



Changes in Volatile Compounds of Virgin Olive Oil Flavored with Essential Oils During Thermal and Photo-Oxidation

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Received: 26 March 2020 / Accepted: 27 November 2020 / Published online: 6 January 2021

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Abstract

The effects of four essential oils from peppermint (*Micromeria fruticosa*), oregano (*Origanum onites*), thyme (*Thymus vulgaris*), and laurel (*Laurus nobilis*) on the volatile compounds of olive oil were determined. The concentration of essential oils was 0.05% (v/w) and flavored olive oil samples were stored for 45 days at 60 °C and room temperature for thermal oxidation and photo-oxidation under fluorescent light, respectively. Control and flavored olive oils were analyzed after 15, 30, and 45th days to track the changes in their volatile compound contents using HS-SPME/GC-MS technique. Higher concentrations of diversified volatile components were detected under thermal oxidation conditions rather than photo-oxidation. According to thermal oxidation results, the E-2-hexenal values of control and flavored oils with peppermint essential oil were higher at the end of 30 days storage, while flavoring with essential oil of *Thymus vulgaris* resulted in the highest E-2-hexenal value for photo-oxidation. Results indicated that the main components of essential oil transferred into olive oil samples. Carvacrol was present in flavored oils with oregano and thyme. Eucalyptol and pulegone were determined as major components in flavored oils with laurel and peppermint essential oils, respectively. In both oxidation methods, these volatile components remained stable and little or no loss was observed.

Keywords Flavored olive oil · Essential oil · Volatile compounds · Thermal oxidation · Photo-oxidation · GC-MS

Introduction

Extra virgin olive oil (EVOO) plays an important role in the Mediterranean diet. EVOO contains high amounts of fatty acids in addition to such minor components as phenolics, sterols, tocopherols, squalene, and volatile compounds. The positive impacts of minor compounds present in EVOO to human health have been evidenced by several clinical works (Francisco et al. 2019; Romani et al. 2019). Besides health benefits, EVOO has a unique aroma that originates from the

mixture of volatile and non-volatile components. The quality of EVOO is mainly affected by its overall composition having such groups as fatty acids, sterols, tocopherols, and other minor compounds. Besides, volatile components also play an important role in the quality of EVOO in terms of the preferences of consumers (Genovese et al. 2019). The characteristic sensory attributes for EVOO are fruity and green, and these attributes are strongly related to C5 and C6 compounds which are formed by the enzymatic oxidation of olive oils, especially through the lipoxygenase pathway during olive oil processing (Angerosa et al. 2001; Kalua et al. 2007). The presence or absence of particular volatile compounds contributes to the overall EVOO aroma.

Consumers prefer functional foods that are beneficial to them. Consumption of these foods ensures some health benefits for some diseases (Sloan 2000). In recent years, new functional products based on olive oil flavored with herbs or spices have been placed on market shelves. Today, different types of flavored oils are available on the market. Vegetables, herbs, spices, mushrooms, fruits, nuts, and some aroma compounds have been used in flavored olive oil production (Sousa et al. 2015). Besides, most of these aromatizers are reported to increase the oxidative stability of oils (Gambacorta et al. 2007).

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The traditional infusion method is one of the several methods used in the production of flavored olive oil. The main disadvantage of this method is the long infusion times (up to months) at room conditions to successfully produce flavored olive oil. Besides, undesirable substances such as waxes can be also transferred from the aromatic plant to olive oil which may change the sensorial and stability attributes of olive oils during shelf life (Baiano et al. 2010). On the other hand, aromatic plants are mostly seasonal and not available throughout the year (Akçar and Gümüskesen 2011). To avoid these problems, olive oil can be flavored by the use of the essential oils (Moldão-Martins et al. 2004) as an alternative approach. Although there are many studies using spices and herbs in aromatized oil production, the number of studies regarding the use of essential oils in flavoring olive oils is limited. Ayadi et al. (2009) studied physicochemical changes in flavored oils with rosemary, lavender, sage, menthe, basil, lemon, and thyme at thermal conditions (60 and 130 °C). Damechki et al. (2001) investigated the changes in the physicochemical characteristics and oxidative stability of olive oils flavored with oregano and rosemary. Moldão-Martins et al. (2004) studied response surface methodology (RSM) to determine the suitable ratio of *Mentha piperita* L. and *Thymus mastichina* subsp. *mastichina* essential oils in olive oil depending on sensory analysis. A study by Arcoleo et al. (2009) evaluated the effects of cold-pressed lemon essential oil on sensory properties of olive oil as well as the shelf life. Asensio et al. (2013) studied the effects of oregano essential oils on the oxidative stability of olive oils stored in dark and light conditions. In another study by Taoudiat et al. (2018), the effects of *Laurus nobilis* essential oil on the oxidative stability of olive oils packaged with brown and transparent glass or PET were investigated during photo-oxidation conditions.

This study aimed to determine the effects of such essential oils as peppermint (*Micromeria fruticosa*), oregano (*Origanum onites*), thyme (*Thymus vulgaris*), and laurel (*Laurus nobilis*) on the volatile components of olive oil during thermal and photo-oxidation.

Materials and Methods

Essential Oils and Preparation of Flavored Olive Oils

The four essential oils, peppermint (*Micromeria fruticosa*), oregano (*Origanum onites*), thyme (*Thymus vulgaris*), and laurel (*Laurus nobilis*) were purchased from the local markets in Turkey. The essential oils were kept at -18 ± 0.5 °C in glass vials in a dark place. Flavored oils were prepared by adding essential oils (0.025 mL) to 50 g of olive oil samples. These oils were then transferred to transparent and dark glass bottles (100 cc) for thermal storage conditions and light-induced storage, respectively. Flavored oil samples are coded

as follows: MF: flavored oil with peppermint essential oil, OO: flavored oil with oregano essential oil, TV: flavored oil with thyme essential oil, and LN: flavored oil with laurel essential oil.

Gas Chromatography/Mass Spectrometry Analysis of Essential Oils

Essential oil compounds were characterized by a Shimadzu GC-2010 Plus model (Shimadzu Corporation, Kyoto, Japan) gas chromatograph equipped with a Shimadzu QP2010 Plus model mass spectrometer using electron impact. An electron ionization system with ionization energy of 70 eV was used within a scan range of 40–300 amu for GC-MS analysis. Separation of the compounds was achieved by using a Rxi 5Sil-MS-GC column (30 m × 0.25 mm ID × 0.25 μm) (Restek Corporation, Bellefonte, USA). Helium was used as the carrier gas with a flow rate of 1.61 mL/min. The temperatures of the ion source and the MS transfer line were 200 and 250 °C, respectively. The oven temperature program was initially set at 40 °C for 2 min, then raised to 250 °C at a rate of 4 °C/min, and finally held isothermal for 5 min. Then, 0.20 μL of each sample was injected into the equipment using split mode with a split ratio of 10:1. The compound identification was performed by the comparison of mass spectra and linear retention indexes (LRI) of the compounds with authentic standards and published data. Also, their mass spectra were also compared by such MS libraries as Wiley, NIST and FFNSC.

Analysis of Volatile Compounds in EVOO Samples

The volatile fractions of EVOO samples were analyzed by headspace solid-phase microextraction/gas chromatography–mass spectrometer (HS–SPME/GC–MS). Two grams of sample was weighed in 20 mL screw cap vials and placed in a vial heater at 45 °C for 15 min of equilibration time. Volatile compounds were adsorbed onto an SPME fiber Carboxen/polydimethylsiloxane (CAR/PDMS; 75 μm) (Supelco, Bellefonte, USA). The sampling time was 45 min at 45 °C. The desorption of volatile compounds was completed by injecting to the GC operated under the same conditions as described above. GC/MS analyses of the volatiles were carried out by a Shimadzu GC-2010 Plus (Shimadzu Corporation, Kyoto, Japan) model gas chromatograph with a Rxi 5Sil-MS-GC column (30 m, 0.25 mm ID, 0.25 μm) (Restek, Bellefonte, USA) equipped with a Shimadzu QP2010 Plus (Shimadzu Corporation, Kyoto, Japan) mass spectrometer using electron impact (ionization energy: 70 eV). Retention indices of each volatile compound were calculated using a homologous series of C7–C30 *n*-alkanes injected under identical conditions with the samples. The components were identified regarding their LRI relative to

n-alkanes and matching data in such MS libraries as Wiley, NIST, FFNSC. Also, the fragmentation pattern of the mass spectra were compared with data published in the literature. The relative amounts of detected volatile compounds were expressed in arbitrary units (AU).

Oxidation Methods

Thermal Oxidation

The flavored olive oil and control samples were heated to 60 °C in the oven. The volatile compounds which were released from oil samples were analyzed at the end of every 15 days up to the end of the 45th day of storage under accelerated thermal oxidation conditions.

Photo-Oxidation

The oil samples were placed in a lightbox (Test T742 LBD, Turkey) equipped with cool white fluorescent light sources. The fluorescent radiation intensity was about 3000 lx. Volatile compounds in the oil samples were analyzed at the end of every 15 days and the whole storage period for photo-oxidation was 45 days.

Statistical Analysis

All samples were analyzed in duplicates. Data were expressed in terms of mean \pm standard deviation. Significant differences were determined using ANOVA in combination with a Duncan test with a significance level of 95% by using Minitab (Version 16, Minitab Inc., USA) statistical software.

Results and Discussion

Essential Oil Composition

The compositions of essential oils from peppermint (*Micromeria fruticosa*), oregano (*Origanum onites*), thyme (*Thymus vulgaris*), and laurel (*Laurus nobilis*) analyzed by GC-MS were exhibited in the [supplementary material](#). The major compound detected in oregano and thyme oil was carvacrol (69.95 and 57.28% for oregano and thyme oils, respectively). In a previous study (Iten et al. 2009), carvacrol was also reported as the major volatile constituent of essential oil obtained from *Thymus vulgaris*. It was determined that laurel essential oil contained two major compounds namely camphor (43.03%) and eucalyptol (1,8-cineol) (35.06%), followed by sabinene (4.21%), α -terpinyl acetate (4.20%), α -pinene (2.92%), and β -pinene (2.36%). Although the high eucalyptol content of *Laurus nobilis* essential oil agreed with data from the literature (Simić et al. 2004), some studies are reporting the

low camphor content in *Laurus nobilis* (Jemâa et al. 2012; Shokoohinia et al. 2014) in contrast to our study. Results indicated that the most abundant compounds of peppermint (*Micromeria fruticosa*) oil were pulegone (65.30%) and p-menthone (14.49%). Pulegone was also reported as the dominant compound of *M. fruticosa* in previous studies (Güllüce et al. 2004; Arslan 2012).

Thermal Oxidation

The identification data of volatile compounds in olive oil samples during thermal treatment at 60 °C are shown in Table 1. During thermal oxidation of olive oil samples, 24 components were identified in the control sample, while flavored olive oils contained numerous volatile compounds. The number of identified compounds was 38 in LN and TV samples, followed by OO with 37 compounds and MF with a minimum number of identified components of 27.

Changes in the volatile compounds of olive oil samples are presented in Table 2. During the thermal oxidation process, the initial volatiles such as hexanal, E-2-hexenal, Z-2-penten-1-ol, Z-3-hexen-1-ol, E-2-hexen-1-ol, octen-3-ol, and Z-3-hexenyl acetate, which contribute to the aroma of olive oil, disappeared first and then appeared as volatile oxidation products as their amounts increased with the ongoing oxidation. In analyzed oils, E-2-hexenal and hexanal were the main volatile compounds determined during oxidation of olive oils as compared with the previously reported data for vegetable oils treated with heat (Kiralan and Ramadan 2016; Wang et al. 2019). The highest E-2-hexenal values in C and MF samples after 30 days storage were 140.45×10^6 AU and 187.10×10^6 AU, respectively, while E-2-hexenal content of LN reached a maximum value of 33.81×10^6 AU at the end of 15 days storage. After reaching the maximum level, E-2-hexenal content started to decrease in the samples mentioned above. However, there was a slight increase in OO and TV samples during thermal oxidation conditions. Although not as much as E-2-hexenal, hexanal also showed an increasing trend during storage. Among volatile aldehydes, the presence of other saturated compounds, such as pentanal, heptanal, octanal, and nonanal, were also detected. Pentanal appeared only in C, OO, and MF samples after 15 days of storage. There was an increase in heptanal and nonanal contents of all analyzed samples, while a similar trend was observed for octanal in samples except the TV sample. 3-Methyl-2-butenal, which is a branched aldehyde, was present in all samples through the oxidation process.

Alcohols such as pentanol, hexanol, and octanol were also detected in the analyzed samples. Hexanol, an aroma compound of olive oil, was detected in fresh samples and subsequently disappeared with ongoing oxidation. Pentanol and octanol were absent only in MF and LN samples, respectively.

Table 1 Identified volatile compounds of olive oil samples at thermal and photo-oxidation experiments

KI ^a _{cal}	KI ^b _{lit}	Method of identification ^c	Library match similarity (%)	Compound ^b	Thermal oxidation					Photo-oxidation				
					C	OO	MF	LN	TV	C	OO	MF	LN	TV
624	656 ¹	MS, KI	96	2-Butenal	+	+	+	+	+	+	+	+	+	+
696	696 ¹	MS, KI	95	Pentanal	+	+	+	-	-	+	+	+	+	+
751	751 ²	MS, KI	96	E-2-Pentenal	+	+	+	+	+	+	+	+	-	+
753	761 ³	MS, KI	96	Pentanol	+	+	-	+	+	-	-	-	+	-
767	767 ²	MS, KI	93	Z-2-penten-1-ol	+	-	-	-	-	+	-	-	-	-
788	788 ⁴	MS, KI	94	3-methyl-2-butenal	+	+	+	+	+	+	+	+	+	-
801	802 ¹	MS, KI	96	Hexanal	+	+	+	+	+	+	+	+	+	+
814	806 ⁵	MS, KI	98	E-2-octene	-	-	+	+	+	-	-	+	+	-
850	850 ²	MS, KI	96	E-2-hexenal	+	+	+	+	+	+	+	+	+	+
853	853 ⁶	MS, KI	95	Z-3-Hexen-1-ol	+	+	+	+	+	+	+	+	+	+
861	861 ²	MS, KI	91	E-2-Hexen-1-ol	+	+	+	+	+	+	+	+	+	+
867	867 ⁶	MS, KI	95	Hexanol	+	+	+	+	+	+	+	+	+	+
898	888 ³	MS, KI	84	2-Heptanone	+	+	-	+	+	-	-	-	-	-
900	900 ²	MS, KI	97	Nonane	-	-	-	+	+	-	-	-	+	+
906	906 ²	MS, KI	94	Heptanal	+	-	-	-	-	+	+	+	+	+
927	928 ⁴	MS, KI	91	α-Thujene	-	+	-	+	+	-	+	-	+	-
933	933 ²	MS, KI	97	α-Pinene	-	-	-	+	+	-	-	-	+	+
956	956 ²	MS, KI	96	E-2-Heptenal	+	+	+	+	+	+	+	+	+	+
972	972 ⁷	MS, KI	97	Sabinene	-	-	-	+	-	-	-	-	+	-
974	974 ⁷	MS, KI	96	β-Pinene	-	-	-	+	-	-	-	-	+	-
978	978 ³	MS, KI	97	Octen-3-ol	+	+	+	+	+	+	-	-	+	+
986	989 ¹	MS, KI	95	6-Methyl-5-hepten-2-one	-	+	-	-	+	+	+	+	+	+
991	991 ⁷	MS, KI	92	β-Myrcene	-	+	-	+	+	-	+	+	+	+
991	991 ¹	MS, KI	97	2-pentyl- Furan	-	+	-	+	+	-	-	-	-	-
1006	1004 ¹	MS, KI	95	Octanal	+	+	+	+	-	+	-	+	+	-
1008	1008 ²	MS, KI	93	Z-3-Hexenyl acetate	+	+	+	+	+	+	+	+	+	+
1013	1014 ¹	MS, KI	95	E,E-2,4-Heptadienal	-	+	-	+	-	-	-	-	-	-
1018	1015 ⁷	MS, KI	96	α- Terpinene	-	+	-	+	+	-	-	-	-	-
1025	1023 ¹	MS, KI	97	p- Cymene	-	+	+	+	+	-	+	+	+	+
1030	1027 ¹	MS, KI	94	Limonene	-	+	+	+	+	-	+	+	+	+
1032	1030 ⁷	MS, KI	96	Eucalyptol	-	-	-	+	-	-	-	-	+	-
1038	1036 ⁸	MS, KI	92	3-Octen-2-one	+	+	+	+	+	+	-	+	+	-
1058	1061 ⁴	MS, KI	94	γ- Terpinene	-	+	-	+	+	-	+	-	+	+
1059	1057 ⁸	MS, KI	96	E-2-Octenal	+	+	+	+	+	+	-	+	+	-
1072	1072 ¹	MS, KI	89	3,5-Octadien-2-one	-	+	-	-	+	-	-	-	-	-
1076	1072 ⁷	MS, KI	96	Octanol	+	+	+	-	+	-	-	-	-	-
1093	1090 ⁹	MS, KI	91	2-Nonanone	+	+	-	-	+	-	-	-	-	-
1101	1098 ¹	MS, KI	94	Linalool	-	+	-	+	+	-	+	-	+	+
1107	1107 ²	MS, KI	95	Nonanal	+	+	+	+	+	+	+	+	+	+
1158	1164 ¹⁰	MS, KI	97	p-Menthone	-	-	+	-	-	-	-	+	-	-
1163	1163 ⁹	MS, KI	94	E-2-Nonenal	+	+	+	+	+	-	-	-	-	-
1174	1177 ¹¹	MS, KI	93	Isopulegone	-	-	+	-	-	-	-	+	-	-
1180	1184 ¹¹	MS, KI	96	Terpinen-4-ol	-	+	-	+	+	-	+	-	+	+
1198	1192 ⁴	MS, KI	96	α-Terpineol	-	-	-	+	+	-	+	-	+	+
1218	1218 ⁸	MS, KI	96	E,E-2,4-nonadienal	-	+	-	-	-	-	-	-	-	-
1241	1238 ¹²	MS, KI	97	Pulegone	-	-	+	-	-	-	-	+	-	-
1265	1262 ¹¹	MS, KI	93	E-2-decenal	+	+	+	+	+	+	+	-	+	+
1267	1255 ¹²	MS, KI	91	Piperitone	-	-	+	-	-	-	-	+	-	-
1288	1290 ¹³	MS, KI	92	Thymol	-	-	-	-	+	-	-	-	-	+
1299	1298 ¹³	MS, KI	97	Carvacrol	-	+	-	-	+	-	+	-	-	+
1322	1322 ⁷	MS, KI	95	E,E-2,4-Decadienal	-	+	+	+	+	-	-	-	-	-
1347	1341 ¹¹	MS, KI	96	Piperitenone	-	-	-	-	-	-	-	+	-	-
1349	1345 ¹⁴	MS, KI	97	α-Terpinyl acetate	-	-	-	+	-	-	-	-	+	-
1376	1364 ¹⁵	MS, KI	93	Piperitenone oxide	-	-	-	-	-	-	-	+	-	-

^a KI_{cal} Kovats index calculated for Rxi 5Sil capillary column (30 m × 0.25 mm ID × 0.25 μm) installed on a GC equipped with a mass-selective detector

^b KI_{lit} Kovats index reported in literature

^c Methods of identification: MS mass spectrum comparison using Wiley, NIST, and FFNSC libraries, KI: Kovats index in agreement with literature value (¹ Araujo et al. 2007; ² Cevik et al. 2016; ³ Wu et al. 2014a; ⁴ Petrović et al. 2017; ⁵ Mallia et al. 2008; ⁶ Karagoz et al. 2017; ⁷ Koutidou et al. 2017; ⁸ Liu et al. 2017; ⁹ Wu et al. 2014b; ¹⁰ Bishr and Salama 2018; ¹¹ Stojanović et al. 2019; ¹² Padalia et al. 2011; ¹³ Santos et al. 2015; ¹⁴ Lei et al. 2010; ¹⁵ Singh et al. 2020)

Table 2 Changes in volatile compounds of olive oil samples during thermal oxidation

Compound	Storage (days)	C	OO	MF	LN	TV
2-Butenal	0	0.14 ± 0.04b	0.00 ± 0.00d	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00b
	15	0.70 ± 0.07ab	0.58 ± 0.02c	0.62 ± 0.03b	1.66 ± 1.34	0.08 ± 0.04b
	30	0.71 ± 0.41ab	1.71 ± 0.22b	0.46 ± 0.01b	1.74 ± 0.33	0.36 ± 0.00a
	45	1.06 ± 0.16a	2.04 ± 0.08a	1.44 ± 0.18a	1.70 ± 0.00	0.43 ± 0.05a
Pentanal	0	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00c	–	–
	15	2.25 ± 0.19	1.40 ± 0.01	1.58 ± 0.02a	–	–
	30	0.00 ± 0.00	0.00 ± 0.00	0.85 ± 0.04b	–	–
	45	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00c	–	–
E-2-Pentenal	0	0.10 ± 0.01c	0.00 ± 0.00d	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00c
	15	0.84 ± 0.04b	0.82 ± 0.01c	0.63 ± 0.21b	0.73 ± 0.23a	0.12 ± 0.01b
	30	0.20 ± 0.08c	1.10 ± 0.0b	0.16 ± 0.01c	1.07 ± 0.08a	0.33 ± 0.03a
	45	0.99 ± 0.04a	1.48 ± 0.06a	1.45 ± 0.01a	0.95 ± 0.02a	0.36 ± 0.01a
Pentanol	0	0.00 ± 0.00b	0.00 ± 0.00b	–	0.25 ± 0.03	0.00 ± 0.00c
	15	0.63 ± 0.15b	0.29 ± 0.03ab	–	0.50 ± 0.32	0.00 ± 0.00c
	30	0.39 ± 0.39b	0.77 ± 0.04a	–	0.86 ± 0.30	0.13 ± 0.00b
	45	1.42 ± 0.30a	0.97 ± 0.51a	–	0.69 ± 0.09	0.42 ± 0.01a
Z-2-penten-1-ol	0	0.06 ± 0.00	–	–	–	–
	15	0.00 ± 0.00	–	–	–	–
	30	0.00 ± 0.00	–	–	–	–
	45	0.00 ± 0.00	–	–	–	–
3-Methyl-2-butenal	0	0.00 ± 0.00b	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00b
	15	0.37 ± 0.14a	0.39 ± 0.03b	0.16 ± 0.08b	0.48 ± 0.28a	0.00 ± 0.00b
	30	0.14 ± 0.08b	0.79 ± 0.04a	0.12 ± 0.01bc	0.52 ± 0.01a	0.18 ± 0.04a
	45	0.55 ± 0.01a	0.76 ± 0.06a	0.84 ± 0.05a	0.55 ± 0.01a	0.15 ± 0.01a
Hexanal	0	4.34 ± 0.08b	2.38 ± 0.12d	3.45 ± 0.05b	2.15 ± 0.09c	5.62 ± 0.47a
	15	17.15 ± 2.35a	12.27 ± 0.06a	13.96 ± 1.44a	6.26 ± 0.43b	1.38 ± 0.00c
	30	4.06 ± 1.71b	11.29 ± 0.20b	5.07 ± 0.29b	8.02 ± 0.51a	2.27 ± 0.04b
	45	11.24 ± 0.21a	8.91 ± 0.00c	14.84 ± 0.87a	6.55 ± 0.23b	2.65 ± 0.09b
E-2-octene	0	–	–	2.51 ± 0.07	1.32 ± 0.18	3.87 ± 0.54
	15	–	–	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00
	30	–	–	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00
	45	–	–	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00
E-2-hexenal	0	4.38 ± 0.03b	1.15 ± 0.04d	1.58 ± 0.06b	1.46 ± 0.12	1.04 ± 0.13a
	15	3.67 ± 0.15b	2.88 ± 0.08a	5.12 ± 0.67b	33.81 ± 0.33	1.11 ± 0.14a
	30	140.45 ± 10.77a	2.37 ± 0.08b	187.10 ± 5.87a	1.62 ± 0.45	0.70 ± 0.03b
	45	1.04 ± 0.10b	1.62 ± 0.17c	2.26 ± 0.31b	1.33 ± 0.33	0.31 ± 0.01c
Z-3-Hexen-1-ol	0	1.86 ± 0.07a	0.62 ± 0.00a	0.85 ± 0.06b	0.72 ± 0.07	0.83 ± 0.13a
	15	0.91 ± 0.04b	0.68 ± 0.06a	1.21 ± 0.03a	0.00 ± 0.00	0.31 ± 0.01b
	30	0.00 ± 0.00	0.16 ± 0.01b	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00c
	45	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00c
E-2-hexen-1-ol	0	0.29 ± 0.01	0.20 ± 0.01a	0.19 ± 0.00a	0.19 ± 0.01	0.28 ± 0.07
	15	0.00 ± 0.00	0.04 ± 0.01b	0.11 ± 0.00b	0.00 ± 0.00	0.04 ± 0.00
	30	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00
	45	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00
Hexanol	0	0.32 ± 0.02a	0.17 ± 0.01a	0.18 ± 0.04a	0.17 ± 0.00	0.39 ± 0.11
	15	0.16 ± 0.03b	0.06 ± 0.01b	0.14 ± 0.01a	0.00 ± 0.00	0.04 ± 0.00
	30	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00	0.00 ± 0.00
	45	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00	0.00 ± 0.00
2-Heptanone	0	0.00 ± 0.00b	0.00 ± 0.00c	–	0.00 ± 0.00c	0.00 ± 0.00
	15	0.00 ± 0.00b	0.00 ± 0.00c	–	0.13 ± 0.10bc	0.00 ± 0.00
	30	0.23 ± 0.23b	0.36 ± 0.04b	–	0.40 ± 0.18ab	0.00 ± 0.00
	45	1.38 ± 0.77a	0.56 ± 0.01a	–	0.54 ± 0.04a	0.27 ± 0.10
Nonane	0	–	–	–	1.35 ± 0.08	2.88 ± 0.57
	15	–	–	–	0.00 ± 0.00	0.00 ± 0.00
	30	–	–	–	0.00 ± 0.00	0.00 ± 0.00
	45	–	–	–	0.00 ± 0.00	0.00 ± 0.00
Heptanal	0	0.03 ± 0.01b	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00d
	15	0.45 ± 0.17ab	0.36 ± 0.00b	0.42 ± 0.04b	0.30 ± 0.12ab	0.05 ± 0.00c
	30	0.49 ± 0.33ab	0.76 ± 0.04a	0.22 ± 0.01bc	0.62 ± 0.28a	0.08 ± 0.01b
	45	1.06 ± 0.46a	0.79 ± 0.09a	1.40 ± 0.20a	0.43 ± 0.01ab	0.18 ± 0.01a
α-Thujene	0	–	0.09 ± 0.00b	–	0.07 ± 0.01a	0.37 ± 0.04
	15	–	0.13 ± 0.01a	–	0.06 ± 0.00a	0.00 ± 0.00
	30	–	0.00 ± 0.00c	–	0.00 ± 0.00b	0.00 ± 0.00
	45	–	0.00 ± 0.00c	–	0.00 ± 0.00b	0.00 ± 0.00
α-Pinene	0	–	–	–	0.14 ± 0.04c	0.08 ± 0.01

Table 2 (continued)

Compound	Storage (days)	C	OO	MF	LN	TV
E-2-Heptenal	15	–	–	–	0.34 ± 0.01a	0.08 ± 0.01
	30	–	–	–	0.26 ± 0.06ab	0.03 ± 0.04
	45	–	–	–	0.21 ± 0.02bc	0.02 ± 0.00
	0	0.05 ± 0.01c	0.00 ± 0.00d	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00c
	15	1.62 ± 0.38bc	2.39 ± 0.11c	0.62 ± 0.11c	4.09 ± 2.46a	0.23 ± 0.04c
Sabinene	30	2.14 ± 1.29b	7.28 ± 0.34b	1.66 ± 0.16b	6.10 ± 0.21a	1.60 ± 0.27b
	45	7.28 ± 0.16a	8.39 ± 0.23a	9.76 ± 0.54a	6.42 ± 0.35a	2.10 ± 0.03a
	0	–	–	–	0.42 ± 0.11ab	–
	15	–	–	–	0.75 ± 0.14a	–
	30	–	–	–	0.42 ± 0.25ab	–
β-Pinene	45	–	–	–	0.30 ± 0.04b	–
	0	–	–	–	0.11 ± 0.03b	–
	15	–	–	–	0.33 ± 0.06ab	–
	30	–	–	–	0.38 ± 0.10a	–
Octen-3-ol	45	–	–	–	0.35 ± 0.12ab	–
	0	0.05 ± 0.01c	0.06 ± 0.02d	0.23 ± 0.01c	0.00 ± 0.00b	0.19 ± 0.01
	15	0.00 ± 0.00bc	0.20 ± 0.03c	0.96 ± 0.07a	0.00 ± 0.00b	0.11 ± 0.01
	30	0.29 ± 0.20b	0.37 ± 0.02a	0.37 ± 0.03b	0.37 ± 0.16a	0.11 ± 0.01
	45	0.70 ± 0.02a	0.27 ± 0.03b	0.98 ± 0.05a	0.36 ± 0.12a	0.18 ± 0.00
6-Methyl-5-hepten-2-one	0	–	0.00 ± 0.00b	–	–	0.00 ± 0.00
	15	–	0.24 ± 0.05a	–	–	0.00 ± 0.00
	30	–	0.18 ± 0.01a	–	–	0.08 ± 0.02
	45	–	0.17 ± 0.01a	–	–	0.00 ± 0.00
	0	–	0.91 ± 0.02b	–	0.32 ± 0.07	0.00 ± 0.00
β-Myrcene	15	–	1.82 ± 0.18a	–	0.00 ± 0.00	1.08 ± 0.10
	30	–	0.00 ± 0.00c	–	0.00 ± 0.00	0.76 ± 0.16
	45	–	0.00 ± 0.00c	–	0.00 ± 0.00	0.00 ± 0.00
	0	–	0.00 ± 0.00b	–	0.00 ± 0.00c	0.00 ± 0.00
	15	–	0.00 ± 0.00b	–	0.00 ± 0.00c	0.00 ± 0.00
2-pentyl-furan	30	–	2.91 ± 0.07ab	–	1.93 ± 0.35b	0.00 ± 0.00
	45	–	5.31 ± 2.14a	–	3.90 ± 0.19a	1.75 ± 0.19
	0	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00	–
	15	0.43 ± 0.36	0.30 ± 0.09c	0.32 ± 0.04b	0.72 ± 0.59	–
	30	0.61 ± 0.49	0.79 ± 0.01b	0.37 ± 0.04b	0.80 ± 0.06	–
Z-3-Hexenyl acetate	45	1.88 ± 1.18	1.43 ± 0.28a	2.63 ± 0.51a	0.70 ± 0.01	–
	0	0.19 ± 0.01	0.16 ± 0.02b	0.12 ± 0.00	0.15 ± 0.02	0.00 ± 0.00
	15	0.25 ± 0.18	0.31 ± 0.10a	0.26 ± 0.00	0.48 ± 0.40	0.13 ± 0.01
	30	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00
	45	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00
E.E-2,4-Heptadienal	0	–	0.00 ± 0.00d	–	0.00 ± 0.00	–
	15	–	0.47 ± 0.03a	–	0.59 ± 0.48	–
	30	–	0.26 ± 0.04b	–	0.18 ± 0.01	–
	45	–	0.14 ± 0.01c	–	0.00 ± 0.00	–
	0	–	0.36 ± 0.06a	–	0.08 ± 0.01	0.77 ± 0.01a
α-Terpinene	15	–	0.21 ± 0.01b	–	0.00 ± 0.00	0.25 ± 0.02b
	30	–	0.00 ± 0.00c	–	0.00 ± 0.00	0.00 ± 0.00c
	45	–	0.00 ± 0.00c	–	0.00 ± 0.00	0.00 ± 0.00c
	0	–	3.59 ± 0.59b	0.56 ± 0.08a	2.25 ± 0.49a	13.86 ± 1.14a
	15	–	5.67 ± 0.11a	0.12 ± 0.01b	1.61 ± 0.51ab	3.29 ± 0.20b
p-cymene	30	–	2.73 ± 0.03bc	0.00 ± 0.00b	0.83 ± 0.59b	2.30 ± 0.04bc
	45	–	1.48 ± 0.96c	0.00 ± 0.00b	0.48 ± 0.01b	0.91 ± 0.03c
	0	–	0.98 ± 0.03a	0.54 ± 0.07a	0.94 ± 0.20a	3.68 ± 0.54
	15	–	0.79 ± 0.06b	0.61 ± 0.05a	0.61 ± 0.17ab	0.28 ± 0.01
	30	–	0.00 ± 0.00c	0.00 ± 0.00b	0.31 ± 0.22b	0.21 ± 0.01
Eucalyptol	45	–	0.00 ± 0.00c	0.00 ± 0.00b	0.21 ± 0.01b	0.00 ± 0.00
	0	–	–	–	2.89 ± 0.81b	–
	15	–	–	–	5.35 ± 0.18a	–
	30	–	–	–	4.52 ± 0.81ab	–
	45	–	–	–	4.13 ± 0.08ab	–
3-Octen-2-one	0	0.00 ± 0.00b	0.00 ± 0.00b	0.00 ± 0.00b	0.00 ± 0.00b	0.00 ± 0.00c
	15	0.23 ± 0.01b	0.25 ± 0.04b	0.11 ± 0.04b	0.38 ± 0.04a	0.09 ± 0.03b
	30	0.27 ± 0.12b	0.64 ± 0.04a	0.18 ± 0.00b	0.56 ± 0.11a	0.22 ± 0.01a
	45	0.86 ± 0.15a	0.79 ± 0.17a	0.73 ± 0.23a	0.57 ± 0.09a	0.29 ± 0.05a
	0	–	1.05 ± 0.18a	–	0.14 ± 0.00	1.83 ± 0.11
γ-Terpinene	15	–	0.92 ± 0.01a	–	0.00 ± 0.00	0.54 ± 0.05

Table 2 (continued)

Compound	Storage (days)	C	OO	MF	LN	TV
E-2-Octenal	30	–	0.00 ± 0.00b	–	0.00 ± 0.00	0.00 ± 0.00
	45	–	0.00 ± 0.00b	–	0.00 ± 0.00	0.00 ± 0.00
	0	0.00 ± 0.00b	2.61 ± 0.42b	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00c
	15	0.31 ± 0.11b	6.66 ± 0.15a	0.09 ± 0.01c	1.58 ± 1.27ab	0.00 ± 0.00c
	30	2.65 ± 2.21ab	0.00 ± 0.00c	2.34 ± 0.26b	4.19 ± 2.38a	1.06 ± 0.13b
3,5-Octadien-2-one	45	5.53 ± 1.67a	0.00 ± 0.00c	8.03 ± 0.66a	4.04 ± 0.28a	3.09 ± 0.01a
	0	–	0.00 ± 0.00c	–	–	0.00 ± 0.00c
	15	–	0.38 ± 0.02a	–	–	0.19 ± 0.04b
	30	–	0.33 ± 0.03a	–	–	0.29 ± 0.04a
	45	–	0.19 ± 0.04b	–	–	0.00 ± 0.00c
Octanol	0	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00b	–	0.00 ± 0.00
	15	0.00 ± 0.00	0.00 ± 0.00	0.00 ± 0.00b	–	0.00 ± 0.00
	30	0.00 ± 0.00	0.00 ± 0.00	0.25 ± 0.04b	–	0.00 ± 0.00
	45	0.40 ± 0.016	0.27 ± 0.12	0.75 ± 0.29a	–	0.25 ± 0.02
	0	0.00 ± 0.00	0.00 ± 0.00b	–	–	0.00 ± 0.00
2-Nonanone	15	0.00 ± 0.00	0.00 ± 0.00b	–	–	0.00 ± 0.00
	30	0.00 ± 0.00	0.10 ± 0.04b	–	–	0.00 ± 0.00
	45	0.40 ± 0.011	0.36 ± 0.06a	–	–	0.07 ± 0.03
	0	–	1.06 ± 0.31ab	–	0.12 ± 0.01	1.32 ± 0.00a
	15	–	1.45 ± 0.04a	–	0.00 ± 0.00	1.04 ± 0.03b
Linalool	30	–	0.72 ± 0.01b	–	0.00 ± 0.00	0.70 ± 0.04c
	45	–	0.24 ± 0.03c	–	0.00 ± 0.00	0.17 ± 0.00d
	0	0.08 ± 0.01	0.15 ± 0.01c	0.00 ± 0.00b	0.13 ± 0.00b	0.00 ± 0.00
	15	1.02 ± 0.06	1.29 ± 0.16b	0.51 ± 0.02b	1.17 ± 0.13a	0.46 ± 0.14
	30	1.14 ± 0.57	1.68 ± 0.07b	0.58 ± 0.04b	1.16 ± 0.04a	0.71 ± 0.06
p-Menthone	45	2.68 ± 1.76	2.37 ± 0.46a	3.28 ± 0.64a	1.18 ± 0.05a	0.62 ± 0.06
	0	–	–	0.25 ± 0.05c	–	–
	15	–	–	1.49 ± 0.11a	–	–
	30	–	–	0.37 ± 0.04c	–	–
	45	–	–	0.66 ± 0.13b	–	–
E-2-Nonenal	0	0.00 ± 0.00	0.00 ± 0.00b	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00c
	15	0.00 ± 0.00	0.00 ± 0.00b	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00c
	30	0.72 ± 0.66	0.44 ± 0.02b	0.35 ± 0.09b	0.59 ± 0.36b	0.09 ± 0.03b
	45	1.66 ± 1.32	1.71 ± 0.36a	2.02 ± 0.04a	1.37 ± 0.01a	0.60 ± 0.03a
	0	–	–	0.17 ± 0.02b	–	–
Isopulegone	15	–	–	0.67 ± 0.06a	–	–
	30	–	–	0.00 ± 0.00c	–	–
	45	–	–	0.00 ± 0.00c	–	–
	0	–	0.15 ± 0.06b	–	0.58 ± 0.21	0.11 ± 0.00b
	15	–	0.24 ± 0.00a	–	0.61 ± 0.16	0.14 ± 0.00a
Terpinen-4-ol	30	–	0.13 ± 0.01b	–	0.33 ± 0.25	0.11 ± 0.01b
	45	–	0.00 ± 0.00c	–	0.21 ± 0.01	0.00 ± 0.00c
	0	–	–	–	0.20 ± 0.06a	–
	15	–	–	–	0.26 ± 0.03a	–
	30	–	–	–	0.00 ± 0.00b	–
α-Terpineol	45	–	–	–	0.00 ± 0.00b	–
	0	–	–	5.05 ± 0.71b	–	–
	15	–	–	12.19 ± 0.47a	–	–
	30	–	–	1.97 ± 0.18c	–	–
	45	–	–	2.85 ± 0.57c	–	–
E.E-2,4-nonadienal	0	–	0.00 ± 0.00c	–	–	–
	15	–	0.00 ± 0.00c	–	–	–
	30	–	0.08 ± 0.01b	–	–	–
	45	–	0.21 ± 0.01a	–	–	–
	0	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00b	0.00 ± 0.00b	0.00 ± 0.00
E-2-Decenal	15	0.00 ± 0.00	0.09 ± 0.01c	0.00 ± 0.00b	0.40 ± 0.31b	0.00 ± 0.00
	30	1.99 ± 1.89	0.87 ± 0.04b	1.18 ± 0.42b	1.69 ± 1.31ab	0.00 ± 0.00
	45	3.37 ± 2.68	4.31 ± 0.40a	3.72 ± 0.90a	2.74 ± 0.10a	2.15 ± 0.31
	0	–	–	0.00 ± 0.00	–	–
	15	–	–	0.23 ± 0.06	–	–
Piperitenone	30	–	–	0.39 ± 0.10	–	–
	45	–	–	0.00 ± 0.00	–	–
	0	–	–	–	–	0.26 ± 0.10a
	15	–	–	–	–	0.06 ± 0.01b
	30	–	–	–	–	0.08 ± 0.01b

Table 2 (continued)

Compound	Storage (days)	C	OO	MF	LN	TV
Carvacrol	45	–	–	–	–	0.00 ± 0.00b
	0	–	2.55 ± 1.23ab	–	–	2.29 ± 0.52ab
	15	–	4.16 ± 0.00a	–	–	2.67 ± 0.11a
	30	–	3.79 ± 0.26ab	–	–	2.69 ± 0.21a
	45	–	2.30 ± 0.09b	–	–	0.77 ± 0.01b
E,Z-2,4-Decadienal	0	–	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00c
	15	–	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00c
	30	–	0.47 ± 0.06b	0.18 ± 0.06b	0.42 ± 0.06b	0.13 ± 0.07b
	45	–	1.08 ± 0.25a	0.51 ± 0.01a	0.91 ± 0.04a	0.34 ± 0.02a
	0	–	–	–	0.73 ± 0.30	–
α-Terpinyl acetate	15	–	–	–	1.14 ± 0.24	–
	30	–	–	–	0.91 ± 0.40	–
	45	–	–	–	0.94 ± 0.05	–
	0	–	–	–	–	–

Results (mean ± standard deviation) are expressed as means of total ion current (TIC) area units ($\times 10^{-6}$)

Mean values within each column for the same storage time followed by the same letters are not significantly different at $P < 0.05$

The levels of these alcohols were increased at the later stages of oxidation.

Ketones such as E-2-octene, 2-heptanone, 2-nonane, 6-methyl-5-hepten-2-one, 3-octen-2-one, and 3,5-octadien-2-one were detected in the headspace of the analyzed oils, and some of these components are known to be linked with oxidation processes.

Except for E-2-hexenal, 2-butenal, E-2-pentenal, E-2-heptenal, E-2-octenal, E-2-nonenal, and E-2-decenal were also detected. Especially, E-2-heptenal, E-2-octenal, and E-2-decenal were detected as the most abundant 2-alkenals in olive oil samples during heating. The contents of E-2-heptenal and E-2-decenal were similar in control and flavored oil samples except TV sample which exhibited lower values than those of other samples. A similar trend for E-2-octenal was observed; however, the highest value of this compound in OO was 6.66×10^6 AU after 15 days of storage, and then an immediate decrease was observed. In literature, the presence of 2-alkenals in olive oil samples treated at high temperatures was also evidenced by other studies (Gómez-Alonso et al. 2004; Issaoui et al. 2011; Poyato et al. 2014).

The members of 2,4-alkadienals group namely 2,4-heptadienal, 2,4-nonadienal, and 2,4-decadienal were not identified in the control sample. However, E,E-2,4-nonadienal was identified only in OO sample, while E,E-2,4-heptadienal was identified in both OO and LN samples. Besides, E,E-2,4-decadienal was identified in all flavored samples after 30 days of storage. Although some vegetable oils include small amounts of alkadienals, oxidized oils—in particular—contain higher amounts of these compounds whose concentrations can be related to fatty acids. Oils that are rich in oleic acid were reported to contain these compounds in fewer levels than high linoleic acid-containing oils (Wang et al. 2019).

During thermal oxidation, only one furan namely 2-pentylfuran, a typical degradation compound of linoleic

acyl groups, formed in OO, LN, and TV samples. At the end of 45 days of storage, the content of this compound in OO, LN, and TV samples reached up to 5.31, 3.90, and 1.75×10^6 AU, respectively. The presence of 2-pentylfuran was also reported in the headspace of the edible oil samples during oxidation (Morales et al. 1997; Xu et al. 2017). Our results were in good agreement with Issaoui et al. (2011) stating that 2-pentylfuran was absent in heated olive oils, while this compound appeared in thyme-flavored olive oil during thermal oxidation.

Other compounds such as terpenes, which are not normally present in non-oxidized virgin olive oil, were detected at varying concentrations in flavored oils. α-Thujene, α-pinene, sabinene, β-pinene, β-myrcene, α-terpinene, p-cymene, limonene, eucalyptol, γ-terpinene, linalool, p-menthone, 4-terpineol, α-terpineol, thymol, carvacrol, isopulegone, pulegone, piperitenone, and α-terpinyl acetate were identified in analyzed samples. These are major compounds present in essential oils and are transferred from essential oil to olive oil samples. Issaoui et al. (2011) also reported that some compounds (e.g., carvacrol, α-thujene, α-pinene, myrcene, α-terpinene, γ-terpinene, and 4-terpineol) with aromatic and antioxidant properties were released to olive oil. Carvacrol is the major component of *Origanum onites* and *Thymus vulgaris* essential oils and it was detected at high levels in OO and TV samples. This volatile compound showed oxidative stability against thermal oxidation, and a similar trend was also reported for thyme-flavored olive oils at 100 °C (Issaoui et al. 2011). Pulegone, which was transferred from *Micromeria fruticosa* essential oil, was also detected in the MF sample. Pulegone was the major component of *M. fruticosa* essential oil (65.30%). Among terpenes, p-cymene was detected in all flavored oils with a good transfer behavior to olive oil samples. The highest concentration of p-cymene was 13.86×10^6 AU in the TV sample, while its lowest level (0.56×10^6 AU) was

Table 3 Changes in volatile compounds of olive oil samples during photo-oxidation

Compound	Storage (days)	C	OO	MF	LN	TV
2-Butenal	0	0.14 ± 0.04b	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00b
	15	0.06 ± 0.03c	0.03 ± 0.02c	0.07 ± 0.01	0.03 ± 0.01bc	0.03 ± 0.01ab
	30	0.08 ± 0.01bc	0.09 ± 0.01b	0.14 ± 0.03	0.09 ± 0.01b	0.09 ± 0.02ab
	45	0.24 ± 0.01a	0.42 ± 0.01a	0.76 ± 0.62	0.82 ± 0.04a	0.13 ± 0.07a
Pentanal	0	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00c	0.00 ± 0.00b
	15	0.66 ± 0.04a	0.40 ± 0.01a	0.85 ± 0.11a	0.43 ± 0.06ab	0.32 ± 0.01a
	30	0.61 ± 0.03a	0.39 ± 0.01a	0.86 ± 0.01a	0.50 ± 0.03a	0.34 ± 0.05a
	45	0.50 ± 0.01b	0.29 ± 0.00b	0.59 ± 0.08b	0.33 ± 0.02b	0.00 ± 0.00b
E-2-Pentenal	0	0.10 ± 0.01a	0.00 ± 0.00b	0.00 ± 0.00c	–	0.00 ± 0.00b
	15	0.05 ± 0.01b	0.03 ± 0.01a	0.06 ± 0.01b	–	0.03 ± 0.01a
	30	0.04 ± 0.00b	0.03 ± 0.00a	0.05 ± 0.00b	–	0.04 ± 0.01a
	45	0.04 ± 0.00b	0.00 ± 0.00b	0.10 ± 0.00a	–	0.00 ± 0.00b
Pentanol	0	–	–	–	0.25 ± 0.03	–
	15	–	–	–	0.00 ± 0.00	–
	30	–	–	–	0.00 ± 0.00	–
	45	–	–	–	0.00 ± 0.00	–
Z-2-penten-1-ol	0	0.06 ± 0.00a	–	–	–	–
	15	0.06 ± 0.03a	–	–	–	–
	30	0.04 ± 0.01ab	–	–	–	–
	45	0.00 ± 0.00b	–	–	–	–
3-Methyl-2-butenal	0	0.00 ± 0.00b	0.00 ± 0.00	0.00 ± 0.00b	0.00 ± 0.00	–
	15	0.00 ± 0.00b	0.00 ± 0.00	0.00 ± 0.00b	0.00 ± 0.00	–
	30	0.02 ± 0.01b	0.00 ± 0.00	0.05 ± 0.01b	0.03 ± 0.01	–
	45	0.14 ± 0.01a	0.25 ± 0.03	0.53 ± 0.32a	0.41 ± 0.11	–
Hexanal	0	4.34 ± 0.08a	2.38 ± 0.12a	3.45 ± 0.05a	2.15 ± 0.09	5.62 ± 0.47a
	15	1.78 ± 0.21b	0.93 ± 0.18b	2.05 ± 0.04bc	1.03 ± 0.15	0.83 ± 0.03b
	30	1.66 ± 0.08b	0.92 ± 0.06b	1.73 ± 0.13c	1.05 ± 0.15	0.73 ± 0.01b
	45	1.67 ± 0.11b	1.02 ± 0.01b	2.47 ± 0.33b	1.66 ± 0.35	0.00 ± 0.00c
E-2-octene	0	–	–	2.51 ± 0.07a	1.32 ± 0.18	–
	15	–	–	0.01 ± 0.00b	0.00 ± 0.00	–
	30	–	–	0.00 ± 0.00b	0.05 ± 0.00	–
	45	–	–	0.00 ± 0.00b	0.00 ± 0.00	–
E-2-hexenal	0	4.38 ± 0.03a	1.15 ± 0.04c	1.58 ± 0.06c	1.46 ± 0.12a	1.04 ± 0.13b
	15	2.32 ± 0.20b	1.77 ± 0.26a	3.09 ± 0.19a	1.74 ± 0.22a	1.87 ± 0.02b
	30	2.11 ± 0.03b	1.62 ± 0.02ab	2.47 ± 0.18b	1.63 ± 0.04a	1.68 ± 0.04b
	45	1.57 ± 0.14c	1.37 ± 0.07bc	1.48 ± 0.25c	0.78 ± 0.08b	54.97 ± 13.10a
Z-3-Hexen-1-ol	0	1.86 ± 0.07	0.62 ± 0.00	0.85 ± 0.06a	0.72 ± 0.07a	0.83 ± 0.13a
	15	0.82 ± 0.21	0.56 ± 0.24	0.74 ± 0.15ab	0.60 ± 0.19a	0.55 ± 0.01b
	30	0.80 ± 0.07	0.57 ± 0.12	0.94 ± 0.09a	0.54 ± 0.03a	0.59 ± 0.01b
	45	0.62 ± 0.04	0.37 ± 0.00	0.46 ± 0.13b	0.20 ± 0.04b	0.00 ± 0.00c
E-2-hexen-1-ol	0	0.29 ± 0.01a	0.20 ± 0.01a	0.19 ± 0.02a	0.19 ± 0.01c	0.28 ± 0.07a
	15	0.13 ± 0.02b	0.09 ± 0.04b	0.17 ± 0.01a	0.10 ± 0.02b	0.10 ± 0.01b
	30	0.13 ± 0.01b	0.08 ± 0.01b	0.13 ± 0.01ab	0.09 ± 0.01b	0.09 ± 0.01b
	45	0.08 ± 0.01c	0.05 ± 0.01b	0.08 ± 0.04b	0.00 ± 0.00a	0.00 ± 0.00b
Hexanol	0	0.32 ± 0.02a	0.17 ± 0.01a	0.18 ± 0.04	0.17 ± 0.00a	0.39 ± 0.11a
	15	0.18 ± 0.01b	0.10 ± 0.04b	0.18 ± 0.03	0.11 ± 0.03ab	0.09 ± 0.00b
	30	0.18 ± 0.01b	0.08 ± 0.01b	0.15 ± 0.04	0.10 ± 0.04b	0.07 ± 0.01b
	45	0.10 ± 0.01c	0.00 ± 0.00c	0.09 ± 0.05	0.00 ± 0.00c	0.00 ± 0.00b
Nonane	0	–	–	–	1.35 ± 0.08	2.88 ± 0.57
	15	–	–	–	0.00 ± 0.00	0.00 ± 0.00
	30	–	–	–	0.00 ± 0.00	0.00 ± 0.00
	45	–	–	–	0.00 ± 0.00	0.00 ± 0.00
Heptanal	0	0.03 ± 0.01	–	–	–	–
	15	0.00 ± 0.00	–	–	–	–
	30	0.00 ± 0.00	–	–	–	–
	45	0.00 ± 0.00	–	–	–	–
α-Thujene	0	–	0.09 ± 0.00a	–	0.07 ± 0.01a	–
	15	–	0.08 ± 0.02a	–	0.05 ± 0.01b	–
	30	–	0.06 ± 0.00a	–	0.03 ± 0.00b	–
	45	–	0.00 ± 0.00b	–	0.00 ± 0.00c	–
α-Pinene	0	–	–	–	0.14 ± 0.04b	0.08 ± 0.01b
	15	–	–	–	0.26 ± 0.01a	0.09 ± 0.01b
	30	–	–	–	0.23 ± 0.00a	0.09 ± 0.01b
	45	–	–	–	0.22 ± 0.00a	0.13 ± 0.00a
E-2-Heptenal	0	0.05 ± 0.01a	0.00 ± 0.00c	0.00 ± 0.00	0.00 ± 0.00c	0.00 ± 0.00c

Table 3 (continued)

Compound	Storage (days)	C	OO	MF	LN	TV
Sabinene	15	0.20 ± 0.02b	0.16 ± 0.01bc	0.29 ± 0.00	0.17 ± 0.02bc	0.14 ± 0.03bc
	30	0.47 ± 0.03c	0.46 ± 0.01b	0.75 ± 0.01	0.50 ± 0.06b	0.47 ± 0.03a
	45	1.34 ± 0.08d	2.01 ± 0.23a	3.06 ± 2.49	3.35 ± 0.23a	0.19 ± 0.11b
	0	–	–	–	0.42 ± 0.11b	–
	15	–	–	–	0.68 ± 0.01a	–
β-Pinene	30	–	–	–	0.61 ± 0.04a	–
	45	–	–	–	0.52 ± 0.04ab	–
	0	–	–	–	0.11 ± 0.03b	–
	15	–	–	–	0.20 ± 0.01a	–
	30	–	–	–	0.18 ± 0.01a	–
Octen-3-ol	45	–	–	–	0.17 ± 0.01a	–
	0	0.05 ± 0.01b	–	–	0.00 ± 0.00	0.19 ± 0.01a
	15	0.00 ± 0.00c	–	–	0.00 ± 0.00	0.21 ± 0.02a
	30	0.02 ± 0.00c	–	–	0.00 ± 0.00	0.23 ± 0.01a
	45	0.10 ± 0.01a	–	–	0.30 ± 0.02	0.00 ± 0.00b
6-Methyl-5-hepten-2-one	0	0.00 ± 0.00d	0.00 ± 0.00d	0.00 ± 0.00	0.00 ± 0.00d	0.00 ± 0.00c
	15	0.10 ± 0.01c	0.12 ± 0.01c	0.00 ± 0.00	0.09 ± 0.01c	0.11 ± 0.01b
	30	0.19 ± 0.03b	0.18 ± 0.00b	0.00 ± 0.00	0.14 ± 0.01b	0.17 ± 0.01a
	45	0.31 ± 0.01a	0.32 ± 0.01a	0.54 ± 0.18	0.33 ± 0.03a	0.00 ± 0.00c
	0	–	0.91 ± 0.22b	0.00 ± 0.00c	0.32 ± 0.07ab	0.00 ± 0.00d
β-Myrcene	15	–	1.32 ± 0.17a	0.21 ± 0.01a	0.39 ± 0.05a	2.02 ± 0.09a
	30	–	1.14 ± 0.04ab	0.23 ± 0.04a	0.32 ± 0.06ab	1.73 ± 0.10b
	45	–	0.85 ± 0.06b	0.13 ± 0.02b	0.19 ± 0.01b	0.27 ± 0.11c
	0	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
	15	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
Octanal	30	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
	45	0.03 ± 0.00	–	0.09 ± 0.02	0.10 ± 0.04	–
	0	0.19 ± 0.01a	0.16 ± 0.02	0.12 ± 0.00c	0.15 ± 0.02	0.00 ± 0.00
	15	0.19 ± 0.01a	0.18 ± 0.01	0.23 ± 0.04a	0.15 ± 0.04	0.19 ± 0.00
	30	0.20 ± 0.01a	0.17 ± 0.00	0.20 ± 0.01ab	0.17 ± 0.01	0.19 ± 0.00
Z-3-Hexenyl acetate	45	0.15 ± 0.01b	0.16 ± 0.00	0.15 ± 0.03bc	0.11 ± 0.01	0.00 ± 0.00
	0	–	3.59 ± 0.59a	0.56 ± 0.08a	2.25 ± 0.49a	13.86 ± 1.14a
	15	–	2.85 ± 0.16ab	0.05 ± 0.01b	2.08 ± 0.04a	4.46 ± 0.01b
	30	–	2.88 ± 0.05ab	0.04 ± 0.00b	1.99 ± 0.04a	4.65 ± 0.11b
	45	–	2.40 ± 0.05b	0.00 ± 0.00b	1.22 ± 0.17b	1.42 ± 0.47c
Limonene	0	–	0.98 ± 0.03a	0.54 ± 0.07a	0.94 ± 0.20a	3.68 ± 0.54a
	15	–	0.37 ± 0.04b	0.43 ± 0.01ab	0.80 ± 0.08a	0.48 ± 0.01b
	30	–	0.40 ± 0.06b	0.42 ± 0.02ab	0.84 ± 0.07a	0.52 ± 0.01b
	45	–	0.18 ± 0.05c	0.34 ± 0.07b	0.42 ± 0.11b	0.13 ± 0.18b
	0	–	–	–	2.89 ± 0.81	–
Eucalyptol	15	–	–	–	3.90 ± 0.01	–
	30	–	–	–	3.55 ± 0.19	–
	45	–	–	–	3.49 ± 0.07	–
	0	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
	15	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
3-Octen-2-one	30	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
	45	0.04 ± 0.01	–	0.07 ± 0.04	0.09 ± 0.01	–
	0	–	1.05 ± 0.18a	–	0.14 ± 0.04a	1.83 ± 0.11a
	15	–	1.09 ± 0.11a	–	0.10 ± 0.00a	1.58 ± 0.01b
	30	–	0.96 ± 0.01a	–	0.10 ± 0.01a	1.52 ± 0.04b
γ-Terpinene	45	–	0.41 ± 0.04b	–	0.00 ± 0.00b	0.38 ± 0.06c
	0	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
	15	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
	30	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00	–
	45	0.03 ± 0.01	–	0.11 ± 0.10	0.27 ± 0.17	–
(E)-2-Octenal	0	–	1.06 ± 0.31a	–	0.12 ± 0.01b	1.32 ± 0.00b
	15	–	1.03 ± 0.04ab	–	0.14 ± 0.00a	1.51 ± 0.01a
	30	–	0.86 ± 0.04ab	–	0.10 ± 0.00c	1.27 ± 0.01b
	45	–	0.61 ± 0.02b	–	0.00 ± 0.00d	0.45 ± 0.10c
	0	0.08 ± 0.01b	0.15 ± 0.01c	0.00 ± 0.00c	0.13 ± 0.00	0.00 ± 0.00c
Nonanal	15	0.17 ± 0.04ab	0.18 ± 0.02bc	0.17 ± 0.03b	0.16 ± 0.03	0.20 ± 0.01a
	30	0.20 ± 0.02a	0.20 ± 0.01b	0.20 ± 0.01b	0.22 ± 0.00	0.23 ± 0.01a
	45	0.24 ± 0.06a	0.30 ± 0.01a	0.26 ± 0.02a	0.28 ± 0.02	0.08 ± 0.04b
	0	–	–	0.25 ± 0.05b	–	–
	15	–	–	0.52 ± 0.01a	–	–

Table 3 (continued)

Compound	Storage (days)	C	OO	MF	LN	TV
Isopulegone	30	–	–	0.54 ± 0.01a	–	–
	45	–	–	0.48 ± 0.04a	–	–
	0	–	–	0.17 ± 0.02b	–	–
	15	–	–	0.35 ± 0.04a	–	–
	30	–	–	0.35 ± 0.00a	–	–
Terpinen-4-ol	45	–	–	0.26 ± 0.08ab	–	–
	0	–	0.15 ± 0.06	–	0.58 ± 0.21	0.11 ± 0.00b
	15	–	0.20 ± 0.01	–	0.70 ± 0.04	0.24 ± 0.01a
	30	–	0.18 ± 0.01	–	0.63 ± 0.01	0.21 ± 0.01a
	45	–	0.16 ± 0.02	–	0.43 ± 0.06	0.09 ± 0.01b
α -Terpineol	0	–	0.00 ± 0.00b	–	0.20 ± 0.06	0.00 ± 0.00b
	15	–	0.07 ± 0.02a	–	0.29 ± 0.01	0.10 ± 0.02a
	30	–	0.07 ± 0.03a	–	0.27 ± 0.02	0.09 ± 0.01a
	45	–	0.08 ± 0.00a	–	0.20 ± 0.02	0.00 ± 0.00b
	0	–	–	5.05 ± 0.71c	–	–
Pulegone	15	–	–	15.52 ± 0.75a	–	–
	30	–	–	12.83 ± 0.14ab	–	–
	45	–	–	10.22 ± 2.33b	–	–
	0	0.00 ± 0.00	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00
	15	0.00 ± 0.00	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00
Dec-2(E)-enal	30	0.03 ± 0.01	0.00 ± 0.00	–	0.00 ± 0.00	0.00 ± 0.00
	45	0.03 ± 0.04	0.11 ± 0.01	–	0.17 ± 0.04	0.07 ± 0.00
	0	–	–	0.00 ± 0.00	–	–
	15	–	–	0.06 ± 0.02	–	–
	30	–	–	0.05 ± 0.01	–	–
Piperitone	45	–	–	0.00 ± 0.00	–	–
	0	–	–	–	–	0.26 ± 0.10a
	15	–	–	–	–	0.06 ± 0.01b
	30	–	–	–	–	0.05 ± 0.01b
	45	–	–	–	–	0.00 ± 0.00b
Thymol	0	–	2.55 ± 1.23b	–	–	2.29 ± 0.52b
	15	–	3.16 ± 0.51ab	–	–	3.72 ± 0.19a
	30	–	3.76 ± 0.39ab	–	–	3.88 ± 0.25a
	45	–	4.71 ± 0.11a	–	–	3.33 ± 0.57ab
	0	–	–	0.00 ± 0.00c	–	–
Carvacrol	15	–	–	0.44 ± 0.06a	–	–
	30	–	–	0.40 ± 0.01ab	–	–
	45	–	–	0.33 ± 0.05b	–	–
	0	–	–	–	0.73 ± 0.30	–
	15	–	–	–	1.16 ± 0.04	–
α -Terpinyl acetate	30	–	–	–	1.20 ± 0.19	–
	45	–	–	–	1.10 ± 0.00	–
	0	–	–	0.00 ± 0.00b	–	–
	15	–	–	0.12 ± 0.02a	–	–
	30	–	–	0.10 ± 0.01a	–	–
Piperitenone oxide	45	–	–	0.09 ± 0.00a	–	–

Results (mean ± standard deviation) are expressed as means of total ion current (TIC) area units ($\times 10^{-6}$)

Mean values within each column for the same storage time followed by the same letters are not significantly different at $P < 0.05$

analyzed in MF. Eucalyptol (1,8-cineol), which is a major component of *L. nobilis* essential oil (35.06%), was only detected in the LN sample. Also, eucalyptol remained stable during the thermal oxidation process.

Photo-Oxidation

According to the headspace analysis of oil samples, more than 20 volatile compounds were formed during storage (Table 1). The highest number of identified compounds was 33 in the

LN sample, while it was the minimum in the control sample with 20 components. The identified volatile compounds in olive oils during photo-oxidation are presented in Table 3. The major volatile compounds were hexanal and E-2-hexenal before the storage period. These compounds generally decreased during photo-oxidation in all samples with one exception; TV had the highest value of E-2-hexenal (54.97×10^6 AU) at the end of 45 days of storage. The amount of E-2-heptenal increased with prolonged storage time under photo-oxidation conditions. At the end of 45 days of storage, the

level of E-2-heptanal increased up to 3.35×10^6 AU in the LN sample, while it reached only 0.19×10^6 AU in the TV sample. E-2-heptanal was considered as a photo-oxidation product in a previous report demonstrating that it was formed and reached higher levels as oxidation occurred in such oils as apricot and plum oils which are also rich in oleic acid like olive oil (Kiralan et al. 2018). As similar to our results, Kanavouras et al. (2004) demonstrated that E-2-heptanal was more abundant in olive oil stored in the light. Such aldehydes as 2-butenal, pentanal, E-2-pentenal, 3-methyl-2-butenal, heptanal, E-2-octenal, nonanal, and E-2-decenal were also identified in the headspace of oil samples. Among these compounds, 2-butenal, pentanal, and nonanal appeared as volatile oxidation compounds in all oil samples during photo-oxidation. E-2-pentenal, 3-methyl-2-butenal, and E-2-decenal did not form in LN, TV, and MF. Heptanal was only identified in the control sample and not detected in any other group. Most of these volatile compounds increased with photo-oxidation and their levels reached the maximum at the end of the storage period.

Alcohols including pentanol, Z-3-hexen-1-ol, E-2-hexen-1-ol, hexanol, and octen-3-ol were determined in the analyzed samples. The concentrations of pentanol, Z-3-hexen-1-ol, E-2-hexen-1-ol, and hexanol were decreased or these compounds disappeared during storage. The level of octen-3-ol increased in two photo-oxidized samples, C and LN, at the end of 45 days of storage, while this compound disappeared in the TV sample at the end of the storage period.

Regarding ketones, E-2-octene, 6-methyl-5-hepten-2-one, and 3-octen-2-one were identified. E-2-octene was the only detected volatile compound in two samples (MF and LN) at the beginning and this compound also disappeared at the end of the oxidation process. A similar trend was observed in a previous study by Lee and Min (2010). The researchers reported that 2-octene decreased dramatically in the linoleic acid system stored under light. 6-Methyl-5-hepten-2-one was detected as a result of the oxidation and the level of this volatile compound was higher in oil samples at the end of storage, except TV sample. 3-Octen-2-one was present in C, MF, and LN samples only at the end of the storage. Lee and Min (2010) reported that 3-octen-2-one was only identified in one sample mixed with chlorophyll during light storage.

α -Thujene, α -pinene, sabinene, β -pinene, β -myrcene, *p*-cymene, limonene, eucalyptol, γ -terpinene, linalool, *p*-menthone, 4-terpineol, α -terpineol, thymol, carvacrol, isopulegone, pulegone, piperitone, piperitenone, piperitenone oxide, and α -terpinyl acetate were identified as terpene compounds in samples. Eucalyptol was only identified in the LN sample at levels ranging between 2.89 – 3.90×10^6 AU. Similarly, carvacrol also remained stable during storage. *p*-Menthone, isopulegone, pulegone, piperitone, piperitenone, and piperitenone oxide were also identified in the MF sample and most of them were stable during oxidation. Besides, the

concentrations of other compounds such as α -thujene, α -pinene, sabinene, β -pinene, β -myrcene, 4-terpineol, *p*-cymene, and α -terpineol did not significantly change in contrast to the main volatile compounds found in essential oils during oxidation.

Conclusion

Flavored oils have been used in such food processing techniques as cooking where heat is available. Besides, the oxidative effects of light have been studied on the valuable volatile compounds which contribute to the aroma of olive oils as well as antioxidant activity. To our best knowledge, no literature reports were available regarding the effects of thermal and photo-oxidation conditions on the volatile compounds of olive oils flavored with essential oils. Results indicated that E-2-hexenal formed at higher concentrations in oils during both oxidation processes. Besides, E-2-heptenal was another important volatile oxidation product and could be used as an indicator for thermal and photo-oxidation. On the other hand, the characteristic volatile compounds that were transferred from essential oils exhibited stable behavior against the oxidation mechanism which was a result of their antioxidant properties. The outcomes of this study are thought to be useful for the improvements in the gourmet oil industry as well as further research in this field.

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s12161-020-01926-w>.

Funding This study has not been funded by any commercial or not-for-profit organizations.

Compliance with Ethical Standards

Conflict of Interest Sunduz Sezer Kiralan declares that she has no conflict of interest. Sermin Goksu Karagoz declares that she has no conflict of interest. Gulcan Ozkan declares that she has no conflict of interest. Mustafa Kiralan declares that he has no conflict of interest. Onur Ketenoglu declares that he has no conflict of interest.

Ethical Approval Not applicable.

Informed Consent Informed consent not applicable.

References

- Akçar H, Gümüskesen A (2011) Sensory evaluation of flavored extra virgin olive oil. *Gıda* 36(5):249–254
- Angerosa F, Mostallino R, Basti C, Vito R (2001) Influence of malaxation temperature and time on the quality of virgin olive oils. *Food Chem* 72(1):19–28. [https://doi.org/10.1016/S0308-8146\(00\)00194-1](https://doi.org/10.1016/S0308-8146(00)00194-1)

- Araújo HC, Lacerda MEG, Lopes D, Bizzo HR, Kaplan MAC (2007) Studies on the aroma of maté (*Ilex paraguariensis* St. Hil.) using headspace solid-phase microextraction. *Phytochem Anal* 18:469–474. <https://doi.org/10.1002/pca.1002>
- Arcoleo G, Indovina MC, Varvaro G, Lanza C, Mazzaglia A (2009) Improving olive oil shelf life with lemon essential oil. *Chem Eng Trans* 17:849–854
- Arslan M (2012) Effects of intra-row spacing on herbage yield, essential oil content and composition of *Micromeria fruticosa*. *Farmacia* 60(6):925–931
- Asensio CM, Nepote V, Grosso NR (2013) Consumers' acceptance and quality stability of olive oil flavored with essential oils of different oregano species. *Int J Food Sci Technol* 48(11):2417–2428. <https://doi.org/10.1111/ijfs.12233>
- Ayadi MA, Grati-Kamoun N, Attia H (2009) Physico-chemical change and heat stability of extra virgin olive oils flavoured by selected Tunisian aromatic plants. *Food Chem Toxicol* 47(10):2613–2619. <https://doi.org/10.1016/j.fct.2009.07.024>
- Baiano A, Gambacorta G, La Notte E (2010) Aromatization of olive oil. *Transworld research Network* 37/661(2):1–29
- Bishr MM, Salama OM (2018) Inter and intra GC-MS differential analysis of the essential oils of three *Mentha* species growing in Egypt. *Future J Pharm Sci* 4(1):53–56. <https://doi.org/10.1016/j.fjps.2017.08.003>
- Cevik S, Ozkan G, Kiralan M (2016) Optimization of malaxation process of virgin olive oil using desired and undesired volatile contents. *LWT-Food Sci Technol* 73:514–523. <https://doi.org/10.1016/j.lwt.2016.06.058>
- Damechki M, Sotiropoulou S, Tsimidou M (2001) Antioxidant and pro-oxidant factors in oregano and rosemary gourmet olive oils. *Grasas Aceites* 52(3–4):207–213. <https://doi.org/10.3989/gya.2001.v52.i3-4.359>
- Francisco V, Ruiz-Fernández C, Lahera V, Lago F, Pino J, Skaltsounis L, González-Gay MA, Mobasher A, Gómez R, Scotecce M, Gualillo O (2019) Natural molecules for healthy lifestyles: oleocanthal from extra virgin olive oil. *J Agric Food Chem* 67(14):3845–3853. <https://doi.org/10.1021/acs.jafc.8b06723>
- Gambacorta G, Faccia M, Pati S, Lamacchia C, Baiano A, La Notte E (2007) Changes in the chemical and sensorial profile of extra virgin olive oils flavored with herbs and spices during storage. *J Food Lipids* 14(2):202–215. <https://doi.org/10.1111/j.1745-4522.2007.00080.x>
- Genovese A, Caporaso N, Leone T, Paduano A, Mena C, Perez-Jimenez MA, Sacchi R (2019) Use of odorant series for extra virgin olive oil aroma characterisation. *J Sci Food Agric* 99(3):1215–1224. <https://doi.org/10.1002/jsfa.9293>
- Gómez-Alonso S, Salvador MD, Fregapane G (2004) Evolution of the oxidation process in olive oil triacylglycerol under accelerated storage conditions (40–60°C). *J Am Oil Chem Soc* 81:177–184. <https://doi.org/10.1007/s11746-004-0878-7>
- Güllüce M, Sökmen M, Şahin F, Sökmen A, Adigüzel A, Özer H (2004) Biological activities of the essential oil and methanolic extract of *Micromeria fruticosa* (L) Druce ssp *serpyllifolia* (Bieb) PH Davis plants from the eastern Anatolia region of Turkey. *J Sci Food Agric* 84(7):735–741. <https://doi.org/10.1002/jsfa.1728>
- Issaoui M, Flamini G, Hajaj ME, Cioni PL, Hammami M (2011) Oxidative evolution of virgin and flavored olive oils under thermo-oxidation processes. *J Am Oil Chem Soc* 88:1339–1350. <https://doi.org/10.1007/s11746-011-1800-5>
- Iten F, Saller R, Abel G, Reichling J (2009) Additive antimicrobial effects of the active components of the essential oil of *Thymus vulgaris*-chemotype carvacrol. *Planta Med* 75(11):1231–1236. <https://doi.org/10.1055/s-0029-1185541>
- Jemâa JMB, Tersim N, Toudert KT, Khouja ML (2012) Insecticidal activities of essential oils from leaves of *Laurus nobilis* L. from Tunisia, Algeria and Morocco, and comparative chemical composition. *J Stored Prod Res* 48:97–104. <https://doi.org/10.1016/j.jspr.2011.10.003>
- Kalua CM, Allen MS, Bedgood DR Jr, Bishop AG, Prenzler PD, Robards K (2007) Olive oil volatile compounds, flavor development and quality: a critical review. *Food Chem* 100(1):273–286. <https://doi.org/10.1016/j.foodchem.2005.09.059>
- Kanavouras A, Hernandez-Munoz P, Coutelieres F, Selke S (2004) Oxidation-derived flavor compounds as quality indicators for packaged olive oil. *J Am Oil Chem Soc* 81:251. <https://doi.org/10.1007/s11746-004-0891-x>
- Karagoz SG, Yilmazer M, Ozkan G, Carbonell-Barrachina AA, Kiralan M, Ramadan MF (2017) Effect of cultivar and harvest time on C₆ and C₅ volatile compounds of Turkish olive oils. *Eur Food Res Technol* 243:1193–1200. <https://doi.org/10.1007/s00217-016-2833-7>
- Kiralan M, Ramadan MF (2016) Volatile oxidation compounds and stability of safflower, sesame and canola cold-pressed oils as affected by thermal and microwave treatments. *J Oleo Sci* 65(10):825–833. <https://doi.org/10.5650/jos.ess16075>
- Kiralan M, Kayahan M, Kiralan SS, Ramadan MF (2018) Effect of thermal and photo oxidation on the stability of cold-pressed plum and apricot kernel oils. *Eur Food Res Technol* 244:31–42. <https://doi.org/10.1007/s00217-017-2932-0>
- Koutidou M, Grauwet T, Van Loey A, Acharya P (2017) Impact of processing on odour-active compounds of a mixed tomato-onion puree. *Food Chem* 228:14–25. <https://doi.org/10.1016/j.foodchem.2017.01.135>
- Lee J, Min DB (2010) Analysis of volatile compounds from chlorophyll photosensitized linoleic acid by headspace solid-phase microextraction (HS-SPME). *Food Sci Biotechnol* 19:611–616. <https://doi.org/10.1007/s10068-010-0086-y>
- Lei H, Wang Y, Liang F, Su W, Feng Y, Guo X, Wang N (2010) Composition and variability of essential oils of *Platycladus orientalis* growing in China. *Biochem Syst Ecol* 38(5):1000–1006. <https://doi.org/10.1016/j.bse.2010.09.018>
- Liu J, Li S, Zhang A, Zhao W, Liu Y, Zhang Y (2017) Volatile profiles of 13 foxtail millet commercial cultivars (*Setaria italica* Beauv.) from China. *Cereal Chem* 94:170–176. <https://doi.org/10.1094/CCHEM-01-16-0007-R>
- Mallia S, Piccinalli P, Rehberger B, Badertscher R, Escher F, Schlichtherle-Cemy H (2008) Determination of storage stability of butter enriched with unsaturated fatty acids/conjugated linoleic acids (UFA/CLA) using instrumental and sensory methods. *Int Dairy J* 18(10–11):983–993. <https://doi.org/10.1016/j.idairyj.2008.05.007>
- Moldão-Martins M, Beirão-da-Costa S, Neves C, Cavaleiro C, Salgueiro L, Beirão-da-Costa ML (2004) Olive oil flavored by the essential oils of *Mentha×piperita* and *Thymus mastichina* L. *Food Qual Prefer* 15(5):447–452. <https://doi.org/10.1016/j.foodqual.2003.08.001>
- Morales MT, Rios JJ, Aparicio R (1997) Changes in the volatile composition of virgin olive oil during oxidation: flavors and off-flavors. *J Agric Food Chem* 45(7):2666–2673. <https://doi.org/10.1021/jf960585+>
- Padalia RC, Verma RS, Chanotiya CS (2011) Variability in volatile terpenoid compositions of peppermint cultivars and some wild accession from northern India. *J Essent Oil Res* 23(2):29–33. <https://doi.org/10.1080/10412905.2011.9700444>
- Petrović GM, Stamenković JG, Kostevski IR, Stojanović GS, Mitić VD, Zlatković BK (2017) Chemical composition of volatiles; antimicrobial, antioxidant and cholinesterase inhibitory activity of *Chaerophyllum aromaticum* L. (Apiaceae) essential oils and extracts. *Chem Biodivers* 14:e1600367. <https://doi.org/10.1002/cbdv.201600367>
- Poyato C, Ansorena D, Navarro-Blasco I, Astiasarán I (2014) A novel approach to monitor the oxidation process of different types of

- heated oils by using chemometric tools. *Food Res Int* 57:152–161. <https://doi.org/10.1016/j.foodres.2014.01.033>
- Romani A, Ieri F, Urciuoli S, Noce A, Marrone G, Nediani C, Bernini R (2019) Health effects of phenolic compounds found in extra-virgin olive oil, by-products, and leaf of *Olea europaea* L. *Nutrients* 11(8):1776. <https://doi.org/10.3390/nu11081776>
- Santos NO, Mariane B, Lago JHG, Sartorelli P, Rosa W, Soares MG, Da Silva AM, Lorenzi H, Vallim MA, Pascon RC (2015) Assessing the chemical composition and antimicrobial activity of essential oils from Brazilian plants—*Eremanthus erythropappus* (Asteraceae), *Plectranthus barbatus*, and *P. amboinicus* (Lamiaceae). *Molecules* 20(5):8440–8452. <https://doi.org/10.3390/molecules20058440>
- Shokoohinia Y, Yegdaneh A, Amin G, Ghannadi A (2014) Seasonal variations of *Laurus nobilis* L. leaves volatile oil components in Isfahan, Iran. *Res J Pharmacogn* 1(3):1–6
- Simić A, Soković MD, Ristić M, Grujić-Jovanović S, Vukojević J, Marin PD (2004) The chemical composition of some Lauraceae essential oils and their antifungal activities. *Phytother Res* 18(9):713–717. <https://doi.org/10.1002/ptr.1516>
- Singh N, Singh HP, Batish DR, Kohli RK, Yadav SS (2020) Chemical characterization, phytotoxic, and cytotoxic activities of essential oil of *Mentha longifolia*. *Environ Sci Pollut Res* 27:13512–13523. <https://doi.org/10.1007/s11356-020-07823-3>
- Sloan AE (2000) The top ten functional food trends. *Food Technol* 54(4):33–62
- Sousa A, Casal S, Malheiro R, Lamas H, Bento A, Pereira JA (2015) Aromatized olive oils: influence of flavoring in quality, composition, stability, antioxidants, and antiradical potential. *LWT-Food Sci Technol* 60(1):22–28. <https://doi.org/10.1016/j.lwt.2014.08.026>
- Stojanović NM, Randjelović PJ, Mladenović MZ, Ilić IR, Petrović V, Stojiljković N, Ilić S, Radulović NS (2019) Toxic essential oils, part VI: acute oral toxicity of lemon balm (*Melissa officinalis* L.) essential oil in BALB/c mice. *Food Chem Toxicol* 133:110794. <https://doi.org/10.1016/j.fct.2019.110794>
- Taoudiat A, Djenane D, Ferhat Z, Spigno G (2018) The effect of *Laurus nobilis* L. essential oil and different packaging systems on the photo-oxidative stability of *Chemlal* extra-virgin olive oil. *J Food Sci Technol* 55(10):4212–4222. <https://doi.org/10.1007/s13197-018-3357-x>
- Wang Y, Zhu M, Mei J, Luo S, Leng T, Chen Y, Nie S, Xie M (2019) Comparison of furans formation and volatile aldehydes profiles of four different vegetable oils during thermal oxidation. *J Food Sci* 84(7):1966–1978. <https://doi.org/10.1111/1750-3841.14659>
- Wu W, Tao N, Gu S (2014a) Characterization of the key odor-active compounds in steamed meat of *Coilia ectenes* from Yangtze River by GC–MS–O. *Eur Food Res Technol* 238:237–245. <https://doi.org/10.1007/s00217-013-2098-3>
- Wu N, Gu S, Tao N, Wang X, Ji S (2014b) Characterization of important odorants in steamed male Chinese mitten crab (*Eriocheir sinensis*) using gas chromatography-mass spectrometry-olfactometry. *J Food Sci* 79:C1250–C1259. <https://doi.org/10.1111/1750-3841.12511>
- Xu L, Yu X, Li M, Chen J, Wang X (2017) Monitoring oxidative stability and changes in key volatile compounds in edible oils during ambient storage through HS-SPME/GC–MS. *Int J Food Prop* 20(sup3):S2926–S2938. <https://doi.org/10.1080/10942912.2017.1382510>

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