**RESEARCH ARTICLE** 



## Cytotoxic T lymphocyte antigen-4 (CTLA-4) expression in chordoma and tumor-infiltrating lymphocytes (TILs) predicts prognosis of spinal chordoma

G.  $He^{1} \cdot X$ .  $Liu^{1} \cdot X$ .  $Pan^{1} \cdot Y$ .  $Ma^{2} \cdot X$ .  $Liu^{1}$ 

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### Abstract

**Purpose** Chordoma is a rare tumor of the skeletal system that is characterized by a high recurrence rate and treatment resistance. Given the common finding of immune dysregulation in chordoma, immunotherapy has emerged as potential treatment option. As an important immune checkpoint regulator, we evaluated cytotoxic T-lymphocyte antigen-4 (CTLA-4) expression and its prognostic significance for patients with chordoma of the spine.

**Methods** CTLA-4 expression was analyzed immunohistochemically in 32 chordoma tissues and 14 nucleus pulposus tissues to examine the specificity of CTLA-4 expression in chordoma. Univariate log-rank analysis was used to evaluate the association of CTLA-4 expression in tumor cells and tumor-infiltrating lymphocytes (TILs) with survival. Cox multivariate analysis was used to identify independent factors of survival.

**Results** Positive CTLA-4 expression was observed in all of the TILs and tumor cell cytoplasm, and partial in the membrane or in both the membrane and nucleus, with a markedly higher positivity rate than that observed in normal nucleus tissues. Higher CTLA-4 expression in the tumor but not in TILs was significantly associated with shorter continuous disease-free survival (CDFS) and overall survival (OS). CTLA-4 expression in tumor cells and TILs were independent predictors for CDFS, whereas only tumor cell expression was a significant predictor of OS. Furthermore, the combination of CTLA-4 expression in the tumor and TILs had higher prognostic value.

Conclusions Targeting CTLA-4 may be a potential novel therapeutic strategy for chordoma patients.

Keywords CTLA-4 · Immunohistochemistry · Chordoma · TILs · Prognosis

### Abbreviations

Continuous disease-free survival
Cytotoxic T lymphocyte antigen 4
Diaminobenzidine
Favorable CTLA-4 expression profile
Immunohistochemistry

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X. Liu xglius@vip.sina.com

<sup>1</sup> Department of Orthopedics, North Garden Street, Peking University Third Hospital, No. 49, Haidian District, Beijing 100191, People's Republic of China

<sup>2</sup> The Center for Pain Medicine, North Garden Street, Peking University Third Hospital, No. 49, Haidian District, Beijing 100191, China

LSS	Lumbar spinal stenosis
LDH	Lumbar disk herniation
OS	Overall survival
PD-1	Programmed cell death protein 1

TILs Tumor-infiltrating lymphocytes

### Introduction

Chordoma is an extremely rare malignant mesenchymal tumor of the skull base and axial skeleton that is considered to originate from notochordal remnants. Chordoma is typically characterized by slow growth and local invasion, accounting for 1–4% of primary malignant bone tumors and up to 20% of all spine tumors [1]. Current clinical treatments for chordoma include en-bloc resection combined with radiotherapy before and after surgery [2]. However, the predilection site of chordoma proximal to vital neurovascular structures and invasive growth into surrounding

soft tissue pose a challenge in achieving complete resection, ultimately leading to tumor recurrence [3, 4]. Moreover, in contrast to other tumors that are relatively sensitive to chemotherapy and radiotherapy, chordoma frequently shows drug resistance and radiotherapy insensitivity, hindering successful treatment [5]. Therefore, it is critical to obtain a better understanding of the biological properties of chordoma so as to develop novel therapeutic strategies that can improve treatment outcomes for patients.

Tumor-related immune dysregulation has recently been identified as a key feature of chordoma, leading to the consideration of immunotherapy as a promising therapeutic strategy [5]. Among available immunotherapy strategies, immune checkpoint blockades have shown good success in the treatment of various malignancies [6]. The mostly common and important immune checkpoint molecules include programmed cell death protein 1 (PD-1) and cytotoxic T lymphocyte antigen 4 (CTLA-4), and clinical trials of monoclonal antibodies targeting PD-1 and CTLA-4 have shown effectiveness for a variety of tumors [7–9]. Recent studies have also reported associations of the expression of PD-1 or its ligand PD-L1 with the metastatic potential of chordoma [10, 11]. However, to our knowledge, no study has explored the prognostic value of CTLA-4 expression in chordoma and its corresponding tumor-infiltrating lymphocytes (TILs) to date.

CTLA-4 is a well-known immune checkpoint protein that is commonly expressed on T cells, which is a CD28 homolog but shows much higher binding affinity for B7 ligands [12], and is mainly localized in the intracellular compartment of resting naive T cells. However, upon T cell activation, CTLA-4 expression is upregulated and the protein is translocated to the cell surface where it outcompetes with CD28 for B7 ligands, thereby antagonizing the effector T cell response by disturbing the cell cycle and cytokine release [13, 14]. Therefore, the relative binding of CTLA-4 and CD28 to B7 determines whether T cells will be activated or silenced, and results in regulating the immune environment.

CTLA-4 expression has also been found in various types of tumor tissues, including tumors of the hematological system [15] and solid tumors [16]. The differential expression of CTLA-4 in tumors and TILs compared to normal cells suggests that CTLA-4 could be a potential cancer marker; however, related results are controversial. Most studies indicated that high CTLA-4 expression in tumors and TILs was associated with a poorer patient prognosis, including for patients with breast cancer [17], nasopharyngeal carcinoma [18], and thymoma [19], suggesting that CTLA-4 overexpression may facilitate tumors in evading the host immune response. However, other studies showed that high CTLA-4 expression in tumors and TILs was related with a good clinical outcome and survival rate for patients with mesothelioma [20] and non-small cell lung cancer [21], indicating that CTLA-4 overexpression may enhance anti-tumor immunity. These contradictory findings might indicate different roles of CTLA-4 in different tumor types, leading to tumor-specific effects on clinical outcome.

Considering this background, we sought to identify whether CTLA-4 expression can be a prognostic marker and potential therapeutic target for chordoma. Toward this end, we evaluated the CTLA-4 expression pattern in chordoma and normal nucleus pulposus tissues, and investigated the association between CTLA-4 expression and survival time as well as clinicopathological features of patients with chordoma of the spine.

### **Patients and methods**

#### Patients and data collection

A total of 32 chordoma patients (23 men and 9 women) with a mean age of 50.53 years (range, 11–70 years) were enrolled in the study. All patients had undergone primary tumor resection at the Department of Orthopedics, Peking University Third Hospital from 2006 to 2019. 14 normal nucleus pulposus tissues as control samples were acquired from patients with lumbar spinal stenosis or lumbar disk herniation. Inclusion criteria were pathologically confirmed chordoma with availability of paraffin-embedded specimens of the primary tumor and relatively complete follow-up data. Patients were excluded if they had undergone any anti-cancer treatments prior to surgery, such as chemotherapy or radiotherapy, which may have an influence on the expression of proteins interest.

We retrospectively reviewed the clinicopathological characteristics, including age, gender, tumor size, tumor location, tumor stage, tumor type, surrounding muscle invasion, recurrence, extent of TILs, and Ki67 staining index. Resected tumor specimens were evaluated by anatomic pathologists and data were recorded based on the Enneking principles [22]. Surrounding muscle invasion of the tumor was verified by magnetic resonance images and histological analysis, and tumor recurrence was diagnosed according to clinical symptoms and imaging examination during followup [23]. Events were defined as the first evidence of local recurrence or distant metastasis for continuous disease-free survival (CDFS) and as death related to any cause for overall survival (OS). Patients were followed up every 3 months for the first 2 years, at 6-month intervals for the third year, and then annually until death. All patients were followed until May 2019 and observations were censored when a patient was disease-free or alive at the time of the last clinical follow-up (May 2019).

#### Immunohistochemistry

Paraffin-embedded tissue Sects. (5  $\mu$ m thick) were dewaxed with xylene and subsequently hydrated with various concentrations of ethanol, followed by antigen heat repair with high-pressure Tris- ethylenediaminetetraacetic acid buffer. The sections were incubated with 3% H<sub>2</sub>O<sub>2</sub> for 15 min to prevent activity of endogenous peroxidase and then blocked with 10% normal goat serum at room temperature for 30 min to avoid interference of non-specific antigens. The sections were incubated with primary anti-CTLA-4 antibody (Abcam, ab134090, 1:300 dilution) overnight at 4 °C, followed by incubation with the secondary antibody (EnVision K500711, Dako, Denmark) at 37 °C for 30 min. The antibody portion of the sections was visualized by 3,3'-diaminobenzidine staining for 40 s and then the nucleus was counterstained with hematoxylin.

### Immunohistochemistry scoring

Two independent pathologists blinded to the clinical data evaluated the density of TILs along with CTLA-4 expression on tumor cells and TILs for each section. The immunohistochemistry staining result was scored based on the immunoreactive score (IRS) system, which takes both the intensity and percentage of stained cells into consideration. The intensity of stained cells was scored as 0 (none), 1 (weak), 2 (moderate), and 3 (strong), and the proportion of stained cells was scored as 0 (0–5%), 1 (5–25%), 2 (25–50%), and 3 (> 50%). The final scores (ranging from 0 to 12) were obtained by multiplying the intensity and percentage values [21]. The result for CTLA-4 expression was defined as negative (score 0-1), weak (score 2-4), moderate (score 5-8), and strong (score 9-12). Samples with negative and weak scores were classified in the low CTLA-4 expression group, whereas those with moderate and strong scores were classified in the high CTLA-4 expression group. The extent of TILs was assessed as absent, rare/few, moderate, and prominent. The groups with absent and rare/few TILs were considered as low intensity, whereas those with moderate and prominent TILs were considered as high intensity.

#### **Statistical analysis**

Pearson's Chi-squared test was applied to analyze the association of CTLA-4 expression in tumor tissues and TILs with the clinicopathological characteristics of chordoma patients. Univariate analysis was performed based on the Kaplan–Meier curve (log-rank test) for associations with CDFS and OS. Multivariate analysis was performed with a Cox proportional hazard model to assess whether CTLA-4 expression independently predicted patient outcome (CDFS and OS) after controlling for the significant or highly clinically related factors based on univariate analysis. SPSS 20.0 software (SPSS Inc., Chicago, IL, USA) was used for all statistical analyses, and a two-sided p value < 0.05 was considered to indicate a statistically significant effect.

### Results

# CTLA-4 expression on tumor and normal control tissues

The clinicopathological characteristics of the patients included in the study are summarized in Supplementary Table S1. TILs were detected in all the tumor samples with 7 (21.9%), rare/few in 6 samples (18.8%), moderate in 10 samples (31.3%), and prominent in 9 samples (28.1%) (Fig. 1), and CTLA-4 staining in TILs was low in 9 samples (28.1%), weak in 5 samples (15.6%), moderate in 7 samples (21.9%), and strong in 11 samples (34.4%) (Fig. 2a–d). CTLA-4 expression in the tumor was scored as low in 5 patients (15.6%), weak in 10 patients (31.3%), moderate in 2 patients (6.3%), and strong in 15 patients (46.9%) (Fig. 2e–h). In addition, among the 32 chordoma samples, CTLA-4 expression was observed either as focal dots or scattered patterns in chordoma cells with



Fig. 1 Representative illustration of tumor-infiltrating lymphocytes (TILs) in chordoma tissues by HE staining. a Absent; b Rare/few; c Moderate; d Prominent.400×magnification



**Fig.2** CTLA-4 expression in TILs and chordoma tissues. TILs: **a** low CTLA-4 staining; **b** weak CTLA-4 staining; **c** moderate CTLA-4 staining; **d** strong CTLA-4 staining; Chordoma tissues: **e** 

low CTLA-4 staining; **b** weak CTLA-4 staining; **c** moderate CTLA-4 staining; **d** strong CTLA-4 staining.  $400 \times \text{magnification}$ . Inset: a further magnification of the small black square

heterogeneous localization as follows: cytoplasm (15.6%), cytoplasm/membrane (25.0%), and cytoplasm/membrane/ nucleus (59.4%) (Fig. 3). Furthermore, significantly higher CTLA-4 expression was observed on chordoma tissues than that on the nucleus pulposus tissues (p = 0.014; Fig. 4, Supplementary Table S4).

# Association of CTLA-4 expression with clinicopathological characteristics

As shown in Table 1, Pearson's Chi-square analysis revealed that high CTLA-4 expression in the tumor (p = 0.005) and TILs (p = 0.016) was significantly



**Fig. 3** Characteristics of CTLA-4 expression on chordoma tissues. **a** CTLA-4 staining in cytoplasmic, **b** CTLA-4 staining in cytoplasmic combined with a membrane staining, **c** CTLA-4 staining in cytoplasmic combined with both membrane and nucleus staining; **d** CTLA-4

staining in chordoma tissues in a focal pattern; **e** CTLA-4 staining in chordoma tissues in a scattered pattern.  $400 \times \text{magnification}$ . Inset: a further magnification of the small black square





associated with recurrence. Moreover, high CTLA-4 expression in TILs (p = 0.002) but not in chordoma tissues (p = 0.513) was significantly correlated with the extent of TILs. However, there was no significant correlations of CTLA-4 expression in tumor cells or TILs with other clinicopathological characteristics such as surrounding muscle invasion, tumor stage, or Ki67 staining index.

# Association of prognostic factors and CTLA-4 expression with CDFS and OS

Kaplan–Meier log-rank analysis demonstrated that high CTLA-4 expression in the tumor was significantly associated with a poor CDFS ( $\chi^2 = 8.548$ , p = 0.003; Table 2, Fig. 5a) and OS ( $\chi^2 = 5.592$ , p = 0.018; Table 3, Fig. 5c), whereas CTLA-4 expression in TILs was not associated with





Fig. 5 Kaplan–Meier survival curves of the correlation between CTLA-4 expression and prognosis. (a, c) CDFS and OS of patients with high CTLA-4 expression were both shorter than that of patients

with low CTLA-4 expression in tumor cells;  $(\mathbf{b}, \mathbf{d})$  CDFS and OS of patients in the FCEP group was longer than that of the other groups

Clinicopathological factors

Sacral vertebra/Cervical or thoracic or lumbar vertebra Surrounding muscle invasio

Age (years)  $\leq 50$ > 50 Sex Male Female Tumor size  $\leq 5 \text{ cm}$ >5 cm Tumor location

Yes No Recurrence Yes No

IB

III

Low

High

Low

High

Tumor stage

Tumor type Typical

Dedifferentiated

Extent of TILs

Ki67 staining index

14

16

1

15

2

6

11

11

6

	CTLA-4 expression on tumor		<i>p</i> -value	CTLA-4 express	<i>p</i> -value	
	$\overline{\text{Low}(n\!=\!15)}$	High $(n=17)$		Low (n = 14)	High $(n=18)$	
	7	8	0.982	7	8	0.755
	8	9		7	10	
	12	11	0.337	12	11	0.125
	3	6		2	7	
	5	8	0.430	6	7	0.821
	10	9		8	11	
	4	7	0.388	3	8	0.174
	11	10		11	10	
n						
	7	5	0.314	4	8	0.358
	8	12		10	10	
	10	3	0.005	0	4	0.016

5

14

0

13

1

10

4

12

2

0.340

0.522

0.513

0.152

14

17

0

14

4

3

15

12

6

CDFS ( $\chi^2 = 1.773$ , p=0.183; Table 2) or OS ( $\chi^2 = 0.664$ , p = 0.415; Table 3). Among the other clinicopathological factors, only recurrence was significantly related to CDFS  $(\chi^2 = 4.872; p = 0.027; \text{ Table 2})$ . Multivariate analysis showed that CTLA-4 expression in tumor cells remained an independent predictor for CDFS (p = 0.007, hazard ratio [HR] = 4.393; Table 2) and OS (p = 0.026, HR = 3.319; Table 3). Moreover, after controlling for other variables, CTLA-4 expression in TILs showed a strong association with CDFS (p = 0.038, HR = 3.409; Table 2) but was still not associated with OS (p = 0.183, HR = 1.773; Table 3).

5

15

0

12

3

7

8

13

2

### Association of combined CTLA-4 expression in tumor cells and TILs with patient survival

Previous studies have highlighted the importance of combined CTLA-4 expression in tumor cells and TILs for

predicting the prognosis of tumor patients [17, 24]. Therefore, we further divided the patients into four subgroups according to various combinations of CTLA-4 expression in tumor cells and TILs: Group 1 (CTLA-4<sup>high</sup> tumor cells CTLA-4<sup>high</sup> TILs), Group 2 (CTLA-4<sup>high</sup> tumor cells CTLA-4<sup>low</sup> TILs), Group 3 (CTLA-4<sup>low</sup> tumor cells CTLA-4<sup>high</sup> TILs), and Group 4 (CTLA-4<sup>low</sup> tumor cells CTLA-4<sup>low</sup> TILs). CDFS and OS were significantly longer in Group 4 (89.563 and 91.188 months, respectively) than those in Group 1 (36.727 and 59.623 months), Group 2 (14.444 and 31.833 months), and Group 3 (41.071 and 75.743 months) (Supplementary Table S2), indicating that high CTLA-4 expression in both tumor cells and TILs is strongly associated with a poor prognosis.

0.370

0.244

0.002

0.217

Factors	CDFS				
	Univariate analysis		Multivariate analysis		
	$\chi^2$	<i>p</i> -value	HR	<i>p</i> -value	
CTLA-4 expression on tumor	8.548	0.003	4.393	0.007	
CTLA-4 expression on TILs	1.773	0.183	3.409	0.038	
FECP group	7.221	0.007	6.561	0.017	
Age	0.667	0.414			
Sex	0.007	0.953			
Tumor size	0.482	0.487			
Tumor location	0.678	0.407			
Surrounding muscle invasion	1.604	0.205			
Recurrence	4.872	0.027			
Tumor stage	0.035	0.853			
Tumor type	0.207	0.649			
Extent of TILS	0.254	0.110			
Ki67 staining index	0.003	0.955			

 
 Table 2
 Univariate and multivariate Cox proportional hazard analyses of prognostic factors for continuous disease-free survival time and overall survival time of spinal chordoma patients

 
 Table 3
 Univariate and multivariate Cox proportional hazard analyses of prognostic factors for overall survival time of spinal chordoma patients

Factors	OS				
	Univari analysis	ate	Multivariate analysis		
	χ2	<i>p</i> -value	HR	<i>p</i> -value	
CTLA-4 expression on tumor	5.592	0.018	3.319	0.026	
CTLA-4 expression on TILs	0.664	0.415	1.773	0.183	
FECP group	4.213	0.040	4.185	0.060	
Age	1.149	0.284			
Sex	0.303	0.582			
Tumor size	0.177	0.732			
Tumor location	0.172	0.679			
Surrounding muscle invasion	0.162	0.687			
Recurrence	1.898	0.168			
Tumor stage	0.005	0.945			
Tumor type	0.001	0.980			
Extent of TILS	0.476	0.490			
Ki67 staining index	0.026	0.871			

### Association of combined CTLA-4 expression in tumor cells and TILs with clinicopathological characteristics and prognosis

Based on the results described above, we defined Group 4 as the favorable CTLA-4 expression profile (FCEP) group, and Groups 1, 2, and 3 were combined as the Other

group. Pearson's Chi-squared analysis showed no significant differences in clinical characteristics between these two groups except for recurrence (p = 0.002) and extent of TILs (p = 0.022) (Supplementary Table S3). Univariate logrank analysis showed that the CDFS ( $\chi^2 = 7.221$ , p = 0.007; Table 2; Fig. 5b) and OS ( $\chi^2 = 4.213$ , p = 0.04; Table 3, Fig. 5d) of the FCEP group were longer than those of the Other group. Multivariate analysis further demonstrated that the FCEP group was an independent predictor for improved CDFS (HR=6.561, p=0.017; Table 2) and OS (HR=4.185, p=0.06; Table 3).

### Discussion

Chordoma is a rare bone tumor characterized by local invasion and poor prognosis [25]. Currently, maximal tumor resection combined with adjuvant radiotherapy is considered the standard treatment for chordoma patients. However, frequent recurrence remains a major limitation to effective chordoma treatment despite wide resection and regular radiotherapy [26]. Therefore, searching for molecular biomarkers that can specifically target chordoma is an important research direction to improve the prediction of prognosis and development of molecular targeted drugs for chordoma patients.

In recent years, an increasing number of molecular biomarkers have been evaluated with respect to potential associations with prognosis of patients with spinal chordoma [27]. However, most of these prognostic biomarkers showed no significant associations after controlling for other potentially confounding factors in multivariate analysis. Thus, we sought to identify new biological markers that can independently predict the prognosis of chordoma patients. We found heterogeneous localization patterns of CTLA-4 expression in chordoma cells in accordance with previous studies [17, 28]. Our finding is consistent with the commonly accepted view that CTLA-4 is mainly localized in the sub-membrane vesicles (cytoplasm) similar to the Golgi apparatus, and will cycle from intracellular stores to a specific position upon activation. However, the relationship between the heterogeneous localization patterns of CTLA-4 expression and the prognosis of chordoma patients requires further investigation to better understand its role in cancer development and progression.

Although previous studies have demonstrated CTLA-4 overexpression in the tumor microenvironment, the relationship of CTLA-4 expression in tumor cells and TILs with patient prognosis remains controversial. The majority of related studies indicated that high CTLA-4 expression was associated with a poor prognosis of tumor patients, and antibodies for targeting CTLA-4 or in combination with other therapies could enhance the tumor-killing effect and

improve the survival rate of patients [29-31]. However, other studies conversely reported that CTLA-4 overexpression was related with an improved prognosis [32, 33]. Such discrepancy among studies suggests that CTLA-4 expression might have distinct clinical outcomes in different tumor types. In our study, both univariate and multivariate analyses demonstrated that high CTLA-4 expression in chordoma cells was associated with a poor CDFS and OS, suggesting that CTLA-4 overexpression in the tumor and TILs might result in an immunosuppressive tumor microenvironment. Two recent studies in breast cancer and esophageal cancer demonstrated that CTLA-4 expression in the tumor combined with that in TILs was a stronger predictor for patient prognosis [17, 24]. Similarly, in the present study, although tumor CTLA-4 expression did not differ from that in TILs, the HR for CDFS and OS of the FECP group (characterized by low CTLA-4 expression in both the tumor and TILs) was much higher than that of other groups, suggesting that the combination of high CTLA-4 expression in chordoma and TILs is also an even stronger poor predictor for chordoma patient prognosis.

Considering these results, we speculate that CTLA-4 expression in chordoma tumor cells or TILs may synergistically damage the function of activated tumor-specific T cells. In addition, CTLA-4 expression was much more frequent and stronger in chordoma than that in normal nucleus pulposus tissues, indicating that CTLA-4 overexpression might contribute to tumorigenesis. Nevertheless, the specific function of CTLA-4 and the underlying regulatory mechanism in chordoma require further experimental clarification.

### Conclusion

To our knowledge, this is the first study to demonstrate an association of CTLA-4 expression in chordoma tissues and TILs with patient prognosis. Our results revealed that high CTLA-4 expression in tumor tissues or TILs was related with poor survival of chordoma patients and their combination was an even stronger poor prognostic predictor (see Schematic 1). These findings suggest that CTLA-4 blockade might be a promising immunotherapeutic strategy for treatment of chordoma patients. Nevertheless, as this was a single-center study with a relatively small sample and lack of functional assays, further in vitro and in vivo studies are needed to verify the role of CTLA-4 in chordoma. In particular, multicenter studies with larger patient cohorts will help to obtain a more comprehensive understanding of the biological function and underlying mechanism of abnormal CTLA-4 expression in chordoma and its clinical relevance.

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Author contributions Conceived and designed the experiments: GH, XL. Performed the experiments: GH, XP, XL, YM. Analyzed the data: GH, XP, XL, XL. Wrote the paper: GH, XL.

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### **Compliance with ethical standards**

**Conflict of interest** The authors declare that they have no conflict of interest.

**Ethical approval** The study was approved by the ethics committee of Peking University Third Hospital (No. IRB00006761-2016048).

**Consent to participate** Informed consent for use of their specimens and data in this study was obtained from all adult patients or written parental consent was provided for minors before the study was conducted.

Availability of data and materials The data and materials are available from the corresponding author on reasonable request.

### References

- Matsumoto T, Imagama S, Ito Z, Imai R, Kamada T, Shimoyama Y, Matsuyama Y, Ishiguro N. Total spondylectomy following carbon ion radiotherapy to treat chordoma of the mobile spine. Bone Joint J. 2013;95(10):1392–5.
- Park L, Delaney TF, Liebsch NJ, Hornicek FJ, Goldberg S, Mankin H, Rosenberg AE, Rosenthal DI, Suit HD. Sacral chordomas: impact of high-dose proton/photon-beam radiation therapy combined with or without surgery for primary versus recurrent tumor. Int J Radiat Oncol Biol Phys. 2006;65(5):1514–21.
- Tzortzidis F, Elahi F, Wright D, Natarajan SK, Sekhar LN. Patient outcome at long-term follow-up after aggressive microsurgical resection of cranial base chordomas. Neurosurgery. 2006;59(2):230–7.
- Kayani B, Hanna SA, Sewell MD, Saifuddin A, Molloy S, Briggs TW. A review of the surgical management of sacral chordoma. Eur J Surg Oncol. 2014;40(11):1412–20.
- Patel SS, Schwab JH. Immunotherapy as a potential treatment for chordoma: a review. Curr Oncol Rep. 2016;18(9):55.
- Bordon Y. Immunotherapy: Checkpoint parley. Nat Rev Cancer. 2015;15(1):3.
- Thomas A, Hassan R. CTLA-4 blockade in mesothelioma: ineffective or reason for optimism? Lancet Oncol. 2017;18(9):1150–1.
- Boutros C, Tarhini A, Routier E, Lambotte O, Ladurie FL, Carbonnel F, Izzeddine H, Marabelle A, Champiat S, Berdelou A, Lanoy E, Texier M, Libenciuc C, Eggermont AM, Soria JC, Mateus C, Robert C. Safety profiles of anti-CTLA-4 and anti-PD-1 antibodies alone and in combination. Nat Rev Clin Oncol. 2016;13(8):473–86.
- Migliorini D, Mach N, Aguiar D, Vernet R, Landis BN, Becker M, McKee T, Dutoit V, Dietrich PY. First report of clinical responses to immunotherapy in 3 relapsing cases of chordoma after failure of standard therapies. Oncoimmunology. 2017;6(8):e1338235.

- Feng Y, Shen J, Gao Y, Liao Y, Cote G, Choy E, Chebib I, Mankin H, Hornicek F, Duan Z. Expression of programmed cell death ligand 1 (PD-L1) and prevalence of tumor-infiltrating lymphocytes (TILs) in chordoma. Oncotarget. 2015;6(13):11139–49.
- Feng Y, Shen J, Gao Y, Liao YG, Cote G, Choy E, Chebib I, Mankin H, Hornicek F, Duan ZG. Expression of programmed cell death ligand 1 (PD-L1) and prevalence of tumor-in ltrating lymphocytes (TILs) in chordoma. Oncotarget. 2015;6(13):11139–49.
- Brunet JF, Denizot F, Luciani MF, Roux-Dosseto M, Suzan M, Mattei MG, Golstein P. A new member of the immunoglobulin superfamily–CTLA-4. Nature. 1987;328(6127):267–70.
- 13. Greenwald RJ, Latchman YE, Sharpe AH. Negative co-receptors on lymphocytes. Curr Opin Immunol. 2002;14(3):391–6.
- 14. Sharpe AH, Freeman GJ. The B7-CD28 superfamily. Nat Rev Immunol. 2002;2(2):116–26.
- Mittal AK, Chaturvedi NK, Rohlfsen RA, Gupta P, Joshi AD, Hegde GV, Bociek RG, Joshi SS. Role of CTLA4 in the proliferation and survival of chronic lymphocytic leukemia. PLoS ONE. 2013;8(8):e70352.
- Menon S, Shin S, Dy G. Advances in cancer immunotherapy in solid tumors. Cancers (Basel). 2016. https://doi.org/10.3390/cance rs8120106.
- Yu H, Yang J, Jiao S, Li Y, Zhang W, Wang J. Cytotoxic T lymphocyte antigen 4 expression in human breast cancer: implications for prognosis. Cancer Immunol Immunother. 2015;64(7):853–60.
- Huang PY, Guo SS, Zhang Y, Lu JB, Chen QY, Tang LQ, Zhang L, Liu LT, Zhang L, Mai HQ. Tumor CTLA-4 overexpression predicts poor survival in patients with nasopharyngeal carcinoma. Oncotarget. 2016;7(11):13060–8.
- Santoni G, Amantini C, Morelli MB, Tomassoni D, Santoni M, Marinelli O, Nabissi M, Cardinali C, Paolucci V, Torniai M, Rinaldi S, Morgese F, Bernardini G, Berardi R. High CTLA-4 expression correlates with poor prognosis in thymoma patients. Oncotarget. 2018;9(24):16665–777.
- Roncella S, Laurent S, Fontana V, Ferro P, Franceschini MC, Salvi S, Varesano S, Boccardo S, Vigani A, Morabito A, Canessa PA, Giannoni U, Rosenberg I, Valentino A, Fedeli F, Merlo DF, Ceppi M, Riggio S, Romani M, Saverino D, Poggi A, Pistillo MP. CTLA-4 in mesothelioma patients: tissue expression, body fluid levels and possible relevance as a prognostic factor. Cancer Immunol Immunother. 2016;65(8):909–17.
- 21. Salvi S, Fontana V, Boccardo S, Merlo DF, Margallo E, Laurent S, Morabito A, Rijavec E, Dal Bello MG, Mora M, Ratto GB, Grossi F, Truini M, Pistillo MP. Evaluation of CTLA-4 expression and relevance as a novel prognostic factor in patients with non-small cell lung cancer. Cancer Immunol Immunother. 2012;61(9):1463–72.
- Samson IR, Springfield DS, Suit HD, Mankin HJ. Operative treatment of sacrococcygeal chordoma. A review of twenty-one cases. J Bone Joint Surg Am. 1993;75(10):1476–84.
- 23. Meng T, Yin H, Li B, Li Z, Xu W, Zhou W, Cheng M, Wang J, Zhou L, Yang X, Liu T, Yan W, Song D, Xiao J. Clinical features

and prognostic factors of patients with chordoma in the spine: a retrospective analysis of 153 patients in a single center. Neuro Oncol. 2015;17(5):725–32.

- Zhang XF, Pan K, Weng DS, Chen CL, Wang QJ, Zhao JJ, Pan QZ, Liu Q, Jiang SS, Li YQ, Zhang HX, Xia JC. Cytotoxic T lymphocyte antigen-4 expression in esophageal carcinoma: implications for prognosis. Oncotarget. 2016;7(18):26670–9.
- 25. Walcott BP, Nahed BV, Mohyeldin A, Coumans JV, Kahle KT, Ferreira MJ. Chordoma: current concepts, management, and future directions. Lancet Oncol. 2012;13(2):e69–76.
- Zou MX, Lv GH, Wang XB, Li J. Letter: factors predicting recurrence after resection of clival chordoma using variable surgical approaches and radiation modalities. Neurosurgery. 2017;81(2):E28–E31.
- Ferraresi V, Nuzzo C, Zoccali C, Marandino F, Vidiri A, Salducca N, Zeuli M, Giannarelli D, Cognetti F, Biagini R. Chordoma: clinical characteristics, management and prognosis of a case series of 25 patients. BMC Cancer. 2010;10:22.
- Paulsen EE, Kilvaer TK, Rakaee M, Richardsen E, Hald SM, Andersen S, Busund LT, Bremnes RM, Donnem T. CTLA-4 expression in the non-small cell lung cancer patient tumor microenvironment: diverging prognostic impact in primary tumors and lymph node metastases. Cancer Immunol Immunother. 2017;66(11):1449–611.
- Zhao Y, Yang W, Huang Y, Cui R, Li X, Li B. Evolving roles for targeting CTLA-4 in cancer immunotherapy. Cell Physiol Biochem. 2018;47(2):721–34.
- Wang C, Xu L, Liang C, Xiang J, Peng R, Liu Z. Immunological responses triggered by photothermal therapy with carbon nanotubes in combination with anti-CTLA-4 therapy to inhibit cancer metastasis. Adv Mater. 2014;26(48):8154–62.
- Smyth MJ, Yagita H, McArthur GA. Combination anti-CTLA-4 and anti-RANKL in metastatic melanoma. J Clin Oncol. 2016;34(12):e104–e106106.
- 32. Erfani N, Mehrabadi SM, Ghayumi MA, Haghshenas MR, Mojtahedi Z, Ghaderi A, Amani D. Increase of regulatory T cells in metastatic stage and CTLA-4 over expression in lymphocytes of patients with non-small cell lung cancer (NSCLC). Lung Cancer. 2012;77(2):306–11.
- 33. Kim JW, Nam KH, Ahn SH, Park DJ, Kim HH, Kim SH, Chang H, Lee JO, Kim YJ, Lee HS, Kim JH, Bang SM, Lee JS, Lee KW. Prognostic implications of immunosuppressive protein expression in tumors as well as immune cell infiltration within the tumor microenvironment in gastric cancer. Gastric Cancer. 2016;19(1):42–52.

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