

Groundwater Discharge as a Source of Dissolved Carbon and Greenhouse Gases in a Subtropical Estuary

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Abstract Groundwater may be highly enriched in dissolved carbon species, but its role as a source of carbon to coastal waters is still poorly constrained. Exports of deep and shallow groundwater-derived dissolved carbon species from a small subtropical estuary (Korogoro Creek, Australia, latitude -31.0478°, longitude 153.0649°) were quantified using a radium isotope mass balance model (²³³Ra and ²²⁴Ra, natural groundwater tracers) under two hydrological conditions. In addition, air-water exchange of carbon dioxide and methane in the estuary was estimated. The highest carbon inputs to the estuary were from deep fresh groundwater in the wet season. Most of the dissolved carbon delivered by groundwater and exported from the estuary to the coastal ocean was in the form of dissolved inorganic carbon (DIC; 687 mmol m^{-2} estuary day⁻¹; 20 mmol m⁻² catchment day⁻¹, respectively), with a large export of alkalinity (23 mmol m^{-2} catchment day⁻¹). Average water to air flux of CO₂ (869 mmol m⁻² day⁻¹) and CH₄ (26 mmol m⁻² day⁻¹) were 5- and 43-fold higher, respectively, than the average global evasion in estuaries due to the large input of CO2- and CH4enriched groundwater. The groundwater discharge contribution to carbon exports from the estuary for DIC, dissolved organic carbon (DOC), alkalinity, CO2, and CH4 was 22, 41, 3, 75, and 100 %, respectively. The results show that CO₂ and CH₄ evasion rates from small subtropical estuaries surrounded

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² National Marine Science Centre, Southern Cross University, PO Box 4321, Coffs Harbour NSW 2450, Australia by wetlands can be extremely high and that groundwater discharge had a major role in carbon export and evasion from the estuary and therefore should be accounted for in coastal carbon budgets.

Keywords Submarine groundwater discharge · Surface water–groundwater interaction · Carbon dioxide · Methane · Permeable sediments · Radon · Cavity ring down spectrometry · Mangrove · Greenhouse gases

Introduction

Estuarine ecosystems provide a major pathway for carbon to travel across the land–ocean interface and are considered an important component of the global carbon cycle (Rodríguez-Murillo et al. 2015; Seitzinger et al. 2005). Estuarine carbon input and transformation processes include material exchange with surrounding environments (Hans et al. 2011) (in particular coastal wetlands (Cai 2011)), high rates of primary productivity and respiration (Bianchi 2007), benthic-pelagic coupling (Maher and Eyre 2010), freshwater inputs (Dixon et al. 2014), atmospheric exchange (Borges and Abril 2011), and groundwater discharge (Liu et al. 2014). Of these processes, groundwater exchange is probably the most poorly resolved and has received little attention until recently.

Groundwater discharge/porewater exchange has been suggested to influence carbon cycling in estuarine environments (Santos et al. 2012a; Faber et al. 2014; Wang et al. 2015). Groundwater inputs to coastal waters may be volumetrically small when compared to surface water inputs; however, due to the high concentrations of dissolved constituents, groundwater can be a major pathway for dissolved material exports between terrestrial and coastal systems (Burnett et al. 2006; Maher et al. 2013a; Liu et al. 2014). Submarine groundwater

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discharge (SGD) is defined as the exchange of groundwater between land and ocean regardless of its composition and scale and may be fresh terrestrial, recirculated seawater, or a combination of both (Moore 2010). Fresh groundwater delivers new water along with dissolved constituents, while recirculated saline groundwater can deliver large amounts of recycled carbon and nutrients to surface waters (Weinstein et al. 2007; Gleeson et al. 2013; Liu et al. 2014). The fresh and recirculated marine components of SGD are often well mixed, thus making quantification of their relative contribution to coastal waters complex.

Previous studies exploring SGD inputs of dissolved carbon have reported that groundwater discharge from aquifers to the estuarine and coastal zones may be a significant source of CO_2 and CH_4 to the atmosphere (Cai 2011; Atkins et al. 2013; Call et al. 2015). Additionally, pCO_2 and CH_4 in groundwater can be orders of magnitude higher than atmospheric values (Gagan et al. 2002; Cai 2003; Call et al. 2015). However, estuarine carbon budgets have rarely assessed the importance of groundwater discharge as a driver of CO_2 , dissolved inorganic carbon (DIC), and dissolved organic carbon (DOC). The results from these limited studies clearly demonstrate that groundwater inputs of dissolved carbon can be significant and warrant quantification in coastal carbon budgets (Porubsky et al. 2014; Wang et al. 2015).

Most previous studies related to groundwater-derived carbon cycling have been conducted in the USA and Australia; however, there is still a lack of data from a variety of ecosystems, including estuaries in the southern hemisphere (Cai 2003; Liu et al. 2012; Maher et al. 2013a). Prior studies have attempted to estimate groundwater-derived fluxes of DIC (Atkins et al. 2013), DOC (Goñi and Gardner 2003; Kim et al. 2011), alkalinity (Stewart et al. 2015; Cyronak et al. 2014), DIC and DOC (Santos et al. 2012a; Maher et al. 2013a; Porubsky et al. 2014), or DIC and alkalinity (Santos et al. 2015; Liu et al. 2014; Wang et al. 2015) into coastal waters. To our knowledge, no previous study has attempted to simultaneously quantify groundwater-derived inputs of the four main dissolved carbon species (DIC, DOC, CO₂, and CH₄) and alkalinity into an estuary, and the relative importance of groundwater carbon inputs to atmospheric evasion of CO_2 and CH_4 and carbon exports to the coastal ocean.

We hypothesize that groundwater discharge into a small subtropical estuary will be a significant pathway for carbon exported to the coastal ocean. We tested this hypothesis by performing time series measurements of carbon parameters (DIC, DOC, pCO_2 , pCH_4), alkalinity, and ²²²Rn and Ra isotopes in a tidal estuary that has a simple geometry. Our objectives were to (1) estimate atmospheric CO₂ and CH₄ evasion rates from the estuary, (2) calculate groundwater-derived inputs to the estuary of the four main dissolved carbon species and alkalinity under contrasting hydrological conditions (wet season and dry season), (3) quantify the relative importance of meteoric fresh and recirculated saline groundwater-derived carbon inputs to the estuary, and (4) determine the relative importance of groundwater in the total carbon and alkalinity exports from the estuary to the coastal ocean. This study builds on a companion paper reporting a detailed radon and radium isotope investigation in the same system (Sadat-Noori et al. 2015).

Materials and Method

Site Description

The investigation was carried out in Korogoro Creek (latitude -31.04781°, longitude 153.06492°), a small (5 km long, ~20-25 m wide, average depth 0.9 m, area $116,160 \text{ m}^2$) subtropical tidal estuary in Hat Head, NSW, Australia (Fig. 1). The estuary has a small catchment (18 km²) which is low lying and subject to flooding by seawater during spring tides. The estuary has a residence time of around 1 day and is normally flushed during each tidal cycle, with ocean water penetrating the lower 4 km of the estuary at high tide (Ruprecht and Timms 2010). The region has an average annual rainfall of 1490 mm and experiences a mild subtropical climate all year round. January and July have the highest (26.9 °C) and lowest (11.2 °C) monthly mean air temperatures, respectively. Rainfall is highest from February to March (175.2 mm month⁻¹) and lowest from July to September (71 mm month⁻¹) (http://www.bom.gov.au). Our most downstream station was located at the mouth of the estuary (-31.057624°, 153.056151°) at a sandy beach environment, while the rest of the stations were surrounded by fringing mangrove vegetation.

Experimental Design

Two field campaigns were carried out over two hydrologically distinct seasons (wet and dry). The first set of time series data collection (wet season) was conducted from 08:45 am March 25 to 10:15 am March 27, 2013, while the second field campaign (dry season) was carried out from 08:30 pm June 6 to 10:00 am June 10, 2013. Both field campaigns were conducted around the spring tides. However, the tidal range between the semidiurnal tides varied. During the wet season, the tidal range was similar between the semidiurnal tides (~1.2 m), while in the dry season, the tidal range varied between ~0.6 and 1 m (Fig. 2) over the semidiurnal tides.

During the first field campaign, we deployed automatic high frequency time series monitoring stations at four approximately equally spaced sites (~1 to 1.5 km apart) along the length of the estuary (Fig. 1). During the second field campaign, two time series monitoring stations were deployed. The station at the mouth of the estuary, here after referred to as **Fig. 1** Study site (Had Head, NSW, Australia) displaying the time series stations (*green points*) and groundwater sampling locations (*red points*). The distance between stations 1 and 2 and 2 and 3 is 1.5 km, while stations 3 and 4 are 1 km apart. The *dark box* represents the study site. The entire estuary length was fringed by mangroves. All carbon species were measured in Station 1, while only pCO_2 data are available for stations 2–4. Image from Google Earth



"downstream station" continually monitored salinity, temperature, dissolved oxygen, current velocity, pCO_2 , pCH_4 , and ²²²Rn, during the two field campaigns, while the other stations measured the same suite of parameters with the exception of pCH_4 . A time series of discrete samples for DIC, DOC, and TAlk were also collected from the downstream station every hour over two consecutive tidal cycles (25 h) during both sampling campaigns. Groundwater samples were collected throughout the catchment (Fig. 1) by installing shallow piezometers (Charette and Allen 2006) and by sampling deep monitoring wells in the region to characterise the groundwater endmember concentration. A mass balance model was developed to evaluate fresh and saline groundwater-derived carbon and alkalinity fluxes, atmospheric exchange rates of CO_2 and CH_4 , and to quantify carbon exports from the estuary.

Surface Water Time Series Observations

A calibrated Hydrolab automatic logger was used to measure pH (± 0.02 units), salinity (± 0.02 ppt), dissolved oxygen ($\pm 0.2 \text{ mg L}^{-1}$), and water temperature ($\pm 0.10 \text{ °C}$), at 15-min intervals, at all stations, during both sampling campaigns. Depth loggers (CTD divers; Schlumberger Water Services) measured estuary depth ($\pm 0.01 \text{ m}$), at 10-min intervals at each of the four stations. Wind speed data were obtained online (www.wunderground.com) from a weather station at 10-m height located at South West Rocks, about 15 km away from the study site. An acoustic Doppler current profiler (ADCP; Sontek Argonaut) was installed in the middle of the estuary at the downstream site to measure current velocity and direction of flow averaged over 10-min intervals. This was combined with time-specific cross-sectional area (adjusted for tidal height) to obtain 10-min discharge estimates assuming that

currents across the channel were homogenous. At the other three time series stations, current velocity was measured using Starflow Ultrasonic Doppler Flow Recorders. The estuary cross section was measured at high tide using a depth gauge at 2-m width intervals.

For determining ²²²Rn concentrations, hereafter referred to as "radon," a radon-in-air monitor modified for radon-inwater (RAD 7, Durridge Co.) was used (Burnett et al. 2010 and references therein). Radon was measured every 10 min for about 40 h in the wet season and 60 h in the dry season. At the downstream station during both seasons, a cavity ring down spectrometer (Picarro G2201-i) coupled to a showerhead equilibrator was used to measure pCO_2 and pCH_4 at ~1 Hz (Maher et al. 2013b) with data averaged over 1-min intervals. The equilibrated air is continuously pumped in a closed-loop from the headspace of the equilibrator chamber through desiccant (Drierite), the cavity ring down spectrometer, a RAD7 and then back to the equilibrator. For measuring pCO_2 at other stations, a Li-Cor 820 CO2 analyzer coupled to a RAD7 radon monitor was used (Santos et al. 2012b). CH₄ partial pressure was converted to concentrations based on the solubility coefficient calculated as a function of temperature and salinity (Wiesenburg and Guinasso 1979) to allow for easy comparison with previous studies that generally use CH₄ concentration rather than partial pressure.

Discrete samples were collected using a sample-rinsed 60ml polyethylene syringe every hour for about 25 h from the downstream station in both seasons. Samples collected for DIC and DOC concentrations were filtered through 0.7 μ m Whatman GF/F filters into acid-rinsed, milli-q rinsed, precombusted (4 h, 400 °C) 40-ml volatile organic carbon borosilicate vials containing 100 μ l of saturated HgCl₂, without any headspace or bubbles. Samples for alkalinity (TAlk) Fig. 2 Surface water time series data from the downstream station located at the mouth of the estuary in the wet (March) and dry (June) seasons. The radon data are reported in our companion paper (Sadat-Noori et al. 2015)



were filtered through 0.7 μ m Whatman GF/F and collected in 30-ml polycarbonate vials. The samples were stored on ice until returning to the lab where they were stored at 4 °C until analysis.

Groundwater Sampling

Groundwater sampling was performed at the same time as surface water sampling in both field campaigns. Samples were collected using a push point piezometer system (Charette and Allen 2006). The tubing was thoroughly flushed with the sample prior to collecting each sample. DOC, DIC, and TAlk were sampled as per surface water methods described earlier. For radon, shallow wells ranging between 0.5 and 2 m deep were dug adjacent to the estuary near each time series station (Fig. 1), using a handheld auger at low tide. PVC pipes with 50-cm-long slotted screens were installed to allow groundwater infiltrate into the pipe. In addition, deep (5–21 m) monitoring wells installed by the NSW Office of Water located across the catchment were also sampled (Fig. 1). A peristaltic pump was used to take samples after the wells were purged (three times the casing volume). Groundwater samples are the same as those used for radium isotope and radon concentrations in Sadat-Noori et al. (2015).

Samples for CO_2 and CH_4 were collected in gas-tight 250ml bottles, overflowing at least three times the bottle volume, to which 200 μ L of saturated HgCl₂ solution was added. A calibrated handheld YSI multiprobe was used to determine pH, temperature, DO, and salinity for each groundwater sample. A total of 27 groundwater samples were collected. Sixliter gas-tight HDPE plastic bottles were used to collect samples for groundwater radon analysis. Each six-liter bottle was connected to a RAD7 radon monitor and given at least 2 h to achieve an air-water radon equilibrium with <5 % uncertainty following well established protocols (Lee and Kim 2006). After radon analysis, the water was filtered through magnesium impregnated acrylic fibers for radium analysis (Peterson et al. 2009). Groundwater samples were classified in two classes; deep (5 m) and shallow (5 m) based on the radium data in the companion paper (Sadat-Noori et al. 2015).

Analytical Methods

Groundwater CO₂ and CH₄ samples were analyzed via a headspace method using a Picarro G2201-i as described by Gatland et al. (2014). Analysis for DIC and DOC concentrations were carried out following the wet oxidation method (St-Jean 2003) with an OI Aurora 1030 W interfaced with a Therma Delta V+ Isotope Ratio Mass Spectrometer (IRMS) (Maher and Eyre 2011). TAlk $(\pm 0.2 \%)$ was measured by Gran Titration using a Metrohm automatic titrator and 0.01 M HCl standardized to Dickson Certified Reference Material (Batch 111). Free CO_2 within the estuary was determined by using the DIC and TAlk pair, and also the pCO_2 and TAlk pair, using version 25 of the CO₂SYS program (Pelletier et al. 2007) with the carbonic acid disassociation constants from Millero et al. (2006) and the KHSO₄ constant from Dickson (1990). Radium samples were collected at the downstream time series station every hour for 30 h in wet and dry seasons, and a Radium Delayed Coincidence Counter (RaDeCC) was used for measuring ²²³Ra and ²²⁴Ra based on Moore and Arnold (1996). Radium data and estimated groundwater discharge rates are presented in a companion paper (Sadat-Noori et al. 2015).

Calculations

The CO_2 and CH_4 atmospheric exchange were estimated following Wanninkhof (1992):

$$Flux = k \alpha (C_{water} - C_{air}) \tag{1}$$

where C_{water} and C_{air} are the partial pressure of CO₂ or CH₄ in the water column and in air, respectively, in units of µatm; α is the solubility coefficient, calculated as a function of temperature and salinity using the constants of Weiss (1974) for CO₂ and Wiesenburg and Guinasso (1979) for CH₄, *k* is the gas transfer velocity at the water–air interface (m day⁻¹). The atmospheric *p*CO₂ and *p*CH₄ were assumed to be constant at an average of 400 and 1.8 µatm, respectively. We used an empirical equation which estimates transfer velocity as a function of water depth, current, and wind speed, which are the dominant sources of water turbulence in estuarine systems (Borges et al. 2004):

$$k_{600} = 1 + 1.719 W^{0.5} D^{-0.5} + 2.58 U_{10}$$
⁽²⁾

where k_{600} is the transfer velocity (normalized to a Schmidt number of 600), W is the water current (cm s⁻¹), D is water depth (m), and U_{10} is the wind speed at a height of 10 m (m s⁻¹). The Schmidt number is defined as the ratio between the kinematic viscosity to mass diffusivity. All k_{600} values were corrected for the Schmidt number of CO₂ and CH₄ at in situ temperatures and salinities (Wanninkhof 1992), assuming a linear relationship between salinities of 0 and 35. The main uncertainty associated with quantifying air–water gas exchange results from the estimation of gas transfer velocity (k). Most previous studies have used empirical equations which calculate transfer velocity only as a function of wind speed. The model used in this study incorporates current and wind induced turbulence at the air–water interface.

The hourly estuarine export (ebb tide) and import (flood tide) of the four dissolved carbon species (DIC, DOC, free CO_2 , and CH_4) and total alkalinity was estimated by multiplying hourly discharge rates by the carbon species concentration. Daily averages where calculated by integrating export and import rates over two tidal cycles then dividing by total time for the two tidal cycles (~25 h) to get an hourly rate, and multiplying by 24 (hours in 1 day) to obtain a daily rate. Groundwater carbon fluxes were calculated by multiplying the corresponding daily volumetric groundwater discharge in each season obtained from Sadat-Noori et al. (2015) by the median concentration of different carbon species in groundwater following Eq. (3):

$$GW_{C flux} = GW_{dis.} \times GW_{med. C conc.}$$
(3)

where $GW_{C\ flux}$ is groundwater carbon fluxes, $GW_{dis.}$ is groundwater discharge in each season and $GW_{med.\ C\ conc.}$ is the median concentration of different carbon species in groundwater. The median concentration was used due to the nonnormal distribution of the groundwater endmember concentrations.

A nonsteady state radium mass balance was applied to quantify fresh and saline discharging groundwater into the estuary. The model details and results are presented in a companion paper (Sadat-Noori et al. 2015). Briefly, concentrations of radium in surface water are converted into net fluxes of groundwater, discharging into the estuary over a 24-h diel cycle. Inputs to the model were groundwater, upstream ²²³Ra input flux during flood tide, diffusion from sediments, and desorption from suspended sediments while outputs consisted of ²²³Ra downstream output flux during ebb tides and the ²²³Ra decay.

Results

Hydrological Conditions

Contrasting hydrological conditions occurred during each field campaign. Two months prior to the time series measurements in March (wet season), the area received 612 mm of rainfall. The June 2013 (dry season) time series deployment had base flow conditions with only 57 mm of rain in the 2 months prior to field campaign. As a result, the groundwater level during March was 100 cm higher than in June. Based on the rainfall events of the area and for simplicity, we describe the first and second field campaigns as wet and dry seasons, respectively. Surface freshwater discharge (i.e., net freshwater discharge out of the mouth of the estuary) was $3 \text{ m}^3 \text{ s}^{-1}$ in the wet season and decreased to 2.2 $\text{m}^3 \text{s}^{-1}$ in the dry season. Wet season had an average surface water temperature of 25.9 °C compared to 19.4 °C in the dry season. Wind speeds were on average 3.1 and 1.7 m s^{-1} during the wet and dry seasons, respectively (Fig. 2). Tidal range was ~1.2 m in the wet season while in the dry season the tidal range varied between 1 and 0.6 m (Fig. 2). Salinity showed a tidal trend and ranged from nearly fresh (1) to saline (up to 35) over a tidal cycle (Fig. 2). Salinity increased rapidly during the start of the flood tide just taking 2.5 h to reach 34 and dropped more slowly during ebb tide taking about 5 h to reach minimum values. Similar salinity trends were observed in both campaigns.

Groundwater Observations and Discharge Rates

Shallow and deep groundwater dissolved carbon concentrations were highly variable (Table 1). Median DIC, DOC, and TAlk in shallow groundwater were 1.1, 1.2, and 1.5 times higher than deep groundwater. Median pCO_2 in deep samples (21,109 µatm) was similar to median pCO_2 in shallow samples (20,924 µatm), while median CH₄ concentration was 6.6 times higher in the deep samples (53 µM) than shallow (8 µM).

The discharging groundwater into the estuary surface water was separated into shallow saline and deep fresh groundwater components. Depth was used as a separation factor rather than salinity because in a tidal estuary with a short resident time salinity may not truly represent the spatial groundwater distribution along the estuary. For example, tidal pumping at the downstream station would cause high salinities in shallow groundwater, while the salinity in groundwater samples at station 4 were never higher than 5. Moreover, the 5 m indicator used as a separation was based on the fact that radium concentration was generally higher in samples collected below 5 m (see Sadat-Noori et al. 2015).

Separate discharge rates were estimated based on ²²³Ra and ²²⁴Ra for wet and dry seasons and deep and shallow ground-water discharge. An average of the ²²³Ra and ²²⁴Ra rates was

used to calculate seasonal deep and shallow groundwater discharge rates which were then used to calculate groundwaterderived carbon fluxes entering estuary surface water (refer to Sadat-Noori et al. 2015 for groundwater discharge calculations). In the wet season, groundwater-derived DIC from fresh deep groundwater was 1.8 times higher than saline shallow groundwater-derived DIC. DOC and alkalinity derived from fresh deep groundwater was 1.7- and 1.3-fold higher than saline shallow groundwater-derived DOC (Table 2). Groundwater-derived free CO2 and CH4 were 2.1- and 14fold higher from fresh deep groundwater compared to shallow. In the dry season, DIC, DOC, alkalinity, and free CO2 from saline shallow GW were 6.2, 6.3, 8.8, and 5.3 times higher than the fresh deep groundwater fluxes while CH₄ fluxes were similar from both deep and shallow fluxes. All estimates of groundwater-derived carbon inputs to the estuary were higher in the wet season (Table 2).

Estuary Surface Water Time Series Measurements

Dissolved oxygen (DO) followed a clear tidal trend with the highest values at high tide (Fig. 2). DO reached 100 % saturation at high tide in both seasons and dropped to 50 and 30 % at low tide during the dry and wet seasons, respectively. Radon, pCO_2 and CH₄ followed a tidal trend in both seasons with the highest concentrations being recorded at low tide and lowest at high tide (Fig. 2). Radon concentrations ranged from 0.6 to 180.2 Bq m⁻³ with an average of 50.5 Bq m⁻³ in wet season and varied between 6.2 and 209.1 Bq m⁻³ with an average of 86.5 Bq m⁻³ in dry season, respectively (Fig. 2).

DOC followed a tidal trend with high concentrations at low tide and low concentrations at high tide (Fig. 2), while DIC and TAlk concentrations displayed an opposite tidal trend (high concentrations at high tide). DIC concentrations ranged from 809 to 2151 μ M with an average of 1558 μ M in wet conditions and 1500 μ M in the dry season (Fig. 2). DOC ranged from 38 to 2158 μ M with an average of 931 μ M in the wet season and 668 μ M in the dry season. Alkalinity varied between 840 and 2347 μ M with a similar average in both seasons.

Carbon dioxide was the only carbon species with observations in multiple stations along the estuary. At the downstream station, pCO_2 followed a tidal trend and was 1.5 times higher in the wet season compared to the dry season (Fig. 3). Average pCO_2 at stations 2, 3, and 4 was about 14,000 µatm in the wet season. Average pCO_2 for station 2 in the dry was 9549 µatm. Maximum pCO_2 in surface waters was 25,130 µatm in the wet season (station 2) and 16,764 µatm in the dry season (Fig. 3). CH₄ concentrations (station 1 only) ranged from 5 nM (high tide) to about 3 µM (low tide), while in the dry season, CH₄ varied between 5 nM (high tide) to 4.8 µM (low tide).

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CO₂ and CH₄ Water to Air Fluxes

Stations 1, 2, 3, and 4 had average gas transfer velocities (k_{600}) values of 4.6, 3.0, 4.4, and 3.1 m day⁻¹, respectively, in the wet season while stations 1 and 2 had k_{600} values of 1.7 and 2.9 m day⁻¹, respectively, in the dry season. In the wet season, CO2 fluxes from the lower, mid and upper estuary were estimated to be 573 (station 1 average), 1505 (average of station 2 and 3), and 1650 mmol m^{-2} day⁻¹ (average of stations 3 and 4), respectively. In the dry season, the average was 220 for station 1 and 1300 mmol m^{-2} day⁻¹ for station 2 (no upper estuary values in dry season, due to vandalism). Wet and dry seasons had an integrated average CO_2 flux of 1128 and 620 mmol m⁻² day⁻¹ (Table 3). CH₄ fluxes from the downstream static ıd 43 mmol m^{-2} day⁻¹ in wet and dry sea ely (Table 3).

Latitude

Depth

Salinity

Longitude

Table 1 Groundwater observations

Date

Sample ID

											(µM)	
shallow												
GW 1	7/06/2013	S31.057	E153.056	1.5	30.9	8.5	25	2707	2055	271	3	0.02
GW 2	7/06/2013	S31.057	E153.056	1.5	25.7	8.5	1599	2617	2101	182	4	0.2
GW 3	8/06/2013	S31.057	E153.056	1.8	17.9	8.2	941	2057	1880	564	11	1
GW 4	12/06/2013	S31.000	E153.032	4	0.1	5.0	365	188	3345	5936	3146	152
GW 5	8/06/2013	S31.0071	E153.032	4	0.1	4.9	105	15	1333	6510	1295	14
GW 6	9/06/2013	S31.305	E153.038	2.5	0.2	4.6	27	22	1531	7728	1486	112
GW 7	9/06/2013	S31.056	E153.033	2	0.1	4.6	266	72	1455	4968	1372	31
GW 8	10/06/2013	S31.042	E153.034	2	2.2	6.6	605	1054	1404	2844	350	3
GW 9	10/06/2013	S31.042	E153.042	1.5	1.5	5.3	834	316	2254	1442	1934	5
GW 10	10/06/2013	S31.042	E153.042	2	0.1	4.9	250	24	672	776	636	0.1
GW 11	10/06/2013	S31.050	E153.042	1.5	14.8	5.8	470	960	2296	545	1335	10
GW 12	10/06/2013	S31.050	E153.050	1.2	0.3	6.7	587	1211	1488	2089	278	13
GW 13	10/06/2013	S31.050	E153.042	1.5	1.8	5.9	444	439	1157	1144	716	0.3
GW 14	11/06/2013	S31.069	E153.032	4.5	0.1	5.2	159	133	1386	533	1246	42
Median						5.6	405	378	1509	1293	981	8
ST error						0.4	115	260	173	700	237	12
Deep												
GW 15	12/06/2013	S30.997	E153.025	5.5	0.1	4.4	574	5	2159	7710	2118	116
GW 16	12/06/2013	S30.999	E153.027	16	0.1	5.2	1403	240	2204	1068	1958	172
GW 17	8/06/2013	S31.029	E153.026	10	0.2	4.7	209	9	931	6194	901	0.1
GW 18	8/06/2013	S31.007	E153.026	18	0.1	6.2	247	843	1569	370	725	54
GW 19	8/06/2013	S31.021	E153.024	6	0.1	6.0	135	845	2196	10230	1350	0.1
GW 20	8/06/2013	S31.021	E153.024	21	0.1	5.8	868	485	1402	320	929	177
GW 21	8/06/2013	S31.056	E153.036	11.7	0.2	4.7	181	26	1195	4985	1150	27
GW 22	9/06/2013	S31.053	E153.037	7.5	0.1	6.8	561	662	823	420	162	12
GW 23	11/06/2013	S31.068	E153.033	20	0.1	5.3	240	145	1190	51	1040	281
GW 24	11/06/2013	S31.068	E153.033	10.8	0.1	6.5	451	881	1293	950	412	53
GW 25	11/06/2013	S31.091	E153.013	10.5	0.1	5.5	103	271	1496	161	1222	84
GW 26	11/06/2013	S31.008	E153.025	11.7	0.1	5.3	501	152	1330	1330	995	42
GW 27	12/06/2013	S31.020	E153.024	5.5	0.1	4.8	416	35	1049	6704	999	30
Median						5.3	416	240	1330	1068	999	53
ST error						0.2	100	96	130	982	148	23
otal												
Median						5.4	416	271	1455	1144	999	27
ST error						0.1	75	148	111	587	140	14

²²²Rn

Alkalinity

pН

CH₄

DOC

Free

DIC

	Wet	Dry
Shallow groundwater		
SGD (m ³ s ^{-1})	$0.33 {\pm} 0.09$	$0.19 {\pm} 0.23$
DIC (mmol $m^{-2} day^{-1}$)	376±376	213±129
DOC (mmol $m^{-2} day^{-1}$)	322±322	183 ± 521
Alkalinity (mmol $m^{-2} day^{-1}$)	94±94	53±193
Free CO ₂ (mmol $m^{-2} day^{-1}$)	244±244	138 ± 177
$CH_4 \text{ (mmol } m^{-2} \text{ day}^{-1}\text{)}$	2 ± 2	1 ± 9
Deep groundwater		
SGD (m ³ s ^{-1})	$0.71 {\pm} 0.25$	$0.03 {\pm} 0.01$
DIC (mmol $m^{-2} day^{-1}$)	702 ± 97	34±97
DOC (mmol $m^{-2} day^{-1}$)	564±730	29±730
Alkalinity (mmol $m^{-2} day^{-1}$)	127±71	6±71
Free CO ₂ (mmol $m^{-2} day^{-1}$)	527±110	26±110
$CH_4 \text{ (mmol } m^{-2} \text{ day}^{-1}\text{)}$	28±17	1 ± 17
Total (deep + shallow)		
SGD (m ³ s ^{-1})	$1.04{\pm}0.13$	0.22 ± 0.13
DIC (mmol $m^{-2} day^{-1}$)	1131±83	243±83
DOC (mmol $m^{-2} day^{-1}$)	889±436	191 ± 587
Alkalinity (mmol $m^{-2} day^{-1}$)	211 ± 110	45±148
Free CO ₂ (mmol $m^{-2} day^{-1}$)	777±104	167±139
$CH_4 \text{ (mmol } m^{-2} \text{ day}^{-1}\text{)}$	20±10	4±14

SGD rates reported in Sadat-Noori et al. (2015) were used. Estuary area $116,160 \text{ m}^2$; catchment area $18,000,000 \text{ m}^2$

Estuarine Carbon Export

The Hat Head estuary exported on average 20 ± 4 and 9 ± 2 mmol C m⁻² of catchment day⁻¹ of DIC and DOC,

Fig. 3 Time series of partial pressure of CO_2 from the four stations in the wet season and two stations in the dry season

respectively, to the coastal ocean (Table 4) based on the two field campaigns. DIC, DOC, TAlk, and free CO₂ exports were 35, 80, 30, and 93 %, higher in the wet season compared to the dry season, while CH₄ was 50 % higher in the dry season. Average alkalinity export $(23\pm5 \text{ mmol m}^{-2} \text{ of}$ catchment day⁻¹) was similar to DIC and approximately six times higher than free CO₂ $(4\pm1 \text{ mmol C m}^{-2} \text{ of}$ catchment day⁻¹) and several orders of magnitude higher than CH₄ (0.005±0.001 mmol C m⁻² of catchment day⁻¹).

Discussion

Carbon Data Integrity

As we over-constrained the carbonate system, we investigate the reliability of our measured DIC data by comparing measured DIC concentrations with those calculated from pCO_2 and TAlk (Fig. 4). Our comparison showed that calculated DIC concentrations were on average 9 % higher than measured DIC concentration in 90 % of the surface water samples. However, for fresh groundwater DIC samples, calculated and measured concentrations showed a closer agreement (Fig. 4b). The lower measured DIC concentrations may be due to CO_2 losses through contact to air at the time of sampling and filtration, and/or, due to an overestimation of calculated DIC due to the contribution of organic acids to the TAlk pool (Hunt et al. 2011; Abril et al. 2015). In spite of these uncertainties, the estimated estuary DIC export rates are similar using both calculated and measured DIC, with only 8 and 6 % higher export rate using the calculated DIC values in the wet and dry season, respectively (Table 4).



Season	Average gas	CO ₂ atmospheric fluxes		GW-derived CO ₂ CH ₄ atmosphe		eric fluxes	GW-derived CH ₄
	transfer velocity m day ^{-1}	mmol day ⁻¹	mmol $m^{-2} day^{-1}$	mmol $m^{-2} day^{-1}$	mmol day ⁻¹	mmol $m^{-2} day^{-1}$	mmol $m^{-2} day^{-1}$
Wet (March)	3.0	13.1×10^{7}	1127.7	777	0.2×10^{7}	17.2	20
Dry (June)	2.3	7.2×10^{7}	619.8	167	0.5×10^{7}	43.0	4
Average	2.7	10.1×10^{7}	869.4	472	0.3×10^{7}	25.8	12

Table 3 Average gas transfer velocity used in calculations (Eq. (2)), CO_2 and CH_4 evasion from the estuary (Eq. (1)), and groundwater-derived CO_2 evasion (Eq. (3)) in areal units of mmol m⁻² of estuary day⁻¹ and estuary-wide units of mmol day⁻¹

Carbon Water to Air Fluxes

Water to air CO₂ and CH₄ fluxes over the study period show that Hat Head estuary was a source of CO_2 (620 to 1128 mmol m^{-2} day⁻¹) and CH₄ to the atmosphere (17 to 43 mmol m^{-2} day⁻¹) (Table 3). Atkins et al. (2013) reported similarly high CO_2 fluxes of 800 mmol m⁻² day⁻¹ in the upper North Creek Estuary, NSW, Australia, with a smaller k value of 2.8 m day⁻¹. Frankignoulle et al. (1998) found that nine European estuaries had a mean CO₂ flux of 170 mmol m⁻² day⁻¹ using a k value of 1.9 m day⁻¹. The average CO₂ emission from ten Brazilian estuaries was reported to be 55 \pm 45 mmol m⁻² day⁻¹ with pCO₂ varying between 168 and 8638 µatm (Noriega and Araujo 2014). We calculated fluxes based on empirical models of k similar to Atkins et al. (2013) and Noriega and Araujo (2014), while Frankignoulle et al. (1998) used the floating chamber method. The global average pCO_2 in upper estuaries is estimated to be $3033\pm$ 1078 µatm with a corresponding atmospheric CO₂ flux of $188\pm70 \text{ mmol m}^{-2} \text{ day}^{-1}$ (Chen et al. 2012). In our case, the CO_2 water to air flux from the upper estuary $(1650 \text{ mmol m}^{-2} \text{ day}^{-1})$ was an order of magnitude higher than the estimated global average. The high fluxes in this study are likely to be directly related to groundwater inputs (see below).

Several previous studies have utilized a fixed time series measuring station usually located at the mouth of the estuary to estimate CO₂ and/or CH₄ flux for the entire area of the estuary (Bouillon et al. 2007; Maher et al. 2013a). While time series measurements have the advantage of capturing temporal variation with very high resolution, they may not be representative of estuary-wide fluxes due to the inability to account for spatial variation. Maher et al. (2015) suggested that multiple time series stations or a combination of both time series and survey methods may be required to adequately constrain the variability of estuarine CO₂ and CH₄ fluxes at the estuary scale. Here, we simultaneously deployed four fixed time series stations approximately 1.5 km apart along the length of the estuary (~5 km) to cover both temporal and spatial variability, thus providing a more robust estimate of the estuary-wide CO₂ dynamics. A similar sampling strategy could not be applied to CH₄ and other carbon species due to logistic reasons.

Our multistation approach demonstrated the importance of spatial variability in estuarine pCO_2 , when calculating estuary-wide fluxes. We estimated CO2 fluxes using four automated measuring stations in the wet season and two stations in the dry season. If we would have only used a single station at the mouth of the estuary, CO₂ fluxes upscaled to the entire estuary would be underestimated by 50 and 65 % during the wet and dry seasons, respectively (573 and 220 mmol $m^{-2} day^{-1}$, for wet and dry seasons), however, still an order of magnitude higher than the global average estimate for lower estuaries (19–59 mmol $m^{-2} day^{-1}$) (Borges and Abril 2011; Cai 2011; Chen et al. 2012). This was calculated following Eq. (1) by assuming that the downstream station represented a partial pressure of CO_2 and k value for the entire estuary (i.e., the downstream flux was applied to the entire estuary area). By using the four stations, the estuary could be fragmented, with each section having a site-specific set of partial pressures and piston velocities. On the other hand, if sampling had been conducted only in the upstream section of the estuary, CO₂ fluxes would be overestimated considerably. This clearly demonstrates the importance of collecting spatial data from the lower, middle, and upper parts of estuarine systems to be able to estimate a more realistic water-air flux of CO₂ as suggested by previous studies (Wang and Cai 2004; Cai 2011; Maher et al. 2015).

Average CH_4 fluxes from Hat Head estuary were high, averaging 26 mmol m⁻² day⁻¹ (Table 3). These fluxes are about 43 times higher than the higher end of average global methane flux estimates, from tidal estuaries, which range between 0.04 and 0.6 mmol m⁻² day⁻¹ (Borges and Abril 2011). Ferrón et al. (2007) and Zhang et al. (2008) reported an annual average CH_4 flux of 0.66 and 0.61 mmol m⁻² day⁻¹ from tidal estuaries in Bay of Cádiz, SW Spain, and Changjiang, China, while Nirmal Rajkumar et al. (2008) reported CH_4 fluxes of 3.6 mmol m⁻² day⁻¹ from an estuarine system (Adyar) in India. Maher et al. (2015) found CH_4 fluxes in a subtropical Australian estuary to be 0.57 mmol m⁻² day⁻¹, and Linto et al. (2014) and Call et al. (2015) reported CH_4 fluxes from tidal mangrove estuaries to be 0.35 and 0.21 mmol m⁻² day⁻¹, respectively.

Average CO₂ atmospheric fluxes were 1.8 times higher in wet (1128 mmol $m^{-2} day^{-1}$) than the dry seasons

		DIC	DOC	Alkalinity	Free CO ₂	CH_4
Wet season	mmol day ⁻¹	$43.5 \times 10^7 \pm 8.6 \times 10^7 (48 \times 10^7)*$	$25.5 \times 10^7 \pm 5.0 \times 10^7$	$48.7 \times 10^7 \pm 9.6 \times 10^7$	$13.8 \times 10^7 \pm 2.6 \times 10^7$	$0.009 \times 10^7 \pm 0.002 \times 10^7$
	$mmol m^{-2} day^{-1}$	$24.1\pm4.7~(26)~*$	14.1 ± 2.7	27.0±5.3	7.6±1.4	0.005 ± 0.001
	Shallow GW contribution (mmol day ⁻¹)	$43.6 \times 10^6 \pm 43.6 \times 10^6$	$37.4 \times 10^{6} \pm 37.4 \times 10^{6}$	$10.9 \times 10^{6} \pm 10.9 \times 10^{6}$	$28.4\!\times\!10^6\!\pm\!28.4\!\times\!10^6$	$2.2 \times 10^5 \pm 2.2 \times 10^5$
	Deep GW contribution (mmol day ⁻¹)	$81.5 \times 10^{6} \pm 11.2 \times 10^{6}$	$65.5 \times 10^{6} \pm 84.8 \times 10^{6}$	$70.5 \times 10^{6} \pm 82.5 \times 10^{6}$	$61.3 \times 10^{6} \pm 12.8 \times 10^{6}$	$3.2 \times 10^{6} \pm 2.0 \times 10^{6}$
Dry season	mmol day ⁻¹	$28.2 \times 10^7 \pm 5.6 \times 10^7 (29 \times 10^7)^*$	$5.1 \times 10^{7} \pm 1.0 \times 10^{7}$	$34.2 \times 10^7 \pm 6.8 \times 10^7$	$0.9 \times 10^7 \pm 1.8 \times 10^6$	$0.02 \times 10^7 \pm 4.0 \times 10^4$
	$\mathrm{mmol}\ \mathrm{m}^{-2}\ \mathrm{day}^{-1}$	15.6 ± 3.1 (16) *	2.8 ± 0.5	19.0 ± 3.7	$0.5 {\pm} 0.1$	0.01 ± 0.01
	Shallow GW contribution (mmol day ⁻¹)	$24.7 imes 10^6 \pm 14.9 imes 10^6$	$21.2\!\times\!10^6\!\pm\!60.1\!\times\!10^6$	$6.1 \times 10^{6} \pm 22.4 \times 10^{6}$	$16.1\!\times\!10^6\!\pm\!20.5\!\times\!10^6$	$0.1 \times 10^{6} \pm 1.0 \times 10^{6}$
	Deep GW contribution (mmol day ⁻¹)	$4.0 \times 10^6 \pm 1.1 \times 10^6$	$3.2 \times 10^6 \pm 8.4 \times 10^6$	$0.7 \times 10^{6} \pm 8.2 \times 10^{6}$	$19.8 \times 10^{6} \pm 83.4 \times 10^{6}$	$0.2 \times 10^{6} \pm 1.0 \times 10^{6}$
Average	mmol day ⁻¹	$35.8 \times 10^7 \pm 7.0 \times 10^7$	$15.3 \times 10^7 \pm 3.0 \times 10^7$	$41.4 \times 10^{7} \pm 8.2 \times 10^{7}$	$7.3 \times 10^7 \pm 1.4 \times 10^7$	$0.01 \times 10^7 \pm 0.002 \times 10^7$
	$\mathrm{mmol}\ \mathrm{m}^{-2}\ \mathrm{day}^{-1}$	19.8 ± 3.8	8.5 ± 1.6	23.0±4.5	$4.0 {\pm} 0.7$	0.005 ± 0.001
	Total GW contribution (Shallow+deep) (mmol day ⁻¹)	$77.0 \times 10^{6} \pm 31.9 \times 10^{6}$	$63.7 \times 10^{6} \pm 98.4 \times 10^{6}$	$16.3 \times 10^{6} \pm 18.8 \times 10^{6}$	$35.1 \times 10^6 \pm 58.8 \times 10^6$	$0.5 \times 10^{6} \pm 2.1 \times 10^{6}$

 $(620 \text{ mmol m}^{-2} \text{ dav}^{-1})$ (Table 3). This difference may be related to higher temperatures in the wet season (summer) and subsequent higher rates of in situ respiration, and/or nitrification which has a net effect of decreasing alkalinity and pH and therefore increasing pCO_2 (Frankignoulle et al. 1996; Gazeau et al. 2005; Borges and Abril 2011; Maher and Eyre 2012). Interestingly, water to air CH₄ fluxes where higher in the dry $(43 \text{ mmol m}^{-2} \text{ day}^{-1})$ than the wet season (17 mmol $m^{-2} day^{-1}$), driven by higher concentrations (Fig. 2) rather than higher transfer velocities (Table 3). This is in spite of higher groundwater inputs in the wet season (Table 2). This may be due to seasonal differences in methane oxidation rate (Abril and Iversen 2002) or alternative sources or production rates of CH₄ within the estuary during the two seasons. Further studies would be required to assess the factors controlling seasonal variability in surface water CH₄ dynamics.

While CO₂ emissions may dominate carbon gaseous fluxes, CH₄ emissions could have a greater impact on global warming potential of the system (Gatland et al. 2014). Although CH₄ losses to the atmosphere were smaller than CO_2 , CH_4 is a more potent greenhouse gas compared to CO₂, therefore, accounting only for CO₂ evasion in systems where there may be high CH₄ emissions, could result in an underestimation, in terms of global warming potential of the system (Gatland et al. 2014; Panneer Selvam et al. 2014; Neubauer and Megonigal 2015). Here, after CH₄ fluxes were converted into CO₂-equivalent emission estimates assuming a 100 year CH₄ sustained-flux global warming potential [i.e., 1 kg CH₄=45 kg of CO₂; Neubauer and Megonigal (2015)], CH₄ accounted for ~50 % of CO₂-equivalent emissions from the estuary for both seasons, and therefore was significant in terms of greenhouse gas emissions.

Carbon Surface Water Exports

mmol day⁻¹ / 116160 m²)]. Estuary area: 116160 m²; Catchment area: 18,000,000 m²

*Values in bracket indicate calculated DIC using TAlk and pCO_2

 $day^{-1} / (35.8 \times 10^7)$

Table 5 presents estimates of DIC and DOC export to coastal waters from small estuarine and large riverine systems. In a review paper, Cai (2011) reported the global riverine DOC export rate to be 246 Tg year⁻¹. Bauer and Bianchi (2011) also reported a similar global oceanic DOC export rate $(250 \text{ Tg year}^{-1})$. Based on the world wide surface areas of estuaries which is 1.05×10^{12} m² (Cai 2011), global DOC export from estuarine systems is estimated to be 0.64 g C m⁻² day⁻¹. Hat Head estuary exported 15.5 g C m⁻² of estuary day^{-1} of DOC, which is 24 times higher than the global estuarine DOC export estimate. DOC export per unit area from Hat Head estuary was also higher than much larger riverine systems (Striegl et al. 2007; Cai et al. 2008). Based on Table 5, DOC exports from systems of a similar small size to Hat Head estuary are higher; however, it should be noted that exports reported in Adame and Lovelock (2011), Bergamaschi et al. (2012), Maher et al. (2013a), Wang and

Fig. 4 Calculated versus measured DIC in surface water and groundwater from the downstream station located at the mouth of the estuary in wet and dry seasons. The *line* represents the 1:1 ratio



Cai (2004), and Winter et al. (1996) are from mangrove or salt marsh systems which essentially have a minimal catchment area to water area ratio, thereby inflating the mmol C m^{-2} catchment day $^{-1}$ (i.e., essentially all the catchment is intertidal). Moreover, DIC exports from the small Hat Head estuary were much higher than larger riverine systems on a catchment area basis (Table 5). Hat Head estuary DIC yield (i.e., export per unit of catchment area) were four times higher than the Gulf of Trieste catchment (Tamše et al. 2014), 11-40 times higher than the Yukon River (Striegl et al. 2007), 16-55 times higher than the six largest Arctic Rivers (Tank et al. 2012) and two orders of magnitude higher than DIC exports reported for the Chena River in Alaska (Cai et al. 2008) and the Guadalquivir Estuary, Spain (De La Paz et al. 2007). In comparison with smaller estuaries, Hat Head DIC yield was two orders of magnitude higher than the York River estuary (Raymond et al. 2000) and comparable with intertidal mangrove and saltmarsh systems (Wang and Cai 2004; Bouillon et al. 2008; Maher et al. 2013a; Winter et al. 1996).

The source of DIC is possibly from the surrounding mangroves, as Hat Heat estuary has an extensive mangrove environment throughout the estuary (Fig. 1). Mangrove environments tend to have high DIC export rates (Bouillon et al. 2008; Miyajima et al. 2009; Maher et al. 2013a). Moreover, a DIC versus salinity scatter graph (Fig. 5) showed a slight concave upward trend (at least in the dry season) which suggests mid-estuary inputs of DIC (perhaps from mangrove groundwater). DOC, however, had a conservative or sink nature in relation to salinity (Fig. 5), which suggests an upstream source (likely the freshwater wetlands or groundwater, see also Sanders et al. 2015) with some loss during estuarine transport (respiration, photomineralization, or flocculation). Our mass balance approach suggests significant groundwater inputs of all dissolved carbon species, yet the traditional salinity mixing model approach does not indicate a clear source. Wang et al. (2015) found a significant groundwater source of DIC in the Jiulong estuary (China), yet salinity versus DIC indicated conservative mixing. The authors suggested that the diffuse nature of SGD-derived solute inputs may lead to no deviation from conservative mixing, which may also be the case in Hat Head estuary, and other estuarine systems that have large areas of diffuse groundwater input along the estuary.

Alkalinity export to the coastal ocean ranged from 19 to 27 mmol m^{-2} of catchment day⁻¹ in dry and wet seasons, respectively (Table 4). Previous studies have reported alkalinity exports, from larger systems; however, studies with time series sampling for calculating alkalinity exports from tidal estuaries are still scarce. Faber et al. (2014) identified that DIC export was mostly alkalinity in a mangrove- and seagrass-dominated tidal creek in southeast Australia. They reported export rates ranging from 140 to 460 mmol m^{-2} of water area day⁻¹, which was an order of magnitude lower than alkalinity export estimates from Hat Head estuary $(3564 \text{ mmol m}^{-2} \text{ of water area day}^{-1})$. Santos et al. (2015) reported groundwater-derived alkalinity exports 4.7 times higher than Hat Head from tidal flats in the Wadden Sea (Germany) where porewater alkalinity concentrations are extremely high at 20 mM. Alkalinity production has a significant influence on the global carbon budgets by affecting the oceanic carbonate system. In the case of alkalinity production, carbon is not lost to the atmosphere as CO₂ and is exported to the ocean and acts as a buffer which facilitates the uptake of extra CO₂ (Faber et al. 2014).

Table 5 shows that a general negative correlation may exist between dissolved carbon yield and catchment area and that small estuarine systems have the ability to deliver more dissolved organic carbon to the coastal ocean compared to larger riverine systems on a catchment area basis. This is mainly due to the shorter residence time of estuaries which reduces the potential for biogeochemical processes to modify the quantity and composition of organic matter. This highlights the importance of studying small systems with a short residence time, such as Hat Head, to obtain a better quantitative understating of global carbon exports to the ocean.

Groundwater-Derived Carbon Fluxes

We could not collect deep groundwater samples during the wet season, and shallow samples were only collected from

Location	Description	Country	Catchment size (km ²)	DIC export	DOC export	Reference
Hat Head	Tidal estuary	Australia	18	19.8	8.5	This study (2015)
Duplin River, on Sapelo Island	Marsh-dominated estuary	USA	2	24.9*		Wang and Cai (2004)
Southwest Florida coast	Mangrove-dominated estuary	USA	230		41.1	Bergamaschi et al. (2012)
Southern Moreton Bay	Mangrove-dominated estuary	Australia	0.4	250.0	25.0	Maher et al. (2013a)
Average from different mangrove systems	Micro-tidal mangrove	USA			16.7	Adame and Lovelock (2011)
York River estuary-Chesapeake Bay	River estuary	USA	4	0.67		Raymond et al. (2000)
Swartkops estuary	Salt marsh estuary	South Africa	4	247.3	23.5	Winter et al. (1996)
Jiulong River estuary	River estuary	China	71	2.4 - 3.9		Wang et al. (2015)
Range of different mangrove systems	Mangrove estuaries		1992	254.0	34.2	Bouillon et al. (2008)
Chena River, Alaska	River	USA	5200	0.9	0.3	Cai et al. (2008)
Guadalquivir Estuary (SW Iberian Peninsula)	River estuary	Spain	58,000	0.2		de la Paz et al. (2007)
Gulf of Trieste (North Adriatic)	River	Italy	500	5.0		Tamše et al. (2014)
Yukon River	River	Canada-USA	853,300	0.5-1.7	0.25-0.4	Striegl et al. (2007)
6 Arctic rivers	River	Europe	10,900,000	0.35-1.2		Tank et al. (2012)

Table 5 Previous studies estimating DIC and DOC export (yield) to coastal waters from small and large estuaries

Units are mmol C m⁻² of catchment day⁻¹



Fig. 5 DIC, DOC, alkalinity, pCO_2 , and CH₄ versus radon, salinity, and depth in estuary surface water in wet (March) and dry (June) seasons from the downstream station located at the mouth of the estuary. *Lines* indicate the theoretical conservative mixing

the most downstream station during the wet season field campaign. We acknowledge the limitations with this approach. However, shallow samples collected at the downstream station were similar during both seasons (averages within 10%), and previous studies have found that deep groundwater has relatively stable composition (Dhar et al. 2008; Chapagain et al. 2010). Further, shallow groundwater only dominates inputs during the dry season (Sadat-Noori et al. 2015), when we have adequate sampling coverage throughout the estuary to constrain the shallow groundwater endmember. Considering the uncertainty in shallow groundwater composition during the wet season (i.e., we have used the dry season data to estimate this), we have assigned a 100 % uncertainty to this term in our calculations (Table 4). The relative contributions of deep and shallow groundwater carbon inputs during wet and dry seasons basically follows the groundwater discharge rates with deep groundwater dominating carbon inputs in the wet season and shallow groundwater delivering more carbon in the dry season. Groundwater fluxes of each carbon species could not be calculated for each individual section during each season due to the lack of groundwater and surface water samples in the upper reaches of the estuary. Surface water samples were only collected at the downstream station for carbon parameters other than pCO_2 .

Another limitation to our groundwater fluxes is that the average flux presented here only considers the differences between the wet and dry season while other factors such as differences in spring-neap tide cycles and annual temperature variability may influence the groundwater discharge flux (de Sieyes et al. 2008; Constantz et al. 1994) and estuarine carbon fluxes. Moreover, tidal variability can also influence groundwater discharge rates and consequently carbon fluxes, as tidal pumping releases shallow saline groundwater into the estuary (Call et al. 2015, Maher et al. 2015; Santos et al. 2009). Tidal pumping was the dominant source of groundwater discharge in the dry season making the shallow saline groundwater. Therefore, some of the differences that were observed may be due to differences in tidal pumping.

Most of the carbon input to surface waters via groundwater was in the form of DOC and DIC and the smallest portion was contributed by CH_4 (1 %) in both seasons (Fig. 6). The total groundwater-derived DIC flux entering surface waters was 4.6 times higher in the wet season compared to dry. While the total (deep+shallow) average groundwater-derived DIC fluxes from both seasons ($687 \pm 117 \text{ mmol m}^{-2}$ of estuary day⁻¹) were comparable to previous studies by Santos et al. (2012a), Dorsett et al. (2011), and Cai (2003) (see Table 6), higher DIC fluxes have been reported in salt marshes/estuaries (Moore et al. 2006) and freshwater tidal creeks (Atkins et al. 2013) (Table 6). The total (deep+shallow) average (wet and dry) groundwater-derived DOC fluxes found here $(540\pm731 \text{ mmol m}^{-2} \text{ day}^{-1})$ are high, being at least 3-fold higher than previous studies which have reported groundwater-derived DOC fluxes ranging from 21 to 170 mmol m^{-2} day⁻¹ (Santos et al. 2012a; Maher et al. 2013a; Porubsky et al. 2014) (Table 6). These high fluxes are related to the high DOC concentrations in groundwater (Table 1).

 pCO_2 in the water column was positively correlated with radon (Fig. 5), indicating that groundwater is a source of free CO_2 and/or [H⁺]. The strong relationship between pCO_2 , CH₄, and radon (groundwater tracer) during both wet and dry season field campaigns (Fig. 5), in addition to the high groundwater pCO₂ and CH₄ concentrations, implies groundwater was a major driver of surface water pCO_2 and CH_4 in Hat Head estuary. This influence could either be directly by discharging pCO_2 - and CH₄-enriched groundwater into the estuary or indirectly by groundwater delivering DOC, which can boost ecosystem respiration and increase pCO_2 (Maher et al. 2015). The later process is further supported by the DOC versus radon plot (Fig. 5) which shows a positive correlation. The salinity mixing plots (Fig. 5) show that pCO_2 and CH₄ have a concave trend with salinity indicating an upstream input likely being groundwater discharge from the upper reaches as this area was found to be a groundwater hotspot in a concurrent study by Sadat-Noori et al. (2015).

Flux calculations offer stronger evidence that groundwater plays a major role in delivering greenhouse gasses to surface waters. The fluxes obtained from the groundwater mass balance approach show that groundwater-derived free CO₂ and CH₄ inputs can account for a large proportion (54 % for CO₂ and 46 % for CH₄) of the observed atmospheric fluxes (Table 3). Previous studies have also found groundwater to be a major driver of surface water pCO₂ and CH₄ (Faber et al. 2014; Macklin et al. 2014; Call et al. 2015). Atkins et al. (2013) reported that groundwater-derived CO₂ fluxes into a flood plain creek estuary, NSW, Australia, averaged 1622 mmol m⁻² day⁻¹, a value twice as high as atmospheric CO₂ evasion in the area, and 1.5 times larger than CO₂ fluxes in our case. Conversely, Porubsky et al. (2014) stated groundwaterderived CH₄ fluxes were 0.8 mmol m⁻² day⁻¹ in Okatee estuary in the USA, 15 times smaller than fluxes from Hat Head estuary.

Radon also had a positive correlation with DOC especially in the wet season, but not as clear as pCO_2 and CH₄, likely due to multiple processes driving surface water DOC dynamics in this system. For example, radon and DOC may have the same source (groundwater) but have different loss pathways as radon is a gas and its major loss pathway is atmospheric evasion, while DOC maybe lost through respiration, flocculation, and photomineralization. This creates a decoupling between radon and DOC which may not be so apparent as in the radon vs CO₂ and CH₄ plots (Fig. 5) and explains the stronger correlation between radon and the greenhouse gases.

Estuarine CO₂ is driven by biological (productivity/respiration), hydrological (groundwater inputs and mixing of riverine and oceanic waters), and physical (temperature-driven groundwater convection and wind-driven evasion) process (Borges and Abril 2011; Maher et al. 2015). Here, the contribution of groundwater to CO₂ loss (evasion and export (Tables 3 and 4)) from the estuary was on average 31 % over both seasons. The groundwater contribution to CH₄ loss (evasion and export) was around 46 %, with significant differences in wet and dry seasons. This can be explained through the difference in the groundwater discharge rate between wet and dry seasons (Table 2), where the groundwater contribution is a function of groundwater discharge (which varied significantly in different hydrological conditions at our site) and groundwater end-member concentration (which was assumed to be the same during both seasons).

Dissolved carbon is transported from groundwater to the estuary and then the atmosphere (CO_2 and CH_4) and the ocean (DIC and DOC). Figure 3 illustrates this for CO_2 , where oceanic waters enter the estuary with the flood tide, becomes enriched in dissolved carbon within the estuary due to mixing with groundwater (and upstream wetland surface water) and leaves the system via ebb tide flow and atmospheric evasion. By investigating the CO_2 versus salinity plot for the 4 stations (Fig. 7), we found hysteresis occurring at some stations with different CO_2 values at the same salinity during the ebb and flood tide. For instance, observations at station 3 in the wet season showed that as the salinity starts to decrease from flood

Fig. 6 Average (wet and dry season) portions of carbon species derived by groundwater and losses from the mouth of the estuary assuming that alkalinity fluxes are related to carbonate alkalinity



tide, CO_2 remains low for 8 h. This continues until brackish waters occupy the station (salinity ~8), implying that the initial fresh water entering the estuary is fresh surface water relatively low in CO_2 . A subsequent increase in CO_2 values implies the input of groundwater during the ebb tide, indicating a delayed groundwater input to the estuary. This interpretation is supported by the CO_2 versus radon scatter that shows no hysteresis (Fig. 5).

Total average groundwater-derived CH₄ fluxes ($12\pm 17 \text{ mmol m}^{-2} \text{ day}^{-1}$) (Table 2) were much higher than groundwater-derived CH₄ fluxes from a small tidal river estuary, in Okatee, USA (0.9 mmol m⁻² day⁻¹, Porubsky et al. 2014). They were also three orders of magnitude higher than CH₄ fluxes on the Florida Gulf Coast (Cable et al. 1996).

Groundwater-derived CH_4 fluxes were accountable for almost all the export from the estuary, and much of the free CO_2 export could be attributed to groundwater inputs (Table 4) indicating the major role of groundwater in CH_4 and dissolved CO_2 transported between terrestrial and aquatic environments.

Average DIC and DOC export from the estuary (3081 ± 602 and 1317 ± 258 mmol m⁻² day⁻¹, respectively) were 4.4- and 2.4-fold higher than groundwater-derived DIC (687 ± 117 m m o 1 m⁻² d a y⁻¹) a n d DOC (540 ± 731 mmol m⁻² day⁻¹), indicating that groundwater can account for almost half of the dissolved organic carbon export from the estuary. In other words, the average contribution of groundwater to DOC export in both seasons was ~41 % while groundwater contributed ~22 % to DIC export with

Location	System description	GW-derived DIC fluxes	GW-derived DOC fluxes	GW-derived TAlk fluxes	GW-derived CH ₄ fluxes	GW trancing method	Reference
Hat Head, Australia	Tidal estuary	687.3	540.2	128.1	12.5	Ra isotopes	This study (2015)
Jiulong River estuary, China	River estuary	121-897		91–748		Ra isotopes	Wang et al. (2015)
Moreton Bay, Australia	Embayment	153	36	161		Ra isotopes	Stewart et al. (2015)
Wadden Sea, Germany	Tidal flats		42	1344		²²² Rn	Santos et al. (2015)
Heron Island, Australia	Coral reef				0.01	²²² Rn	O'Reilly et al. (2015)
Okatee Estuary, USA	Salt marsh/estuary	1079	64		0.8	Ra isotopes	Porubsky et al (2014)
North Creek, Australia	Fresh water tidal creek	1810				²²² Rn	Atkins et al. (2013)
Moreton Bay, Australia	Mangrove tidal creek	250	24			^{222}Rn and δ ^{13}C	Maher et al. (2013a)
Heron Island, Australia	Permeable carbonate sediments	1.6–18.8		5.1-8.8		²²² Rn	Cyronak et al. (2013)
Yarra River, Australia	Salt wedge estuary	349	21			²²² Rn	Santos et al. (2012a)
Indian River Lagoon, USA	Coastal lagoon	120–340				$\delta^{13}C$ and $\delta^{14}C$	Dorsett et al. (2011)
West coast of Florida, USA	Sandy beach		19–27			Ra isotopes	Santos et al. (2009)
Okatee Estuary, USA	Salt marsh/estuary	1963	170			Ra isotopes	Moore et al. (2006)
North Inlet, USA	Salt marsh/estuary	171				Ra isotopes	Cai (2003)
South Carolina, USA	Tidal creek		50			Piezometer	Goñi and Gardner (2003)

Table 6 Previous studies estimating groundwater derived DIC, DOC, TAlk, and CH₄ using natural tracers

Units are in mmol m^{-2} of estuary day⁻¹. Table updated from Atkins et al. (2013)



Fig. 7 CO_2 versus salinity scatter plots in wet and dry seasons at the four stations along the estuary showing hysteresis

Fig. 8 Conceptual diagram of the study area summarizing flux estimates±standard error of groundwater-derived, estuary export and atmospheric evasion of carbon from the mouth of the estuary in (a) wet and (b) dry seasons. Groundwater discharge rates are in $m^{-3} s^{-1}$, and all other parameters are in units of 10^4 mol day⁻¹ (estuary area= 116,160 m² and catchment area= 18 km²)

considerable differences in wet and dry seasons (Table 4). Maher et al. (2013a) also found that groundwater advection was a dominant pathway for DOC export and was responsible for 90 % of DOC export in a mangrove tidal estuary. Faber et al. (2014) reported that 90 % of the carbon loss from an estuary system was from groundwater DIC advection while DOC only accounted for 5 %. Liu et al. (2014) reported that groundwater DIC fluxes were 11–71 times higher than the combined input of local rivers, suggesting that SGD was the dominant source of DIC to the southwest Florida Shelf, USA. Wang et al. (2015) found that SGD input of DIC to the Jiulong River estuary in China was the equivalent to between 25 and 110 % of riverine DIC exports.

Groundwater had a minor contribution to TAlk input into the estuary (~3 %) in both seasons (Table 4). Average alkalinity export from estuary was almost 28-fold higher than groundwater-derived alkalinity inputs, suggesting that processes other than groundwater input are driving alkalinity export from this system. This alkalinity is thought to come from sulfate reduction in shallow porewaters (Faber et al. 2014); however, our groundwater sampling resolution was not adequate to capture this process (alkalinity may be produced in the upper cm of sediments, while our groundwater samples were taken from areas deeper than 1 m).

To summarize the key findings of this study, we present a conceptual diagram (Fig. 8) that illustrates the (1) groundwater discharge rates, (2) flux estimates of groundwater-derived



carbon into the estuary, (3) estuary carbon export, and (4) carbon atmospheric evasion in wet and dry conditions in the whole estuary system. The large variability observed in groundwater discharge and carbon loss rates over a relatively short time scale indicates the need for more frequent measurements to be carried to assess the influence of groundwater on carbon cycling. Nevertheless, it is clear that this small wetland-surrounded subtropical estuary has a high carbon yield (in terms of both oceanic export and air–water exchange), and groundwater carbon inputs play a major role in estuarine carbon cycling in this system. Combined with the recent literature, this investigation demonstrates that groundwater may play a major role in estuarine carbon dynamics.

Conclusions

The Hat Head estuary had a high area normalized export rate of DIC $(3081\pm602 \text{ mmol m}^{-2} \text{ day}^{-1})$, DOC $(1317\pm$ 258 mmol m^{-2} day⁻¹), and TA1k (3564 ± 705 mmol $m^{-2} day^{-1}$) to the coastal ocean and groundwaterderived carbon inputs were a significant component of this carbon export. Groundwater contribution to carbon loss from the estuary for DIC, DOC, TAlk, free CO2 and CH4 was found to be approximately 22, 41, 3, 75, and 100 %, respectively. The average estuary-wide CO2 and CH4 evasion rates were 870 ± 174 and 26 ± 5 mmol m⁻² day⁻¹ (some of the highest estuarine fluxes reported yet), and groundwater discharge accounted for 54 and 46 % of these evasions, respectively. Our observations indicate that small estuarine systems with a short residence time can pump more carbon to the coastal ocean compared to some larger riverine systems on a catchment area basis, and that groundwater exchange may deliver large amounts of carbon to surface estuarine waters.

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