

## MYC oncogene in myeloid neoplasias

M. Dolores Delgado · Marta Albajar ·  
M. Teresa Gomez-Casares · Ana Batlle ·  
Javier León

Received: 6 July 2012 / Accepted: 24 July 2012 / Published online: 22 August 2012  
© Federación de Sociedades Españolas de Oncología (FESEO) 2012

**Abstract** MYC is a transcription factor that regulates many critical genes for cell proliferation, differentiation, and biomass accumulation. MYC is one of the most prevalent oncogenes found to be altered in human cancer, being deregulated in about 50 % of tumors. Although MYC deregulation has been more frequently associated to lymphoma and lymphoblastic leukemia than to myeloid malignancies, a body of evidence has been gathered showing that MYC plays a relevant role in malignancies derived from the myeloid compartment. The myeloid leukemogenic activity of MYC has been demonstrated in different murine models. Not surprisingly, MYC has been found to be amplified or/and deregulated in the three major types of myeloid neoplasms: acute myeloid leukemia, myelodysplastic syndromes, and myeloproliferative neoplasms, including chronic myeloid leukemia. Here, we review the recent literature describing the involvement of MYC in myeloid tumors.

**Keywords** MYC · Myeloid neoplasia · Acute myeloid leukemia · Chronic myeloid leukemia · Myelodysplastic syndromes

### Myelopoiesis and MYC impact on myeloid cell proliferation and differentiation

Hematopoietic stem cells differentiate along two major lineages. The lymphoid lineages give rise to B- and T-lymphocytes, as well as natural killer cells. The myeloid lineage is much more complex, with at least six major differentiated circulating cells (erythrocytes, monocyte/macrophages, eosinophils, neutrophils, basophils, and megakaryocyte/platelets) each of them very different from the other cell types in its function in the organism, its number in peripheral blood, and its morphology and gene expression profile [1–3]. Myeloid lineage commitment relies on timely activation of appropriate transcription factors and silencing of others, which is the result from a network of extracellular signals [4, 5].

MYC is a transcription factor of the helix-loop-helix-leucine zipper family. MYC forms dimers with MAX protein and binds to E-box sequences in the regulatory regions of around one thousand target genes (Fig. 1a) [6]. MYC is involved in a number of fundamental functions for the cell biology, such as control of proliferation and differentiation, energy production, cell size, and many others (Fig. 1b) [7–9]. Not surprisingly, deregulation of MYC activities contributes to the tumor phenotype. MYC is one of the most prevalent oncogenes in human cancer, and particularly in hematopoietic neoplasias [10, 11]. In fact, in human cancer, MYC deregulation was first observed in Burkitt lymphoma, as well as in chemically induced murine plasmacytomas [12]. Virtually all cases of these

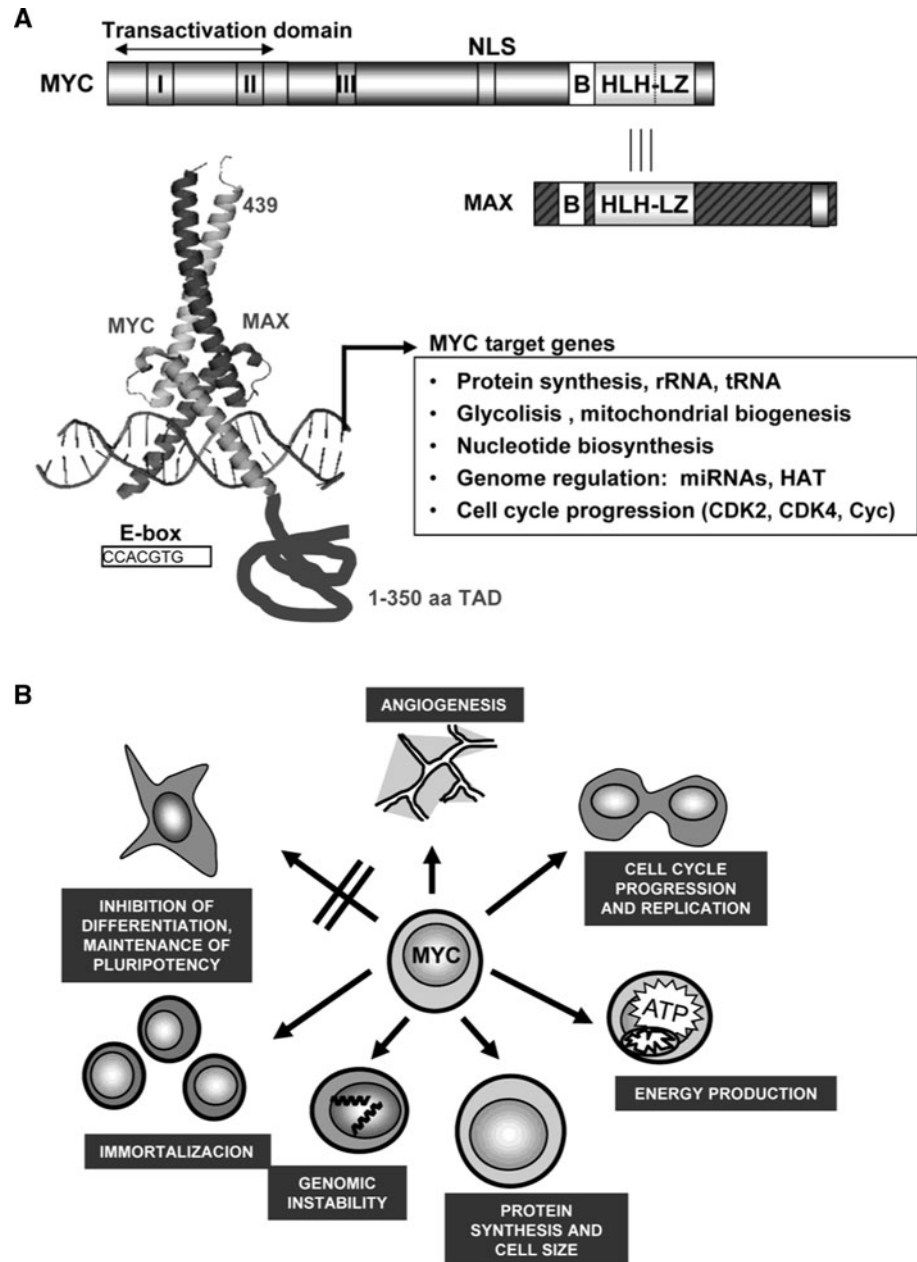
---

M. D. Delgado · M. Albajar · A. Batlle · J. León (✉)  
Group of Transcriptional Control and Cancer, Departamento de Biología Molecular, Facultad de Medicina, Instituto de Biomedicina y Biotecnología de Cantabria (IBBTEC), Universidad de Cantabria, CSIC, SODERCAN, Avda Cardenal Herrera Oria s/n, 39011 Santander, Spain  
e-mail: leonj@unican.es

M. Albajar · A. Batlle  
Servicio de Hematología, Hospital Universitario Marqués de Valdecilla, and IFIMAV, Santander, Spain

M. T. Gomez-Casares  
Servicio de Hematología and Unidad de Investigación, Hospital Dr. Negrín, Las Palmas, Spain

**Fig. 1** MYC structure and functions. **a** Scheme showing the main features of MYC transcription factor: the N-terminal transactivation domain (TAD), conserved MYC box-I, II, and III, nuclear localization signal (NLS), and the basic-helix-loop-helix-leucine zipper (B-HLH-LZ) DNA binding and dimerization domain. Heterodimers MYC-MAX bind E-box sequences at the regulatory regions of its target genes (the most representative indicated in the *square*). **b** MYC is involved in many cellular functions through the regulation (activation or repression) of its target genes. MYC favors the cell cycle progression by activating cyclin-dependent kinases (CDKs) and repressing CDK inhibitors; MYC regulates metabolic enzymes involved in the ATP production; MYC has a role in protein synthesis and cell size through the control of ribosomal genes among others; MYC induces genomic instability and cell immortalization; the inhibition of cell differentiation, maintenance of pluripotency, and induction of angiogenesis are important MYC functions. Elevated MYC levels (by gene amplification, chromosomal translocation, overexpression, and protein stabilization) deregulate such functions, contributing to tumorigenesis



lymphoma show a translocation affecting the MYC locus [13]. From then on, MYC translocations have been found in a significant fraction of high-grade lymphomas and multiple myeloma, and MYC overexpression has also been reported in numerous types of lymphoma and lymphoblastic leukemia [14].

In contrast to lymphoid tumors, MYC involvement in myeloid cancer has received much less attention. Nonetheless, MYC oncogene was first discovered as the oncogene carried by retrovirus that induced a myeloid neoplasm in chicken, i.e., myelocytomatosis, and MYC was named after this tumor [15]. Moreover, the inhibition of myeloid cell differentiation was one of the first biological effects

described for MYC (reviewed in Ref. [9]). In 1986, three reports showed that MYC blocked the chemically induced erythroid differentiation of Friend murine erythroleukemia (MEL) cells [16–18]. We also showed that MYC inhibited the erythroid differentiation of human myeloid cells such as K562 derived from chronic myeloid leukemia (CML) [19], and found that MYC blocks erythroid differentiation in genetically-defined models where erythroid differentiation is induced by p27<sup>KIP1</sup> [20]. In this model, MYC impairs the up-regulation of many erythroid-specific genes, as well as that of transcription factors that determine erythroid lineage differentiation (including GATA1 and NFE2) but, strikingly, it does not reverse the proliferation

arrest and the repression of CDK activity mediated by p27<sup>KIP1</sup>. In a complementary approach, using the leukemia K562 and U937 cells, it has been shown that MYC is not down-regulated when cells are growth-arrested but not differentiated [21, 22]. Enforced MYC expression also blocks the monocytic differentiation of the AML-derived cell line U937 [23]. The fact that MYC can block differentiation of myeloid leukemia cells models in culture is consistent with its involvement in myeloid leukemia. However, MYC activity is not universally linked to differentiation inhibition of myeloid cells, as MYC enhances the retinoic acid-induced differentiation of a promyelocytic leukemia cell line (NB4) [24].

### MYC roles in the determination of myeloid lineages in vivo

The results described above are consistent with the in vivo observed data. The effects of MYC deletion in the myeloid compartment have been recently reported in the MYC conditional knock-out mice. *Myc*<sup>-/-</sup> mice show significant thrombocytosis, severe anemia, and grossly decreased neutrophil/monocyte numbers [25]. Thus MYC induces opposite effects in the differentiation of megakaryocytic versus monocyte and erythroid lineages. Moreover, these effects of MYC deletion in vivo on hematopoietic lineages correlates well with MYC expression in mouse hematopoietic cells: cells expressing higher levels of MYC such as granulocyte/monocyte precursors, common lymphocyte progenitors, and erythrocytic blasts are significantly reduced while cells expressing lower MYC levels (hematopoietic stem cells (HSC), megakaryocyte/erythroid precursors, and megakaryocytes) are less affected or are increased in number [25].

Megakaryocytes from *Myc*<sup>-/-</sup> mice are significantly smaller in size and are lower in ploidy than those of control mice; as a result, fewer platelets are produced by each megakaryocyte. However, due to the increase in megakaryocytic number, a significant increase in platelet number was observed in *Myc*<sup>-/-</sup> mice. The involvement of MYC in megakaryocytic differentiation is confirmed in transgenic mice with MYC overexpression in the megakaryocytic lineage, where a *Myc* transgene is under the control of the platelet factor-4 promoter. These mice show an increase in low-ploidy megakaryocytes due to enhanced proliferation and survival, along with the blocking of differentiation [26].

In conclusion, MYC plays a pivotal role in regulating the normal hematopoiesis, and thus, it is not surprising that MYC is frequently deregulated in human leukemia and lymphoma. Such deregulation would destroy this balance and transform hematopoietic cells by stimulating proliferation and blocking terminal differentiation.

### In vivo models for MYC-induced myeloid leukemia: engraftment of retrovirally mediated expression of MYC in hematopoietic precursors

Several reports illustrate the leukemogenic effects of MYC in hematopoietic cells after the enforced expression in murine hematopoietic precursors via retroviral infection, although the induced neoplasia varies depending on the virus and the cells type targeted by the virus. Unfractionated bone marrow cells retrovirally transduced with *Myc*, using different vectors and experimental settings, resulted in development of acute myeloid leukemia (AML) [27–29]. In one study, the MYC-transduced fetal liver cells transduced with another MYC vector induced a long-latency lymphoma [30].

The infection of bone marrow progenitors in a p53<sup>-/-</sup> background also resulted in lymphoma [31]. Interestingly, upon culturing in vitro, cells derived from these lymphomas underwent myeloid differentiation, but then switched from myeloid to lymphoid lineage and induced B cell lymphomas when returning to in vivo conditions [32]. In another study, mice bone marrow cells transduced with another member of the MYC family, MYCN (also called N-Myc), developed monoclonal and transplantable AML [33]. In a parallel report, it was shown that when the bone marrow from lethally irradiated cells was repopulated with bone marrow cells expressing *Myc*, the mice developed an AML-like disease [29]. In this model, the co-expression of several antiapoptotic genes of the BCL2 family accelerated leukemogenesis but did not change the myeloid phenotype of the leukemia [29]. In conclusion, these experimental models indicate that high MYC levels switch normal hematopoiesis into myeloid leukemia. The analysis of the AML induced in some of these models showed high expression of the antiapoptotic genes of the BCL2 family including the MCL1 gene. Moreover, MCL1 haploinsufficiency abrogated AML development in this model [28]. The results suggest that abrogation of apoptosis in MYC-targeted cells is probably necessary for the development of an aggressive AML. This has been recently confirmed in another report, in which retroviral transduction of MYC in purified (Lin<sup>-</sup>) murine HSC results in the development of both myeloid and T-lymphoid tumors within 2 months after transplantation. Interestingly, co-expression of MYC with BCLXL or BCL2 resulted in almost immediate development of AML-like disease, but not lymphoma (at the expense of other hematopoietic lineages) [34].

### In vivo models for MYC-induced myeloid leukemia: transgenic mice developing myeloid neoplasia

Although most of the available literature on murine models of MYC-induced tumors is focused on MYC-induced

lymphoma, there are also transgenic models that demonstrate the carcinogenic potential of MYC in the myeloid compartment. Mice carrying the human MYC proto-oncogene under the control of the murine GATA-1 promoter (an erythroid-specific gene) developed an early onset erythroleukemia [35]. Several transgenic mice lines have been generated carrying the MYC gene under the control of the Vav promoter (which is active in all hematopoietic cells lineages and hematopoietic precursors) showing different MYC expression levels [36]. Interestingly, the tumor lineage varied with the different MYC expression levels achieved in the transgenic mice. For instance, aggressive T-cell lymphomas were the predominant tumor arising in transgenic mice lines showing the highest MYC expression [36]. In contrast, most tumors in the mouse lines expressing lower MYC levels were late-onset monocytic tumors [37]. It is noteworthy that just a two-fold decrease in MYC levels switched the phenotype from T-cell tumors to monocytic tumors. Thus, relatively low MYC levels can transform monocyte-macrophages but are insufficient to transform T-lymphocytes [37]. These data and other gathered in the literature strongly suggest that relatively small increases in MYC are sufficient to promote leukemia.

### MYC in human myeloproliferative neoplasms

Consistent with the data gathered in animal models (see above), MYC deregulation has been found in human myeloid neoplasias. Three major groups of clonal myeloid diseases have been described: acute myeloid leukemia, myelodysplastic syndromes, and myeloproliferative neoplasms [38]. Myeloproliferative neoplasms (MPN) are characterized by an enhanced proliferation of one or more of myeloid, erythroid, or megakaryocytic lineages. Chronic myeloid leukemia (CML) affects annually 1–2 cases per 100,000 individuals. CML is a proliferative clonal disorder of hematopoietic stem cells that results primarily in the expansion of mature myeloid cells that retains a capacity for differentiation. Untreated CML progress from the initial stage, termed chronic phase, to a blastic crisis phase which is similar to acute leukemia. Other frequent MPNs are essential thrombocythemia (ET), polycythemia vera (PV), and primary myelofibrosis (PMF).

The Philadelphia chromosome and the resulting fusion Bcr-Abl oncogene is the universal molecular marker of CML and has a central role in the disease etiology [39]. Bcr-Abl up-regulates MYC expression [40, 41] and MYC cooperates with Bcr-Abl in transformation [42–44]. Moreover, Bcr-Abl provokes the phosphorylation of MYC in Ser-62. This phosphorylation renders a more stable MYC protein. A proteomic analysis revealed that MYC-Ser62 was largely dephosphorylated after dasatinib

treatment [45]. This result suggests that these kinase inhibitors would reduce the levels of MYC protein in the leukemic cell.

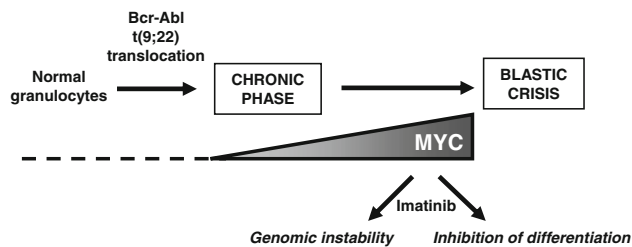
Studies performed with a small number of cases, showed that MYC mRNA levels are elevated in CML-blastic crisis [46, 47] and in chronic phase CML versus healthy bone marrow samples [48, 49]. We have recently observed the up-regulation of MYC expression during CML progression [50]. Moreover, high levels of MYC along the disease correlate with poorer response to imatinib, and higher MYC mRNA levels at diagnosis tend to correlate with a worse response to imatinib [50]. Essentially the same results have been recently reported in a smaller cohort of CML patients, showing that MYC protein is elevated at diagnosis in patients destined to progress to blastic crisis [51]. It is also of note that trisomy 8 and gain at 8q24 (where MYC maps) are among the most frequent cytogenetic alterations in CML [52, 53], although their correlation with MYC expression is yet unknown.

It has been recently shown that at the time of diagnosis of CML, patients who will later progress to blast crisis have significantly higher levels of CIP2A protein (cancerous inhibitor of PP2A) [51]. CIP2A inhibits PP2A-mediated dephosphorylation of MYC at Ser-62, as pSer62-MYC is stabilized against degradation [54]. Although this should be confirmed in bigger cohorts of patients, these results indicate that CML progression is not only a consequence of elevated mRNA levels but also of MYC protein stabilization [51].

Misregulation of the activity of a specific group of ATP-binding cassette transporters (ABC) is responsible for reducing the intracellular concentration of drugs in leukemic cells. CD34<sup>+</sup> hematopoietic cell precursors of CML patients overexpress ABC transporters, and this overexpression is, at least in part, due to MYC, as ABC is a direct target of MYC [55]. This MYC activity could also contribute to CML transformation.

Why MYC should be deregulated during the CML progression? CML progression to blastic crisis is associated to cell survival, genomic instability, and differentiation arrest [39, 56]. We have shown that enforced MYC expression in CML-derived cells as K562, results in aberrant DNA synthesis under imatinib stress [50] and blocks imatinib-mediated differentiation [57], suggesting that MYC may contribute to CML transformation acting at those two levels (Fig. 2).

MYC mRNA is overexpressed in bone marrow cells from ET [58]. On the contrary, MYC overexpression is not found in the PV [58]. This is striking as more than 95 % of PV patients carry an activating mutation in the tyrosine kinase JAK2, and JAK2 has been reported to induce MYC expression in cell lines [40, 59]. Clearly, an analysis of STAT activation and the transcription factors phosphorylated by JAKs, is lacking in PV, making it difficult to obtain the interpretation of these results.



**Fig. 2** MYC is overexpressed in chronic myeloid leukemia at diagnosis, and in blastic crisis. In CML-derived cells (K562 cell line) treated with imatinib, MYC induces genetic instability [50] and block of erythroid differentiation [57]

### MYC in human myelodysplastic syndrome (MDS) and acute myeloid leukemia (AML)

Acute myeloid leukemia (AML) is characterized by the proliferation and accumulation of immature hematopoietic cells in the bone marrow and blood. AML is a heterogeneous group of neoplasms affecting the myeloid lineage. Altogether, their incidence worldwide is about 2–3 cases in 100,000 individuals per year, and is reported to be the highest in Australia, Western Europe, and United States. The former FAB (French-American-British) classification system of AML (M0 to M7 subtypes, attending to the differentiation type and stage) has been superseded by the World Health Organization (WHO) classification, that identifies 15 diseases characterized by clinical presentation and recurrent chromosomal aberrations [38]. Understandably, most molecular studies to date are carried out with a relatively small number of cases of each myeloid disease or do not make distinctions between the different entities. Therefore, the information on *MYC* expression in myeloid leukemia actually refers to a range of related diseases, and therefore its involvement in a particular myeloid neoplasm has not been properly addressed.

*MYC* amplification and overexpression have been found in AML [62] (Table 1). The overexpression of *MYC* mRNA in bone marrow and peripheral blood in sporadic AML cases patients (as compared with normal bone marrow) was observed early on [60, 61]. Subsequent studies based on microarray hybridizations, and RT-qPCR analysis confirmed this finding and showed that not only *MYC* but also *MYCN* is overexpressed in AML patients [33]. These data are in agreement with the observation that *MYC* is up-regulated in radiation-induced AML in mice [62]. *MYC* expression also appeared elevated in a microarray-based study in 5 AML samples, and was validated by RT-qPCR [63]. In contrast, *MYC* has not been detected as a major overexpressed gene in other microarray-based studies on AML samples [64–66]. However, it must be noted, first, that microarray-based studies are done at the mRNA level,

and thus changes in *MYC* protein level are not evaluated. Second, if *MYC* up-regulation is moderate (e.g., two-fold with respect to controls), it might be filtered out by the statistical analysis of microarray data. It should be mentioned, however, that a two-fold expression change of *MYC* may be relevant. For example, as noted above, transgenic mice lines with low *MYC* expression in the hematopoietic precursors develop myeloid leukemia; whereas, high *MYC* expression results in T cell lymphomas [37]. Also, a mere two-fold change means a major difference for *MYC*-dependent transformation of fibroblasts and mouse embryonic stem cells [67, 68] as well as in transgenic animals, where *MYC* dosage can be modulated [69]. This finding reveals that the impact of *MYC* in human leukemia is difficult to evaluate in the clinical setting, as two-fold increase in gene expression is usually not considered as significant when measuring mRNA expression in the clinical laboratories; whereas, as commented above, it could mean a difference in cell transformation. Finally, the great diversity of AML commented above makes the analysis of homogeneous sample cohorts difficult.

The mechanisms that lead to *MYC* overexpression in these cases are essentially unknown. Recurrent translocations in AML generate fusion proteins that are leukemogenic transcription factors, [38] and at least three of these (*RUNX1-ETO*, *PML-RAR $\alpha$* , and *PLZF-RAR $\alpha$* ) induce *MYC* expression. Thus, *MYC* could be a downstream target of these oncogenes [70, 71]. On the other side, *MYC* might be mediating the effects of some of these oncogenic translocation products. For example, it has been shown that *MYC* mediates the block in granulocytic differentiation brought about by *MLL-ENL* [72].

Cytogenetic hallmarks of gene amplification in tumors are double minute (dmin) chromosomes and homogeneously staining regions (hsr). In contrast to other tumors, both double minute chromosomes and homogeneously staining regions containing amplified regions are rare in AML. Although *MYC* amplification in AML is infrequent (less than 1 % of all cytogenetically abnormal leukemias, <http://cgap.nci.nih.gov/Chromosomes/Mitelman>), hsr and dmin including the region of 8q24 where *MYC* maps have been consistently described in AML and 8q24 is one of the most common amplified regions in dmin [73–82]. It has been suggested that a high *MYC* gene copy number does not necessarily result in higher *MYC* expression [79, 83]. However, recent reports show that *MYC* amplification is associated with higher *MYC* expression, with disease progression and poor prognosis [75]. Also, CD34<sup>+</sup> cells from MDS patients with trisomy 8 showed up-regulation of *MYC* mRNA [84]. This situation is in agreement with the correlation between *MYC* amplification and expression observed in lymphoma [85].

**Table 1** Summary of MYC alterations in myeloid malignancies

Leukemia	MYC deregulation	References
Chronic myeloid leukemia (CML)	MYC mRNA overexpression over healthy cells	[48, 49]
	High MYC mRNA and protein at diagnosis correlated with poor response to imatinib	[50]
	MYC protein elevated at diagnosis associated to progression. Altered MYC phosphorylation	[51]
Essential thrombocythemia (ET)	MYC mRNA overexpression	[58]
MDS	MYC mRNA up-regulation by microarray or RT-PCR	[84]
	MYC amplification (in dmin and hsr)	[86] [77, 78, 81, 84]
Acute myeloid leukemia (AML)	MYC amplification (in dmin)	[73–82]
	MYC mRNA overexpression by microarray analysis	[63]
AML (w/o translocations)	MYC mRNA overexpression by microarray (20 %)	[98]
AML (pediatric)	MYCN overexpression (24–40 %)	[33, 65]
AML (therapy-related)	MYC mRNA overexpression	[99]

The myelodysplastic syndromes (MDS) are also a heterogeneous group of clonal hematologic disorders characterized by both an aberrant differentiation process with morphologic evidence of marrow dysplasia in one or more of the three major myeloid cell types and an increased ineffective proliferation of the myeloid precursors in bone marrow, leading to cytopenia(s) and to an enhanced risk of transformation to an AML. Gene expression profiles of CD34<sup>+</sup> cells from MDS patients showed MYC as one of the most up-regulated genes in these patients [86]. Many patients with MDS, especially those with increased myeloblasts counts and adverse cytogenetic prognostic markers, will develop AML. MYC amplification has been reported with low frequencies in MDS [77, 78, 81, 84] (Table 1) but its prognostic impact is uncertain.

### MYC as a pharmacological target in leukemia

As most transcription factors, MYC is not an easy drugable target. However, using dominant negative forms (OmoMyc), it has been shown that in vivo OmoMyc expression abolishes the MYC-mediated skin carcinogenesis in animal models [87]. Moreover, it has been shown that MYC blocks the K-Ras and T-antigen-dependent carcinogenesis in lung and pancreas, respectively [88–90]. This result suggests that MYC inhibition would be a sensible antitumor approach even for tumors that show no MYC deregulation. Several small molecules have been described as MYC inhibitors, although none of them are in clinical use at the moment. Most of them impair the MYC-MAX interaction, thus inhibiting the transcriptional

activity of MYC [11]. This has the caveat that some of the MYC activities such as DNA replication, RNA pol II elongation, or CAP addition do not depend on the DNA binding activity of MYC [91, 92]. On the other hand, MYC requires other proteins for transformation, which may not be essential in normal cell physiology. Thus, a number of secondary targets capable of synthetic lethal interaction with MYC have been described, and thus proposed as putative pharmacological targets for MYC-driven tumors. This is the case of proteins as WRN [93], aurora kinase [94], CHK1 kinase [95] or CSKN1e kinase [96]. A different and recent approach is to block MYC expression by impairing BET bromodomain protein, a strategy shown to be effective in AML [97]. In summary, in vitro and in vivo models demonstrate a critical role of MYC in normal and malignant myelopoiesis. Therefore, the pharmacological targeting of MYC in myeloid leukemias is nowadays a challenge.

**Acknowledgments** This work, in the laboratory of the authors, was provided by grants SAF11-23796 and ISCIII-RETIC RD06/0020/0017 to JL, and FIS 11/00397 to MDD.

**Conflict of interest** The authors declare that they have no conflict of interest regarding the publication of this manuscript.

### References

- Iwasaki H, Akashi K (2007) Myeloid lineage commitment from the hematopoietic stem cell. *Immunity* 26:726–740
- Orkin SH, Zon LI (2008) Hematopoiesis: an evolving paradigm for stem cell biology. *Cell* 132:631–644
- Tsiftoglou AS, Bonovolias ID, Tsiftoglou SA (2009) Multilevel targeting of hematopoietic stem cell self-renewal, differentiation and apoptosis for leukemia therapy. *Pharmacol Ther* 122:264–280

4. Miranda-Saavedra D, Gottgens B (2008) Transcriptional regulatory networks in haematopoiesis. *Curr Opin Genet Dev* 18:530–535
5. Kim SI, Bresnick EH (2007) Transcriptional control of erythropoiesis: emerging mechanisms and principles. *Oncogene* 26:6777–6794
6. Dang CV, O'Donnell KA, Zeller KI, Nguyen T, Osthus RC et al (2006) The c-Myc target gene network. *Semin Cancer Biol* 16:253–264
7. Oster SK, Ho CS, Soucie EL, Penn LZ (2002) The myc oncogene: marvelousLY complex. *Adv Cancer Res* 84:81–154
8. Eilers M, Eisenman RN (2008) Myc's broad reach. *Genes Dev* 22:2755–2766
9. Leon J, Ferrandiz N, Acosta JC, Delgado MD (2009) Inhibition of cell differentiation: a critical mechanism for MYC-mediated carcinogenesis? *Cell Cycle* 8:1148–1157
10. Nesbit CE, Tersak JM, Prochownik EV (1999) MYC oncogenes and human neoplastic disease. *Oncogene* 18:3004–3016
11. Vita M, Henriksson M (2006) The Myc oncoprotein as a therapeutic target for human cancer. *Semin Cancer Biol* 16:318–330
12. Varmus HE (1984) The molecular genetics of cellular oncogenes. *Annu Rev Genet* 18:553–612
13. Sanchez-Beato M, Sanchez-Aguilera A, Piris MA (2003) Cell cycle deregulation in B-cell lymphomas. *Blood* 101:1220–1235
14. Delgado MD, Leon J (2010) Myc roles in hematopoiesis and leukemia. *Genes Cancer* 1:605–616
15. Sheiness D, Bishop JM (1979) DNA and RNA from uninfected vertebrate cells contain nucleotide sequences related to the putative transforming gene of avian myelocytomatosis virus. *J Virol* 31:514–521
16. Coppola JA, Cole MD (1986) Constitutive c-myc oncogene expression blocks mouse erythroleukaemia cell differentiation but not commitment. *Nature* 320:760–763
17. Prochownik EV, Kukowska J (1986) Deregulated expression of c-myc by murine erythroleukaemia cells prevents differentiation. *Nature* 322:848–850
18. Dmitrovsky E, Kuehl WM, Hollis GF, Kirsch IR, Bender TP et al (1986) Expression of a transfected human c-myc oncogene inhibits differentiation of a mouse erythroleukaemia cell line. *Nature* 322:748–750
19. Delgado MD, Lerga A, Canelles M, Gomez-Casares MT, Leon J (1995) Differential regulation of Max and role of c-Myc during erythroid and myelomonocytic differentiation of K562 cells. *Oncogene* 10:1659–1665
20. Acosta JC, Ferrandiz N, Bretones G, Torrano V, Blanco R et al (2008) Myc inhibits p27-induced erythroid differentiation of leukemia cells by repressing erythroid master genes without reversing p27-mediated cell cycle arrest. *Mol Cell Biol* 28:7286–7295
21. Ryan KM, Birnie GD (1997) Cell-cycle progression is not essential for c-Myc to block differentiation. *Oncogene* 14:2835–2843
22. Gomez-Casares MT, Delgado MD, Lerga A, Crespo P, Quincoces AF et al (1993) Down-regulation of c-myc gene is not obligatory for growth inhibition and differentiation of human myeloid leukemia cells. *Leukemia* 7:1824–1833
23. Bahram F, Wu S, Oberg F, Luscher B, Larsson LG (1999) Posttranslational regulation of Myc function in response to phorbol ester/interferon-gamma-induced differentiation of v-Myc-transformed U-937 monoblasts. *Blood* 93:3900–3912
24. Uribesalvo I, Buschbeck M, Gutierrez A, Teichmann S, Demajo S et al (2011) E-box-independent regulation of transcription and differentiation by MYC. *Nat Cell Biol* 13:1443–1449
25. Guo Y, Niu C, Breslin P, Tang M, Zhang S et al (2009) c-Myc-mediated control of cell fate in megakaryocyte-erythrocyte progenitors. *Blood* 114:2097–2106
26. Thompson A, Zhang Y, Kamen D, Jackson CW, Cardiff RD et al (1996) Deregulated expression of c-myc in megakaryocytes of transgenic mice increases megakaryopoiesis and decreases polyploidization. *J Biol Chem* 271:22976–22982
27. Luo H, Li Q, O'Neal J, Kreisel F, Le Beau MM et al (2005) c-Myc rapidly induces acute myeloid leukemia in mice without evidence of lymphoma-associated antiapoptotic mutations. *Blood* 106:2452–2461
28. Xiang Z, Luo H, Payton JE, Cain J, Ley TJ et al (2010) Mcl1 haploinsufficiency protects mice from Myc-induced acute myeloid leukemia. *J Clin Invest* 120:2109–2118
29. Beverly LJ, Varmus HE (2009) MYC-induced myeloid leukemogenesis is accelerated by all six members of the antiapoptotic BCL family. *Oncogene* 28:1274–1279
30. Hemann MT, Bric A, Teruya-Feldstein J, Herbst A, Nilsson JA et al (2005) Evasion of the p53 tumour surveillance network by tumour-derived MYC mutants. *Nature* 436:807–811
31. Yu D, Thomas-Tikhonenko A (2002) A non-transgenic mouse model for B-cell lymphoma: in vivo infection of p53-null bone marrow progenitors by a Myc retrovirus is sufficient for tumorigenesis. *Oncogene* 21:1922–1927
32. Yu D, Allman D, Goldschmidt MH, Atchison ML, Monroe JG et al (2003) Oscillation between B-lymphoid and myeloid lineages in Myc-induced hematopoietic tumors following spontaneous silencing/reactivation of the EBF/Pax5 pathway. *Blood* 101:1950–1955
33. Kawagoe H, Kandilci A, Kranenburg TA, Grosveld GC (2007) Overexpression of N-Myc rapidly causes acute myeloid leukemia in mice. *Cancer Res* 67:10677–10685
34. Hogstrand K, Hejll E, Sander B, Rozell B, Larsson LG et al (2012) Inhibition of the intrinsic but not the extrinsic apoptosis pathway accelerates and drives MYC-driven tumorigenesis towards acute myeloid leukemia. *PLoS ONE* 7:e31366
35. Skoda RC, Tsai SF, Orkin SH, Leder P (1995) Expression of c-MYC under the control of GATA-1 regulatory sequences causes erythroleukemia in transgenic mice. *J Exp Med* 181:1603–1613
36. Smith DP, Bath ML, Harris AW, Cory S (2005) T-cell lymphomas mask slower developing B-lymphoid and myeloid tumours in transgenic mice with broad haematopoietic expression of MYC. *Oncogene* 24:3544–3553
37. Smith DP, Bath ML, Metcalf D, Harris AW, Cory S (2006) MYC levels govern hematopoietic tumor type and latency in transgenic mice. *Blood* 108:653–661
38. Vardiman JW, Brunning RD, Arber DA, LeBeau MM, Porwit A et al (2008) Introduction and overview of the classification of the myeloid neoplasms. In: Swerdlow SH, Campo E, Harris NL, Jaffe ES, Pileri SA et al (eds) WHO classification of tumours of haematopoietic and lymphoid tissues, 4th edn. International Agency for Research on Cancer, Lyon, pp 18–37
39. Melo JV, Barnes DJ (2007) Chronic myeloid leukaemia as a model of disease evolution in human cancer. *Nat Rev Cancer* 7:441–453
40. Xie S, Lin H, Sun T, Arlinghaus RB (2002) Jak2 is involved in c-Myc induction by Bcr-Abl. *Oncogene* 21:7137–7146
41. Gomez-Casares MT, Vaque JP, Lemes A, Molero T, Delgado MD et al (2004) C-myc expression in cell lines derived from chronic myeloid leukemia. *Haematologica* 89:241–243
42. Lugo TG, Witte ON (1989) The BCR-ABL oncogene transforms Rat-1 cells and cooperates with v-myc. *Mol Cell Biol* 9:1263–1270
43. Sawyers CL, Callahan W, Witte ON (1992) Dominant negative MYC blocks transformation by ABL oncogenes. *Cell* 70:901–910
44. Afar DE, Goga A, McLaughlin J, Witte ON, Sawyers CL (1994) Differential complementation of Bcr-Abl point mutants with c-Myc. *Science* 264:424–426
45. Pan C, Olsen JV, Daub H, Mann M (2009) Global effects of kinase inhibitors on signaling networks revealed by quantitative phosphoproteomics. *Mol Cell Proteomics* 8:2796–2808
46. Handa H, Hegde UP, Kotelnikov VM, Mundle SD, Dong LM et al (1997) Bcl-2 and c-myc expression, cell cycle kinetics and apoptosis during the progression of chronic myelogenous leukemia from diagnosis to blastic phase. *Leuk Res* 21:479–489
47. Beck Z, Bacsi A, Kovacs E, Kiss J, Kiss A et al (1998) Changes in oncogene expression implicated in evolution of chronic granulocytic leukemia from its chronic phase to acceleration. *Leuk Lymphoma* 22:3952–3963
48. Nowicki MO, Pawlowski P, Fischer T, Hess G, Pawlowski T et al (2003) Chronic myelogenous leukemia molecular signature. *Oncogene* 22:3952–3963
49. Diaz-Blanco E, Bruns I, Neumann F, Fischer JC, Graef T et al (2007) Molecular signature of CD34(+) hematopoietic stem and progenitor cells of patients with CML in chronic phase. *Leukemia* 21:494–504
50. Albajar M, Gomez-Casares MT, Llorca J, Mauleon I, Vaque JP et al (2011) MYC in chronic myeloid leukemia: induction of aberrant DNA synthesis and association with poor response to imatinib. *Mol Cancer Res* 9:564–576
51. Lucas CM, Harris RJ, Giannoudis A, Copland M, Slupsky JR et al (2011) Cancerous inhibitor of PP2A (CIP2A) at diagnosis of chronic myeloid leukemia is a critical determinant of disease progression. *Blood* 117:6660–6668
52. Johansson B, Fioretos T, Mitelman F (2002) Cytogenetic and molecular genetic evolution of chronic myeloid leukemia. *Acta Haematol* 107:76–94
53. Brazma D, Grace C, Howard J, Melo JV, Holyoke T et al (2007) Genomic profile of chronic myelogenous leukemia: imbalances associated with disease progression. *Genes Chromosomes Cancer* 46:1039–1050
54. Sears RC (2004) The life cycle of C-myc: from synthesis to degradation. *Cell Cycle* 3:1133–1137
55. Porro A, Iraci N, Soverini S, Diolaiti D, Gherardi S et al (2011) c-MYC oncoprotein dictates transcriptional profiles of ATP-binding cassette transporter genes in chronic myelogenous leukemia CD34 + hematopoietic progenitor cells. *Mol Cancer Res* 9:1054–1066
56. Quintas-Cardama A, Cortes J (2009) Molecular biology of bcr-abl1-positive chronic myeloid leukemia. *Blood* 113:1619–1630
57. Gomez-Casares MT, Garcia-Alegria E, Lopez-Jorge CE, Ferrandiz N, Blanco R, et al. (2012) MYC antagonizes the differentiation induced by imatinib in chronic myeloid leukemia cells through downregulation of p27KIP1. *Oncogene*. doi: 10.1038/onc.2012.246
58. Theophile K, Buesche G, Kreipe H, Bock O (2008) The expression levels of telomerase catalytic subunit hTERT and oncogenic MYC in essential thrombocythemia are affected by the molecular subtype. *Ann Hematol* 87:263–268
59. Watanabe S, Itoh T, Arai K (1996) JAK2 is essential for activation of c-fos and c-myc promoters and cell proliferation through the human granulocyte-macrophage colony-stimulating factor receptor in BA/F3 cells. *J Biol Chem* 271:12681–12686
60. Ferrari S, Narni F, Mars W, Kaczmarek L, Venturelli D et al (1986) Expression of growth-regulated genes in human acute leukemias. *Cancer Res* 46:5162–5166
61. Calabretta B, Venturelli D, Kaczmarek L, Narni F, Talpaz M et al (1986) Altered expression of G1-specific genes in human malignant myeloid cells. *Proc Natl Acad Sci USA* 83:1495–1498
62. Hirouchi T, Takabatake T, Yoshida K, Nitta Y, Nakamura M et al (2008) Upregulation of c-myc gene accompanied by PU.1 deficiency in radiation-induced acute myeloid leukemia in mice. *Exp Hematol* 36:871–885

63. Court EL, Smith MA, Avent ND, Hancock JT, Morgan LM et al (2004) DNA microarray screening of differential gene expression in bone marrow samples from AML, non-AML patients and AML cell lines. *Leuk Res* 28:743–753
64. Valk PJ, Verhaak RG, Beijen MA, Erpelinck CA, Barjesteh van Waalwijk van Doorn-Khosrovani S et al (2004) Prognostically useful gene-expression profiles in acute myeloid leukemia. *N Engl J Med* 350:1617–1628
65. Ross ME, Mahfouz R, Onciu M, Liu HC, Zhou X et al (2004) Gene expression profiling of pediatric acute myelogenous leukemia. *Blood* 104:3679–3687
66. Stirewalt DL, Meshinchi S, Kopecky KJ, Fan W, Pogossova-Agadjanya EL et al (2008) Identification of genes with abnormal expression changes in acute myeloid leukemia. *Genes Chromosomes Cancer* 47:8–20
67. Bazarov AV, Adachi S, Li SF, Mateyak MK, Wei S et al (2001) A modest reduction in c-myc expression has minimal effects on cell growth and apoptosis but dramatically reduces susceptibility to Ras and Raf transformation. *Cancer Res* 61:1178–1186
68. Baudino TA, McKay C, Pendeveille-Samain H, Nilsson JA, Maclean KH et al (2002) c-Myc is essential for vasculogenesis and angiogenesis during development and tumor progression. *Genes Dev* 16:2530–2543
69. Murphy DJ, Junttila MR, Pouyet L, Karnezis A, Shchors K et al (2008) Distinct thresholds govern Myc's biological output in vivo. *Cancer Cell* 14:447–457
70. Muller-Tidow C, Steffen B, Cauvet T, Tickenbrock L, Ji P et al (2004) Translocation products in acute myeloid leukemia activate the Wnt signaling pathway in hematopoietic cells. *Mol Cell Biol* 24:2890–2904
71. Rice KL, Hormaeche I, Doulatov S, Flatow JM, Grimwade D et al (2009) Comprehensive genomic screens identify a role for PLZF-RARalpha as a positive regulator of cell proliferation via direct regulation of c-MYC. *Blood* 114:5499–5511
72. Schreiner S, Birke M, Garcia-Cuellar MP, Zilles O, Greil J et al (2001) MLL-ENL causes a reversible and myc-dependent block of myelomonocytic cell differentiation. *Cancer Res* 61:6480–6486
73. Sait SN, Qadir MU, Conroy JM, Matsui S, Nowak NJ et al (2002) Double minute chromosomes in acute myeloid leukemia and myelodysplastic syndrome: identification of new amplification regions by fluorescence in situ hybridization and spectral karyotyping. *Genes Chromosomes Cancer* 34:42–47
74. Slovak ML, Ho JP, Pettenati MJ, Khan A, Douer D et al (1994) Localization of amplified MYC gene sequences to double minute chromosomes in acute myelogenous leukemia. *Genes Chromosomes Cancer* 9:62–67
75. Rayeroux KC, Campbell LJ (2009) Gene amplification in myeloid leukemias elucidated by fluorescence in situ hybridization. *Cancer Genet Cytogenet* 193:44–53
76. O'Malley F, Rayeroux K, Cole-Sinclair M, Tong M, Campbell LJ (1999) MYC amplification in two further cases of acute myeloid leukemia with trisomy 4 and double minute chromosomes. *Cancer Genet Cytogenet* 109:123–125
77. Thomas L, Stamberg J, Gojo I, Ning Y, Rapoport AP (2004) Double minute chromosomes in monoblastic (M5) and myeloblastic (M2) acute myeloid leukemia: two case reports and a review of literature. *Am J Hematol* 77:55–61
78. Mathew S, Lorsbach RB, Shearer P, Sandlund JT, Raimondi SC (2000) Double minute chromosomes and c-MYC amplification in a child with secondary myelodysplastic syndrome after treatment for acute lymphoblastic leukemia. *Leukemia* 14:1314–1315
79. Storlazzi CT, Fioretos T, Surace C, Lonoce A, Mastroianni A et al (2006) MYC-containing double minutes in hematologic malignancies: evidence in favor of the episome model and exclusion of MYC as the target gene. *Hum Mol Genet* 15:933–942
80. Bruyere H, Sutherland H, Chipperfield K, Hudoba M (2010) Concomitant and successive amplifications of MYC in APL-like leukemia. *Cancer Genet Cytogenet* 197:75–80
81. Bajaj R, Xu F, Xiang B, Wilcox K, Diadamo AJ et al (2011) Evidence-based genomic diagnosis characterized chromosomal and cryptic imbalances in 30 elderly patients with myelodysplastic syndrome and acute myeloid leukemia. *Mol Cytogenet* 4:3
82. Micale L, Augello B, Daniele G, Macchia G, L'Abbate A et al (2011) Amplification of the G allele at SNP rs6983267 in 8q24 amplicons in myeloid malignancies as cause of the lack of MYC overexpression? *Blood Cells Mol Dis* 47:259–261
83. Paulsson K, Lassen C, Kuric N, Billstrom R, Fioretos T et al (2003) MYC is not overexpressed in a case of chronic myelomonocytic leukemia with MYC-containing double minutes. *Leukemia* 17:813–815
84. Sloand EM, Pfannes L, Chen G, Shah S, Solomou EE et al (2007) CD34 cells from patients with trisomy 8 myelodysplastic syndrome (MDS) express early apoptotic markers but avoid programmed cell death by up-regulation of anti-apoptotic proteins. *Blood* 109:2399–2405
85. Stasik CJ, Nitta H, Zhang W, Mosher CH, Cook JR et al (2010) Increased MYC gene copy number correlates with increased mRNA levels in diffuse large B-cell lymphoma. *Haematologica* 95:597–603
86. Vasikova A, Belickova M, Budinska E, Cermak J (2010) A distinct expression of various gene subsets in CD34 + cells from patients with early and advanced myelodysplastic syndrome. *Leuk Res* 34:1566–1572
87. Soucek L, Nasi S, Evan GI (2004) Omomyc expression in skin prevents Myc-induced papillomatosis. *Cell Death Differ* 11:1038–1045
88. Soucek L, Whitfield J, Martins CP, Finch AJ, Murphy DJ et al (2008) Modelling Myc inhibition as a cancer therapy. *Nature* 455:679–683
89. Fukazawa T, Maeda Y, Matsuoka J, Yamatsuji T, Shigemitsu K et al (2010) Inhibition of Myc effectively targets KRAS mutation-positive lung cancer expressing high levels of Myc. *Anticancer Res* 30:4193–4200
90. Sodr NM, Swigart LB, Karnezis AN, Hanahan D, Evan GI et al (2011) Endogenous Myc maintains the tumor microenvironment. *Genes Dev* 25:907–916
91. Meyer N, Penn LZ (2008) Reflecting on 25 years with MYC. *Nat Rev Cancer* 8:976–990
92. Cole MD, Cowling VH (2008) Transcription-independent functions of MYC: regulation of translation and DNA replication. *Nat Rev Mol Cell Biol* 9:810–815
93. Moser R, Toyoshima M, Robinson K, Gurley KE, Howie HL et al (2012) MYC-driven tumorigenesis is inhibited by WRN syndrome gene deficiency. *Mol Cancer Res* 10:535–545
94. Yang D, Liu H, Goga A, Kim S, Yuneva M et al (2010) Therapeutic potential of a synthetic lethal interaction between the MYC proto-oncogene and inhibition of aurora-B kinase. *Proc Natl Acad Sci USA* 107:13836–13841
95. Murga M, Campaner S, Lopez-Contreras AJ, Toledo LI, Soria R, Montaña MF, D'Artista L, Schleker T, Guerra C, Garcia E, Barbacid M, Hidalgo M, Amati B, Fernandez-Capetillo O (2011) Exploiting oncogene-induced replicative stress for the selective killing of Myc-driven tumors. *Nat Struct Mol Biol* 18:1331–1335
96. Toyoshima M, Howie HL, Imakura M, Walsh RM, Annis JE et al (2012) Functional genomics identifies therapeutic targets for MYC-driven cancer. *Proc Natl Acad Sci USA* 109:9545–9550
97. Delmore JE, Issa GC, Lemieux ME, Rahl PB, Shi J et al (2011) BET bromodomain inhibition as a therapeutic strategy to target c-Myc. *Cell* 146:904–917
98. Larramendy ML, Niini T, Elonen E, Nagy B, Ollila J et al (2002) Overexpression of translocation-associated fusion genes of FGFR1, MYC, NPML, and DEK, but absence of the translocations in acute myeloid leukemia. A microarray analysis. *Haematologica* 87:569–577
99. Qian Z, Fernald AA, Godley LA, Larson RA, Le Beau MM (2002) Expression profiling of CD34+ hematopoietic stem/progenitor cells reveals distinct subtypes of therapy-related acute myeloid leukemia. *Proc Natl Acad Sci USA* 99:14925–14930