

Nitrogen Use Efficiency for Sugarcane-Biofuel Production: What Is Next?

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Abstract Land area devoted to sugarcane (Saccharum spp.) production in Brazil has increased from 2 million to 10 million ha over the past four decades. Studies have shown that, from an environmental perspective, the transformation of nitrogen (N) fertilizers into $N₂O$ gases can offset the advantages gained by replacing fossil fuels with biofuels. Our objectives here were to review recent developments in N management for sugarcane-biofuel production and assess estimates of N use efficiency (NUE) and N losses based on future scenarios, as well as for life-cycle assessments of bioenergy production. Approximately 60 % of N-based fertilizer applied to sugarcane fields in Brazil is recovered by plants and soils, whereas N losses to leaching and N_2O emissions can average 5.6 and 1.84 % of the total applied N, respectively. Maintenance of trash, rotation with N-fixing legume species, and optimization of byproducts usage have potential for reducing the N requirements of sugarcane cultivation in Brazil. Moreover, the development of sugarcane genotypes with higher NUEs, along with management systems that consider soil capacity of mineralization, is required for improving the NUE of sugarcane. Strategies to maintain N as NH_4^+ in sugarcane-cropped soils also have the potential to reduce N losses and enhance NUE. The development of second-

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generation biofuels is important for increasing biofuel production while simultaneously maintaining N rates and improving NUE, and sugarcane systems in Brazil show potential for sustainable biofuel production with low N rates and limited $N₂O$ losses. Reducing N rates in sugarcane fields is thus necessary for improving sugarcane-based biofuel production and reducing its environmental impacts.

Keywords Review \cdot Ethanol \cdot NUE \cdot N₂O \cdot Nitrogen fertilizer

Introduction

Production of biofuel from sugarcane has numerous advantages over that from other crops such as maize, wheat, and sugar beets, which include lower energy demand over the course of the production process and highly positive energy balances [[1](#page-12-0)–[4\]](#page-12-0). This feedstock has a high photosynthetic efficiency in tropical regions, resulting in high biomass production, which, in association with the re-use of its byproducts, has environmental and economic advantages over fossil fuels [[2](#page-12-0), [5,](#page-12-0) [6\]](#page-12-0).

Cultivation of feedstock is generally viewed as controversial due to the uncertainty regarding greenhouse-gas (GHG) emission reductions and its potential competition with land use for biodiversity and food [\[7](#page-12-0)]. N_2O is an important N component for the net GHG balance of biofuels, as N_2O is 300fold more potent than $CO₂$ as a GHG and, therefore, a small increase in N_2O emissions resulting from additional fertilizer use can offset large $CO₂$ reductions through the replacement of fossil fuels by biofuels [[7,](#page-12-0) [8\]](#page-12-0).

Production of ethanol from sugarcane is one of the most robust GHG-saving options based on first-generation biofuel production [\[9](#page-12-0)]. Optimization of NUE and development of second-generation technologies are also options for increasing

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sustainability of biofuel production [\[7\]](#page-12-0). Second-generation ethanol production by enzymatic hydrolysis of trash or bagasse is under development in Brazil, with potential to increase ethanol production per area without increasing application of N-based fertilizers. Brazil uses less N in its sugarcane production systems than all other large-scale producers worldwide [\[32](#page-13-0)]. Recent developments in N management for sugarcane production in Brazil include in situ quantification of N losses from leaching, denitrification, volatilization, and a series of N-response trials in the main areas of intense sugarcane cultivation.

Given their tendency to increase N rates, the rapid expansion of green-cane trash blanketing (GCTB) systems has posed challenges to traditional N management practices, especially if increasing N rates do not directly enhance yields in light of the positive correlation between fertilizer N rates and degree of N losses to the environment [\[7](#page-12-0)]. Reviewing recently developed N response curves for growing conditions in Brazil will be helpful to ascertain whether there is an increase in N response under GCTB cultivation in the country—information that is critical for the development of effective N management strategies.

Brazil is currently the largest sugarcane producer in the world, and the area under sugarcane cultivation is expected to continue to expand as domestic demand and consumption of ethanol products increase over the coming decades. As this expansion will occur primarily in areas with low fertility soils,—degraded grasslands, for instance—substantial N inputs will be required to generate high yields. Although bioenergy production has the potential to reduce $CO₂$ emissions from the burning of fossil fuels, these crops also bring the issues relating to increasing N requirements for generating high crop yields and the environmental concerns associated with the use of synthetic fertilizers to the attention of the scientific community.

Our objectives were therefore to review recent developments in N management for sugarcane-ethanol production and to provide reliable estimates of NUE and N losses that can be useful to future assessments of bioenergy production. The review discusses the most up-to-date information about the preferred forms of N uptake by sugarcane, NUE, in situ measurements of N losses, and the effects of trash and crop rotation on N requirements, as well as N-response trials for the development of N management guidelines for GCTB systems. Finally, we highlight the main advances required in terms of N management strategies to advance sugarcane production while reducing N usage.

Sugarcane in Brazil: Current Scenario

Sugarcane production in Brazil generates over 1 million jobs directly and 3.6 million indirectly, and a foreign exchange of around US\$107 billion per year [\[10](#page-12-0)]. Approximately 45 % of

the total energy matrix in Brazil is composed of renewable energy sources [\[11,](#page-12-0) [12](#page-12-0)], with 42 % of renewable sources consisting of sugarcane biomass (used for ethanol or electricity production), 28 % hydroelectric, 20 % wood, and 10 % deriving from other sources [[13](#page-12-0)]. Brazilian sugarcane biofuel production creates six times as many employment opportunities as does the country's petroleum sector [[14\]](#page-12-0).

Of the 27 million ha of sugarcane produced worldwide, the largest share, at 10.2 million ha, is found in Brazil, followed by India (5.1 million ha) and China (1.8 million ha) [\[15](#page-12-0)]. The government of Brazil developed the National Alcohol Program (Proálcool) in 1975 to promote large-scale substitution of petrol-derived motor fuels with ethanol, which consequently drove the rapid increase in the Brazilian sugarcane industry [[2\]](#page-12-0). As a result, the area under sugarcane cultivation in Brazil expanded from 2 million ha in 1975 to 10.2 million ha in 20[1](#page-2-0)3 (Fig. 1) and annual production increased from 91 million Mg to 768 million Mg while the national average productivity increased from 46 to 75 Mg ha^{-1} .

Some projections indicate that the land area under sugarcane cultivation in Brazil will reach 11.5 million ha by 2024, with sugarcane production rising to an estimated 884 million Mg [\[13\]](#page-12-0). From 2015 to 2024, sugar production could increase from 39 to 48 million Mg and ethanol production from 30 to 42.5 billion L [\[13](#page-12-0)]. Another advantage that the sugarcane industry in Brazil has compared to that in other countries is the flexibility that most sugarcane mills have for producing sugar or ethanol in the same plant, thereby allowing rapid transition from one product to another in response to changing economic conditions or market demand. Presently, around 53 % of sugarcane milled in Brazil is allocated for ethanol production, with the remaining 47 % intended for sugar production, by 2024, however, it is estimated that closer to 60 % of sugarcane will be processed for ethanol production following the expected increase in domestic consumption [[13](#page-12-0)].

Approximately 90 % of the country's sugarcane yield is generated in southeastern Brazil, whereas São Paulo is the single largest producer state, with 56 % of national production [\[16](#page-12-0)]. The shift from burned to GCTB systems occurred rapidly over the past decade in Brazil. Approximately 66 % of the cultivated area in the state of São Paulo was burned in 2006, causing environmental problems due to emissions of air pollutants (particulate matter, carbon monoxide, and hydrocarbons) and GHG emissions (CH₄ and N_2O), as well as affecting local fauna and flora. In 2014, 84 % of the cultivated area in the state was harvested under the GCTB system [[17\]](#page-12-0). The preservation of sugarcane residues on the soil surface minimizes GHG emissions from the burning process and may promote carbon (C) storage in soils, and, over the long term, may also result in lower amounts of N application to the crop due to enhanced N release through trash mineralization, thus reducing GHG emissions from fertilizer N sources [[18](#page-12-0)]. Maintaining crop residues over soil surfaces brings new

Fig. 1 Evolution of cultivated area, production and average yield of sugarcane in Brazil from 1975 to 2013. Source: FAOSTAT $[15]$ $[15]$

challenges, however. The main difficulties caused by a thick layer of trash, when compared with sugarcane fields that were previously burned, related to the incorporation of fertilizers [\[19\]](#page-12-0), loss of N due to $NH₃$ volatilization from surfaceapplied urea [\[20](#page-12-0), [21](#page-12-0)], and a higher incidence of pests [[22\]](#page-12-0). Maintenance of trash on the soil surface may also delay sugarcane sprouting during the winter in colder regions, subsequently affecting yields [\[23\]](#page-12-0).

In addition to struggling with problems associated with maintenance of surface trash, the Brazilian sugar and ethanol sectors have also sought to increase energy generation in an effort to boost profitability. One of the strategies they have adopted includes the use of trash left in the fields following mechanical harvesting. Sugarcane trash can be burned in boilers to generate steam for electricity production (cogeneration) or it can be used for the production of secondgeneration ethanol [[24\]](#page-12-0). Considering the large-scale production of cellulosic ethanol from ground bagasse and crop residues (i.e., trash), an important topic of discussion is how to effectively manage the sugarcane residues in the fields [[25\]](#page-12-0). Related issues to be addressed include the proportion of trash to be removed from soil for this purpose and how much trash should be left in the fields to provide physical, chemical, and biological benefits to the soil-plant system.

Nitrogen Forms and Nitrogen Use Efficiency by Sugarcane

The main forms of available N for plant uptake in the soil are ammonium (NH_4^+) and nitrate (NO_3^-), which are either generated as products of the mineralization of soil organic matter (SOM) or added in the form of mineral fertilizers [[26\]](#page-12-0). In environments where N availability is limited, plants and microorganisms are also able to absorb intact amino acid, small peptides, and urea [[27](#page-12-0)–[29](#page-12-0)], as reported by Vinall et al. [\[30](#page-13-0)] in a study demonstrating that sugarcane plants take up some amino acids. However, it is generally accepted that mineral forms are the main sources of N to plants in agricultural systems.

Research on sugarcane using a 15 N-labeled nutrient solution has shown that N recovery is higher when the nutrient was provided as NH_4^+ than NO_3^- [[31,](#page-13-0) [32](#page-13-0)]. However, considering the short-term assessment performed by Robinson et al. [\[32](#page-13-0)], further research is required to ascertain the preferential forms of N for sugarcane over the entirety of the crop cycle under field conditions $(\sim 12 \text{ months})$. This is more apparent considering the rapid conversions of NH₄⁺ into NO₃⁻ through nitrification that occur under field conditions, as reported by D'Andréa [[33\]](#page-13-0), who assessed gross nitrification rates in 17 soils under sugarcane cultivation in São Paulo. If the observation made by Robinson et al. [[32](#page-13-0)] occurs, to a large extent, under field conditions, the use of nitrification inhibitors such as dicyandiamide (DCD) and 3,4-dimethylpyrazole phosphate (DMPP) may be a temporary alternative for attenuating the conversion rate from NH_4^+ into NO_3^- by soil-nitrifying bacteria [[34\]](#page-13-0) and, thus, may enhance sugarcane NUE.

Several studies using the 15 N tracer through isotope dilution methods were undertaken in Brazil over the past several decades. In such studies, three indices are usually reported: (1) percentage of the fertilizer N assimilated by plants (i.e., NUE); (2) percentage of the fertilizer N remaining in the soil (NDFS); and (3) percentage of the total N content deriving from fertilizer in plants (NDFP). Values of sugarcane NUE in Brazil range between 7 and 40 % (average of 26 %) [\[35](#page-13-0)–[40\]](#page-13-0), whereas NDFS varies between 23 and 37 % (average of 32 %) [\[38,](#page-13-0) [40](#page-13-0)–[42\]](#page-13-0). Together, the proportion of the fertilizer N taken up by plants and remaining in the soil (e.g., immobilized, soluble, or sorbed) is $~58$ %, with losses by leaching, volatilization, denitrification, runoff, and NH_3 and N_2O emissions via the

leaves accounting for the remaining 42 % or so. Furthermore, foliar emissions of ammonia occur in senescent tissues via the stomata due to decreases in the NH_3 compensation point [[43\]](#page-13-0), and it seems to be the main pathway of N loss during advanced growth stages [\[44\]](#page-13-0), in which losses as high as 90 kg N ha^{-1} have been reported in sugarcane fields [[36](#page-13-0)].

NDFP at the time of crop harvest normally varies between 10 and 28 % [\[35](#page-13-0), [37,](#page-13-0) [40,](#page-13-0) [41](#page-13-0), [45](#page-13-0)–[47\]](#page-13-0), which means that 28 % of the N found in the plant (with the exception of the roots in most cases) derived from fertilizers and 72 % from other sources, such as SOM mineralization, biological N fixation (BNF), dry and wet deposition, and so on. A common finding in such studies is that NDFP levels are higher during initial growth stages and decrease through the final growing stages. Franco et al. [[46](#page-13-0)] and Vieira-Megda et al. [\[47](#page-13-0)], for instance, observed that NDFP in sugarcane reached 60–70 % in the initial stages of the crop cycle but had dropped to 10–20 % at the time of harvesting, which demonstrates the importance of N fertilizer application for crop growth in the initial stages and the importance of soil N reserves for the remaining period. Immediate release of soluble N fertilizers explains optimal N fertilizer recovery by crops immediately following fertilizer application [\[48](#page-13-0)]; during crop growth, however, N derived from SOM proportionally increases as the root system expands, which does not occur with N from fertilizers [\[46](#page-13-0)]. This finding indicated that SOM, and not fertilizers, is the main source of N for sugarcane over the course of the entire crop cycle, as previously reported [[46](#page-13-0)–[49](#page-13-0)]. As such, soil N supply from mineralization must be considered in N management systems where increasing NUE is a primary goal.

Strategies for improving sugarcane NUE include adoption of best fertilizer management practices, which will be discussed later in this review. Reducing N rates without affecting yield is an alternative to increasing NUE in sugarcane systems, but a soil-based N management approach must be developed to determine under what conditions N fertilization can be reduced. Otto et al. [[49](#page-13-0)], for example, designated sites were as highly, moderately or non-responsive to N fertilization. Although N fertilization should not be reduced in highly responsive sites to avoid yield limitations, it can be substantially reduced (from 100 kg ha⁻¹ to 50 kg ha⁻¹ N, for instance) in moderately responsive sites, or even eliminated in nonresponsive sites. The strategy of reducing N rates while maintaining yields was proven to be the best option for increasing sustainability of biofuels production [\[50\]](#page-13-0).

Another means of improving sugarcane NUE in Brazil is through the use of more N-efficient sugarcane genotypes. Identification and quantification studies have demonstrated the NUE-related differences among different sugarcane genotypes [\[51](#page-13-0), [52\]](#page-13-0), with some genotypes being highly efficient in using available N to produce biomass, whereas others are capable of storing N during the initial growth stages [[32,](#page-13-0) [51](#page-13-0)–[53\]](#page-13-0). Evaluating genotypic differences in terms of NUE by sugarcane is an important strategy for enhancing NUE and hence decreasing the environmental risks that arise from N fertilization. Over the long term, breeding programs should focus on developing genotypes with improved NUE, thereby allowing growers to choose genotypes that deliver high yields with low N demand. However, in terms of genetically modified genotypes, current attention is focused mainly on breeding resistance to herbicides, insects, drought-tolerance, and high sugar content [\[54](#page-13-0)] rather than improving NUE. As such, further research on NUE-related genotypic differences among sugarcane genotypes is greatly needed.

Nitrogen Losses in Sugarcane Fields

Globally, approximately 2.5 Tg of fertilizer N is applied to sugarcane fields every year, representing 2 % of total global fertilizer N use [\[32\]](#page-13-0). The magnitude of N losses and the low NUE of sugarcane translate into significant financial and environmental costs [\[55](#page-13-0), [56](#page-13-0)], and there is growing concern about the economic and environmental sustainability of the increasing dependency on fertilizer N for the production of bioenergy crops [\[7\]](#page-12-0).

Of the top producing nations worldwide, sugarcane cropped in Brazil uses N fertilizer more efficiently, given the low N rates applied, the relatively high yields, and the favorable growing conditions. Robinson et al. [\[32](#page-13-0)], for example, reported that N-application rates are considerably lower in Brazil (60-100 kg N ha⁻¹ y⁻¹), than in India (150-400 kg N ha⁻¹ y⁻¹) and China (100–755 kg N ha⁻¹ y⁻¹), furthermore noting that Brazil produces approximately 42 % of the world's supply of sugarcane but consumes only 25 % of the N used for sugarcane production, whereas India and China collectively produce 31 % of global sugarcane and consume 50 % of N fertilizer applied to sugarcane fields. Nitrogen rates were reduced from 200 to 160 kg N ha⁻¹ yr⁻¹ in sugarcane fields in Australia [\[57\]](#page-13-0) following an unprecedented scrutiny of the environmental impacts of N application rates and N losses attributed to the country's sugarcane industry [[55\]](#page-13-0). Consequently, efforts have been made to develop best management practices aimed at improving NUE and reducing N losses in Australian sugarcane fields [\[57\]](#page-13-0).

Following N fertilizer application, microbial oxidation of NH_4^+ to NO_3^- in the soil releases H^+ , resulting in soil acidification over the long term [[58](#page-13-0)]. Nitrate can also be leached from the root zone since soil colloids and SOM do not hold $NO₃⁻$. Leaching of $NO₃⁻$ is often accompanied by basic cations such as Ca, Mg, and K, resulting in $H⁺$ concentrations in the soil solution that increase acidification [\[59](#page-13-0)]. Soil acidification is of great concern to sugarcane producers in locations with more acidic soils, as it limits yields [\[60](#page-13-0)]. Leaching of $NO₃⁻$ can result in environmental problems, especially in regard to water quality; the magnitude of $NO₃⁻$ losses varies, depending on soil texture, soil charges in the subsoil, water

balance, and N-management practices (rates, timing, and method used; e.g., single or split N-fertilizer application), with the highest rates of leaching most likely to occur on coarsetextured, free-draining soils following periods of heavy precipitation [\[61\]](#page-13-0).

To compare the influence of soil properties and management practices on the potential of leaching losses in sugarcane soils, we will compare the production systems in Brazil and Australia. Nitrogen rates adopted in Brazil are lower than in Australia, both in the plant cane and ratoon [[32\]](#page-13-0). The sugarcane-cultivated area in Australia is largely located within wet tropical and humid subtropical climates separated by areas of unsuitable soils or unreliable rainfall [\[62\]](#page-14-0). In some areas, frequent flooding may occur during the wet season [\[55](#page-13-0)]. Not surprisingly, sugarcane cultivation was ranked as the largest source of anthropogenic dissolved mineral N in the Great Barrier Reef catchments [\[63](#page-14-0)]. In Brazil, sugarcane is cultivated mainly in deep, well-drained, and highly weathered soils [\[64](#page-14-0)], and the expansion of sugarcane in Brazil will occur mainly in areas of similar conditions. In subsurface horizons of highly weathered soils with low SOM, iron (Fe) and aluminum (Al) oxides prevail and, depending on soil pH, development of anion exchange capacity can sorb anions, such as $NO₃⁻$ [[65](#page-14-0)]. Therefore, given the comparatively deep and weathered soils, only limited leaching of fertilizer N has been reported for the sugarcane production systems of Brazil (Table 1) and, thus, only relatively small amounts of $NO_3^$ enter water tables. Reductions in N leaching losses were also observed for sugarcane productions systems in Argentina when moderate N rates were applied to the crop [\[70\]](#page-14-0).

Denitrification is defined as the conversion of NO_3 ⁻ to gaseous forms of N (nitric oxide—NO, nitrous oxide—N₂O, or di-nitrogen— N_2) by microorganisms under anaerobic conditions [[71\]](#page-14-0). Among these three N gases, special attention shall be given to N_2O , a GHG with a global-warming potential 298-fold higher than that of $CO₂$ [\[63](#page-14-0)]. Emissions of N₂O have raised concerns over the past several years in terms of the sustainability of biofuel production [[7](#page-12-0)–[9](#page-12-0), [72](#page-14-0)–[74\]](#page-14-0). The

Table 1 Leaching of N from native soil and ¹⁵N-labeled fertilizer in studies performed in sugarcane fields in Brazil

Crop cycle N rates		Losses by leaching from		Reference
	kg ha ⁻¹	Native soil-N kg ha ⁻¹	Fertilizer-N %	
Plant cane	0, 30, 60, 90	5.0, 2.5, 4.3, 3.91	θ	[66]
Plant cane	120	18.3	0.02	[67]
Plant cane	120	1.1	0.05	[68]
Ratoon	0, 100, 150	3.9, 34.3, 21.6	22.5	[69]
Average		10.5	5.6	

Mean values of sugarcane harvested with prior burning and under the green cane trash blanketing system

Intergovernmental Panel on Climate Change (IPCC) has estimated that 1 % of applied N fertilizers is converted into N_2O in agricultural systems [\[75](#page-14-0)], but this factor could be even higher if indirect emissions are also taken into account. For example, Crutzen et al. [\[8](#page-12-0)], in deriving N_2O emission factors from global N budgets, fixed N inputs, and atmospheric concentrations of N_2O , calculated an overall emission factor closer to 3–5 %. Using these values in the estimations, they found that the production of biofuels from rapeseed and corn may, in fact, contribute to global warming via increased N_2O emissions as opposed to having a mitigating effect by substituting for fossil fuels. In the study of Crutzen et al. [[8\]](#page-12-0), ethanol from sugarcane was the only environmentally friendly option from the perspective of GHG emissions, even considering the relatively high emission factor of his estimations. However, N_2O emissions from sugarcane fields are highly heterogeneous [\[76](#page-14-0)–[81\]](#page-14-0): a series of in situ measurements of $N₂O$ emissions developed over the past several years show emissions factors both higher and much lower than the values reported by Crutzen et al. [\[8\]](#page-12-0) depending on the site.

High $N₂O$ emissions can be expected from waterlogged soils with high C and/or NO_3^- contents, and high temperature [\[76](#page-14-0)]. GCTB systems also promote higher N_2O emissions, as they promote soil-moisture retention and increased microbial activity [[77](#page-14-0)]. In sugarcane systems in Australia, it has been estimated that from 1 to 6.7 % of N fertilizer can be converted into $N₂O$ [[76](#page-14-0)]; under favorable conditions, denitrification losses can be as high as 21 % of fertilizer N [[78](#page-14-0), [79](#page-14-0)]. In a review, Lisboa et al. [[79\]](#page-14-0) calculated an emission factor of 3.9 % (without subtracting background values) using data from Australia, Hawaii, and Brazil. Measurements of denitrification losses under sugarcane cultivation in other large-scale producers (e.g., China and India) are notably lacking.

Rates of denitrification are lower on the well-drained and low-C-content soils on which sugarcane in Brazil are typically grown; Soares et al. [[80\]](#page-14-0) reported N_2O emissions of 0.7– 0.75 % over two consecutive seasons under sugarcane cultivation in the state of São Paulo, for instance. Emissions of 1.1 % of N fertilizer applied over GCTB were observed, but combined vinasse application boosted N_2O emissions to 3 % [\[81](#page-14-0)]. Soil and climatic conditions following fertilizer application may affect the magnitude of losses, as demonstrated by Signor et al. [\[82](#page-14-0)], who found that losses of ammonium nitrate reached 1.2–1.5 % and that of urea 0.3–1.1 % at one experimental site, and under conditions more conducive to losses (e.g., maintenance of trash, rainfall events after fertilizer application), these values were as high as 2.9–6.7 % for urea and 0.8–13.0 % for ammonium nitrate [\[82\]](#page-14-0). Based on data compiled from recent studies carried out under Brazilian field conditions, an average 1.84 % emission factor was estimated (Table [2](#page-5-0)). In addition, replacing fossil fuels with ethanol production from sugarcane reduces overall GHG emissions even at N₂O emission levels of 3–5 % [\[8](#page-12-0)].

Table 2 In situ measurements of N2O emissions in sugarcane fields in Brazil

Crop cycle	Fertilizer management ^a	N rates kg ha ⁻¹	N_2O emissions ^b $\%$	Reference
Plant cane	Urea	60	1.11	[81]
Plant cane	Urea $+$ press mud	122	1.10	[81]
Plant cane	Urea $+$ vinasse	87	2.65	[81]
Plant cane	Urea + press $mud + vinasse$	149	1.56	[81]
Ratoon (Piracicaba)	Urea	60, 90, 120, 180	0.8, 1.33, 6.21, 12.95	[82]
Ratoon (Piracicaba)	Ammonium nitrate	60, 120, 180	1.22, 1.53, 1.22	[82]
Ratoon (Goianésia)	Urea	60, 90, 120, 180	2.85, 3.59, 6.76, 4.31	$[82]$
Ratoon (Goianésia)	Ammonium nitrate	60, 120, 180	1.10, 0.63, 0.31	[82]
Ratoon	Urea	120	0.69 (y1) / 0.75 (y2)	[80]
Ratoon	$Urea + DCD$	120	0.04 (y1) / 0.14 (y2)	[80]
Ratoon	Urea + DMPP	120	0.01(y1)/0.00(y2)	[80]
Ratoon	Urea + $DCD-R$	120	0.07(y2)	[80]
Ratoon	$Urea + DMPP-R$	120	0.05(y2)	[80]
Ratoon	PSCU	120	0.93 (y1) / 1.09 (y2)	[80]
Ratoon	Urea	120	0.68, 0.96, 0.76, 2.03°	[81]
Ratoon	Urea $+$ vinasse	142	0.59, 1.19, 1.89, 3.03°	[81]
Average			1.84	

^a DCD, dicyandiamide; DMPP, 3,4-dimethylpyrazole phosphate; PSCU, polymer-S coated urea; R, nitrification inhibitor reapplied in the same plot of previous cycle

 b y1, year 1; y2; year 2</sup>

^c Respectively for 0, 7, 14, and 21 Mg ha⁻¹ of trash

Nitrogen losses via denitrification may be reduced if adequate fertilization strategies are adopted, such as application rates based on the variable N demand of plants, as well as the ability of soil to supply N [[83\]](#page-14-0). Another strategy for mitigating $N₂O$ emissions from N fertilization is by adding nitrification inhibitors directly to the fertilizers, thereby reducing rates of microbial oxidation of NH_4^+ to NO_3^- [\[34\]](#page-13-0) and consequently reducing substrate availability for denitrifying bacteria. This strategy is interesting given that the addition of these inhibitors has been shown to suppress $N₂O$ emissions from urea by 90 % under Brazilian conditions [[80\]](#page-14-0). However, while DCD and DMPP exhibited similar capacities for reducing N_2O emissions, little potential for reducing $N₂O$ emissions was found for controlled-release fertilizers [[80\]](#page-14-0).

Considering the in situ measurements of leaching, denitrification, N uptake, immobilization, and volatilization losses under Brazilian field conditions that have been made available over the past several years, it is possible to develop a simplified N budget in the soil-plant system, as demonstrated in Fig. [2](#page-6-0). Such generalizations can be useful in establishing future scenarios or life-cycle assessment of sugarcane production, but care must be taken in interpreting these data; for example, the mean value of 5.6 % for leaching losses does not include losses from native soil N (Table [1](#page-4-0)). Emissions of N₂O are affected by numerous factors, such as the sources and rates of N [\[82\]](#page-14-0) and soil moisture [\[84\]](#page-14-0) and, thus, a single emission factor cannot be applied universally to all production systems. Volatilization losses can be much higher than the value reported when urea is applied over GCTB [[20](#page-12-0), [21](#page-12-0)] and can be much lower when non-amidic sources are used [\[20,](#page-12-0) [85](#page-14-0)]. Such limitations must be considered when forming broader generalizations from the estimates shown in Fig. [2.](#page-6-0)

Influence of Trash Blanket on Nitrogen Fertilization

Sugarcane cultivated in GCTB production systems adds approximately 10–20 Mg ha^{-1} (dry mass) of crop residues annually, composed mainly of dry leaves and sugarcane tops [\[6,](#page-12-0) [88](#page-14-0), [89\]](#page-14-0), depending on yield and genotype characteristics. The amount of nutrients returned to the soil through trash also varies, ranging from 39–72 kg N ha⁻¹, 4–23 kg P ha⁻¹, 35–173 kg K ha⁻¹, 9– 81 kg Ca ha⁻¹, 6-26 kg Mg ha⁻¹, and 7-15 kg S ha⁻¹ [\[90](#page-14-0)-[92](#page-14-0)]. In addition, the trash layer that accumulates over the soil surface influences sugarcane production by affecting yield, fertilizer management, weed control, soil erosion, and SOM dynamics [\[6\]](#page-12-0).

Under the field conditions that occur in Brazil, trash can be strongly reduced (~73 % depletion) as a result of decomposition by microorganisms within 3 years after its initial deposition, and the nutrients released in larger amounts are K, Ca, and N [\[89\]](#page-14-0). However, given the high C/N ratio of the trash (varying from 80 to 100), its decomposition and subsequent release of N are low in one crop season. Simulations performed by Robertson and Thorburn [[93\]](#page-14-0) using data from Australia determined that organic

Fig. 2 Fate of N from fertilizer in sugarcane-cropping soils based on measurements under Brazilian field conditions. Microbial immobilization: values gathered from Gava et al. [\[42](#page-13-0)] (37 %), Vitti [\[38](#page-13-0)] (32 %), Basanta et al. [\[41\]](#page-13-0) (29 %), and Faroni [\[40\]](#page-13-0) (29 %). Plant uptake: values gathered from Trivelin et al. [\[36](#page-13-0)] (12 %), Gava et al. [\[37\]](#page-13-0) (17 %), Vitti [\[38\]](#page-13-0) (26 %), Franco et al. [\[39\]](#page-13-0) (28 %), Faroni [[40](#page-13-0)] (37 %), and Trivelin et al. [[35\]](#page-13-0) (40 %); 3) NH₃ volatilization: values gathered from Costa et al. [\[20\]](#page-12-0) (24 %), Cantarella et al. [[85\]](#page-14-0) (8 %), and Mariano et al. [[21\]](#page-12-0) (25 %). Leaching: values gathered from Table [1](#page-4-0). N_2O emissions: values gathered from Table [2](#page-5-0). Other pathways: N derived from fertilizer not accounted in previous pathways (e.g., losses by runoff and foliar emissions of $NH₃$ and $N₂O$)

C and total N in the soil could increase by 8–15 % and 9–24 %, respectively, whereas soil mineral N could increase by 37 kg ha^{-1} y^{-1} under GCTB, taking 20–30 years for the soils to reach a new steady state. According to their results, N fertilizer application should not be reduced within the first 6 years after adoption of GCTB, but small reductions may be possible in the long term (> 15 years). In a simulation performed for southeastern Brazil, Trivelin et al. [\[94\]](#page-14-0) found that GCTB would increase soil N stocks and N recovery by sugarcane, reaching a dynamic equilibrium after 40 years, with N recovery by plants reaching ~40 kg N ha⁻¹ y⁻¹.

The effects of GCTB on sugarcane yield are complex, with research studies reporting both negative [[23,](#page-12-0) [41](#page-13-0), [95](#page-15-0)] and positive [[36](#page-13-0), [96](#page-15-0)] effects on yield. In colder regions, trash reduces shoot sprouting from ratoon crops [\[97\]](#page-15-0), which may account for some of the observed negative results. One option for avoiding decreasing yields in such regions is to remove trash mulch from crop rows but maintain it between rows, as demonstrated by Campos et al. [\[23\]](#page-12-0). In warmer climates, trash promotes higher yields by helping to conserve soil moisture and decreasing soil temperatures [[98](#page-15-0)].

The large-scale adoption of GCTB in Brazil triggered two significant changes in fertilization management. The first involved alterations in the methods and sites of application; incorporation of fertilizer midway between rows, as was typically done in the previous management system when residues were largely absent, was now hindered by the trash layers that cover the soil surface [\[99](#page-15-0)], and for this reason, producers began to adopt the practice of surface applications in ratoon. The second modification refers to a trend in increasing N rates under GCTB as compared to the previous system (manual harvest with prior burning), to avoid yield losses associated with N immobilization during trash mineralization by microorganisms.

Another recent issue that has arisen in the sugarcane industry is the increasing interest in removing the trash blankets to use in the production of electricity or second-generation ethanol [\[24\]](#page-12-0), a topic that is being widely discussed in the literature in Brazil [\[26\]](#page-12-0). Considering the advantages of GCTB for N cycling [\[94\]](#page-14-0), removing trash may directly affect the sustainability of the agroecosystem over the long term. Although trash does not effectively supply N in the first growing season, its contribution to SOM maintenance is twice that of N fertilizers and, in addition, the long-term effect of crop residues in providing mineral N to the crop is greater than that of N fertilizers [[48](#page-13-0)]. Therefore, research is required to assess the impacts of trash-blanket removal on sugarcane productivity under different climatic conditions, as well as on C and N storage in soils.

Influence of Crop Rotation on Nitrogen Fertilization

Cultivation of N-fixing legumes (sunn hemp species: Crotalaria juncea, Crotalaria spectabilis, and Crotalaria ochroleuca; peanuts: Arachis hypogaea L.; and soybean: Glycine max L.) in rotation with sugarcane is becoming a common practice, especially in the southeastern region of Brazil. The main benefits of such rotations include improving N supply [[100](#page-15-0)–[103\]](#page-15-0), weed control [[104\]](#page-15-0), reduction in the population of nematodes [\[105\]](#page-15-0), and erosion control, and increasing yields [[100](#page-15-0), [102](#page-15-0), [106\]](#page-15-0).

Legumes typically accumulate large amounts of N and K, which are essential for sugarcane. The amount of N fixed from the atmosphere by bacteria associated with legumes may vary from 30 to 200 kg N ha^{-1} [\[106](#page-15-0)–[108](#page-15-0)]. However, the amount of N in legume residues available for subsequent crops depends on the internal processes of N-cycling in the soil, such as mineralization and immobilization, as well as whether legumes are cultivated exclusively for biomass production and hence will be returned to the soil—or as grain crops such as soybeans and peanuts, which reduce the N contribution to the system owing to nutrient removal as a result of harvest. In general, the amount of N fertilizer applied to a cycle of plant cane might be reduced, or even suppressed, when rotation with N-fixing legumes is performed [\[109\]](#page-15-0). Nevertheless,

sugarcane growers normally maintain the usual N rates that are applied on ratoon crops, neglecting possible residual effects of N fixed by legumes on subsequent ratoon. Park et al. [\[110](#page-15-0)] simulated the residual effect of N fixed by soybean (in the absence of grain harvesting) during a sugarcane-growing season in Australia. They observed a potential for reduction of 100 % of N in the plant cane, and 60, 25, and 10 % in subsequent ratoon crops, respectively. The cultivation of legume crops in rotation with sugarcane certainly has the potential to reduce N rates under Brazilian conditions, based on the results of a study by Otto et al. [[49\]](#page-13-0) in which no response of ratoon to N fertilization in fields managed with legume rotation was found. However, the effects of legume crop rotation on the N requirements of subsequent crop cycles (ratoon) have yet to be examined in Brazil, even for the Crotalaria species that are being widely adopted in the country.

Improvements in Sugarcane Nitrogen Fertilization: Recent Developments in Brazil

Nitrogen Rates

There is a perception in Brazil that plant cane cycles show limited responses to N fertilization [\[111](#page-15-0)]. Recent studies have reported positive responses, but N rates that delivered high yields or net returns still range in the relatively low 40– 60 kg N ha^{-1} [\[112](#page-15-0)]. There is a clear relationship between soil tillage operations and plant cane responsiveness to N. When fields are established following conventional tillage operations (e.g., deep plowing, subsoiling, or disking), plant cane responsiveness to N is limited [[108](#page-15-0)], whereas responsiveness increases under reduced tillage practices [[113\]](#page-15-0). Such results indicate a clear relationship between conditions that enhance soil N mineralization and decrease the N requirements of sugarcane [\[114,](#page-15-0) [115](#page-15-0)], demonstrating that soil N mineralization plays a key role in supplying N to plants. Based on such findings, we expect that the usual N rates adopted for plant cane in Brazil will not be modified over the medium term.

Research undertaken in Brazil has suggested that BNF by diazotrophic bacteria might be an important source of N to sugarcane [\[116,](#page-15-0) [117\]](#page-15-0); Urquiaga et al. [\[117](#page-15-0)], for instance, estimated that sugarcane can obtain at least 40 kg N ha^{-1} through BNF. Five species of diazotrophic bacteria [[118](#page-15-0)] have been used in studies under greenhouse and field conditions, with a mix of positive and negative results. On the positive side, Schultz et al. [\[119](#page-15-0), [120\]](#page-15-0) obtained similar yields between inoculated and fertilized treatments in one site but not in another. However, Cantarella et al. [\[121](#page-15-0)] did not find positive results of inoculation in a comprehensive 13-site-years study in São Paulo. Evidence of BNF was also not observed for sugarcane in Australia [[122](#page-15-0)] or South Africa [\[123\]](#page-15-0). More recent evidence has shown that the benefits may be associated with the

production of plant-growth promoting substances rather than BNF [\[120](#page-15-0), [124](#page-15-0), [125](#page-15-0)]. These results indicate a gap between the evidence of BNF obtained in the past (usually using micropropagated sugarcane) and studies performed under field conditions. Further research is required that focuses on transforming inoculation into a practical alternative for sugarcane production and its potential for reducing N requirements.

Ratoon N fertilization has been shown to improve sugarcane yields [\[126](#page-15-0), [127](#page-16-0)] and can be effective in circumstances of limited soil disturbance, which results in lower soil N mineralization. However, the N rates adopted in Brazil are substantially lower than are the N rates adopted in other large-scale producers [\[32\]](#page-13-0). In São Paulo, for example, official recommendations for N fertilization on ratoon crops vary between 60 and 120 kg N ha^{-1} , depending on the expected yield concept [\[128\]](#page-16-0). Such recommendations were formulated from experiments conducted at sites subjected to burning (i.e., no trash maintenance) and have resulted in an empirical and widely adopted recommendation of an N fertilizer requirement of 1.0 kg N per Mg of stalk.

Currently, ~80 % of the area under sugarcane cultivation in Brazil is harvested under GCTB systems [\[17](#page-12-0), [129](#page-16-0)]. These systems promote alterations in soil C and N cycling, which may require further adjustment in the N recommendation system for sugarcane. On the one hand, the amount of trash maintained overtop of the soil can promote immobilization of N fertilizer during trash decomposition and may require an increase in the recommended N rates for burned sugarcane. On the other hand, the release of N from trash along with increases in soil C and N stocks over time may serve to reduce demand for N fertilization in GCTB systems. In the short term, no reduction in N rates are expected [\[93](#page-14-0), [94](#page-14-0)], whereas in the medium- to long-term, gradual release of trash N may enhance soil N availability to sugarcane [[89,](#page-14-0) [130](#page-16-0)–[132\]](#page-16-0).

It is not yet clear whether the GCTB system will require increases or reductions in N rates as compared to the previous burning system, but growers have shifted to an increase in N rates to avoid yield losses due to N deficiencies. The official recommendation of 1.0 kg N per Mg of stalk produced has been empirically replaced by the 1.2 kg N per Mg factor adopted by the majority of growers in southeastern Brazil. This approach is similar to the 1.2 factor used in the Six Easy Steps nutrient management program [\[133](#page-16-0)], and higher than the 1.0 factor used in the N Replacement System [\[127\]](#page-16-0), both of which were developed for application under Australian conditions.

Aiming to elucidate the need for increased N rates adopted in GCTB systems, we surveyed 45 field trials carried out under the GCTB system over the past several decades in Brazil. We recorded the results of check-plot yields, and calculated the yield increase to N fertilization in each single trial (Table [3](#page-8-0)), and ranked the sites as non-responsive to N fertilization (no significant effect of N fertilization in the original study), moderately responsive (from 0 to 25 % yield increase),

Table 3 Survey of N-response curve trials under the green cane trash blanketing system in Brazil

^a Check plot yield, stalk yield obtained in the control treatment (no-N applied)

 b Calculated as ((high yielding treatment—check plot yield)/check plot yield x 100)</sup>

^c Non-responsive (yield increases =0); moderate (yield increases between 0–25 %); high (yield increases >25 %)

and highly responsive $(> 25\%$ vield increase). An N requirement adjustment to sugarcane production in Brazil would be expected if most sites showed high levels of response to N fertilization after being shifted from burning to a GCTB system. However, as the results presented in Fig. 3 show, 75 % of the sites were non-responsive or moderately responsive to N fertilization, whereas only 25 % of the sites were highly responsive to N fertilization. These results suggest that increasing N fertilization will probably not result in yield increases in most cases, and that the change from 1.0 to 1.2 kg N per Mg of stalk produced requires further revision.

More interestingly, plotting data of N responsiveness against the check-plot yield for each site revealed a linear negative correlation ($r = -0.49$; $P < 0.001$), indicating a clear trend toward reduction in N responsiveness as check-plot yield increases (Fig. 3). The high check-plot yield indicates a high availability of mineral N or easily mineralizable organic N fractions in the soil. This is expected, given the gradual release of N in GCTB systems [\[94](#page-14-0)], the optimization of press mud and vinasse usage by growers, and the expansion of areas using legume rotation. Such management approaches have the potential to increase soil N availability and reduce sugarcane response to N, as demonstrated by Otto et al. [[49](#page-13-0)]. In contrast, low check-plot yields are indicative of plants growing in conditions of low soil N availability (limited N conditions) and, thus, exhibit significant responses to N additions. Indeed, the majority of sugarcane grown in Brazil is not subjected to limited N conditions (considering the byproducts usage, legume rotation, and usual practices of N fertilization) and, thus, it is expected that moderate responses to N fertilization of sugarcane will remain in the future.

Even with the low N rates adopted for sugarcane ratoon in Brazil as compared to other countries, lower profitability for growers may result from the N fertilization practice in situations where there is no substantial return on the investment and due to costs associated with overuse of N fertilizer, which

Fig. 3 Yield increases due to N fertilization (a) and relationship between check plot yield and yield response (b) as obtained in 45 N-response curve trials carried out under GCTB systems in Brazil. In each single trial reported on Table [3](#page-8-0), check plot yield represents the control treatment (no-N applied); increase in yield due to N fertilization was obtained as [(high yielding treatment – check plot yield) check plot yield \times 100]; yield response was further categorized as non-responsive (yield increases =0), moderately responsive (yield increases between 0–25 %), and highly responsive (yield increases >25 %). Dotted line in panel A represents the boundary line (yield increase =25 %) between moderate and highly responsive sites. Dotted lines in panel B indicates the curve fitting to a linear model for check plot yield and yield increase data

may also result in significant environmental impacts [\[7](#page-12-0)]. Regarding leaching losses, for example, whereas Ghiberto et al. [[67,](#page-14-0) [68\]](#page-14-0) reported limited N losses in the plant cane, N leaching in the ratoon crop cycle represented 22.5 % of the total N applied as fertilizer (100 kg N ha⁻¹) [[69\]](#page-14-0). Higher leaching losses are expected to occur in ratoons given the proximity between the period of fertilizer application and the rainy season. This is of particular concern considering that $NO₃⁻$ leaching is usually highest at the onset of the wet season, when both water drainage and NO_3 ⁻ are present simultaneously [\[68](#page-14-0), [70](#page-14-0)]. Moreover, higher N rates also have the potential to increase N_2O emissions [\[82](#page-14-0)].

Fertilizer Nitrogen Sources

Urea is widely used as a fertilizer; in Brazil, it represents 66 % of N fertilizer consumption [\[142](#page-16-0)], despite the high potential for losses through volatilization [[85\]](#page-14-0). However, reconciling the trend toward increasing use of urea and the large-scale adoption of GCTB systems with surface application of fertilizers presents a challenge, as surface-applied urea on trash blankets may result in increased $NH₃$ volatilization losses ranging between 24 to 37 % of applied N [[21,](#page-12-0) [85](#page-14-0), [86](#page-14-0)].

One possible strategy for reducing $NH₃$ losses from surface-applied urea is through the use of urease inhibitors. Under field conditions, Cantarella et al. [[85](#page-14-0)] observed that NBPT [N-(N-butyl) thiophosphoric triamide]-treated urea reduced N losses via volatilization by 15–78 % when compared with urea-only losses. Faria et al. [\[87\]](#page-14-0), in an assessment of Band Cu-coated urea as well as S-coated urea as additional alternatives to reduce the volatilization process, found that although the coatings delayed loss peaks, accumulated N losses were similar to those reported for conventional granulated urea. Such results and concerns regarding the effectiveness of urea inhibitors in systems with large amounts of trash overtop the soil surface (such as sugarcane) is a limiting factor for their large-scale adoption by sugarcane growers in Brazil. For this reason, sources less subject to volatilization losses (i.e., ammonium nitrate and ammonium sulfate) are preferred to urea for superficial application, especially in dry periods. Adjustments in the quantity of NBPT to be added to urea under high-trash conditions, or improvements in the inhibitors themselves, are required before widespread use of urea-based fertilizer for sugarcane production in Brazil can be initiated.

Nitrification inhibitors can also be added to urea- or ammonium-based fertilizers in order to delay microbial oxidation of NH_4^+ to NO_3^- . There are three main reasons to expect positive effects of using nitrification inhibitors in sugarcane cropped soils: (1) $NO₃⁻$ is subjected to leaching in sugarcane fields in some circumstances $[68]$; (2) $NO₃⁻$ is subjected to denitrification losses, especially under GCTB and high-moisture conditions [[84\]](#page-14-0); and (3) the $NO₃⁻$ form is generally the most prevalent in sugarcane-cropped soils, and

sugarcane has been shown to preferentially absorb $\mathrm{NH_4}^+$ rather than $NO₃⁻$ [[32\]](#page-13-0). DCD has been the most commonly used product to inhibit nitrification [\[143,](#page-16-0) [144\]](#page-16-0), despite the availability of alternative products, such as DMPP. Soares et al. [[80\]](#page-14-0), for instance, reported that DMPP- or DCD-treated urea reduced N_2O losses from urea by 90 %. However, adopting urease and nitrification inhibitors simultaneously may lead to increased volatilization losses due to the retention of high NH4 ⁺ concentrations in the soil [[86\]](#page-14-0) and, therefore, should be avoided. Research evaluating the effects of nitrificationinhibitor treated urea in sugarcane systems is needed.

The use of ammonium chloride is not common in Brazil, and some studies have shown limitations in its effectiveness as a fertilizer for sugarcane. Similar levels of efficiency for ammonium chloride, ammonium nitrate, ammonium sulfate, and urea in the first year of application to sugarcane have been reported [[137\]](#page-16-0), but assessment of the residual effects of fertilizers on subsequent cycles suggested that ammonium chloride linearly reduced crop yield when compared to other N sources, most likely due to the cumulative salt effect of chloride ions [\[145](#page-16-0)]. Similarly, Mariano et al. [\[44](#page-13-0)] reported a low availability of mineral N in soils and a lower N content in sugarcane following ammonium chloride application.

Slow- or controlled-release fertilizers (also known as "enhanced efficiency fertilizers") were developed that supply nutrients in a more gradual manner to better match the variations in plant nutritional demands. Organic or synthetic compounds are used as coating materials, but these types of fertilizers generally have positive or no effects when compared to conventional N sources [[34](#page-13-0)]. In spite of favorable results in other crops [\[146\]](#page-16-0), no studies were found regarding the effectiveness of such fertilizers in sugarcane systems in Brazil.

Application Methods

Fertilizer application methods require further evaluation, as most experiments involving sugarcane are restricted to comparisons of fertilizer sources or application rates [[147](#page-16-0)]. Due to growing concerns about GHG emissions, several studies have been carried out to examine how N application methods may influence N_2O emissions [[148](#page-16-0)]. Incorporated application of N fertilizer to soils at depths of 5 cm might reduce N_2O emis-sions, especially in humid climates [\[149\]](#page-16-0). Lower N_2O emissions were also observed when fertilizer granules were placed 10-cm deep, as deeper applications increase the residence time of N_2O in soil and, thus, lower losses to the atmosphere [\[150\]](#page-16-0). Incorporation of N fertilizers into the soil also has the potential to eliminate NH_3 volatilization losses from urea [[45,](#page-13-0) [151](#page-16-0)] which is of particular interest given that reducing volatilization losses has the additional benefit of increasing NUE [\[152,](#page-16-0) [153\]](#page-16-0).

In the burning systems used in the past, fertilizers were normally incorporated into the soil during cultivation.

Cultivation was typically performed using a deep tillage cultivator that breaks up the soil between crop rows [\[154\]](#page-16-0), making it possible to incorporate fertilizer in the middle-row. However, incorporation of fertilizer in crops under GCTB systems is more difficult [[99\]](#page-15-0), and it requires specialized equipment and climatic conditions to ensure successful operations. Moreover, incorporation of fertilizer has been shown to result in similar or even lower yields compared to single-side surface-banding application [\[155](#page-16-0), [156\]](#page-16-0).

In terms of fertilizer placement, Bianchini et al. [[19](#page-12-0)] observed that banded application resulted in higher yields than did broadcast application, whereas Vitti et al. [\[151\]](#page-16-0) found no differences between the two in terms of yield. These results suggest that, so long as N fertilizers are not subject to $NH₃$ volatilization losses, there is little need for their incorporation in areas under GCTB, and that localized application (usually performed 20-cm distant from the crop row) has advantages because roots are concentrated in the superficial soil layer (0– 20 cm) and adjacent to the sugarcane row [\[157](#page-16-0), [158](#page-16-0)]. However, development of the specialized equipment that would allow for incorporated application next to the crop row has the potential to maintain sugarcane yield (due to proximity to the root system) and permits the use of ureabased fertilizers by mitigating NH_3 and N_2O emissions.

Timing of Fertilizer Application

In plant cane, N fertilization is usually applied at the bottom of the furrow, at depths of 0.15–0.30 m. Consequently, N losses caused by volatilization are insignificant even when urea is applied [[151](#page-16-0)]. Despite official recommendations for split application of N on plant cane [\[128\]](#page-16-0), in most cases N is applied once at planting due to uncertainties regarding yield gains when using a split application approach [[159](#page-16-0)]. However, in some cases, growers are beginning to split N application following K application coupled with land-leveling (performed to facilitate mechanized harvesting), which is carried out at 90–120 days following planting. The results of split N application in such operations are still unclear and must be investigated.

There is asynchrony between the time of fertilizer application and crop nutritional demand in the ratoon crop cycle. In southeastern Brazil, sugarcane harvest occurs from April to November, and fertilizers are usually applied right after harvesting, but \sim 75 % of the total biomass and N content accumulated by sugarcane in the ratoon cycle occurs between December and March [\[45,](#page-13-0) [46,](#page-13-0) [160\]](#page-17-0). Thus, further research examining the viability of split application of N in the ratoon crop cycle is needed, as the evidence for the benefits of split application derive from studies of fertigated sugarcane, in which N is applied according to crop demand, resulting in high responsiveness to N [\[161](#page-17-0), [162\]](#page-17-0). The use of slow- or controlled-release fertilizers may serve a similar purpose.

The development of specialized equipment enabling variable rate application creates new opportunities for the timing of fertilizer application using information obtained from canopy sensors [\[163\]](#page-17-0). The use of close remote sensing (optical sensors attached to agricultural machinery) assumes that leaf spectral properties, such as reflectance and transmittance, are affected by N deficiency [[164](#page-17-0)]. The normalized difference vegetation index (NDVI) is commonly used to estimate N nutrition, yield, and chlorophyll content in leaves [\[165](#page-17-0)–[167\]](#page-17-0). Depending on the methodology adopted, the use of NDVI facilitates the development of recommendations for variablerate application of fertilizers, in either real-time or by following recommendation maps [\[168](#page-17-0)]. However, there are few studies that have assessed the effectiveness of using optical sensors (such as NVDI) on variable-rate N application for sugarcane and, thus, further research on calibrating vegetation indices with crop yields is required [\[169\]](#page-17-0) prior to large-scale adoption of variable-rate technology in sugarcane fields.

Concluding Remarks

Moderate N rates are generally used for sugarcane production in Brazil when compared to other major sugarcane-producing countries. Volatilization is one of the primary N-loss pathways in Brazilian sugarcane systems, but concerns about leaching loss and $N₂O$ emissions that result from biofuel production are rising. Here, we found that approximately 60 % of the fertilizer N is recovered by plants and soils throughout the crop cycle, while leaching losses and $N₂O$ emissions may reach as high as 5.6 and 1.84 % of the applied N, respectively.

The increasing shift from burning to non-burning (GCTB) systems in Brazil has led to modifications in the country's Nmanagement strategies, with an increase in N rates from 1.0 kg N per Mg of stalk to 1.2 kg N per Mg of stalk being adopted by most growers. However, our review demonstrates that 75 % of the sites examined were non-responsive or only moderately responsive to N, indicating that increases in N rates are unlikely to promote higher yields, further reducing NUE. More interestingly, high-yielding systems exhibited very limited response to N fertilization, whereas lowyielding systems were highly responsive. Such findings indicate that sugarcane growing under conditions of an adequate supply of N require lower N rates.

This review identified strategies that can be adopted to enhance NUE for sugarcane-biofuel production. Such strategies include (i) reduction in N-fertilizer usage by most largescale sugarcane producers in order to optimize plant uptake and reduce losses. Savings in N fertilizers without reduction on yield can be ensured by optimizing byproduct usage, growth of legume crops in rotation, and trash maintenance in sugarcane fields; (ii) adoption of best-management practices, including strategies to maintain N as NH_4^+ in soils, as well as

adopting fertilizer and application methods that minimize losses; (iii) developing new N-recommendation systems that do not exclusively focus on the expected yield concept; (iv) developing second-generation biofuel production that increases ethanol production per unit area without requiring additional N fertilizer; and (v) evaluating N-internal use efficiency in breeding programs in order to propagate genotypes with improved NUE. All of these issues must be addressed in Brazil and other large-scale producing countries to deliver high sugarcane yields with minimum N usage and reduced environmental impacts.

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References

- 1. Pimentel D, Patzek T (2007) Ethanol production: energy and economic issues related to U.S. and Brazilian sugarcane. Nat Resour Res 16:235–242. doi[:10.1007/s11053-007-9049-2](http://dx.doi.org/10.1007/s11053-007-9049-2)
- 2. Goldemberg J (2007) Ethanol for a sustainable energy future. Science 315:808–810. doi:[10.1126/science.1137013](http://dx.doi.org/10.1126/science.1137013)
- 3. Renouf MA, Wegener MK, Nielsen LK (2008) An environmental life cycle assessment comparing Australian sugarcane with US corn and UK sugar beet as producers of sugars for fermentation. Biomass Bioenergy 32:1144–1155. doi[:10.1016/j.biombioe.2008.](http://dx.doi.org/10.1016/j.biombioe.2008.02.012) [02.012](http://dx.doi.org/10.1016/j.biombioe.2008.02.012)
- 4. Smeets E, Jungiger M, Faaij A, Walter A, Dolsan P, Turkemburg W (2008) The sustainability of Brazilian ethanol—an assessment of the possibilities of certified production. Biomass Bioenergy 16: 192–123. doi[:10.1016/j.biombioe.2008.01.005](http://dx.doi.org/10.1016/j.biombioe.2008.01.005)
- 5. Nass LL, Pereira PAA, Ellis D (2007) Biofuels in Brazil: an overview. Crop Sci 47:2228–2237. doi[:10.2135/cropsci2007.03.0166](http://dx.doi.org/10.2135/cropsci2007.03.0166)
- 6. Leal MRLV, Galdos MV, Scarpare FV, Seabra JEA, Walter A, Oliveira COF (2013) Sugarcane straw availability, quality, recovery and energy use: a literature review. Biomass Bioenergy 53:11– 19. doi:[10.1016/j.biombioe.2013.03.007](http://dx.doi.org/10.1016/j.biombioe.2013.03.007)
- 7. Erisman JW, Van Grinsven H, Leip A, Mosier A, Bleeker A (2010) Nitrogen and biofuels; an overview of the current state of knowledge. Nutr Cycl Agroecosyst 86:211–223. doi[:10.1007/](http://dx.doi.org/10.1007/s10705-009-9285-4) [s10705-009-9285-4](http://dx.doi.org/10.1007/s10705-009-9285-4)
- 8. Crutzen PJ, Mosier AR, Smith KA, Winiwarter W (2008) N₂O release from agrobiofuel production negates global warming reduction by replacing fossil fuels. Atmos Chem Phys 8:389–395. doi[:10.5194/acp-8-389-2008](http://dx.doi.org/10.5194/acp-8-389-2008)
- 9. Smeets E, Bouwman L, Stehfest E, Van Vuuren DP, Posthuma A (2009) Contribution of N_2O to the green-house gas balance of first-generation biofuels. Glob Chang Biol 15:1–23. doi:[10.1111/](http://dx.doi.org/10.1111/j.1365-2486.2008.01704.x) [j.1365-2486.2008.01704.x](http://dx.doi.org/10.1111/j.1365-2486.2008.01704.x)
- 10. Neves MF, Trombin VG, Kalaki RB, Gerbasi T, Rodrigues JM, Canto F, Simprini ES, Rovanhol P, Consoli MH (2014) A dimensão do setor sucroenergético: mapeamento e quantificação da safra 2013/2014. Markstrat, Fundace, FEA-RP/USP, Ribeirão Preto 45p
- 11. CGEE (2009) Centro de Gestão e Estudos Estratégicos. Bioetanol combustível: uma oportunidade para o Brasil. [http://www.cgee.](http://www.cgee.org.br/publicacoes/bietanol.php) [org.br/publicacoes/bietanol.php](http://www.cgee.org.br/publicacoes/bietanol.php). Accessed 2 Feb 2016
- 12. Unica (2014) União da agroindústria canavieira. [http://www.](http://www.unica.com.br) [unica.com.br.](http://www.unica.com.br) Accessed 24 April 2015
- 13. OECD (2015) Food and agriculture organization of the United Nations. OECD-FAO agricultural outlook 2015. OECD Publishing, Paris. doi[:10.1787/agr_outlook-2015-en](http://dx.doi.org/10.1787/agr_outlook-2015-en) Accessed 6 January 2016
- 14. Moraes M, Costa C, Guilhoto J, Souza L, Oliveira F (2010) Social externalities of fuels. In: Souza ELL, Macedo IC (eds) Ethanol and bioeletricity: sugarcane in the future of energy matrix. UNICA, Sao Paulo, pp. 44–75
- 15. FAOSTAT (2015) Food and agriculture organization of the United Nations. Statistics Division. <http://faostat3.fao.org/home/E>. Accessed 18 April 2015
- 16. CONAB (2015) Companhia nacional de abastecimento. Acompanhamento da safra brasileira. Cana-de-açúcar. Safra 2015–16. Segundo levantamento. Agosto de 2015. Monitoramento Agrícola. [http://www.conab.gov.br/OlalaCMS/](http://www.conab.gov.br/OlalaCMS/uploads/arquivos/15_08_13_15_58_44_boletim_cana_portugues_-_2o_lev_-_15-16.pdf) [uploads/arquivos/15_08_13_15_58_44_boletim_cana_](http://www.conab.gov.br/OlalaCMS/uploads/arquivos/15_08_13_15_58_44_boletim_cana_portugues_-_2o_lev_-_15-16.pdf) [portugues_-_2o_lev_-_15-16.pdf.](http://www.conab.gov.br/OlalaCMS/uploads/arquivos/15_08_13_15_58_44_boletim_cana_portugues_-_2o_lev_-_15-16.pdf) Accessed 9 Sept 2015
- 17. Protocolo Agroambiental do Setor Sucroenergético (2014) Relatório consolidado 2007/08–2013/14. [http://www.ambiente.](http://www.ambiente.sp.gov.br/etanolverde/files/2015/02/Protocolo-Agroambiental-do-Setor-Sucroenerg%C3%A9tico-Relat%C3%B3rio-consolidado-RV.pdf) [sp.gov.br/etanolverde/files/2015/02/Protocolo-Agroambiental](http://www.ambiente.sp.gov.br/etanolverde/files/2015/02/Protocolo-Agroambiental-do-Setor-Sucroenerg%C3%A9tico-Relat%C3%B3rio-consolidado-RV.pdf)[do-Setor-Sucroenerg%C3%A9tico-Relat%C3%B3rio](http://www.ambiente.sp.gov.br/etanolverde/files/2015/02/Protocolo-Agroambiental-do-Setor-Sucroenerg%C3%A9tico-Relat%C3%B3rio-consolidado-RV.pdf)[consolidado-RV.pdf](http://www.ambiente.sp.gov.br/etanolverde/files/2015/02/Protocolo-Agroambiental-do-Setor-Sucroenerg%C3%A9tico-Relat%C3%B3rio-consolidado-RV.pdf). Accessed 18 July 2015
- 18. Cerri CEP, Galdos MV, Carvalho JLN, Feigl B, Cerri CC (2013) Quantifying soil carbon stocks and greenhouse gas fluxes in the sugarcane agrosystem: point of view. Sci Agric 70:361–368. doi: [10.1590/S0103-90162013000500011](http://dx.doi.org/10.1590/S0103-90162013000500011)
- 19. Bianchini A, Valadão Junior DD, Rosa RP, Colhado F, Daros RF (2014) Soil chiseling and fertilizer location in sugarcane ratoon cultivation. Eng Agric 34:57–65. doi:[10.1590/S0100-](http://dx.doi.org/10.1590/S0100-69162014000100007) [69162014000100007](http://dx.doi.org/10.1590/S0100-69162014000100007)
- 20. Costa MCG, Vitti GC, Cantarella H (2003) Volatilização de N-NH3 de fontes nitrogenadas em cana-de-açúcar colhida sem despalha a fogo. R Bras Ci Solo 27:631–637. doi[:10.1590/](http://dx.doi.org/10.1590/S0100-06832003000400007) [S0100-06832003000400007](http://dx.doi.org/10.1590/S0100-06832003000400007)
- 21. Mariano E, Trivelin PCO, Vieira MX, Leite JM, Otto R, Franco HCJ (2012) Ammonia losses estimated by an open collector from urea applied to sugarcane straw. R Bras Ci Solo 36:411–419. doi[:10.1590/S0100-06832012000200010](http://dx.doi.org/10.1590/S0100-06832012000200010)
- 22. Dinardo-Miranda LL, Fracasso JL (2013) Sugarcane straw and the populations of pests and nematodes. Sci Agric 70:305–310. doi[:10.1590/S0103-90162013000500012](http://dx.doi.org/10.1590/S0103-90162013000500012)
- 23. Campos LHF, Carvalho SJP, Christoffoleti PJ, Fortes C, Silva JS (2010) Sistemas de manejo da palhada influenciam acúmulo de biomassa e produtividade da cana-de-açúcar (var. RB855453). Acta Sci-Agron 32:345–350. doi[:10.4025/actasciagron.v32i2.](http://dx.doi.org/10.4025/actasciagron.v32i2.3703) [3703](http://dx.doi.org/10.4025/actasciagron.v32i2.3703)
- 24. Sordi RA, Manechini C (2013) Utilization of trash: a view from the agronomic and industrial perspective. Sci Agric 70:1–2. doi[:10.1590/S0103-90162013000500002](http://dx.doi.org/10.1590/S0103-90162013000500002)
- 25. Cantarella H, Cerri CEP, Carvalho JLN, Magalhães PSG (2013) How much sugarcane trash should be left on the soil? Sci Agric 70:1–2. doi[:10.1590/S0103-90162013000500001](http://dx.doi.org/10.1590/S0103-90162013000500001)
- 26. Jansson SL, Persson J (1982) Mineralization and immobilization of soil nitrogen. In: Stevenson FJ (ed) Nitrogen in agricultural soils. American Society of Agronomy, Madison, pp. 229–252
- 27. Schimel JP, Bennett J (2004) Nitrogen mineralization: challenges of a changing paradigm. Ecology 85:591–602. doi[:10.1890/03-](http://dx.doi.org/10.1890/03-8002) [8002](http://dx.doi.org/10.1890/03-8002)
- 28. Jones DL, John RH, Willet VB, Farrar JF, Hodge A (2005) Dissolved organic nitrogen uptake by plants—an important N uptake pathway? Soil Biol Biochem 37:413–423. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.soilbio.2004.08.008) [soilbio.2004.08.008](http://dx.doi.org/10.1016/j.soilbio.2004.08.008)
- 29. Kuzyakov Y, Xu X (2013) Competition between roots and microorganisms for nitrogen: mechanisms and ecological relevance. New Phytol 198:656–669. doi:[10.1111/nph.12235](http://dx.doi.org/10.1111/nph.12235)
- 30. Vinall K, Schmidt S, Brackin R, Lakshmanan P, Robinson N (2012) Amino acids are a nitrogen source for sugarcane. Funct Plant Biol 39:503–511. doi[:10.1071/FP12042](http://dx.doi.org/10.1071/FP12042)
- 31. De Armas R, Valadier MH, Champigny ML, Lamaze T (1992) Influence of ammonium and nitrate on the growth and photosynthesis of sugarcane. J Plant Physiol 140:531–535. doi[:10.1016/](http://dx.doi.org/10.1016/S0176-1617(11)80783-2) [S0176-1617\(11\)80783-2](http://dx.doi.org/10.1016/S0176-1617(11)80783-2)
- 32. Robinson N, Brackin R, Soper KVF, Gamage JHH, Paungfoo-Lonhienne C, Rennenberg H, Lakshmanan P, Schmidt S (2011) Nitrate paradigm does not hold up for sugarcane. PLoS One 6: e19045. doi:[10.1371/journal.pone.0019045](http://dx.doi.org/10.1371/journal.pone.0019045)
- 33. D'Andréa MS (2014) Técnica da diluição do isótopo ¹⁵N para determinação da amonificação e nitrificação brutas de N em solos cultivados com cana-de-açúcar e braquiária. Thesis, University of São Paulo
- 34. Chien SH, Prochnow LI, Cantarella H (2009) Recent developments of fertilizer production and use to improve nutrient efficiency and minimize environmental impacts. Adv Agron 102:267– 322. doi:[10.1016/S0065-2113\(09\)01008-6](http://dx.doi.org/10.1016/S0065-2113(09)01008-6)
- 35. Trivelin PCO, Victoria RL, Rodrigues JCS (1995) Aproveitamento por soqueira de cana-de-açúcar de final de safra do nitrogênio da aquamônia-15 N e ureia-15 N aplicado ao solo em complemento à vinhaça. Pesq Agrop Brasileira 30:1375– 1385
- 36. Trivelin PCO, Oliveira MW, Vitti AC, Gava GJC, Bendassolli JA (2002) Nitrogen losses of applied urea in the soil-plant system during two sugarcane cycles. Pesq Agrop Brasileira 37:193–201. doi[:10.1590/S0100-204X2002000200011](http://dx.doi.org/10.1590/S0100-204X2002000200011)
- 37. Gava GJC, Trivelin PCO, Vitti AC, Oliveira MW (2003) Recuperação do nitrogênio (15 N) da ureia e da palhada por soqueira de cana-de-açúcar (Saccharum spp.). R Bras Ci Solo 27:621–630. doi:[10.1590/S0100-06832003000400006](http://dx.doi.org/10.1590/S0100-06832003000400006)
- 38. Vitti AC (2003) Adubação nitrogenada da cana-de-açúcar (soqueira) colhida mecanicamente sem a queima prévia: manejo e efeito na produtividade. Thesis, University of São Paulo
- 39. Franco HCJ, Trivelin PCO, Faroni CE, Vitti AC, Otto R (2008) Aproveitamento pela cana-de-açúcar da adubação nitrogenada de plantio. R Bras Ci Solo 32:2763–2770. doi[:10.1590/S0100-](http://dx.doi.org/10.1590/S0100-06832008000700021) [06832008000700021](http://dx.doi.org/10.1590/S0100-06832008000700021)
- 40. Faroni CE (2008) Eficiência agronômica das adubações nitrogenadas de plantio e após o primeiro corte avaliada na primeira soqueira de cana-de-açúcar. Thesis, University of São Paulo
- 41. Basanta MV, Dourado Neto D, Reichardt K, Bacchi OOS, Oilveira JCM, Trivelin PCO, Timm LC, Tominaga TT, Correchel V, Cássaro FAM, Pires LF, Macedo JR (2003) Management effects on nitrogen recovery um a sugarcane crop grown in Brazil. Geoderma 116:235–248. doi[:10.1016/S0016-](http://dx.doi.org/10.1016/S0016-7061(03)00103-4) [7061\(03\)00103-4](http://dx.doi.org/10.1016/S0016-7061(03)00103-4)
- 42. Gava GJC, Trivelin PCO, Vitti AC, Oliveira MW (2002) Balanço do nitrogênio da ureia (15 N) no sistema solo-cana-de-açúcar (cana-soca). In: Congresso Nacional dos Técnicos Açucareiros e Alcooleiros do Brasil, 8. STAB, Recife p 245–251
- 43. Farquhar GD, Firth PM, Wetselaar R, Weir B (1980) On the gaseous exchange of ammonia between leaves and the environment: determination of the ammonia compensation point. Plant Physiol 66:710–714. doi:[10.1104/pp.66.4.710](http://dx.doi.org/10.1104/pp.66.4.710)
- 44. Mariano E, Leite JM, Megda MXV, Torres-Dorante L, Trivelin PCO (2015) Influence of nitrogen form supply on soil mineral nitrogen dynamics, nitrogen uptake, and productivity of sugarcane. Agron J 107:641–650. doi:[10.2134/agronj14.0422](http://dx.doi.org/10.2134/agronj14.0422)
- 45. Gava GJC, Trivelin PCO, Oliveira MW, Penatti CP (2001) Crescimento e acúmulo de nitrogênio em cana-de-açúcar cultivada em solo coberto com palhada. Pesq Agrop Brasileira 36:347–1354. doi:[10.1590/S0100-204X2001001100004](http://dx.doi.org/10.1590/S0100-204X2001001100004)
- 46. Franco HCJ, Otto R, Faroni CE, Vitti AC, Oliveira ECA, Trivelin PCO (2011) Nitrogen in sugarcane derived from fertilizer under Brazilian field conditions. Field Crop Res 121:29–41. doi:[10.](http://dx.doi.org/10.1016/j.fcr.2010.11.011) [1016/j.fcr.2010.11.011](http://dx.doi.org/10.1016/j.fcr.2010.11.011)
- 47. Vieira-Megda MX, Mariano E, Leite JM, Franco HCJ, Vitti AC, Megda MM, Khan SA, Mulvaney RL, Trivelin PCO (2015) Contribution of fertilizer nitrogen to the total nitrogen extracted by sugarcane under Brazilian field conditions. Nutr Cycl Agroecosyst 101:241–257. doi[:10.1007/s10705-015-9676-7](http://dx.doi.org/10.1007/s10705-015-9676-7)
- 48. Dourado Neto D, Powlson D, Abu Bakar R, Bacchi OOS, Basanta MV, Thi Cong P, Keerthisinghe S, Ismaili M, Rahman SM, Reichardt K, Safwat MSA, Sangakkara R, Timm LC, Wang JY, Zagal E, Van Kessel C (2010) Multiseason recoveries of organic and inorganic nitrogen-15 in tropical cropping systems. Soil Sci Soc Am J 74:139–152. doi:[10.2136/sssaj2009.0192](http://dx.doi.org/10.2136/sssaj2009.0192)
- 49. Otto R, Mulvaney RL, Khan SA, Trivelin PCO (2013) Quantifying soil nitrogen mineralization to improve fertilizer nitrogen management of sugarcane. Biol Fertil Soils 49:893–904. doi:[10.1007/s00374-013-0787-5](http://dx.doi.org/10.1007/s00374-013-0787-5)
- 50. Ogle SM, McCarl BA, Baker J, Del Grosso SJ, Adler PR, Paustian K, Parton WJ (2015) Managing the nitrogen cycle to reduce greenhouse gas emissions from crop production and biofuel expansion. Mitig Adapt Strateg Glob Chang:1–16. doi[:10.1007/s11027-015-](http://dx.doi.org/10.1007/s11027-015-9645-0) [9645-0](http://dx.doi.org/10.1007/s11027-015-9645-0)
- 51. Robinson N, Fletcher A, Whan A, Critchley C, Wirén N, Von Lakshmanan P, et al. (2007) Sugarcane genotypes differ in internal nitrogen use efficiency. Funct Plant Biol 34:1122–1129. doi[:10.](http://dx.doi.org/10.1071/FP07183) [1071/FP07183](http://dx.doi.org/10.1071/FP07183)
- 52. Whan A, Robinson N, Lakshmanan P, Schmidt S, Aitken K (2010) A quantitative genetics approach to nitrogen use efficiency in sugarcane. Funct Plant Biol 37:448–454. doi[:10.1071/FP09260](http://dx.doi.org/10.1071/FP09260)
- 53. Robinson N, Fletcher A, Whan A, Vinall K, Brackin R, Lakshmanan P, Schmidt S (2008) Sustainable sugarcane production systems: reducing plant nitrogen demand. Proc Aust Soc Sugar Cane Technol 30:212–219
- Arruda P (2012) Genetically modified sugarcane for bioenergy generation. Curr Opin Biotechnol 23:315–322. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.copbio.2011.10.012) [copbio.2011.10.012](http://dx.doi.org/10.1016/j.copbio.2011.10.012)
- 55. Skocaj DM, Y E, Schroeder BL (2013) Nitrogen management guidelines for sugarcane production in Australia: can these be modified for wet tropical conditions using seasonal climate forecasting? Springer Sci Rev 1:51–71. doi:[10.1007/s40362-013-](http://dx.doi.org/10.1007/s40362-013-0004-9) [0004-9](http://dx.doi.org/10.1007/s40362-013-0004-9)
- 56. Thorburn PJ, Jakku E, Webster AJ, Everingham YL (2011a) Agricultural decision support systems facilitating co-learning: a case study on environmental impacts of sugarcane production. Int J Agric Sustain 9:1–12. doi:[10.1080/14735903.2011.582359](http://dx.doi.org/10.1080/14735903.2011.582359)
- 57. Wood AW, Schroeder BL, Dwyer R (2010) Opportunities for improving the efficiency of use of nitrogen fertiliser in the Australian sugar industry. Proc Aust Soc Sugar Cane Technol 32:221–233
- 58. Schroder JL, Zhang H, Girma K, Raun WR, Penn CJ, Payton ME (2011) Soil acidification from long-term use of nitrogen fertilizers on winter wheat. Soil Sci Soc Am J 75:957–964. doi[:10.2136/](http://dx.doi.org/10.2136/sssaj2010.0187) [sssaj2010.0187](http://dx.doi.org/10.2136/sssaj2010.0187)
- 59. Brady NC, Weil RR (2007) The nature and properties of soils, 14th edn. Prentice Hall, Pearson, New York
- 60. Crusciol CAC, Foltran R, Rossato OB, Mccray JM, Rossetto R (2014) Effects of surface application of calcium-magnesium silicate and gypsum on soil fertility and sugarcane yield. R Bras Ci Solo 38:1843–1854. doi[:10.1590/S0100-06832014000600019](http://dx.doi.org/10.1590/S0100-06832014000600019)
- 61. Chen D, Suter H, Islam A, Edis R, Freney JR, Walker CN (2008) Prospects of improving efficiency of fertiliser nitrogen in Australian agriculture: a review of enhanced efficiency fertilisers. Aust J Soil Res 46:289–301. doi[:10.1071/SR07197](http://dx.doi.org/10.1071/SR07197)
- 62. Kingston G (2000) Climate and the management of sugarcane. In: Hogarth DM, Allsopp PG (eds) Manual of canegrowing. Bureau of Sugar Experiment Stations, Brisbane, pp. 7–25
- 63. Waterhouse J, Brodie J, Lewis S, Mitchell A (2012) Quantifying the sources of pollutants in the great barrier reef catchments and the relative risk to reef ecosystems. Mar Pollut Bull 65:394–406. doi[:10.1016/j.marpolbul.2011.09.031](http://dx.doi.org/10.1016/j.marpolbul.2011.09.031)
- 64. Prado H (2005) Ambientes de produção de cana-de-açúcar na região Centro-Sul do Brasil. Encarte do Informações Agronômicas 110:12–17 [https://wwwipninet/ppiweb/](https://www.ipni.net/ppiweb/brazil.nsf/87cb8a98bf72572b8525693e0053ea70/7759ddc6878ca7eb83256d05004c6dd1/FILE/Enc12-17-110.pdf) [brazilnsf/87cb8a98bf72572b8525693e0053ea70/](https://www.ipni.net/ppiweb/brazil.nsf/87cb8a98bf72572b8525693e0053ea70/7759ddc6878ca7eb83256d05004c6dd1/FILE/Enc12-17-110.pdf) [7759ddc6878ca7eb83256d05004c6dd1/\\$FILE/Enc12-17-](https://www.ipni.net/ppiweb/brazil.nsf/87cb8a98bf72572b8525693e0053ea70/7759ddc6878ca7eb83256d05004c6dd1/FILE/Enc12-17-110.pdf) [110pdf.](https://www.ipni.net/ppiweb/brazil.nsf/87cb8a98bf72572b8525693e0053ea70/7759ddc6878ca7eb83256d05004c6dd1/FILE/Enc12-17-110.pdf) Acessed 22 Jan 2016
- 65. Fontes MPF, Alleoni LRF (2006) Electrochemical attributes and availability of nutrients, toxic elements, and heavy metals in tropical soils. Sci Agric 63:589–608. doi:[10.1590/S0103-](http://dx.doi.org/10.1590/S0103-90162006000600014) [90162006000600014](http://dx.doi.org/10.1590/S0103-90162006000600014)
- 66. Oliveira MW, Trivelin PCO, Boaretto AE, Muraoka T, Mortatti J (2002) Leaching of nitrogen, potassium, calcium and magnesium in sandy soil cultivated with sugarcane. Pesq Agrop Brasileira 37: 861–868. doi[:10.1590/S0100-204X2002000600016](http://dx.doi.org/10.1590/S0100-204X2002000600016)
- 67. Ghiberto PJ, Libardi PL, Brito AS, Trivelin PCO (2009) Leaching of nutrients from a sugarcane crop growing on an Ultisol in Brazil. Agric Water Manag 96:1443–1448. doi[:10.1016/j.agwat.2009.04.](http://dx.doi.org/10.1016/j.agwat.2009.04.020) [020](http://dx.doi.org/10.1016/j.agwat.2009.04.020)
- 68. Ghiberto PJ, Libardi PL, Brito AS, Trivelin PCO (2011) Components of the water balance in soil with sugarcane crops. Agric Water Manag 102:1–7. doi[:10.1016/j.agwat.2011.09.010](http://dx.doi.org/10.1016/j.agwat.2011.09.010)
- 69. Ghiberto PJ, Libardi PL, Trivelin PCO (2015) Nutrient leaching in an Ultisol cultivated with sugarcane. Agric Water Manag 31:141– 149. doi:[10.1016/j.agwat.2014.09.027](http://dx.doi.org/10.1016/j.agwat.2014.09.027)
- 70. Portocarrero RA, Acreche MM (2014) Nitrate leaching in an Argiudoll cultivated with sugarcane. Sugar Tech 16:329–332. doi[:10.1007/s12355-013-0287-9](http://dx.doi.org/10.1007/s12355-013-0287-9)
- 71. Denmead OT, Macdonald BCT, Bryant G, Reilly RJ, Griffith DWT, Stainlay W, White I, Melville MD (2005) Gaseous nitrogen losses from acid sulfate sugarcane soils on the coastal lowlands. Aust Soc Sugar Cane Technol 27:211–219. doi[:10.1016/S1002-](http://dx.doi.org/10.1016/S1002-0160(11)60118-5) [0160\(11\)60118-5](http://dx.doi.org/10.1016/S1002-0160(11)60118-5)
- 72. Ammann C, Sprig C, Fischer C, Leifeld J, Nefler A (2007) Interactive comment to P. Crutzen et al. Atmos Chem Phys Discuss 7:79–81
- 73. Rauh S (2007) Interactive comment to P. Crutzen et al. Atmos Chem Phys Discuss 7:6–9
- 74. Creutzig F, Ravindranath NH, Berndes G, Bolwig S, Bright R, Cherubini F, Chum H, Corbera E, Delucchi M, Faaij A, Fargione J, Haberl H, Heath G, Lucon O, Plevin R, Popp A, Robledo-Abad C, Rose S, Smith P, Stromman A, Suh S, Masera O (2015) Bioenergy and climate change mitigation: an assessment. Glob Chang Biol Bioenergy 7:916–944. doi:[10.1111/gcbb.](http://dx.doi.org/10.1111/gcbb.12205) [12205](http://dx.doi.org/10.1111/gcbb.12205)
- 75. IPCC (2007) Intergovernmental panel on climate change. Climate change: synthesis report. In: Core Writing Team, Pachauri RK, Reisinger A (eds) Contribution of working groups I, II and III to the fourth assessment report of the intergovernmental panel on climate change. IPCC, Geneva
- 76. Allen DE, Kingston G, Rennenberg H, Dalal RC, Schmidt S (2010) Effect of nitrogen fertilizer management and waterlogging on nitrous oxide emission from subtropical sugarcane soils. Agric Ecosyst Environ 136:209–217. doi:[10.1016/j.agee.2009.11.002](http://dx.doi.org/10.1016/j.agee.2009.11.002)
- 77. Weier KL, Rolston DE, Thorburn PJ (1998) The potential for N losses via denitrification beneath a green cane trash blanket. Proc Aust Soc Sugar Cane Technol 20:169–175
- 78. Denmead OT, Macdonald BCT, Bryant G, Naylor T, Wilson S, Griffith DWT, Wang WJ, Salter B, White I, Moody PW (2010) Emissions of methane and nitrous oxide from Australian

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sugarcane soils. Agric For Meteorol 150:748–756. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.agrformet.2009.06.018) [agrformet.2009.06.018](http://dx.doi.org/10.1016/j.agrformet.2009.06.018)

- 79. Lisboa CC, Butterbach-Bahl K, Mauder M, Kiese R (2011) Bioethanol production from sugarcane and emissions of greenhouse. Glob Chang Biol Bioenergy 3:277–292. doi:[10.1111/j.](http://dx.doi.org/10.1111/j.1757-1707.2011.01095.x) [1757-1707.2011.01095.x](http://dx.doi.org/10.1111/j.1757-1707.2011.01095.x)
- 80. Soares JR, Cantarella H, Vargas VP, Carmo JB, Martins AA, Souza RM, Andrade CA (2015) Enhanced-efficiency fertilizers in nitrous oxide emissions from urea applied to sugarcane. J Environ Qual 44:423–430. doi[:10.2134/jeq2014.02.0096](http://dx.doi.org/10.2134/jeq2014.02.0096)
- 81. Carmo JB, Filoso S, Zotelli LC, Souza Neto ERD, Pitombo LM, Duarte Neto PJ, Vargas VP, Andrade CA, Gava GJC, Rossetto R (2013) Infield greenhouse gas emissions from sugarcane soils in Brazil: effects from synthetic and organic fertilizer application and crop trash accumulation. Glob Chang Biol Bioenergy 5:267–280. doi[:10.1111/j.1757-1707.2012.01199.x](http://dx.doi.org/10.1111/j.1757-1707.2012.01199.x)
- 82. Signor D, Cerri CEP, Conant R (2013) N₂O emissions due to nitrogen fertilizer applications in two regions of sugarcane cultivation in Brazil. Environ Res Lett 8:1–9. doi[:10.1088/1748-9326/](http://dx.doi.org/10.1088/1748-9326/8/1/015013) [8/1/015013](http://dx.doi.org/10.1088/1748-9326/8/1/015013)
- 83. Borjesson P (2009) Good or bad bioethanol from a greenhouse gas perspective—what determines this? Appl Energy 86:589–594. doi[:10.1016/j.apenergy.2008.11.025](http://dx.doi.org/10.1016/j.apenergy.2008.11.025)
- 84. Vargas VP, Cantarella H, Martins AA, Soares JR, Do Carmo JB, De Andrade CA (2014) Sugarcane crop residue increases N_2O and CO₂ emissions under high soil moisture conditions. Sugar Tech 16:174–179. doi:[10.1007/s12355-013-0271-4](http://dx.doi.org/10.1007/s12355-013-0271-4)
- 85. Cantarella H, Trivelin PCO, Contin TLM, Dias FLF, Rossetto R, Marcelino R, Coimbra RB, Quaggio JA (2008) Ammonia volatilisation from urease inhibitor-treated urea applied to sugarcane trash blankets. Sci Agric 65:397–401. doi[:10.1590/S0103-](http://dx.doi.org/10.1590/S0103-90162008000400011) [90162008000400011](http://dx.doi.org/10.1590/S0103-90162008000400011)
- 86. Soares JR, Cantarella H, Menegale MLD (2012) Ammonia volatilization losses from surface-applied urea with urease and nitrification inhibitors. Soil Biol Biochem 52:82–89. doi:[10.1016/j.](http://dx.doi.org/10.1016/j.soilbio.2012.04.019) [soilbio.2012.04.019](http://dx.doi.org/10.1016/j.soilbio.2012.04.019)
- 87. Faria LA, Nascimento CAC, Ventura BP, Florim GP, Luz PHC, Vitti GC (2014) Hygroscopicity and ammonia volatilization losses from nitrogen sources in coated urea. R Bras Ci Solo 38:942–948. doi[:10.1590/S0100-06832014000300026](http://dx.doi.org/10.1590/S0100-06832014000300026)
- 88. Thorburn P, Meier EA, Collins K, Roberton FA (2012) Changes in soil carbon sequestration, fractionation and soil fertility in response to sugarcane residue retention are site-specific. Soil Tillage Res 120:99–111. doi:[10.1016/j.still.2011.11.009](http://dx.doi.org/10.1016/j.still.2011.11.009)
- 89. Fortes C, Trivelin PCO, Vitti AC (2012) Long term decomposition of sugarcane harvest residues in Sao Paulo state Brazil. Biomass Bioenergy 42:189–198. doi[:10.1016/j.biombioe.2012.03.011](http://dx.doi.org/10.1016/j.biombioe.2012.03.011)
- 90. Oliveira MW, Trivelin PCO, Kingston G, Barbosa MHP, Vitti AC (2002) Decomposition and release of nutrients from sugarcane trash in two agricultural environments in Brazil. In: Conference of the Australian Society of Sugar Cane Technologists. Carns, Proceedings 24:1–10
- 91. Meier EA, Thorburn PJ, Wegener MK, Basford KE (2006) The availability of nitrogen from sugarcane trash on contrasting soils in the wet tropics of North Queensland. Nutr Cycl Agroecosyst 75: 101–114. doi[:10.1007/s10705-006-9015-0](http://dx.doi.org/10.1007/s10705-006-9015-0)
- 92. Hemwong S, Toomsan B, Cadisch G, Limpinuntana V, Vityakon P, Patanothai A (2009) Sugarcane residue management and grain legume crop effects on N dynamics, N losses and growth of sugarcane. Nutr Cycl Agroecosyst 83:135–151. doi:[10.1007/s10705-](http://dx.doi.org/10.1007/s10705-008-9209-8) [008-9209-8](http://dx.doi.org/10.1007/s10705-008-9209-8)
- 93. Robertson FA, Thorburn PJ (2007) Management of sugarcane harvest residues: consequences for soil carbon and nitrogen. Aust J Soil Res 45:13–23. doi[:10.1071/SR06080](http://dx.doi.org/10.1071/SR06080)
- 94. Trivelin PCO, Franco HCJ, Otto R, Ferreira DA, Vitti AC, Fortes C, Faroni CE, Oliveira ECA, Cantarella H (2013) Impact of

sugarcane trash on fertilizer requirements for São Paulo, Brazil. Sci Agric 70:345–352. doi[:10.1590/S0103-90162013000500009](http://dx.doi.org/10.1590/S0103-90162013000500009)

- 95. Prado RM, Pancelli M (2008) Resposta de soqueiras de cana-deaçúcar em sistema de colheita sem queima à aplicação de nitrogênio. Bragantia 67:951–959. doi:[10.1590/S0006-](http://dx.doi.org/10.1590/S0006-87052008000400018) [87052008000400018](http://dx.doi.org/10.1590/S0006-87052008000400018)
- 96. Wood AW (1991) Management of crop following green harvesting of sugarcane in North Queensland. Soil Tillage Res 20:69–85. doi[:10.1016/0167-1987\(91\)90126-I](http://dx.doi.org/10.1016/0167-1987(91)90126-I)
- 97. Landel MGA, Scarpari MS, Xavier MA, Anjos IA, Baptista AS, Aguiar CL, Silva DN, Bidoia MAP, Brancalião SR, Bressiani JÁ, Campos MF, Miguel PEM, Silva TN, Silva VHP, Souza Anjos LO, Ogata BH (2013) Residual biomass potential of commercial and pre-commercial sugarcane cultivars. Sci Agric 70:299–304. doi[:10.1590/S0103-90162013000500003](http://dx.doi.org/10.1590/S0103-90162013000500003)
- 98. Oliveira MW, Trivelin PCO, Gava GJC, Penatti CP (1999) Sugarcane trash degradation. Sci Agric 56:803–809. doi[:10.](http://dx.doi.org/10.1590/S0103-90161999000400006) [1590/S0103-90161999000400006](http://dx.doi.org/10.1590/S0103-90161999000400006)
- 99. Magalhães PSG, Nogueira LAH, Cantarella H, Rossetto R, Franco HCJ, Braunbeck OA (2012) Agro-industrial technological paths in sustainability of sugarcane bioenergy by center of strategic studies and management. CGEE, Brasília, Brasil
- 100. Caceres NT, Alcarde JC (1995) Adubação verde com leguminosas em rotação com cana-de-açúcar (Saccharum ssp). STAB – Açúcar Álcool e Subprodutos 13:16–20
- 101. Garside AL, Berthelsen JE, Richards CL, Toovey LM (1996) Fallow legumes on the wet tropical coast: some species and management options. P Aust Soc Sugar Cane Technol 18:202–208
- 102. Shoko MD, Pieterse PJ, Zhou M (2009) Effect of soybean (Glycine max) as a breakcrop on the cane and sugar yield of sugarcane. Sugar Tech 11:252–257
- 103. Ambrosano EJ, Trivelin PCO, Cantarella H, Ambrosano GMB, Schammas EA, Muraoka T, Rossi F (2011b) 15 N-labeled nitrogen from green manure and ammonium sulfate utilization by the sugarcane ratoon. Sci Agric 68:361–368. doi[:10.1590/S0103-](http://dx.doi.org/10.1590/S0103-90162011000300014) [90162011000300014](http://dx.doi.org/10.1590/S0103-90162011000300014)
- 104. Cheruiyot EK, Mumera LM, Nakhone LN, Mwonga SM (2003) Effect of legume-managed fallow on weeds and soil nitrogen in following maize (Zea mays L.) and wheat (Triticum aestivum L.) crops in the Rift Valley highlands of Kenya. Aust J Exp Agric 43: 597–604. doi[:10.1071/EA02033](http://dx.doi.org/10.1071/EA02033)
- 105. Dinardo-Miranda LL, Fracasso JV (2009) Spatial distribution of plant-parasitic nematodes in sugarcane fields. Sci Agric 66:188– 194. doi:[10.1590/S0103-90162009000200007](http://dx.doi.org/10.1590/S0103-90162009000200007)
- 106. Ambrosano EJ, Cantarella H, Ambrosano GMB, Schammas EA, Dias FLF, Rossi F, Trivelin PCO, Muraoka T, Sachs RCC, Azcon R (2011a) Produtividade da cana-de-açúcar após o cultivo de leguminosas. Bragantia 70:810–818. doi:[10.1590/S0006-](http://dx.doi.org/10.1590/S0006-87052011000400012) [87052011000400012](http://dx.doi.org/10.1590/S0006-87052011000400012)
- 107. Herridge DF, Peoples MB, Boddey RM (2008) Global inputs of biological nitrogen fixation in agricultural systems. Plant Soil 311: 1–18. doi:[10.1007/s11104-008-9668-3](http://dx.doi.org/10.1007/s11104-008-9668-3)
- 108. Silva GTA, Matos LV, Nobrega PD, Campello EFC, Resende AS (2008) Chemical composition and decomposition rate of plants used as green manure. Sci Agric 65:298–305. doi[:10.1590/](http://dx.doi.org/10.1590/S0103-90162008000300010) [S0103-90162008000300010](http://dx.doi.org/10.1590/S0103-90162008000300010)
- 109. Garside AL, Berthelsen JE, Richards CL (1997) Effect of fallow history on cane and sugar yield of a following plant cane crop. P Aust Soc Sugar Cane Technol 19:80–86
- 110. Park SE, Webster TJ, Horan HL, James AT, Thorburn PJ (2010) A legume rotation crop lessens the need for nitrogen fertiliser throughout the sugarcane cropping cycle. Field Crop Res 119: 331–341. doi[:10.1016/j.fcr.2010.08.001](http://dx.doi.org/10.1016/j.fcr.2010.08.001)
- 111. Cantarella H, Trivelin PCO, Vitti AC (2007) Nitrogênio e enxofre na cultura da cana de açúcar. In: Yamada T, Abdalla SRS, Vitti GC

(eds) Nitrogênio e enxofre na agricultura Brasileira. International Plant Nutrition Institute, Piracicaba, pp. 355–412

- 112. Franco HCJ, Trivelin PCO, Faroni CE, Vitti AC, Otto R (2010) Stalk yield and technological attributes of planted cane as related to nitrogen fertilization. Sci Agric 67:579–590. doi:[10.1590/](http://dx.doi.org/10.1590/S0103-90162010000500012) [S0103-90162010000500012](http://dx.doi.org/10.1590/S0103-90162010000500012)
- 113. Fortes C, Trivelin PCO, Vitti AC, Otto R, Franco HCJ, Faroni CE (2013a) Stalk and sucrose yield in response to nitrogen fertilization of sugarcane under reduced tillage. Pesq Agropec Bras 48:88–96. doi:[10.1590/S0100-204X2013000100012](http://dx.doi.org/10.1590/S0100-204X2013000100012)
- 114. Salcedo IH, Sampaio EVSB, Alves GD (1985) Mineralização do carbono e do nitrogênio em solo cultivado com cana-de-açúcar. R Bras Ci Solo 9:33–38
- 115. Thorburn PJ, Dart IK, Biggs IJ, Baillie CP, Smith MA, Keating BA (2003) The fate of nitrogen applied to sugarcane by trickle irrigation. Irrig Sci 22:201–209. doi[:10.1007/s00271-003-0086-2](http://dx.doi.org/10.1007/s00271-003-0086-2)
- 116. Boddey RM, Urquiaga S, Alves BJR, Reis VM (2003) Endophytic nitrogen fixation in sugarcane: present knowledge and future applications. Plant Soil 252:139–149. doi:[10.1023/](http://dx.doi.org/10.1023/A:1024152126541) [A:1024152126541](http://dx.doi.org/10.1023/A:1024152126541)
- 117. Urquiaga S, Xavier RP, Morais RF, Batista RB, Schultz N, Leite JM, Sá JM, Barbosa KP, Resende AS, Alves BJR, Boddey RM (2012) Evidence from field nitrogen balance and $15N$ natural abundance data of the contribution of biological N2 fixation to Brazilian sugarcane varieties. Plant Soil 356:5–21. doi[:10.1007/](http://dx.doi.org/10.1007/s11104-011-1016-3) [s11104-011-1016-3](http://dx.doi.org/10.1007/s11104-011-1016-3)
- 118. Oliveira ALM, Canuto EL, Urquiaga S, Reis VM, Baldani JI (2006) Yield of micropropagated sugarcane varieties in different soil types following inoculation with endophytic diazotrophic bacteria. Plant Soil 284:23–32. doi:[10.1007/s11104-006-0025-0](http://dx.doi.org/10.1007/s11104-006-0025-0)
- 119. Schultz N, Morais RF, Silva JA, Baptista RB, Oliveira RP, Leite JM, Pereira W, Carneiro Junior JB, Alves BJR, Baldani JI, Boddey RM, Urquiaga S, Reis VM (2012) Avaliação agronômica de variedades de cana-de-açúcar inoculadas com bactérias diazotróficas e adubadas com nitrogênio. Pesq Agrop Brasileira 47:261–268. doi[:10.1590/S0100-204X2012000200015](http://dx.doi.org/10.1590/S0100-204X2012000200015)
- 120. Schultz N, Silva JA, Sousa JS, Monteiro RC, Oliveira RP, Chaves VA, Pereira WS, Marinete F, Baldani JI, Boddey RM, Reis VM, Urquiaga S (2014) Inoculation of sugarcane with diazotrophic bacteria. R Bras Ci Solo 38:407–414. doi:[10.1590/S0100-](http://dx.doi.org/10.1590/S0100-06832014000200005) [06832014000200005](http://dx.doi.org/10.1590/S0100-06832014000200005)
- 121. Cantarella H, Montezano ZF, Joris HAW, Vitti AC, Rossetto R, Gava GJC, Dias FLF, Urquiaga S, Reis VM (2014) Nitrogen fertilization and inoculation of sugarcane with diazotrophic bacteria: 13-site-year of field results. Proceedings of the 2nd Brazilian BioEnergy Science and Technology Conference, Campos do Jordão
- 122. Biggs IM, Stewart GR, Wilson JR, Critchley C (2002) 15 N natural abundance studies in Australian commercial sugarcane. Plant Soil 238:21–30. doi[:10.1023/A:1014280420779](http://dx.doi.org/10.1023/A:1014280420779)
- 123. Hoefsloot G, Termorshuizen AJ, Watt DA, Cramer MD (2005) Biological nitrogen fixation is not a major contributor to the nitrogen demand of a commercially grown South African sugarcane cultivar. Plant Soil 277:85–96. doi:[10.1007/s11104-005-2581-0](http://dx.doi.org/10.1007/s11104-005-2581-0)
- 124. Lin L, Li Z, Hu C, Zhang X, Chang S, Yang L, Li Y, An Q (2012) Plant growth-promoting nitrogen-fixing enterobacteria are in association with sugarcane plants growing in Guangxi, China. Microbes Environ 27:391–398. doi:[10.1264/jsme2.ME11275](http://dx.doi.org/10.1264/jsme2.ME11275)
- 125. Santi C, Bogusz D, Franchie C (2013) Biological nitrogen fixation in non-legume plants. Ann Bot 111:743–767. doi:[10.1093/aob/](http://dx.doi.org/10.1093/aob/mct048) [mct048](http://dx.doi.org/10.1093/aob/mct048)
- 126. Rossetto R, Dias FLF, Landell MGA, Cantarella H, Tavares S, Vitti AC, Perecin D (2010) N and K fertilization of sugarcane ratoons harvested without burning. Proc Int Soc Sugar Cane Tech 27:1–8
- 127. Thorburn PJ, Biggs JS, Webster AJ, Biggs IM (2011b) An improved way to determine nitrogen fertiliser requirements of sugarcane crops to meet global environmental challenges. Plant Soil 339:51–67. doi[:10.1007/s11104-010-0406-2](http://dx.doi.org/10.1007/s11104-010-0406-2)
- 128. Spironello A, Raij B, Penatti CP, Cantarella H, Morelli JLM, Orlando Filho J, Landell MGA, Rossetto R (1997) Sugarcane crop. In: van Raij B, Cantarella H, Quaggio JA, Furlani AMC (eds) Recomendations of fertilizer and lime to São Paulo state, 2nd edn. Instituto Agronômico, Campinas
- 129. Galdos MV, Cerri CC, Lal R, Bernoux M, Feigl B, Cerri CEP (2010) Net greenhouse gas fluxes in Brazilian ethanol production systems. Glob Chang Biol Bioenergy 2:37–44. doi[:10.1111/j.](http://dx.doi.org/10.1111/j.1757-1707.2010.01037.x) [1757-1707.2010.01037.x](http://dx.doi.org/10.1111/j.1757-1707.2010.01037.x)
- 130. Vitti AC, Franco HCJ, Trivelin PCO, Ferreira DA, Otto R, Fortes C, Faroni CE (2011) Nitrogênio proveniente da adubação nitrogenada e de resíduos culturais na nutrição da cana-planta. Pesq Agrop Brasileira 46:287–293. doi:[10.1590/S0100-](http://dx.doi.org/10.1590/S0100-204X2011000300009) [204X2011000300009](http://dx.doi.org/10.1590/S0100-204X2011000300009)
- 131. Fortes C, Vitti AC, Otto R, Ferreira DA, Franco HCJ, Trivelin PCO (2013b) Contribution of nitrogen from sugarcane harvest residues and urea for crop nutrition. Sci Agric 70:313–320. doi: [10.1590/S0103-90162013000500005](http://dx.doi.org/10.1590/S0103-90162013000500005)
- 132. Ferreira DA, Franco HCJ, Otto R, Vitti AC, Fortes C, Faroni CE, Garside AL, Trivelin PCO (2015) Contribution of N from green harvest residues for sugarcane nutrition in Brazil. Glob Chang Biol Bioenergy. doi:[10.1111/gcbb.12292](http://dx.doi.org/10.1111/gcbb.12292)
- 133. Schroeder BL, Wood AW, Sefton M, Hurney AP, Skocaj DM, Stainlay T, Moody PW (2010) District yield potential: an appropriate basis for nitrogen guidelines for sugarcane production. Proc Aust Soc Sugar Cane Technol 32:193–209
- 134. Mariano E (2015) Predição da necessidade de fertilizante nitrogenado pela cana-de-açúcar e reações do nitrogênio orgânico e mineral dissolvidos em palha e solo de canaviais. University of São Paulo, Thesis
- 135. Vale DW, Prado RM, Pancelli MA (2009) Análise econômica da adubação nitrogenada em soqueiras de cana-de-açúcar. STAB-Açúcar Álcool e Subprodutos 28:32–34
- 136. Fortes C (2010) Produtividade de cana-de-açúcar em função da adubação nitrogenada e da decomposição da palhada em ciclos consecutivos. Thesis, University of São Paulo
- 137. Vieira MX, Trivelin PCO, Franco HCJ, Otto R, Faroni CE (2010) Ammonium chloride as nitrogen source in sugarcane harvested without burning. R Bras Ci Solo 34:1165–1174. doi[:10.1590/](http://dx.doi.org/10.1590/S0100-06832010000400016) [S0100-06832010000400016](http://dx.doi.org/10.1590/S0100-06832010000400016)
- 138. Castro SGQ (2012) Sistemas de manejo em colheita e cultivo associados à adubação nitrogenada da cana-de-açúcar: aspectos qualiquantitativos da produção. Thesis, University of São Paulo State
- 139. Penatti CP, Araújo JV, Forti JA, Ribeiro R (2001) Doses de vinhaça e nitrogênio aplicadas em cana-soca durante quatro safras em solo LV—Usina São José da Estiva. STAB-Açúcar Álcool e Subprodutos 19:38–41
- 140. Castro SGQ, Barbosa JC, Castro SAQ, Mutton MA (2012) Aplicação diferenciada de nitrogênio em soqueira de cana-deaçúcar. In: Reunião Brasileira de Fertilidade do Solo e Nutrição de Plantas, 30, Maceió, Anais... Maceió, SBCS, UFAL, CD-ROM
- 141. Orlando Filho J, Rodella AA, Beltrame JÁ, Lavorenti NA (1999) Doses, fontes e formas de aplicação de nitrogênio em cana-deaçúcar. STAB Açúcar Álcool e Subprodutos 17:39–41
- 142. Anda (2015) Associação nacional para difusão de adubos. Anuário estatístico do setor de fertilizantes, 2013. [http://anda.](http://anda.org.br/multimidia/capa_e_indice_do_ae_20130001.pdf) [org.br/multimidia/capa_e_indice_do_ae_20130001.pdf.](http://anda.org.br/multimidia/capa_e_indice_do_ae_20130001.pdf) Accessed 24 April 2015
- 143. Moir JL, Cameron KC, Di HJ (2007) Effects of the nitrification inhibitor dicyandiamide on soil mineral N, pasture yield, nutrient

uptake and pasture quality on a grazed pasture system. Soil Use Manag 23:111–120. doi[:10.1111/j.1475-2743.2006.00078.x](http://dx.doi.org/10.1111/j.1475-2743.2006.00078.x)

- 144. Zaman M, Saggar S, Blennerhassett JD, Singh J (2009) Effect of urease and nitrification inhibitors on N transformation, gaseous emissions of ammonia and nitrous oxide, pasture yield and N uptake in grazed pasture system. Soil Biol Biochem 41:1270– 1280. doi:[10.1016/j.soilbio.2009.03.011](http://dx.doi.org/10.1016/j.soilbio.2009.03.011)
- 145. Vieira-Megda MX, Trivelin PCO, Franco HCJ, Otto R, Vitti AC (2012) Eficiência agronômica de adubos nitrogenados em soqueira de cana-de-açúcar colhida sem queima. Pesq Agrop Brasileira 47:1681-1690. doi:[10.1590/S0100-](http://dx.doi.org/10.1590/S0100-204X2012001200002) [204X2012001200002](http://dx.doi.org/10.1590/S0100-204X2012001200002)
- 146. Shoji S, Delgado JA, Mosier A, Miura Y (2001) Use of controlled release fertilizers and nitrification inhibitors to increase nitrogen use efficiency and to conserve air and water quality. Commun Soil Sci Plant Anal 32:1051–1070. doi:[10.1081/CSS-100104103](http://dx.doi.org/10.1081/CSS-100104103)
- 147. Penatti CP (2014) Adubação da cana-de-açúcar: 30 anos de experiência. Ottoni, Itu
- 148. Liu XJ, Mosier AR, Halvorson AD, Zhang FS (2005) Tillage and nitrogen application effects on nitrous and nitric oxide emissions from irrigated corn fields. Plant Soil 276:235–249. doi[:10.1007/](http://dx.doi.org/10.1007/s11104-005-4894-4) [s11104-005-4894-4](http://dx.doi.org/10.1007/s11104-005-4894-4)
- 149. Van Kessel C, Venterea R, Six J, Adviento-Borbe MA, Linquist B, Groenigen KJV (2013) Climate, duration, and N placement determine N2O emissions in reduced tillage systems: a meta-analysis. Glob Chang Biol 19:33–44. doi[:10.1111/j.1365-2486.2012.](http://dx.doi.org/10.1111/j.1365-2486.2012.02779.x) [02779.x](http://dx.doi.org/10.1111/j.1365-2486.2012.02779.x)
- 150. Chapuis-Lardy L, Wrage N, Metay A, Chotte J-L, Bernoux M (2007) Soils a sink for N_2O ? A review. Glob Chang Biol 13:1– 17. doi:[10.1111/j.1365-2486.2006.01280.x](http://dx.doi.org/10.1111/j.1365-2486.2006.01280.x)
- 151. Vitti AC, Trivelin PCO, Gava GJC, Franco HCJ, Bologna IR, Faroni CE (2007) Produtividade da cana-de-açúcar relacionada à localização de adubos nitrogenados aplicados sobre os resíduos culturais em canavial sem queima. R Bras Ci Solo 31:491–498. doi[:10.1590/S0100-06832007000300009](http://dx.doi.org/10.1590/S0100-06832007000300009)
- 152. Amaducci MT, Barbanti L, Venturi G (2006) Comparing application methods for N-fertilizer in the sugar beet crop. Ital J Agron 1: 51–61. doi[:10.4081/ija.2006.51](http://dx.doi.org/10.4081/ija.2006.51)
- 153. Beres BL, Harker KN, Clayton GW, Bremer E, O'Donovan JT, Blackshaw RE, Smith AM (2010) Influence of N fertilization method on weed growth, grain yield and grain protein concentration in no-till winter wheat. Can J Plant Sci 90:637–644. doi[:10.](http://dx.doi.org/10.4141/CJPS10037) [4141/CJPS10037](http://dx.doi.org/10.4141/CJPS10037)
- 154. Blair N (2000) Impact of cultivation and sugarcane green trash management on carbon fractions and aggregate stability for a chromic Luvisol in Queensland. Soil Tillage Res 55:183–191. doi[:10.1016/S0167-1987\(00\)00113-6](http://dx.doi.org/10.1016/S0167-1987(00)00113-6)
- 155. Manechini C (2000) Cultivo mecânico da soqueira de cana colhida sem queimar (condensado da experiência acumulada). Relatório Interno Coopersucar - Projeto Cana crua, Piracicaba
- 156. Castro SGQ, Franco HJC, Mutton MA (2014) Harvest managements and cultural practices in sugarcane. R Bras Ci Solo 38:299– 306. doi:[10.1590/S0100-06832014000100030](http://dx.doi.org/10.1590/S0100-06832014000100030)
- 157. Smith DM, Inman-Bamber NG, Thorburn PJ (2005) Growth and function of the sugarcane root system. Field Crop Res 92:169– 183. doi:[10.1016/j.fcr.2005.01.017](http://dx.doi.org/10.1016/j.fcr.2005.01.017)
- 158. Otto R, Silva AP, Franco HCJ, Oliveira ECA, Trivelin PCO (2011) High soil penetration resistance reduces sugarcane root system development. Soil Tillage Res 117:201–210. doi:[10.1016/j.still.](http://dx.doi.org/10.1016/j.still.2011.10.005) [2011.10.005](http://dx.doi.org/10.1016/j.still.2011.10.005)
- 159. Demattê JLI (2005) Cultura da cana-de-açúcar: recuperação e manutenção da fertilidade dos solos. Encarte Técnico Informações Agronômicas 111. [http://www.ipni.net/publication/](http://www.ipni.net/publication/ia-brasil.nsf/0/80EF811848745EAF83257AA1006B29C1/FILE/Encarte%20111.pdf) [ia-brasil.nsf/0/80EF811848745EAF83257AA1006B29C1/](http://www.ipni.net/publication/ia-brasil.nsf/0/80EF811848745EAF83257AA1006B29C1/FILE/Encarte%20111.pdf) [\\$FILE/Encarte%20111.pdf](http://www.ipni.net/publication/ia-brasil.nsf/0/80EF811848745EAF83257AA1006B29C1/FILE/Encarte%20111.pdf). Accessed 12 Dec 2015
- 160. Oliveira ECA (2011) Balanço nutricional da cana-de-açúcar relacionado à adubação nitrogenada. Thesis, University of São Paulo
- 161. Uribe RAM, Gava GJC, Saad JCC, Kolln OT (2013) Ratoon sugarcane yield integrated drip-irrigation and nitrogen fertilization. Eng Agric 33:1124–1133. doi:[10.1590/S0100-](http://dx.doi.org/10.1590/S0100-69162013000600005) [69162013000600005](http://dx.doi.org/10.1590/S0100-69162013000600005)
- 162. Kolln OT, Gava GJC, Cantarella H, Franco HCJ, Uribe RAM, Pannuti LER, Trivelin PCO (2015) Fertigated sugarcane yield and carbon isotope discrimination $(\Delta 13C)$ related to nitrogen nutrition. Sug Tech 1–10. doi: [10.1007/s12355-015-0384-z](http://dx.doi.org/10.1007/s12355-015-0384-z)
- 163. Molin JP, Frasson FR, Amaral LR, Povh FP, Salvi JV (2010) Capacidade de um sensor ótico em quantificar a resposta da cana-de-açúcar a doses de nitrogênio. R Bras Eng Agric Amb 14: 1345–1349. doi:[10.1590/S1415-43662010001200014](http://dx.doi.org/10.1590/S1415-43662010001200014)
- 164. Teal RK, Tubana B, Girma K, Freeman KW, Arnall DB, Walsh O, Raun RW (2006) In-season prediction of corn grain yield potential using normalized difference vegetation index. Agron J 98:1488– 1494. doi:[10.2134/agronj2006.0103](http://dx.doi.org/10.2134/agronj2006.0103)
- 165. Eitel JUH, Long DS, Gessler PE, Hunt ER (2008) Combined spectral index to improve ground-based estimates of nitrogen status in dryland wheat. Agron J 100(6):1694–1702. doi[:10.2134/](http://dx.doi.org/10.2134/agronj2007.0362) [agronj2007.0362](http://dx.doi.org/10.2134/agronj2007.0362)
- 166. Wu C, Niu Z, Tang Q, Huang W (2008) Estimating chlorophyll content from hyperspectral vegetation indices: modeling and validation. Agric For Meteorol 148:1230–1241. doi[:10.1016/j.](http://dx.doi.org/10.1016/j.agrformet.2008.03.005) [agrformet.2008.03.005](http://dx.doi.org/10.1016/j.agrformet.2008.03.005)
- 167. Blackmer TM, Schepers JS, Varvel GE, Walter-Shea EA (1996) Nitrogen deficiency detection using reflected shortwave radiation from irrigated corn canopies. Agron J 88:1–5. doi[:10.2134/](http://dx.doi.org/10.2134/agronj1996.00021962008800010001x) [agronj1996.00021962008800010001x](http://dx.doi.org/10.2134/agronj1996.00021962008800010001x)
- 168. Amaral LR, Molin JP, Portz G, Finazzi FB, Cortinove L (2015) Comparison of crop canopy reflectance sensors used to identify sugarcane biomass and nitrogen status. Precis Agric 16:15–28. doi[:10.1007/s11119-014-9377-2](http://dx.doi.org/10.1007/s11119-014-9377-2)
- 169. Amaral LR, Molin JP, Schepers JS (2015) Algorithm for variablerate nitrogen application in sugarcane based on active crop canopy sensor. Agron J 107:1513–1523. doi:[10.2134/agronj14.0494](http://dx.doi.org/10.2134/agronj14.0494)