



# Anaerobic Digestion of Agri-Food Wastes for Generating Biofuels

Chunjie Gong<sup>1</sup> · Ankit Singh<sup>2</sup> · Pranjali Singh<sup>3</sup> · Archana Singh<sup>4</sup>

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**Abstract** Presently, fossil fuels are extensively employed as major sources of energy, and their uses are considered unsustainable due to emissions of obnoxious gases on the burning of fossil fuels, which can lead to severe environmental complications, including human health. To tackle these issues, various processes are developing to waste as a feed to generate eco-friendly fuels. The biological production of fuels is considered to be more beneficial than physicochemical methods due to their environmentally friendly nature, high rate of conversion at ambient physiological conditions, and less energy-intensive. Among various biofuels, hydrogen (H<sub>2</sub>) is considered as a wonderful due to high calorific value and generate water molecule as end product on the burning. The H<sub>2</sub> production from biowaste is demonstrated, and agri-food waste can be potentially used as a feedstock due to their high biodegradability over lignocellulosic-based biomass. Still, the H<sub>2</sub> production is uneconomical from biowaste in fuel competing market because of low yields and increased capital and operational expenses. Anaerobic digestion is widely used for waste management and the generation of value-added products. This article is highlighting the

valorization of agri-food waste to biofuels in single (H<sub>2</sub>) and two-stage bioprocesses of H<sub>2</sub> and CH<sub>4</sub> production.

**Keywords** Anaerobic digestion · Agri-food waste · Biohydrogen · Biomethane · Integrative bioprocesses · Value-added products

## Introduction

Nature is progressing via sustainable mechanisms. Therefore living organisms are strongly harmonized through environmental changes. Energy utilization is significantly increasing in developed countries as compared to developing countries, and nearly 15% of the World's population is consuming over half of the total energy consumption [1, 2]. The exponential increase of world populations (~ 7.9 billion) in the past few decades is pressuring too much burden for sustainable development. Primarily, we rely on fossil-based sources to fulfill its increasing energy demands in societal and industrial areas [3–5]. The economic development hinders due to deteriorating stocks of non-renewable energy-based assets. An alternative to these energy sources, biofuels-based energy sources such as hydrogen (H<sub>2</sub>) [6–8], biogas mainly methane (CH<sub>4</sub>) [9, 10], ethanol [11], methanol [12–15], and biodiesel [16, 17], are more helpful to minimize the emission of harmful gases via the burning of fossil fuels and also their eco-friendly nature. A large quantum biowaste(s) is generated through our daily lives and various human activities [18–20]. Thus, the utilization of biowaste(s) for generating useful for various kinds of biomolecules such as biofuels [21–25], biopolymers such polyhydroxyalkanoates (PHAs) [26–30], and bioelectricity [31, 32]. Biological processes have been proved more beneficial for biotransformation applications

✉ Archana Singh  
archana.singh7891@gmail.com

<sup>1</sup> National “111” Center for Cellular Regulation and Molecular Pharmaceutics, Key Laboratory of Fermentation Engineering (Ministry of Education), Hubei University of Technology, Wuhan 430068, People's Republic of China

<sup>2</sup> National Institute of Technology, Sikkim 737139, India

<sup>3</sup> National Institute of Technology, Warangal 506004, India

<sup>4</sup> Department of Bioinformatics, Mahila Mahavidyalaya, Banaras Hindu University, Varanasi 21005, India

than physical or chemical methods, which are primarily considered high energy-intensive processes [33–38]. Further, the biocatalyst's properties can be significantly improved through genetic and protein engineering or related synthetic approaches for their potential applications [39–44]. Also, biological-derived products, materials or microbes themselves can be potentially applied in the area of microbial pathogenesis to improve microbes, human, and plants health [45–53]. Recent pandemic arising due to viral infection is a significant influence of human thinking for better management of population sustainability and environmental issues research over other non-related areas [54–56].

The energy resources-based predictions suggested that coal deposits will be utilized over the next Century. In contrast, petroleum-based deposits will be used up within few decades [10, 57]. Also, the environmental worsening is a significant concern, which is significantly associated with the extensive uses of these non-renewable energies. Renewable energy resources are vital for sustainable development [10, 58–61]. However, from the past few decades, alternative energy sources to fossil fuels are recognized as a significant area of research. The production of biofuels, especially  $H_2$  can be more beneficial using biowaste as a low-cost feed over costly pure sugar [1, 19, 62–64]. The various approaches have been used for the utilization of biowaste(s)-based feedstocks to produce biofuels such as  $H_2$  and  $CH_4$  [10]. The production of these biofuels is extensively studied using various biowastes from agricultural, municipal, industrial, and synthetic origins. The production of  $H_2$  is largely demonstrated using mixed cultures (MCs) over pure cultures as an inoculum due to their better metabolism [10, 65, 66]. The use of integrative processes such as  $H_2$  production followed by  $CH_4$  can be adopted at a large scale to improve the bioprocess economy. This article presents the status of the production of biofuels from agri-food waste in single- and two-stage. Further, the bioprocess improvement strategies for sustainable development have been discussed.

## Biowastes

Globally, a significant advancement in life-routine and industrialization has generated a severe problem by accumulating various kinds of waste (including biowastes) and their negative environmental impact [3, 67, 68]. The challenges of their management have earned considerable public and political recognition in current times. Therefore, minimization of wastes generation through their management is highly recommended for sustainable development. In addition, we are highly relying on unsustainable fossil fuels-based energy sources that can lead to environmental

pollution via the emission of harmful gases, and they (fossil fuels) may be depleted in the following centuries. However, in the past few decades, the generation of biofuels such as  $H_2$ ,  $CH_4$ , methanol, ethanol, and biodiesel is demonstrated as an alternative to fossil fuels [9, 11, 17, 69]. The production of biofuels from biowastes can be carried out to solve these waste management issues and biofuels and the environmental benefits. The primary sources of wastes can distribute in various groups based on their origin, such as agricultural, industrial, municipal, and biomedical [1, 19]. The quantum of wastes can be varied at regional and cultural levels. Despite the numerous environmental regulations and rules, a small level has been accomplished primarily in developing countries to minimize the generation of wastes [10]. In recent times, the generation of wastes in Indian major cities is escalating high rate ( $\sim 1.5\%$ ) of total wastes quantum [2]. However, the handling of a large quantum of waste is needed through practical methods in an economical manner. Various wastes management technologies have been used, includes—(1) AD, (2) composting, (3) incineration, (4) landfilling, (5) recycling, and (6) dumping (especially in the open) [10]. These methods can be used individually or in combination for effective waste management and showed some benefits over each other. The brief benefits of different waste management methods such as landfilling and dumping (open) are widely adopted globally, contributing up to 80% of the total waste management methods presented in Table 1 [10]. In contrast, AD and composting are equally used with very low combined contributions of 10–12% that is equal to the recycling method. The agri-food waste such as cereals (no-edible parts), fruits, and vegetables are generated in considerable amounts in markets. These kinds of biowaste are highly biodegradable that can be easily managed via their valorization to value-added products or other environmental applications [66, 70–72]. However, biowastes-based generation of biofuels is considered to be potentially applicable technologies for sustainable development.

## Biofuels Production from Biowastes

The major biofuels such as  $H_2$  [8, 10],  $CH_4$  [10, 73], methanol [74, 75], ethanol [11], biodiesel [17], production is expected to reduce global warming. These are probable to take fundamental developments in biofuels production [10]. The biofuels production are broadly classified into four generations: (1) 1st generation—this type of biofuels (biodiesel, bioethanol, biogas) is produced largely from agricultural-based crops, sugarcane, sugar beet, wheat, rice, corn, and sunflower through hydrolysis and fermentation, (2) 2nd generation—this generation of fuels are produced

**Table 1** The management procedures for valorization or disposal of wastes

Process	Contribution (%)	Benefits
Anaerobic digestion	6.30	Provide renewable energy (biogas) and/to generate electricity Reduce pollution, smell, pathogens, and weed seeds Conservation of agricultural land Generate fertilizer
Composting	5.05	Embolden microorganisms to produce humus (nutrient-filled materials) Soil enrichments and conquer plant infections and pests Decrease methane emissions Reduce chemical fertilizers requirement
Incineration	6.45	Reduce waste quantity, and efficient waste management Generation of energy and pollution reduction It prevents methane generation and operated in any weather Reduce harmful microbes and chemicals
Landfilling	37.4	Advanced landfills are eco-friendly, and an excellent energy source An easy method to keep clean city and town Helpful to manage all kinds of wastes Economical
Dumping (in open)	32.2	The simplest method and requires a small area Very economical Convenient Source for shelter and nutrients
Recycling	12.6	Provide a livable environment for a sustainable future Reduce quantity for waste management by other methods Conserve natural resources Improve economy and save energy

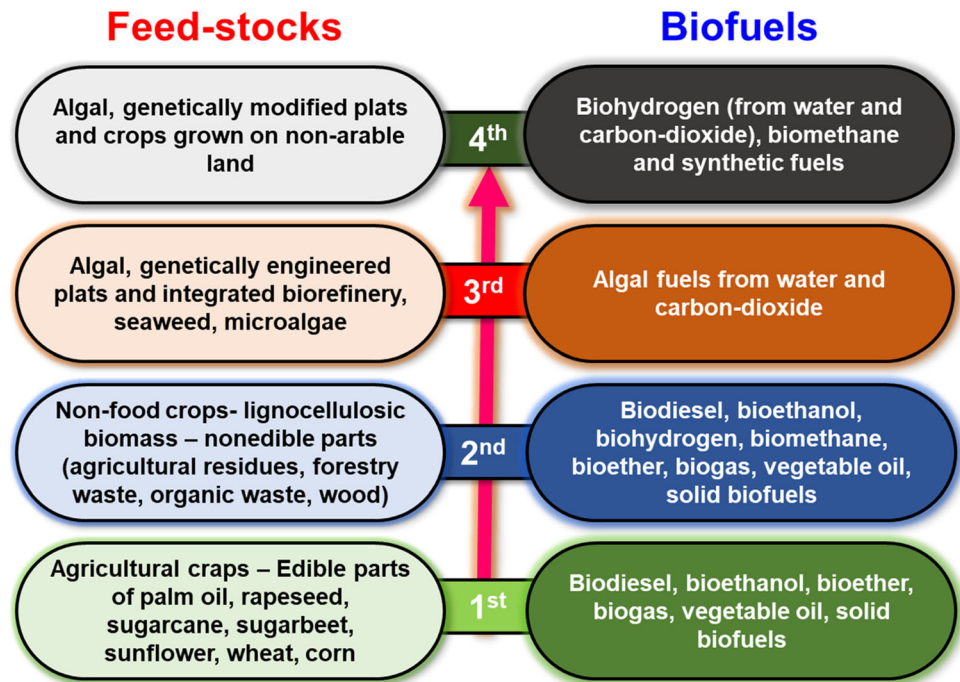
using non-edible plant parts, (3) 3rd generation—biofuels such as ethanol and biodiesel were produced via photosynthetic algae and genetically engineered plants through biochemical and thermochemical bioprocesses, and (4) 4th generation—this type of biofuels are produced through advanced photobiological solar or electric fuels (Fig. 1). The main drawback of this generation of fuels is a conflict of “food vs. fuel” [76].

The selection of suitable fuel for future uses can meet different criteria such as (1) convenient in transportation, (2) safe to use, (3) easily transform to another form of energy, (4) environmentally friendly nature, (5) high utilization efficiency, and (6) inexpensive to use [1, 10]. Among various available biofuels based on the above criteria, H<sub>2</sub> can be considered as a wonder fuel for sustainable development. Biologically H<sub>2</sub> has been produced from numerous microbes by using cheap raw materials such as biowastes. Lignocellulose-based biowastes are abundantly accessible [77]. Due to their complex nature, the pretreatment of biowastes is considered a suitable approach to produce soluble sugars for easy utilization towards biofuels (H<sub>2</sub>) through fermentation. Primarily, lignocellulosic

biowastes are consists of cellulose, hemicellulose and lignin. However, the hydrolysis of biomass largely depends on the type of pretreatment approaches due to significant variations in their compositions. The different pretreatments of biomass approaches have been used to generate fermentable sugars, includes physical (microwave and pyrolysis), chemical (acidic and alkaline), (3) physical-chemical-based (ultra-sonication and steam explosion), and (4) biological (microbial and enzymatic) [11, 76]. In the case of enzymatic pretreatment of biowastes, the following cellulase, xylanase,  $\beta$ -glucosidase, and laccase can be used for direct hydrolysis or to decrease the toxicity of hydrolysate [8, 78–81].

The biological pretreatment methods can be considered as eco-friendly as compared to physical, chemical, or their combinations [10, 76]. Still, the economic H<sub>2</sub> production from biowaste is challenging due to partial utilization of feed and bioprocess scaling-up. Also, the present production cost of H<sub>2</sub> through biological routes is higher than available energy sources. In general, the integrative approaches are proved more beneficial for value-added bioproducts that can improve the process economy.

**Fig. 1** The generations of biofuels production from various feed-stocks



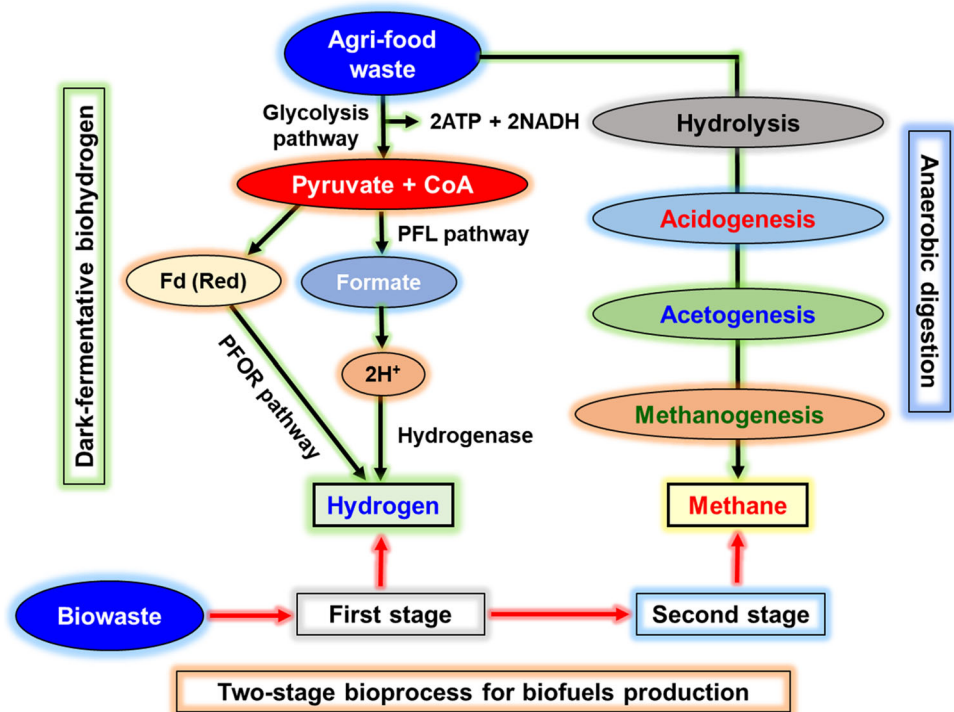
Various integrative approaches such as  $H_2$  followed photo fermentative  $H_2$ ,  $CH_4$  or PHAs have been reported [10, 21, 66]. The utilization of PHAs for the biotechnological applications can be more useful because of their novel therapeutic uses such as antimicrobial, tissue engineering, and drugs carrier [82–86]. Also, the techno-economics analysis suggested that these integrative processes will be more desirable over single-stage  $H_2$  production from sugars or biowastes [10, 21].

### Anaerobic Digestion

AD is considered one of the oldest bioprocesses for wastes utilization. Biowastes are very complex; thus, different strategies such as AD have been employed for their valorization to useful bioproducts such as  $H_2$  and  $CH_4$  [10, 57]. AD is a multi-step process and primarily carried to utilize complex materials such as biowastes using indigenous microbial populations or externally added cultures. The AD is carried out in four steps that are classified as (a) hydrolysis, (b) acidogenesis, (c) acetogenesis and (d) methanogenesis [10]. In the AD 1st step, the biowaste (complex organics) are hydrolyzed to simple sugars, fatty and amino acids by hydrolytic enzymes such as amylase, cellulase, protease, and lipase activity of microbial cultures. This group of cultures is known as hydrolytic fermentation bacteria, and they provide hydrolyzed substrates to the next step of the bacterial population (Acidogenesis). At the 2nd step of acidogenesis (fastest step in the AD), the

partially hydrolyzed substrate was further broken down by enzymatic reaction of cultures. Acidogenic bacteria are very fast growing with lower than an hour of doubling time and especially generates volatile fatty acids (VFAs), and gases, includes  $H_2$ , carbon dioxide ( $CO_2$ ), and ammonia. The 3rd stage of AD is known as acetogenesis, and during this stage, largely acetic acid is produced by acetogens along with  $H_2$  and  $CO_2$ . This stage microbial population is slow-growing with a more significant doubling time about 50-fold higher to acidogens (2nd stage). Thus, this stage's success primarily depends on cooperation between their microbial populations to achieve better efficiency. The 4th stage of AD is known as methanogenesis and is considered the terminal stage of AD (Fig. 2). At this step, methanogens are producing  $CH_4$  directly from acetate or  $H_2$  and  $CO_2$  mixture as a biogas [10, 18]. Methanogens are phylogenetically diverse groups of unique bacteria that are called archaeobacteria. Through the AD of biowastes, the biological oxygen demand, as well as chemical oxygen demand (COD), can be significantly reduced, and this process can all offer various environmental, and socio-economic benefits via the generation of renewable fuels. In addition to numerous benefits, AD can exhibit limitations such as strict anaerobic conditions requirement susceptible towards even low presence of oxygen amount) concentrations, and slow metabolic activities of methanogens [1, 10]. Apart from  $H_2$  and  $CH_4$ , the VFAs generated during the acidogenesis stage in AD can be potentially used to produce PHAs.

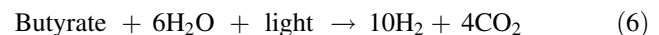
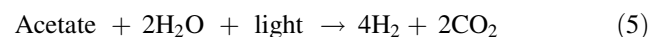
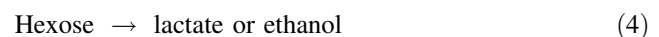
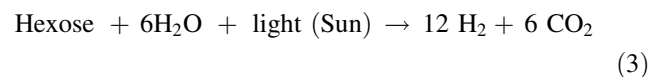
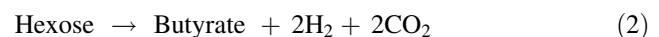
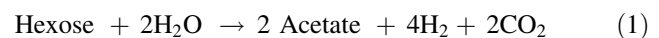
**Fig. 2** Bioprocess illustrations for the first-stage (hydrogen) and second-stage (hydrogen and methane) biofuels production from agri-food waste



### Biohydrogen Producers and Their Biodiversity

Among various candidates, H<sub>2</sub> is recognized as a promising future fuel due to its high caloric energy (141.9 MJ/kg) and non-polluting potential [10]. The H<sub>2</sub> can be produced using natural gases, biomass, coal, and fossil fuels. In the present scenario, ~ 90% of H<sub>2</sub> is produced through fossil-fuels [1, 10]. Biologically produced H<sub>2</sub> showed benefits like moderate production conditions, and an environmental-friendly bioprocess over various physicochemical processes [65]. The biological methods to produce H<sub>2</sub>, include—dark-fermentation (DF), photo-fermentation, photolysis, and electrochemical processes. The fermentative H<sub>2</sub> generation is a novel aspect, and it is considered suitable when biowaste is used as feed. H<sub>2</sub> production is occurred by hydrogenases through excess protons release via reversible reaction of H<sub>2</sub> ↔ 2H<sup>+</sup> + 2e<sup>-</sup> [1–3]. Based on the type of metal contents, hydrogenases are categorized into [Fe–Fe]- (naturally involves for H<sub>2</sub> generation), [NiFe]- (such as uptake-hydrogenases, bidirectional cytoplasmic-hydrogenases, cytoplasmic H<sub>2</sub> sensors and cyanobacterial uptake-hydrogenases, and H<sub>2</sub>-evolving hydrogenases), and [Fe]-containing enzymes. The metabolic pathway of H<sub>2</sub> involves the generation of pyruvate from glucose via Embden–Meyerhof–Parnas cycle or glycolytic pathway [3]. Further, formate is produced from pyruvate through pyruvate formate lyase. The generation of H<sub>2</sub> involved different pathways into facultative (such as *Escherichia* via hydrogenase and formate-dehydrogenase)

and strict anaerobic (like *Clostridium* through pyruvate ferredoxin oxidoreductase (POR) and H<sub>2</sub>-POR) organisms (Fig. 2). In photo-fermentation H<sub>2</sub> evolution occurs in bacterial by nitrogenase via capturing solar energy [19]. The biotransformation of hexose to H<sub>2</sub> by dark- and photo-fermentative organisms are demonstrated as following from Eqs. 1, 2, 3, 4, 5, 6 [1, 10]:



The taxonomically diverse microbes have been used to generate H<sub>2</sub>—(1) Archaea such as *Methanobacterium*, *Pyrococcus* and *Methylotrophs*; (2) Actinobacteria such as *Mycobacterium*; (3) Cyanobacteria like *Anabaena*, *Calothrix*, *Nostoc*, and *Spirulina*; (4) Firmicutes such as *Bacillus*, *Clostridium*, *Caldicellulosiruptor*, and *Frankia*; (5) Bacteroidetes or Chlorobi like *Acetomicrobium*, *Chlorobium*, and *Bacteroides*; (6) Thermotogae such as *Thermotoga*; (7) Fusobacteria like *Fusobacteriia*; (8) Alpha-proteobacteria such as *Rhizobium*, *Rhodobacter*, and *Rhodospseudomonas*; (9) Beta-proteobacteria like *Alcaligenes* and *Rubrivivax*; (10) Delta-proteobacteria such as

*Desulfovibrio*; (11) Epsilon-proteobacteria like *Campylobacter*; and (12) Gamma-proteobacteria like *Azotobacter*, *Enterobacter*, *Escherichia*, *Pseudomonas*, *Citrobacter* and *Klebsiella* [1]. Overall, along with a few unique H<sub>2</sub>-producers, a lower H<sub>2</sub> production to stoichiometric yield has been described. In DF production, H<sub>2</sub>-producers like *Bacillus*, *Clostridium*, *Caldicellulosiruptor*, and *Enterobacter* have shown yield ~ 3.8 mol of H<sub>2</sub>/mol of glucose [19]. Whereas photo-fermentative H<sub>2</sub>-producers like *Rhodobacter* and *Rhodospseudomonas* have reported yield ~ 9.0 mol of H<sub>2</sub>/mol of hexose [10, 87]. The key benefits are associated with DF over photo-fermentative include—lower energy input, and high production efficiency. The fermentative H<sub>2</sub> yield can be improved by various approaches such as (1) pretreatment of biowaste as feed, (2) uses of nanoparticles and metal ions, (3) use of selective defined MCs (DMCs) over pure culture, (4) co-digestion of feed, (5) use of metabolically engineered H<sub>2</sub>-producers. The H<sub>2</sub>-produces can be engineered to eliminate lactate dehydrogenase, uptake hydrogenase, or fumarate reductase encoding genes [1, 10]. These genetically modified H<sub>2</sub>-producers are limited by the fact that H<sub>2</sub> production is associated with undesirable influences such as lower yield and poor utilization of feed [1, 10, 88]. Overall, H<sub>2</sub> production by engineered microbes can be boosted through inhibition of H<sub>2</sub> production competitive pathways, designing unique pathways, or over-expressing genes related to H<sub>2</sub>-production [10]. Alternatively, the uses of immobilization of biocatalysts (either cell-free or cell-based systems especially enzymes) are well stabilized to improve various biotransformations [89–97]. Numerous kinds of support such as solids and polymeric materials have been used to developed efficient biocatalysts especially whole cells [14, 36, 67, 77]. Additionally, the uses of low-cost supports such as lignocellulosic-derived biowastes can be more beneficial for economical biotransformation over costly polymers. However, immobilized H<sub>2</sub>-producers can be potentially enhanced H<sub>2</sub> yield over free cells, especially under continuous culture conditions [2, 77]. Nanomaterials play a crucial role in biohydrogen production and improved yield up to sixfold as compared to control [10, 64]. Also, nanomaterials exhibit selective antimicrobial properties towards specific organisms that potentially can be effectively employed for the enrichment of H<sub>2</sub>-producers in mixed populations containing non-producers [64, 98, 99].

## Biofuels Production from Agri-Food Wastes

### Single-Stage Biohydrogen

The maximum 2 and 4 mol/mol of glucose can be produced through the generation of acetate and butyrate as soul metabolite intermediates, respectively [1, 19]. In contrast, H<sub>2</sub> generation is inhibited in the fermentative conversion of hexose to lactate or ethanol. From the past few decades, primarily various initiatives carried out to identify efficient H<sub>2</sub>-producers with desirable features to use diverse kinds of feed. Broadly, undefined MCs (UMCs) have been adopted to produce H<sub>2</sub> from biowaste over pure cultures due to their higher substrate specificity and stability towards undesirable changes during fermentation like pH and feed. Still, lower H<sub>2</sub> yields are achieved to 4 mol/mol of hexose because of the generation of undesirable metabolite intermediates such as butyrate, propionate, lactate, and ethanol instead of acetate [1, 10]. The production of H<sub>2</sub> is highly varied by the composition of feed. The agricultural-based food wastes composition for cellulose, hemicellulose, and lignin are presented in Table 2. The cellulosic (cellulose and hemicellulose) and lignin contents are highly varied among wastes. However, the production of H<sub>2</sub> is mainly dependent on the cellulosic content of wastes and the potential of H<sub>2</sub>-producers to metabolize them directly or after pretreatment [100].

The H<sub>2</sub> production under batch and continuous culture conditions from various agricultural-based food waste has been shown in Table 3. Under batch conditions, the H<sub>2</sub> production of ranges from 8.3 L/kg of COD to 239 L/kg of feed [101, 102]. Whereas H<sub>2</sub> yields from 54.0 L/kg of total solids (TS) to 635 L/kg of volatile solids (VS) under continuous mode [21, 103]. Overall, these studies suggested that the continuous mode of H<sub>2</sub> production is more beneficial to achieve nearly 6.5-folds better H<sub>2</sub> yield than the batch culture conditions. Agri-food wastes such as *Agave tequilana* bagasse, cheese whey, rice husk, sugar beet, and sugarcane molasses reported H<sub>2</sub> yield of 0.92–2.10 mol/mol of glucose [104–108]. Among these feeds and cultures, the association of molasses and *Caldicellulosiruptor saccharolyticus* DSM 8903 founded more beneficial to achieve a maximum yield of 2.10 mol/mol of glucose over other cultures either in pure form (*Bacillus cereus* and *Clostridium thermocellum* DSMZ 1313) or anaerobic sludge as UMCs and different biowastes combinations. Under batch mode, the combination of potato peels with *Parageobacillus thermoglucosidasius* KCTC 33,548 and DMCs resulted in yields up to 0.83 L/L of feed and 92 L/kg of TS, respectively [5, 59, 109]. In contrast, potato starch founded more beneficial to achieve higher production of 151 L/kg of feed

**Table 2** Cellulose, hemicellulose, and lignin composition of few agricultural origin wastes

Agricultural waste	Cellulose	Hemicellulose	Lignin	Others
Rice husk	35.1–41.1	17.6–38.3	18.8–26.6	11.8–22.5
Banana peels	11.5–44.0	18.4–25.5	8.05–9.80	29.5–53.3
Barley bran	37.1–44.1	30.4–34.9	19.8–25.5	8.20–19.4
Sugarcane bagasse	39.2–58.2	9.20–25.8	13.4–18.4	16.6–19.2
Apple pomace	36.0–42.5	11.0–18.8	19.0–23.7	15.0–34.0
Cassava	38.8–56.5	7.2–12.6	11.8–12.2	18.7–42.8
Olive husk	31.9–36.4	21.9–26.8	26.0–26.5	10.8–19.7

[110]. The supplementation of glucose to pea-shells hydrolysate recorded high production up to 75.0 L/kg of TS over pea-shells (microbially hydrolyzed) as compared to pea-shells with yields of 65.0 L/kg of TS [20, 21]. Also, the organic fraction of municipal solid waste (OFMSW) showed quite similar H<sub>2</sub> production of 62.5 L/kg of VS under batch-mode by UMCs [111]. These findings suggested that a suitable combination of feed and H<sub>2</sub>-producing cultures can be desirable to achieve a high yield.

The vegetables, fruit, and cheese whey mixture exhibited ~ 44-folds lower H<sub>2</sub> yields to those recorded of 553 L/kg of VS from mixed fruit wastes (Table 3) [101, 103]. The combinations of the various agri-biowastes mixture (two to six different combinations) along with corresponding controls proved beneficial to produce H<sub>2</sub> by DMCs and the high H<sub>2</sub> production varied between 54.0 and 102 L/kg of TS [5, 59, 62]. Similarly, higher H<sub>2</sub> yield of 166, and 199 L/kg of feed from cassava and sweet potato-based starch, respectively, were also reported [110]. In contrast, an association of banana, grape, melon, orange to MCs noted maximum H<sub>2</sub> yields up to 403 L/kg of VS under continuous mode [103]. In batch mode, the H<sub>2</sub> production from food waste recorded a higher production of 220 L/kg of feed over 35.1, 93.4, and 119 L/kg of feed from cassava pulp, cheese whey, and cassava waste, respectively [101, 112, 113]. Based on yield among the various agri-food wastes, apple waste can be potentially utilized as a suitable feed for commercial biohydrogen production in the near future.

### Two-Stage Process of Biohydrogen and Methane

In general, under single-stage DF H<sub>2</sub> production the partial valorization of biowaste has occurred and bioprocess seems less economical due to the maximum H<sub>2</sub> yield achievable of only 33% to total theoretical production of 12 mol/mol of hexose [1, 19]. Therefore, to improve DF process efficiency, various over integrating approaches have been demonstrated to produce value-added biofuels biomolecules (H<sub>2</sub>, CH<sub>4</sub>, butanol, and biodiesel) and eco-friendly biodegradable polymers (PHAs) a substitute to manmade plastics [10, 70]. Thus, such integrative bioprocesses as the

biorefinery approach can endorse better management of wastes and environmental pollution along with the generation of various renewable products. The combination of DF H<sub>2</sub> generation followed with AD to produce CH<sub>4</sub> can achieve almost complete utilization of feed (Fig. 2). [1, 10]. The two-stage integrative generation of H<sub>2</sub> and CH<sub>4</sub> from agri-food wastes is presented in Table 4. Generally, the MCs inoculum employed at the H<sub>2</sub> production stage requires pretreatment like heat to enrich H<sub>2</sub>-produces and minimize methanogens (H<sub>2</sub> consumers). In contrast, the CH<sub>4</sub> stage inoculum can be directly used as an inoculum instead of any initial pretreatment (naturally selected). Giordano et al. demonstrated integrative production of 177 L of H<sub>2</sub>/kg of COD, and 243 L of CH<sub>4</sub>/kg of COD from wheat (Common and durum), mashed and steamed peels of potato, respectively. These findings suggested that feed can significantly altered the production of H<sub>2</sub> and CH<sub>4</sub> by granular sludge [114]. In contrast, a quite similar production of H<sub>2</sub> and CH<sub>4</sub> was recorded from potato peels and rice by anaerobic sludge as inoculum [115].

Agri-food pure wastes such as banana peels, beans, cassava residues, potato, cheese whey, and sugarcane bagasse reported H<sub>2</sub> and CH<sub>4</sub> up to 253 and 507 L/kg of total VS (TVS), respectively (Table 4) [116–120]. In contrast, a lower H<sub>2</sub> up to 87.5 L/L of feed and higher CH<sub>4</sub> yields up to 570 L/kg of COD observed from OFMSW as a mixture of agri-food to other type wastes [121–123]. A quite comparable production of 223 L of H<sub>2</sub>/kg of VS and 277 L of CH<sub>4</sub>/kg of VS was noted from rice residue and *Chlorella pyrenoidosa* [124]. The association of food wastes resulted in yields of H<sub>2</sub> and CH<sub>4</sub>—(1) up to 218 and 432 L/kg of VS by anaerobic sludge, and (2) 135 and 510 L/kg of VS by seed sludge as inoculum, respectively [125–127]. Similarly, food waste in different combinations to olive husk, garden, and activated sludge produced up to 87.0 NL of H<sub>2</sub>/kg of VS and 682 L of CH<sub>4</sub>/kg of TS [128–130]. Significant variations are observed to integrative generation of H<sub>2</sub> and CH<sub>4</sub> from agri-food wastes that can be associated with compositions of sugars in feed, fermentation conditions, mode of production, and types of inoculums (Table 2). Overall, among the other agri-food wastes, potato, and a mixture of vegetables to other wastes

**Table 3** Biohydrogen generation by dark-fermentation of various agri-food wastes

Agri-biowaste	Culture	Biohydrogen		Reference
		Mode	Yield	
Agave <i>tequilana</i> bagasse	Anaerobic sludge	Continuous	1.53 mol/mol of hexose	[105]
Agri-biowaste mixtures	Defined mixed cultures (DCMs)	Batch	54.0–102 L/kg of TS	[5, 62]
Apple	Mixed culture	Continuous	635 L/kg of VS	[103]
Apple pomace	DCMs	Batch	60.0–83.0 L/kg of TS	[5, 59]
Banana	Mixed culture	Continuous	403 L/kg of VS	[103]
Cassava pulp	Soil-based mixed culture	Batch	35.1 L/kg of feed	[113]
Cassava starch	Anaerobic sludge	Batch	166 L/kg of starch	[110]
Cassava waste	Cattle dung	Batch	119 L/kg of feed	[112]
Cheese whey	Anaerobic sludge	Continuous	1.97 mol/mol of hexose	[106]
	Mixed culture	Batch	93.4 L/kg of COD	[101]
Corn starch	Anaerobic sludge	Batch	177 L/kg of starch	[110]
Grape	Mixed culture	Continuous	384 L/kg of VS	[103]
Date fruit waste	<i>Enterobacter aerogenes</i> ATCC 13,408	Batch	144–239 L/kg of feed	[102]
Food waste	Cattle dung	Batch	220 L/kg of feed	[112]
Melon	Mixed culture	Continuous	352 L/kg of VS	[103]
Mixed fruit wastes	Mixed culture	Continuous	553 L/kg of VS	
Onion-peels	DCMs	Batch	56.0–86.0 L/kg of TS	[5, 59]
Orange	Mixed culture	Continuous	403 L/kg of VS	[103]
The organic fraction of municipal solid waste	Anaerobic digestion sludge	Batch	62.5 L/kg of VS	[111]
Pea-shells	DMCs	Batch	65.0 L/kg of TS	[20]
Pea-shells hydrolysate	DMCs	Batch	75.0 L/kg of TS	[21]
	DMCs	Continuous	54.0 L/kg of TS	
Potato peels	<i>Parageobacillus thermoglucosidasius</i> KCTC 33,548	Batch	0.83 L/L	[109]
	DCMs	Batch	64.0–92.0 L/kg of TS	[5, 59]
Potato starch	Anaerobic sludge	Batch	151 L/kg of starch	[110]
Rice husk	<i>Bacillus cereus</i>		1.37 mol/mol of hexose	[107]
Sugar beet molasses	<i>Caldicellulosiruptor saccharolyticus</i> DSM 8903	Batch	2.10 mol/mol of hexose	[104]
Sugarcane bagasse	<i>Clostridium thermocellum</i> DSMZ 1313	Batch	0.92 mol/mol of hexose	[108]
Sweet potato starch	Anaerobic sludge	Batch	199 L/kg of starch	[110]
Vegetable and fruits	Mixed culture	Batch	8.3 L/kg of COD	[101]
Vegetable, fruit, and cheese whey	Mixed culture	Batch	12.5 L/kg of COD	

recorded maximum productions of 253 L of H<sub>2</sub>/kg of TVS and 730 L/kg of VS at first and second stages of integrative bioprocess, respectively [120, 130]. Thus, these wastes combinations can be more beneficial to produced higher H<sub>2</sub> and CH<sub>4</sub> in the future. Additionally, the lower H<sub>2</sub> generation at the first-stage of the integrative bioprocess can be improved via the uses of DMCs, novel culture or genetically engineered culture over UMCs [10, 132, 133].

## Conclusions and Prospects

The key challenges of H<sub>2</sub> production are associated with the costly sugars as primary feed and lower H<sub>2</sub> yield to 4 mol/mol of hexose under DF conditions. Biowastes, including agri-food wastes, are desirable alternative low-cost feed to produce biohydrogen. However, the available quantum of these wastes is highly variable, especially in



**Table 4** Two-stage bioprocesses for hydrogen and methane production from various agri-food wastes

Agri-waste	Stage I—Biohydrogen		Stage II—Biomethane		Reference
	Culture	Yield	Culture	Yield	
Banana peels	Anaerobic sludge	210 L/kg of VS	Anaerobic sludge	284 L/kg of VS	[117]
Bean waste	Seed sludge	152 L/kg of TVS	Seed sludge	463 L/kg of TVS	[120]
Cassava residues	Mixed culture	118 L/kg of TS	Mixed culture	308 L/kg of TS	[119]
Cheese whey	Anaerobic sludge	137 L/kg of COD	Anaerobic sludge	250 L/kg of COD	[116]
Common wheat	Granular sludge	47.0 L/kg of COD	Granular sludge	202 L/kg of COD	[114]
Durum wheat	Granular sludge	76.0 L/kg of COD	Granular sludge	243 L/kg of COD	
Food waste	Anaerobic sludge	215 L/kg of COD	Anaerobic sludge	311 L/kg of COD	[125]
		218 L/kg of VS	Anaerobic sludge	432 L/kg of VS	[127]
Food and olive husk	Seed sludge	135 L/kg of VS	Seed sludge	510 L/kg of VS	[126]
	Anaerobic sludge	87.0 NL/kg of VS	Anaerobic sludge	505 NL/kg of VS	[128]
Food waste and activated sludge	Seed sludge	76.8 L/kg of VS	Seed sludge	148 L/kg of VS	[130]
Garden and food	<i>C. saccharolyticus</i> DSM 8903	46.0 L/kg of TS	Anaerobic sludge	682 L/kg of TS	[129]
Mashed potato	Granular sludge	177 L/kg of COD	Granular sludge	207 L/kg of COD	[114]
Organic fraction of municipal solid waste	Anaerobic sludge	24.0 L/kg of VS	Anaerobic sludge	570 L/kg of VS	[122]
		87.5 L/L	Mixed culture	241 L/L	[123]
		41.7 L/kg of VS	Anaerobic sludge	300 L/kg of VS	[121]
Potato	Seed sludge	253 L/kg of TVS	Seed sludge	507 L/kg of TVS	[120]
Potato peels	Anaerobic sludge	103 L/kg of VS	Anaerobic sludge	237 L/kg of VS	[115]
Rice	Anaerobic sludge	125 L/kg of VS	Anaerobic sludge	232 L/kg of VS	
Rice residue and <i>Chlorella pyrenoidosa</i>	Anaerobic sludge	223 L/kg of VS	Anaerobic sludge	277 L/kg of VS	[124]
Steam potato peeling	Granular sludge	134 L/kg of COD	Granular sludge	183 L/kg of COD	[114]
Sugarcane bagasse	Cow dung	93.4 L/kg of VS	Anaerobic sludge	222 L/kg of VS	[118]
Vegetables and other wastes mixture	Seed sludge	79.4 L/kg of VS	Seed sludge	730 L/kg of VS	[131]

cases of seasonal waste that can be a limiting factor for sustainable H<sub>2</sub> production via environmentally friendly technologies. Also, feedstocks (biowastes) mobilization is a vital concern to produce from biomass. Due to the complex nature of biowastes and variations in their cellulosic contents can also influence biohydrogen production. Thus, the utilization of biowastes (type) can impact overall prospects of their use such as the production of H<sub>2</sub> through DF and CH<sub>4</sub> via AD. These obstacles may be undertaken

through the development of cost-effective biowastes pre-treatment techniques via focusing on the improvement of bioprocess efficiency by the valorization of waste to increase H<sub>2</sub> yield. The bioprocess-based technologies to produce H<sub>2</sub> are in different levels of developmental stages. In typical, various studies have been focused on H<sub>2</sub> production bioprocess through—(1) reduce capital investments, different operational expenses, including maintenance), revenue (profits either directly or indirectly)

and cost of the product ( $H_2$ ), and (2) improvement of technical efficiency of  $H_2$  production such as (1) use of inexpensive-pretreatment methods for hydrolysis of bio-waste to fermentable sugars, (2) screening efficient and novel  $H_2$ -producers, (3) use of metal and nanoparticles to influence biocatalytic activity especially hydrogenases, (4) co-digestion of biowastes to improve nutrition balance as suitable feed, (5) use of selective consortia of DMCs instead of pure or UMCs (it requires additional pretreatment to the elimination of  $CH_4$ -producers and enrichment of  $H_2$ -producers) to improve metabolization of feed towards  $H_2$ , (6) selection of desirable reactor type and (7) metabolic engineering of biocatalysts. In the current scenario, still, the  $H_2$  production cost is substantially high in addition to the uses of biowaste as low-cost feed due to higher capital and operation costs. The integration of pure or MCs-based bioprocesses from agri-food wastes can be more economically desirable to produce  $H_2$  followed by value-added products at the second stage such as (1)  $H_2$  via photo-fermentative, (2)  $CH_4$  through AD, (3) PHAs, or (4) electricity production.

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#### Declarations

**Conflicts of interest** The authors declare no conflict of interest.

#### References

- Kalia VC, Purohit HJ (2008) Microbial diversity and genomics in aid of bioenergy. *J Ind Microbiol Biotechnol* 35:403–419. <https://doi.org/10.1007/s10295-007-0300-y>
- Patel SKS (2010) Studies on biodiversity of hydrogen producers and enhancement of dark fermentative hydrogen production process. Ph.D. Thesis, University of Pune, India. <http://hdl.handle.net/10603/3269>
- Kumar P, Patel SKS, Lee J-K et al (2013) Extending the limits of *Bacillus* for novel biotechnological applications. *Biotechnol Adv* 31:1543–1561. <https://doi.org/10.1016/j.biotechadv.2013.08.007>
- Prakash J, Sharma R, Patel SKS et al (2018) Biohydrogen production by co-digestion of domestic wastewater and biodiesel industry effluent. *PLoS ONE* 13:e0199059. <https://doi.org/10.1371/journal.pone.0199059>
- Patel SKS, Ray S, Prakash J et al (2019) Co-digestion of biowastes to enhance biological hydrogen process by defined mixed bacterial cultures. *Indian J Microbiol* 59:154–160. <https://doi.org/10.1007/s12088-018-00777-8>
- Kumar P, Sharma R, Ray S et al (2015) Dark fermentative bioconversion of glycerol to hydrogen by *Bacillus thuringiensis*. *Bioresour Technol* 182:383–388. <https://doi.org/10.1016/j.biortech.2015.01.138>
- Kondaveeti S, Kim IW, Otari S et al (2019) Co-generation of hydrogen and electricity from biodiesel process effluents. *Int J Hydrogen Energy* 44:27285–27296. <https://doi.org/10.1016/j.ijhydene.2019.08.258>
- Patel SKS, Gupta RK, Das D et al (2021) Continuous biohydrogen production from poplar biomass hydrolysate by a defined bacterial mixture immobilized on lignocellulosic materials under non-sterile conditions. *J Clean Prod* 287:125037. <https://doi.org/10.1016/j.jclepro.2020.125037>
- Patel SKS, Gupta RK, Kondaveeti S et al (2020) Conversion of biogas to methanol by methanotrophs immobilized on chemically modified chitosan. *Bioresour Technol* 315:123791. <https://doi.org/10.1016/j.biortech.2020.123791>
- Patel SKS, Das D, Kim SC et al (2021) Integrating strategies for sustainable conversion of waste biomass into dark-fermentative hydrogen and value-added products. *Renew Sust Energ Rev* 150:111491. <https://doi.org/10.1016/j.rser.2021.111491>
- Kumar V, Patel SKS, Gupta RK et al (2019) Enhanced saccharification and fermentation of rice straw by reducing the concentration of phenolic compounds using an immobilization enzyme cocktail. *Biotechnol J* 14:1800468. <https://doi.org/10.1002/biot.201800468>
- Mardina P, Li J, Patel SKS et al (2016) Potential of immobilized whole-cell *Methylocella tundræ* as biocatalyst for methanol production from methane. *J Microbiol Biotechnol* 26:1234–1241. <https://doi.org/10.4014/jmb.1602.02074>
- Patel SKS, Mardina P, Kim SY et al (2016) Biological methanol production by a type II methanotroph *Methylocystis bryophila*. *J Microbiol Biotechnol* 26:717–724. <https://doi.org/10.4014/jmb.1601.01013>
- Patel SKS, Selvaraj C, Mardina P et al (2016) Enhancement of methanol production from synthetic gas mixture by *Methylosinus sporium* through covalent immobilization. *Appl Energy* 171:383–391. <https://doi.org/10.1016/j.apenergy.2016.03.022>
- Patel SKS, Kondaveeti S, Otari SV et al (2018) Repeated batch methanol production from a simulated biogas mixture using immobilized *Methylocystis bryophila*. *Energy* 145:477–485. <https://doi.org/10.1016/j.energy.2017.12.142>
- Kumar A, Park GD, Patel SKS et al (2019)  $SiO_2$  microparticles with carbon nanotube-derived mesopores as an efficient support for enzyme immobilization. *Chem Eng J* 359:1252–1264. <https://doi.org/10.1016/j.cej.2018.11.052>
- Otari SV, Patel SKS, Kalia VC et al (2020) One-step hydrothermal synthesis of magnetic rice straw for effective lipase immobilization and its application in esterification reaction. *Bioresour Technol* 302:122887. <https://doi.org/10.1016/j.biortech.2020.122887>
- Kalia VC, Kumar A, Jain SR et al (1992) Biomethanation of plant materials. *Bioresour Technol* 41:209–212. [https://doi.org/10.1016/0960-8524\(92\)90003-G](https://doi.org/10.1016/0960-8524(92)90003-G)
- Patel SKS, Kumar P, Kalia VC (2012) Enhancing biological hydrogen production through complementary microbial metabolisms. *Int J Hydrogen Energy* 37:10590–10603. <https://doi.org/10.1016/j.ijhydene.2012.04.045>
- Patel SKS, Singh M, Kumar P et al (2012) Exploitation of defined bacterial cultures for production of hydrogen and polyhydroxybutyrate from pea-shells. *Biomass Bioenergy* 36:218–225. <https://doi.org/10.1016/j.biombioe.2011.10.027>
- Patel SKS, Kumar P, Singh S et al (2015) Integrative approach to produce hydrogen and polyhydroxybutyrate from biowaste using defined bacterial cultures. *Bioresour Technol* 176:136–141. <https://doi.org/10.1016/j.biortech.2014.11.029>
- Patel SKS, Kumar P, Singh M et al (2015) Integrative approach for biohydrogen and polyhydroxyalkanoates production. In: Kalia VC (ed) *Microbial factories, waste treatment*. Volume 1. Springer, New Delhi, pp. 73–85. [https://doi.org/10.1007/978-81-322-2598-0\\_5](https://doi.org/10.1007/978-81-322-2598-0_5)

23. Patel SKS, Jeong J-H, Mehariya S et al (2016) Production of methanol from methane by encapsulated *Methylosinus sporium*. *J Microbiol Biotechnol* 26:2098–2105. <https://doi.org/10.4014/jmb.1608.0805>
24. Patel SKS, Mardina P, Kim D et al (2016) Improvement in methanol production by regulating the composition of synthetic gas mixture and raw biogas. *Bioresour Technol* 218:202–208. <https://doi.org/10.1016/j.biortech.2016.06.065>
25. Lee J-K, Patel SKS, Sung BH et al (2020) Biomolecules from municipal and food industry wastes: an overview. *Bioresour Technol* 298:122346. <https://doi.org/10.1016/j.biortech.2020.122346>
26. Singh M, Kumar P, Patel SKS et al (2013) Production of polyhydroxyalkanoate co-polymer by *Bacillus thuringiensis*. *Indian J Microbiol* 53:77–83. <https://doi.org/10.1007/s12088-012-0294-7>
27. Patel SKS, Singh M, Kalia VC (2011) Hydrogen and polyhydroxybutyrate producing abilities of *Bacillus* spp. from glucose in two stage system. *Indian J Microbiol* 51:418–423. <https://doi.org/10.1007/s12088-011-0236-9>
28. Kumar P, Singh M, Mehariya S et al (2014) Ecobiotechnological approach for exploiting the abilities of *Bacillus* to produce co-polymer of polyhydroxyalkanoate. *Indian J Microbiol* 54:151–157. <https://doi.org/10.1007/s12088-014-0457-9>
29. Kumar P, Ray S, Patel SKS et al (2015) Bioconversion of crude glycerol to polyhydroxyalkanoate by *Bacillus thuringiensis* under non-limiting nitrogen conditions. *Int J Biol Macromol* 78:9–16. <https://doi.org/10.1016/j.ijbiomac.2015.03.046>
30. Kalia VC, Patel SKS, Shanmugam R et al (2021) Polyhydroxyalkanoates: trends and advances towards biotechnological applications. *Bioresour Technol* 326:124737. <https://doi.org/10.1016/j.biortech.2021.124737>
31. Kondaveeti S, Pagolu R, Patel SKS et al (2019) Bioelectrochemical detoxification of phenolic compounds during enzymatic pre-treatment of rice straw. *J Microbiol Biotechnol* 29:1760–1768. <https://doi.org/10.4014/jmb.1909.09042>
32. Kondaveeti S, Patel SKS, Pagolu R et al (2019) Conversion of simulated biogas to electricity: sequential operation of methanotrophic reactor effluents in microbial fuel cell. *Energy* 189:116309. <https://doi.org/10.1016/j.energy.2019.116309>
33. Panday D, Patel SKS, Singh R et al (2019) Solvent-tolerant acyltransferase from *Bacillus* sp. APB-6: purification and characterization. *Indian J Microbiol* 59:500–507. <https://doi.org/10.1007/s12088-019-00836-8>
34. Kala A, Kamra DN, Agarwal N et al (2020) Insights into metatranscriptome, and CAZymes of buffalo rumen supplemented with blend of essential oils. *Indian J Microbiol* 60:485–493. <https://doi.org/10.1007/s12088-020-00894-3>
35. Kondaveeti S, Patel SKS, Woo J et al (2020) Characterization of cellobiohydrolases from *Schizophyllum commune* KMJ820. *Indian J Microbiol* 60:160–166. <https://doi.org/10.1007/s12088-019-00843-9>
36. Devi N, Patel SKS, Kumar P et al (2021) Bioprocess scale-up for acetohydroxamic acid production by hyperactive acyltransferase of immobilized *Rhodococcus pyridinivorans*. *Catal Lett*. <https://doi.org/10.1007/s10562-021-03696-4>
37. Goderska K (2021) Biosynthesis of lactobionic acid in whey-containing medium by microencapsulated and free bacteria of *Pseudomonas taetrolensis*. *Indian J Microbiol* 61:315–323. <https://doi.org/10.1007/s12088-021-00944-4>
38. Muneeswaran G, Patel SKS, Kondaveeti S et al (2021) Biotin and Zn<sup>2+</sup> increase xylitol production by *Candida tropicalis*. *Indian J Microbiol* 61:331–337. <https://doi.org/10.1007/s12088-021-00960-4>
39. Kim TS, Patel SKS, Selvaraj C et al (2016) A highly efficient sorbitol dehydrogenase from *Gluconobacter oxydans* G624 and improvement of its stability through immobilization. *Sci Rep* 6:33438. <https://doi.org/10.1038/srep33438>
40. Ramachandran P, Jagtap SS, Patel SKS et al (2016) Role of the non-conserved amino acid Asparagine 285 in the glycone-binding pocket of *Neosartorya fischeri* β-glucosidase. *RSC Adv* 6:48137–48144. <https://doi.org/10.1039/c5ra28017f>
41. Selvaraj C, Krishnasamy G, Jagtap SS et al (2016) Structural insights into the binding mode of D-sorbitol with sorbitol dehydrogenase using QM-polarized ligand docking and molecular dynamics simulations. *Biochem Eng J* 114:244–256. <https://doi.org/10.1016/j.bej.2016.07.008>
42. Gao H, Li J, Sivakumar D et al (2019) NADH oxidase from *Lactobacillus reuteri*: A versatile enzyme for oxidized cofactor regeneration. *Int J Biol Macromol* 123:629–636. <https://doi.org/10.1016/j.ijbiomac.2018.11.096>
43. Kim J-S, Patel SKS, Tiwari MK et al (2020) Phe-140 determines the catalytic efficiency of arylacetamidase from *Alcaligenes faecalis*. *Int J Mol Sci* 21:7859. <https://doi.org/10.3390/ijms21217859>
44. Pagolu R, Singh R, Shanmugam R et al (2021) Site-directed lysine modification of xylanase for oriented immobilization onto silicon dioxide nanoparticles. *Bioresour Technol* 331:125063. <https://doi.org/10.1016/j.biortech.2021.125063>
45. Kumar P, Koul S, Patel SKS et al (2015) Heterologous expression of quorum sensing inhibitory genes in diverse organisms. In: Kalia VC (ed) *Quorum sensing vs quorum quenching: a battle with no end in sight*, Springer, pp. 343–356. [https://doi.org/10.1007/978-81-322-1982-8\\_28](https://doi.org/10.1007/978-81-322-1982-8_28)
46. Otari SV, Patel SKS, Jeong JH et al (2016) A green chemistry approach for synthesizing thermostable antimicrobial peptide-coated gold nanoparticles immobilized in an alginate biohydrogel. *RSC Adv* 6:86808–86816. <https://doi.org/10.1039/c6ra1488k>
47. Otari SV, Kumar M, Anwar MZ et al (2017) Rapid synthesis and decoration of reduced graphene oxide with gold nanoparticles by thermostable peptides for memory device and photothermal applications. *Sci Rep* 7:10980. <https://doi.org/10.1038/s41598-017-10777-1>
48. Otari SV, Pawar SH, Patel SKS et al (2017) *Canna edulis* leaf extract-mediated preparation of stabilized silver nanoparticles: Characterization, antimicrobial activity, and toxicity studies. *J Microbiol Biotechnol* 27:731–738. <https://doi.org/10.4014/jmb.1610.10019>
49. Kalia VC, Patel SKS, Kang YC et al (2019) Quorum sensing inhibitors as antipathogens: biotechnological applications. *Biotechnol Adv* 37:68–90. <https://doi.org/10.1016/j.biotechadv.2018.11.006>
50. Otari SV, Patel SKS, Kalia VC et al (2019) Antimicrobial activity of biosynthesized silver nanoparticles decorated silica nanoparticles. *Indian J Microbiol* 59:379–382. <https://doi.org/10.1007/s12088-019-00812-2>
51. Kalia VC, Gong C, Patel SKS et al (2021) Regulation of plant mineral nutrition by signal molecules. *Microorganisms* 9:774. <https://doi.org/10.3390/microorganisms9040774>
52. Kalia VC, Patel SKS, Cho B-K et al (2021) Emerging applications of bacteria as anti-tumor agents. *Sem Cancer Biol*. <https://doi.org/10.1016/j.semcancer.2021.05.012>
53. Parasuraman P, Devadatha B, Sarma VV et al (2020) Inhibition of microbial quorum sensing mediated virulence factors by *Pestalotiopsis sydowiana*. *J Microbiol Biotechnol* 30:571–582. <https://doi.org/10.4014/jmb.1907.07030>
54. Patel SKS, Lee J-K, Kalia VC (2020) Deploying biomolecules as anti-COVID-19 agents. *Indian J Microbiol* 60:263–268. <https://doi.org/10.1007/s12088-020-00893-4>

55. Prakash O, Nimonkar Y, Desai D (2020) A recent overview of microbes and microbiome preservation. *Indian J Microbiol* 60:297–309. <https://doi.org/10.1007/s12088-020-00880-9>
56. Rishi P, Thakur K, Vij S et al (2020) Diet, gut microbiota and COVID-19. *Indian J Microbiol* 60:420–429. <https://doi.org/10.1007/s12088-020-00908-0>
57. Kalia VC, Joshi AP (1995) Conversion of waste biomass (pea-shells) into hydrogen and methane through anaerobic digestion. *Bioresour Technol* 53:165–168. [https://doi.org/10.1016/0960-8524\(95\)00077-R](https://doi.org/10.1016/0960-8524(95)00077-R)
58. Patel SKS, Kumar P, Mehariya S et al (2014) Enhancement in hydrogen production by co-cultures of *Bacillus* and *Enterobacter*. *Int J Hydrogen Energy* 39:14663–14668. <https://doi.org/10.1016/j.ijhydene.2014.07.084>
59. Patel SKS, Lee JK, Kalia VC (2017) Dark-fermentative biological hydrogen production from mixed biowastes using defined mixed cultures. *Indian J Microbiol* 57:171–176. <https://doi.org/10.1007/s12088-017-0643-7>
60. Patel SKS, Singh R, Kumar A et al (2017) Biological methanol production by immobilized *Methylocella tundrae* using simulated biohythane as a feed. *Bioresour Technol* 241:922–927. <https://doi.org/10.1016/j.biortech.2017.05.160>
61. Arora K, Kaur P, Kumar P et al (2021) Valorization of wastewater resources into biofuels and value-added products using microalgal system. *Front Energy Res* 9:646571. <https://doi.org/10.3389/fenrg.2021.646571>
62. Patel SKS, Lee J-K, Kalia VC (2016) Integrative approach for producing hydrogen and polyhydroxyalkanoate from mixed wastes of biological origin. *Indian J Microbiol* 56:293–300. <https://doi.org/10.1007/s12088-016-0595-3>
63. Patel SKS, Lee JK, Kalia VC (2018) Beyond the theoretical yields of dark-fermentative biohydrogen. *Indian J Microbiol* 58:529–530. <https://doi.org/10.1007/s12088-018-0759-4>
64. Patel SKS, Lee JK, Kalia VC (2018) Nanoparticles in biological hydrogen production: an overview. *Indian J Microbiol* 58:8–18. <https://doi.org/10.1007/s12088-017-0678-9>
65. Porwal S, Kumar T, Lal S et al (2008) Hydrogen and polyhydroxybutyrate producing abilities of microbes from diverse habitats by dark fermentative process. *Bioresour Technol* 99:5444–5451. <https://doi.org/10.1016/j.biortech.2007.11.011>
66. Patel SKS, Kalia VC (2013) Integrative biological hydrogen production: an overview. *Indian J Microbiol* 53:3–10. <https://doi.org/10.1007/s12088-012-0287-6>
67. Patel SKS, Jeon MS, Gupta RK et al (2019) Hierarchical macroporous particles for efficient whole-cell immobilization: application in bioconversion of greenhouse gases to methanol. *ACS Appl Mater Interfaces* 11:18968–18977. <https://doi.org/10.1021/acsami.9b03420>
68. Patel SKS, Gupta RK, Kumar V et al (2020) Biomethanol production from methane by immobilized cocultures of methanotrophs. *Indian J Microbiol* 60:318–324. <https://doi.org/10.1007/s12088-020-00883-6>
69. Patel SKS, Kalia VC, Joo JB et al (2020) Biotransformation of methane into methanol by methanotrophs immobilized on coconut coir. *Bioresour Technol* 297:122433. <https://doi.org/10.1016/j.biortech.2019.122433>
70. Singh M, Patel SKS, Kalia VC (2009) *Bacillus subtilis* as potential producer for polyhydroxyalkanoates. *Microb Cell Fact* 8:38. <https://doi.org/10.1186/1475-2859-8-38>
71. Purohit HJ (2019) Aligning microbial biodiversity for valorization of biowastes: conception to perception. *Indian J Microbiol* 59:391–400. <https://doi.org/10.1007/s12088-019-00826-w>
72. Imam A, Kanaujia PK, Ray A et al (2021) Removal of petroleum contaminants through bioremediation with integrated concepts of resource recovery: a review. *Indian J Microbiol* 61:250–261. <https://doi.org/10.1007/s12088-021-00928-4>
73. Patel SKS, Gupta RK, Kalia VC et al (2021) Integrating anaerobic digestion of potato peels to methanol production by methanotrophs immobilized on banana leaves. *Bioresour Technol* 323:124550. <https://doi.org/10.1016/j.biortech.2020.124550>
74. Patel SKS, Kumar V, Mardina P et al (2018) Methanol production from simulated biogas mixtures by co-immobilized *Methylococcus methanica* and *Methylocella tundrae*. *Bioresour Technol* 263:25–32. <https://doi.org/10.1016/j.biortech.2018.04.096>
75. Patel SKS, Shanmugam R, Kalia VC et al (2020) Methanol production by polymer-encapsulated methanotrophs from simulated biogas in the presence of methane vector. *Bioresour Technol* 304:123022. <https://doi.org/10.1016/j.biortech.2020.123022>
76. Kumari D, Singh R (2018) Pretreatment of lignocellulosic wastes for biofuel production: a critical review. *Renew Sustain Energy Rev* 90:877–891. <https://doi.org/10.1016/j.rser.2018.03.111>
77. Patel SKS, Purohit HJ, Kalia VC (2010) Dark fermentative hydrogen production by defined mixed microbial cultures immobilized on ligno-cellulosic waste materials. *Int J Hydrogen Energy* 35:10674–10681. <https://doi.org/10.1016/j.ijhydene.2010.03.025>
78. Patel SKS, Choi SH, Kang YC et al (2016) Large-scale aerosol-assisted synthesis of biofriendly Fe<sub>2</sub>O<sub>3</sub> yolk-shell particles: a promising support for enzyme immobilization. *Nanoscale* 8:6728–6738. <https://doi.org/10.1039/C6NR00346J>
79. Patel SKS, Choi SH, Kang YC et al (2017) Eco-friendly composite of Fe<sub>3</sub>O<sub>4</sub>-reduced graphene oxide particles for efficient enzyme immobilization. *ACS Appl Mater Interfaces* 9:2213–2222. <https://doi.org/10.1021/acsami.6b05165>
80. Kumar A, Patel SKS, Madan B et al (2018) Immobilization of xylanase using a protein-inorganic hybrid system. *J Microbiol Biotechnol* 28:638–644. <https://doi.org/10.4014/jmb.1710.10037>
81. Patel SKS, Gupta RK, Kumar V et al (2019) Influence of metal ions on the immobilization of β-glucosidase through protein-inorganic hybrids. *Indian J Microbiol* 59:370–374. <https://doi.org/10.1007/s12088-019-0796-z>
82. Bhatia SK, Wadhwa P, Bhatia RK, et al. (2019) Strategy for biosynthesis of polyhydroxyalkanoates polymers/copolymers and their application in drug delivery. In: Kalia VC (ed) *Biotechnological applications of polyhydroxyalkanoates*. Springer, Singapore, pp. 13–34. [https://doi.org/10.1007/978-981-13-3759-8\\_2](https://doi.org/10.1007/978-981-13-3759-8_2)
83. Kalia VC, Ray S, Patel SKS, et al. (2019) The dawn of novel biotechnological applications of polyhydroxyalkanoates. In: Kalia VC (ed) *Biotechnological applications of polyhydroxyalkanoates*. Springer, Singapore, pp. 1–11. [https://doi.org/10.1007/978-981-13-3759-8\\_1](https://doi.org/10.1007/978-981-13-3759-8_1)
84. Kalia VC, Ray S, Patel SKS, et al. (2019) Applications of polyhydroxyalkanoates and their metabolites as drug carriers. In: Kalia VC (ed) *Biotechnological applications of polyhydroxyalkanoates*. Springer, Singapore, pp. 35–48. [https://doi.org/10.1007/978-981-13-3759-8\\_3](https://doi.org/10.1007/978-981-13-3759-8_3)
85. Patel SKS, Sandeep K, Singh M, et al. (2019) Biotechnological application of polyhydroxyalkanoates and Their Composites as anti-microbial agents. In: Kalia VC (ed) *Biotechnological applications of polyhydroxyalkanoates*. Springer, Singapore, pp. 207–225. [https://doi.org/10.1007/978-981-13-3759-8\\_8](https://doi.org/10.1007/978-981-13-3759-8_8)
86. Ray S, Patel SKS, Singh M, et al. (2019) Exploiting polyhydroxyalkanoates for tissue engineering. In: Kalia VC (ed) *Biotechnological applications of polyhydroxyalkanoates*.

- Springer, Singapore, pp. 271–282. [https://doi.org/10.1007/978-981-13-3759-8\\_10](https://doi.org/10.1007/978-981-13-3759-8_10)
87. Azwar MY, Hussain MA, Abdul-Wahab AK (2014) Development of biohydrogen production by photobiological, fermentation and electrochemical processes: a review. *Renew Sustain Energy Rev* 31:158–171. <https://doi.org/10.1016/j.rser.2013.11.022>
  88. Penfold DW, Forster CF, Macaskie LE (2003) Increased hydrogen production by *Escherichia coli* strain HD701 in comparison with the wild-type parent strain MC4100. *Enzym Microb Technol* 33:185–189. [https://doi.org/10.1016/S0141-0229\(03\)00115-7](https://doi.org/10.1016/S0141-0229(03)00115-7)
  89. Patel SKS, Kalia VC, Choi JH et al (2014) Immobilization of laccase on SiO<sub>2</sub> nanocarriers improves its stability and reusability. *J Microbiol Biotechnol* 24:639–647. <https://doi.org/10.4014/jmb.1401.01025>
  90. Anwar MZ, Kim DJ, Kumar A et al (2017) SnO<sub>2</sub> hollow nanotubes: a novel and efficient support matrix for enzyme immobilization. *Sci Rep* 7:15333. <https://doi.org/10.1038/s41598-017-15550-y>
  91. Patel SKS, Otari SV, Kang YC et al (2017) Protein-inorganic hybrid system for efficient his-tagged enzymes immobilization and its application in L-xylulose production. *RSC Adv* 7:3488–3494. <https://doi.org/10.1039/c6ra24404a>
  92. Kumar A, Kim I-W, Patel SKS et al (2018) Synthesis of protein-inorganic nanohybrids with improved catalytic properties using Co<sub>3</sub>(PO<sub>4</sub>)<sub>2</sub>. *Indian J Microbiol* 58:100–104. <https://doi.org/10.1007/s12088-017-0700-2>
  93. Patel SKS, Otari SV, Li J et al (2018) Synthesis of cross-linked protein-metal hybrid nanoflowers and its application in repeated batch decolorization of synthetic dyes. *J Hazard Mater* 347:442–450. <https://doi.org/10.1016/j.jhazmat.2018.01.003>
  94. Patel SKS, Anwar MZ, Kumar A et al (2018) Fe<sub>2</sub>O<sub>3</sub> yolk-shell particles-based laccase biosensor for efficient detection of 2,6-dimethoxyphenol. *Biochem Eng J* 132:1–8. <https://doi.org/10.1016/j.bej.2017.12.013>
  95. Otari SV, Patel SKS, Kim S-Y et al (2019) Copper ferrite magnetic nanoparticles for the immobilization of enzyme. *Indian J Microbiol* 59:105–108. <https://doi.org/10.1007/s12088-018-0768-3>
  96. Patel SKS, Choi H, Lee J-K (2019) Multi-metal based inorganic–protein hybrid system for enzyme immobilization. *ACS Sustainable Chem Eng* 7:13633–13638. <https://doi.org/10.1021/acssuschemeng.9b02583>
  97. Patel SKS, Gupta RK, Kim S-Y et al (2021) *Rhus vernicifera* laccase immobilization on magnetic nanoparticles to improve stability and its potential application in bisphenol A degradation. *Indian J Microbiol* 61:45–54. <https://doi.org/10.1007/s12088-020-00912-4>
  98. Patel SKS, Kim JH, Kalia VC et al (2019) Antimicrobial activity of amino-derivatized cationic polysaccharides. *Indian J Microbiol* 59:96–99. <https://doi.org/10.1007/s12088-018-0764-7>
  99. Otari SV, Shinde VV, Hui G et al (2019) Biomolecule-entrapped SiO<sub>2</sub> nanoparticles for ultrafast green synthesis of silver nanoparticle-decorated hybrid nanostructures as effective catalysts. *Ceram Int* 45:5876–5882. <https://doi.org/10.1016/j.ceramint.2018.12.054>
  100. Kee SH, Chiongson JBV, Saludes JP et al (2021) Bioconversion of agro-industry sourced biowaste into biomaterials via microbial factories - A viable domain of circular economy. *Environ Pollut* 271:116311. <https://doi.org/10.1016/j.envpol.2020.116311>
  101. Niño-Navarro C, Chairez I, Christen P et al (2020) Enhanced hydrogen production by a sequential dark and photo fermentation process: effects of initial feedstock composition, dilution and microbial population. *Renew Energy* 147:924–936. <https://doi.org/10.1016/j.renene.2019.09.024>
  102. Rambabu K, Bharath G, Banat F et al (2021) Ferric oxide/date seed activated carbon nanocomposites mediated dark fermentation of date fruit wastes for enriched biohydrogen production. *Int J Hydrogen Energy* 46:16631–16643. <https://doi.org/10.1016/j.ijhydene.2020.06.108>
  103. Akinbomi J, Taherzadeh MJ (2015) Evaluation of fermentative hydrogen production from single and mixed fruit wastes. *Energies* 8:4253–4272. <https://doi.org/10.3390/en8054253>
  104. Ozgur E, Mars AE, Peksel B et al (2010) Biohydrogen production from beet molasses by sequential dark and photofermentation. *Int J Hydrogen Energy* 35:511–517. <https://doi.org/10.1016/j.ijhydene.2009.10.094>
  105. Contreras-Dávila CA, Méndez-Acosta HO, Méndez-Acosta L et al (2017) Continuous hydrogen production from enzymatic hydrolysate of Agave tequilana bagasse: effect of the organic loading rate and reactor configuration. *Chem Eng J* 313:671–679. <https://doi.org/10.1016/j.cej.2016.12.084>
  106. Colombo B, Calvo MV, Sciarria TP et al (2019) Biohydrogen and polyhydroxyalkanoates (PHA) as products of a two-steps bioprocess from deproteinized dairy wastes. *Waste Manag* 95:22–31. <https://doi.org/10.1016/j.wasman.2019.05.052>
  107. Dinesh GH, Nguyen DD, Ravindran B et al (2020) Simultaneous biohydrogen (H<sub>2</sub>) and bioplastic (poly-β-hydroxybutyrate-PHB) productions under dark, photo, and subsequent dark and photo fermentation utilizing various wastes. *Int J Hydrogen Energy* 45:5840–5853. <https://doi.org/10.1016/j.ijhydene.2019.09.036>
  108. Ahmad Q-A, Manzoor M, Chaudhary A et al (2021) Bench-scale fermentation for second generation ethanol and hydrogen production by *Clostridium thermocellum* DSMZ 1313 from sugarcane bagasse. *Environ Prog Sustainable Energy* 40:e13516. <https://doi.org/10.1002/ep.13516>
  109. Singhvi M, Maharjan A, Thapa A et al (2021) Nanoparticle-associated single step hydrogen fermentation for the conversion of starch potato waste biomass by thermophilic *Parageobacillus thermoglucosidasius*. *Bioresour Technol* 337:125490. <https://doi.org/10.1016/j.biortech.2021.125490>
  110. Ren H-Y, Liu B-F, Kong F et al (2015) Sequential generation of hydrogen and lipids from starch by combination of dark fermentation and microalgal cultivation. *RSC Adv* 5:76779–76782. <https://doi.org/10.1039/C5RA15023J>
  111. Alavi-Borazjani SA, da Cruz Tarelho LA, Capela MI (2021) Parametric optimization of the dark fermentation process for enhanced biohydrogen production from the organic fraction of municipal solid waste using Taguchi method. *Int J Hydrogen Energy* 46:21372–21382. <https://doi.org/10.1016/j.ijhydene.2021.04.017>
  112. Zong W, Yu R, Zhang P et al (2009) Efficient hydrogen gas production from cassava and food waste by a two-step process of dark fermentation and photo-fermentation. *Biomass Bioenergy* 33:1458–1463. <https://doi.org/10.1016/j.biombioe.2009.06.008>
  113. Pason P, Tachaapaikoon C, Panichnumsin P et al (2020) One -step biohydrogen production from cassava pulp using novel enrichment of anaerobic thermophilic bacteria community. *Biocatal Agri Biotechnol* 27:101658. <https://doi.org/10.1016/j.bcab.2020.101658>
  114. Giordano A, Cantù C, Spagni A (2011) Monitoring the biochemical hydrogen and methane potential of the two-stage dark fermentative process. *Bioresour Technol* 102:4474–4479. <https://doi.org/10.1016/j.biortech.2010.12.106>
  115. Dong L, Zhenhong Y, Yongming S et al (2011) Anaerobic fermentative co-production of hydrogen and methane from an organic fraction of municipal solid waste. *Energy Sources A* 33:575–585. <https://doi.org/10.1080/15567030903117653>

116. Cota-Navarro CB, Carillo-Reyes J, Davila-Vazquez G et al (2011) Continuous hydrogen and methane production in a two-stage cheese whey fermentation system. *Water Sci Technol* 64:367–374. <https://doi.org/10.2166/wst.2011.631>
117. Nathao C, Sirisukpoka U, Pisutpaisal N (2014) Production of hydrogen and methane from banana peel by two phase anaerobic fermentation. *Energy Procedia* 50:702–710. <https://doi.org/10.1016/j.egypro.2014.06.086>
118. Kumari S, Das D (2015) Improvement of gaseous energy recovery from sugarcane by dark fermentation followed by biomethanation process. *Bioresour Technol* 192:354–363. <https://doi.org/10.1016/j.biortech.2015.07.038>
119. Jiang H, Qin Y, Gadow SI et al (2018) Bio-hythane production from cassava residue by two-stage fermentative process with recirculation. *Bioresour Technol* 247:769–775. <https://doi.org/10.1016/j.biortech.2017.09.102>
120. Salem AH, Mietzel T, Brunstermann R et al (2018) Two-stage anaerobic fermentation process for bio-hydrogen and bio-methane production from pre-treated organic wastes. *Bioresour Technol* 265:399–406. <https://doi.org/10.1016/j.biortech.2018.06.017>
121. Yeshanew MM, Paillet F, Barrau C et al (2018) Co-production of hydrogen and methane from the organic fraction of municipal solid waste in a pilot scale dark fermenter and methanogenic biofilm reactor. *Front Environ Sci* 6:41. <https://doi.org/10.3389/fenvs.2018.00041>
122. Bolzonella D, Micolucci F, Battista F et al (2020) Producing biohythane from urban organic wastes. *Waste Biomass Valor* 11:2367–2374. <https://doi.org/10.1007/s12649-018-00569-7>
123. Kumar CP, Rena X, Meenakshi A et al (2019) Bio-hythane production from organic fraction of municipal solid waste in single and two stage anaerobic digestion processes. *Bioresour Technol* 294:122220. <https://doi.org/10.1016/j.biortech.2019.122220>
124. Sun C, Xia A, Fu Q et al (2019) Effects of pretreatment and biological acidification on fermentative hydrogen and methane co-production. *Energy Convers Manag* 185:431–441. <https://doi.org/10.1016/j.enconman.2019.01.118>
125. Wongthanate J, Mongkarothai K (2018) Enhanced thermophilic bioenergy production from food waste by a two-stage fermentation process. *Int J Recycl Org Waste Agricul* 7:109–116. <https://doi.org/10.1007/s40093-018-0196-8>
126. Algapani DE, Qiao W, Ricci M et al (2019) Bio-hydrogen and bio-methane production from food waste in a two-stage anaerobic digestion process with digestate recirculation. *Renew Energy* 130:1108–1115. <https://doi.org/10.1016/j.renene.2018.08.079>
127. Yuan T, Bian S, Ko JH et al (2019) Enhancement of hydrogen production using untreated inoculum in two-stage food waste digestion. *Bioresour Technol* 282:189–196. <https://doi.org/10.1016/j.biortech.2019.03.020>
128. Pagliaccia P, Gallipoli A, Gianico A et al (2016) Single stage anaerobic bioconversion of food waste in mono and co-digestion with olive husks: impact of thermal pretreatment on hydrogen and methane production. *Int J Hydrogen Energy* 41:905–915. <https://doi.org/10.1016/j.ijhydene.2015.10.061>
129. Abreu AA, Tavares F, Alves MM et al (2019) Garden and food waste co-fermentation for biohydrogen and biomethane production in a two-step hyperthermophilic-mesophilic process. *Bioresour Technol* 278:180–186. <https://doi.org/10.1016/j.biortech.2019.01.085>
130. Liu X, Li R, Ji M (2019) Effects of two-stage operation on stability and efficiency in co-digestion of food waste and waste activated sludge. *Energies* 12:2748. <https://doi.org/10.3390/en12142748>
131. Farhat A, Miladi B, Hamdi M et al (2018) Fermentative hydrogen and methane co-production from anaerobic co-digestion of organic wastes at high loading rate coupling continuously and sequencing batch digesters. *Environ Sci Pollut Res* 25:27945–27958. <https://doi.org/10.1007/s11356-018-2796-2>
132. Kalia VC, Lal S, Ghai R et al (2003) Mining genomic databases to identify novel hydrogen producers. *Trends Biotechnol* 21:152–156. [https://doi.org/10.1016/S0167-7799\(03\)00028-3](https://doi.org/10.1016/S0167-7799(03)00028-3)
133. Adesra A, Srivastava VK, Varjani S (2021) Valorization of dairy wastes: Integrative approaches for value added products. *Indian J Microbiol* 61:270–278. <https://doi.org/10.1007/s12088-021-00943-5>

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