

The Early Life Origins of Cardiovascular Disease

Robert Murray · Keith M. Godfrey · Karen A. Lillycrop

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Abstract Cardiovascular disease continues to impose a high societal and economic burden. Although it occurs primarily in later life, there is strong evidence that it originates in early life. The nutritional environment that an unborn child is exposed to can heavily influence later disease risk, with nutritional exposures altering organ development and programming metabolic changes that are then maintained during the life course. Epigenetic changes induced by the early life environment are thought to be a key mechanism by which these early life events influence subsequent disease risk. Here, we review the emerging role of epigenetics in the development of cardiovascular disease.

Keywords Cardiovascular disease · Early life origins · Epigenetic changes · Epigenetics

Introduction

Non-communicable diseases (NCDs) such as diabetes, cardiovascular disease, obesity and cancer collectively represent one

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R. Murray · K. M. Godfrey
Faculty of Medicine, Academic Unit of Human Development and Health, Southampton, UK

K. M. Godfrey
MRC Lifecourse Epidemiology Unit and NIHR Southampton Biomedical Research Centre, University Hospital Southampton NHS Foundation Trust, University of Southampton, Southampton, UK

K. A. Lillycrop (✉)
Centre for Biological Sciences, Faculty of Natural and Environmental Sciences, Institute of Developmental Sciences, Southampton General Hospital, University of Southampton, Tremona Road, Southampton SO16 6YD, UK
e-mail: kal@soton.ac.uk

of the greatest challenges to global health in the twenty-first century [1]. Globally, NCDs kill 38 million people a year, accounting for approximately 8 out of every 10 deaths in the developed world [2]. NCDs are not restricted to industrialized countries; they are also the leading cause of death in both low and middle income countries [3]. Countries undergoing rapid socio-economic improvement are especially vulnerable, as while the burden of infectious diseases decreases with advances in health care provision, the incidence of NCDs has increased rapidly along with the adoption of more sedentary lifestyles and consumption of westernized diets [4, 5]. Together, these factors are predicted to drive a 17 % increase in NCD levels over the next decade [3]. This book chapter will explore the role of the early life environment in the development of cardiovascular disease, and examine the growing evidence that epigenetic changes underlie this link between early life and later CVD disease risk.

Aetiology of CVD

Cardiovascular disease (CVD) is the single largest cause of death among NCDs, responsible for almost a third of all NCD deaths [2]. CVD encompasses a class of diseases involving both the heart and the wider circulatory system, and includes cardiac hypertrophy, damage to blood vessels, endothelial dysfunction and arterial narrowing that can lead to heart failure, stroke and myocardial infarction. Atherosclerosis and hypertension are considered the two main underlying causes of CVD [6]. There are a number of risk factors associated with CVD including family history and age as well as a number of modifiable risk factors which include obesity, hypertension, high cholesterol concentrations, physical inactivity, diabetes, poor nutrition and smoking [7, 8].

Obesity is the largest modifiable risk factor for CVD [8]. Obesity levels are increasing at a dramatic rate: global obesity levels having doubled since the 1980s, with 55 % of European adults now overweight and 20 % clinically obese [3]. Obesity increases metabolic load due to increased blood volume

servicing the greater adipose mass, leading to increased pre-load on the heart, while afterload is increased by arterial stiffness, with obese adults often exhibiting both concentric and eccentric hypertrophy [6, 9]. Obesity is also thought to account for as much as 70 % of essential hypertension [10]. Obesity among women of child-bearing age has also steadily increased over the last 20 years, with some western countries now reporting that two thirds of potential mothers are overweight and a third obese [11, 12]. This is of particular concern as there is mounting evidence that exposure to maternal obesity in utero can increase adult CVD risk. Consistent with the rise in maternal obesity, obesity among children is also increasing globally with some western countries reporting that a third of children are overweight, of whom half are clinically obese [2]. Childhood CVD should be rare, but is increasing along with childhood obesity. Obese children exhibit several physiological signs of cardiovascular dysfunction, independent of obesity related co-morbidities, with altered cardiac structure [13] and increased regional deformation of the left ventricle [14•]. By adolescence, obese children display both diastolic and systolic dysfunction, along with reduced ventricular strain rate [15, 16]. Childhood obesity also promotes endothelial dysfunction leading to atherosclerotic plaque formation, hypertrophy and ultimately, cardiac dysfunction [9, 17].

The Early Life Environment and the Risk of NCDs

Research into CVD and its co-morbidities initially focused on the disease in adulthood, but there is now increasing evidence that CVD may originate in early life [18]. The early life environment has been shown to greatly influence future health, with nutrition during pregnancy and in early post-natal life impacting upon organ development and metabolic regulation, influencing later disease risk [19].

The initial link between adult CVD risk and the early life environment came from a Norwegian study that found an association between CVD in middle age and under nutrition and poverty during childhood and adolescence [20]. Later work in Britain by David Barker and colleagues expanded on these observations by comparing birth measurements with health in middle age. Here, they found a strong correlation between low birth weight (LBW) and coronary heart disease (CHD), as well as diabetes, increased systolic blood pressure and hyperlipidemia [21–24]. Subsequent cohort studies have confirmed these associations, with LBW linked to an increase in CVD risk factors in childhood [25•, 26–28], leading to greater mortality risk from stroke and CHD in adult life [27, 29, 30].

While these epidemiological studies demonstrated the relationship between foetal growth and CVD risk, studies carried out on people whose mothers were exposed to the Dutch Hunger Winter, a famine that occurred in the Netherlands in 1944–1945, revealed the importance of maternal nutrition to

the offspring's health in later life. In these studies, individuals born to mothers exposed to famine during the periconceptual period up to the first trimester of pregnancy had an increased risk of CVD and obesity in middle age, whereas individuals that were exposed to famine in the later stages of gestation showed an increased risk of developing insulin resistance and hypertension in adulthood [31, 32]. Such findings also suggest that the timing of the dietary constraint may be important and may determine which organ system is affected.

Postnatal growth trajectory has been linked to CVD risk, with evidence that rapid catch up growth among those born with a low birth weight impacts upon endothelial function and subsequent atherosclerosis [6, 33] leading to hypertension and coronary heart disease in adulthood [26, 34].

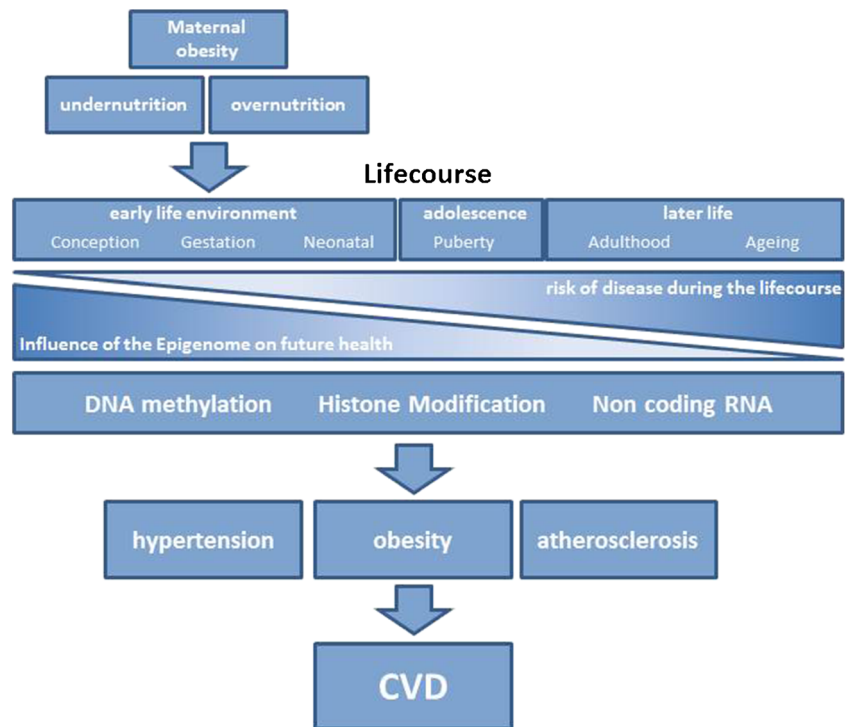
A number of studies have also shown a J- or U-shaped relationship between birth weight (BW) and disease risk with babies born at the highest BW also being at increased risk of developing CVD and other NCDs [33, 35–39] (Fig. 1). Maternal obesity and weight gain during pregnancy have also been associated with subsequent obesity [40] and an increase in offspring systolic blood pressure [41]. Further studies have also linked parental BMI and offspring cardiovascular risk factors such as increased diastolic and systolic blood pressure, elevated insulin levels and lowered HDL [8, 42•, 43], and more specifically, shown an association between increased maternal BMI during pregnancy and later death from cardiovascular events [26, 44•]. Maternal obesity has been shown to influence the BMI of the grandchildren [45], implying a transgenerational effect that has far reaching implications for the health of future generations [38]. Maternal obesity may affect the developing foetus through intrauterine interactions and while confounding factors in human studies prevent inferences about the mechanisms of disease, animal models have allowed investigation of the potential mechanisms underlying these associations.

Animal Models of Nutritional Programming

Animal models have sought to replicate findings from human epidemiological studies that show an association between the quality of the early life environment and future disease risk. Models of maternal undernutrition, such as global dietary and protein restrictions, as well as models of maternal overnutrition such as high fat diets and maternal obesity, have resulted in offspring that exhibit persistent metabolic changes often leading to features similar to human cardio-metabolic disease such as obesity, insulin resistance, hypertension and raised serum cholesterol levels in later life [46].

Maternal exposure to a protein-restricted diet alters offspring metabolism, leads to impaired glucose homeostasis [47], increased fat deposition, altered feeding behaviour towards a preference for high-fat foods [48, 49] and vascular

Fig. 1 The association between maternal diet on subsequent offspring CVD risk. Birth weight is often used as a surrogate for an adverse intrauterine environment. Being born either underweight or overweight is associated with an increased CVD risk, effecting cardiovascular health through the development of hypertension and atherosclerosis. There is, however, much evidence that developmental influences can programme increased CVD risk without necessarily affecting birth weight



dysfunction and hypertension [50]. Pre-natal undernutrition, particularly during the late intrauterine period, also results in restricted kidney development that reduces final nephron numbers, which disrupts kidney natriuresis [51–53].

The induction of persistent changes to the metabolism and physiology of the offspring by perturbations in maternal diet is accompanied by changes in the expression of key metabolic regulators [54–56]. For example, maternal protein restriction has been shown to directly impact upon offspring vascular function, promoting hypertension [50, 57] and increasing oxidative stress; the latter is a key inducer of endothelial dysfunction, leading to atherosclerosis and is accompanied by the upregulation of enzymes that produce reactive oxygen species, while at the same time, a decrease in antioxidant activity [58, 59]. Maternal protein restriction has also been shown to alter adrenal gene expression, with offspring exhibiting decreased expression of type II adrenal receptors, reducing the capacity for negative feedback that lowers blood pressure [53, 60, 61], while adrenal type I receptor density was increased, upregulating production of adrenal aldosterone, which raises blood pressure by promoting nephron sodium and water reabsorption [60, 62]. Maternal exposure to global dietary restriction also influences blood pressure by inducing long-term changes in the expression of 11β -hydroxysteroid dehydrogenase type 2 (HSD11 β), a key regulator of active glucocorticoid levels that play a central role in regulation of blood pressure [63, 64].

With the increasing prevalence of obesity, particularly among mothers of child-bearing age, several animal models

have been used to examine the effects of exposure to a western style maternal diet enriched in fat and sugar or the effects of maternal obesity on the later health of the offspring. Offspring of mice fed an obesogenic diet during pregnancy and lactation developed endothelial dysfunction and hypertension, and also exhibited increased fasting insulin and plasma glucose levels as well as a dysregulation of appetite [65]. Exposure to a maternal high fat diet can also induce structural changes in the foetal heart, with offspring having an accelerated cardiac growth rate leading to impaired baseline cardiac function and cardiac hypertrophy. At a molecular level, maternal obesity triggered re-expression of cardiac foetal genes in the offspring that altered cardiac muscle structure [66]. Exposure to maternal obesity also alters the regulation of the renin-angiotensin system, which regulates blood pressure and water balance; Guberman and colleagues found that exposure to a maternal high fat diet in a rat model stimulated adipose angiotensin (AGT) production, causing hypertension in the offspring. Additionally, adipose production of AGT stimulated lipogenesis, resulting in increased adiposity that altered adipose tissue angiogenesis and smooth muscle tone, further exacerbating the animals' hypertensive state [67].

Developmental Plasticity

The influence of the early life environment on later phenotype may in part reflect predictive adaptive responses that allow

organisms to adjust their development in response to environmental cues, to aid fitness or survival [68]. This developmental plasticity allows change and adaptation to the environment within a set developmental window, after which changes can persist throughout the life course. Early life adaptation could result in later disease, if an organism adapts to one environment but is then exposed to a different environment post birth, creating a mismatch [69]. Mismatches between pre-natal and post-natal environments may be a key factor behind the rapid rise of NCDs in developing countries, as they undergo rapid urbanization and sudden shifts in dietary composition [70–72].

Epigenetics Mechanisms

Genetic differences between individuals cannot explain the inter-individual differences in NCD disease risk [73]. Firstly, the overall proportion of risk variance accounted for by genetic variation accounts for less than 5 % of the observed differences [74, 75], and secondly, fixed genomic variation cannot explain the observed flexibility of a developing organism to physiologically adapt to the early life environment [19]. Epigenetic mechanisms have the potential to provide both the variability and adaptability required to change the developmental programme in response to environmental cues, and then maintain these changes throughout the life course. Epigenetics is thought to be the key mechanism underpinning the developmental origins of NCDs [73].

‘Epigenetic modifications are stably inherited through cell division without alteration of the DNA sequence’, and provide a large degree of control over a gene’s transcriptional state. The main components of the epigenetic regulatory apparatus, DNA methylation, histone modifications and non-coding RNAs work together to control access to the underlying DNA sequence, determining gene transcription and ultimately defining the role of each cell within the body; DNA methylation at regulatory regions, often upstream of a gene’s transcriptional start site, is generally associated with gene repression. The presence of DNA methylation causes recruitment of repressive protein complexes that prevent gene transcription [76]. DNA methylation alters during early development; gametes exhibit high levels of methylation, but shortly after fertilization, global methylation levels decrease, reaching their lowest levels around the time of blastocyst implantation. De novo methylation then occurs within the inner cell mass giving rise to cell lineage-specific methylation patterns that are maintained in differentiated cells [77, 78].

Epigenetics and the Early Life Environment

‘There is an increasing body of evidence suggesting that the early life environment can alter the epigenome’, and that once

these changes have occurred in early life, they are then maintained during the life course. Some of the first clear examples of maternal diet altering DNA methylation in the offspring came from studies of Agouti viable yellow (Avy) mice. Here, ‘supplementation of the maternal diet with dietary methyl donors and cofactors (folic acid, vitamin B12, ‘choline and betaine) shifted the coat colour of the offspring from yellow (agouti) to brown (pseudo-agouti)’ due to increased methylation of the agouti gene [79–82].

Subsequent animal studies have shown that alterations in maternal macronutrient intake can alter the epigenetic regulation of key metabolic genes. For example, a maternal protein-restricted diet-induced altered DNA methylation and expression of genes involved in fat metabolism (PPAR α) and stress response (glucocorticoid receptor (GR)) in the liver of both juvenile [55, 56] and adult offspring [54]. Furthermore, these epigenetic changes in PPAR α and GR also led to alterations in the activity of their downstream target genes and specific physiological processes.

Altered epigenetic regulation of the renin-angiotensin system, important for kidney natriuresis and a potential cause of hypertension, has been observed in animal models. Maternal protein restriction programmed increased AGTR1 levels in the offspring; this was accompanied by a decrease in DNA methylation at key regulatory sequences within the gene promoter region, suggesting that altered epigenetic regulation was behind the increase in receptor numbers, driving the observed hypertension [60, 83]. In addition to altered receptor expression patterns that favour hypertension, maternal undernourishment also alters the epigenetic regulation of angiotensin converting enzyme (ACE1), the enzyme responsible for converting angiotensin to its active form. Increased levels of ACE1 in mice exposed to maternal protein restriction was accompanied by a reduction in DNA methylation within the gene’s promoter and upregulation of miRNAs associated with positive regulation of ACE1 [84].

Over feeding in early life has also been shown to induce hypermethylation of the proopiomelanocortin (POMC) promoter, preventing the upregulation of POMC expression that normally occurs after feeding, promoting satiety. POMC methylation perturbed the normal physiological response to high plasma levels of both leptin and insulin, contributing to the development of obesity in the offspring [85].

Human studies have also found epigenetic changes associated with perturbations in early life nutrition. In studies from the Dutch Hunger Winter, periconceptual famine exposure was associated with altered DNA methylation across the imprint control region of IGF2 in individuals whose mothers had been exposed to famine compared to their non-exposed siblings, while exposure to famine in late gestation showed no altered methylation at the same region [86]. Examination of further genes found the same pattern: periconceptual famine exposure was ‘associated with small DNA methylation

changes within multiple loci (including leptin, IL-10, MEG3 and ABCA3) while a later exposure did not alter DNA methylation [87]. The detection of these epigenetic marks associated with famine exposure in peripheral blood 60 years after famine exposure supports the findings from animal models that early life nutrition can induce epigenetic changes in the offspring that persist long after the nutritional constraint has been removed, and that these epigenetic changes may underlie the long-term changes seen in metabolism and disease risk. Consistent with these findings, Godfrey et al. have shown that the methylation level of a CpG site within the promoter of the RXRA gene in the umbilical cord predicted greater than 25 % of the variation in fat mass in children aged 9 years [88] with replication of the finding in a second group of children aged 6 years. Furthermore, methylation of PGC1A in whole blood of children aged 5–7 years was found to predict adiposity in teenagers [89]. Such findings not only support that hypothesis that developmentally induced epigenetic marks make a significant contribution to later phenotype but also suggest that the detection of epigenetic marks even in peripheral tissue may allow identification of individuals at increased risk of chronic disease in later life before the onset of clinical disease’.

Conclusions

Cardiovascular disease is increasingly viewed as not simply developing in adulthood, but rather as a group of disorders that become apparent in adulthood, but which have their origins in the early life environment [90]. Undernutrition or exposure to an obesogenic environment pre-birth has a great impact upon an individual’s future health, with nutritional exposure altering development of the cardiovascular system and kidneys, as well as triggering wide-ranging alterations to gene expression of many critical factors that regulate blood pressure and metabolism. Many of the changes brought about by nutrition in early life are thought to be mediated by epigenetic alterations that regulate how a gene is expressed, with epigenetic changes maintaining this new pattern of expression during an individual’s life course, thereby setting them on the path to future disease.

While many factors contribute to an individual’s CVD risk, it is the central role of the early life environment and the cyclical nature of disease risk whereby maternal obesity and diet impact upon the health of the next generation that in turn passes on the burden of disease, which must be addressed [5]. Pregnancy and early infancy represents a window of opportunity to promote healthier diets and increased physical activity, with dietary interventions during pregnancy shown to reduce CVD risk associated with undernourishment [7, 19, 91, 92]. Studies on maternal weight loss through bariatric surgery prior to pregnancy have found that transmission of obesity to the

next generation can be avoided or reduced [93], suggesting that the transgenerational impact of obesity can be blunted by nutritional intervention. The importance of early life influences on an individual’s future cardiovascular wellbeing is central to any health and prevention strategy that seeks to ameliorate the burgeoning rates of cardiovascular-linked mortality, and to tackle what has been described as the most important global health issue of this century [2]. Such preemptive strategies are possible, practical and will provide long-term benefits for public health.

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Compliance with Ethics Guidelines

Conflict of Interest Robert Murray and Karen Lillycrop have no conflicts of interest. Keith Godfrey reports other from Nestle Nutrition Institute, grants from Abbott Nutrition and Nestec, outside the submitted work; In addition, Dr. Godfrey has a patent phenotype prediction pending, a patent predictive use of CpG methylation pending, and a patent maternal nutrition composition pending.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

References

1. Singhal A. The global epidemic of noncommunicable disease: the role of early-life factors. *Nestle Nutr Inst Workshop Ser.* 2014;78: 123–32.
2. WHO. Noncommunicable diseases country profiles. 2014.
3. WHO. Global status report on noncommunicable diseases. 2010.
4. Ramachandran A, Snehalatha C. Rising burden of obesity in Asia. *J Obes.* 2010; 2010.
5. Hanson MA. Developmental origins of obesity and non-communicable disease. *Endocrinol Nutr Org Soc Esp Endocrinol Nutr.* 2013;60 Suppl 1:10–1.
6. Kelishadi R, Poursafa P. A review on the genetic, environmental, and lifestyle aspects of the early-life origins of cardiovascular disease. *Curr Probl Pediatr Adolesc Health Care.* 2014;44(3):54–72.
7. Fall CH. Fetal programming and the risk of noncommunicable disease. *Indian J Pediatr.* 2013;80 Suppl 1:S13–20.
8. Blackmore HL, Ozanne SE. Maternal diet-induced obesity and offspring cardiovascular health. *J Dev Origins Health Dis.* 2013;4(5): 338–47.
9. Cote AT, Harris KC, Panagiotopoulos C, Sandor GG, Devlin AM. Childhood obesity and cardiovascular dysfunction. *J Am Coll Cardiol.* 2013;62(15):1309–19.
10. Kannel WB, Garrison RJ, Dannenberg AL. Secular blood pressure trends in normotensive persons: the Framingham Study. *Am Heart J.* 1993;125(4):1154–8.
11. Flegal KM, Carroll MD, Kit BK, Ogden CL. Prevalence of obesity and trends in the distribution of body mass index among US adults, 1999–2010. *JAMA.* 2012;307(5):491–7.

12. Heslehurst N, Rankin J, Wilkinson JR, Summerbell CD. A nationally representative study of maternal obesity in England, UK: trends in incidence and demographic inequalities in 619 323 births, 1989–2007. *Int J Obes*. 2010;34(3):420–8.
13. Dhuper S, Abdullah RA, Weichbrod L, Mahdi E, Cohen HW. Association of obesity and hypertension with left ventricular geometry and function in children and adolescents. *Obesity*. 2011;19(1):128–33.
14. Black D, Bryant J, Peebles C, Davies L, Inskip H, Godfrey K, et al. Increased regional deformation of the left ventricle in normal children with increased body mass index: implications for future cardiovascular health. *Pediatr Cardiol*. 2014;35(2):315–22. *This study examined the link between BMI and CVD risk factors in children, finding that a higher BMI associated with an increases in: systolic blood pressure, heart rate, cardiac output, and left ventricular diastolic mass/strain rate.*
15. Mahfouz RA, Dewedar A, Abdelmoneim A, Hossien EM. Aortic and pulmonary artery stiffness and cardiac function in children at risk for obesity. *Echocardiography*. 2012;29(8):984–90.
16. Di Salvo G, Pacileo G, Del Giudice EM, Natale F, Limongelli G, Verrengia M, et al. Abnormal myocardial deformation properties in obese, non-hypertensive children: an ambulatory blood pressure monitoring, standard echocardiographic, and strain rate imaging study. *Eur Heart J*. 2006;27(22):2689–95.
17. Ozdemir O, Hizli S, Abaci A, Agladioglu K, Aksoy S. Echocardiographic measurement of epicardial adipose tissue in obese children. *Pediatr Cardiol*. 2010;31(6):853–60.
18. Godfrey KM, Inskip HM, Hanson MA. The long-term effects of prenatal development on growth and metabolism. *Semin Reprod Med*. 2011;29(3):257–65.
19. Langley-Evans SC. Nutrition in early life and the programming of adult disease: a review. *J Hum Nutr Diet Off J Br Diet Assoc*. 2014.
20. Forsdahl A. Are poor living conditions in childhood and adolescence an important risk factor for arteriosclerotic heart disease? *Br J Prev Soc Med*. 1977;31(2):91–5.
21. Barker DJ, Osmond C. Low birth weight and hypertension. *BMJ*. 1988;297(6641):134–5.
22. Barker DJ, Winter PD, Osmond C, Margetts B, Simmonds SJ. Weight in infancy and death from ischaemic heart disease. *Lancet*. 1989;2(8663):577–80.
23. Hales CN, Barker DJ, Clark PM, Cox LJ, Fall C, Osmond C, et al. Fetal and infant growth and impaired glucose tolerance at age 64. *BMJ*. 1991;303(6809):1019–22.
24. Barker DJ, Hales CN, Fall CH, Osmond C, Phipps K, Clark PM. Type 2 (non-insulin-dependent) diabetes mellitus, hypertension and hyperlipidaemia (syndrome X): relation to reduced fetal growth. *Diabetologia*. 1993;36(1):62–7.
25. Jaddoe VW, de Jonge LL, Hofman A, Franco OH, Steegers EA, Gaillard R. First trimester fetal growth restriction and cardiovascular risk factors in school age children: population based cohort study. *BMJ*. 2014;348:g14. *This article describes a recent human study on 1184 children that provide strong evidence for the link between fetal growth in early pregnancy and the development of cardiovascular risk factors.*
26. Forsen T, Eriksson JG, Tuomilehto J, Osmond C, Barker DJ. Growth in utero and during childhood among women who develop coronary heart disease: longitudinal study. *BMJ*. 1999;319(7222):1403–7.
27. Phillips DI, Barker DJ, Fall CH, Seckl JR, Whorwood CB, Wood PJ, et al. Elevated plasma cortisol concentrations: a link between low birth weight and the insulin resistance syndrome? *J Clin Endocrinol Metabol*. 1998;83(3):757–60.
28. Skilton MR, Evans N, Griffiths KA, Harmer JA, Celermajer DS. Aortic wall thickness in newborns with intrauterine growth restriction. *Lancet*. 2005;365(9469):1484–6.
29. Eriksson JG, Forsen T, Tuomilehto J, Osmond C, Barker DJ. Early growth and coronary heart disease in later life: longitudinal study. *BMJ*. 2001;322(7292):949–53.
30. Eriksson JG, Forsen T, Tuomilehto J, Winter PD, Osmond C, Barker DJ. Catch-up growth in childhood and death from coronary heart disease: longitudinal study. *BMJ*. 1999;318(7181):427–31.
31. Roseboom T, de Rooij S, Painter R. The Dutch famine and its long-term consequences for adult health. *Early Hum Dev*. 2006;82(8):485–91.
32. Painter RC, Roseboom TJ, Bleker OP. Prenatal exposure to the Dutch famine and disease in later life: an overview. *Reprod Toxicol*. 2005;20(3):345–52.
33. Dearden L, Ozanne SE. The road between early growth and obesity: new twists and turns. *Am J Clin Nutr*. 2014;100(1):6–7.
34. Barker DJ, Eriksson JG, Forsen T, Osmond C. Fetal origins of adult disease: strength of effects and biological basis. *Int J Epidemiol*. 2002;31(6):1235–9.
35. Pettitt DJ, Jovanovic L. Birth weight as a predictor of type 2 diabetes mellitus: the U-shaped curve. *Curr Diab Rep*. 2001;1(1):78–81.
36. Ong KK. Size at birth, postnatal growth and risk of obesity. *Horm Res*. 2006;65 Suppl 3:65–9.
37. Evagelidou EN, Giapros VI, Challa AS, Cholevas VK, Vartholomatos GA, Siomou EC, et al. Prothrombotic state, cardiovascular, and metabolic syndrome risk factors in prepubertal children born large for gestational age. *Diabetes Care*. 2010;33(11):2468–70.
38. Poston L. Maternal obesity, gestational weight gain and diet as determinants of offspring long term health. *Best Pract Res Clin Endocrinol Metab*. 2012;26(5):627–39.
39. Curhan GC, Willett WC, Rimm EB, Spiegelman D, Ascherio AL, Stampfer MJ. Birth weight and adult hypertension, diabetes mellitus, and obesity in US men. *Circulation*. 1996;94(12):3246–50.
40. Reynolds RM, Osmond C, Phillips DI, Godfrey KM. Maternal BMI, parity, and pregnancy weight gain: influences on offspring adiposity in young adulthood. *J Clin Endocrinol Metabol*. 2010;95(12):5365–9.
41. Oken E, Taveras EM, Kleinman KP, Rich-Edwards JW, Gillman MW. Gestational weight gain and child adiposity at age 3 years. *Am J Obstet Gynecol*. 2007;196(4):322–8.
42. Cooper R, Pinto Pereira SM, Power C, Hypponen E. Parental obesity and risk factors for cardiovascular disease among their offspring in mid-life: findings from the 1958 British Birth Cohort Study. *Int J Obes*. 2013;37(12):1590–6. *This article describes the intergenerational influence of parental obesity, using data from 9328 individuals a strong association was observed between parental obesity and offspring CVD risk, even after adjusting for lifestyle and socioeconomic factors.*
43. Kalk P, Guthmann F, Krause K, Relle K, Godes M, Gossing G, et al. Impact of maternal body mass index on neonatal outcome. *Eur J Med Res*. 2009;14(5):216–22.
44. Reynolds RM, Allan KM, Raja EA, Bhattacharya S, McNeill G, Hannaford PC, et al. Maternal obesity during pregnancy and premature mortality from cardiovascular event in adult offspring: follow-up of 1 323 275 person years. *BMJ*. 2013;347:f4539. *This study used birth records of 37,709 people to show a strong association between maternal obesity and subsequent offspring CVD-linked mortality in later life.*
45. Murrin CM, Kelly GE, Tremblay RE, Kelleher CC. Body mass index and height over three generations: evidence from the Lifeways cross-generational cohort study. *BMC Public Health*. 2012;12:81.
46. Bertram CE, Hanson MA. Animal models and programming of the metabolic syndrome. *Br Med Bull*. 2001;60(1):103–21.
47. Burns SP, Desai M, Cohen RD, Hales CN, Iles RA, Germain JP, et al. Gluconeogenesis, glucose handling, and structural changes in livers of the adult offspring of rats partially deprived of protein during pregnancy and lactation. *J Clin Invest*. 1997;100(7):1768–74.

48. Bellinger L, Lilley C, Langley-Evans SC. Prenatal exposure to a maternal low-protein diet programmes a preference for high-fat foods in the young adult rat. *Br J Nutr.* 2004;92(3):513–20.
49. Bellinger L, Sculley DV, Langley-Evans SC. Exposure to undernutrition in fetal life determines fat distribution, locomotor activity and food intake in ageing rats. *Int J Obes.* 2006;30(5):729–38.
50. Torrens C, Poston L, Hanson MA. Transmission of raised blood pressure and endothelial dysfunction to the F2 generation induced by maternal protein restriction in the F0, in the absence of dietary challenge in the F1 generation. *Br J Nutr.* 2008;100(4):760–6.
51. Langley-Evans SC, Sherman RC, Welham SJ, Nwagwu MO, Gardner DS, Jackson AA. Intrauterine programming of hypertension: the role of the renin-angiotensin system. *Biochem Soc Trans.* 1999;27(2):88–93.
52. Van Abeelen AF, Veenendaal MV, Painter RC, De Rooij SR, Thangaratinam S, Van Der Post JA, et al. The fetal origins of hypertension: a systematic review and meta-analysis of the evidence from animal experiments of maternal undernutrition. *J Hypertens.* 2012;30(12):2255–67.
53. McMullen S, Gardner DS, Langley-Evans SC. Prenatal programming of angiotensin II type 2 receptor expression in the rat. *Br J Nutr.* 2004;91(1):133–40.
54. Burdge GC, Slater-Jefferies J, Torrens C, Phillips ES, Hanson MA, Lillycrop KA. Dietary protein restriction of pregnant rats in the F0 generation induces altered methylation of hepatic gene promoters in the adult male offspring in the F1 and F2 generations. *Br J Nutr.* 2007;97(3):435–9.
55. Lillycrop KA, Slater-Jefferies JL, Hanson MA, Godfrey KM, Jackson AA, Burdge GC. Induction of altered epigenetic regulation of the hepatic glucocorticoid receptor in the offspring of rats fed a protein-restricted diet during pregnancy suggests that reduced DNA methyltransferase-1 expression is involved in impaired DNA methylation and changes in histone modifications. *Br J Nutr.* 2007;97(6):1064–73.
56. Lillycrop KA, Phillips ES, Torrens C, Hanson MA, Jackson AA, Burdge GC. Feeding pregnant rats a protein-restricted diet persistently alters the methylation of specific cytosines in the hepatic PPAR α promoter of the offspring. *Br J Nutr.* 2008;100(2):278–82.
57. Langley SC, Jackson AA. Increased systolic blood pressure in adult rats induced by fetal exposure to maternal low protein diets. *Clin Sci (Lond).* 1994;86(2):217–22.
58. Tarry-Adkins JL, Martin-Gronert MS, Fernandez-Twinn DS, Hargreaves I, Alfaradhi MZ, Land JM, et al. Poor maternal nutrition followed by accelerated postnatal growth leads to alterations in DNA damage and repair, oxidative and nitrosative stress, and oxidative defense capacity in rat heart. *FASEB J Off Publ Fed Am Soc Exp Biol.* 2013;27(1):379–90.
59. Nascimento L, Freitas CM, Silva-Filho R, Leite AC, Silva AB, da Silva AI, et al. The effect of maternal low-protein diet on the heart of adult offspring: role of mitochondria and oxidative stress. *Appl Physiol Nutr Metabol Physiol Appl Nutr Metabol.* 2014;39(8):880–7.
60. Bogdarina I, Welham S, King PJ, Burns SP, Clark AJ. Epigenetic modification of the renin-angiotensin system in the fetal programming of hypertension. *Circ Res.* 2007;100(4):520–6.
61. Tsukuda K, Mogi M, Iwanami J, Min LJ, Jing F, Ohshima K, et al. Influence of angiotensin II type 1 receptor-associated protein on prenatal development and adult hypertension after maternal dietary protein restriction during pregnancy. *J Am Soc Hypertens JASH.* 2012;6(5):324–30.
62. Briet M, Schiffrin EL. Aldosterone: effects on the kidney and cardiovascular system. *Nat Rev Nephrol.* 2010;6(5):261–73.
63. Gluckman PD, Lillycrop KA, Vickers MH, Pleasants AB, Phillips ES, Beedle AS, et al. Metabolic plasticity during mammalian development is directionally dependent on early nutritional status. *Proc Natl Acad Sci U S A.* 2007;104(31):12796–800.
64. Woodall SM, Johnston BM, Breier BH, Gluckman PD. Chronic maternal undernutrition in the rat leads to delayed postnatal growth and elevated blood pressure of offspring. *Pediatr Res.* 1996;40(3):438–43.
65. Samuelsson AM, Matthews PA, Argenton M, Christie MR, McConnell JM, Jansen EH, et al. Diet-induced obesity in female mice leads to offspring hyperphagia, adiposity, hypertension, and insulin resistance: a novel murine model of developmental programming. *Hypertension.* 2008;51(2):383–92.
66. Blackmore HL, Niu Y, Fernandez-Twinn DS, Tarry-Adkins JL, Giussani DA, Ozanne SE. Maternal diet-induced obesity programs cardiovascular dysfunction in adult male mouse offspring independent of current body weight. *Endocrinology.* 2014;155(10):3970–80. *This article reports on the findings of a mouse model of maternal obesity. Offspring of obese mothers developed cardiovascular dysfunction despite being fed a healthy diet and having a similar pattern of postnatal growth to controls.*
67. Guberman C, Jellyman JK, Han G, Ross MG, Desai M. Maternal high-fat diet programs rat offspring hypertension and activates the adipose renin-angiotensin system. *Am J Obstet Gynecol.* 2013;209(3):8. *This study reported on the results of a rat model of maternal obesity and its effects upon the RAS system in offspring adipose tissue. They found that exposure to maternal obesity increased expression of both angiotensin and ACE in adipose tissue, driving further adipogenesis while also causing hypertension.*
68. Hanson M, Godfrey KM, Lillycrop KA, Burdge GC, Gluckman PD. Developmental plasticity and developmental origins of non-communicable disease: theoretical considerations and epigenetic mechanisms. *Prog Biophys Mol Biol.* 2011;106(1):272–80.
69. Bateson P, Barker D, Clutton-Brock T, Deb D, D'Udine B, Foley RA, et al. Developmental plasticity and human health. *Nature.* 2004;430(6998):419–21.
70. Norris SA, Osmond C, Gigante D, Kuzawa CW, Ramakrishnan L, Lee NR, et al. Size at birth, weight gain in infancy and childhood, and adult diabetes risk in five low- or middle-income country birth cohorts. *Diabetes Care.* 2012;35(1):72–9.
71. Bavdekar A, Yajnik CS, Fall CH, Bapat S, Pandit AN, Deshpande V, et al. Insulin resistance syndrome in 8-year-old Indian children: small at birth, big at 8 years, or both? *Diabetes.* 1999;48(12):2422–9.
72. Huffman MD, Prabhakaran D, Osmond C, Fall CH, Tandon N, Lakshmy R, et al. Incidence of cardiovascular risk factors in an Indian urban cohort results from the New Delhi birth cohort. *J Am Coll Cardiol.* 2011;57(17):1765–74.
73. Gluckman PD, Hanson MA. Developmental and epigenetic pathways to obesity: an evolutionary-developmental perspective. *Int J Obes (London).* 2008;32 Suppl 7:S62–71.
74. Peterson RE, Maes HH, Lin P, Kramer JR, Hesselbrock VM, Bauer LO, et al. On the association of common and rare genetic variation influencing body mass index: a combined SNP and CNV analysis. *BMC Genomics.* 2014;15:368. *This article reports on a large scale SNP study that used a whole genome genotyping approach to examine the genetic basis for variation in BMI and concludes that genetic variation only accounts for 3.2% of observed variance, with this increasing to 11.5% when copy number variation was added into the model.*
75. Speliotes EK, Willer CJ, Berndt SI, Monda KL, Thorleifsson G, Jackson AU, et al. Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat Genet.* 2010;42(11):937–48.
76. Miranda TB, Jones PA. DNA methylation: the nuts and bolts of repression. *J Cell Physiol.* 2007;213(2):384–90.
77. Fulka H, St John JC, Fulka J, Hozak P. Chromatin in early mammalian embryos: achieving the pluripotent state. *Differentiation.* 2008;76(1):3–14.

78. Li E. Chromatin modification and epigenetic reprogramming in mammalian development. *Nat Rev Genet.* 2002;3(9):662–73.
79. Morgan HD, Sutherland HG, Martin DI, Whitelaw E. Epigenetic inheritance at the agouti locus in the mouse. *Nat Genet.* 1999;23(3):314–8.
80. Rakyán VK, Blewitt ME, Druker R, Preis JI, Whitelaw E. Metastable epialleles in mammals. *Trends Genet TIG.* 2002;18(7):348–51.
81. Waterland RA, Travisano M, Tahiliani KG. Diet-induced hypermethylation at agouti viable yellow is not inherited transgenerationally through the female. *FASEB J Off Publ Fed Am Soc Exp Biol.* 2007;21(12):3380–5.
82. Waterland RA, Jirtle RL. Transposable elements: targets for early nutritional effects on epigenetic gene regulation. *Mol Cell Biol.* 2003;23(15):5293–300.
83. Bogdarina I, Haase A, Langley-Evans S, Clark AJ. Glucocorticoid effects on the programming of AT1b angiotensin receptor gene methylation and expression in the rat. *PLoS One.* 2010;5(2):e9237.
84. Goyal R, Goyal D, Leitzke A, Gheorghe CP, Longo LD. Brain renin-angiotensin system: fetal epigenetic programming by maternal protein restriction during pregnancy. *Reprod Sci.* 2010;17(3):227–38.
85. Plagemann A, Harder T, Brunn M, Harder A, Roepke K, Wittrock-Staar M, et al. Hypothalamic proopiomelanocortin promoter methylation becomes altered by early overfeeding: an epigenetic model of obesity and the metabolic syndrome. *J Physiol.* 2009;587(Pt 20):4963–76.
86. Heijmans BT, Tobi EW, Stein AD, Putter H, Blauw GJ, Susser ES, et al. Persistent epigenetic differences associated with prenatal exposure to famine in humans. *Proc Natl Acad Sci U S A.* 2008;105(44):17046–9.
87. Tobi EW, Lumey LH, Talens RP, Kremer D, Putter H, Stein AD, et al. DNA methylation differences after exposure to prenatal famine are common and timing- and sex-specific. *Hum Mol Genet.* 2009;18(21):4046–53.
88. Godfrey KM, Sheppard A, Gluckman PD, Lillycrop KA, Burdge GC, McLean C, et al. Epigenetic gene promoter methylation at birth is associated with child's later adiposity. *Diabetes.* 2011;60(5):1528–34.
89. Clarke-Harris R, Wilkin TJ, Hosking J, Pinkney J, Jeffery AN, Metcalf BS, et al. PGC1alpha promoter methylation in blood at 5–7 years predicts adiposity from 9 to 14 years (EarlyBird 50). *Diabetes.* 2014;63(7):2528–37. *This study identified DNA methylation changes within the promoter of PGC1a in children that were predictive of later adiposity and showed the functional relevance of methylation at these specific CpG sites as binding sites for the PBX-1 or HOXB9 transcription factors when methylated.*
90. West NA, Crume TL, Maligie MA, Dabelea D. Cardiovascular risk factors in children exposed to maternal diabetes in utero. *Diabetologia.* 2011;54(3):504–7.
91. Bhutta ZA, Ahmed T, Black RE, Cousens S, Dewey K, Giugliani E, et al. What works? Interventions for maternal and child undernutrition and survival. *Lancet.* 2008;371(9610):417–40.
92. Fall C. Maternal nutrition: effects on health in the next generation. *Indian J Med Res.* 2009;130(5):593–9.
93. Kral JG, Biron S, Simard S, Hould FS, Lebel S, Marceau S, et al. Large maternal weight loss from obesity surgery prevents transmission of obesity to children who were followed for 2 to 18 years. *Pediatrics.* 2006;118(6):e1644–9.