

# A brief history of *Frankia* and actinorhizal plants meetings

## 1. Introduction

Meetings of various scopes exist that address various needs of the scientific community. There are meetings of large microbiological societies such as the ASM that occur every year with thousands of participants and several sessions in parallel. These constitute a unique opportunity to get overviews of recent developments in several subfields, in concepts and in technological improvements. At the other end of the spectrum are small meetings such as the '*Frankia* and actinorhizal plants meetings'. These gather people who almost all know each other and have interacted with one another in the past because they work on biological objects with major unusual peculiarities. Such is the case of the '*Frankia* and actinorhizal plants' meetings that have taken place for the last 35 years.

## 2. Main events

Since the first description of swellings on the roots of alder and Russian olive and the supposed presence of a bacterium (Brunchorst 1886), there have been hundreds of attempts at isolation of the bacterium causing them. If the bacterium causing nodules in Legumes have been described at about the same time as *Frankia*, the fast-growing *Rhizobium* was cultivated much more rapidly (Beijerinck 1888). A very exhaustive review (Baker and Torrey 1979) has been done of published isolation attempts of *Frankia* with the proposed identification of the microbes recovered that highlighted the tremendous variety of microorganisms recovered, ranging from fungi to bacteria, including mycoplasma, proteobacteria and actinobacteria.

Many of these isolates were tested for their ability to fulfill Koch's postulates and for an in vitro morphology comparable to that seen inside the nodules. In retrospect, only the claims made by Pommer (1956, 1959) are valid but the strain was not disseminated and subsequently lost. All other isolates either did not have the expected morphology or else took months to cause nodules on their host plant roots, raising the possibility that contaminations may have occurred. These unfounded claims created a need for standards to decide when to call an isolation a success and an isolate a *Frankia*. When Torrey and his co-workers (Callaham *et al.* 1978) obtained an infective isolate from sweet fern (*Comptonia peregrina*) that had the expected morphological features, they felt the need to interact with their colleagues, convince them of the validity of their claim and create a dynamic cycle of interactions. Another factor was the growing recognition that actinorhizals played an ecological role commensurate with that played by Legumes. That was the backdrop for the organization of the first *Frankia* meeting that took place in Harvard Forest, MA, in April 1978, organized by JG Torrey and JD Tjepkema. The proceedings of the meeting were published as a special issue of the *Botanical Gazette* (Torrey and Tjepkema 1979). This meeting grouped about 35 scientists from the US and Canada. Many papers published in that special issue detailed the course of nodulation (Benson and Eveleigh 1979), the extended host range of the isolate relative to that obtained with crushed nodule (Lalonde 1979), or modifications of the tedious isolation procedure that was thought at the time to be critical (Baker *et al.* 1979), as well as argued the case for commercial or ecological applications of actinorhizal plants in forestry (Gordon and Dawson 1979; Klemmedson 1979).

The second meeting took place the following year in 1979 in Corvallis, OR organized by JC Gordon, CT Wheeler and DA Perry and reflected the need felt to monitor rapid progress done after the first meeting (Gordon *et al.* 1979). It reunited 62 attendees, all from the US except 7 from Canada and 1 from Sweden. The meeting was dedicated to Dr G Bond, FRS (1906–1988, professor of botany, University of Glasgow,

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UK), who had been a pioneer on physiology of actinorhizal plants and who had retired in 1976. Many strains were reported from a variety of hosts, ultrastructure of nodules from several actinorhizal plants was described and a new actinorhizal plant was announced, the herbaceous *Datisca* from California and Pakistan (Winship and Chaudhary 1979). Several papers emphasized applications of actinorhizals such as large scale production of inoculant (Lalonde and Calvert 1979) or industrial uses of red alder (Resch 1980). Published in *Nature* shortly afterward was the demonstration that contrary to *Rhizobium*, *Frankia* could develop vesicles and fix nitrogen in vitro (Tjepkema *et al.* 1980).

The third meeting took place in Madison, WI in 1982 organized by JG Torrey, JD Tjepkema and their local host J Ensign and the proceedings were published in the *Canadian Journal of Botany* (Torrey and Tjepkema 1983). Sixty scientists exchanged on various subjects such as *Frankia* plasmids with an eye on genetic transformation, ecology of host plants for industrial applications or isolation of DNA for future species delineation and biotechnological work. Host genera from which *Frankia* isolates had been cultured and shown to fulfill Koch's postulates (and form nitrogenase) included *Alnus*, *Casuarina*, *Comptonia*, *Elaeagnus*, *Hippophae* and *Myrica* and synthetic medium were compared (Tisa *et al.* 1983). Also a strain from *Casuarina* was shown to be able to infect its original host plant (Diem *et al.* 1983) after the previous mind-boggling isolation of an 'atypical' strain that grew well but was not infective on *Casuarina* (Gauthier *et al.* 1981) but shown to be infective on another host plant, *Hippophae*.

The fourth meeting took place in Wageningen in The Netherlands in 1983 organized by ADL Akkermans, D Baker, K Huss-Danell and JD Tjepkema and the proceedings were published in *Plant & Soil* in 1984 (Akkermans *et al.* 1984). The meeting was dedicated to Dr A Quispel (1960–1983, professor of experimental botany, University of Leiden, The Netherlands) who had been a pioneer on attempts to isolate *Frankia*, on the occasion of his retirement. A large part of the proceedings was devoted to create groups among the increasing number of strains (Lechevalier 1984) with the development of physiology-based tools (Horriere 1984; Lechevalier and Ruan 1984). The diversity of strains was analysed regarding efficiency (Sellstedt and Huss-Danell 1984). Questions on the physiology of the interaction also surfaced with the demonstration of the synthesis of auxin by *Frankia* (Wheeler *et al.* 1984).

The fifth meeting took place in Quebec City in Canada in August 1984, it was organized by M Lalonde, C Camiré and JO Dawson and the proceedings were published in *Plant & Soil* in 1985 (Dawson *et al.* 1985). The event reunited 86 participants, including 25 from Canada, 4 from China, 1 from Finland, 8 from France, 1 from the Netherlands and 3 from Spain. Reported work concerned grouping of strains based on DNA (An *et al.* 1985; Simonet *et al.* 1985) and attempts to increase the scale of inoculated seedlings (Périnet *et al.* 1985; Stowers and Smith 1985) with an aim to commercial or large-scale applications.

The sixth meeting took place in Umeå in Sweden in August 1986 organized by K Huss-Danell, A-S Hahlin, A Sellstedt, KR Sundström and P-A Vikman, and the proceedings were published in *Physiologia Plantarum* in 1987 (Huss-Danell and Wheeler 1987). The event reunited 55 participants from 13 countries. Reported work concerned groupings of strains based on various approaches such as host specificity (Baker 1987), or total proteins (Gardes and Lalonde 1987), developments in physiology such as hydrogen production (Sellstedt and Winship 1987) or discussion on sporulation of *Frankia* in vitro and in nodules (Torrey 1987). There was also a very much discussed presentation on protoplast fusion between *Streptomyces* and *Frankia*, that was published elsewhere (Prakash and Cummings 1988), which raised the possibility of genetic work but this result unfortunately could not be reproduced and the putative fusant strains obtained were never transmitted to other laboratories for independent testing.

The seventh meeting took place in Storrs, CT, in August 1988 organized by DR Benson and LJ Winship (Benson and Winship 1989), and the proceedings were published in *Plant & Soil* in 1989. The event reunited about 100 participants. Reported work concerned ultrastructure, with among other findings that multiple lipid monolayers served to resist oxygen penetration into the nitrogenase-rich *Frankia* vesicle interior, lipids that were later shown to be hopanoids (Berry *et al.* 1993). Molecular ecology was expanding with the use of synthetic oligonucleotide probes that were developed to identify *Frankia* strains (Hahn *et al.* 1989). There were also applied forestry work such as alder-walnut interplanting assays (Paschke *et al.* 1989) or a study on the influence of mycorrhizae on symbiotic performance of alder (Chatarpaul *et al.* 1989).

The eighth meeting took place in Lyon, France in September 1991 organized by P Normand, MP Fernandez, P Simonet and AM Domenach, and the proceedings were published in *Acta Oecologica* in 1992 (Normand *et al.* 1992). The event reunited about 80 participants from 18 countries and was the occasion to

honor 4 famous retirees, Yvon Dommergues (ORSTOM, France), Mary P Lechevalier and Hubert P Lechevalier (Waksman Institute, Rutgers University, NJ) and John G Torrey (Harvard University, MA). Reported work concerned the demonstration that *Frankia* could thrive in non-host rhizospheres (Paschke and Dawson 1992), attempts at electroporation (Courmoyer and Normand 1992), and applied work to select salt-tolerant *Casuarina* provenances (Girgis *et al.* 1992).

The ninth meeting took place in Waikato, New Zealand in September 1993 organized by W Silvester and S Harris and the proceedings were published in *Soil Biology and Biochemistry* in 1994 (Silvester and Harris 1994) with a tribute to the recently deceased John G. Torrey (Baker and Berry 1994). The meeting saw various works such as the immunolocalization of nitrogenase and hydrogenase to vesicles (Sellstedt and Mattsson 1994), siderophore synthesis (Aronson and Boyer 1994) or more applied ones such as the use of alder to revegetate polluted mine spoils (Lumini *et al.* 1994).

The tenth meeting took place in Davis, CA in August 1995 organized by AM Berry and DD Myrold and the proceedings were published in *Physiologia Plantarum* in 1997 (Berry and Myrold 1997). The meeting saw various work such as most prominently the characterization of several host plant nodulin genes (Gherbi *et al.* 1997; Guan *et al.* 1997; Pawlowski 1997) after the previous publication of the upregulated *Alnus* protease (Goetting-Minesky and Mullin 1994; Ribeiro *et al.* 1995), the beginning of phylogenetic work on actinorhizal plants (Swensen and Mullin 1997) and on the root hair deforming factor synthesized by *Frankia* (Van Ghelue *et al.* 1997) and compounds synthesized by the plant that modified the physiology of *Frankia* (Benoit and Berry 1997). There was also a rising perception of the similarities between the actinorhizal and the legume symbioses, with an emphasis on the need to use a common language, for instance replacing 'atypical' by Nod<sup>-</sup> or 'encapsulation' by infection thread (Akkermans and Hirsch 1997).

The eleventh meeting took place in Urbana, IL in August 1997 organized by JO Dawson and the proceedings were published in the *Canadian Journal of Botany* in 1999 (Dawson 1999). There were 45 attendees from 12 countries. The meeting saw several work on molecular ecology (Clawson *et al.* 1999; Jeong and Myrold 1999; Lumini and Bosco 1999; Ritchie and Myrold 1999), as well as ultrastructural work detailing the *Datisca-Coriaria* nodule type and arguing for the existence of infection threads (Berg 1999a, b; Berg *et al.* 1999) and classical *Frankia* physiology such as a study of the antibiotic resistance pattern of strains (Tisa *et al.* 1999).

The twelfth meeting took place in Carry-le-Rouet, France in June 2001 organized by P Normand, N Alloisio, AM Domenach, I Navarro and MP Fernandez, with the proceedings published in *Plant & Soil* in 2003 with JO Dawson and K Pawlowski as co-editors (Normand 2003). There were 80 attendees from 14 countries. The meeting saw work on a new antibiotic from *Frankia*, frankiamide (Haansuu *et al.* 2001), synthesis *in vitro* of the auxin PAA by *Frankia* (Hammad *et al.* 2003), germination of *Frankia* spores (Krumholz *et al.* 2003) as well as the use of  $\delta^{15}\text{N}$  to monitor nitrogen fluxes in Glacier Bay (Kohls *et al.* 1994) and evolutionary significance of host and *Frankia* DNA sequences (Varghese *et al.* 2003).

The thirteenth meeting took place in Durham, NH, in June 2005 organized by LS Tisa, with the proceedings published in the *Symbiosis* journal in 2005 (Tisa 2005). The meeting saw the first presentation of the genomes that had been initiated at the Genoscope and at the JGI the year before and would only be published two years later (Normand *et al.* 2007). There was also a presentation of work on the physiology and genes coding hemoglobins of *Frankia* (Niemann *et al.* 2005; Schwintzer and Tjepkema 2005).

The fourteenth meeting took place in Umeå, Sweden, in July 2006 organized by A Sellstedt, which proceedings were published in *Physiologia Plantarum* in 2007 with P Normand and JO Dawson as co-editors (Sellstedt *et al.* 2007). Among the works reported, there was a demonstration that a defense-related chitinase-encoding gene (*cgchi3*) was specifically activated in nodules as compared with uninoculated control roots (Fortunato *et al.* 2007), that coping with reactive oxygen species was critical (Tavares *et al.* 2007) and a first proteomic approach on *Frankia* (Alloisio *et al.* 2007). There was a workshop organized to get to work on the web site housing the three *Frankia* genomes (<https://www.genoscope.cns.fr/agc/microscope/home/index.php>) available at the time, which would eventually result in many in the field exploiting the genomes data in various ways.

The fifteenth meeting took place in Bariloche, Argentina in October 2008 organized by LG Wall, L Gabbarini, L Imanishi, E Chaia, M Solans and G Vobis, which proceedings were published in *Symbiosis* in 2010 with LG Wall, E Chaia and JO Dawson as co-editors (Wall *et al.* 2010). There were 34 attendees from 9 countries. The special *Symbiosis* issue was dedicated to the memory of YR Dommergues, from the

overseas French Research Institute (ORSTOM, then IRD) who had been a major contributor to actinorhizal biology, and in particular an indefatigable promoter of *Casuarina* fundamental research and applied uses such as a 500km × 500m filao plantation in Sénégal to supply firewood and stabilize the seafront sand dune. Among the works reported, there was a functional proteomics of *Frankia* from field nodules (Mastrorunzio and Benson 2010), the exploration of diversity of *Frankia* and non-*Frankia* actinobacteria in nodules (Ghodhbane-Gtari *et al.* 2010), work on genetic transformation of host plants (Svistoonoff *et al.* 2010) that permitted to show the presence of a Sym kinase governing symbiosis establishment in a manner similar to what happens in Legumes (Gherbi *et al.* 2008), and an evaluation of the contribution of actinorhizal plants to increased soil N fertility (Chaia and Myrold 2010).

The sixteenth meeting took place in Porto, Portugal, in September 2010, organized by A Ribeiro, C Santos, F Tavares and P Santos. The proceedings were published in two separate journals, *Archives of Microbiology* for the microbial papers (Santos and Tavares 2012) and in *Functional Plant Biology* for the plant papers (Ribeiro *et al.* 2011). There were 30 attendees from 9 countries. Among the reported works were transcriptomics EST studies that yielded a global view of host plants response to *Frankia* (Berry *et al.* 2011; Hocher *et al.* 2011), exploration of actinorhizal fruits for their medicinal properties (Goyal *et al.* 2011), taxonomic resolution of *Myrica* sp. (Yanthan *et al.* 2011) and an exploration of the reaction of actinorhizal plants to global change (Tobita *et al.* 2011). On the bacterial side, there was a discussion of the controversial presence of nitrogenase in other actinobacteria (Gtari *et al.* 2012), or the development of a high-throughput system to study the physiology of *Frankia* (Furnholm *et al.* 2012).

The seventeenth meeting, initially due to take place in Tunisia had to be relocated due to the turbulences this country and its neighbors undergo in the wake of the ‘Arab Spring’. Dr Arvind K Misra was kind enough to substitute and set up the meeting in April 2013 in Shillong, India, for which his colleagues are all grateful.

### 3. Conclusion

The visibility of research on *Frankia* and actinorhizal plants, and hence the funding we get depends on several factors, prominent among which are these meetings and publications in high profile journals. A Web of Science search retrieved with keyword ‘Frankia’ 1 paper in Nature, 1 paper in Science and 5 in PNAS-USA, while keyword ‘Rhizobium’ retrieved 91 papers in Nature, 35 in Science, and 234 in PNAS-USA. This difference is in large part due to the absence of a genetic system in *Frankia*, to the fact so many of our strains have not been given a species name but not only. As a community, we must try again and again, using the new ‘-omics’ approaches but also more classic approaches such as highly resolutive spectrometry and of course the actinorhizal host plants transformation methodology that has been so fruitful so far. And most importantly, we must keep interacting together using one of the tools at our disposal, the *Frankia* and actinorhizal plants meetings.

### References

- Akkermans ADL, Baker DD, Huss-Danell K and Tjepkema JD 1984 Preface. *Plant Soil* **78** ix–x
- Akkermans ADL and Hirsch AM 1997 A reconsideration of terminology in *Frankia* research: a need for congruence. *Physiol. Plant.* **99** 574–578
- Alloisio N, Félix S, Maréchal J, Pujic P, Rouy Z, Vallenet D, Medigue C and Normand P 2007 *Frankia alni* proteome under nitrogen-fixing and nitrogen-replete conditions. *Physiol. Plant.* **13** 440–453
- An C, Riggsby W and Mullin B 1985 Restriction pattern analysis of genomic DNA of *Frankia* isolates. *Plant Soil* **87** 43–48
- Aronson D and Boyer G 1994 Growth and siderophore formation in six iron limited strains of *Frankia*. *Soil Biol. Biochem.* **26** 561–567
- Baker D 1987 Relationships among pure-cultured strains of *Frankia* based on host specificity. *Physiol. Plant.* **70** 245–248
- Baker D, Kidd G and Torrey JG 1979 Separation of actinomycete nodule endophytes from crushed nodule suspensions by Sephadex fractionation. *Bot. Gaz.* **140S** S49–S51

- Baker D and Torrey J 1979 The isolation and cultivation of actinomycetous root nodule endophytes; in *Symbiotic nitrogen fixation in the management of temperate forests* (eds) JC Gordon, CT Wheeler, DA Perry and OR Corvallis (Oregon State University: Forest Research Laboratory) pp 38–56
- Baker DD and Berry A 1994 A tribute to John G. TORREY 1921–1993. *Soil Biol. Biochem.* **26** vii–viii
- Beijerinck MW 1888 Die Bacterien der Papilionaceen-Knöllchen. *Bot. Zeitung* **46** 725–735
- Benoit LF and Berry AM 1997 Flavonoid-like compounds from seeds of red alder (*Alnus rubra*) influence host nodulation by *Frankia* (Actinomycetales). *Physiol. Plant.* **99** 588–593
- Benson D and Eveleigh D 1979 Ultrastructure of the nitrogen-fixing symbiont of *Myrica pensylvanica* L. (bayberry) root nodules. *Bot. Gaz.* **140S** S15–S21
- Benson DR and Winship LJ 1989 Preface. *Plant Soil* **181** viii–ix
- Berg RH 1999a Cytoplasmic bridge formation in the nodule apex of actinorhizal root nodules. *Can. J. Bot.* **77** 1351–1357
- Berg RH 1999b *Frankia* forms infection threads. *Can. J. Bot.* **77** 1327–1333
- Berg RH, Langenstein B and Silvester WB 1999 Development in the *Datisca-Coriaria* nodule type. *Can. J. Bot.* **77** 1334–1350
- Berry AM, Harriott OT, Moreau RA, Osman SF, Benson DR and Jones AD 1993 Hopanoid lipids compose the *Frankia* vesicle envelope, presumptive barrier of oxygen diffusion to nitrogenase. *Proc. Nat. Acad. Sci. USA* **90** 6091–6094
- Berry AM, Mendoza-Herrera A, Guo Y-Y, Hayashi J, Persson T, Barabote RD, Demchenko K, Zhang S and Pawlowski K 2011 New perspectives on nodule nitrogen assimilation in actinorhizal symbioses. *Funct. Plant Biol.* **38** 645–652
- Berry AM and Myrold DD 1997 Proceedings of the tenth international conference on *Frankia* and actinorhizal plants. *Physiol. Plant.* **99** 564
- Brunchorst J 1886 Über einige Wurzelanschwellungen, besonders diejenigen von *Alnus*, und den Elaeagnaceen. *Unters. bot. Inst. Tübingen* **2** 151–177
- Callaham D, Del Tredici P and Torrey J 1978 Isolation and cultivation *in vitro* of the actinomycete causing root nodulation in *Comptonia*. *Science* **199** 899–902
- Chaia EE and Myrold DD 2010 Variation of <sup>15</sup>N natural abundance in leaves and nodules of actinorhizal shrubs in Northwest Patagonia. *Symbiosis* **50** 97–105
- Chatarpaul L, Chakravarty P and Subramaniam P 1989 Studies in tetrapartite symbioses. I. Role of ecto- and endomycorrhizal fungi and *Frankia* on the growth performance of *Alnus incana*. *Plant Soil* **118** 145–150
- Clawson M, Gawronski J and Benson DR 1999 Dominance of *Frankia* strains in stands of *Alnus incana* subsp. *rugosa* and *Myrica pensylvanica*. *Can. J. Bot.* **77** 1203–1207
- Cournoyer B and Normand P 1992 Electroporabilization of *Frankia* intact cells to plasmid DNA. *Acta Oecologica* **13** 369–378
- Dawson JO 1999 Foreword. *Can. J. Bot.* **77**. doi:10.1139/cjb7709foreword
- Dawson JO, Camire C and Lalonde M 1985 Preface. *Plant Soil* **87** xi–xii
- Diem HG, Gauthier D and Dommergues Y 1983 An effective strain of *Frankia* from *Casuarina* sp. *Can. J. Bot.* **61** 2815–2821
- Fortunato A, Santos P, Gracxa I, Gouveia M, Martins S, Ricardo C, Pawloski K and Ribeiro A 2007 Isolation and characterization of cgchi3, a nodule-specific gene from *Casuarina glauca* encoding a class III chitinase. *Physiol. Plant.* **130** 418–426
- Furnholm T, Beauchemin N and Tisa LS 2012 Development of a semi-high-throughput growth assay for the filamentous actinobacteria *Frankia*. *Arch. Microbiol.* **194** 13–20
- Gardes M and Lalonde M 1987 Identification and subgrouping of *Frankia* strains using sodium dodecyl sulfate-polyacrylamide gel electrophoresis. *Physiol. Plant.* **70** 237–244
- Gauthier D, Diem H and Dommergues Y 1981 *In vitro* nitrogen fixation by two actinomycete strains isolated from *Casuarina* nodules. *Appl. Environ. Microbiol.* **41** 306–308
- Gherbi H, Duhoux E, Franche C, Pawlowski K, Nasser A, Berry AM and Bogusz D 1997 Cloning of a full-length symbiotic hemoglobin cDNA and *in situ* localization of the corresponding mRNA in *Casuarina glauca* root nodule. *Physiol. Plant.* **99** 608–616
- Gherbi H, Markmann K, Svistoonoff S, Estevan J, Autran D, Giczey G, Auguy F, Peret B, Laplaze L, Franche C, Parniske M and Bogusz D 2008 SymRK defines a common genetic basis for plant root endosymbioses with arbuscular mycorrhiza fungi, rhizobia, and *Frankia* bacteria. *Proc. Natl. Acad. Sci. USA* **105** 4928–4932
- Ghodhbane-Gtari F, Essoussi I, Chattaoui M and Chouaia B 2010 Isolation and characterization of non-*Frankia* actinobacteria from root nodules of *Alnus glutinosa*, *Casuarina glauca* and *Elaeagnus angustifolia*. *Symbiosis* **50** 51–57
- Girgis M, Ishac Y, Diem H and Dommergues Y 1992 Selection of salt tolerant *Casuarina glauca* and *Frankia*. *Acta Oecologica* **13** 443–451

- Goetting-Minesky MP and Mullin BC 1994 Differential gene expression in an actinorhizal symbiosis evidence for a nodule-specific cysteine proteinase. *Proc. Natl. Acad. Sci. USA* **91** 9891–9895
- Gordon JC and Dawson JO 1979 Potential uses of nitrogen-fixing trees and shrubs in commercial forestry. *Bot. Gaz.* **140S** S88–S90
- Gordon JC, Wheeler CT and Perry DA 1979 Introduction; in *Symbiotic nitrogen fixation in the management of temperate forests* (eds) JC Gordon, CT Wheeler, DA Perry and OR Corvallis (Oregon State University: Forest Research Laboratory) p 1
- Goyal AK, Basistha BC, Sen A and Middha SK 2011 Antioxidant profiling of *Hippophae salicifolia* growing in sacred forests of Sikkim, India. *Funct. Plant Biol.* **138** 697–701
- Gtari M, Ghodhbane-Gtari F, Nouioui I, Beauchemin N and Tisa LS 2012 Phylogenetic perspectives of nitrogen-fixing actinobacteria. *Arch. Microbiol.* **194** 3–11
- Guan C, Akkermans ADL, van Kammen A, Bisseling T and Pawlowski K 1997 ag13 is expressed in *Alnus glutinosa* nodules in infected cells during endosymbiont degradation in the nodule pericycle. *Physiol. Plant.* **99** 601–607
- Haansuu JP, Klika KD, Soderholm PP, Ovcharenko VV, Pihlaja K, Haahtela KK and Vuorela PM 2001 Isolation and biological activity of frankiamide. *J. Ind. Microbiol. Biotechnol.* **27** 62–66
- Hahn D, Dorsch M, Stackebrandt E and Akkermans ADL 1989 Synthetic oligonucleotide probes for identification of *Frankia* strains. *Plant Soil* **118** 211–219
- Hammad Y, Nalin R, Marechal J, Fiasson K, Pepin R, Berry AM, Normand P and Domenach A-M 2003 A possible role for phenylacetic acid (PAA) in *Alnus glutinosa* nodulation by *Frankia*. *Plant Soil* **254** 193–205
- Hocher V, Alloisio N, Auguy F, Fournier P, Doumas P, Pujic P, Gherbi H, Queiroux C, Da Silva C, Wincker P, Normand P and Bogusz D 2011 Transcriptomics of actinorhizal symbioses reveals homologs of the whole common symbiotic signaling cascade. *Plant Physiol.* **156** 1–12
- Horriere F 1984 In vitro physiological approach to classification of *Frankia* isolates of 'the *Alnus* group' based on urease, protease and  $\beta$ -glucosidase activities. *Plant Soil* **78** 7–13
- Huss-Danell K and Wheeler CT 1987 *Frankia* and actinorhizal plants. *Physiol. Plant.* **70** 235
- Jeong S-C and Myrold DD 1999 Genomic fingerprinting of *Frankia* microsymbionts from *Ceanothus* copopulations using repetitive sequences and polymerase chain reactions. *Can. J. Bot.* **77** 1220–1230
- Klemmedson JO 1979 Ecological importance of actinomycete-nodulated plants in the western United States. *Bot. Gaz.* **140S** S91–S96
- Kohls S, van Kessel C, Baker D, Grigal D and Lawrence D 1994 Assessment of  $N_2$  fixation and N cycling by *Dryas* along a chronosequence within the forelands of the Athabasca Glacier, Canada. *Soil Biol. Biochem.* **26** 623–632
- Krumholz GD, Chval MS, McBride MJ and Tisa LS 2003 Germination and physiological properties of *Frankia* spores. *Plant Soil* **254** 57–67
- Lalonde M 1979 Immunological and ultrastructural demonstration of nodulation of the European *Alnus glutinosa* (L.) Gaertn. host plant by an actinomycetal isolate from the North American *Comptonia peregrina* (L.) Coult. root nodule. *Bot. Gaz.* **140S** S35–S43
- Lalonde M and Calvert H 1979 Production of *Frankia* hyphae and spores as an infective inoculant for *Alnus* species; in *Symbiotic nitrogen fixation in the management of temperate forests* (eds) JC Gordon, CT Wheeler, DA Perry and OR Corvallis (Oregon State University: Forest Research Laboratory) pp 95–110
- Lechevalier M 1984 The taxonomy of the genus *Frankia*. *Plant Soil* **78** 1–6
- Lechevalier M and Ruan J 1984 Physiology and chemical diversity of *Frankia* spp. isolated from nodules of *Comptonia peregrina* (L.) Coult. and *Ceanothus americanus* L. *Plant Soil* **78** 15–22
- Lumini E and Bosco M 1999 Polymerase chain reaction - restriction fragment length polymorphisms for assessing and increasing biodiversity of *Frankia* culture collections. *Can. J. Bot.* **77** 1261–1269
- Lumini E, Bosco M, Puppi G, Isopi R, Frattegiani M, Buresti E and Favilli F 1994 Field performance of *Alnus cordata* Loisel (italian alder) inoculated with *Frankia* and VA-mycorrhizal strains in mine spoil afforestation plots. *Soil Biol. Biochem.* **26** 659–661
- Mastrorunzio JE and Benson DR 2010 Wild nodules can be broken: proteomics of *Frankia* in field-collected root nodules. *Symbiosis* **50** 13–26
- Niemann JM, Tjepkema JD and Tisa LS 2005 Identification of the truncated hemoglobin gene in *Frankia*. *Symbiosis* **39** 83–90
- Normand P 2003 Introduction. *Plant Soil* **254** vii
- Normand P, Fernandez MP, Simonet P and Domenach AM 1992 Introduction to the proceedings of the 8<sup>th</sup> *Frankia* and Actinorhizal Plants congress. *Acta Oecologica* **13** 367–368
- Normand P, Lapierre P, Tisa LS, Gogarten JP, Alloisio N, Bagnarol E, Bassi CA, Berry AM, et al. 2007 Genome characteristics of facultatively symbiotic *Frankia* sp. strains reflect host range and host plant biogeography. *Genome Res.* **17** 7–15
- Paschke M, Dawson J and David M 1989 Soil nitrogen mineralization in plantations of *Juglans nigra* interplanted with actinorhizal *Elaeagnus umbellata* or *Alnus glutinosa*. *Plant Soil* **118** 33–42

- Paschke MW and Dawson JO 1992 *Frankia* abundance in soils beneath *Betula nigra* and other non-actinorhizal woody plants. *Acta Oecologica* **13** 407–416
- Pawlowski K 1997 Nodule-specific gene expression. *Physiol. Plant.* **99** 617–631
- Périnet P, Brouillette J, Fortin J and Lalonde M 1985 Large scale inoculations of actinorhizal plants with *Frankia*. *Plant Soil* **87** 175–183
- Pommer E 1956 Beiträge zur Anatomie und Biologie der Wurzelknöllchen von *Alnus glutinosa* Gaertn. *Flora* **14** 603–634
- Pommer E 1959 Über die Isolierung des Endophyten aus den Wurzelknöllchen *Alnus glutinosa* Gaertn. und über erfolgreiche Re-Infektionsversuche. *Ber. Deutsch Botan. Gesell.* **72** 138–150
- Prakash R and Cummings B 1988 Creation of novel nitrogen-fixing actinomycetes by protoplast fusion of *Frankia* and *Streptomyces*. *Plant Mol. Biol.* **10** 281–289
- Resch H 1980 Utilization of Red Alder in the pacific Northwest. *For. Prod. J.* **30** 21–26
- Ribeiro A, Akkermans ADL, van Kammen A, Bisseling T and Pawlowski K 1995 A nodule-specific gene encoding a subtilisin-like protease is expressed in early stages of actinorhizal nodule development. *Plant Cell* **7** 785–794
- Ribeiro A, Berry AM, Pawlowski K and Santos P 2011 Actinorhizal plants. *Funct. Plant Biol.* **38** v–vii
- Ritchie NJ and Myrold DD 1999 Phylogenetic placement of uncultured *Ceanothus* microsymbionts using 16S rRNA gene sequences. *Can. J. Bot.* **77** 1208–1213
- Santos CL and Tavares F 2012 A step further on *Frankia* biology. *Arch. Microbiol.* **194** 1–2
- Schwintzer CR and Tjepkema JD 2005 Effect of oxygen concentration on growth and hemoglobin production in *Frankia*. *Symbiosis* **39** 77–82
- Sellstedt A and Huss-Danell K 1984 Nitrogen fixation and relative efficiency of nitrogenase in *Alnus incana* grown in different cultivation systems. *Plant Soil* **78** 147–158
- Sellstedt A and Mattsson U 1994 Hydrogen metabolism in *Casuarina* *Frankia* immunolocalization of nitrogenase and hydrogenase. *Soil Biol. Biochem.* **26** 583–592
- Sellstedt A, Normand P and Dawson JO 2007 *Frankia* – the friendly bacteria – infecting actinorhizal plants. *Physiol. Plant.* **130** 315–317
- Sellstedt A and Winship L 1987 Hydrogen metabolism of *Casuarina* root nodules: A comparison of two inoculum sources. *Physiol. Plant* **70** 367–372
- Silvester W and Harris S 1994 Preface. *Soil Biol. Biochem.* **26** v
- Simonet P, Normand P, Moiroud A and Lalonde M 1985 Restriction enzyme digestion patterns of *Frankia* plasmids. *Plant Soil* **87** 49–60
- Stowers M and Smith J 1985 Inoculation and production of container-grown red alder seedlings. *Plant Soil* **87** 153–160
- Svistoonoff S, Gherbi H, Nambiar-Veetil M and Zhong C 2010 Contribution of transgenic Casuarinaceae to our knowledge of the actinorhizal symbioses. *Symbiosis* **50** 3–11
- Swensen SM and Mullin BC 1997 Phylogenetic relationships among actinorhizal plants. The impact of molecular systematics and implications for the evolution of actinorhizal symbioses. *Physiol. Plant.* **99** 565–573
- Tavares F, Santos CL and Sellstedt A 2007 Reactive oxygen species in legume and actinorhizal nitrogen-fixing symbioses: the microsymbiont's responses to an unfriendly reception. *Physiol. Plant.* **130** 344–356
- Tisa L, McBride M and Ensign JC 1983 Studies of growth and morphology of *Frankia* strains EAN1pec, Eu1c, Cp11, and ACN1AG. *Can. J. Bot.* **61** 2768–2773
- Tisa LS 2005 Preface. *Symbiosis* **39** 59
- Tisa LS, Chval MS, Krumholz GD and Richards J 1999 Antibiotic resistance patterns of *Frankia* strains. *Can. J. Bot.* **77** 1257–1260
- Tjepkema J, Ormerod W and Torrey JG 1980 Vesicle formation and acetylene reduction activity in *Frankia* sp. Cp11 cultured in defined nutrient media. *Nature* **287** 633–635
- Tobita H, Uemura A, Kitao M, Kitaoka S, Maruyama Y and Utsugi H 2011 Effects of elevated atmospheric carbon dioxide, soil nutrients and water conditions on photosynthetic and growth responses of *Alnus hirsuta*. *Funct. Plant Biol.* **138** 702–710
- Torrey JG 1987 Endophyte sporulation in root nodules of actinorhizal plants. *Physiol. Plant.* **70** 279–288
- Torrey JG and Tjepkema JD 1979 Symbiotic nitrogen fixation in actinomycete-nodulated plants. Preface and program. *Bot. Gaz.* **140S** Si–Sv
- Torrey JG and Tjepkema JD 1983 International conference on the biology of *Frankia*. Introduction. *Can. J. Bot.* **61** 2765–2767
- Varghese R, Chauhan VS and Misra AK 2003. Evolutionary implications of nucleotide sequence relatedness between *Alnus nepalensis* and *Alnus glutinosa* and also between corresponding *Frankia* microsymbionts. *Plant Soil* **254** 219–227
- Van Ghelue M, Lovaas E, Ringo E and Solheim B 1997 Early interactions between *Alnus glutinosa* and *Frankia* strain Ar13. Production and specificity of root hair deformation factor(s). *Physiol. Plant.* **99** 579–587

- Wall LG, Chaia E and Dawson JO 2010 Special Volume devoted to the 15th International *Frankia* and Actinorhizal Plant Meeting. *Symbiosis* **50** 1–2
- Wheeler CT, Crozier A and Sandberg G 1984 The biosynthesis of indole-3-acetic acid by *Frankia*. *Plant Soil* **78** 99–104
- Winship L and Chaudhary A 1979 Nitrogen fixation by *Datisca glomerata* : a new addition to the list of actinorhizal diazotrophic plants; in *Symbiotic nitrogen fixation in the management of temperate forests* (eds) JC Gordon, CT Wheeler, DA Perry and OR Corvallis (Oregon State University: Forest Research Laboratory) p 485
- Yanthan M, Biate D and Misra AK 2011 Taxonomic resolution of actinorhizal *Myrica* species from Meghalaya (India) through nuclear rDNA sequence analyses. *Funct. Plant Biol.* **38** 738–746

PHILIPPE NORMAND

*Université Lyon 1, Université de Lyon, CNRS, Ecologie Microbienne UMR5557*

*Villeurbanne 69622 cedex, France*

*(Email, Philippe.Normand@univ-lyon1.fr)*