# TECHNICAL REPORT

# Stability and alignment of MCC/IMS devices

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## Introduction

There are numerous analytical detection methods available for human breath investigations. The major spectrometric methods used are proton transfer reaction mass spectrometry (PRT-MS) [\[1](#page-4-0)–[5\]](#page-4-0), selected ion flow tube mass spectrometry (SIFT-MS) [\[6](#page-4-0)–[10](#page-4-0)], solid phase micro extraction-gas chromatography coupled to mass spectrometry (SPME-GC/ MS) [[11](#page-4-0)–[14\]](#page-4-0), and multi-capillary column coupled to ion mobility spectrometry (MCC/IMS) [[15](#page-4-0)–[20](#page-4-0)]. In all cases mentioned, a non–invasive and an easy method for early diagnosis or therapy monitoring are developed by identifying disease-specific biomarkers in the breath of patients.

Obviously, if the results of these studies are to be valid, consideration of sampling techniques, and the comparison of the inhaled and the exhaled air are some fundamental requirements. Small changes within the room air can affect the difference between room air and exhaled breath depending

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P. Favrod e-mail: p.favrod@kist-europe.de on the method selected, for example, patients suffering lung cancer and healthy volunteers. Therefore, in the present paper a study of the stability of two volatiles present in room air ion mobility spectrometry measurements over 6 months is reported, and also the variability of these two volatiles for different MCC/IMS devices is also described.

The present paper considers the influence of temperature and other parameters with small variations in the instrumentation. General remarks could be found in [\[21](#page-4-0), [22](#page-4-0)], the influence of temperature in [\[23](#page-4-0)–[32\]](#page-4-0), pressure in [\[33](#page-5-0)], electric field in [[34](#page-5-0), [35\]](#page-5-0), carrier gas flow [\[23](#page-4-0), [31](#page-4-0)], geometrical parameters [\[36](#page-5-0), [37\]](#page-5-0). Therefore, we discuss the inter-comparison of different instruments. Theoretical aspects and in-depth explanations regarding the operating conditions of the MCC/IMS were discussed in the literature mentioned above. The present paper deals with practical points that are to be considered while intercomparing different instruments and show, that a proper alignment is really needed for such purpose.

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#### Material and methods

The multi-capillary column—ion mobility spectrometer (MCC/ IMS) used for this study was a BreathDiscovery (B&S Analytik, Dortmund, Germany). The major measurement parameters are summarized in Table 1. The multi-capillary column (MCC) is an OV-5 type (Multichrom, Novosibirsk, Russia) and is used as the pre-separation unit. In this MCC, the analytes of the sample are sent through 1,000 parallel capillaries, each with an inner diameter of 40 μm and a film thickness of 200 nm. The total diameter of the separation column is 3 mm. Once pre-separated, the analytes present in the sample of 10 mL and the carrier gas reach the ion mobility spectrometer where they are ionized with a 555 MBq  $^{63}$ Ni β-radiation.

Once the analytes are ionized, when the shutter opens they can pass through the drift region where they are differentiated by mobility and then they are detected using a Faraday plate. In this study only the ions in positive mode were considered.

The signals/peaks were characterized using the software Visual Now (B&S Analytik, Dortmund Germany), which is described elsewhere [[19](#page-4-0), [38](#page-5-0)–[40\]](#page-5-0). All analytes are characterized



Fig. 1 MCC/IMS chromatogram for a measurement of room air using IMS-1 device. The studied compounds are highlighted inside purple circles: Benzothiazol and Nonanal

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by their position (in the chromatograms) with the inverse reduced mobilities  $(1/K_0$ -value) and retention time (RT-value) and no peak height intensity was considered.

# Results and discussion

# Stability—MCC/IMS peaks variations

Studies of Room Air with MCC/IMS show a wide variety of peaks depending on the location of the device and the day and time. So, to compare different Chromatograms, there is a need to have at least one peak that is always in the chromatogram to align the different measurement data. In this line, we have found two analytes that are always in the room air chromatograms analyzed: Benzothiazol and Nonanal as can be seen in Fig. [1](#page-1-0).

To assure that both compounds are found in all room air chromatograms, measurements over 6 months giving up to more than 2000 measurements for IMS-1 device have been analyzed. Figure 2 shows the histograms of the inverse of the reduced mobility and the retention time distributions for Benzothiazol and Nonanal. It can be easily seen that Benzothiazol follows a Gaussian curve for both  $1/K_0$  ( $R^2$ = 0.95) and RT ( $R^2$ =0.93). While, Nonanal does not follow this behavior. This is due to the fact that Benzothiazol evaporates from the plastic compounds of the MCC/IMS, while Nonanal



Fig. 3  $1/K_0$  and RT distributions along the time for Benzothiazol and Nonanal for IMS-1 device

Table 2 Mean values of RT and  $1/K_0$  of the studied MCC/IMS devices

	<b>Benzothiazol</b>		Nonanal		
	$1/K_0$ (Vs/cm <sup>2</sup> )	RT(s)	$1/K_0$ (Vs/cm <sup>2</sup> )	RT(s)	
$IMS-1$			$0.575 \pm 0.001$ $127 \pm 1$ $0.741 \pm 0.002$ $57 \pm 2$		2066 files
$IMS-2$			$0.571 \pm 0.001$ $144 \pm 2$ $0.734 \pm 0.002$ $61 \pm 1$		127 files
$IMS-3$			$0.574 \pm 0.002$ $148 \pm 3$ $0.734 \pm 0.002$ $64 \pm 1$		43 files
mean	0.573		0.739		

comes from the air-conditioning system and it can be present in the room air sometimes or not.

Furthermore, we can take a look into the distribution of the 1/  $K_0$  and RT over time (Fig. [3](#page-2-0)), where the mean values can be found in Table 2 for IMS-1 where is also showed with the standard deviation range. The analysis for Benzothiazol and Nonanal shows that for the  $1/K_0$ , there is a declining linear tendency of the inverse reduced mobility value, while for the RT there is a constant linear tendency and almost all results are within the range of standard deviation. Variations in the mobility can be explained due to the appearance of  $M(H<sub>2</sub>O)<sub>n</sub>H<sup>+</sup>$  like cluster ions formed from small variations of humidity content of the carrier gas. The standard deviation for Nonanal is the double that of Benzothiazol for both  $1/K_0$  and RT, as can be seen in Fig. [3](#page-2-0).

### Alignment—MCC/IMS devices variations

Once considered the stability of the devices, we focused on the alignment between three different MCC/IMS devices for Benzothiazol and Nonanal. The box-and-whisker plots have been used to show the variation in the inverse reduced mobility and also in the RT for the studied devices (Fig. 4). The results are summarized in Table 2. The expected  $1/K_0$  values have to be similar for all three devices, as it is obtained. However for the retention time, it is expected that for different devices with

different oldness the RT values may vary more sustainably. Thus, the way to adjust the temperature of the MCC will be different. Small changes in the temperature adjustment and place of measurement of the temperature (IMS-1 and IMS-2/IMS-3 are different in that point of measurement) will affect the retention time scale. So, the results show the importance to align the data of different devices especially for the RT. In addition, more sensitive regulation of temperature over the MCC should be considered. As shown in Fig. 4, linear alignment on the Retention time scale is possible.

From the analyzed VOC's, Benzothiazol has a Gaussian behavior in RT-values and a linear tendency over time. The older the device, (i.e. multi-capillay column) the sooner the analytes elute out from the MCC, i.e., the lower RT for the analytes. Also Benzothiazol is from the materials of which the MCC/IMS is built, so we will have it in all the chromatograms either from Room Air or from Breath Analysis, so all these properties make Benzothiazol a suitable peak for the alignment of different devices in positive mode.

It should be noted, that the software package of B&S Analytik, VisualNow includes functionality for alignment on both the inverse reduced mobilitiy and the retention time scales.

#### Summary

The present paper shows, that for investigations of time series using a single instrument and multi-centric studies including the comparison of spectra and IMS-chromatograms from different instruments a real need exists for alignment on the inverse reduced mobilitiy and the retention time scales. By comparison of three different instruments the influence of measuring and adjusting the temperature as well as the temperature range allowed by the regulation circle was observed to be small but considerable. On the retention time scale a linear alignment is sufficient. For the drift time scale further investigations are needed, but the variations were rather small in this dimension.



Fig. 4 Box-and-whisker plots of the  $1/K_0$  and RT of Benzothiazol and Nonanal for three studied MCC/IMS devices

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#### References

- 1. Thekedar B, Szymczak W, Hoellriegl V, Hoeschen C, Oeh U (2009) Investigations on the variability of breath gas sampling using PTR-MS. J Breath Res 3:1–11
- 2. Schwarz K, Filipiak W, Amann A (2009) Determining concentration patterns of volatile compounds in exhaled breath by PTR-MS. J Breath Res 3:1–15
- 3. Wisthaler A (2004) PTR-MS: a new tool for the rapid detection and quantification of VOCs in air at ultra-trace levels. Institut für Ionenphysik, Leopold-Franzens-Universität Innsbruck, Innsbruck
- 4. Lindinger W, Hansel A, Jordan A (1998) On-Line Monitoring of Volatile Organic Compounds at pptv Levels by Means of Proton-Transfer-Reaction Mass Spectrometry ( PTR-MS). - Medical Applications, Food Control and Environmental Research. Int J Mass Spectr Ion Proc 173:191–241
- 5. Lindinger W, Hansel A, Jordan A (1998) Proton-transfer-reaction mass spectrometry (PTR-MS): on-line monitoring of volatile organic compounds at pptv levels. Chem Soc Rev 27:347– 354
- 6. Smith D, Spanel P, Enderby B, Lenney W, Turner C, Davies SJ (2010) Isoprene levels in the exhaled breath of 200 healthy pupils within the age range 7–18 years studied using SIFT-MS. J Breath Res 4:1–7
- 7. Enderby B, Lenney W, Brady M, Emmett C, Spanel P, Smith D (2009) Concentrations of some metabolites in the breath of healthy children aged 7–18 years measured using selected ion flow tube mass spectrometry (SIFT-MS). J Breath Res 3:1–11
- 8. Spanel P, Smith D (2008) Quantification of trace levels of the potential cancer biomarkers formaldehyde, acetaldehyde and propanol in breath by SIFT-MS. J Breath Res 2:1–10
- 9. Spanel P, Dryahina K, Smith D (2007) The concentration distributions of some metabolites in the exhaled breath of young adults. J Breath Res 1:1–8
- 10. Dryahina K, Polasek M, Spanel P (2004) A selected ion flow tube, SIFT, study of the ion chemistry of H3O+, NO+ and O2+• ions with several nitroalkanes in the presence of water vapour. Int J Mass Spectrom 239:57–65
- 11. Ligor M, Ligor T, Bajtarevic A, Ager C, Pienz M, Klieber M, Denz H, Fiegl M, Hilbe W, Weiss W, Lukas P, Jamnig H, Hackl M, Buszewski B, Miekisch W, Schubert J, Amann A (2009) Determination of volatile organic compounds in exhaled breath of patients with lung cancer using solid phase microextraction and gas chromatography mass spectrometry. Clin Chem Lab Med 47(5):550– 560
- 12. Buszewski B, Ulanowska A, Ligor T, Denderz N, Amann A (2009) Analysis of exhaled breath from smokers, passive smokers and

non-smokers by solid-phase microextraction gas chromatography/ mass spectrometry. Biomed Chromatogr 23(5):551–556

- 13. Ligor T, Ligor M, Amann A, Ager C, Bachler M, Dzien A, Buszewski B (2008) The analysis of healthy volunteers' exhaled breath by the use of solid-phase microextraction and GC-MS. J Breath Res 2:1–8
- 14. Schubert JK, Miekisch W, Fuchs P, Scherzer N, Lord H, Pawliszyn J, Mundkowski RG (2007) Determination of antibiotic drug concentrations in circulating human blood by means of solid phase micro-extraction. Clin Chim Acta 386(1–2):57–62
- 15. Baumbach JI (2006) Process analysis using ion mobility spectrometry. Anal Bioanal Chem 384(5):1059–1070
- 16. Jünger M, Bödeker B, Baumbach JI (2010) Peak assignment in multi-capillary column—ion mobility spectrometry using comparative studies with gas chromatography—mass spectrometry for exhalred breath analysis. Anal Bioanal Chem 396(1):471– 482
- 17. Maddula S, Blank L, Schmid A, Baumbach JI (2009) Detection of volatile metabolites of Escherichia coli by multi capillary column coupled ion mobility spectrometry. Anal Bioanal Chem 394 (3):791–800
- 18. Baumbach JI (2009) Ion Mobility Spectrometry coupled with Multi-Capillary Columns for Metabolic Profiling of Human Breath. J Breath Res 3:1–16
- 19. Bödeker B, Vautz W, Baumbach JI (2008) Peak Comparison in MCC/IMS—Data—Searching for potential biomarkers in human breath data. Int J Ion Mobil Spec 11(1):89–93
- 20. Baumbach JI, Westhoff M (2006) Ion mobility spectrometry to detect lung cancer and airway infections. Spectrosc Eur 18(6):22–27
- 21. Borsdorf H, Mayer T, Zarejousheghani M, Eiceman GA (2011) Recent Developments in Ion Mobility Spectrometry. Appl Spectrosc Rev 46(6):472–521. doi[:10.1080/05704928.2011.582658](http://dx.doi.org/10.1080/05704928.2011.582658)
- 22. Eiceman GA, Karpas Z (2005) Ion Mobility Spectrometry, 2nd edn. CRC, Boca Raton
- 23. Perl T, Bödecker B, Jünger M, Nolte J, Vautz W (2010) Alignment of retention time obtained from multicapillary column gas chromatography used for VOC analysis with ion mobility spectrometry. Anal Bioanal Chem 397(6):2385–2394
- 24. Viitanen AK, Mattila T, Makela JM, Marjamaki M, Anttalainen O, Keskinen J (2008) Experimental study of the effect of temperature on ion cluster formation using ion mobility spectrometry. Atmos Res 90(2–4):115–124
- 25. Barnett DA, Belford M, Dunyach J-J, Purves RW (2007) Characterization of a Temperature-Controlled FAIMS System. J Am Soc Mass Spectr 18(9):1653–1663
- 26. Jia J, Guo HY, Gao XG, He XL, Li JP (2006) Effects of drift tube temperature on ion mobility spectra. Chin J Anal Chem 34 (12):1783–1786
- 27. Tabrizchi M (2004) Temperature effects on resolution in ion mobility spectrometry. Talanta 62(1):65-70
- 28. Tabrizchi M (2002) Temperature Corrections for Ion Mobility Spectrometry. Int J Ion Mobil Spec 5(1):59–62
- 29. Ewing RG, Miller CJ (2001) The effects of temperature on the detection of volatile vapors emitted from explosives using ion mobility spectrometry. Int J Ion Mobil Spec 3(1):56
- 30. Eiceman GA, Nazarov EG, Rodriguez JE (2001) Chemical class information in ion mobility spectra at low and elevated temperatures. Anal Chim Acta 433:53–70
- 31. Eiceman GA, Nazarov EG, Rodriguez JE, Bergloff JF (1998) Positive Reactant Ion Chemistry for Analytical, High Temperature Ion Mobility Spectrometry (IMS): Effects of Electric Field of the Drift Tube and Moisture, Temperature, and Flow of the Drift Gas. Int J Ion Mobil Spec 1(1):28–37
- 32. Chen YH, Hill HH Jr, Wittmer DP (1996) Thermal effects on electrospray ionization ion mobility spectrometry. Int J Mass Spectr Ion Proc 154:1–13
- <span id="page-5-0"></span>33. Eiceman G, Nazarov E, Rodriguez J, Stone J (2001) Analysis of a drift tube at ambient pressure: Models and precise measurements in ion mobility spectrometry. Rev Sci Instrum 72(9):3610–3621
- 34. Borsdorf H, Mayer T (2010) Electric field dependence of ion mobilities of aromatic compounds with different ionic mass and different functional groups. Int J Ion Mobil Spec 13(3–4):103–108
- 35. Soppart O, Baumbach JI (2000) Comparison of electric fields within drift tubes for ion mobility spectrometry. Meas Sci Technol 11 (10):1473–1479
- 36. Guevremont R, Purves R (2005) Comparison of experimental and calculated peak shapes for three cylindrical geometry FAIMS

prototypes of differing electrode diameters. J Am Soc Mass Spectr 16(3):349–362

- 37. Young D, Eiceman GA, Breach J, Brittain AH, Thomas CLP (2002) Automated control and optimisation of ion mobility spectrometry responses using a sheath-flow inlet. Anal Chim Acta 463(2):143–154
- 38. Bödeker B, Baumbach J (2009) Analytical description of IMS-signals. Int J Ion Mobil Spec 12(3):103–108. doi:[10.1007/s12127-009-0024-y](http://dx.doi.org/10.1007/s12127-009-0024-y)
- 39. Bödeker B, Vautz W, Baumbach JI (2008) Peak Finding and Referencing in MCC/IMS—Data. Int J Ion Mobil Spec 11(1–4):83–88
- 40. Bödeker B, Vautz W, Baumbach JI(2008) Visualisation of MCC/IMS— Data. Int J Ion Mobil Spec 11(1):77–82