

Electrochemical Determination of Carbendazim in Food Samples Using an Electrochemically Reduced Nitrogen-Doped Graphene Oxide-Modified Glassy Carbon Electrode

Yu Ya^{1,2} · Cuiwen Jiang^{1,2} · Leixing Mo^{1,2} · Tao Li^{1,2} · Liping Xie^{1,2} · Jie He^{1,2} · Li Tang^{1,2} · Dejiao Ning^{1,2} · Feiyan Yan^{1,2}

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Abstract Nitrogen-doped graphene oxide (NGO) was synthesized via pyrolysis of graphene oxide and urea and was characterized by transmission electron microscopy (TEM) and Xray photoelectron spectroscopy (XPS). An electrochemically reduced nitrogen-doped graphene oxide-modified glassy carbon electrode (ERNGO/GCE) was developed for the determination of carbendazim (CBZ) in food samples. The surface morphology of the modified electrode was characterized by scanning electron microscopy (SEM). Cyclic voltammetry and electrochemical impedance spectroscopy were employed to demonstrate the large electrode surface and fast electron transfer of the ERNGO/GCE. Electrochemical behaviors of CBZ at different electrodes were studied by voltammetry. Experimental results showed that the ERNGO/GCE achieved better performance for the electrochemical oxidation of CBZ than either the bare glassy carbon electrode (GCE) or the nitrogen-doped graphene oxide-modified GCE (NGO/GCE). Under optimized conditions, the ERNGO/GCE exhibited a wide linearity of 5.0~850 μ g/L with a detection limit of 1.0 μ g/L (signal-to-noise ratio = 3). Application of our proposed method in food products was shown to be practical and reliable.

Keywords Nitrogen-doped graphene oxide · Electrochemically reduced · Carbendazim · Electroanalysis

Feiyan Yan yanfeiyan2014@126.com

Introduction

Carbendazim (CBZ) is a broad-spectrum fungicide. It has been widely used in the prevention and control of various diseases that affect crops, fruits, and vegetables (Devi et al. 2015; Thomidis et al. 2009). Recently, toxicology studies suggested that CBZ can have potentially harmful effect on human health (Farag et al. 2011; Rama et al. 2014). Additionally, CBZ can persist for a long time in the environment, crops, fruits, and vegetables (Lewandowska and Walorczyk 2010). Therefore, developing an accurate, sensitive, and simple method for the determination of CBZ has practical significance for the environment and human health. Different analytical methods had been adopted for CBZ detection, including chromatography (Chen et al. 2015a; Qin et al. 2016; Subhani et al. 2013), spectrophotometry (Chen et al. 2015b; Pourreza et al. 2015; Strickland and Batt 2009), and chemiluminescence (Liao and Xie 2006; Llorent-Martínez et al. 2013). In addition, electroanalytical methods have been used in CBZ detection and have particular advantages such as high sensitivity, ease of operation, short analysis time, and low cost. Many different kinds of materials were used as electrode modifier for CBZ electrochemical detection, owing to the poor response of CBZ at the unmodified electrode. Among these materials, carbon-based nanomaterials are the most commonly used types. A variety of carbon nanotube-based electrochemical sensors, decorated with anionic surfactant (Petroni et al. 2016), mesoporous silica (Razzino et al. 2015), and graphene oxide (Luo et al. 2013), had been fabricated toward electrochemical oxidation of CBZ. We described an advanced electrochemical sensor for the determination of carbendazim based on pyrrolidinium ionic liquid/ordered mesoporous carbon composite system (Ya et al. 2015). Graphene was used as a promising electrode modifier for the fabrication of novel CBZ electrochemical sensor due to its

¹ Institute for Agricultural Product Quality Safety and Testing Technology, Guangxi Academy of Agricultural Sciences, Nanning 530007, China

² Quality Inspection and Test Center for Sugarcane and Its Product, China Ministry of Agriculture (Nanning), Nanning 530007, China

distinct physical and chemical attributes (Noyrod et al. 2014). Diamond electrode, one of the carbon-based material electrodes, is also a powerful tool for the electroanalysis of CBZ (França et al. 2012; Lima et al. 2016). Meanwhile, other research had presented carbendazim quantification by electrochemical methods using various modified electrodes, such as hetropolyacid montmorillonite clay-modified electrode (Manisankar et al. 2006), carboxylic group functionalized poly (3,4-ethylenedioxythiophene) mimic electrode (Yao et al. 2014), and zeolite-modified electrode (Maximiano et al. 2016).

Graphene is a two-dimensional layer of carbon atoms with an extended honeycomb network and has been the subject of much attention since its first artificial exfoliation in 2004 (Novoselov et al. 2004). It is widely used in electroanalysis because of its extraordinary physical and chemical properties, such as its high surface area, high thermal conductivity, high chemical stability, and rapid charge carrier mobility (Li et al. 2015; Wei et al. 2016). Chemical doping with heteroatoms, such as N, B, or P, can be used to tailor the electronic and chemical properties of graphene (Wang et al. 2014). Among these elements, nitrogen is considered to be an excellent dopant for graphene because of its comparable atomic size and number of valence electrons. Nitrogen doped in graphene can improve the electrical conductivity and stability of graphene and also leads to the formation of an increased activation region on the graphene surface (Usachov et al. 2011). Thus, nitrogen-doped graphene has great potential application in the field of electroanalysis (Shao et al. 2010; Wang et al. 2010).

Graphene is limited in electrochemical applications because of its irreversible agglomeration (Kuila et al. 2012). It is known that graphene oxide has good dispersibility in many solvents because of the polar oxygens that render graphene oxide strongly hydrophilic. However, because of the oxygencontaining groups, the electrical conductivity of the modified interface decreases, and this may limit the direct application of graphene oxide in the field of electroanalysis (Chen et al. 2012). At the same time, oxygen-containing groups of graphene oxide can be reduced, thus restoring a more perfect graphene structure with planar conjugation and further improving the conductivity of graphene oxide (Liu et al. 2014; Qiao et al. 2015). In this work, we describe a sensitive, reliable, and simple electrochemical method for the detection of CBZ based on an electrochemically reduced nitrogen-doped graphene oxide-modified glassy carbon electrode (ERNGO/ GCE). It is noted that the oxidation signal of CBZ improves significantly for ERNGO/GCE compared to the nitrogendoped graphene oxide-modified glassy carbon electrode (NGO/GCE). This is attributed to the large electrically active area and excellent electrical conductivity of the ERNGO/ GCE.

Experimental

Reagents and Apparatus

Carbendazim (99.0 %) was obtained from Dr. Ehrenstorfer GmbH Company (Augsburg, Germany). Potassium ferrocyanide and potassium hexacyanoferrate were purchased from Sigma Aldrich (St. Louis, MO, USA). Graphite powder, sodium nitrate, potassium permanganate, sulfuric acid, and hydrogen peroxide were supplied from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). All other reagents were of analytical grade and were used as received. A phosphate buffer solution (PBS) was prepared from KOH and 0.1 M H_3PO_4 , and the pH was monitored using a pH meter. All aqueous solutions were prepared in deionized water.

Electrochemical experiments were performed on a CHI760E electrochemical workstation (Chenhua Instrument Co. Ltd., China). A conventional three-electrode system was used for all electrochemical experiments. A bare or modified glassy carbon electrode (GCE) of 3 mm diameter was the working electrode, a saturated calomel electrode was used as the reference electrode, and a platinum wire electrode was used as the auxiliary electrode. X-ray photoelectron spectroscopy (XPS) analysis was performed using a VG MultiLab2000 spectrometer (ThermoVG scientific, UK). Transmission electron microscopy (TEM) images were obtained on a Tecnai G2 F20 S-Twin microscope (FEI, USA). The surface morphology of the electrodes was characterized by a S-3400N scanning electron microscope (SEM) (Hitachi Co., Japan). pH values were measured using a pH-3C precision pH meter (Leici Instruments, China).

Preparation of NGO

NGO was prepared according to a previously reported method (Du et al. 2014). In a typical synthesis, 1.0 graphite powder, 0.50 g sodium nitrate, and 50 mL sulfuric acid were mixed in ice-water bath. Nine grams of potassium permanganate was slowly added into the mixture in 2 h. Then, 1.0 g melamine was put into suspension obtained above. After stirring at 30 °C for 12 h, 46 mL hot deionized water was added into the suspension dropwise. This process was performed at 90 °C for 1 h. Subsequently, 20 mL of hydrogen peroxide was added into the suspension. Afterward, 30 min of ultrasonication was taken with the power of 200 W. The resulting suspension was centrifuged at 4000 rmp for 20 min to obtain mud-like material. The obtained material was washed with deionized water and ethanol for several times. Then, the product was dried at 40 °C for 2 days to get NGO.

Preparation of Modified Electrode

The GCE was polished to a mirror-like surface with a 0.05 μ m alumina slurry and was ultrasonically washed in a 1:1 nitric acid solution, alcohol, and deionized water for 5 min, respectively. The GCE was then dried with a high-purity nitrogen steam. One milligram of NGO was dispersed in 1.0 mL of ethanol by ultrasonication for 1 h to form a homogenous suspension. Then, 5 μ L of the obtained suspension was dipped onto the surface of the GCE and dried at room temperature; the obtained modified electrode was denoted as NGO/GCE. The ERNGO/GCE was fabricated by the following procedure: a cyclic voltammetric scan was run from 0 to -1.0 V with a scan rate of 25 mV/s using NGO/GCE as the working electrode in 0.1 M KNO₃ followed by sweeping 20 segments for efficient reduction of NGO.

Analytical Procedure

For the determination of CBZ, the three-electrode system was immersed in 10-mL aliquots of pH 4.0 PBS containing different concentrations of CBZ. After 240 s of accumulation, differential pulse voltammetry (DPV) measurements were recorded from 0.50 to 1.20 V, and the oxidation peak currents at 0.90 V were measured for CBZ with the following parameters: a potential increase of 0.004 V, an amplitude of 0.05 V, a pulse width of 0.05 s, and a pulse interval of 0.5 s.

Results and Discussion

Characterization of NGO

The XPS spectra of NGO are presented in Fig. 1. The widescan (Fig. 1a) and high-resolution spectra (Fig. 1b) of NGO reveal the presence of nitrogen in *N*-graphene, indicating that nitrogen was successfully doped into the graphene oxide. Figure 1c shows the TEM images of NGO. The typical wrinkle and flake of the graphene sheet are clearly observed in the image. Such wrinkled structure is highly beneficial in maintaining a high surface area on the electrode since the sheets cannot readily collapse back to a graphitic structure (Du et al. 2011).

Characterization of Modified Electrodes

The surface morphologies of the bare GCE, NGO/GCE, and ERNGO/GCE were characterized by SEM as shown in Fig. 2a, b, c, respectively. As seen in these images, the surface of the bare GCE is smooth and virtually featureless. On the surface of the NGO/GCE, the typical wrinkled feature is slightly visible, indicating that the NGO successfully modified the surface of the GCE. After electrochemical reduction, the wrinkled structure is clearly observed on the surface of the ERNGO/GCE, showing higher surface roughness. This surface structure of the ERNGO/GCE is highly beneficial in maintaining a high surface area on the electrode (Sun and Wang 2015).





Fig. 2 SEM images of bare GCE (**a**), NGO/GCE (**b**), and ERNGO/GCE (**c**)



The electrochemical characterizations of the different electrodes in the presence of an external redox probe $[Fe(CN)_6]^3$ ^{-/4} were carried out using cyclic voltammetry and electrochemical impedance spectroscopy. Figure 3a shows the cyclic voltammograms of 5.0 mmol/L $[Fe(CN)_6]^{3-/4-}$ for the bare GCE (curve a), the NGO/GCE (curve b), and the ERNGO/ GCE (curve c) at a scan rate of 50 mV/s. A pair of redox peaks for $[Fe(CN)_6]^{3-/4-}$ is shown for the bare GCE with a peak-topeak separation ($\triangle E_n$) of 167 mV. For the NGO/GCE, both the cathodic and anodic peak currents increased slightly, and the ΔE_p value decreased to 101 mV, which indicated high elctrocatalytic activity of the NGO. While the $\triangle E_p$ exhibited by the ERNGO/GCE was 67 mV, compared to the NGO/ GCE, the redox peak current increased significantly for the ERNGO/GCE. These results demonstrate that the electrochemical reduction can greatly improve the electrical conductivity and electrocatalytic activity of the NGO. The effect of the scan rate was assessed using the Randles-Sevcik formula (Zhou et al. 2013):

$$I_{\rm p} = 2.69 \times 10^5 D^{1/2} n^{3/2} A v^{1/2} C$$

where I_p refers to the peak current (μ A), D is the diffusion coefficient (cm² s⁻), n is the electron-transfer number, A is the electroactive surface area of the electrode (cm²), v is the scan rate (V/s), and C is the concentration of [Fe(CN)₆]^{3-/4-} (mmol/L). Here, $D = 0.76 \times 10^{-5}$, n = 1, v = 0.050, and C = 5.0. As calculated from this formula, the electroactive areas of the bare GCE, NGO/GCE, and ERNGO/GCE are 0.055, 0.071, and 0.092 cm², respectively. These results indicate that the electrochemical reduction of NGO effectively improved the electroactive area of the electrode surface.

Figure 3b shows the spectra from electrochemical impedance spectroscopy represented as Nyquist plots for the bare GCE (curve a), NGO/GCE (curve b), and ERNGO/GCE (curve c) in 5.0 mmol/L [Fe(CN)₆]^{3-/4-}. A clear semicircle is observed on the bare GCE and NGO/GCE. However, the ERNGO/GCE exhibits a nearly straight line. The







Fig. 4 Cyclic voltammetry curves for 300 μ g/L CBZ in pH 4.0 PBS at the GCE (*a*), NGO/GCE (*b*), and ERNGO/GCE (*c*)

electrochemical impedance spectroscopy result confirms that electrochemical reduction can further improve the conductivity of NGO.

Electrochemical Behaviors of CBZ at Different Electrodes

Electrochemical behaviors of CBZ at different electrodes were investigated with cyclic voltammograms, and these are shown in Fig. 4. As illustrated in Fig. 4, a very weak anodic peak was found for the bare GCE (curve a). In contrast, a well-defined anodic peak with significantly increasing peak current was observed for the NGO/GCE (curve b), indicating that NGO has remarkable enhancing effects on the electrochemical oxidation of CBZ. At the same time, a sharp anodic peak can be seen for the ERNGO/GCE (curve c). The peak current of CBZ for the ERNGO/GCE was onefold higher than that for the NGO/GCE (curve b). This result may be attributed to two factors: (1) ERNGO provides a larger electroactive area which increases the loading amount of CBZ at the modified electrode surface and (2) the excellent conductivity of ERNGO can accelerate electron transfer between the modified electrode and CBZ in solution.

Effect of Scan Rate

The linear sweep voltammetry technique was used to probe the effect of the scan rate on the electrochemical oxidation of CBZ. Figure 5a shows the relationships between the scan rate and the current responses for 150 μ g/L of CBZ at the ERNGO/GCE. The current response increased linearly with the scan rate in the range of 25~200 mV/s and can be expressed as the following: I_p (μA) = 0.03406 v (mV/s) + 1.977(r = 0.997). This finding indicates that the electrochemical oxidation of CBZ at the ERNGO/GCE is an adsorptioncontrolled process. In addition, a linear relationship between the oxidation peak potential (E_{pa}) and Napierian logarithm of scan rate $(\ln v)$ is also observed in the scan rate range of 25~200 mV/s (Fig. 5b). A linear calibration curve with a regression equation of E_{pa} (V) = 0.9980 + 0.02477 ln v (V/s) (r = 0.996) was obtained. For an adsorption-controlled and totally irreversible electrode process, the relationship between $E_{\rm pa}$ and $\ln v$ is expressed through the Laviron's equation (Laviron 1974):

$$E_{\rm pa} = E^0 + \frac{RT}{[anF]} \ln \left[\frac{RTk}{anF} \right] + \left[\frac{RT}{anF} \right] \ln k$$

where E_{pa} is the oxidation peak potential (V), E^0 is the formal potential, *R* is the universal gas constant (8.314 J/mol K), *T* is the temperature (298 k), α is the electron-transfer coefficient, *n* is the electron-transfer number, *F* is the Faraday constant (96,480 C/mol), and *v* is the scan rate (V/s). In our study, αn is calculated to be 1.0367. Usually, α is assumed as 0.5 in a totally irreversible electrode process. The obtained for *n* is 2.07 \approx 2.0. So, two electrons are involved in the electrochemical oxidation of CBZ.



Fig. 5 a The relationship between the current response for $150 \mu g/L$ CBZ at the ERNGO/GCE in pH 4.0 PBS and the scan rate. b The plot of oxidation peak potential of CBZ versus Napierian logarithm of scan rate



Fig. 6 a Effect of pH on peak current and E_{pa} of 300 μ g/L CBZ at the ERNGO/GCE. **b** The effect of accumulation time on the peak current for 300 μ g/L CBZ at the ERNGO/GCE

Effect of pH and Accumulation Time

The influence of pH on the current response for 300 μ g/L of CBZ at the ERNGO/GCE was investigated in the pH range from 2.0 to 9.0 using DPV (Fig. 6a). The results indicate that solution pH influences the oxidation peak current of CBZ. The peak current increased with decreasing pH in the range of 9.0 to 4.0 and decreased at lower pH values. Thus, a pH value of 4.0 was adopted for subsequent analytical experiments. The relationship between E_{pa} and pH was also depicted in Fig. 6a. As we can see, $E_{\rm pa}$ linearly shifted negatively with the increasing of pH value, which suggested that protons participated in the electrochemical oxidation of CBZ directly. A good linear relationship between E_{pa} and pH was constructed with a linear regression equation expressed as $E_{pa} = 1.15-0.064$ pH (r = 0.997). The slope value of -64 mV/pH was close to the theoretical value of -59 mV/pH, indicating that the number of electron transfer was equal to the proton number involved in the electrochemical oxidation of CBZ. Thus, the electrochemical oxidation of CBZ involves two electrons and two protons, which is in agreement with the literature reported. (Yao et al. 2014). The electrochemical oxidation of CBZ can be elucidated in Scheme 1.

The influence of accumulation time on current responses of CBZ was investigated in the range of 60 to 360 s (Fig. 6b). Peak currents increased rapidly up to240 s and changed slightly when accumulation time was longer than 240 s. Therefore, an accumulation time of 240 s was chosen for all of the experiments.

Scheme 1 Electrochemical oxidation of CBZ

Analytical Performance of ERNGO/GCE

Under the optimized conditions, DPV was used to electrochemically determine CBZ on the ERNGO/GCE. Figure 7 gives the DPV responses at different concentrations of CBZ. It is clear that, with an increase in CBZ concentration, the current response increased gradually. The current response showed good linear correlation with CBZ concentration from 5.0 to 850 µg/L. The regression equation for this relationship was I_p (µA) = 0.02258C (µg/L) + 0.5972 (r = 0.997). The detection limit was calculated as 1.0 µg/L (signal-to-noise ratio = 3).

Reproducibility and Stability

The reproducibility of the ERNGO/GCE was evaluated by repeated measurement of 150 μ g/L of CBZ. Six different modified electrodes showed a relative standard deviation (RSD) of 5.4 %, and for one electrode, the RSD was 4.2 % for six repeated measurements. The stability of the ERNGO/GCE was investigated after long-term storage in a refrigerator at 4 °C for 1 month, and 90.1 % of the original oxidation peak current was retained. These results indicate that the fabricated modified electrode had good reproducibility and stability.

Interference Study

The selectivity of the ERNGO/GCE was investigated by determination of 100 μ g/L of CBZ in the presence of different common interferences. In this study, the tolerance limit was defined as the molar ratio of potential interferences/CBZ that



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Fig. 7 a DPV curves of CBZ at the ERNGO/GCE in pH 4.0 PBS with different concentrations (from a to i: 5.0, 20, 40, 80, 100, 200, 400, 800, and $850 \mu g/L$, respectively). **b** Linear relationship between the current response and the CBZ concentration

caused the ± 5.0 % change in the current response of CBZ. The results revealed that more than 5000-fold sucrose, glucose, and fructose;1000-fold Na⁺, K⁺, Ca²⁺, Mg²⁺, Cu²⁺, Zn²⁺, Fe³⁺, Al³⁺, Cl⁻, NO₃⁻, CO₃²⁻, and SO₄²⁻; and 200-fold methyl parathion, ethyl parathion, vitamin B₁, B₂, C, and E, Laspartic acid, glutamic acid, serine, thiophanate, thiophanatemethyl, thiabendazole, endodsulfan, ametryn, fenamiphos, and benomyl did not interfere with the determination of CBZ. As shown in Fig. 8, the peak current of 100 μ g/L CBZ at the ERNGO/GCE in the absence (curve a) and presence (curve b) of pesticide mixtures containing 200-fold of methyl parathion, ethyl parathion, thiophanate, thiophanatemethyl, thiabendazole, endodsulfan, ametryn, fenamiphos, and benomyl was about the same. The result suggests that the ERNGO/GCE has good selectivity.

Practical Applications



To evaluate the feasibility of the developed method, the ERNGO/GCE was used to measure the concentration of CBZ in Dendrobium candidum samples by using a standard

Fig. 8 DPV curves of 100 µg/L CBZ at the ERNGO/GCE in the absence (a) and presence (b) of pesticide mixtures

addition method. One gram of D. candidum power was added to 20 mL of ethanol, and the mixture was sonicated for 5 h. The mixture was then separated by centrifugation at 4000 rpm for 5 min. The resulting ethanol solution was evaporated to dryness using a mild nitrogen stream, and the residue was reconstituted in 1.0 mL of ethanol. One hundred microliters of this ethanol solution was injected into 10 mL of pH 4.0 PBS, and then the measurement was made by the method described above. The results are shown in Table 1, and it can be seen that the results were satisfactory. Recovery was in the range of 90.5~94.8 %, indicating that the described method is adequate for practical application in detecting CBZ in real samples.

Conclusions

NGO was successfully synthesized by a simple method and was used as a modifier to the GCE. A novel electrochemical method for the determination of CBZ was developed based on the ERNGO/GCE. As an advanced sensor electrode, the ERNGO/GCE exhibited better electrochemical performance than both the bare GCE and NGO/GCE. This was attributed to the large electroactive area and its good conductivity. The ERNGO/GCE has the advantages of a simple preparation, an easily renewable surface, and a low cost. It is thus expected to have further applications in the electroanalytical field.

Table 1 Results for the determination of CBZ in Dendrobium candidum samples

Added (μ g/L)	Found ($\mu g/L$)	Recovery (%)
20.0	18.1	90.5
50.0	47.6	95.2
100	97.3	97.3
400	379	94.8
	Added (µg/L) 20.0 50.0 100 400	Added (μg/L) Found (μg/L) 20.0 18.1 50.0 47.6 100 97.3 400 379

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Compliance with Ethical Standards

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Conflict of Interest Yu Ya declares that he has no conflict of interest. Cuiwen Jiang declares that he has no conflict of interest. Leixing Mo declares that he has no conflict of interest. Tao Li declares that he has no conflict of interest. Liping Xie declares that he has no conflict of interest. Jie He declares that he has no conflict of interest. Li Tang declares that he has no conflict of interest. Dejiao Ning declares that he has no conflict of interest. Feiyan Yan declares that he has no conflict of interest.

Ethical Approval This article does not contain any studies with human participants or animals performed by any of the authors.

Informed Consent Not applicable.

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