Preconcentration and Trace Determination of Chromium Using Modified Ionic Liquid Cold-Induced Aggregation Dispersive Liquid–Liquid Microextraction: Application to Different Water and Food Samples

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Abstract An efficient microextraction procedure based on modified ionic liquid cold-induced aggregation dispersive liquid–liquid microextraction (M-IL-CIA-DLLME) was developed for trace determination of chromium in water and food samples by flame atomic absorption spectrometry (FAAS), and it was used for speciation of Cr(III) and Cr (VI) in water samples by using $Na₂SO₃$ as the reducing agent. A mixture of water-immiscible 1-hexyl-3-methylimidazolium hexafluorophosphate ([Hmim][PF_6]) ionic liquid (IL) (microextraction solvent) and ethanol (disperser solvent) were directly injected into a heated aqueous solution containing bis(2-methoxy benzaldehyde) ethylene diimine as a Schiff's base ligand (chelating agent), hexafluorophosphate (NaP F_6 ; as a common ion) and Cr(III). Afterwards, the solution was placed in an ice-water bath and a cloudy solution was formed due to a considerable decrease of IL solubility. After centrifuging, the sedimented phase containing enriched analyte was determined by FAAS. Under the optimum conditions, the calibration graph was linear over the range of 2–50 μ gL⁻¹ with limit of detection of 0.7 μg L−¹ . The accuracy of the present methodology was tested by recovery experiments and by analyzing a certified reference material. Relative standard deviation (RSD %) was 2.7 % for Cr(III). The proposed method was successfully applied for trace determination of chromium in water and food samples.

Keywords Modified ionic liquid cold-induced aggregation dispersive liquid–liquid microextraction .Chromium .Flame atomic absorption spectrometry . Ionic liquid

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Introduction

Chromium shows various oxidation states from 0 to $+6$, but the most stable forms are $+3$, and $+6$, which exist in equilibrium in water. Cr(III) and Cr(VI) show different properties and toxicities. Cr(VI) is 100 times more toxic than Cr (III) and there are no specific antidotes. Cr(VI) is very carcinogenic and causes skin allergy and some diseases, such as dermatitis, dermal necrosis and dermal corrosion (Gad [1989](#page-7-0)), while Cr(III) is a necessary nutrient and has a significant task in transferring sugar, proteins and fat (Anderson [1989](#page-7-0)). Heavy metals are potentially toxic and may cause health problems to humans via the food chain. Maximum permissible levels (MCL) in drinking water as recommended by United States Environmental Protection Agency (US-EPA) and World Health Organization (WHO) are 50 μ gL⁻¹.

In recent years, speciation methods are becoming more and more popular. Different methods have been used for speciation of chromium and other heavy metals, such as precipitation (Duran et al. [2011](#page-7-0); Soylak et al. [2011](#page-7-0); Uluozlu et al. [2009](#page-8-0)), liquid–liquid extraction (Agrawal and Sharma [2005](#page-7-0)), cloud point extraction (Matos et al. [2009\)](#page-7-0), dispersive liquid–liquid microextraction (DLLME; Ying et al. [2011;](#page-8-0) Chen et al. [2010](#page-7-0)), and solid phase extraction (SPE; Tuzen et al. [2006](#page-8-0); Ghaedi et al. [2008;](#page-7-0) Monasterio et al. [2009\)](#page-7-0). Different analytical methods have been applied for recognizing chromium, such as flame atomic absorption spectrometry (FAAS; Shemirani et al. [2003a](#page-7-0)), inductively coupled plasma-atomic emission spectrometry (Liang et al. [2006\)](#page-7-0), inductively coupled plasma-mass spectrometry (Sun et al. [2006](#page-8-0); Pantsar-Kallio and Manninen [1999\)](#page-7-0), electrothermalatomic absorption spectrometry (Hao et al. [2010\)](#page-7-0), electrochemistry (Kiptoo et al. [2004;](#page-7-0) Kuban et al. [2003\)](#page-7-0), fluorometry (Hosseini and Belador [2009](#page-7-0)), chemiluminescence

(Yuan et al. [2008\)](#page-8-0), and UV–Vis spectrophotometry (Mahmoud et al. [2008\)](#page-7-0).

In order to increase the concentration of analyte and reduce matrix effects, development of sample pretreatment methods prior to analysis is necessary. It is well documented that DLLME belongs to one of the best microextraction procedures (Bidari et al. [2007](#page-7-0); Jahromi et al. [2007;](#page-7-0) Zeeb et al. [2010\)](#page-8-0). But the usage of toxic organic solvents in this sample pretreatment procedure is an important problem. In order to completely remove the toxic extraction solvents in preconcentration methods, several sample preparation techniques based on ionic liquids (ILs), such as ionic liquidbased liquid–liquid microextraction (Hirayama et al. [2005](#page-7-0); Berton et al. [2009](#page-7-0)), ionic liquid-based dispersive liquid– liquid microextraction (IL-DLLME; Yao and Anderson [2009;](#page-8-0) Yao et al. [2011;](#page-8-0) Gharehbaghi et al. [2009a](#page-7-0); Abdolmohammad-Zadeh and Sadeghi [2010](#page-7-0); Yousefi and Shemirani [2010](#page-8-0)), cold-induced aggregation microextraction (CIAME; Baghdadi and Shemirani [2008](#page-7-0); Gharehbaghi et al. [2009b](#page-7-0)), ionic liquid-based single-drop microextraction (Vidal et al. [2010](#page-8-0); Aguilera-Herrador et al. [2008;](#page-7-0) Zhou and Ye [2008\)](#page-8-0), temperature-controlled ionic liquid dispersive liquid phase microextraction (Zhou et al. [2008](#page-8-0)), have been applied. In recent years, CIAME and IL-DLLME have shown excellent results in the case of preconcentration and isolation. In CIAME procedure, IL is dispersed through the sample using a relatively high temperature and afterwards, the IL is aggregated by applying a low temperature. On the contrary, in IL-DLLME method the microextraction solvent is dispersed into the sample by applying a disperser solvent. In comparison with CIAME, IL-DLLME is more efficient for dispersing the microextraction phase through the sample thus significantly improves the extraction time and the extraction efficiency. It is well established that the solubility of ILs in aqueous phase depends on temperature changes. As a result, CIAME requires less amount of IL to complete the extraction process. Therefore, combination of these two microextraction methods (ionic liquid cold-induced aggregation dispersive liquid–liquid microextraction) can offer all the advantages mentioned above (Zhang et al. [2010\)](#page-8-0). Based on the results obtained in our previous studies, performance of microextraction procedures based on ILs depends on variations in the ionic strength of the sample solution (Zeeb and Sadeghi [2011;](#page-8-0) Zeeb et al. [2011\)](#page-8-0). It is well documented that solubility of ILs increases as the ionic strength of the aqueous solution increases. To solve this problem, a common ion of the IL was dissolved in the sample solution. As a result, solubility of the IL significantly decreased and volume of the settled phase was not affected by variations of the salt content of the sample. In the present work, bis(2 methoxybenzaldehyde) ethylene diimine is a Schiff's base ligand, which acts as a complexing agent. The structure of this ligand is shown in (Fig. 1).

Fig. 1 Structure of Schiff's base bis(2-methoxybenzaldehyde) ethylene diimine

In the present study, combination of M-IL-CIA-DLLME and FAAS was applied for trace determination of chromium in water and food samples, and it was used for speciation and trace determination of two oxidation states of Cr in water samples with satisfactory results.

Experimental

Apparatus

A Shimadzu AA-6300 (Kyoto, Japan, www.shimadzu.com) flame atomic absorption spectrometer equipped with deuterium background correction, chromium hollow-cathode lamp (Hamamatsu Photonics, Shizuoka, Japan, [www.hamamatsu.](http://www.hamamatsu.com) [com\)](http://www.hamamatsu.com) and air-acetylene flame was used for the measurement of chromium. The operational conditions are summarized in Table 1. The pH values were measured with a Metrohm pH meter (Herisau, Switzerland, www.metrohm-ag.com). Test tubes with a conic bottom were used for extraction and centrifuging vessels in order to trap and remove the sedimented phase. A 1-mL Hamilton syringe (Reno, NV, USA, [www.](http://www.hamiltoncompany.com) [hamiltoncompany.com](http://www.hamiltoncompany.com)) was used. A centrifuge (centurion scientific Ltd model: 1020D, West Sussex, England,

Table 1 Instrumental and experimental conditions for chromium determination

M-IL-CIA-DLLME parameters	Selected conditions	
Amount of [Hmim][PF_6] (mg)	90	
Amount of NaP F_6	200	
pH	10	
Volume of disperser solvent (μL)	500	
Chelating agent concentration (M)	10^{-4}	
Extraction temperature $(^{\circ}C)$	40	
Instrumental parameters	Selected conditions	
Slit (nm)	0.5	
Spectral band pass (nm)	0.5	
Wavelength (nm)	357.9	
Air flow rate $(Lmin^{-1})$	8	
Acetylene $(Lmin^{-1})$	2.6	

[www.centurionscientific.net\)](http://www.centurionscientific.net) was used to accelerate separation of the phases. An adjustable 1,000 μL micropipette (Hamilton, Germany) was used to take an appropriate amount of organic solvent.

Reagents and Materials

All used reagents were of analytical grade with high purity. 1-Hexyl-3-methylimidazolium hexafluorophosphate [Hmim][PF_6], acetone, acetonitrile, methanol, ethanol, sodium hexafluorophosphate (NaP F_6), Na₂SO₃, ascorbic acid, and all used salts were purchased from Merck Co. (Darmstadt, Germany, [www.merck.de\)](http://www.merck.de). Cr(III) and Cr(VI) stock solutions $(1,000 \text{ mgL}^{-1})$ were prepared from Cr(NO₃)₃.9H₂O and $K_2Cr_2O_7$, respectively. The working standard solutions were prepared daily by a suitable stepwise dilution of the stock solution with deionized water. Double deionized water was used throughout the procedure. A solution of 250 mgmL⁻¹ NaPF₆ was prepared by dissolving an appropriate amount of NaP F_6 in deionized water. The pH of the solution was adjusted to 10 using NH_3/NH_4 ⁺ buffer (0.1 M). In order to reduce loss of adsorption, since small amounts of chromium may be adsorbed on the surface of the vessel, the glassware were soaked in 10– 20 % nitric acid solution overnight, and then rinsed with deionized water. To avoid a source of contamination, high quality nitric acid was used. Stock solutions of interfering ions were prepared from their high purity compounds. Bis(2-methoxybenzaldehyde) ethylene diimine and reducing agent were dissolved in ethanol and double deionized water, respectively. The chelating-agent solution was prepared by dissolving 0.458 g of bis(2-methoxybenzaldehyde) ethylene diimine in 100 mL of ethanol (the molecular weight of chelating agent is 296 g/mol).

Synthesis of Schiff's Base $[(CH₃)₂Salen]$

Bis(2-methoxybenzaldehyde) ethylene diimine was synthesized according to the following procedure. 0.3 g of ethylenediamine was dissolved in 40 mL of ethanol. Afterwards, the obtained solution was transferred into a 250-mL flask. In the next section, 1.36 g of 2-methoxybenzaldehyde was dissolved in 30 mL of ethanol and then this solution was added dropwise to the first obtained solution, which was under reflux condition. After cooling the resulting solution, it was filtered and then recrystallized with ethanol for its purification. Afterwards, the solution was vacuum-dried for 12 h (Shemirani et al. [2003b](#page-7-0)).

Sample Preparation

Preparation of Water Samples

Tap water was freshly collected from our laboratory, after allowing it to flow for 1 min. River water was collected from Karaj River (Alborz, Iran) and Chamran River (Shiraz, Iran). These samples were collected in PTFE containers, which were soaked in concentrated $HNO₃$ and washed with deionized water. In order to remove suspended particles, they were centrifuged for 10 min at 2,000 rpm and then filtered through 0.45 μm membrane filter. To avoid the losses of chromium, since chromium can be adsorbed onto container's wall, the samples and solutions were kept in diluted nitric or hydrochloric acid (0.05 M) below pH 1.5. In acidic conditions, reduction of Cr(VI) to Cr(III) is favored, which is an unpleasant reaction while performing the speciation. Therefore, all the samples were analyzed as soon as possible (maximum recommended interval of time for storage is 8 h).

Preparation of Food Samples

Non-fat long life liquid cow's milk was purchased from the local market. The liquid sample (0.50 mL) and 5 mL of $HNO₃$ were mixed in a glass beaker at a fairly low temperature and then heated to dryness. Then it was left at room temperature in order to cool down. Then 3.0 mL of H_2O_2 (30 %) was added and the above described procedure was repeated to obtain nearly 0.50 mL of the sample solution. After cooling down to room temperature, the solution was transferred into a 50-mL volumetric flask and diluted to the mark with deionized water. The filtration of the digested solution is not necessary because there is no particle in the suspension to cause any difficulty in the absorption measurement by FAAS. Black tea (0.5 g) and green tea (0.5 g) were weighted and then transferred into a ceramic vessel. Concentrated nitric acid (6 mL) was added to this sample, and after 15 min it was heated for 5 min at 150 °C. Afterward, the resulting solution was cooled and 3 mL of hydrogen peroxide was added. Then, this solution was heated to dryness at 200 °C, and the residue was dissolved in nitric acid. The solution was transferred into a flask and neutralized with NaOH. After neutralization step, the obtained sample was diluted to 50 mL and analyzed using the proposed procedure. 1 g of wheat flour was dried to stable weight at 200 °C, put into a muffle furnace, heated to 400 °C and kept at this temperature for 8 h. The residue was then cooled, treated by 8.0 mL of concentrated nitric acid and 2 mL of 30 % (w/v) H₂O₂. The obtained solution was kept in furnace for 4 h at 400 °C. Afterwards, 4 mL of concentrated hydrochloric acid and 3 mL of 70 $\%$ (w/v) perchloric acid were added to this and evaporated to fumes. Finally, the resulting material was transferred to a volumetric flask, and diluted to 50 mL.

General Analytical Procedure

In M-IL-CIA-DLLME method, 30 mL of the sample or standard solution containing Cr(III) and 10^{-4} M bis(2methoxybenzaldehyde) ethylene diimine (Schiff's base ligand, as a chelating agent) was adjusted to pH 10.0 in a test tube. 0.8 mL of NaPF₆ (250 mgmL⁻¹) was then added to this. Afterwards, the tube was transferred into a thermo stated bath and kept at 40 °C for 5 min. A solution containing 500 μL of ethanol and 90 mg of 1-hexyl-3-methylimidazolium hexafluorophosphate [Hmim][PF_6] was injected into the sample using a 1.0-mL syringe. Afterward, the resulting solution was cooled in an ice-water bath for 5 min and a cloudy condition was formed due to a considerable decrease of IL solubility. The solution was centrifuged for 5 min at 4,000 rpm. After removal of the aqueous solution, the sedimented phase (about $9 \mu L$) in the glass test tube was dissolved in a diluting agent (0.5 mL of 0.1 M $HNO₃$ in ethanol) and introduced into the FAAS by conventional aspiration.

Speciation Procedure

Total chromium was determined by addition of 10^{-3} M $Na₂SO₃$ to the sample solution and therefore Cr(VI) was converted to Cr(III). After 40 min, the solution was diluted with ultra pure water in a calibrated flask, and then the pH was adjusted to 10 with NaOH. Finally, Cr(III) was determined as indicated in the general procedure. The concentration of Cr(VI) was obtained as the difference between the total chromium and Cr(III).

Results and Discussion

Elemental Analysis of Chelating Agent

Elemental analysis of bis(2-methoxybenzaldehyde) ethylene diimine gave the following percentages C 72.90 %, H 6.71 %, and N 9.44 %.

Selection of Reducing Agent

In order to examine the effect of reducing agent, 10^{-3} M $Na₂SO₃$ and ascorbic acid (10 %w/v) were chosen as reducing reagents to reduce Cr(VI) to Cr(III). One hundred percent efficiency was obtained by $Na₂SO₃$, while a reduction of 67 % was observed when ascorbic acid solution was used as reducing agent. Therefore, $Na₂SO₃$ was used as reducing agent in the following experiment.

Selection of IL and Disperser Solvent

The IL applied in this microextraction must be liquid and show suitable hydrophobicity. In this type of microextraction procedures, the density of IL must be high enough to let the dispersed droplets of IL settle to the bottom of the test tube. Short alkyl chain imidazolium-based ILs containing CI^- , BF_4^- , and $CF_3SO_3^-$ show hydrophilic behavior and ILs containing PF_6^- and $(CF_3SO_2)_2N^-$ show hydrophobic behavior. IL must be inexpensive. ILs containing $(CF_3SO_2)_2N^-$ are relatively expensive and those containing PF_6^- are fairly inexpensive. According to the above criteria and the cost of ILs, $[Hmim][PF_6]$ was selected as an extraction phase. Miscibility of the dispersive solvent in the extraction phase and the aqueous sample is a critical point for the selection of the disperser solvent in M-IL-CIA-DLLME procedure. Various disperser solvents including acetone, ethanol, acetonitrile, and methanol were chosen as disperser solvents. The results revealed that the type of the organic solvent had no significant effect on the performance of the microextraction method. Due to less toxicity of ethanol, it was selected for all experiments.

Effect of pH

Type and concentration of chromium species are dependent on different processes, such as hydrolysis, complexation, redox and adsorption reactions. Bis(2-methoxybenzaldehyde) ethylene diimine forms a selective complex with Cr (III). Stability of a metal–chelate complex and extraction of Cr(III) is highly dependent on pH. The influence of pH on the M-IL-CIA-DLLME was studied over the pH range of 3- 13. At low pH protons can compete with the metal for the coordination sites of a ligand, and in basic media of the sample solution, more stable complex formation and displacement reaction can occur. In the absence of complexing agent, Cr(III) exists as hexa-aqua chromium (III), in basic media $Cr(OH)₂⁺$ is converted to $Cr(OH)₃$, which is the dominant form in pH 4–10. At pH 10, the highest absor-bance was obtained (Fig. [2](#page-4-0)), because formation of $Cr(OH)$ ₃ was complete and selective hydrophobic complex was formed between bis(2-methoxy benzaldehyde) ethylene diimine and Cr(III). Therefore Cr(III) can be extracted into the fine droplets of IL and consequently, Cr(III) ions are separated and preconcentration takes place. Cr(III) shows amphoteric behavior in pH≥10 and forms a soluble complex $[Cr(OH)₄]$. While the extraction efficiency of Cr(III) is quantitative at pH 10, the extraction efficiency of Cr(VI) is rather low. In order to separate $Cr(III)$ from $Cr(VI)$ and determine the concentration of $Cr(III)$, $pH=10$ was selected for further studies.

Effect of Chelating Agent Concentration

The effect of different concentrations of bis(2-methoxybenzaldehyde) ethylene diimine on the recovery of Cr(III) was examined. Low concentration of the chelating agent was not enough to react with chromium. By increasing the concentration of chelating agent, the recovery increased. In

Fig. 2 Effect of pH on the analytical signals. Experimental conditions: Cr (III) [or Cr (VI)] concentration 50 $\mu g L^{-1}$, IL amount 90 mg, NaPF₆ amount 200 mg, volume of disperser solvent 500 μL, chelating agent concentration 10−⁴ M, and extraction temperature 40 °C

concentrations greater than 10^{-4} M, the signal decreased because the excess amount of ligand dissolves the hydrophobic metal–chelate (Fig. 3). The optimum concentration of the chelating agent was 10^{-4} M.

Effect of Diluting Agent

Because of high viscosity of ILs, they must be diluted with a diluting agent prior to the introduction into the FAAS instrument. Different solvents including methanol, ethanol, acetone, and acetonitrile were examined in order to select a diluting agent that can completely dissolve the IL phase and provide the highest signals. The best analytical performance was achieved by using ethanol. Thus, in order to dissociate the complex and release Cr(III) into the solution, 0.5 mL of 0.1 M $HNO₃$ in ethanol was used as a diluting

Fig. 3 Optimization of chelating agent concentration. Experimental conditions: chromium concentration 50 $\mu g L^{-1}$, IL amount 90 mg, $NaPF₆$ amount 200 mg, volume of disperser solvent 500 μ L, extraction temperature 40 °C, and pH 10

agent in all experiments. It should be noted that appropriate dilution could decrease the matrix effect.

Effect of IL Amount and Volume of Disperser Solvent

The amount of $[Hmim][PF_6]$ IL that is used as an extractor in this sample preparation method is an important parameter to obtain high signals of chromium. Therefore, the microextraction system was carefully evaluated in order to obtain the highest analytical responses. The variation in the recovery as a function of the amount of IL that was added to 30.0 mL of the sample was studied in the range of 10- 120 mg (Fig. 4). The results obtained in this experiment show that the analytical responses increase as the amount of IL increases and then start to decrease. Therefore, 90 mg of IL was selected as the optimum value. Effect of the volume of disperser solvent (ethanol) on the signals was tested in the range of 400 to 1,000 μL. In the studied range, no appreciable change in analytical signals was observed. As a result, 500 μL of ethanol was chosen for all experiments.

Effect of Common Ion Amount and Ionic Strength

Each ion of IL can be dissolved in the studied sample solution as a common ion, in order to decrease IL solubility and subsequently to increase performance of the system. In the present experiment, $NaPF_6$ was selected as a common ion source and its influence on the analytical responses was tested in the range of 0-300 mg (Fig. [5](#page-5-0)). These results reveal that the analytical signals increase as the amount of $NaPF_6$ increases and then remain nearly constant. Therefore, 200 mg of this agent was used for the rest of the work. Based on the results obtained in our previous studies (Zeeb and Sadeghi [2011](#page-8-0); Zeeb et al. [2011\)](#page-8-0), in sample pretreatment methods based on the application of ILs, the volume of the enriched phase depends on the salt concentration of the

Fig. 4 Influence of IL amount on the extraction performance. Experimental conditions: chromium concentration 50 $\mu g L^{-1}$, NaPF₆ amount 200 mg, volume of disperser solvent 500 μL, chelating agent concentration 10−⁴ M, extraction temperature 40 °C, and pH 10

Fig. 5 Effect of common ion (PF_6^-) on the extraction efficiency. Experimental conditions: chromium concentration 50 $\mu g L^{-1}$, IL amount 90 mg, volume of disperser solvent 500 μL, chelating agent concentration 10−⁴ M, extraction temperature 40 °C, and pH 10

sample solution, which is due to the dependence of ILs' solubility upon the ionic strength of the aqueous solution. In order to solve the mentioned problem, a common ion of IL was dissolved in the sample solution. NaCl was chosen as an electrolyte to study influence of the ionic strength on the system performance. This factor was tested within the range of 0–35 % (w/v) . The results obtained in this experiment revealed that phase separation occurred up to 30 % of NaCl. It can be concluded that in the presence of common ion of IL, the performance of the presented sample pretreatment method is not affected by variations in the ionic strength of the studied sample.

Effect of Temperature

Temperature effect on the microextraction performance was tested in the range of 30–50 °C. This experiment showed that the highest signals were obtained at 40 °C. Thus, 40 °C was chosen as the optimum value for all experiments. After injection of a binary solution consisting of disperser and extraction solvent, solutions were cooled in the temperature range of 0–25 °C. The results revealed that when the temperature decreased, the sensitivity of analytical signals

Table 2 Analytical characteristics of M-IL-CIA-DLLME for determination of chromium

Analytical feature	
$2 - 50$	
0.9984	
0.7	
2.7	
60	
30	
$<$ 6	

increased due to a decrease in $[Hint] [PF_6]$ solubility. Thus, a temperature of 0 °C was used as the optimal one.

Interferences

In order to demonstrate selectivity of the proposed method for determining chromium and competition between interfering ions, effect of addition of transition metal ions, such as Pb²⁺, Zn²⁺, Cd²⁺, Co²⁺, Ni²⁺, and Cu²⁺, to a 50 µgL⁻¹ Cr (III) upon complexation with complexing agent were examined at ratios of 1:1, 1:10 and 1:100. But of course, a high concentration of chromium can decrease this competition effect. Bis(2-methoxybenzaldehyde) ethylene diimine is selective enough and interfering ions with a specific tolerance limit did not change the signal more than ± 5 %. In this study, chromium was quantitatively extracted and coexisting ions did not interfere with the extraction of chromium.

Analytical Figures of Merits

Several analytical parameters, such as repeatability, linearity, correlation of coefficient, and detection limit under optimized conditions were studied and a good linear regression between the absorbance and the concentration was found. The analytical characteristics of the optimized method are summarized in Table 2.

Table 3 Application of M-IL-CIA-DLLME-FAAS to the determination of Cr(III) in water and food samples

Samples	Certified value (μgg^{-1})	Added Cr $(\mu g g^{-1})$	Found Cr $(\mu gg^{-1})^a$	Recovery $(\%)$
GBW 07605 Tea Black tea	0.8		0.8 ± 0.0	100
			0.6 ± 0.0	
		\overline{c}	2.5 ± 0.1	96.2
Green tea			0.5 ± 0.0	
		\overline{c}	2.2 ± 0.1	88
Wheat flour			0.4 ± 0.0	
		$\overline{2}$	2.3 ± 0.1	95.8
Samples		Added Cr $(\mu g L^{-1})$	Found Cr $(\mu g L^{-1})$	Recovery $(\%)$
Cow's milk			ND^b	
		5°	4.6 ± 0.2	92
River water $(1)^d$			2.2 ± 0.1	
		5	7.4 ± 0.3	102.8
River water (2) ^d			2.9 ± 0.1	
		5	7.4 ± 0.4	93.7

^a Mean \pm standard deviation (n=3).

b Not detected.

^c This concentration was added to the diluted sample solution.

 d Samples (1) and (2) were obtained form Karaj River (Alborz, Iran) and Chamran River (Shiraz, Iran), respectively.

Table 4 Speciation and determination of Cr(III) and Cr(VI) using M-IL-CIA-DLLME-FAAS

^a Mean \pm standard deviation (n=3). ^b Bottled mineral water was pur-

chased from local markets. ^c Not detected.

Analytical Applications

To validate the accuracy of the proposed method, extraction efficiency of chromium in different samples was studied. Different samples, such as wheat flour, green tea, black tea, cow's milk, tap water and river water were analyzed by the present methodology to determine chromium concentration. The accuracy of the proposed analytical method was tested by means of recovery experiments and analysis of certified reference material (GBW 07605 Tea). The obtained results are shown in Table [3](#page-5-0). In addition, the proposed method was applied for determination and speciation of chromium in tap water and mineral water (Table 4). These results show the accuracy and validity of the combined analytical methodology for trace determination and speciation of chromium in water and food samples.

Comparison with Other Methods

The figures of merit of the presented method (i.e., LOD, LR, and RSD) were compared with some of the previously reported methods and the results are shown in Table 5. As can be seen, the method has a relatively low LOD, wide dynamic range and good reproducibility. The results obtained in this study indicate the fact that M-IL-CIA-DLLME-FAAS is a sensitive, safe, environmentally friendly, and reproducible technique. The method developed in this work is recommended as a suitable alternative to expensive instrumental methods for pre-concentration, speciation, and determination of chromium in real samples. The usage of toxic organic solvents in previously reported sample pretreatment procedure such as DLLME is an important problem. On the contrary, in the present approach, $[Hmim][PF_6]$ was selected as a green microextraction phase and an alternative to traditional volatile organic solvents. In addition, the performance of the present sample preparation method was not affected by variations of the ionic strength of the sample.

Conclusion

In this work a new method has been developed for preconcentration and speciation of trace amounts of chromium by M-IL-CIA-DLLME combined with FAAS. The proposed technique provides a simple, inexpensive, rapid, sensitive, reliable, safe, environmentally friendly, and less tedious tool for quantitation. A Schiff's base ligand reacts selectively with Cr(III) in the presence of foreign ions. The extraction

a Limit of detection

^b Relative standard deviation

^c Electrothermal-atomic absorption spectrometry

^d Inductively coupled plasma-atomic emission spectrometry

time is very short because the equilibrium state is achieved quickly. High extraction recoveries were obtained for a small volume of only 30 mL. The presented method was applied to determination of chromium in environmental water and food samples with satisfactory results.

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