REVIEW ARTICLE

Association of susceptible genetic markers and autoantibodies in rheumatoid arthritis

VASANTH KONDA MOHAN¹, NALINI GANESAN^{1*} and RAJASEKHAR GOPALAKRISHNAN²

¹Department of Biochemistry and ²Sri Ramachandra Hospital, Sri Ramachandra University, Porur, Chennai 600 116, India

Abstract

Rheumatoid arthritis (RA) is a chronic autoimmune disorder of unknown aetiology resulting in inflammation of the synovium, cartilage and bone. The disease has a heterogeneous character, consisting of clinical subsets of anti-citrullinated protein antibody (ACPA)-positive and APCA-negative disease. Although, the pathogenesis of RA is incompletely understood, genetic factors play a vital role in susceptibility to RA as the heritability of RA is between 50 and 60%, with the human leukocyte antigen (HLA) locus accounting for at least 30% of overall genetic risk. Non-HLA genes, i.e. tumour necrosis factor- α (*TNF*- α) within the MHC (major histocompatibility complex) have also been investigated for association with RA. Although, some contradictory results have originated from several studies on *TNF*- α gene, the data published so far indicate the possible existence of *TNF*- α gene promoter variants that act as markers for disease severity and response to treatment in RA. The correlation of HLA and non-HLA genes within MHC region is apparently interpreted. A considerable number of confirmed associations with RA and other autoimmune disease susceptibility loci including peptidylarginine deiminase type 4 (*PADI4*), protein tyrosine phosphatase non-receptor type 22 (*PTPN22*), signal transducer and activator of transcription (*STAT4*), cluster of differentiation 244 (*CD244*) and cytotoxic T lymphocyte-associated antigen 4 (*CTLA4*), located outside the MHC have been reported recently. In this review, we aim to give an update on recent progress in RA genetics, the importance of the combination of *HLA-DRB1* alleles, non-HLA gene polymorphism, its detection and autoantibodies as susceptibility markers for early RA disease.

[Vasanth K. M., Nalini G. and Rajasekhar G. 2014 Association of susceptible genetic markers and autoantibodies in rheumatoid arthritis. J. Genet. 93, 597–605]

Introduction

Aetiology of rheumatoid arthritis (RA) remains unclear, but that the autoimmunity in RA is represented by the presence of 'rheumatoid factor' (RF) was recognized in patients with RA over 50 years ago (Rose *et al.* 1948). The RF assay, in its current manifestation, remains suboptimal as a diagnostic test, as it lacks sensitivity (54–88%) and specificity (48–92%) (Weinblatt and Schur 1980; Schellekens *et al.* 2000; Bizzaro *et al.* 2001; Bas *et al.* 2002; Saraux *et al.* 2002). In contrast, the identification of anti-citrullinated peptide antibodies (ACPA) and the advancement of commercial tests based on recognition by anti-cyclic citrullinated peptide (anti-CCP) antibodies seem to play a pivotal role in the pathogenesis of RA as they are highly specific (Schellekens *et al.* 2000); RA can be detected years before the onset of symptoms (Berglin *et al.* 2004; Nielen *et al.* 2004).

Genetic contribution to RA pathogenesis has been predicted to be $\sim 60\%$, and the human leukocyte antigen (HLA) region has consistently shown the strongest genetic association with RA (MacGregor et al. 2000). Several studies have shown that shared epitope (SE) alleles are associated with anti-CCP-positive RA but not with anti-CCP-negative RA (Huizinga et al. 2005; Verpoort et al. 2005). Almost 30 years after the designating of HLA alleles as a risk factor for RA, non-HLA genes within the major histocompatibility complex (MHC) have also been examined for association with RA. The genes for tumour necrosis factor (TNF) lie within the MHC and have been a focus of intense interest, given the affirmation that TNF- α plays a central role in the inflammatory cascade in affected joints and the striking efficacy of TNF- α antagonists as therapeutic agents. Results of such analysis, some studies report an association of particular TNF markers with RA (Danis et al. 1995; Mulcahy et al. 1996; van Krugten et al. 1999; Tuokko et al. 2001; Waldron-Lynch et al. 2001), while others find no differences (Wilson et al. 1995; Field et al. 1997; Vinasco et al. 1997;

^{*}For correspondence. E-mail: nalinisrmc@gmail.com.

Keywords. allele; anti-CCP; MHC; shared epitope; SNP.

Yen *et al.* 2001; Low *et al.* 2002). However, it is likely that *HLA-DRB1* is not the only RA susceptibility gene in the MHC domain.

A large number of ratified associations with RA susceptibility genes apart from MHC have been reported lately. Since 2007, there has been an explosion in the number of RA susceptibility genes identified and confirmed in wellpowered cohorts (Begovich et al. 2004; Bowes and Barton 2008; Coenen and Gregersen 2009; Kochi et al. 2009; Plant et al. 2010). It was not until 2003, nevertheless, that the gene peptidylarginine deiminase type 4 (PADI4) was identified in a Japanese population as a second risk factor for RA (Suzuki et al. 2003). The discovery of PADI4 as a risk factor was followed by the introduction of protein tyrosine phosphatase non-receptor type 22 (PTPN22) in 2004 (Begovich et al. 2004; Carlton et al. 2005; Gregersen 2005). A year later, in 2005, cytotoxic T lymphocyte-associated antigen 4 (CTLA4) was found during a candidate gene analysis (Plenge et al. 2005). In 2007, by means of a candidate gene advent (Kurreeman et al. 2007), a novel genetic risk factor was identified in the 9q33 region of the genome containing TRAF1/C5; it was also detected concurrently in a genome wide study (Plenge et al. 2007a, b). In 2007, the signal transducer and activator of transcription (STAT4) gene region on chromosome 2q gene was associated with RA pathogenesis (Remmers et al. 2007). SNPs in cluster of differentiation 244 (CD244) were found to be associated with susceptibility to RA and systemic lupus erythematosus in a Japanese cohort (Suzuki et al. 2008), but not in Korean population (Cho et al. 2009). In the light of these findings, the role of genetics in RA is explored in this article.

Evidence supporting a genetic component in RA

There is extensive evidence of a role for genetic factors in RA. The investigation of monozygotic (MZ) twins revealed increased concordance rates of RA compared to dizygotic twins. The MZ concordance rate for RA is four times higher than the dizygotic (DZ) twin concordance rate, signifying a heritability of 40–60% (Lawrence 1969; Aho *et al.* 1986; Silman *et al.* 1993). The overall MZ twin concordance rate is 12–15%. These twin analyses support an upper limit to the genetic contribution to RA. An interesting study found smoking to be a predictor primarily in the subset of patients with RA-associated *HLA-DRB1* genotypes, illustrating that the genetic and environmental factors could interact in predisposing to RA (Padyukov *et al.* 2004).

Susceptibility HLA genes within the MHC region

Association of HLA–DRB1 SE alleles in RA: In 1987, Gregerson et al. first elaborated a connecting hypothesis for the association of different HLA-DRB1 specificities associated with RA, termed the 'SE hypothesis'. In this they hypothesized an association between RA and HLA-DRB1-SE, including DRB1*04 and DRB1*01 alleles. They showed that RA is associated with specific *HLA-DRB1* (DRB1) alleles that encode a conserved sequence of amino acids, (⁷⁰QRRAA⁷⁴, ⁷⁰RRRAA⁷⁴ or ⁷⁰QKRAA⁷⁴) consist of residues 70–74 in the third hyper variable region (HVR3) of the DRb1 chain (Gregersen *et al.* 1987). These residues constitute a helical domain forming one side of the antigen binding site, a site likely to affect antigen presentation.

The SE hypothesis assumes that these specific class II molecules are directly involved in the pathogenesis of RA. The mechanism underlying SE–RA association is ambiguous. Familiar hypotheses attribute it to presentation of arthritogenic antigens (Wucherpfennig and Strominger 1995), or T-cell repertoire selection (Bhayani and Hedrick 1991). However, it should be indicated that data supporting antigenspecific responses as the primary event in RA are indecisive. Additionally, various non-RA human diseases (Weyand *et al.* 1994; Tait *et al.* 1995), and experimental animal models of autoimmunity (Ollier *et al.* 2001) have shown to be associated with SE-coding alleles as well. Moreover, the antigen-presentation hypothesis is arduous to reconcile with SE allele-dose effects on RA penetrance and disease severity (Holoshitz and Ling 2007).

Association of HLA–DRB1 alleles with serum anti-CCP antibody: Lately, studies have proved that there is a significant association between the SE and RA in RA patients who are anti-CCP antibody-positive (Huizinga *et al.* 2005; Irigoyen *et al.* 2005; Michou *et al.* 2008; Ding *et al.* 2009). For example, a significantly greater prevalence of anti-CCP antibodies was found in those who carried two SE alleles than those with one or none (85, 58 and 30%, respectively). Hence, the SE seems to predispose to anti-CCP positive RA and the development of ACPAs may be a path variable to explain the association of the SE with RA susceptibility and/or severity.

Modelling studies have shown that the SE P4 pocket of the *HLA-DRB1* gene should bind citrulline more efficaciously than arginine. Studies in mice carrying the *HLA-DRB1**0401 transgene have evidenced that converting a residue from arginine to citrulline leads to enhanced T-cell activation and increased binding of peptides by the SE (Hill *et al.* 2003). Hence, it has been hypothesized that smoking leads to elevated citrullination of proteins. Carriage of SE alleles in this environmental background increases susceptibility to RA because they bind citrullinated peptides more strongly and stimulate an exaggerated T-cell response. The exaggerated T-cell response, in turn, may drive the increased autoantibody production by B-cells, containing anti-CCP antibodies, seen in RA (Auger *et al.* 2005; van der Helm-van Mil *et al.* 2006).

van der Woude *et al.* (2009) showed that ACPA-positive and ACPA-negative RA have a similar heritability of 66%. This means that genetic predisposition also plays an important role in the pathogenesis of ACPA-negative RA, for which most individual genetic risk factors remain to be identified.

Non-HLA genes within the MHC region

TNF- α : Non-HLA genes within the MHC have also been examined for association with RA. Genes for tumour necrosis factor (TNF) lies in the class III region of MHC, ~250 kb centromeric of the human leukocyte antigen (HLA)-B locus and 850 kb telomeric of *HLA-DR*. Until now, five polymorphisms have been described within the *TNF-* α gene. Four polymorphisms consist of a guanine (G) to adenine (A) transition at positions -376, -308, -238 and -163 (Wilson *et al.* 1993), and the fifth polymorphism consists of a cytosine (C) insertion in a C-stretch starting at position +70 (Brinkman *et al.* 1995). All known *TNF-* α polymorphisms are situated in the inner region of the gene that is central to the transcriptional regulation of *TNF-* α expression. Single base alterations in such regions may have dynamic effects on gene expression (Matsuda *et al.* 1992).

One of the most studied results suggest that $TNF-\alpha$ gene promoter polymorphisms influence the outcome of this chronic disease. Despite this evidence, the value of genotyping RA patients in order to define their clinical course will remain unproven until a proper prospective evaluation of this cohort of patients validates this hypothesis.

Genetic risk factor located outside the MHC region

PAD14: The genetic variant, *PAD14* gene is located on chromosome 1 (1p36). The *PAD14* gene encodes the type 4 peptidylarginine deiminase enzyme, which catalyses the posttranslational modification of arginine to citrulline, producing citrullinated proteins (table 1; figure 1) (Vossenaar *et al.* 2004). The mechanism by which *PAD14* genotype may influence RA susceptibility has not yet been annotated. Antibodies to these citrullinated peptides are

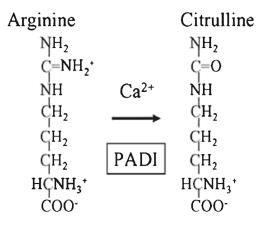


Figure 1. Peptidylarginine deiminase enzyme catalyses the posttranslational modification of arginine to citrulline, generating citrullinated proteins.

extremely specific for RA and usually precede the development of disease, advocating an essential role in RA pathogenesis. *PADI4* was the first non-HLA genetic risk factor known to be associated with RA, especially in Japanese population (Suzuki *et al.* 2003). Association has also been observed in Korean and North American populations (Plenge *et al.* 2005; Kang *et al.* 2006). Studies in Spanish, Swedish and UK populations declared no evidence for association of *PADI4* with RA (Caponi *et al.* 2005; Martinez *et al.* 2005). The strongest association has been authenticated for single nucleotide polymorphism (SNP) located in intron 3 (341-15A \rightarrow T) of *PADI4*, called *PADI4_94* (rs2240340) and a meta-analysis revealed a significant association between RA and the PADI4_94 SNP in Asian community (Takata *et al.* 2008).

Table 1. Cytogenetic loci of RA susceptibility genes and their function.

Gene	Location	Function
HLA-DRB1	6p21.3	Encodes cell surface antigens that present proteins to the T-lymphocytes
TNF-α	6p21.3	Plays a central role in the inflammatory cascade in affected joints and the striking efficacy of TNF- α antagonists as therapeutic agents
PADI4	1p36.13	Encodes enzymes responsible for the conversion of arginine residues to citrulline residues. This gene may play a role in granulocyte and macrophage development leading to inflammation and immune response
PTPN22	1p13.2	Encodes a protein tyrosine phosphatase which is expressed primarily in lymphoid tissues. This enzyme is involved in several signalling pathways associated with the immune response
STAT4	2q32.2-q32.3	Provides instructions for a protein that acts as a transcription factor, which means that it attaches (binds) to specific regions of DNA and helps control the activity of certain genes
		Encodes a transcription factor for signals from certain cytokines
TRAF1-C5	9q33-34	Involved in signalling pathways that play a role in cell proliferation and differentiation, apoptosis, bone remodelling and activation or inhibition of cytokines
CD244	1q23.3	One of the molecules that activates or inhibits natural killer cells have indicated that they play critical roles in the immune system and in autoimmune diseases
CTLA4	2q33	Member of the immunoglobulin superfamily and encodes a protein which transmits an inhibitory signal to T-cells
TNFAIP3	6q23	Encodes a cytoplasmatic zinc-finger protein that inhibits NFKB (nuclear factor of kappa light polypeptide gene enhancer in B-cells) activation and TNF-mediated apoptosis
SPRED2	2p14	Involved in regulating CD45+ cells via the Ras-MAP kinase pathway

Possible other explanations for this disparity are the presence of other genetic or environmental factors that interact with the genetic factor in a specific population, thereby affecting disease susceptibility or enrichment of genetic variants in one population but not in the other. Although the peptidylarginine deiminases are involved in the generation of ACPA, there is no compelling evidence supporting PADI4 genotypes associated with ACPA levels - or ACPA-positive disease in appropriate (van der Helm-van Mil and Huizinga 2008). Recently in India, Panati et al. (2012) investigated polymorphism in exon 4 (padi4 104, [rs1748033]) of PADI4 showed significant association of 'C' allele with RA in the study population (P = 0.0008). Polymorphism in exon 3 (padi4 92, [rs874881]) also exhibited moderate association with the disease (P = 0.075). However, no association of the disease was found with the SNPs padi4 89 (rs11203366) and padi4_90 (rs11203367) in exon 2 of PADI4.

PTPN22: The minor allele of a nonsynonymous SNP (rs2476601, 1858C \rightarrow T, R620W) in the *PTPN22* gene, positioned on chromosome 1p13, has been found to be associated with RA. The PTPN22 R620W was increased in RA patients versus healthy controls in studies of multiple North American and European Caucasian populace, but not in Koreans (Kochi et al. 2009). The PTPN22 gene is a compelling biological candidate for engrossment in autoimmune diseases. It has been found to be expressed in a class of immunologically relevant tissues (Begovich et al. 2004) and encodes the intracellular protein lymphoid tyrosine phosphatase (LYP). Protein tyrosine phosphatases (PTPs) play a crucial role in signal transduction and are integral in the T-cell antigen receptor (TCR) signalling pathway. LYP itself is known to be an effective inhibitor of T-cell activation (table 1) (Hill et al. 2002).

STAT4: The signal transducer and activator of transcription 4 (*STAT4*) gene is stationed on the long (q) arm of chromosome 2 between positions 32.2 and 32.3 is another non-*MHC* gene associated with RA pathogenesis (Remmers *et al.* 2007). The JAK/STAT pathway is the signalling target of a multitude of cytokines that are thought to perform biologically significant roles in rheumatoid synovial inflammation (Walker and Smith 2005). Specifically, STAT4, which encodes *STAT4*, transmits signals induced by several key cytokines, including IL-12, IL-23, and type I interferons (IFNs) (table 1) (Watford *et al.* 2004). In response to cytokines and growth factors, STAT family members are phosphorylated by the receptor-associated kinases, and then form homodimers or heterodimers that translocate to the cell nucleus where they act as transcription activators.

Association of *STAT4* with RA was determined through a combination of linkage and candidate gene studies. In contrast with *HLA-DRB1* and *PTPN22*, the association of *STAT4* with RA is more modest. Four polymorphisms in tight linkage disequilibrium (i.e. rs11889341, rs7574865, rs8179673 and rs10181656) form a susceptibility haplotype which is tagged by the T allele (rs7574865), have the strongest reported association with RA (Remmers et al. 2007). Association of STAT4 variant (rs7574865) with RA was confirmed in patients from European, North American and Asian descent (Zervou et al. 2008; Lee et al. 2010). Europeans appear to have the lowest (21.4%) and Asians the highest (32.0%) prevalence of the rs7574865 variant among the populations studied (Lee et al. 2010). Stratification of RA patients according to the presence of ACPA antibody disclosed a statistically significant association between the rs7574865 variant and RA in both ACPA-positive and ACPA-negative RA patients versus controls (Orozco et al. 2008). In 2012, recent data from Spanish population suggested that patients with early arthritis, who are homozygous for the T allele of rs7574865 in STAT4, may develop a more severe form of the disease with increased disease activity and disability (Lamana et al. 2012).

TRAF1-C5: The TNF receptor-associated factor 1 (TRAF1) and complement component 5 (*C5*) genes are located on chromosome 9 is a member of the TNF receptor-associated factor (TRAF) family, a group of adaptor proteins that bond TNF receptor family members (for example, TNF- α) to downstream signalling (Arch *et al.* 1998). The molecules are involved in signalling pathways that play a role in cell proliferation and differentiation, apoptosis, bone remodelling and activation or inhibition of cytokines (table 1) (Speiser *et al.* 1997). Interestingly, GG homozygotes at the *TRAF1-C5* SNP rs3761847 with RA have a substantially increased risk of death (hazard ratio 3.96, 95% confidence interval 1.24 to 12.6, *P* = 0.02) from malignancy or sepsis, conceivably allowing identification of patients for appropriate screening (Panoulas *et al.* 2009).

CD244: Recent studies on the molecular mechanisms of signalling lymphocyte activation molecule family members, including *CD244*, which is one of the molecules that activates or inhibits natural killer cells have indicated that they play critical roles in the immune system and in autoimmune diseases (table 1) (Veillette 2006). The chromosomal location of a cluster of differentiation 244 (*CD244*) gene is 1q23.1. The Japanese study identified RA susceptibility alleles of two functional SNPs in *CD244* (rs3766379 and rs6682654) that were found to be associated with ~1.5–1.7-fold higher expression levels of *CD244* (Suzuki *et al.* 2008). Newly, determination of the association between SNPs in *CD244* and susceptibility to RA and SLE in a Korean population does not show allelic association with susceptibility to RA (Cho *et al.* 2009).

CTLA4: Genes involved in the regulation of T-cell responses may be primary determinants of susceptibility to RA. *CTLA4* is cytogenetically located in chromosome 2q33 (table 1). Polymorphisms within the *CTLA4* gene appear to

be associated with RA (Lee *et al.* 2003; Zhernakova *et al.* 2005; Suppiah *et al.* 2006), an G \rightarrow A SNP in the 3' untranslated region (CT60; rs3087243) has received more thorough investigation, especially in European populations (Suppiah *et al.* 2006).

A large cohort ($n_{\text{cases/controls}} = 2370/1757$) from the North American Rheumatoid Arthritis Consortium (NARAC) and the Swedish Epidemiological Investigation of Rheumatoid Arthritis (EIRA) collections, provided support for an association of CTLA4 (CT60 allele) with the development of RA, but only in the NARAC cohort (OR = 1.1, 95% CI: 1.0–1.2; P = 0.004) (Plenge *et al.* 2005). When those results were combined with previously published data for CTLA4, it demonstrated continued evidence of association with RA (OR = 1.1, 95% CI: 1.0–1.2; P = 0.01) (Plenge et al. 2005). These earlier results correlated well with a recent meta-analysis which ascertained an association of CTLA4 gene polymorphism with RA in Caucasians (OR = 0.9, $P = 1.8 \times 10-3$) which also reported that CTLA4 embellished the development of ACPA-positive as compared with ACPA-negative RA (Daha et al. 2009). Analogous to HLA-DBR1 SE and PTPN22, these reports clearly indicate that CTLA4 influences the development of RA only in ACCP-positive patients and is supporting the evidence pointing to a divergence in pathology dependent anti-CCP status.

Chromosome 6q23: Two SNPs, rs6920220 (A allele) and rs10499194 (C allele), were found to be independently associated with ACPA+ disease. Both SNPs map to a single linkage disequilibrium block spanning ~60 kb in a region on chromosome 6q23 that lacks known genes or transcripts. The closest genes are oligodendrocyte lineage transcription factor 3 (*OLIG3*) and *tumour necrosis factor* α *-induced protein 3* (*TNFAIP3*). The latter is of potential importance to RA pathogenesis, as the protein *TNFAIP3* acts as a negative regulator of NF- κ B (Wertz *et al.* 2004). So far, however, functional relevance of the reported polymorphisms is unascertained (table 1).

rs6920220 was initially identified in ACPA+ patients with RA (minor allele OR = 1.38) originating from the UK (Wellcome Trust Case Control Consortium 2007). rs10499194 was identified in North American ACPA+ patients (Plenge *et al.* 2007a, b). However, although the intergenic region is certainly associated with RA susceptibility, the involvement of the *TNFAIP3* gene is yet to be confirmed.

Other RA susceptibility genes

A meta-analysis of obtainable data from genomewide association studies of RA exhibited strong evidence for association of the *CD40* gene with RA susceptibility (Raychaudhuri *et al.* 2008). CD40, which is expressed on the surface of B cells, monocytes and dendritic cells, interacts with CD154 on T cells. This interaction is important in immunoglobulin class switching, memory B cell development and germinal centre formation.

An association of SPRED2 locus was reported in an expanded meta-analysis of six genomewide association studies in Caucasian RA samples from the US, UK, Sweden and Canada, all of whom were anti-CCP antibody positive (Stahl *et al.* 2010). The associated SNP mapped to intron 1 of the gene, which is involved in regulating CD45+ cells via the Ras-MAP kinase pathway.

Nongenetic factors

A case–control study in Denmark addressed a large number of environmental factors potentially involved in the aetiology of RA (Pedersen *et al.* 2006). Upon dichotomization of patients with RA according to the presence or absence of anti-CCP antibodies, we show that environmental risk factors differ considerably between anti-CCP-positive and anti-CCP negative RA. One of the most classic genetic risk factors for an autoimmune disease, the shared epitope in RF seropositive RA, is strongly influenced by the presence of a defined environmental risk factor, smoking, in the population at risk.

Similar to genetic associations, a role for environmental factors such as body weight, smoking and blood transfusion make a modest contribution to disease risk and was also confirmed by replicating these findings in large populations (Firestein 1997; Symmons *et al.* 1997).

Detection of HLA and non-HLA polymorphisms

The presence of *HLA-DR4* and *HLA-DRB1* increases the risk of RA 7-fold (Weyand and Goronzy 2000). Long time ago, HLA-A, HLA-B, and HLA-C antigens were identified with the conventional microcytotoxicity test in which well-validated antisera were used. For HLA-DR antigen determinations, enriched suspensions of B-cells were prepared by Legrand *et al.* (1984).

HLA-DRB1 alleles were genotyped by polymerase chain reaction (PCR), sequence-specific oligonucleotide probe hybridization method (SSOPH), which suggests that presence of double *04 SE is associated with a higher risk of developing amyloid A amyloidosis in Japanese patients with RA (Migita *et al.* 2006).

In 2011, Naqi *et al.* genotyped *HLA-DRB1* alleles in Pakistani patients with RA by using low resolution PCR-sequence specific primer (SSP) method and concluded that *HLA-DRB1**04 was expressed with significantly increased frequency in patients with RA. *HLA-DRB1**11 was expressed statistically significantly more in control group as compared to rheumatoid patients indicating a possible protective effect (Naqi *et al.* 2011).

Recently, Mourad and Monem (2013) genotyped *HLA*-*DRB1* alleles by PCR-SSP method. The results indicated that *HLA-DRB1**01, *HLA-DRB1**04, and *HLA-DRB1**010 alleles were related to RA, while the *HLA-DRB1**11 and *HLA-DRB1**13 protect against RA in the Syrian population.

The SSOPH and PCR-SSP approaches, both suffer from the limitations of the need for large numbers of probes or primers and for frequent updating in response to newly published alleles. Direct sequencing is increasingly attractive as a general method for HLA typing.

A number of non-HLA genes outside of the MHC region also exhibit an association with RA. Martinez *et al.* (2005) analysed *PADI4* polymorphism by TaqMan assays and suggested that *PADI4* polymorphisms do not play a role in susceptibility to RA in European population. This study was contrary to findings on Japanese population and concordant with those of previous British and French studies.

Pradhan *et al.* (2012) genotyped *PTPN22* 1858C/T polymorphism by PCR–RFLP method and proposed that there is no direct association between *PTPN22* 1858C/T polymorphism and RA in patients from western India.

Recently, Chang *et al.* (2013) genotyped 17 tag SNPs across the PAD locus using MassARRAY matrix-assisted laser desorption ionization-time-of-flight mass spectrometry system, and suggested that *PADI2* is significantly associated with RA and may be involved in the pathogenesis of the disease.

Conclusions: genetic studies in RA

The last few years have seen tremendous achievements in the identification of RA susceptibility genes. Since 2011, more than 20 robustly associated loci have been found to be involved, mainly with ACPA-positive disease. Still, it is likely that many more remain to be brought to light. Additionally advancements in molecular genetics, such as microarray chips, will allow synchronous large-scale differential identification of thousands of genetic polymorphisms segregating with RA. One of the first genomewide association analyses in RA found several new candidate SNPs for RA and chronic inflammatory arthritis. They performed a replication analysis in an independent subset of SNPs, from which KLF12 emerged as a new candidate susceptibility gene for RA (Julià et al. 2008). In conclusion, a suggested anti-CCP and genetic profile consisting of HLA-DRB1, TNFα, PADI4, PTPN22, STAT4, TRAF1-C5, CD244, CTLA4 and chromosome 6q 23 shows promise to identify those patients who may benefit from a more aggressive treatment regimen or immediate biological biotherapy. The cost of such an extensive panel may be justified by the benefits to the patients and management of disease burden in the long term.

Future research and treatment directions

Although previous case–control studies in different populations have proposed a accessible association of these alleles with RA, controversial results have been disclosed about the significance of genetic variants in affinity with the role of autoantibodies seropositivity in the development of RA. Ethnic differences may play a role in the confliction results among these association studies. However, results show that the selective genotyping approach is more efficient in detecting common variants than detecting rare variants and it is efficient only when the level of declaring significance is not stringent. In summary, the selective genotyping approach is most suitable for detecting common variants in candidate gene-based studies. There is a dearth of such genetic and seropositive association studies in India and we are contriving to undertake such studies (Vasanth and Nalini 2011). Eventually, the contribution of the autoantibodies and genetic markers will lead to an extended knowledge on the pathogenesis of RA and the studied genetic markers will suggest the combined predictive value for RA disease. This knowledge may lead us to make possible earlier preventive strategies as well as the development of more appropriate and effective treatment options in RA.

Acknowledgements

The proposed study is in progress and funded by Department of Science and Technology-Science and Engineering Research Board, New Delhi, India.

References

- Aho K., Koskenvuo M., Tuominen J. and Kaprio J. 1986 Occurrence of rheumatoid arthritis in a nationwide series of twins. *J. Rheumatol.* **13**, 899–902.
- Arch R. H., Gedrich R. W. and Thompson C. B. 1998 Tumor necrosis factor receptor-associated factors (TRAFs)—a family of adapter proteins that regulates life and death. *Genes Dev.* 12, 2821–2830.
- Auger I., Sebbag M., Vincent C., Balandraud N., Guis S., Nogueira L. et al. 2005 Influence of HLA-DR genes on the production of rheumatoid arthritis-specific autoantibodies to citrullinated fibrinogen. Arthritis Rheum. 52, 3424–3432.
- Bas S., Perneger T. V., Kunzle E. and Vischer L. 2002 Comparative study of different enzyme immunoassays for measurement of IgM and IgA rheumatoid factors. *Ann. Rheum. Dis.* 61, 505–510.
- Begovich A. B., Carlton V. E., Honigberg L. A., Schrodi S. J., Chokkalingam A. P., Alexander H. C. *et al.* 2004 A missense single-nucleotide polymorphism in a gene encoding a protein tyrosine phosphatase (PTPN22) is associated with rheumatoid arthritis. *Am. J. Hum. Genet.* **75**, 330–337.
- Berglin E., Padyukov L., Sundin U., Hallmans G., Stenlund H., Van Venrooij W. J. *et al.* 2004 A combination of autoantibodies to cyclic citrullinated peptide (CCP) and HLA-DRB1 locus antigens is strongly associated with future onset of rheumatoid arthritis. *Arthritis Res. Ther.* 6, R303–R308.
- Bhayani H. R. and Hedrick S. M. 1991 The role of polymorphic amino acids of the MHC molecule in the selection of the T cell repertoire. *J. Immunol.* **146**, 1093–1098.
- Bizzaro N., Mazzanti G., Tonutti E., Villalta D. and Tozzoli R. 2001 Diagnostic accuracy of the anti-citrulline antibody assay for rheumatoid arthritis. *Clin. Chem.* 47, 1089–1093.
- Bowes J. and Barton A. 2008 Recent advances in the genetics of RA susceptibility. *Rheumatology* (Oxford) **47**, 399–402.
- Brinkman B. M., Kaijzel E. L., Huizinga T. W., Giphart M. J., Breedveld F. C and Verweij C. L. 1995 Detection of a novel

c-insertion polymorphism within the human tumor necrosis factor alpha gene. *Hum. Genet.* **96**, 493.

- Caponi L., Petit-Teixeira E., Sebbag M., Bongiorni F., Moscato S., Pratesi F. *et al.* 2005 A family based study shows no association between rheumatoid arthritis and the PADI4 gene in a white French population. *Ann. Rheum. Dis.* 64, 587–593.
- Carlton V. E., Hu X., Chokkalingam A. P., Schrodi S. J., Brandon R., Alexander H. C. *et al.* 2005 PTPN22 genetic variation: evidence for multiple variants associated with rheumatoid arthritis. *Am. J. Hum. Genet.* 77, 567–581.
- Chang X., Xia Y., Pan J., Meng Q., Zhao Y. *et al.* 2013 PADI2 is significantly associated with rheumatoid arthritis. *PLoS One* **8**, e81259.
- Cho S. K., Han T. U., Kim K., Bang S. Y., Bae S. C. and Kang C. 2009 *CD244* is not associated with susceptibility to rheumatoid arthritis and systemic lupus erythematosus in a Korean population. *Arthritis Rheum.* **60**, 3153–3154.
- Coenen M. J. and Gregersen P. K. 2009 Rheumatoid arthritis: a view of the current genetic landscape. *Genes Immun.* 10, 101–111.
- Daha N. A., Kurreeman F. A., Marques R. B., Stoeken-Rijsbergen G., Verduijn W., Huizinga T. W. *et al.* 2009 Confirmation of STAT4, IL2/IL21, and CTLA4 polymorphisms in rheumatoid arthritis. *Arthritis Rheum.* **60**, 1255–1260.
- Danis V. A, Millington M., Hyland V., Lawford R., Huang Q. and Grennan D. 1995 Increased frequency of the uncommon allele of a tumour necrosis factor alpha gene polymorphism in rheumatoid arthritis and systemic lupus erythematosus. *Dis. Markers* 12, 127–133.
- Ding B., Padyukov L., Lundström E., Seielstad M., Plenge R. M., Oksenberg J. R. *et al.* 2009 Different patterns of associations with anti-citrullinated protein antibody-positive and anti-citrullinated protein antibody-negative rheumatoid arthritis in the extended major histocompatibility complex region. *Arthritis Rheum.* **60**, 30–38.
- Field M., Gallagher G., Eskdale J., McGarry F., Richards S. D., Munro R. *et al.* 1997 Tumor necrosis factor locus polymorphisms in rheumatoid arthritis. *Tissue Antigens* 50, 303–307.
- Firestein G. S. 1997 Etiology and pathogenesis of rheumatoid arthritis: Textbook of rheumatology, 5th edition, p. 851–897, W. B. Saunders, Philadelphia.
- Gregersen P. K. 2005 Gaining insight into PTPN22 and autoimmunity. Nat. Genet. 37, 1300–1302.
- Gregersen P. K., Silver J. and Winchester R. J. 1987 The shared epitope hypothesis. An approach to understanding the molecular genetics of susceptibility to rheumatoid arthritis. *Arthritis Rheum.* **30**, 1205–1213.
- Hill R. J., Zozulya S., Lu Y. L., Ward K., Gishizky M. and Jallal B. 2002 The lymphoid protein tyrosine phosphatase Lyp interacts with the adaptor molecule Grb2 and functions as a negative regulator of T-cell activation. *Exp. Hematol.* **30**, 237–244.
- Hill J. A., Southwood S., Sette A., Jevnikar A. M., Bell D. A. and Cairns E. 2003 Cutting edge: the conversion of arginine to citrulline allows for a high-affinity peptide interaction with the rheumatoid arthritis-associated HLADRB1*0401 MHC class II molecule. J. Immunol. 171, 538–541.
- Holoshitz J. and Ling S. 2007 Nitric oxide signaling triggered by the rheumatoid arthritis shared epitope: a new paradigm for MHCdisease association. *Ann. New York Acad. Sci.* 1110, 73–83.
- Huizinga T. W., Amos C. I., van der Helm-van Mil A. H., Chen W., van Gaalen F. A., Jawaheer D. *et al.* 2005 Refining the complex rheumatoid arthritis phenotype based on specificity of the HLA-DRB1 shared epitope for antibodies to citrullinated proteins. *Arthritis Rheum.* 52, 3433–3438.
- Irigoyen P., Lee A. T., Wener M.H., Li W., Kern M., Batliwalla F. et al. 2005 Regulation of anti-cyclic citrullinated peptide antibodies in rheumatoid arthritis: contrasting effects of HLA-DR3 and the shared epitope alleles. Arthritis Rheum. 52, 3813–3818.

- Julià A., Ballina J., Canete J. D., Balsa A., Tornero-Molina J., Naranjo A. *et al.* 2008 Genome-wide association study of rheumatoid arthritis in the Spanish population: KLF12 as a risk locus for rheumatoid arthritis susceptibility. *Arthritis Rheum.* 58, 2275–2286.
- Kang C. P., Lee H. S., Ju H., Cho H., Kang C. and Bae S. C. 2006 A functional haplotype of the PADI4 gene associated with increased rheumatoid arthritis susceptibility in Koreans. *Arthritis Rheum.* 54, 90–96.
- Kochi Y., Suzuki A., Yamada R. and Yamamoto K. 2009 Genetics of rheumatoid arthritis: underlying evidence of ethnic differences. *J. Autoimmun.* 32, 158–162.
- Kurreeman F. A., Padyukov L., Marques R. B., Schrodi S. J., Seddighzadeh M., Stoeken-Rijsbergen G. *et al.* 2007 A candidate gene approach identifies the TRAF1/C5 region as a risk factor for rheumatoid arthritis. *PLoS Med.* 4, e278.
- Lamana A., Balsa A., Rueda B., Ortiz A. M., Nuno L., Miranda-Carus M. E. *et al.* 2012 The TT genotype of the STAT4 rs7574865 polymorphism is associated with high disease activity and disability in patients with early arthritis. *PLoS One* 7, e43661.
- Lawrence J. S. 1969 The epidemiology and genetics of rheumatoid arthritis. *Rheumatology* 2, 1–36.
- Lee C. S., Lee Y. J, Liu H. F., Su C. H., Chang S. C., Wang B. R. et al. 2003 Association of CTLA4 gene A-G polymorphism with rheumatoid arthritis in Chinese. *Clin. Rheumatol.* 22, 221–224.
- Lee Y. H., Woo J. H., Choi S. J., Ji J. D. and Song G. G. 2010 Association between the rs7574865 polymorphism of STAT4 and rheumatoid arthritis: a metaanalysis. *Rheumatol. Int.* **30**, 661– 666.
- Legrand L., Lathrop G. M., Marcelli-Barge A., Dryll A., Bardin T., Debeyre N. *et al.* 1984 HLA-DR genotype risks in seropositive rheumatoid arthritis. *Am. J. Hum. Genet.* **36**, 690–699.
- Low A. S., Gonzalez-Gay M. A., Akil M., Amos R. S., Bax D. E., Cannings C. *et al.* 2002 TNF +489 polymorphism does not contribute to susceptibility to rheumatoid arthritis. *Clin. Exp. Rheumatol.* 20, 829–832.
- MacGregor A. J., Snieder H., Rigby A. S., Koskenvuo M., Kaprio J., Aho K. *et al.* 2000 Characterizing the quantitative genetic contribution to rheumatoid arthritis using data from twins. *Arthritis Rheum.* 43, 30–37.
- Martinez A., Valdivia A., Pascual-Salcedo D., Lamas J. R., Fernández-Arquero M., Balsa A. *et al.* 2005 PADI4 polymorphisms are not associated with rheumatoid arthritis in the Spanish population. *Rheumatology* 44, 1263–1266.
- Matsuda M., Sakamoto N. and Fukumaki Y. 1992 Delta-thalassemia caused by disruption of the site for an erythroid-specific transcription factor, GATA-1, in the delta-globin gene promoter. *Blood* **80**, 1347–1351.
- Michou L., Teixeira V. H., Pierlot C., Lasbleiz S., Bardin T., Dieudé P. et al. 2008 Associations between genetic factors, tobacco smoking and autoantibodies in familial and sporadic rheumatoid arthritis. Ann. Rheum. Dis. 67, 466–470.
- Migita K., Nakamura T., Maeda Y., Miyashita T., Origuchi T., Yatsuhashi H. *et al.* 2006 HLA-DRB1*04 alleles in Japanese rheumatoid arthritis patients with AA amyloidosis. *J. Rheumatol.* **33**, 2120–2123.
- Mourad J. and Monem F. 2013 HLA-DRB1 allele association with rheumatoid arthritis susceptibility and severity in Syria. *Rev. Bras. Reumatol.* **53**, 51–56.
- Mulcahy B., Waldron-Lynch F., McDermott M. F., Adams C., Amos C. I., Zhu D. K. *et al.* 1996 Genetic variability in the tumor necrosis factor-lymphotoxin region influences susceptibility to rheumatoid arthritis. *Am. J. Hum. Genet.* 59, 676.
- Naqi N., Ahmed T. A., Malik J. M., Ahmed M. and Bashir M. M. 2011 HLA DRβ1 Alleles in Pakistani patients with rheumatoid arthritis. *J. Coll. Physicians Surg. Pak.* **21**, 727–730.

- Nielen M. M., van Schaardenburg D., Reesink H. W., van de Stadt R. J., van der Horst-Bruinsma I. E., de Koning M. H. *et al.* 2004 Specific autoantibodies precede the symptoms of rheumatoid arthritis: a study of serial measurements in blood donors. *Arthritis Rheum.* **50**, 380–386.
- Ollier W. E., Kennedy L. J., Thomson W., Barnes A. N., Bell S. C. and Bennett D. 2001 Dog MHC alleles containing the human RA shared epitope confer susceptibility to canine rheumatoid arthritis. *Immunogenetics* **53**, 669–673.
- Orozco G., Alizadeh B. Z., Delgado-Vega A. M., González-Gay M. A, Balsa A., Pascual-Salcedo D. *et al.* 2008 Association of STAT4 with rheumatoid arthritis: a replication study in three European populations. *Arthritis Rheum.* 58, 1974–1980.
- Padyukov L., Silva C., Stolt P., Alfredsson L. and Klareskog L. 2004 A gene environment interaction between smoking and shared epitope genes in HLADR provides a high risk of seropositive rheumatoid arthritis. *Arthritis Rheum.* 50, 3085–3092.
- Panati K., Pal S., Rao K. V. and Reddy V. D. 2012 Association of single nucleotide polymorphisms (SNPs) of PADI4 gene with rheumatoid arthritis (RA) in Indian population. *Genes Genet. Syst.* 87, 191–196.
- Panoulas V. F., Smith J. P., Nightingale P. and Kitas G. D. 2009 Association of the TRAF1/C5 locus with increased mortality, particularly from malignancy or sepsis, in patients with rheumatoid arthritis. *Arthritis Rheum.* 60, 39–46.
- Pedersen M., Jacobsen S., Klarlund M., Pedersen B. V., Wiik A., Wohlfahrt J. *et al.* 2006 Environmental risk factors differ between rheumatoid arthritis with and without auto-antibodies against cyclic citrullinated peptides. *Arthritis Res. Ther.* 8, R133.
- Plant D., Flynn E., Mbarek H., Dieudé P., Cornelis F., Arlestig L. et al. 2010 Investigation of potential non-HLA rheumatoid arthritis susceptibility loci in a European cohort increases the evidence for nine markers. Ann. Rheum. Dis. 69, 1548–1553.
- Plenge R. M., Padyukov L., Remmers E. F., Purcell S., Lee A. T., Karlson E. W. *et al.* 2005 Replication of putative candidate-gene associations with rheumatoid arthritis in >4,000 samples from North America and Sweden: association of susceptibility with PTPN22, CTLA4, and PADI4. *Am. J. Hum. Genet.* 77, 1044– 1060.
- Plenge R. M., Cotsapas C., Davies L., Price A. L, de Bakker P. I., Maller J. *et al.* 2007a Two independent alleles at 6q23 associated with risk of rheumatoid arthritis. *Nat. Genet.* **39**, 1477–1482.
- Plenge R. M., Seielstad M., Padyukov L., Lee A. T., Remmers E. F., Ding B. *et al.* 2007b TRAF1-C5 as a risk locus for rheumatoid arthritis—a genomewide study. *N. Engl. J. Med.* 357, 1199–1209.
- Pradhan V. D., Dalvi H., Parsannavar D., Rajadhyaksha A., Patwardhan M. and Ghosh K. 2012 Study of PTPN22 1858C/T polymorphism in rheumatoid arthritis patients from Western India. *Ind. J. Rheumatol.* 7, 130–134.
- Raychaudhuri S., Remmers E. F., Lee A. T, Hackett R., Guiducci C., Burtt N. P. *et al.* 2008 Common variants at CD40 and other loci confer risk of rheumatoid arthritis. *Nat. Genet.* 40, 1216–1223.
- Remmers E. F., Plenge R. M., Lee A. T., Graham R.R., Hom G., Behrens T. W. *et al.* 2007 STAT4 and the risk of rheumatoid arthritis and systemic lupus erythematosus. *N. Engl. J. Med.* 357, 977–986.
- Rose H. M., Ragan C., Pearce E. and Lipman M. O. 1948 Differential agglutination of normal and sensitized sheep erythrocytes by sera of patients with rheumatoid arthritis. *Proc. Soc. Exp. Biol. Med.* 68, 1–6.
- Saraux A., Berthelot J. M., Chales G., Le Henaff C., Mary J. Y., Thorel J. B. *et al.* 2002 Value of laboratory tests in early prediction of rheumatoid arthritis. *Arthritis Rheum.* 47, 155–165.
- Schellekens G. A., Visser H, de Jong B. A., van den Hoogen F. H., Hazes J. M., Breedveld F. C. *et al.* 2000 The diagnostic properties of rheumatoid arthritis antibodies recognizing a cyclic citrullinated peptide. *Arthritis Rheum.* **3**, 155–163.

- Silman A. J., MacGregor A. J., Thomson W., Holligan S., Carthy D., Farhan A. *et al.* 1993 Twin concordance rates for rheumatoid arthritis: results from a nationwide study. *Br. J. Rheumatol.* 32, 903–907.
- Speiser D. E., Lee S. Y., Wong B., Arron J., Santana A., Kong Y. Y. *et al.* 1997 A regulatory role for TRAF1 in antigen-induced apoptosis of T cells. *J. Exp. Med.* 185, 1777–1783.
- Stahl E. A., Raychaudhuri S., Remmers E. F., Xie G., Eyre S., Thomson B. P. *et al.* 2010 Genome-wide association study metaanalysis identifies seven new rheumatoid arthritis risk loci. *Nat. Genet.* 42, 508.
- Suppiah V., O'Doherty C., Heggarty S., Patterson C. C., Rooney M. and Vandenbroeck K. 2006 The CTLA4 +49A/G and CT60 polymorphisms and chronic inflammatory arthropathies in Northern Ireland. *Exp. Mol. Pathol.* **80**, 141–146.
- Suzuki A., Yamada R., Chang X., Tokuhiro S., Sawada T., Suzuki M. *et al.* 2003 Functional haplotypes of PADI4, encoding citrullinating enzyme peptidylarginine deiminase 4, are associated with rheumatoid arthritis. *Nat. Genet.* 34, 395–402.
- Suzuki A., Yamada R., Kochi Y., Sawada T., Okada Y., Matsuda K. *et al.* 2008 Functional SNPs in CD244 increase the risk of rheumatoid arthritis in a Japanese population. *Nat. Genet.* **40**, 1224–1249.
- Symmons D. P., Bankhead C. R., Harrison B. J., Brennan P., Barrett E. M., Scott D. G. *et al.* 1997 Blood transfusion, smoking, and obesity as risk factors for the development of rheumatoid arthritis: results from a primary care-based incident case-control study in Norfolk, England. *Arthritis Rheum.* 40, 1955–1961.
- Tait B. D., Drummond B. P., Varney M. D. and Harrison L. C. 1995 HLA-DRB1*0401 is associated with susceptibility to insulindependent diabetes mellitus independently of the DQB1 locus. *Eur. J. Immunogenet.* 22, 289–297.
- Takata Y., Inoue H., Sato A., Tsugawa K., Miyatake K., Hamada D. et al. 2008 Replication of reported genetic associations of PADI4, FCRL3, SLC22A4 and RUNX1 genes with rheumatoid arthritis: results of an independent Japanese population and evidence from meta-analysis of East Asian studies. J. Hum. Genet. 53, 163–173.
- Tuokko J., Nejentsev S., Luukkainen R., Toivanen A. and Ilonen J. 2001 HLA haplotype analysis in Finnish patients with rheumatoid arthritis. *Arthritis Rheum.* **44**, 315–322.
- van der Helm-van Mil A. H. and Huizinga T. W. 2008 Advances in the genetics of rheumatoid arthritis point to subclassification into distinct disease subsets. *Arthritis Res. Ther.* **10**, 205.
- van der Helm-van Mil A. H., Verpoort K. N., Breedveld F. C., Huizinga T. W., Toes R. E. and de Vries R. R. 2006 The HLA-DRB1 shared epitope alleles are primarily a risk factor for anticyclic citrullinated peptide antibodies and are not an independent risk factor for development of rheumatoid arthritis. *Arthritis Rheum.* 54, 1117–1121.
- van der Woude D., Houwing-Duistermaat J. J., Toes R. E., Huizinga T. W., Thomson W., Worthington J. *et al.* 2009 Quantitative heritability of anti-citrullinated protein antibody-positive and anti-citrullinated protein antibody-negative rheumatoid arthritis. *Arthritis Rheum.* **60**, 916–923.
- van Krugten M. V., Huizinga T. W., Kaijzel E. L., Zanelli E., Drossaers-Bakker K. W., van de Linde P. *et al.* 1999 Association of the TNF +489 polymorphism with susceptibility and radiographic damage in rheumatoid arthritis. *Genes Immun.* **1**, 91– 96.
- Vasanth K. M. and Nalini G. 2011 Usefulness of genetic markers in rheumatoid arthritis. Proceedings of the International Conference on Medical Genetics and Genomics ICMG-2011: December 12–14; Bharathidasan University, Tiruchirappalli, India.
- Veillette A. 2006 Immune regulation by SLAM family receptors and SAP-related adaptors. *Nat. Rev. Immunol.* 6, 56–66.

- Verpoort K. N., van Gaalen F. A., van der Helm-van Mil A. H., Schreuder G. M., Breedveld F. C., Huizinga T. W. *et al.* 2005 Association of HLA-DR3 with anti-cyclic citrullinated peptide antibody-negative rheumatoid arthritis. *Arthritis Rheum.* 52, 3058–3062.
- Vinasco J., Beraun Y., Nieto A., Fraile A., Mataran L., Pareja E. et al. 1997 Polymorphism at the TNF loci in rheumatoid arthritis. *Tissue Antigens* 49, 74–78.
- Vossenaar E. R., Després N., Lapointe E., van der Heijden A., Lora M., Senshu T. *et al.* 2004 Rheumatoid arthritis specific anti-Sa antibodies target citrullinated vimentin. *Arthritis Res. Ther.* 6, R142–R150.
- Waldron-Lynch F., Adams C., Amos C., Zhu D. K., McDermott M. F., Shanahan F. *et al.* 2001 Tumour necrosis factor 5' promoter single nucleotide polymorphisms influence susceptibility to rheumatoid arthritis (RA) in immunogenetically defined multiplex RA families. *Genes Immun.* 2, 82– 87.
- Walker J. G. and Smith M. D. 2005 The Jak-STAT pathway in rheumatoid arthritis. *J. Rheumatol.* **32**, 1650–1653.
- Watford W. T., Hissong B. D., Bream J. H., Kanno Y., Muul L. and O'Shea J. J. 2004 Signaling by IL-12 and IL-23 and the immunoregulatory roles of STAT4. *Immunol. Rev.* **202**, 139–156.
- Weinblatt M. E. and Schur P. H. 1980 Rheumatoid factor detection by nephelometry. *Arthritis Rheum.* 23, 777–779.
- Wellcome Trust Case Control Consortium 2007 Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature* 447, 661–678.
- Wertz I. E., O'Rourke K. M., Zhou H., Eby M., Aravind L., Seshagiri S. *et al.* 2004 De-ubiquitination and ubiquitin ligase domains of A20 downregulate NF-kappaB signalling. *Nature* 430, 694–699.

- Weyand C.M. and Goronzy J. 2000 Association of MHC and rheumatoid arthritis: HLA polymorphisms in phenotypic variants of rheumatoid arthritis. *Arthritis Res.* **2**, 212–216.
- Weyand C. M, Hunder N. N., Hicok K. C., Hunder G. G. and Goronzy J. J. 1994 HLA-DRB1 alleles in polymyalgia rheumatica, giant cell arteritis, and rheumatoid arthritis. *Arthritis Rheum.* 37, 514–520.
- Wilson A. G., de Vries N., Pociot F., di Giovine F. S., van der Putte L. B. and Duff G. W. 1993 An allelic polymorphism within the human tumor necrosis factor alpha promoter region is strongly associated with HLA A1, B8, and DR3 alleles. *J. Exp. Med.* 177, 557–560.
- Wilson A. G, de Vries N., van de Putte L. B. and Duff G. W. 1995 A tumour necrosis factor alpha polymorphism is not associated with rheumatoid arthritis. *Ann. Rheum. Dis.* 54, 601–603.
- Wucherpfennig K. W. and Strominger J. L. 1995 Selective binding of self peptides to disease-associated major histocompatibility complex (MHC) molecules: a mechanism for MHC-linked susceptibility to human autoimmune diseases. J. Exp. Med. 181, 1597–1601.
- Yen J. H., Chen C. J., Tsai W. C., Lin C. H., Ou T. T, Wu C. C. et al. 2001 Tumor necrosis factor promoter polymorphisms in patients with rheumatoid arthritis in Taiwan. J. Rheumatol. 28, 1788–1792.
- Zervou M. I., Sidiropoulos P., Petraki E., Vazgiourakis V., Krasoudaki E., Raptopoulou A. *et al.* 2008 Association of a TRAF1 and a STAT4 gene polymorphism with increased risk for rheumatoid arthritis in a genetically homogeneous population. *Hum. Immunol.* **69**, 567–571.
- Zhernakova A., Eerligh P., Barrera P., Wesoly J. Z., Huizinga T. W., Roep B.O. *et al.* 2005 CTLA4 is differentially associated with autoimmune diseases in the Dutch population. *Hum. Genet.* **118**, 58–66.
- Received 2 July 2013, in revised form 3 January 2014; accepted 5 January 2014 Unedited version published online: 11 June 2014 Final version published online: 23 July 2014