# **REGULAR ARTICLE**



# Specific detection of hypochlorite: a cyanine based turn-on fluorescent sensor

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Abstract. Judiciously designed cyanine based fluorogenic probe (L) can exhibit interesting solvent polarity induced isomerization. The probe displayed a highly selective TURN-ON fluorescence response towards hypochlorite among various reactive oxygen species (ROS) and analytes in a mixed aqueous medium. The sensing process was attributed to the formation of a probe-OCl adduct which results in restricting the donor-acceptor extended conjugation. The detection limit was found to be as low as 3  $\mu$ M. The proposed sensing mechanism is supported by mass spectrometric analysis and HPLC study.

Keywords. Isomerization; fluorescent probe; TURN-ON response; hypochlorite.

# 1. Introduction

Reactive oxygen species (ROS) such as  ${}^{1}O_{2}$ ,  ${}^{\bullet}OH$ ,  $O_{2}^{-}$ ,  $H_2O_2$ , t-BuOOH and HOCl/ClO<sup>-</sup> cause oxidative stress and play vital roles in toxicology as well as pathology.<sup>1,2</sup> Reactive oxygen species (ROS) have very close involvement in diverse biological processes including its alleged role in many diseases like cancer and neurodegenerative disorders.<sup>3-6</sup> Hypochlorous acid (HClO) or its conjugate base hypochlorite (ClO<sup>-</sup>) is a well-known ROS which is commonly used as bleaching agents.<sup>7–9</sup> Hypochlorous acid (HClO)/hypochlorite (ClO<sup>-</sup>) can be generated endogenously from the peroxidation reaction of hydrogen peroxide and chloride ion, catalyzed by the enzyme myeloperoxidase (MPO).<sup>10-14</sup> Importantly, Hypochlorous acid (HClO) can serve as a powerful antimicrobial agent and it exerts a destructive effect on invading bacteria and pathogens to enable a defence mechanism in the immune system.<sup>15,16</sup> However, unregulated ClO- production could lead to the damage of DNA and proteins which might induce several ailments such as cardiovascular diseases, atherosclerosis, osteoarthritis, rheumatoid arthritis, lung injury and

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even various types of cancer.  $^{17-23}$  Hence, the sensitive detection of ClO- with high selectivity is much awaited to understand the biological insight of ClO<sup>-</sup> related diseases.

The fluorescence-based sensing probes  $^{24-33}$  are much more attractive compared to other systems owing to the superiority of fluorescence technique in terms of high sensitivity, rapid response and user friendliness along with its ability to provide non-invasive temporal detection of target analytes in living cells. However, till now there are only a few reports which deal with the sensitive detection of (HClO)/hypochlorite (ClO<sup>-</sup>) using fluorescent probe.<sup>5,34-49</sup> Some mitochondria-targeted fluorescent probes for HClO/ClO<sup>-</sup> have been reported recently which significantly contribute to research related to the role of ClO<sup>-</sup> in physiology and pathology.<sup>50-55</sup> Among them, a number of reported fluorescent probes for (HClO)/hypochlorite (ClO<sup>-</sup>) detection are mostly confined to oxime and imine based probes.<sup>5,34–46,52</sup> But oxime or imine based probes might be vulnerable to acid catalyzed hydrolysis which clearly may curb their application potential in complex biological milieu.<sup>19</sup> Especially inside the cancer cells, wherein the lower pH environment exists, the stability of oxime-based fluorescent probe might become a pertinent issue.<sup>56–60</sup> Hence, cyanine based probes  $^{61,62}$  with high stability could be an alternative system to develop hypochlorite sensor.

Soham Samanta and Senjuti Halder both authors have contributed equally to this study.

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In this context, we would like to report a new cyanine based spiropyran fluorogenic probe which can demonstrate a highly selective TURN-ON fluorescence response towards hypochlorite ( $ClO^{-}$ ) in 1:1 methanolaqueous medium. The sensing process was explained because of the disruption in extended conjugation due to the addition reaction between L and ClO- anion.

#### 2. Experimental

#### 2.1 General information and materials

All the materials for synthesis were purchased from commercial suppliers and used without further purification. The absorption spectra were recorded on a Perkin-Elmer Lamda-25 UV-Vis spectrophotometer using 10 mm path length quartz cuvettes in the range of 300–800 nm wavelength, whereas fluorescence measurements were performed on a Horiba Fluoromax-4 spectrofluorometer using 10 mm path length quartz cuvettes with a slit width of 3 nm at 298 K. The mass spectrum of the probe L was obtained using Waters Q-ToF Premier mass spectrometer. Nuclear magnetic resonance (NMR) spectra were recorded on a Bruker Avance 600 MHz NMR instrument. The chemical shifts were recorded in parts per million (ppm) on the scale. The following abbreviations are used to describe spin multiplicities in <sup>1</sup>H NMR spectra: s = singlet; d = doublet; t = triplet; q = quartet, m = multiplet.

#### 2.2 Synthesis of the probe L

First. 3-ethyl-1,1,2-trimethyl-1H-benzo[e]indol-3-ium was synthesized through the reported procedure as described in our previously published work (first step of Scheme 1).<sup>26,28</sup> Thereafter, 1.0 mmol of this 3-ethyl-1,1,2-trimethyl-1Hbenzo[e]indol-3-ium was taken in a round-bottomed flask and it was dissolved in EtOH by gentle warming. 1.5 mmol of anhydrous sodium acetate was then added to it and the mixture was refluxed for 1 h. Subsequently, 1.1 mmol of 2-hydroxy-1-naphthaldehyde was added to the reaction mixture in the heating condition and it was further refluxed for 6 h (Scheme 1). A colourless crystalline solid product was obtained upon cooling the reaction mixture to the room temperature. The solid product was then filtered, washed thoroughly with cold methanol and dried in a desiccator. Calculated yield: 64%. <sup>1</sup>H NMR [600 MHz, CDCl<sub>3</sub>, TMS, J (Hz),  $\delta$  (ppm)]: 8.06 (1H, d, J = 8.4), 7.95 (1H, d, J = 8.4), 7.81 (1H, d, J = 8.4), 7.76 (2H, m), 7.65 (2H, m), 7.53 (1H, t, J = 7.2), 7.41 (1H, t, J = 7.2), 7.35 (1H, t, J = 7.2), 7.22 (1H, t, J = 7.2), 7.02 (1H, d, J = 9.0), 6.93 (1H, d, J = 9.0),5.87 (1H, t, J = 10.2), 3.46 (2H, m), 1.69 (3H, s), 1.37 (3H, s) 1.22 (3H, t, J = 7.2). <sup>13</sup>C NMR [150 MHz, CDCl<sub>3</sub>, TMS, δ (ppm)]: 152.80, 145.13, 130.46, 130.18, 129.99, 129.67, 129.21, 128.98, 128.81, 128.77, 126.85, 126.34, 125.25, 123.34, 121.45, 121.39, 120.70, 117.90, 117.68, 110.23, 110.08, 105.81, 53.86, 38.05, 24.16, 21.82, 14.93. ESI-MS



Scheme 1. Design and synthesis of probe L.

(positive mode, m/z) Calculated for  $C_{28}H_{25}NO$  (M + H<sup>+</sup>): 392.2009. Found: 392.2051.

#### 2.3 Preparation of ROS and RNS

Various ROS and RNS including <sup>1</sup>O<sub>2</sub>, <sup>•</sup>OH, O<sub>2</sub><sup>-</sup>, H<sub>2</sub>O<sub>2</sub>, t-BuOOH, ClO<sup>-</sup>, NO<sup>-</sup><sub>2</sub>, NO<sup>-</sup><sub>3</sub>, NO and ONOO<sup>-</sup> were prepared according to the following procedures. The exact concentration of hypochlorite (OCl<sup>-</sup>) solution was determined from the extinction coefficient of 350 M<sup>-1</sup>cm<sup>-1</sup> (292 nm) at pH=9.0. Hydroxyl radical (•OH) was produced using Fenton reaction, wherein  $Fe(NH_4)_2(SO_4)_2 \cdot 6H_2O$  was reacted with 10 equivalents of  $H_2O_2$  to generate •OH. The concentration of the prepared •OH was estimated from the concentration of  $Fe^{2+}$ . Commercially available stock of H<sub>2</sub>O<sub>2</sub> was used for the study and concentration of the H2O2 solution was estimated by optical absorbance at 240 nm. Singlet oxygen  $({}^{1}O_{2})$  was generated by adding NaOCl to H2O2 according to the reported procedure.<sup>63</sup> The source of  $NO_2^-$  and  $NO_3^-$  were NaNO<sub>2</sub> and NaNO<sub>3</sub> respectively. Nitric oxide (NO) was prepared from sodium nitro-ferricyanide(III) dihydrate. Peroxynitrite (ONOO<sup>-</sup>) was prepared following the reported method<sup>64</sup> and the concentration of it was estimated by using an extinction coefficient of 1670  $M^{-1}cm^{-1}$  (302 nm).<sup>65</sup> Superoxide ( $O_2^-$ ) is prepared from KO<sub>2</sub>. t-BuOOH was obtained from commercial suppliers.

#### 2.4 UV-Vis and fluorescence spectroscopic studies

Stock solutions of various anions  $(1 \times 10^{-1} \text{ mol.L}^{-1})$  were prepared in methanol or Millipore water depending upon the preferred solubility of the analytes chosen. A stock solution of L  $(5 \times 10^{-3} \text{ mol.L}^{-1})$  was prepared in DMSO. The solution of L was then diluted to  $10 \times 10^{-6} \text{ mol.L}^{-1}$  for spectral studies by taking only 4.0 µL stock solution of L and making the final volume to 2.0 mL by adding 1:1 methanol-water mixed solvent. In fluorescence selectivity experiment, the test samples were prepared by placing appropriate amounts of the stock solutions of the respective anions/analytes into 2.0 mL of probe solution  $(10 \times 10^{-6} \text{ mol.L}^{-1})$ . For UV-visible and fluorescence titration experiments, another set of OCl<sup>-</sup> standard solution having 5.0 mM concentration was prepared by diluting the earlier prepared stock solutions  $(1 \times 10^{-1} \text{ mol.L}^{-1})$  in Millipore water. Quartz optical cells of 1.0 cm path length were filled with 2.0 mL solutions of L for UV-visible and fluorescence titration experiments respectively, to which the newly prepared stock solutions of  $OCI^-$  (5.0 mM) were gradually added using a micropipette as necessary. For fluorescence measurements, excitation was provided at 360 nm and emission was acquired from 380 nm to 650 nm. Spectral data were recorded within 1 min after addition of the analytes except for kinetic studies.

### 2.5 Detection limit

The detection limit was calculated on the basis of the fluorescence titration. The fluorescence emission spectrum of **L** was measured 10 times, and the standard deviation of blank measurement was estimated ( $\lambda_{em} = 435$  nm). To measure the slope, the fluorescence emission at 435 nm was plotted as a function of the concentration of OCl<sup>-</sup> from the titration experiment. The detection limit was then calculated using the following equation:

Detection limit = 
$$3\sigma/k$$
 (1)

Where  $\sigma$  is the standard deviation of blank measurement, and k is the slope between the fluorescence emission intensity versus [OCl<sup>-</sup>].

#### 2.6 Crystallization of L

A small amount of brown-red solid of L was taken in a test tube containing DCM-EtOH mixture and sonicated for 5 min to obtain a clear solution. Then the solution was allowed to evaporate slowly at room temperature and after one week, X-ray mountable plate-shaped orange-red colored crystals appeared at the bottom of the test tube.

2.6a Crystallographic refinement details: Herein, a suitable crystal was selected from the mother liquor and immersed into silicone oil, then mounted on the tip of glass fibre and cemented using epoxy resin. Intensity data for all crystals were collected Mo-K  $\alpha$  radiation ( $\lambda = 0.71073$ Å) at 298(2) K, with increasing  $\omega$  (width of 0.3° per frame) at a scan speed of 6 s/ frame on a Bruker SMART APEX diffractometer equipped with CCD area detector. The data integration and reduction were processed with SAINT<sup>66</sup> software. Empirical absorption correction was applied to the collected reflections with SADABS.<sup>67</sup> The structures were solved by direct methods using SHELXTL-2014 and were refined on F2 by the full-matrix least-squares technique using the SHELXL-2014 program package.<sup>68</sup> Graphics are generated using MERCURY 2.3.69 In all the cases, non-hydrogen atoms are treated anisotropically. The hydrogen atoms are located on a difference Fourier map and refined. In other cases, the hydrogen atoms are geometrically fixed.

CCDC number – 1572275, formula – C<sub>28</sub>H<sub>25</sub>NO, Fw – 391.49, crystal system – monoclinic, space group – *P* 21/c, a = 22.441(4) Å, b = 11.222 (14) Å, c = 18.966(2) Å,  $\alpha =$ 90.00,  $\beta = 114.981(4)$ ,  $\gamma = 90.00$ , V = 4329.5(10) Å<sup>3</sup>, Z = 4,  $D_c = 0.601$  g cm<sup>-3</sup>,  $\mu$  (Mo K  $\alpha$ ) = 0.036 mm<sup>-1</sup>, F(000) = 832.0, T = 298(2) K, Theta<sub>max</sub> = 28.532, total no. of reflections = 75550, independent reflections = 10726, observed reflections = 8457, parameters refined = 274,  $R_1$ ,  $I > 2\sigma(I) = 0.1438$ , w $R_2$ ,  $I > 2\sigma(I) = 0.3210$ , GOF ( $F^2$ ) = 0.993.

# 3. Results and Discussions

# 3.1 Design, synthesis and structural elucidation of the probe L

The probe was synthesized by following a procedure summarized in Scheme 1. Condensation of an indole derivative and 2-hydroxy-1-naphthaldehyde in the presence of anhydrous sodium acetate led to the formation of the probe L with about 64% product yield. The synthesized probe was well-characterized by NMR and MS analysis. The structure of L was further reaffirmed by its crystal structure analysis. The X-ray crystal structure reveals that L exists in spiropyran (SP) ring form rather than merocyanine (MC) form in the solid state. The X-ray single-crystal structure of L showed that it has adopted a non-centrosymmetric molecular arrangement with the polar space group of P 21/c (experimental methods). The crystal packing of the probe L suggested that two different types of conformers are present in a unit cell of L (Figure 1a). Both the types of conformers of L are subjected to eleven short-range  $C - H \cdots \pi$ intermolecular interactions each (Figure 1b-1c). The aromatic naphthyl planes of the two conformers are found to have almost perpendicular orientations (87.7°) (Figure 1d). The packing diagrams of L have been presented in Figure 1e and Figure 1f.

#### 3.2 Isomerization of the probe L

It is well known that spiropyran derivatives can exhibit photoisomerization.<sup>70,71</sup> In the absence of UV irradiation, these types of compounds mainly exist in spiropyran (SP) form and do not show any prominent absorbance in the visible region (Scheme 2). On the contrary, upon UV irradiation it transforms into a merocyanine (MC) form which displays a well-defined absorbance maximum in the visible region.<sup>72</sup> In accordance with this, we found that probe L does not reveal any noticeable absorbance peak in an aqueous medium which clearly indicates that the probe is in the SP form. However, it was noted that when we varied the polarity of the medium by changing the water fraction of a methanol-water mixture, there were interesting changes in the colourimetric behaviour of L (Figure 2).



**Figure 1.** (A) ORTEP molecular structure at 30% probability level of two conformers, Intermolecular  $C - H \cdots \pi$  interactions within the range of 2.814–2.899 Å in (B) conformer 1, (C) conformer 2, (D) Almost perpendicular orientations of aromatic naphthyl planes of two conformer, (E) Packing motif along crystallographic *b* axis depicting wave-like architecture construction by alternative conformer array in symmetry-equivalence (blue-conformer 1, green-conformer 2) and (F) molecular packing in single crystals in symmetry-equivalence as viewed along crystallographic *c* axis.

It was observed that when the water fraction of the methanol-water mixture is high (>70%) the probe does not reveal any absorbance peak above 400 nm (Figure 2). However, increasing the methanol content of the mixed solvent leads to the generation of a new absorbance peak at around 583 nm. Interestingly when the methanol content of the mixed solvent is 50% the absorbance maximum of the probe L at 583 nm increases substantially and reaches the highest absorbance value (Figure 2). However, a further increase in the methanol content from 50% to 100% resulted in the gradual decrease in the absorbance maxima at 583 nm (Figure 2). Hence, the study clearly indicates that the probe preferentially attains merocyanine form when there is 50% methanol and 50% water in a methanol-water mixed solvent. As the merocyanine form of the probe could only comprise a potential nucleophilic site, it is important to carry out the sensing studies of L in an optimal condition wherein the conversion to the MC form from SP form will be maximum. Herein, the probe can be easily transformed into MC form without irradiation of UV light just by changing the water fraction of a methanol-water mixed solvent. Hence, we can utilize the probe for sensing purpose with merely adjusting the ratio of methanol and



Scheme 2. Solvent polarity induced isomerization of the probe L for sensing study.

water in a methanol-water mixed solvent. As the maximum conversion to the MC form is attained at 50% methanol and 50% water in a methanol-water mixed solvent, all the subsequent spectral studies of L regarding its sensing pursuit in 1:1 methanol-aqueous solution were carried out at room temperature.



**Figure 2.** Changes in the absorbance behavior of **L** upon varying the water fraction of the methanol-water mixture of the mixed solvent; Bottom: visual changes.

#### 3.3 UV-Vis spectroscopic study of L

Probe L displayed a strong absorption peak at 583 nm  $(2800 \text{ M}^{-1} \text{ cm}^{-1})$  in 1:1 methanol- aqueous solution at room temperature as shown in Figure 3a. The absorption band at 583 nm is possibly originated from the conjugated cyanine moiety present in L in its MC form and the absorption band might be attributed to the  $\pi - \pi *$ charge transfer. Initially, UV-Vis spectroscopic studies of L were carried out by recording the absorption spectral changes upon the addition of hypochlorite and other reactive oxygen/nitrogen species. The addition of excess (60 equivalents) of various ROS/RNS to L induced a negligible change in the colourimetric behaviour of L except for hypochlorite (Figure S6, Supplementary Information). It was noted that among various ROS/RNS only addition of OCl<sup>-</sup> could substantially reduce the absorbance maximum of L (Figure 3a) at 583 nm. This spectral change was accompanied by a visible color change as violet (only L) experimental solution turned into almost colorless (L+ OCl<sup>-</sup>) solution (Figure 3a inset). No other reactive species was able to yield any substantial change in the spectral behaviour when interacted with L, which clearly indicates the high selectivity of the probe towards OCl<sup>-</sup>. It was also noted that the incremental addition of  $OCl^{-}$  to L (0–60 equivalents) led to a regular decrease in the absorbance maxima of L at 583 nm (Figure 3). So, the UV-Visible spectral studies suggest that the probe L could exhibit good selectivity



**Figure 3.** UV-Visible spectra of L (10  $\mu$ M) in presence of varying concentration of OCl<sup>-</sup> (0–60 equivalents), INSET: Visual change in the color of the solution of L in presence of OCl<sup>-</sup> under daylight.



**Figure 4.** Fluorescence spectra of L  $(10 \,\mu\text{M})$  in 1:1 methanol-aqueous solution in presence of excess (40 equivalents) of various anions, ROS, RNS and bioanalytes.

towards OCl<sup>-</sup>. A fluorescence study was followed in detail to understand the analytical prospect of L.

# 3.4 Fluorescence spectral response of L towards OCl<sup>-</sup>

The selectivity, sensitivity and response time of L toward OCl<sup>-</sup> were investigated using fluorescence spectroscopy. Free probe L was found to be almost non-fluorescent in 1:1 methanol-aqueous solution wherein the merocyanine (MC) form predominates. Hence, upon excitation at 360 nm, L revealed a very low-intensity emission band around 435 nm (Figure 4). However, the fluorescence intensity of the probe L increased



**Figure 5.** Fluorescence spectra of L (10  $\mu$ M) in the presence of varying concentration of OCl<sup>-</sup>; INSET: Changes in the emission intensity at 435 nm with the addition of equivalents of OCl<sup>-</sup>.

more than 10 folds upon addition of 40 equivalents of OCl<sup>-</sup> to L and a well-defined emission maximum was found to emerge at 435 nm (Figure 4). However, negligible changes in fluorescence were observed when L interacted with an excess of other ROS and RNS (Figure 4). Especially, the common interfering ROS/RNS like  ${}^{1}O_{2}$ ,  ${}^{\bullet}OH$ ,  $O_{2}^{-}$ ,  $H_{2}O_{2}$ , *t*-BuOOH, NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup> and ONOO<sup>-</sup> could not bring any noticeable change in the fluorescence intensity of L when added to the solution of L in the similar experimental conditions. Moreover, it was also found that various common anions and bio-analytes including F<sup>-</sup>, Cl<sup>-</sup>,  $Br^{-}, I^{-}, CH_{3}COO^{-}, H_{2}PO_{4}^{-}, PF_{6}^{-}, HCO_{3}^{-}, SO_{4}^{2-}, HSO_{4}^{-}, HSO_{4}^{$  $ClO_4^-$ ,  $SCN^-$ ,  $S_2O_3^{2-}$ ,  $S_2O_8^{2-}$ , pyrophosphate(PPi), cysteine(Cys), homocysteine(Hcy) and glutathione(GSH) hardly induced any change in the fluorescence behavior of L when added to it. Hence, the probe shows an outstanding selectivity towards OCl<sup>-</sup>. It is also worth mentioning, some recent literature have reported that reactive nucleophilic species like SO<sub>2</sub> derivatives and H<sub>2</sub>S can react with various hemi-cyanine dyes containing nucleophilic reaction site.<sup>31,33,73</sup> Hence, it was pertinent to check the selectivity of probe L towards  $SO_2$  derivatives ( $SO_3^{2-}$ ) and  $H_2S$  derivatives ( $HS^{-}$ ). It was noted that even the presence of 200 equivalents of SO<sub>2</sub> derivatives and H<sub>2</sub>S could not bring any noticeable change to the fluorogenic behaviour of the probe L which further reiterated the high selectivity of the probe towards OCl<sup>-</sup> (Figure S7, Supplementary Information). Interestingly, the time-dependent fluorescence study of L revealed that it can produce a noticeable change in the fluorescence intensity within seconds of



Scheme 3. Plausible sensing mechanism.

the addition of OCl<sup>-</sup> which clearly underlines the rapid sensing aptitude of the probe (Figure S8, Supplementary Information). Hence, the probe can sense hypochlorite selectively with a rapid TURN-ON fluorescence response. However, it is very important to get a quantitative appraisal of the interaction between OCl<sup>-</sup> and the probe L to explore the possibility of using the probe in real samples. Thus, a fluorescence titration experiment was performed with varying OCl<sup>-</sup> concentration. It was noted that the gradual incremental addition of OC1<sup>-</sup> to L in 1:1 methanol-aqueous solution at room temperature resulted in a sequential increase of the emission intensity at 435 nm as evident from Figure 5. It should be mentioned here that the detection limits for OCl<sup>-</sup> was determined from the fluorescence titration experiment. The detection limit for OCl- was found to be  $3.0 \times 10^{-6}$  M (Figure S9, Supplementary Information). In addition, since the Cyanine is sensitive to pH, and the solution of NaOCl is alkaline, the following investigation study was performed to explore the effect of pH on the emission intensity of L at 435 nm upon OCl<sup>-</sup> addition (Figure S10, Supplementary Information).

#### 3.5 Mechanism of L responding to OCl<sup>-</sup>

The sensing of the OCl<sup>-</sup> by L can be accredited to the nucleophilic addition reaction between L and OCl<sup>-</sup>. The proposed sensing mechanism is depicted in Scheme 3. Essentially the nucleophilic attack of OCl<sup>-</sup> to the nucleophilic site of L resulted in the formation of the product L-OCl adduct which is responsible for the observed change in the optical behaviour of L. It is worth mentioning that the plausible nucleophilic attack of OCl<sup>-</sup> led to the rupture in the conjugation in L which is reflected in the observed decolouring of the solution of L upon interacting with OCl<sup>-</sup>. To validate the proposed sensing mechanism mass spectrometric study was conducted. The mass spectrometric analysis indicated towards the

formation of L-OCl adduct when L interacted with  $OCl^{-}$  by showing a prominent peak at m/z = 485.6401 (Figure S11, Supplementary Information). Moreover, a reverse phase HPLC study of L carried out in the presence of OCl<sup>-</sup> also further endorsed the formation of L-OCl adduct. It was observed that HPLC of pure L resulted in a single peak with a retention time of 26.93 min. However, when HPLC was run after the addition of excess OCl<sup>-</sup> there was a new prominent peak generated with a retention time of 25.67 min apart from the pure ligand peak which is indicative of the formation of L-OCl adduct (Figure S12, Supplementary Information). Hence, these studies supported the premise that nucleophilic addition of OCl- toward L is key to sensing. However, it was interesting to observe that  $SO_3^{2-}$ , even being a better nucleophile could not produce a similar fluorescence change. This might be explained by the steric effect around the nucleophilic site. Probably owing to the larger size of sulfite compared to hypochlorite, it could not undergo a nucleophilic addition reaction at the specific reaction site.

# 4. Conclusions

In conclusion, a new fluorogenic probe was successfully designed to demonstrate interesting isomerization. The probe demonstrated a highly selective rapid TURN-ON fluorescence response towards hypochlorite in a mixed aqueous medium. The sensing process was explained as a consequence of the probe-OCl adduct formation; which in turn restricted the donor-acceptor extended conjugation to facilitate the TURN-ON fluorescence response. The sensing mechanism is well-corroborated by mass spectroscopic and HPLC study.

# Supplementary Information (SI)

The supplementary information is available at www.ias.ac.in/ chemsci.

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# References

 Dickinson B C and Chang C J 2011 Chemistry and biology of reactive oxygen species in signalling or stress responses *Nat. Chem. Biol.* 7 504

- Finkel T and Holbrook N J 2000 Oxidants, oxidative stress and the biology of ageing *Nature* 408 239
- Lambeth J D 2007 Nox enzymes, ROS, and chronic disease: An example of antagonistic pleiotropy *Free Radic. Biol. Med.* 43 332
- 4. Bove P F and van der Vliet 2006 A Nitric oxide and reactive nitrogen species in airway epithelial signaling and inflammation *Free Radic. Biol. Med.* **41** 515
- Chen X, Wang F, Hyun J Y, Wei T, Qiang J, Ren X, Shin I and Yoon J 2016 Recent progress in the development of fluorescent, luminescent and colorimetric probes for detection of reactive oxygen and nitrogen species *Chem. Soc. Rev.* 45 2976
- Zhang R, Zhao J, Han G, Liu Z, Liu C, Zhang C, Liu B, Jiang C, Liu R, Zhao T, Han M Y and Zhang Z 2016 Real-time discrimination and versatile profiling of spontaneous reactive oxygen species in living organisms with a single fluorescent probe *J. Am. Chem. Soc.* **138** 3769
- 7. Aoki T and Munemori M 1983 Continuous flow determination of free chlorine in water *Anal. Chem.* **55** 209
- 8. Hidalgo E, Bartolome R and Dominguez C 2002 Cytotoxicity mechanisms of sodium hypochlorite in cultured human dermal fibroblasts and its bactericidal effectiveness *Chem.-Biol. Interact.* **139** 265
- 9. Winterbourn C C, Hampton M B, Livesey J H and Kettle A J 2006 Modeling the reactions of superoxide and myeloperoxidase in the neutrophil phagosome: Implications for microbial killing *J. Biol. Chem.* **281** 39860
- Fiedler T J. Davey C A and Fenna R E 2000 X-ray crystal structure and characterization of halide-binding sites of human myeloperoxidase at 1.8 Å resolution *J. Biol. Chem.* 275 11964
- Yap Y W, Whiteman M and Cheung N S 2007 Chlorinative stress: An under appreciated mediator of neurodegeneration? *Cell Signal.* 19 219
- Pattison D I and Davies M J 2006 Evidence for rapid inter- and intramolecular chlorine transfer reactions of histamine and carnosine chloramines: Implications for the prevention of hypochlorous-acid-mediated damage *Biochemistry* 45 8152
- 13. Harrison J and Schultz J 1976 Studies on the chlorinating activity of myeloperoxidase *J. Biol. Chem.* **251** 1371
- 14. Klebanoff S J 2005 Myeloperoxidase: Friend and foe J. *Leukoc. Biol.* **77** 598
- 15. Prokopowicz Z M, Arce F, Biedron R, Chiang C L L, Ciszek M D, Katz R, Nowakowska M, Zapotoczny S, Marcinkiewicz J and Chain B M 2010 Hypochlorous acid: A natural adjuvant that facilitates antigen processing, cross-priming, and the induction of adaptive immunity J. Immunol. **184** 824
- 16. Pattison D I and Davies M J 2001 Absolute rate constants for the reaction of hypochlorous acid with protein side chains and peptide bonds *Chem. Res. Toxicol.* **14** 1453
- 17. Pullar J M, Vissers M C and Winterbourn C C 2000 Living with a killer: The effects of hypochlorous acid on mammalian cell *IUBMB Life* **50** 259
- Sugiyama S, Okada Y, Sukhova G K, Virmani R, Heinecke J W and Libby P 2001 Macrophage myeloperoxidase regulation by granulocyte macrophage colonystimulating factor in human atherosclerosis and implications in acute coronary syndromes *Am. J. Pathol.* 158 879

- Steinbeck M J, Nesti L J, Sharkey P F and Parvizi J 2007 Myeloperoxidase and chlorinated peptides in osteoarthritis: Potential biomarkers of the disease J. Orthop. Res. 25 1128
- Greig F H, Hutchison L, Spickett C M and Kennedy S 2015 Differential effects of chlorinated and oxidized phospholipids in vascular tissue: Implications for neointima formation *Clin. Sci.* **128** 579
- Daugherty A, Dunn J L, Rateri D L and Heinecke J W 1994 Myeloperoxidase, a catalyst for lipoprotein oxidation, is expressed in human atherosclerotic lesions J. *Clin. Invest.* 94 437
- 22. Sugiyama S, Kugiyama K, Aikawa M, Nakamura S, Ogawa H and Libby P 2004 Hypochlorous acid, a macrophage product, induces endothelial apoptosis and tissue factor expression: Involvement of myeloperoxidase-mediated oxidant in plaque erosion and thrombogenesis *Arterioscler. Thromb. Vasc. Biol.* **24** 1309
- 23. Oushiki D, Kojima H, Terai T, Arita M, Hanaoka K, Urano Y and Nagano T 2010 Development and application of a near-infrared fluorescence probe for oxidative stress based on differential reactivity of linked cyanine dyes *J. Am. Chem. Soc.* **132** 2795
- 24. Samanta S, Goswami S, Hoque Md N, Ramesh A and Das G 2014 An aggregation-induced emission (AIE) active probe renders Al(III) sensing and tracking of subsequent interaction with DNA *Chem. Commun.* 50 11833
- 25. S Samanta, T Ray, F Haque and Das G 2016 A turn-on Rhodamine B-indole based fluorogenic probe for selective sensing of trivalent ions *J. Lumin.* **171** 13
- 26. Samanta S, Goswami S, Ramesh A and Das G 2015 A new chemodosimetric probe for the selective detection of trivalent cations in aqueous medium and live cells *J. Photochem. Photobiol. A* **310** 45
- 27. Samanta S, Goswami S, Ramesh A and Das G 2014 A new fluorogenic probe for solution and intra-cellular sensing of trivalent cations in model human cells *Sens. Actuat. B* **194** 120
- Samanta S, Kar C and Das G 2015 Colorimetric and fluorometric discrimination of geometrical isomers (Maleic Acid vs Fumaric Acid) with real-time detection of maleic acid in solution and food additives *Anal. Chem.* 87 9002
- 29. Samanta S, Manna U, Ray T and Das G 2015 An aggregation-induced emission (AIE) active probe for multiple targets: A fluorescent sensor for Zn<sup>2+</sup> and Al<sup>3+</sup> & a colorimetric sensor for Cu<sup>2+</sup> and F<sup>-</sup> Dalton Trans. 44 18902
- Samanta S, Datta B K, Boral M, Nandan A and Das G 2016 A multi-responsive turn-on flurogenic probe to sense Zn<sup>2+</sup>, Cd<sup>2+</sup> and Pb<sup>2+</sup>: left-right-center emission signal swing *Analyst* 141 4388
- 31. Samanta S, Dey P, Ramesh A and Das G 2016 A solo fluorogenic probe for the real-time sensing of  $SO_3^{2-}$  and  $SO_4^{2-}/HSO_4^{-}$  in aqueous medium and live cells by distinct turn-on emission signals *Chem. Commun.* **52** 10381
- 32. Samanta S, Manna U and Das G 2017 White-light emission from simple AIE-ESIPT-excimer tripled single molecular system *New J. Chem.* **41** 1064
- 33. Samanta S, Halder S, Dey P, Manna U, Ramesh A and Das G 2017 A ratiometric fluorogenic probe for the

real-time detection of  $SO_3^{(2-)}$  in aqueous medium: application in a cellulose paper based device and potential to sense  $SO_3^{(2-)}$  in mitochondria *Analyst* **143** 250

- 34. Emrullahoğlu M, Üçüncü M and Karakuş E 2013 A BODIPY aldoxime-based chemodosimeter for highly selective and rapid detection of hypochlorous acid *Chem. Commun.* **49** 7836
- Reja S I, Bhalla V, Sharma A, Kaur G and Kumar M 2014 A highly selective fluorescent probe for hypochlorite and its endogenous imaging in living cells *Chem. Commun.* 50 11911
- Zhan Z, Liu R, Chai L, Li Q, Zhang K and Lv Y 2017 Turn-on fluorescent probe for exogenous and endogenous imaging of hypochlorous acid in living cells and quantitative application in flow cytometry *Anal. Chem.* 89 9544
- 37. Zhang Y R, Liu Y, Feng X and Zhao B X 2017 Recent progress in the development of fluorescent probes for the detection of hypochlorous acid *Sens. Actuat. B* **240** 18
- Qiao L, Nie H, Wu Y, Xin F, Gao C, Jing J and Zhang X 2017 An ultrafast responsive BODIPY-based fluorescent probe for the detection of endogenous hypochlorite in live cells J. Mater. Chem. B 5 525
- Wang X, Wang X, Feng Y, Zhu, Yin H, Guo Q and Meng X 2015 A two-photon fluorescent probe for detecting endogenous hypochlorite in living cells *Dalton Trans*. 44 6613
- 40. Chen W C, Venkatesan P and Wu S P 2015 A turnon fluorescent probe for hypochlorous acid based on HOCl-promoted removal of the C=N bond in BODIPYhydrazone *New J. Chem.* **39** 6892
- Kang J, Huo F, Yue Y, Wen Y, Chao J, Zhang Y and Yin C 2017 A solvent depend on ratiometric fluorescent probe for hypochlorous acid and its application in living cells *Dyes Pigm.* 136 852
- 42. Zhao Y, Li H, Xue Y, Ren Y and Han T 2017 A phenanthroimidazole-based fluorescent probe for hypochlorous acid with high selectivity and its bioimaging in living cells *Sens. Actuat. B* 241 335
- Xu X and Qian Y 2017 A novel pyridyl triphenylamine-BODIPY aldoxime: Naked-eye visible and fluorometric chemodosimeter for hypochlorite *Spectrochim. Acta A* 183 356
- 44. Wang E, Qiao H, Zhou Y, Pang L, Yu F, Zhang J and Ma T 2015 A novel "turn-on" fluorogenic probe for sensing hypochlorous acid based on BODIPY *RSC Adv.* **5** 73040
- 45. Wang B, Chen D, Kambam S, Wang F, Wang Y, Zhang W, Yin J, Chen H and Chen X 2015 A highly specific fluorescent probe for hypochlorite based on fluorescein derivative and its endogenous imaging in living cells *Dyes Pigm.* **120** 22
- 46. Guo B, Nie H, Yang W, Tian Y, Jing J and Zhang X 2016 A highly sensitive and rapidly responding fluorescent probe with a large Stokes shift for imaging intracellular hypochlorite *Sens. Actuat. B* **236** 459
- 47. Chen H, Shang H, Liu Y, Guo R and Lin W 2016 Development of a unique class of spiro-type twophoton functional fluorescent dyes and their applications for sensing and bioimaging Adv. Funct. Mater. 26 8128
- 48. Xie X, Wu T, Wang X, Li Y, Wang K, Zhai Z, Jiao X and Tang B 2018 A two-photon fluorescent probe for

ratiometric visualization of hypochlorous acid in live cells and animals based on a selenide oxidation/elimination tandem reaction *Chem. Commun.* **54** 11965

- 49. Huang Y, Zhang P, Gao M, Zeng F, Qin A, Wu S and Tang B Z 2016 Ratiometric detection and imaging of endogenous hypochlorite in live cells and in vivo achieved by using an aggregation-induced emission (AIE)-based nanoprobe *Chem. Commun.* 52 7288
- Zhou X D and Yu X Q 2015 A ratiometric fluorescent probe for in situ quantification of basal mitochondrial hypochlorite in cancer cells *Chem. Commun.* 51 6781
- Li K, Hou J T, Yang J and Yu X Q 2017 A tumorspecific and mitochondria-targeted fluorescent probe for real-time sensing of hypochlorite in living cells *Chem. Commun.* 53 5539
- 52. Li D, Feng Y, Lin J, Chen M, Wang S, Wang X, Sheng H, Shao Z, Zhu M and Meng X 2016 A mitochondria-targeted two-photon fluorescent probe for highly selective and rapid detection of hypochlorite and its bio-imaging in living cells *Sens. Actuat. B* **222** 483
- 53. Xiao H D, Li J H, Zhao J, Yin G, Quan Y W, Wang J and Wang R Y 2015 A colorimetric and ratiometric fluorescent probe for ClO<sup>-</sup> targeting in mitochondria and its application in vivo J. Mater. Chem. B 3 1633
- 54. Shen B X, Qian Y, Qi Z Q, Lu C G, Sun Q, Xia X and Cui Y P 2017 Near-infrared BODIPY-based two-photon ClO<sup>-</sup> probe based on thiosemicarbazide desulfurization reaction: Naked-eye detection and mitochondrial imaging *J. Mater. Chem. B* 5 5854
- 55. Yuan L, Wang L, Agrawalla B K, Park S J, Zhu H, Sivaraman B, Peng J, Xu Q H and Chang Y T 2015 Development of targetable two-photon fluorescent probes to image hypochlorous acid in mitochondria and lysosome in live cell and inflamed mouse model *J. Am. Chem. Soc.* 137 5930
- Thakur B, Jayakumar S and Sawant S N 2015 Probing extracellular acidity of live cells in real time for cancer detection and monitoring anti-cancer drug activity *Chem. Commun.* 51 7015
- Liu J, Ren J, Bao X, Gao W, Wu C and Zhao Y 2016 pH-Switchable fluorescent probe for spatially-confined visualization of intracellular hydrogen peroxide *Anal. Chem.* 88 5865
- Li S Y, Liu L H, Jia H Z, Qiu W X, Rong L, Cheng H and Zhang X Z 2014 A pH-responsive prodrug for realtime drug release monitoring and targeted cancer therapy *Chem. Commun.* 50 11852

- 59. Raju G S R, Benton L, Pavitra E and Yu J S 2015 Multifunctional nanoparticles: Recent progress in cancer therapeutics *Chem. Commun.* **51** 13248
- Negrini R, Fong W K, Boyd B J and Mezzenga R 2015 pH-responsive lyotropic liquid crystals and their potential therapeutic role in cancer treatment *Chem. Commun.* 51 6671
- 61. Wu Y, Wang J, Zeng F, Huang S, Huang J, Xie H, Yu C and Wu S 2016 Pyrene derivative emitting red or nearinfrared light with monomer/excimer conversion and its application to ratiometric detection of hypochlorite *ACS Appl. Mater. Interfaces* **8** 1511
- 62. Xu J, Yuan H, Qin C, Zeng L and Bao G M 2016 A mitochondria-targeted near-infrared probe for colorimetric and ratiometric fluorescence detection of hypochlorite in living cells *RSC Adv.* **6** 107525
- Li X H, Zhang G X, Ma H M, Zhang D Q, Li J and Zhu D B 2004 4,5-Dimethylthio-4'-[2-(9-anthryloxy)ethylthio] tetrathiafulvalene, a highly selective and sensitive chemiluminescence probe for singlet oxygen *J. Am. Chem. Soc.* 126 11543
- 64. Sun Z, Liu F, Chen Y, Tam P K H and Yang D 2008 A highly specific BODIPY-based fluorescent probe for the detection of hypochlorous acid *Org. Lett.* **10** 2171
- 65. Yu S Y, Hsu C Y, Chen W C, Wei L F and Wu S P 2014 A hypochlorous acid turn-on fluorescent probe based on HOCl-promoted oxime oxidation and its application in cell imaging *Sens. Actuat. B* **196** 203
- 66. Sheldrick G M 1995 SAINT and XPREP 5.1 Siemens Industrial Automation Inc.: Madison, WI
- 67. Sheldrick G M 1999-2003 SADABS, software for Empirical Absorption Correction, University of Gottingen, Institute fur Anorganische Chemie der Universitat, Tammanstrasse 4, D-3400 Gottingen, Germany
- 68. Sheldrick G M 2015 Crystal structure refinement with SHELXL Acta Crystallogr. Sect C: Struct. Chem. 71 3
- 69. Mercury 2.3 Supplied with Cambridge Structural Database; CCDC: Cambridge, UK, 2011–2012
- Liu F and Morokuma K 2013 Multiple pathways for the primary step of the spiropyran photochromic reaction: A CASPT2//CASSCF study J. Am. Chem. Soc. 135 10693
- 71. Be'nard S and Yu P 2000 New spiropyrans showing crystalline-state photochromism *Adv. Mater* **12** 48
- Minkin V I 2004 Photo-, thermo-, solvato-, and electrochromic spiroheterocyclic compounds *Chem. Rev.* 104 2751
- 73. Chen Y, Zhu C, Yang Z, Chen J, He Y, Jiao Y, He W, Qiu L, Cen J and Guo Z 2013 A ratiometric fluorescent probe for rapid detection of hydrogen sulfide in mitochondria *Angew. Int. Ed. Chem.* 52 1688