REVIEW ARTICLE



Altered expression and functional role of ion channels in leukemia: bench to bedside

H. Rafieemehr¹ · A. Samimi² · M. Maleki Behzad³ · M. Ghanavat⁴ · S. Shahrabi⁵

Received: 27 February 2019 / Accepted: 26 May 2019 / Published online: 6 July 2019 © Federación de Sociedades Españolas de Oncología (FESEO) 2019

Abstract

Leukemic cells' (LCs) survival, proliferation, activation, differentiation, and invasiveness/migration can be mediated through the function of cation and anion channels that are involved in volume regulation, polarization, cytoskeleton, and extracellular matrix reorganization. This study will review the expression of ion channels in LCs and their possible function in leukemia progression. We searched relevant literature by a PubMed (2002–2019) of English-language literature using the terms "ion channels", "leukemia", "proliferation", "differentiation", "apoptosis", and "migration". Altered expression and dysfunction of ion channels can have a strong impact on hematopoietic cell and LCs physiology and signaling, which contributes to the vital processes such as proliferation, differentiation, and apoptosis. Indeed, it can be stated that changing expression of ion channels can affect the onset and progression as well as clinical features and therapeutic responses of leukemia via inducing the maintenance of LCs. Since ion channels are membrane proteins, they can be easily accessible in LCs for understanding their influence on leukemia progression. On the other hand, ion channels can be new potential targets for chemotherapeutic agents, which may open a novel clinical and pharmaceutical field in leukemia therapy.

Article highlights

- Ion channels may be implicated in leukemia as the genes subjected to altered expression.
- Expression and function of ion channels can regulate proliferation, activation, differentiation, malignant progression, and invasiveness/migration of leukemic cells.
- Ion channels can be used as pharmacologic targets in leukemia therapy.

Keywords Ion channels · Leukemia · Proliferation · Differentiation · Apoptosis · Migration

S. Shahrabi sshahrabi45@yahoo.com

- ¹ Department of Medical Laboratory Sciences, School of Paramedicine, Hamadan University of Medical Sciences, Hamadan, Iran
- ² Department of Pharmacology and Toxicology, School of Pharmacy, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran
- ³ Thalassemia and Hemoglobinopathy Research Center, Health Research Institute, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

- ⁴ Child Growth and Development Research Center, Isfahan University of Medical Sciences, Isfahan, Iran
- ⁵ Department of Biochemistry and Hematology, Faculty of Medicine, Semnan University of Medical Sciences, Semnan, Iran

Introduction

Ion channels are membrane proteins that play an essential role in preserving the normal physiology of cells by participating in processes such as neurotransmission, heart rate, muscle contraction, insulin secretion, immunity, and cellular proliferation [1]. The most important feature of ion channels is their selective permeability that enables only certain types of ions with a given charge and size to pass through. In addition, the opening and closing of these channels are influenced by various stimuli such as voltage, elongation and pressure applied to the membrane, or by connecting to certain effectors [2]. The trafficking of ions by ion channels creates an electric potential difference on two sides of the membrane, which is necessary to produce various intracellular signals involved in processes such as mitosis and cell migration [2]. Evidence suggests that abnormal expression and function of almost all types of ion channels can contribute to the development of tumors, metastasis, and drug resistance of solid tumors and leukemia [3]. Volatility and mitotic control points that play a significant role in survival and proliferation of tumor cells are processes affected by the aberrant expression and stable properties of ion channels [4–6]. Leukemias are a group of hematological malignancies originating from hematopoietic stem cells (HSCs) of the bone marrow (BM) that are associated with an increase of immature precursors in peripheral blood (PB). These malignancies are clinically divided into acute and chronic leukemias and originate from myeloid and lymphoid lineages [7]. Although cytogenetic alterations such as mutations and translocations are a major cause of leukemia, changes in the function and expression of ion channels that are involved in the control of proliferation, apoptosis, and migration of leukemic cells (LCs) by ions passage can play a crucial role in the progression of leukemias [3]. Ion channels appear to trigger a cascade of intracellular signals affecting vital processes of LC growth through the passage of certain ions capable of producing macromolecular complexes with intracellular proteins [8, 9]. Cl⁻, K⁺, Ca²⁺, and Na⁺ are the most important ions associated with the regulation of physiological processes in cells. It has already been shown that changing expression and function of channels transmitting these ions are major causes of non-differentiation and increased migration of LCs as well as drug resistance in leukemia [10]. However, it has also been shown that targeting ion channels with anti-cancer compounds and modulation of their function can be associated with desirable therapeutic results [11], which highlight the vital role of ion channels in the development and progression of leukemia.

Over the past decade, studies have focused on the role of ion channels in excitatory cells (neurons, cardiac, secretive). A large number of investigations have focused on the role of these channels in the behavior of cancer cells in solid tumors, indicating that overexpression of some of these channels is related to poor prognosis [12–15]. Thus, ion channels can be considered as new biomarkers for the diagnosis and treatment of leukemia. Therefore, in this article, we try to discuss the effects of onset and progression of these malignancies by reviewing the most important changes in the expression and function of ion channels in leukemia.

Expression and functional role of ion channels in HSCs and leukemic stem cells

CD34⁺HSCs are cells capable of propagating and differentiating into various cell types residing in BM niches that can be linked to BM matrix through ion channels [16]. Fluctuations in volume and membrane potential are characteristics of HSCs, which can be associated with the passage of various stages of cell cycle including G1 transition to S phase [17]. Transient receptor potential (TRP) proteins are a family of Ca²⁺ transducer channels that can form a cascade of intracellular signals, including apoptosis-related signals in cells via increasing penetration of Ca²⁺ into HSCs [17, 18]. TRPM2 is a member of this family that is activated in response to oxidative stress and tumor necrosis factor alpha (TNF- α), inducing apoptosis by transferring Ca²⁺ into HSCs as well as increasing the levels of caspase-8, -9, -3, and -7 [18]. It can be inferred that decreasing expression of TRPM2 channels in normal HSCs can be a reason for resistance to apoptosis, uncontrolled proliferation, and transformation into leukemic stem cells (LSCs), which requires further studies to provide convincing evidence in this regard. LSCs are the first malignant cells in leukemia, which have a potent self-renewal and proliferation potential that plays an essential role in the onset and increase of LC populations. Li et al. showed that the human ether-a-go-related (herg) gene encodes a family of voltage-dependent K⁺ channels called HERG K⁺ channels at the surface of CD34⁺/ CD38⁻/CD123^{high} LCs, K562 cells [a chronic myeloid leukemia (CML) cell line], cellular and HL60 leukemia cell lines while normal CD34⁺/CD38⁻ HSCs lack these channels [19, 20]. Although the possible relationship between the expressions of HERG K⁺ channels with occurrence of cytogenetic changes and clinical findings has not yet been determined, the use of the HERG K⁺ channel inhibitors associated with the inhibition of G1/S transition phase of cell cycle has highlighted the role of these ion channels in inducing cell cycle and proliferation of LSCs [19, 20]. In addition, increased expression of HERG1 K⁺ channels on the surface of primary acute myeloid leukemia (AML) blast cells and immature neoplastic B-chronic lymphoblastic leukemia (B-CLL) cells (CD5⁺) shows that the aberrant expression of these ion channels can induce oncogenicity in LSCs

[10]. In this regard, increasing electrical potential of plasma membrane ($V_{\rm m}$) by HERG K⁺ channels has been introduced as a possible mechanism for inducing proliferation in tumor cells. Moreover, studies have shown that the progression of cell cycle and mitosis is strongly related to the number and function of HERG K⁺ channels [21]. However, reducing the expression and blocking HERG1 K⁺ channels in K562, CEM, and U937 by HERG channel blockers such as E-4031 and inducing apoptosis in them are indicative of the fact that these ion channels can be diagnostic biomarkers and potential targets for controlling the progression of leukemia and solid tumors [10, 19, 22, 23].

BM mesenchymal stem cell induction of ion channel expression and migration in LCs

Migration is the most important physiological process of LCs, which is related to the ability of these cells to leave BM and accumulate in PB [24]. BM mesenchymal stem cells (BM-MSCs) that are capable of differentiation to a variety of cells are among the most basic components of BM niches and umbilical cord blood [25–27]. Furthermore, BM-MSCs can be involved in the secretion of chemokines such as stromal cell-derived factor-1 (SDF-1) that play a vital role in migration or homing of LCs by binding to chemokine receptor CXC4 (CXCR4) at the surface of these cells [28]. It has been shown that SDF-1/CXCR4 interaction can induce HERG1 K⁺ channels at the surface of B-ALL LCs, which enhance their anti-apoptotic property (Fig. 1) [29]. The molecular mechanism proposed for this feature is cascade activation of intracellular signals by HERG1 K⁺ channels, which forms the β 1 integrin subunit and CXCR4 complex in B acute lymphoblastic leukemia (B-ALL) while normal B lymphocytes lack this complex [29]. It has been indicated that these complexes are specific for LCs and that their formation activates the intracellular signals of Flt-1-dependent pathway in AML-LCs, leading to increase of their excretion from BM to PB and enhancement of resistance to chemotherapy (Fig. 1) [6, 29]. Similar to B-ALL LCs, the activation of MAP kinase and phosphoinositide 3-kinase (PI3 K)/Akt pathways is a molecular mechanism resulting from CXCR4/SDF-1 interaction and increased expression of HERG1 K⁺ channels with anti-apoptotic effect and drug resistance in AML, CML and CLL [30, 31]. Interestingly, studies have shown that blocking HERG1 K⁺ channels can induce SDF-1 as well as migration of leukemic blasts in HL-60 cell lines [32]. In fact, these findings indicate that overexpression of HERG1 K⁺ channels could be associated with poor prognosis via the development of drug resistance and migration of LCs. Also, expression induction of HERG1 K⁺ channels in LCs has been reported as one of the basic mechanisms for protecting LCs against cytotoxic effects of asparaginase as well as other chemotherapeutic agents such as cytarabine (Ara-C) and etoposide (VP-16) [33]. Therefore, blocking HERG1 K⁺ channels and CXCR4/ SDF-1 interaction, which has been reported in previous studies as an approach to overcome resistance to chemotherapy in AML and ALL [34], may be related to increased LC apoptosis, decreased migration, and drug resistance of these cells. In this regard, various HERG1 inhibitors are known to control the function of HERG1 K⁺ channels (Table 1) [29, 35]. Nevertheless, due to the involvement of HERG1 K⁺ channels in cardiac repolarization potential, there is evidence that the inhibition of these channels can slow down repolarization in the heart muscle and eventually lead to arrhythmia [10], which can account for the limited use of HERG1 K⁺ channel blockers and the meticulousness in choosing different forms of these inhibitors.

Potential effect of K+ channel expression and leukemia progression

The fact that K⁺ channels as key factors in controlling membrane potential play a central role in the growth of cells in both physiological and pathological conditions has been proved in numerous studies [35]. Voltage-dependent channels are divided into $K_V 1 - K_V 12$ types. Human voltage-gated potassium ion channel ether-à-go-go 1 (hEag1 or K_v10.1) is one of the most well-known voltage-dependent K⁺ channels whose expression has been shown to increase in many solid tumors and has been introduced as a poor prognostic factor in these malignancies due to its oncogenicity [36–39]. Furthermore, Agarwal et al. showed that hEag1 expression significantly increases in primary myeloid leukemia, CML, and myelodysplastic syndrome (MDS) [40]. Interestingly, the increase in hEag1 expression in these malignancies is directly related to the age of patients and is accompanied by adverse clinical outcomes such as increased relapse rate and shorter overall survival [40]. Although targeting hEag1 with different types of inhibitors is suggested as a therapeutic approach in cancers (Table 1) [35, 40], it seems that therapeutic intervention to control the molecular mechanisms involved in expression regulation of this gene is an alternative strategy to reduce the adverse effects of hEag1 expression. In this regard, Lin et al. showed that P53/Mir34/ E2f is a molecular pathway that regulates the expression of hEag1 in cells. Defective expression and function of the components of this molecular pathway, including P53 and Mir34, are associated with the overexpression of hEag1 as well as tumor cell proliferation (Fig. 1) [41]. Since methylation has been reported to be responsible for decreased P53 expression and increased LC proliferation [42], demethylation drugs may be useful as a therapeutic strategy to reduce the expression of hEag1 and its adverse effects in leukemia.



Fig. 1 Ion channels in the plasma membrane of LCs and their participation in leukemia progression. In BM, production of SDF-1 by MSCs can cause SDF-1/CXCR4 interaction as well as CXCR4/1integrin complex formation that lead to decreased intracellular K⁺ and LCs apoptosis. On the other hand, activation of CXCR4/1-integrin complex can be associated with resistance to chemotherapeutic agents and LC migration from BM toward PB via stimulating MAPK/PI3 K/Akt and Flt-1-dependent signaling pathways. Activation of RAS–RAF–MEK–ERK signaling pathway can promote K_V1.3 expression as well as apoptosis that lead to K⁺ excretion and induction of cellular contraction. Also, excretion of K⁺ via ClC-3 channels induces plasma membrane expression of AQPs and loss of LC H₂O and apoptosis. While accumulation of AQPs in intracellular space

 $K_v 1.3$ is another voltage-dependent member of K⁺ channel families that is expressed in the internal membrane of mitochondria with a crucial role in the formulation of apoptotic processes. It has been shown that the inhibition of $K_v 1.3$ by Bax and Bak can lead to hyperpolarization of the internal mitochondria membrane and the release of reactive oxygen species (ROS) causing death of cells [43]. In addition, the inhibition of mitochondrial $K_v 1.3$ channels in animal models and human cell lines by three distinct membrane-permeant inhibitors of $K_v 1.3$, namely Psora-4, PAP-1, and clofazimine, reveals the crucial function and

is associated with inhibition of apoptosis, localization of KCa3.1, TRPV5, and TRPV6 can be associated with accumulation of Ca²⁺ in intracellular environment that can promote LC proliferation. Also, proliferation of LCs can be mediated by activation of P53/Mir34/E2f signaling in nucleus that leads to overexpression of hEag1 channel. However, increased proliferation of LCs can inhibit it by P53 methylation. *BM* bone marrow, *MSCs* mesenchymal stem cells, *SDF-1* stromal cell-derived factor 1, *CXCR4* C–X–C chemokine receptor type 4, *HERG1* human ether-a-go-go 1, *hEag1* human voltage-gated potassium ion channel ether-à-go-go 1, *ClC-3* chloride channel 3, *TRPV* transient receptor potential vanilloid, *PB* peripheral blood, *AQP* aquaporin, *LCs* leukemic cells, *VGSCs* voltage-gated sodium channels

expression of these ion channels in cell death even in the absence of Bax and Bak [44]. Another mechanism regarding the association of K_v 1.3 channels with apoptotic process is the stimulation of the function of these channels in cellular membrane by Fas receptors. This mechanism results in K⁺ excretion of cells and induction of cellular contraction, which is one of the first cell responses to the onset of pro-apoptotic processes [45]. Meanwhile, other K⁺ channels such as K_v 3.4 can cause resistance of AML-LCs to radiotherapy and reduce the apoptosis of these cells [46]. Interestingly, the expression and activity of K_v 1.3 channels

Ion channels	Gene name	Cro.	Leukemia	Expression	Function	Inhibitor drug	References
K ⁺ channels							
k11.1 (hERG)	KCNH2	7q36.1	PAML, B-CLL, B-ALL, CML	UP	Inducing cell cycle and proliferation, migration, anti-apoptotic effect and drug resistance	Erythromycin, sertindole, WAY 123, dofetilide, astemizole, roscovitine, E4031	[10, 19, 40]
K10.1 hEag1	KCNH1	1q32.1	PAML, CML, MDS	UP	Inducing cell proliferation and adverse clinical outcomes such as increases relapse rate and shorter overall survival	Astemizole, imipramine, mAb56	[40, 41]
K _v 1.3	KCNA3	1p13.3	B-CLL, AML, ALL	UP	Initiation of apoptotic process	Correolide, Psora-4, clofazimine, benzamides, piperidines, tetraphenyl- porphyrins, dihydrophenanthridines, memantine	[44, 47–49, 86]
$K_v 3.4$	KCNC4	1p13.3	AML	UP	Reduced apoptosis and radioresistance	BDS-I	[46]
K _{2p} 18.1	KCNK18	10q25.3	T-ALL	UP	Inducing proliferation and migration of LCs	Bisindolylmaleimide, Ro-32-0432, and chelerythrine	[58, 87]
Ca2 ⁺ channels))	
kCa3.1	KCNN4	19q13.31	B lymphoma	UP	Associated with increased migration of LCs	TRAM-34	[51]
TRPV2	TRPV2		AML, CML, ALL, CLL	UP	Increased LC proliferation		[99]
TRPV5	TRPV5	7q34 7-24	K562 and Jurkat T cell lines	UP	Increased LC proliferation and resistance to anontosis	Fentamate, capsazepine	[60, 64, 65]
IKPV0	IKPV0	/q.54			aronhohm on		
P2X7	P2X7	12q24.31	AML, B-CLL, ALL, MDS, CMI	UP J	Leukemogenesis, proliferation, resistance to apoptosis	KN62, A-438079	[67, 88]
Na ⁺ channels							
Na1.3	SCN3A	2q24.3	Jurcat leukemic T cell line	UP	Motility and invasiveness of LCs	TTX, flecainide, mexiletine	[70, 74]
Na1.5	SCN5A	3p22.2					
Na1.6	SCN8A	12q13.13					
Na1.7	SCN9A	2q24.3					
Na1.9	SCN11A	3p22.2					
Cl ⁻ channels							
CLC3	CLCN3	4q33	CML	UP	Can be associated with apoptosis and proliferation of LCs	Tamoxifen, 5-nitro-2-(3-phenylpro- pylamino benzoic acid	[81, 85]
Cro chromoson	IID un-red	nletion hE					

 Table 1
 Altered ion channel expression and pharmacological treatment in leukemias

go 1; *KCNA* potassium voltage-gated channel subfamily A, *KCNC* potassium voltage-gated channel subfamily A, *KCNC* potassium voltage-gated channel subfamily K, *PAML* primary acute myeloid leukemia, *CML* chronic myeloid leukemia, *ALL* acute lymphoblastic leukemia, *CLL* chronic lymphoblastic leukemia, *MDS* myelodysplastic syndrome, *LCs* leukemic cells, *TTX* tetrodototic leukemia, *BDS-1* blood depressing substance-1, *KCNN4* potassium-calcium-activated channel subfamily N member 4, *TRPV* transient receptor potential vanilloid, *SCNA* sodium voltage-gated channel subfamily N member 4, *TRPV* transient receptor potential vanilloid, *SCNA* sodium voltage-gated channel channel subfamily N member 4, *TRPV* transient receptor potential vanilloid, *SCNA* sodium voltage-gated channel channel channel channel subfamily N member 4, *TRPV* transient receptor potential vanilloid, *SCNA* sodium voltage-gated channel ch nel alpha, CLC3 chloride channel 3, CLCN3 chloride voltage-gated channel 3 in malignant B-CLL cells are higher in membrane resting potential than in normal cells [47]. The induction of $K_{y}1.3$ expression by oncogenic B-RAF signaling, which is a major component of RAS-RAF-MEK-ERK signal transduction crucial for obtaining survival signals by B-cell receptor (BCR) in malignant B cells, is a molecular mechanism reported to increase the expression and activity of these ionic channels in B-CLL LCs [48]. It is inferred that the induction of expression and function of K_v1.3 channels in cytoplasmic membrane and its inhibition in mitochondrial membrane could change membrane potential and concentration of intracellular potassium to pave the way for the initiation of apoptosis processes. In addition to apoptosis, the expression of $K_y 1.3$ channels is known as a proliferation marker in CLL [49]. KCa3.1 channels or Ca²⁺-activated K⁺ is another potassium channel with low expression levels in normal B lymphocytes and resting CLL cells in contrast to its increased expression in activated CLL cells, suggesting the fact that these channels may be involved in the induction of proliferation in CLL cells [50]. The noteworthy point about K_v1.3 and KCa3.1 is their complexity in the regulation of potassium flow. For example, it has been shown that human Daudi cell line, a B-lymphoma cell line, expresses both K_v1.3 and KCa3.1 channels that interfere with their growth. While selective blocking of KCa3.1 inhibits cell cycle, simultaneous expression of both channels can induce cell cycle and proliferation in this cell line [51]. Therefore, the use of K⁺ channels as therapeutic targets for the induction of apoptosis and reduction of proliferation seems to require the use of compounds that can selectively target these channels in plasma or mitochondria membrane.

Two-pore domain K^+ (K_{2P}) channels are a family of potassium channels that play a role in maintaining the resting potential and depolarization of the membrane. Unlike K_v1.3 channels, the performance of these channels is independent of voltage and is only sensitive to changing physiological parameters such as pH, temperature, membrane stretch, and some intracellular signaling pathways, as well as in response to vital cell processes such as proliferation, differentiation, and apoptosis [52, 53]. It has been shown that four members of this family, including k_{2p}2.1 (TREK-1), K_{2P}3.1 (TASK-1), K_{2P}18.1 (TRESK), and K_{2P} 5.1 (TASK2), are increased in a variety of cancers such as leukemia and are considered as therapeutic targets in many cancers (Table 1) [54, 55]. It has been shown that the expression of these channels at the surface of T lymphocytes plays a significant role in maintaining their osmotic volume [56, 57]. Although little information is available on the expression of K_{2P} channels in humans LCs, it has recently been shown that increased expression of K_{2P}18.1 in plasma membranes of several T-lymphocytic cell lines (Jurkat, JCaM, H9) and T-ALL cells is associated with the proliferation of these cells. This is despite the fact that the expression rate of this potassium channel in resting T lymphocytes is negligible or is not generally

expressed, which may indicate their oncogenicity characteristic [58]. Interestingly, the dependence of TRESK channels on Ca²⁺ is different from that of KCa²⁺ channels. While KCa²⁺ channels are rapidly activated by increased concentration of Ca²⁺ and deactivated in its absence, TRESK channels maintain their activity in the absence of Ca²⁺ for a long time as a result of phosphorylation by compounds such as calcineurin, affecting the survival and function of the cells. Studies have also shown that TRESK can be involved in the expression of different genes and immune dysfunction in T-ALL LCs. This finding suggests that TRESK is a potential therapeutic target for immunomodulation in this malignancy [59].

Calcium activated ion channels in leukemia

Sustained Ca²⁺ flow is an essential mechanism for regulating vital processes, including cytokine production, proliferation, and differentiation in blood cells [60]. The passage of Ca²⁺ through the membrane of blood cells is mediated by two types of Ca²⁺ channels, including Ca²⁺ release-activated Ca^{2+} (CRAC) channels and transient receptor potential (TRP) family proteins [61]. Two members of TRP channels, namely TRP vanilloid type 5 (TRPV5) and TRPV6, are voltage-dependent Ca²⁺ channels that regulate cell proliferation. Also, it has been shown that increasing expression of TRPV5 and TRPV6 channels in K562 cell line results in the activation of Ca² ⁺/calmodulin-dependent kinase II (CaMKII) that can lead to increase of intracellular Ca²⁺ and thus induce proliferation, differentiation, and resistance to apoptosis in these cells [62-64]. Moreover, it has been shown that the expression of TRPV5 and TRPV6, as well as entry of Ca^{2+} , is directly related to the activity and proliferation of lymphocytes (Fig. 1). In fact, the direct relationship between the expression of TRPV5 and TRPV6 with leukemia Jurkat T-cell proliferation may indicate the pathological role of these ion channels in leukemias, especially lymphoid leukemias [65]. There is also widespread evidence indicating that TRPV2 expression is higher in myeloid and lymphoid LCs than in normal cells, which is associated with the growth of these malignant cells [66]. However, the presence of some polymorphisms in TRPM5 gene (another member of the TRP channels) in children with AML and ALL is linked with a reduction in disease progression [66]. These finding suggests that the genetic background of patients can have a significant relevance to ion channels' expression and function. Purinoreceptors (P2X) are a group of specific ATP-dependent Ca²⁺ channels in T lymphocytes with a significant role in mitosis and proliferation of these cells. Studies have shown that this group of Ca²⁺ channels has seven members, three of which (P2X1, P2X4, and P2X7) have been detected at the surface of hematopoietic progenitor cells and different hematopoietic cell lines [67]. Evidence suggests that there is a direct relationship between the expression of P2X7 in BM of AML and CLL patients with leukemogenesis in these malignancies [68]. Although there is a relatively high level of P2X7 expression in patients with ALL, there is no evidence of the origin of LCs (B-ALL or T-ALL cells) [68]. Nonetheless, the results of these studies have shown that P2X7 channels can trigger a network of signaling pathways with a key role in tendency of normal cells toward leukemic form via facilitating the entry of Ca²⁺ into cells. In fact, the expression of P2X7 channels is known as a poor prognostic factor of leukemogenesis characteristic that causes the progresses of leukemia [68].

Sodium channel expression in leukemia

Voltage-gated sodium channels (VGSCs) are membrane proteins that cause Na⁺ to enter the cells in the direction of concentration gradient. Research has shown that increasing expression of VGSCs in breast and prostate cancer cells is associated with an increase in their invasiveness, while inhibiting the expression and function of these ion channels with tetrodotoxin (TTX) in vitro is accompanied with a dramatic reduction in cancer cell metastasis and invasiveness [69, 70]. Studies on Jurkat T lymphocytes have shown that various types of VGSCs, including Na1.3, Na1.5, Na1.6, Na1.7, and Na1.9 channels, are expressed in this cell line and are related to Jurkat cell invasiveness; however, VGSC blockers are able to reduce the expression of these channels as well as Jurkat cell invasiveness by approximately 93% [70]. However, amilorides as blockers of another group of Na⁺ channels called non-voltage-gated sodium (NVGS) channels are not capable of inhibiting their expression and function in K562, U937, and AML-M5 cell lines [71, 72]. Since the expression of VGSCs and NVGS may be associated with invasive cellular behaviors such as secretion, adhesion, and motility in solid tumor cells [73], their expression in normal physiological conditions may be a mechanism for invasive T lymphocytes towards infectious tissues. In contrast, the expression of VGSCs and NVGS in T LCs is a factor in stimulating invasion into the lymph nodes and involvement of these organs. Despite extensive studies on inhibiting Na⁺ channels in solid tumors and the use of various inhibitors for inhibiting these channels (Table 1) [74-76], limited studies have been conducted on the use of such inhibitors in leukemia. Therefore, further studies are needed to confirm the use of Na⁺ blockers in patients with leukemia.

Effect of chloride channels in leukemia progression

Chloride (Cl⁻) channels are a group of membrane channels that play a role in a wide range of biological functions in cells, including regulating cell volume, stimulating the cells, and acidifying their environment [77]. Despite the fact that the precise performance of Cl⁻ channels in blood cells such as lymphocytes and neutrophils showing a high expression of these ion channels than other cells is not specified, investigations have shown that some of the volume-activated Cl⁻ channels are activated under isotonic conditions in ALL Molt4 cell line and play a critical role in maintaining the cellular volume via creating a stable Cl⁻ stream [78]. In addition, Jiang et al. indicated that the use of Cl⁻ channel blockers inhibits LC proliferation and cell cycle arrest in G0/ G1 phase, which may indicate the role of these ion channels in leukemia progression [79]. ClC-3 channel is a Cl⁻ channel that has recently been the focus of much attention in leukemia. According to Kasinathan et al.'s study, ClC-3 channels are accumulated in intracellular resting state, while the expression of these ion channels in plasma membrane increases with oxidation, which could indicate the role of CIC-3 channels in stimulating the oxidation of current anions over the membrane [80]. Additionally, under these conditions, cell membrane permeability to anions such as KCl is increased. Excretion of these anions and H₂O from cells could lead to cellular contractions as well as apoptosis [81]. Indeed, as demonstrated in solid tumors that ClC-3 channels play an important role in apoptosis and proliferation [82, 83], it is inferred that the expression of ClC-3 channel in cytoplasmic membrane of LCs may indicate their tendency to apoptosis while the cytoplasmic expression of this channel leads to LC proliferation. Another mechanism by which Cl⁻ channels are involved in apoptosis and proliferation of LCs is the regulation of volume-regulated chloride currents (VRCCs) by intracellular sigma receptors. Renaudo et al. showed that sigma receptors could modify the expression of VRCC and LC volume by altering the pattern of apoptosis and proliferation [81]. A noteworthy point about VRCCs is the control of their function by aquaporins (AQPs), which has been shown to contribute to the development of chemotherapy resistance in epithelial cancers [84]. Chae et al. showed that increasing the expression of AQP5 due to its effect on cell volume can stimulate proliferation, inhibit apoptosis, and generally improve the CML LCs [85]. Also, it has been shown that higher expression of AQP5 in patients with CML in the accelerated and blastic phases relative to patients in the chronic phase is directly related to imatinib mesylate resistance [85]. Although the exact mechanism of AQPs' function in regulating the expression of VRCCs is unknown, the increase in the expression of AQPs, especially AQP5, may play an essential role in reducing the absorption and sensitivity of LCs to chemotherapeutic agents by H₂O transport as well as disturbance of osmotic balance and intracellular ion concentration.

Discussion

In the past decades, studies on the role of ion channels in leukemias have focused on their level of expression, function, and introduction of these channels as diagnostic and prognostic markers. For example, the first studies on the role of K^+ channels in leukemia showed that channel K_y 11.1 is associated with aggressive AML phenotype and adverse clinical findings, including increasing relapses and shorter overall survival [6]. Subsequently, other studies have indicated that in addition to AML, the expression of other K⁺ channels such as hEag1 (K_v10.1) is strongly associated with increasing risk of recurrence and short-term survival in patients with CML and MDS [40]. K, 11.1 and K, 10.1 are K⁺ channels whose expression is associated with cell cycle progression and proliferation of LCs. Although ion channels are expressed in both normal and neoplastic cells, it seems that in the course of rapid transformations of neoplastic cells, the tendency to change the expression of ion channels increases, but they do not return to normal state due to rapid proliferation. For example, the stable expression of $K_v 1.3$ and $K_v 10.1$ on the surface of LCs has a significant impact on vital LC processes including proliferation, differentiation, migration, and apoptosis by regulating cell volume fluctuations and affecting intracellular signals [41, 50, 89]. These findings suggest that blocking of K⁺ channels can have a significant effect on controlling the progression of leukemias. Intracellular Ca²⁺ signals are an essential mechanism for the survival of normal and cancerous cells, and the extensive expression of Ca²⁺ channels has been shown to increase Ca²⁺ and cell hyperpolarization, which is possibly associated with movement and proliferation of LCs [65, 68]. In addition to direct contribution of ion channels to transportation of ions, K⁺, Na⁺, and Cl⁻ channels can indirectly affect the susceptibility of LCs to chemotherapeutic agents through affecting resting potential and ion gradient change. However, investigations have shown that various drugs such as E4031, a potential blocker for hERG1 channels, alone or in combination with chemotherapy drugs can inhibit the expression and function of these channels in different aspects of LCs physiology in vivo and in vitro to induce its anti-leukemic effects [10, 19, 22]. In fact, these findings suggest that ion channels can be considered as new candidates for overcoming drug resistance in leukemias.

Conclusion

Changing expression and function of ion channels in LCs can affect different aspects of pathophysiology of leukemias by influencing intracellular signaling pathways. Since ion channels are easily accessible membrane proteins, targeting them with effective drug compounds that can selectively articulate their expression and function is an effective way to control patients' clinical conditions and improve their response to treatment. However, we believe that the effectiveness of ion channels as therapeutic goals in leukemias requires extensive clinical studies to understand the exact mechanism of changes in their expression and function in LC survival and maintenance.

Acknowledgements We wish to thank all our colleagues in Allied Health Sciences School, Ahvaz Jundishapur University of Medical Sciences.

Author contributions SS conceived the manuscript and revised it. HR, AS, MMB, MG, and SS wrote the manuscript and prepared the table.

Compliance with ethical standards

Conflict of interest The authors declare no conflict of interest.

Research involving human participants and/or animals This article does not contain any studies with human participants or animals performed by any of the authors.

Informed consent For this type of study, informed consent is not required.

References

- Jean-Yves LG, Halima O-A, Olivier S, Pierre B, Ahmed A, Christophe V. Voltage-gated ion channels, new targets in anti-cancer research. Recent Pat Anticancer Drug Discov. 2007;2(3):189–202.
- 2. Arcangeli A, Becchetti A. Novel perspectives in cancer therapy: targeting ion channels. Drug Resist Updat. 2015;21:11–9.
- Arcangeli A, Crociani O, Lastraioli E, Masi A, Pillozzi S, Becchetti A. Targeting ion channels in cancer: a novel frontier in antineoplastic therapy. Curr Med Chem. 2009;16(1):66–93.
- Turner KL, Sontheimer H. Cl and K + channels and their role in primary brain tumour biology. Philos Trans R Soc Lond B Biol Sci. 2014;369(1638):20130095.
- Becchetti A. Ion channels and transporters in cancer. 1. Ion channels and cell proliferation in cancer. Am J Physiol Cell Physiol. 2011;301(2):C255–65.
- Pillozzi S, Brizzi MF, Bernabei PA, Bartolozzi B, Caporale R, Basile V, et al. VEGFR-1 (FLT-1), β1 integrin, and hERG K + channel for a macromolecular signaling complex in acute myeloid leukemia: role in cell migration and clinical outcome. Blood. 2007;110(4):1238–50.
- Tefferi A, Vardiman J. Classification and diagnosis of myeloproliferative neoplasms: the 2008 World Health Organization criteria and point-of-care diagnostic algorithms. Leukemia. 2008;22(1):14.
- Arcangeli A, Becchetti A. Complex functional interaction between integrin receptors and ion channels. Trends Cell Biol. 2006;16(12):631–9.
- 9. Kaczmarek LK. Non-conducting functions of voltage-gated ion channels. Nat Rev Neurosci. 2006;7(10):761.
- Arcangeli A, Pillozzi S, Becchetti A. Targeting ion channels in leukemias: a new challenge for treatment. Curr Med Chem. 2012;19(5):683–96.

- Becchetti A, Arcangeli A. Integrins and ion channels in cell migration: implications for neuronal development, wound healing and metastatic spread. Adv Exp Med Biol. 2010;674:107–23.
- Lang F, Föller M, Lang K, Lang P, Ritter M, Gulbins E, et al. Ion channels in cell proliferation and apoptotic cell death. J Membr Biol. 2005;205(3):147–57.
- 13. Prevarskaya N, Skryma R, Shuba Y. Ion channels and the hallmarks of cancer. Trends Mol Med. 2010;16(3):107–21.
- Cuddapah VA, Sontheimer H. Ion channels and transporters in cancer. 2. Ion channels and the control of cancer cell migration. Am J Physiol Cell Physiol. 2011;301(3):C541–9.
- D'Amico M, Gasparoli L, Arcangeli A. Potassium channels: novel emerging biomarkers and targets for therapy in cancer. Recent Pat Anticancer Drug Discov. 2013;8(1):53–65.
- Saki N, Abroun S, Hagh MF, Asgharei F. Neoplastic bone marrow niche: hematopoietic and mesenchymal stem cells. Cell J. 2011;13(3):131.
- Park KS, Pang B, Park SJ, Lee Y-G, Bae J-Y, Park S, et al. Identification and functional characterization of ion channels in CD34 + hematopoietic stem cells from human peripheral blood. Mol Cells. 2011;32(2):181–8.
- Zhang W, Hirschler-Laszkiewicz I, Tong Q, Conrad K, Sun S-C, Penn L, et al. TRPM2 is an ion channel that modulates hematopoietic cell death through activation of caspases and PARP cleavage. Am J Physiol Cell Physiol. 2006;290(4):C1146–59.
- Li H, Liu L, Guo L, Zhang J, Du W, Li X, et al. HERG K + channel expression in CD34 +/CD38 -/CD123 high cells and primary leukemia cells and analysis of its regulation in leukemia cells. Int J Lab Hematol. 2008;87(4):387.
- Li H, Liu L, Guo T, Zhang J, Li X, Du W, et al. Expression and functional role of HERG1, K + channels in leukemic cells and leukemic stem cells. J Huazhong Univ Sci Technol Med Sci. 2007;27(3):257–60.
- Urrego D, Tomczak AP, Zahed F, Stühmer W, Pardo LA. Potassium channels in cell cycle and cell proliferation. Philos Trans R Soc Lond B Biol Sci. 2014;369(1638):20130094.
- Glassmeier G, Hempel K, Wulfsen I, Bauer CK, Schumacher U, Schwarz JR. Inhibition of HERG1 K + channel protein expression decreases cell proliferation of human small cell lung cancer cells. Pflug Arch. 2012;463(2):365–76.
- Leanza L, O'Reilly P, Doyle A, Venturini E, Zoratti M, Szegezdi E, et al. Correlation between potassium channel expression and sensitivity to drug-induced cell death in tumor cell lines. Curr Pharm Des. 2014;20(2):189–200.
- Arcangeli A. Ion channels and transporters in cancer. 3. Ion channels in the tumor cell-microenvironment cross talk. Am J Physiol Cell Physiol. 2011;301(4):C762–71.
- Rafieemehr H, Kheirandish M, Soleimani M. Improving the neuronal differentiation efficiency of umbilical cord blood-derived mesenchymal stem cells cultivated under appropriate conditions. Iran J basic Med Sci. 2015;18(11):1100.
- Rafieemehr H, Kheirandish M, Soleimani M. Neural differentiation of human umbilical cord blood derived mesenchymal stem cells. Avicenna J Med Biochem. 2016;4(1):e29066.
- Ravan AP, Goudarzi F, Rafieemehr H, Bahmani M, Rad F, Jafari M, et al. Human umbilical cord-mesenchymal stem cells conditioned medium attenuates CCl4 induced chronic liver fibrosis. Toxin Rev. 2019;38(3):1–12.
- Zeng Z, Samudio IJ, Munsell M, An J, Huang Z, Estey E, et al. Inhibition of CXCR29 with the novel RCP168 peptide overcomes stroma-mediated chemoresistance in chronic and acute leukemias. Mol Cancer Ther. 2006;5(12):3113–21.
- Pillozzi S, Masselli M, De Lorenzo E, Accordi B, Cilia E, Crociani O, et al. Chemotherapy resistance in acute lymphoblastic leukemia requires hERG1 channels and is overcome by hERG1 blockers. Blood. 2011;117(3):902–14.

- Liu P, Ma D, Yu Z, Zhe N, Ren M, Wang P, et al. Overexpression of heme oxygenase-1 in bone marrow stromal cells promotes microenvironment-mediated imatinib resistance in chronic myeloid leukemia. Biomed Pharmacother. 2017;91:21–30.
- Burger JA, Gribben JG. The microenvironment in chronic lymphocytic leukemia (CLL) and other B cell malignancies: insight into disease biology and new targeted therapies. Semin Cancer Biol. 2014;24:71–81.
- 32. Li H, Du YM, Guo L, Jie S, Zhang S, Du W, et al. The role of hERG1 K + channels and a functional link between hERG1 K + channels and SDF-1 in acute leukemic cell migration. Exper Cell Res. 2009;315(13):2256–64.
- Iwamoto S, Mihara K, Downing JR, Pui CH, Campana D. Mesenchymal cells regulate the response of acute lymphoblastic leukemia cells to asparaginase. J Clin Invest. 2007;117(4):1049–57.
- Zeng Z, Shi YX, Samudio IJ, Wang RY, Ling X, Frolova O, et al. Targeting the leukemia microenvironment by CXCR35 inhibition overcomes resistance to kinase inhibitors and chemotherapy in AML. Blood. 2009;113(24):6215–24.
- 35. Wulff H, Castle NA, Pardo LA. Voltage-gated potassium channels as therapeutic targets. Nat Rev Drug Discov. 2009;8(12):982–1001.
- Hemmerlein B, Weseloh RM, de Queiroz MF, Knotgen H, Sanchez A, Rubio ME, et al. Overexpression of Eag1 potassium channels in clinical tumours. Mol Cancer. 2006;5:41.
- Gomez-Varela D, Zwick-Wallasch E, Knotgen H, Sanchez A, Hettmann T, Ossipov D, et al. Monoclonal antibody blockade of the human Eag1 potassium channel function exerts antitumor activity. Cancer Res. 2007;67(15):7343–9.
- Rodriguez-Rasgado JA, Acuna-Macias I, Camacho J. Eagl channels as potential cancer biomarkers. Sensors (Basel). 2012;12(5):5986–95.
- Downie BR, Sanchez A, Knotgen H, Contreras-Jurado C, Gymnopoulos M, Weber C, et al. Eag1 expression interferes with hypoxia homeostasis and induces angiogenesis in tumors. J Biol Chem. 2008;283(52):36234–40.
- 40. Agarwal JR, Griesinger F, Stuhmer W, Pardo LA. The potassium channel Ether a go-go is a novel prognostic factor with functional relevance in acute myeloid leukemia. Mol Cancer. 2010;9:18.
- Lin H, Li Z, Chen C, Luo X, Xiao J, Dong D, et al. Transcriptional and post-transcriptional mechanisms for oncogenic overexpression of ether a go-go K+channel. PLoS One. 2011;6(5):e20362.
- Behzad MM, Shahrabi S, Jaseb K, Bertacchini J, Ketabchi N, Saki N. Aberrant DNA methylation in chronic myeloid leukemia: cell fate control, prognosis, and therapeutic response. Biochem Gene. 2018;56(3):149–75.
- Leanza L, Venturini E, Kadow S, Carpinteiro A, Gulbins E, Becker KA. Targeting a mitochondrial potassium channel to fight cancer. Cell Calcium. 2015;58(1):131–8.
- Leanza L, Henry B, Sassi N, Zoratti M, Chandy KG, Gulbins E, et al. Inhibitors of mitochondrial Kv1.3 channels induce Bax/Bak-independent death of cancer cells. EMBO Mol Med. 2012;4(7):577–93.
- Szabò I, Zoratti M, Gulbins E. Contribution of voltage-gated potassium channels to the regulation of apoptosis. FEBS Lett. 2010;584(10):2049–56.
- Palme D, Misovic M, Schmid E, Klumpp D, Salih HR, Rudner J, et al. Kv.34 potassium channel-mediated electrosignaling controls cell cycle and survival of irradiated leukemia cells. Pflug Arch. 2013;465(8):1209–21.
- Szabo I, Trentin L, Trimarco V, Semenzato G, Leanza L. Biophysical characterization and expression analysis of Kv1.3 potassium channel in primary human leukemic B cells. Cell Physiol Biochem. 2015;37(3):965–78.
- Roskoski R Jr. ERK1/2 MAP kinases: structure, function, and regulation. Pharmacol Res. 2012;66(2):105–43.

- Nguyen W, Howard BL, Neale DS, Thompson PE, White PJ, Wulff H, et al. Use of Kv1.3 blockers for inflammatory skin conditions. Curr Med Chem. 2010;17(26):2882–96.
- Grossinger EM, Weiss L, Zierler S, Rebhandl S, Krenn PW, Hinterseer E, et al. Targeting proliferation of chronic lymphocytic leukemia (CLL) cells through KCa3.1 blockade. Leukemia. 2014;28(4):954–8.
- Wang J, Xu YQ, Liang YY, Gongora R, Warnock DG, Ma HP. An intermediate-conductance Ca(2+)-activated K (+) channel mediates B lymphoma cell cycle progression induced by serum. Pflug Arch. 2007;454(6):945–56.
- 52. Cid LP, Roa-Rojas HA, Niemeyer MI, Gonzalez W, Araki M, Araki K, et al. TASK-2: a K2P K(+) channel with complex regulation and diverse physiological functions. Front Physiol. 2013;4:198.
- Enyedi P, Czirjak G. Molecular background of leak K + currents: two-pore domain potassium channels. Physiol Rev. 2010;90(2):559–605.
- 54. Liu H, Enyeart JA, Enyeart JJ. Potent inhibition of native TREK-1 K + channels by selected dihydropyridine Ca2 + channel antagonists. J Pharmacol Exp Ther. 2007;323(1):39–48.
- 55. Borsotto M, Veyssiere J, Maati MOH, Devader C, Mazella J, Heurteaux C. Targeting two-pore domain K(+) channels TREK-1 and TASK-3 for the treatment of depression: a new therapeutic concept. Br J Pharmacol. 2015;172(3):771–84.
- Meuth SG, Bittner S, Meuth P, Simon OJ, Budde T, Wiendl H. TWIK-related acid-sensitive K + channel 1 (TASK1) and TASK3 critically influence T lymphocyte effector functions. J Biol Chem. 2008;283(21):14559–70.
- Pottosin II, Bonales-Alatorre E, Valencia-Cruz G, Mendoza-Magaña ML, Dobrovinskaya OR. TRESK-like potassium channels in leukemic T cells. Pflug Arch. 2008;456(6):1037–48.
- Sánchez-Miguel DS, García-Dolores F, Flores-Márquez MR, Delgado-Enciso I, Pottosin I, Dobrovinskaya O. TRESK potassium channel in human T lymphoblasts. Biochem Biophys Res Commun. 2013;434(2):273–9.
- Es-Salah-Lamoureux Z, Steele DF, Fedida D. Research into the therapeutic roles of two-pore-domain potassium channels. Trends Pharmacol Sci. 2010;31(12):587–95.
- Semenova SB, Vassilieva IO, Fomina AF, Runov AL, Negulyaev YA. Endogenous expression of TRPV5 and TRPV6 calcium channels in human leukemia K562 cells. Am J Physiol Cell Physiol. 2009;296(5):C1098–104.
- Feske S, Skolnik EY, Prakriya M. Ion channels and transporters in lymphocyte function and immunity. Nat Rev Immunol. 2012;12(7):532.
- 62. Monteith GR, Davis FM, Roberts-Thomson SJ. Calcium channels and pumps in cancer: changes and consequences. J Biol Chem. 2012;287(38):31666–73.
- Lehen'Kyi V, Flourakis M, Skryma R, Prevarskaya N. TRPV6 channel controls prostate cancer cell proliferation via Ca 2+/ NFAT-dependent pathways. Oncogene. 2007;26(52):7380.
- 64. Heise N, Palme D, Misovic M, Koka S, Rudner J, Lang F, et al. Non-selective cation channel-mediated Ca2 + -entry and activation of Ca2 +/calmodulin-dependent kinase II contribute to G2/M cell cycle arrest and survival of irradiated leukemia cells. Cell Physiol Biochem. 2010;26(4–5):597–608.
- 65. Vassilieva IO, Tomilin VN, Marakhova II, Shatrova AN, Negulyaev YA, Semenova SB. Expression of transient receptor potential vanilloid channels TRPV5 and TRPV6 in human blood lymphocytes and Jurkat leukemia T cells. J Membr Biol. 2013;246(2):131–40.
- 66. Morelli BM, Liberati S, Amantini C, Nabiss M, Santoni M, Farfariello V, et al. Expression and function of the transient receptor potential ion channel family in the hematologic malignancies. Curr Mol Pharmacol. 2013;6(3):137–48.

- Feng W, Wang L, Zheng G. Expression and function of P2 receptors in hematopoietic stem and progenitor cells. Stem Cell Investig. 2015;30(2):14.
- Guven Maiorov E, Keskin O, Hatirnaz Ng O, Ozbek U, Gursoy A. Identification of interconnected markers for T-cell acute lymphoblastic leukemia. Biomed Res Int. 2013;2013:210253.
- 69. Roger S, Potier M, Vandier C, Besson P, Le Guennec J-Y. Voltagegated sodium channels: new targets in cancer therapy? Curr Pharm Des. 2006;12(28):3681–95.
- Fraser SP, Diss JK, Lloyd LJ, Pani F, Chioni A-M, George AJ, et al. T-lymphocyte invasiveness: control by voltage-gated Na+channel activity. FEBS Lett. 2004;569(1–3):191–4.
- Sudarikova A, Vassilieva I, Morachevskaya E, Negulyaev YA. Molecular and functional identification of sodium channels in K562 cells. Cell Tissue Biol. 2012;6(5–6):435–41.
- Sudarikova AV, Tsaplina OA, Chubinskiy-Nadezhdin VI, Morachevskaya EA, Negulyaev YA. Amiloride-insensitive sodium channels are directly regulated by actin cytoskeleton dynamics in human lymphoma cells. Biochem Biophys Res Commun. 2015;461(1):54–8.
- Fraser SP, Ozerlat-Gunduz I, Brackenbury WJ, Fitzgerald EM, Campbell TM, Coombes RC, et al. Regulation of voltage-gated sodium channel expression in cancer: hormones, growth factors and auto-regulation. Philos Trans R Soc Lond B Biol Sci. 2014;369(1638):20130105.
- 74. Hesselink JMK. Moving targets in sodium channel blocker development: the case of raxatrigine: from a central NaV1. 3 blocker via a peripheral NaV1. 7 blocker to a less selective sodium channel blocker. J Med Ther. 2017;1(1):1–3.
- Luiz AP, Wood JN. Sodium channels in pain and cancer: new therapeutic opportunities. Adv Pharmacol. 2016;75:153–78.
- Jensen MK, Sakakura T, Abe Y, Takamori H, Takasuna K, Tsurubuchi Y, et al. Use and state dependent Nav1. 5 blockers on QPatch X and in vivo assays. J Pharmacol Toxicol Methods. 2011;1(64):e8.
- 77. Verkman AS, Galietta LJ. Chloride channels as drug targets. Nat Rev Drug Discov. 2009;8(2):153.
- Cao G, Zuo W, Fan A, Zhang H, Yang L, Zhu L, et al. Volumesensitive chloride channels are involved in maintenance of basal cell volume in human acute lymphoblastic leukemia cells. J Membr Biol. 2011;240(2):111–9.
- Jiang B, Hattori N, Liu B, Nakayama Y, Kitagawa K, Inagaki C. Suppression of cell proliferation with induction of p21 by Cl⁻ channel blockers in human leukemic cells. Eur J Pharmacol. 2004;488(1–3):27–34.
- Kasinathan RS, Föller M, Lang C, Koka S, Lang F, Huber SM. Oxidation induces CIC-3-dependent anion channels in human leukaemia cells. FEBS Lett. 2007;581(28):5407–12.
- Renaudo A, L'Hoste S, Guizouarn H, Borgèse F, Soriani O. Cancer cell cycle modulated by a functional coupling between sigma-1 receptors and Cl-channels. J Biol Chem. 2007;282(4):2259–67.
- Zuo W, Zhu L, Bai Z, Zhang H, Mao J, Chen L, et al. Chloride channels involve in hydrogen peroxide-induced apoptosis of PC12 cells. Biochem Biophys Res Commun. 2009;387(4):666–70.
- de Tassigny ADA, Berdeaux A, Souktani R, Henry P, Ghaleh B. The volume-sensitive chloride channel inhibitors prevent both contractile dysfunction and apoptosis induced by doxorubicin through PI3kinase, Akt and Erk 1/2. Eur J Heart Fail. 2008;10(1):39–46.
- 84. Pedersen SF, Hoffmann EK, Novak I. Cell volume regulation in epithelial physiology and cancer. Front Physiol. 2013;4:233.
- Chae YK, Kang SK, Kim MS, Woo J, Lee J, Chang S, et al. Human AQP5 plays a role in the progression of chronic myelogenous leukemia (CML). PLoS One. 2008;3(7):e2594.
- Lowinus T, Heidel FH, Bose T, Nimmagadda SC, Schnöder T, Cammann C, et al. Memantine potentiates cytarabine-induced cell

death of acute leukemia correlating with inhibition of K v 1.3 potassium channels, AKT and ERK1/2 signaling. Cell Commun Signal. 2019;17(1):5.

- Rahm AK, Gierten J, Kisselbach J, Staudacher I, Staudacher K, Schweizer PA, et al. PKC-dependent activation of human K2P18.
 K + channels. Br J Pharmacol. 2012;166(2):764–73.
- Chong J-H, Zheng G-G, Ma Y-Y, Zhang H-Y, Nie K, Lin Y-M, et al. The hyposensitive N187D P2X7 mutant promotes malignant progression in nude mice. J Biol Chem. 2010;285(46):36179–87.

 Gulbins E, Sassi N, Grassme H, Zoratti M, Szabo I. Role of Kv.13 mitochondrial potassium channel in apoptotic signalling in lymphocytes. Biochim Biophys Acta. 2010;1797(6–7):1251–9.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.