page-5-1"></span>

 **Blood cadmium (Quartile)**



<span id="page-6-0"></span>

*β*, efect sizes; 95% *CI*, 95% confdence interval

Model 1: unadjusted model

Model 2: adjusted for age, sex and races

Model 3: adjusted for age, sex and races, educational levels, marriage, RIP, BMI, Fastglucose, HbA1c, systolic blood pressure, diastolic blood pressure, diabetes, hypertension, total cholesterol, serum creatinine, serum uric acid, serum phosphorus, serum calcium, eGFR, alcohol use, triglyceride and total 25-hydroxyvitamin D

Model 4: a further adjustment of smoking relative to Model 3

The bold entries represented statistical signifcance

increased 108% with each unit of cadmium exposure (Table [4](#page-7-0)). When we further adjusted the education level, RIP, diameter, hypertension, serium creativity, serium uric acid, serium phophorus, serium calculus, eGFR, alcohol use, and total 25-hydroxyvitamin D, We still observed a positive association between increased B-Cd and the risk of severe AAC in overweight and obese people (Overweight OR 2.94; 95% CI: 1.01–8.55; *P*=0.048. Obesity OR 3.03; 95% CI: 1.03–8.90; *P* = 0.044). In Model d, a positive association between increased B-Cd and risk of severe AAC was observed only in the obese population, with a 125% increase in the higher risk of severe AAC with increasing concentrations of cadmium per unit (Obesity OR 2.25; 95% CI: 1.04–4.88;  $P = 0.041$  $P = 0.041$  $P = 0.041$ ) (Table 4, Fig. [5](#page-9-0)).

In Model c, the adjusted OR (reference to Quartile 1) was 6.64 (OR 6.64; 95% CI: 1.32–33.44; *P* = 0.025) for Quartile 4, suggesting a positive relationship between higher B-Cd and increased risk of severe AAC with statistical sighificance in overweight population. However, this positive association did not exist in Model d (Table [4,](#page-7-0) Fig. [6\)](#page-10-0).



<span id="page-6-1"></span>

*OR*, odd ratio; *95% CI*, 95% confdence interval

Model 1: unadjusted model

Model 2: adjusted for age, sex and races

Model 3: adjusted for age, sex and races, educational levels, marriage, RIP, BMI, Fastglucose, HbA1c, systolic blood pressure, diastolic blood pressure, diabetes, hypertension, total cholesterol, serum creatinine, serum uric acid, serum phosphorus, serum calcium, eGFR, alcohol use, triglyceride and total 25-hydroxyvitamin D

Model 4: a further adjustment of smoking relative to Model 3

The bold entries represented statistical signifcance

#### Blood cadmium β/OR (95%CI) Model a Good and Model b Model compared to Model d AAC score Continuous Normal weight 0.28 (−0.28, 0.84), *P*=0.301 0.39 (−0.11, 0.88), *P*=0.113 0.05 (−0.40, 0.50), *P*=0.818  $-0.23$  ( $-0.69$ , 0.22), *P*=0.293 Overweight 0.38 (−0.35,1.10), *P*=0.598 0.38 (−0.22, 0.98), *P*=0.187 0.47 (−0.21, 1.16), *P*=0.161 −0.334 (−1.050, 0.382), *P*=0.336 Obesity 0.20 (−0.58, 0.97), *P*=0.282 0.37 (−0.38, 1.11), *P*=0.296 0.40(−0.28, 1.07), *P*=0.233 0.507 (−0.099, 1.112), *P*=0.095 Categorical Normal weight Quartile 1 Reference Reference Reference Reference Quartile 2 0.66 (−0.86, 2.17), *P*=0.363 0.28 (−1.19, 1.74), *P*=0.669 0.47 (−0.69, 1.63), *P*=0.398 0.42 (−0.65, 1.49), *P*=0.415 Quartile 3 0.78 (−0.27, 1.83), *P* = 0.132 −0.09 (−1.22, 1.05), *P*=0.863  $0.27 (-0.72, 1.27)$ *P*=0.568  $0.19$  ( $-0.73$ , 1.11), *P*=0.662 Quartile 4 1.12 (0.20,2.04), *P***=0.021** 0.84 (−0.15, 1.83), *P*=0.086 0.69 (−0.37, 1.76), *P*=0.186 0.393 (−0.71, 1.49), *P*=0.459 Overweight Quartile 1 Reference **Reference** Reference Reference Reference Reference Quartile 2 0.52 (−0.56, 1.61), *P* = 0.314 −0.06 (−1.16, 1.05), *P*=0.910 0.18 (−0.91, 1.27), *P*=0.730  $-0.10$  ( $-1.06$ , 0.86), *P*=0.828 Quartile 3 1.17 ( 0.21, 2.12), *P***=0.021** 0.36 (−0.70, 1.42), *P*=0.446 0.41 (−0.53, 1.36), *P*=0.365 −0.23 (−0.90, 0.43), *P*=0.467 Quartile 4 0.52 (−0.46, 1.50), *P*=0.268 0.23 (−0.81, 1.28), *P*=0.616 0.45 (−0.72, 1.62), *P*=0.425 −0.91 (−2.12, 0.31),  $P = 0.133$ **Obesity** Quartile 1 Reference **Reference** Reference Reference Reference Reference Quartile 2 0.60 (−0.26, 1.45), *P*=0.153 0.16 (−0.78, 1.10), *P*=0.699 0.19 (−0.74, 1.12), *P*=0.668 0.24 (−0.72, 1.19), *P*=0.605 Quartile 3 0.45 (−0.30, 1,19), *P*=0.215 0.02 (−0.87, 0.91), *P*=0.964 0.28 (−0.51, 1.07), *P*=0.460 0.35 (−0.45, 1.16), *P*=0.365 Quartile 4 0.55 (−1.15, 2.24), *P*=0.495 0.53 (−1.27, 2.32), *P*=0.510 0.58 (−1.01, 2.17),  $P = 0.448$ 0.78 (−0.95, 2.52), *P*=0.351 Severe AAC Continuous Normal weight 0.88 (0.50, 1.54), *P*=0.175 1.59 (0.78, 3.24), *P*=0.175 0.51 (0.12, 2.09), *P*=0.324 0.102 (0.014, 0.774), *P***=0.030** Overweight 1.87 (1.00, 3.50), *P*=0.051 2.49 (1.14, 5.45), *P***=0.027** 2.94 (1.01, 8.56), *P***=0.048** 1.36 (0.47, 3.91), *P*=0.551 Obesity 1.45 (0.90, 2.34), *P*=0.118 2.08 (1.05, 4.12), *P***=0.039** 3.03 (1.03, 8.90), *P***=0.044** 2.25 (1.04, 4.88), *P***=0.041** Categorical Normal weight Quartile 1 Reference **Reference** Reference Reference Reference Reference Quartile 2 1.87 (0.40, 8.70), *P*=0.391 0.95 (0.15, 6.21), *P*=0.950 1.47 (0.11, 19.34), *P*=0.755 1.04 (1,00, 10.97), *P*=0.973 Quartile 3 1.96 (0.66, 5.80), *P*=0.202 0.88 (0.19, 4.17), *P*=0.852 1.23 (0.13, 11.83), *P*=0.847 0.78 (0.11, 5.57), *P*=0.795 Quartile 4 1.61 (0.55, 4.69), *P*=0.354 1.99 (0.55, 7.22), *P*=0.249 1.19 (0.14, 10.28), *P*=0.867 0.37 (0.05, 2.86), *P*=0.32 Overweight Quartile 1 Reference Reference Reference Reference Quartile 2 2.95 (0.80, 10.94), *P*=0.097 1.76 (0.46, 6.71), *P*=0.351 2.66 (0.44, 16.06), *P*=0.265 2.12 (0.35, 12.94), *P*=0.389 Quartile 3 4.09 (1.18, 14.25), *P***=0.03** 1.91 (0.56, 6.49), *P*=0.252 2.50 (0.63, 9.96), *P*=0.178 1.44 (0.51, 4.04), *P*=0.464 Quartile 4 4.54 (1.22,16.92), *P***=0.028** 3.60 (0.91, 14.29), *P*=0.064 6.64 (1.32, 33.44), *P***=0.025** 2.09 (0.42, 10.40), *P*=0.344 **Obesity** Quartile 1 Reference **Reference** Reference Reference Reference Reference

#### <span id="page-7-0"></span>**Table 4** Association of blood cadmium level with AAC score and Severe AAC in diferent weight status

| Blood cadmium | $\beta$ /OR (95%CI)  |   |  |                                |
|---------------|--|---|--|--------------------------------|
|               | Model a  | Model b   | Model c  | Model d                        |
| Ouartile 2    | 2.02 (0.41, 10.03), $P = 0.358$ 1.22 (0.22, 6.81), $P = 0.797$ |   | $2.00(0.45, 8.83), P=0.335$                                    | 1.73 (0.44, 6.88), $P = 0.412$ |
| Ouartile 3    | $2.15(0.56, 8.29), P=0.24$                                     | 1.42 (0.35, 5.77), $P = 0.569$  | 2.36 (0.40, 13.87), $P = 0.318$ 1.76 (0.39, 7.92), $P = 0.433$ |                                |
| Quartile 4    |  | 2.03 (0.27, 15.23), $P = 0.457$ 1.95 (0.21, 18.27), $P = 0.502$ 3.53 (0.32, 38.47), $P = 0.278$ 1.57 (0.20, 12.19), $P = 0.644$ |  |                                |

**Table 4** (continued)

Model a: no covariates were adjusted

Model b was adjusted for age, sex and races

Model c was adjusted for age, sex, races, education level, RIP, diabetes, hypertension, serum creatinine, serum uric acid, serum phosphorus, serum calcium, eGFR, alcohol use and total 25-hydroxyvitamin D

Model d is a further adjustment of the smoking status based on model c

The bold entries represented statistical signifcance

## **Discussion**

In our large cross-sectional study that included 1530 participants, we found a possible correlation of B-Cd with AAC scores and severe AAC. In adjusted model 3, we observed that each 1 unit increase in blood cadmium level was associated with a 0.3 unit increase in AAC score, although this correlation disappeared after further adjustment for smoking. In addition to this, we found a positive association between increased B-Cd and increased risk of severe AAC in Model 2 and 3. In the fully adjusted model, we found a positive correlation between increased risk of B-Cd severe AAC only in Quartile 2 with quartiles of blood cadmium levels. We further analyzed the effect of blood cadmium exposure on AAC in people with diferent weight status, and we found In the unadjusted model, we observed a positive correlation between blood cadmium levels and AAC scores in Quartile 3 of B-Cd in the overweight population. In Model a, Model b, and Model c, we observed a positive association between increased B-Cd and the risk of severe AAC in overweight and obese populations. In Model d, a positive association between increased B-Cd and the risk of severe AAC was observed only in the obese population. Therefore, in our clinical work, we should pay attention to cadmium load in AAC patients, especially in those obese population.

Several studies have shown a positive association between cadmium exposure and the risk of cardiovascular disease [\[34–](#page-12-17)[36\]](#page-12-18). One study demonstrated that foodborne cadmium exposure accounted for a considerable burden of coronary heart disease and stroke [\[34](#page-12-17)]. Ma et al. found that elevated serum cadmium concentrations were positively associated with congestive heart failure, coronary heart disease, heart failure and risk of stroke [[35](#page-12-19)]. Björn Fagerberg et al. reported that cadmium exposure was associated with an increased risk of atherosclerotic plaque formation, and with a 40% increase in the prevalence of large plaques [\[36\]](#page-12-18). In a study of young women, cadmium levels were found to be associated with early atherosclerotic vessel wall thickening,

and the authors also found signifcant aortic plaque production in ApoE knockout cadmium-fed mice compared to controls [\[23](#page-12-6)]. In addition, cadmium levels at the site of



<span id="page-8-0"></span>**Fig. 4** Odds ratios (95% confdence intervals) for the association between diferent quartiles of blood cadmium concentrations and AAC scores by weight status subgroups

<span id="page-9-0"></span>**Fig. 5** Odds ratios (95% confdence intervals) for the association between blood cadmium concentrations and AAC scores and severe AAC by weight status subgroups in the fully adjusted model. **A** Odds ratios (95% confdence intervals) for the association between blood cadmium concentrations and severe AAC by weight status subgroups, **B** Odds ratios (95% confdence intervals) for the association between blood cadmium concentrations and severe AAC by weight status subgroups



aortic plaque rupture were twice as high as in other parts of the plaque [[37\]](#page-12-20). Aramjoo H et al. found that blood and hair cadmium levels were correlated with hypertension, with hair being the best biological sample to study the relationship between cadmium exposure and hypertension [[38\]](#page-12-21). A study in a Korean population also showed that high blood cadmium levels were signifcantly associated with high systolic blood pressure, high diastolic blood pressure and increased risk of hypertension in never-smoking women [\[39](#page-12-22)].

The specifc mechanisms linking cadmium exposure to cardiovascular disease are not known. One of the possible mechanisms of vascular calcifcation due to cadmium exposure may be related to oxidative stress and infammation. Cadmium exposure may lead to upregulation of the pro-inflammatory cytokines (IL-6,IL-1 $\beta$ , TNF- $\alpha$ ) and the infammatory markers (C-reactive protein) [[40,](#page-12-23) [41](#page-12-24)]. Cadmium exposure activated the expression of mitogen-activated protein kinase (MAPK) and nuclear factor-kappa B (NFkB) pathways, two major infammatory signaling pathways that are closely associated with the regulation of apoptosis

[\[42](#page-12-25), [43\]](#page-12-26). Elevated levels of pro-infammatory cytokines can activate osteogenic proteins by upregulating NF-kB ligand/ osteoprotegerin expression, thereby favoring osteoblast diferentiation of vascular smooth muscle cells [\[44\]](#page-12-27). Cadmium can cause oxidative stress by inducing an antioxidant response that leads to mitophagy, resulting in the accumulation of reactive oxygen species in vivo [\[45](#page-12-28)]. In turn, elevated oxidative stress can promote vascular calcifcation through mechanisms such as DNA damage and extracellular matrix remodeling [\[46\]](#page-12-29). Oxidative stress is closely related to the phenotype of vascular smooth muscle cells and can increase osteogenic gene expression and extracellular vesicle secretion in vascular smooth muscle cells, thus contributing to osteoblastic diferentiation of vascular smooth muscle cells [\[47](#page-12-30)].

The association between blood cadmium and AAC is stronger in obese and overweight populations. It has been suggested that overweight or obesity signifcantly amplifes the adverse effects of cadmium exposure on the risk of prediabetes [\[48](#page-12-31)]. One study found that high urinary cadmium was signifcantly associated with elevated systolic and diastolic



<span id="page-10-0"></span>**Fig. 6** Odds ratios (95% confdence intervals) for the association between diferent quartiles of blood cadmium concentrations and severe AAC by weight status subgroups

blood pressure in obese people, while such signifcance was not observed in non-obese people [[49](#page-12-32)]. The opposite result was shown in one study, which found a signifcant association between urinary cadmium and hypertension in normal weight participants, but not in overweight or obese participants [[50](#page-12-33)]. Although the exact mechanism responsible for this phenomenon has not been fully elucidated, we speculate that it may be related to increased oxidative stress due to obesity. One study suggested that increased production of reactive oxygen species by leukocytes of those obese individuals may be responsible for increased oxidative damage to lipids and proteins, thus leading to atherosclerosis [[51\]](#page-12-34).

Cadmium, a toxic heavy metal found in soil, seafood, and water, has caused serious global environmental problems and may lead to a heavy socio-economic burden [\[52](#page-12-35), [53](#page-12-36)]. Common causes of cadmium exposure mainly include smoking, traffic emissions and ingestion of food contaminated with cadmium  $[54, 55]$  $[54, 55]$  $[54, 55]$ . It has been found that deficiency of calcium and zinc, two essential metal elements, leads to increased absorption of cadmium by the body, which further

aggravates its toxic efects in the organism [[56\]](#page-13-2). The results of animal experiments also demonstrated that supplementation with zinc ions ameliorated the nephrotoxic efects of cadmium exposure in rats and repaired the expression of tight junction proteins in the kidney [[57](#page-13-3)]. One study found that supplementation with vitamin C reduced cadmium load in liver, kidney, testis, and muscle [\[58](#page-13-4)]. Vitamin D supplementation not only ameliorated the inhibition of osteoblast proliferation due to cadmium exposure, but also alleviated cadmium-induced hepatotoxicity through antioxidant and anti-infammatory pathways [[59,](#page-13-5) [60](#page-13-6)]. Vitamin E has a protective efect against cadmium-induced apoptosis via the Bax/Bcl2 pathway [[61](#page-13-7)]. The synergistic effect among the micronutrients can also alleviate the toxic efects caused by cadmium exposure. Co-administration of vitamin C and vitamin E ameliorated the elevated markers of renal injury and reduced renal oxidative stress markers in rats exposed to cadmium, resulting in a signifcant repair of renal function [[62](#page-13-8)]. The combination of vitamin C, vitamin E, and selenium ameliorated the degenerative changes in the stomach induced by cadmium toxicity [[63](#page-13-9)]. Various chelating agents such as ethylenediaminetetraacetic acid (EDTA) have also been used to treat cadmium toxicity [\[64](#page-13-10), [65\]](#page-13-11). Probiotics have also shown promising potential in reducing cadmium toxicity. Seven probiotics, including Lactobacillus, Bifdobacterium longum, clotting bacillus, Streptococcus valerate, etc., were found to exhibit signifcant therapeutic efects on cadmium toxicity in preclinical studies [[66\]](#page-13-12). The mechanism of the protective efect of probiotics against cadmium toxicity may be due to their ability to reduce intestinal cadmium absorption, avoid tissue cadmium accumulation, and reduce oxidative stress [[67,](#page-13-13) [68\]](#page-13-14). In a randomized controlled trial, it was also found that oral administration of L. plantarum not only reduced cadmium levels in the blood of subjects, but also increased the rate of cadmium excretion in the feces, and this modulating efect may be related to the intestinal microbiota, as the authors found that the protective efect of L. plantarum against cadmium toxicity was diminished after administration of antibiotic treatment [\[69](#page-13-15)]. These fndings above support the development of safe and efective strategies to prevent and control cadmium exposure, such as reducing the bioavailability of cadmium in the body, thus minimizing cadmium exposure, preventing adverse health events and reducing the global economic burden. In addition, the state and government should strengthen environmental monitoring of cadmium, develop relevant policies to help people identify the sources of cadmium contamination and how to avoid cadmium exposure in a timely manner, and conduct a comprehensive health risk assessment to mitigate cadmium exposure in the general population. Cadmium exposure may be a modifable risk factor for the development and progression of abdominal aortic calcifcation, and more research on the mechanisms between cadmium toxicity

and abdominal aortic calcifcation is necessary. Identifcation of molecular targets of cadmium exposure and early biomarkers of cadmium cardiovascular toxicity as well as studies on the interaction between cadmium and other cardiovascular risk factors are also necessary to lay the foundation for the implementation of individualized prevention and control measures.

Our study is based on data from the NHANES database, a national population-based sample, and obtained through standard protocols. Appropriate NHANES sampling weights were considered for all analyses. We also adjusted covariates based on previous studies to reduce confounding bias. However, our study still has some limitations. First, because our study was a cross-sectional study, a causal relationship between blood cadmium levels and AAC could not be obtained, and a longitudinal study with a larger sample size may be needed to further demonstrate this. Second, because the participants in our study were all from the same country, the results of this study may not be generalizable to many other countries around the world. Finally, our study may not be able to exclude the effect of drug use on vascular calcifcation, and we were unable to analyze this association in a broad age group because patients younger than 40 years of age did not have DXA scans and therefore had missing AAC scores.

# **Conclusions**

Blood cadmium concentration was positively correlated with AAC score and the risk of severe AAC. This correlation was more signifcant in overweight and obese populations. Consider the adverse effects of cadmium exposure on the cardiovascular system. We should consider the cadmium load in patients with AAC in our clinical work, especially in overweight and obese populations. Larger prospective studies are needed to assess the exact causality of this relationship.

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**Data Availability** Data described in the manuscript, codebook, and analytic code will be made publicly and freely available without restriction at [www.cdc.gov/nchs/nhanes/](http://www.cdc.gov/nchs/nhanes/)

## **Declarations**

**Ethics Approval and Consent to Participate** The studies involving human participants were reviewed and approved by the NCHS Ethics Review Board. Written informed consent was obtained from all participants in this study.

**Consent for Publication** All authors contributed to the article and approved the submitted version.

**Competing Interests** The authors declare no competing interests.

## **References**

- <span id="page-11-0"></span>1. Villa-Bellosta R. Vascular calcifcation: key roles of phosphate and pyrophosphate. Int J Mol Sci. 2021;22(24):13536.
- <span id="page-11-1"></span>2. Yuan C, Ni L, Zhang C, Hu X, Wu X. Vascular calcifcation: new insights into endothelial cells. Microvasc Res. 2021;134:104105.
- <span id="page-11-2"></span>3. Liu ZH, Yu XQ, Yang JW, Jiang AL, Liu BC, Xing CY, et al. Prevalence and risk factors for vascular calcifcation in Chinese patients receiving dialysis: baseline results from a prospective cohort study. Curr Med Res Opin. 2018;34(8):1491–500.
- <span id="page-11-3"></span>4. Ghosh S, Luo D, He W, Chen J, Su X, Huang H. Diabetes and calcifcation: the potential role of anti-diabetic drugs on vascular calcifcation regression. Pharmacol Res. 2020;158:104861.
- <span id="page-11-4"></span>5. Leopold JA. Vascular calcifcation: mechanisms of vascular smooth muscle cell calcifcation. Trends Cardiovasc Med. 2015;25(4):267–74.
- <span id="page-11-5"></span>6. Chen J, Budoff MJ, Reilly MP, Yang W, Rosas SE, Rahman M, et al. Coronary artery calcifcation and risk of cardiovascular disease and death among patients with chronic kidney disease. JAMA Cardiol. 2017;2(6):635–43.
- <span id="page-11-6"></span>7. Djuric P, Dimkovic N, Schlieper G, Djuric Z, Pantelic M, Mitrovic M, et al. Sodium thiosulphate and progression of vascular calcifcation in end-stage renal disease patients: a double-blind, randomized, placebo-controlled study. Nephrol Dial Transplant. 2020;35(1):162–9.
- <span id="page-11-7"></span>8. Raggi P, Bellasi A, Bushinsky D, Bover J, Rodriguez M, Ketteler M, et al. Slowing progression of cardiovascular calcifcation with SNF472 in patients on hemodialysis: results of a randomized phase 2b Study. Circulation. 2020;141(9):728–39.
- <span id="page-11-8"></span>9. Bendix EF, Johansen E, Ringgaard T, Wolder M, Starup-Linde J. Diabetes and abdominal aortic calcifcation-a systematic review. Curr Osteoporos Rep. 2018;16(1):42–57.
- <span id="page-11-9"></span>10. Zhou Y, Hellberg M, Kouidi E, Deligiannis A, Höglund P, Clyne N. Relationships between abdominal aortic calcifcation, glomerular fltration rate, and cardiovascular risk factors in patients with non-dialysis dependent chronic kidney disease. Clin Nephrol. 2018;90(6):380–9.
- <span id="page-11-10"></span>11. Rahman EU, Chobufo MD, Farah F, Elhamdani A, Khan A, Thompson EA, et al. Prevalence and risk factors for the development of abdominal aortic calcifcation among the US population: NHANES study. Arch Med Sci Atheroscler Dis. 2021;6:e95–101.
- <span id="page-11-11"></span>12. Rodondi N, Taylor BC, Bauer DC, Lui LY, Vogt MT, Fink HA, et al. Association between aortic calcification and total and cardiovascular mortality in older women. J Intern Med. 2007;261(3):238–44.
- <span id="page-11-12"></span>13. Selvan K, Sampathkumar K, Sampath D, Rajiv A. Abdominal aortic calcifcation as prognostic marker in continuous ambulatory peritoneal dialysis. Clin Nephrol. 2022;98(6):267–73.
- <span id="page-11-13"></span>14. Golestani R, Tio R, Zeebregts CJ, Zeilstra A, Dierckx RA, Boersma HH, et al. Abdominal aortic calcifcation detected by dual X-ray absorptiometry: a strong predictor for cardiovascular events. Ann Med. 2010;42(7):539–45.
- <span id="page-11-14"></span>15. Leow K, Szulc P, Schousboe JT, Kiel DP, Teixeira-Pinto A, Shaikh H, et al. Prognostic value of abdominal aortic calcifcation: a systematic review and meta-analysis of observational studies. J Am Heart Assoc. 2021;10(2):e017205.
- <span id="page-11-15"></span>16. Kauppila LI, Polak JF, Cupples LA, Hannan MT, Kiel DP, Wilson PW. New indices to classify location, severity and

progression of calcifc lesions in the abdominal aorta: a 25-year follow-up study. Atherosclerosis. 1997;132(2):245–50.

- <span id="page-12-0"></span>17. Mäkelä S, Asola M, Hadimeri H, Heaf J, Heiro M, Kauppila L, et al. Abdominal aortic calcifcations predict survival in peritoneal dialysis patients. Perit Dial Int. 2018;38(5):366–73.
- <span id="page-12-1"></span>18. Satarug S, Garrett SH, Sens MA, Sens DA. Cadmium, environmental exposure, and health outcomes. Environ Health Perspect. 2010;118(2):182–90.
- <span id="page-12-2"></span>19. Rani A, Kumar A, Lal A, Pant M. Cellular mechanisms of cadmium-induced toxicity: a review. Int J Environ Health Res. 2014;24(4):378–99.
- <span id="page-12-3"></span>20. Genchi G, Sinicropi MS, Lauria G, Carocci A, Catalano A. The efects of cadmium toxicity. Int J Environ Res Public Health. 2020;17(11):3782.
- <span id="page-12-4"></span>21. Eum KD, Lee MS, Paek D. Cadmium in blood and hypertension. Sci Total Environ. 2008;407(1):147–53.
- <span id="page-12-5"></span>22. Li Z, Fan Y, Tao C, Yan W, Huang Y, Qian H, et al. Association between exposure to cadmium and risk of all-cause and causespecifc mortality in the general US adults: a prospective cohort study. Chemosphere. 2022;307(Pt 4):136060.
- <span id="page-12-6"></span>23. Messner B, Knofach M, Seubert A, Ritsch A, Pfaller K, Henderson B, et al. Cadmium is a novel and independent risk factor for early atherosclerosis mechanisms and in vivo relevance. Arterioscler Thromb Vasc Biol. 2009;29(9):1392–8.
- <span id="page-12-7"></span>24. Barregard L, Sallsten G, Harari F, Andersson EM, Forsgard N, Hjelmgren O, et al. Cadmium exposure and coronary artery atherosclerosis: a cross-sectional population-based study of Swedish middle-aged adults. Environ Health Perspect. 2021;129(6):67007.
- <span id="page-12-8"></span>25. Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet. 2014;384(9945):766–81.
- <span id="page-12-9"></span>26. Afshin A, Forouzanfar MH, Reitsma MB, Sur P, Estep K, Lee A, et al. Health efects of overweight and obesity in 195 countries over 25 years. N Engl J Med. 2017;377(1):13–27.
- <span id="page-12-10"></span>27. Guh DP, Zhang W, Bansback N, Amarsi Z, Birmingham CL, Anis AH. The incidence of co-morbidities related to obesity and overweight: a systematic review and meta-analysis. BMC Public Health. 2009;9:88.
- <span id="page-12-11"></span>28. Zipf G, Chiappa M, Porter KS, Ostchega Y, Lewis BG, Dostal J. National health and nutrition examination survey: plan and operations, 1999–2010. Vital Health Stat 1. 201310(56):1–37.
- <span id="page-12-12"></span>29. Westgard JO, Barry PL, Hunt MR, Groth T. A multi-rule Shewhart chart for quality control in clinical chemistry. Clin Chem. 1981;27(3):493–501.
- <span id="page-12-13"></span>30. Górriz JL, Molina P, Cerverón MJ, Vila R, Bover J, Nieto J, et al. Vascular calcifcation in patients with nondialysis CKD over 3 years. Clin J Am Soc Nephrol. 2015;10(4):654–66.
- <span id="page-12-14"></span>31. Fryar CD, Ostchega Y, Hales CM, Zhang G, Kruszon-Moran D. Hypertension prevalence and control among adults: United States, 2015–2016. NCHS Data Brief. 2017;289:1–8.
- <span id="page-12-15"></span>32. Menke A, Casagrande S, Geiss L, Cowie CC. Prevalence of and trends in diabetes among adults in the United States, 1988–2012. JAMA. 2015;314(10):1021–9.
- <span id="page-12-16"></span>33. Johnson CL, Paulose-Ram R, Ogden CL, Carroll MD, Kruszon-Moran D, Dohrmann SM, et al. National health and nutrition examination survey: analytic guidelines, 1999–2010. Vital Health Stat 2. 2013;10(161):1–24.
- <span id="page-12-17"></span>34. Liu J, Li Y, Li D, Wang Y, Wei S. The burden of coronary heart disease and stroke attributable to dietary cadmium exposure in Chinese adults, 2017. Sci Total Environ. 2022;825:153997.
- <span id="page-12-19"></span>35. Ma S, Zhang J, Xu C, Da M, Xu Y, Chen Y, et al. Increased serum levels of cadmium are associated with an elevated risk of cardiovascular disease in adults. Environ Sci Pollut Res Int. 2022;29(2):1836–44.
- <span id="page-12-18"></span>36. Fagerberg B, Barregard L, Sallsten G, Forsgard N, Ostling G, Persson M, et al. Cadmium exposure and atherosclerotic carotid plaques–results from the Malmö diet and Cancer study. Environ Res. 2015;136:67–74.
- <span id="page-12-20"></span>37. Bergström G, Fagerberg B, Sallsten G, Lundh T, Barregard L. Is cadmium exposure associated with the burden, vulnerability and rupture of human atherosclerotic plaques? Plos One. 2015;10(3):e0121240.
- <span id="page-12-21"></span>38. Aramjoo H, Arab-Zozani M, Feyzi A, Naghizadeh A, Aschner M, Naimabadi A, et al. The association between environmental cadmium exposure, blood pressure, and hypertension: a systematic review and meta-analysis. Environ Sci Pollut Res Int. 2022;29(24):35682–706.
- <span id="page-12-22"></span>39. Kwon JA, Park E, Kim S, Kim B. Infuence of serum ferritin combined with blood cadmium concentrations on blood pressure and hypertension: from the Korean National Health and Nutrition Examination Survey. Chemosphere. 2022;288(Pt 1):132469.
- <span id="page-12-23"></span>40. Arikan TA, Kelles M. Plasma selenium and cadmium levels in patients with chronic otitis media in a Turkish population and their relation to infammation markers. Biol Trace Elem Res. 2019;189(1):55–63.
- <span id="page-12-24"></span>41. Hong H, Xu Y, Xu J, Zhang J, Xi Y, Pi H, et al. Cadmium exposure impairs pancreatic β-cell function and exaggerates diabetes by disrupting lipid metabolism. Environ Int. 2021;149:106406.
- <span id="page-12-25"></span>42. Li T, Quan H, Zhang H, Lin L, Ou Q, Chen K. Silencing cyclophilin A improves insulin secretion, reduces cell apoptosis, and alleviates infammation as well as oxidant stress in high glucose-induced pancreatic β-cells via MAPK/NF-kb signaling pathway. Bioengineered. 2020;11(1):1047–57.
- <span id="page-12-26"></span>43. Liu L, Zhao L, Liu Y, Yu X, Qiao X. Rutin ameliorates cadmium-induced necroptosis in the chicken liver via inhibiting oxidative stress and MAPK/NF-κB pathway. Biol Trace Elem Res. 2022;200(4):1799–810.
- <span id="page-12-27"></span>44. Ono T, Hayashi M, Sasaki F, Nakashima T. RANKL biology: bone metabolism, the immune system, and beyond. Infamm Regen. 2020;40:2.
- <span id="page-12-28"></span>45. Park JH, Lee BM, Kim HS. Potential protective roles of curcumin against cadmium-induced toxicity and oxidative stress. J Toxicol Environ Health B Crit Rev. 2021;24(3):95–118.
- <span id="page-12-29"></span>46. Hu CT, Shao YD, Liu YZ, Xiao X, Cheng ZB, Qu SL, et al. Oxidative stress in vascular calcifcation. Clin Chim Acta. 2021;519:101–10.
- <span id="page-12-30"></span>47. Petsophonsakul P, Burgmaier M, Willems B, Heeneman S, Stadler N, Gremse F, et al. Nicotine promotes vascular calcifcation via intracellular Ca2+-mediated, Nox5-induced oxidative stress, and extracellular vesicle release in vascular smooth muscle cells. Cardiovasc Res. 2022;118(9):2196–210.
- <span id="page-12-31"></span>48. Jiang F, Zhi X, Xu M, Li B, Zhang Z. Gender-specifc diferences of interaction between cadmium exposure and obesity on prediabetes in the NHANES 2007–2012 population. Endocrine. 2018;61(2):258–66.
- <span id="page-12-32"></span>49. Vallée A, Gabet A, Grave C, Blacher J, Olié V. Associations between urinary cadmium levels, blood pressure, and hypertension: the ESTEBAN survey. Environ Sci Pollut Res Int. 2020;27(10):10748–56.
- <span id="page-12-33"></span>50. Wang Q, Wei S. Cadmium afects blood pressure and negatively interacts with obesity: fndings from NHANES 1999–2014. Sci Total Environ. 2018;643:270–6.
- <span id="page-12-34"></span>51. Dandona P, Mohanty P, Ghanim H, Aljada A, Browne R, Hamouda W, et al. The suppressive efect of dietary restriction and weight loss in the obese on the generation of reactive oxygen species by leukocytes, lipid peroxidation, and protein carbonylation. J Clin Endocrinol Metab. 2001;86(1):355–62.
- <span id="page-12-35"></span>52. Clemens S, Ma JF. Toxic heavy metal and metalloid accumulation in crop plants and foods. Annu Rev Plant Biol. 2016;67:489–512.
- <span id="page-12-36"></span>53. Godt J, Scheidig F, Grosse-Siestrup C, Esche V, Brandenburg P, Reich A, et al. The toxicity of cadmium and resulting hazards for human health. J Occup Med Toxicol. 2006;1:22.

<span id="page-13-0"></span>54. Böhlandt A, Schierl R, Diemer J, Koch C, Bolte G, Kiranoglu M, et al. High concentrations of cadmium, cerium and lanthanum in indoor air due to environmental tobacco smoke. Sci Total Environ. 2012;414:738–41.

<span id="page-13-1"></span>55. Yang Q, Li Z, Lu X, Duan Q, Huang L, Bi J. A review of soil heavy metal pollution from industrial and agricultural regions in China: pollution and risk assessment. Sci Total Environ. 2018;642:690–700.

- <span id="page-13-2"></span>56. Reeves PG, Chaney RL. Marginal nutritional status of zinc, iron, and calcium increases cadmium retention in the duodenum and other organs of rats fed rice-based diets. Environ Res. 2004;96(3):311–22.
- <span id="page-13-3"></span>57. Jacquillet G, Barbier O, Cougnon M, Tauc M, Namorado MC, Martin D, et al. Zinc protects renal function during cadmium intoxication in the rat. Am J Physiol Renal Physiol. 2006;290(1):F127-137.
- <span id="page-13-4"></span>58. Grosicki A. Infuence of vitamin C on cadmium absorption and distribution in rats. J Trace Elem Med Biol. 2004;18(2):183–7.
- <span id="page-13-5"></span>59. Dal Ulutas A, Turgut Cosan D, Mutlu F. Protective and curative role of vitamin D and hormones on the cadmium-induced inhibition of proliferation of human osteoblast cells. J Basic Clin Physiol Pharmacol. 2020;32(5):995–1000.
- <span id="page-13-6"></span>60. El-Boshy M, Refaat B, Almaimani RA, Abdelghany AH, Ahmad J, Idris S, et al. Vitamin D(3) and calcium cosupplementation alleviates cadmium hepatotoxicity in the rat: enhanced antioxidative and anti-infammatory actions by remodeling cellular calcium pathways. J Biochem Mol Toxicol. 2020;34(3):e22440.
- <span id="page-13-7"></span>61. Amanpour P, Khodarahmi P, Salehipour M. Protective efects of vitamin E on cadmium-induced apoptosis in rat testes. Naunyn Schmiedebergs Arch Pharmacol. 2020;393(3):349–58.
- <span id="page-13-8"></span>62. Milton Prabu S, Shagirtha K, Renugadevi J. Quercetin in combination with vitamins (C and E) improves oxidative stress and renal injury in cadmium intoxicated rats. Eur Rev Med Pharmacol Sci. 2010;14(11):903–14.
- <span id="page-13-9"></span>63. Bolkent S, Sacan O, Yanardag R, Bolkent S. Efects of vitamin E, vitamin C, and selenium on gastric fundus in cadmium toxicity in male rats. Int J Toxicol. 2008;27(2):217–22.
- <span id="page-13-10"></span>64. Lamas GA, Goertz C, Boineau R, Mark DB, Rozema T, Nahin RL, et al. Efect of disodium EDTA chelation regimen on cardiovascular events in patients with previous myocardial infarction: the TACT randomized trial. JAMA. 2013;309(12):1241–50.
- <span id="page-13-11"></span>65. Mousavi A, Pourakbar L, Moghaddam SS. Efects of malic acid and EDTA on oxidative stress and antioxidant enzymes of okra (Abelmoschus esculentus L.) exposed to cadmium stress. Ecotoxicol Environ Saf. 2022;248:114320.
- <span id="page-13-12"></span>66. Bhattacharya S. The role of probiotics in the amelioration of cadmium toxicity. Biol Trace Elem Res. 2020;197(2):440–4.
- <span id="page-13-13"></span>67. Daisley BA, Monachese M, Trinder M, Bisanz JE, Chmiel JA, Burton JP, et al. Immobilization of cadmium and lead by Lactobacillus rhamnosus GR-1 mitigates apical-to-basolateral heavy metal translocation in a Caco-2 model of the intestinal epithelium. Gut Microbes. 2019;10(3):321–33.
- <span id="page-13-14"></span>68. Zhai Q, Wang G, Zhao J, Liu X, Tian F, Zhang H, et al. Protective efects of Lactobacillus plantarum CCFM8610 against acute cadmium toxicity in mice. Appl Environ Microbiol. 2013;79(5):1508–15.
- <span id="page-13-15"></span>69. Zhai Q, Liu Y, Wang C, Zhao J, Zhang H, Tian F, et al. Increased cadmium excretion due to oral administration of lactobacillus plantarum strains by regulating enterohepatic circulation in mice. J Agric Food Chem. 2019;67(14):3956–65.

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