Mechanisms of HSP72 release

ALEXZANDER ASEA

Division of Investigative Pathology, Scott & White Clinic and Texas A&M University System Health Science Center College of Medicine, 2401 South 31st Street, Temple, TX 76508, USA

(Fax, 1 (254) 743 0247; Email, asea@medicine.tamhsc.edu)

Currently two mechanisms are recognized by which heat shock proteins (HSP) are released from cells; a passive release mechanism, including necrotic cell death, severe blunt trauma, surgery and following infection with lytic viruses, and an active release mechanism which involves the non classical protein release pathway. HSPs are released both as free HSP and within exosomes. This review covers recent findings on the mechanism by which stress induces the release of HSP72 into the circulation and the biological significance of circulating HSP72 to host defense against disease.

[Asea A 2007 Mechanisms of HSP72 release; *J. Biosci*. **32** 579–584]

1. Introduction

Heat shock proteins (HSP) were originally described for their roles as chaperones induced by temperature shock as well as various other kinds of stress including environmental (UV radiation, heat shock, heavy metals and amino acids), pathological (bacterial, parasitic infections or fever, inflammation, malignancy or autoimmunity) or physiological stresses (growth factors, cell differentiation, hormonal stimulation or tissue development), that induced a marked increase in intracellular HSP (iHSP) synthesis known as the stress response (Lindquist and Craig 1988). This is achieved by activating the trimerization and nuclear translocation of cytoplasmic heat shock factor-1 (HSF-1) for binding with the heat shock elements (HSE) and consequent transcription of *HSP* genes. By binding unfolded, misfolded or mutated peptides or proteins and transporting them to the endoplasmic reticulum (ER), HSP prevent potential aggregation and/or death. Within the ER the peptides are released in an ATP-dependent fashion and refolded (Fink 1999). Recently an additional role has been ascribed to HSP as danger signals produced and released when cells are under stress and as activators of the immune system. This review will focus mainly on the HSP70 family, which constitutes the most conserved and best studied class of all heat shock proteins. The human HSP70 family consists of at least 12 members (for review *see* Tavaria *et al* 1996). The best known HSP70 family members are the constitutively expressed HSP70 (Hsc70 or HSP73; molecular weight of 73 kDa), the stress inducible HSP70 (HSP70 or HSP72; molecular weight of 72 kDa), the mitochondrial HSP70 (HSP75; molecular weight of 75 kDa), and the endoplasmic reticulum HSP70 (Grp78; molecular weight of 78 kDa). This review will briefly delve on recent advances in our understanding of how stress induces the release of HSP72 into the circulation. I will also discuss the steps required for initiation of an immune response and its biological significance in the context of host defense.

2. Chaperokine activity of extracellular HSP72

Chaperokine, is a term recently coined to better describe the unique function of extracellular HSP72 (eHSP72) as both

Keywords. Cancer; Chaperokine; heat shock proteins; inflammation; receptors, signal transduction

Abbreviations used: APC, antigen presenting cells; eHSP72, extracellular HSP72; Hsc73, constitutive seventy-kilo Dalton heat shock protein; HSP72, inducible seventy-kilo Dalton heat shock protein; IFN-*γ*, interferon-gamma; iHSP72, intracellular HSP72; TLR, toll-like receptors

chaperone and cytokine (Asea 2003 2005). The consequence of binding and signaling is the stimulation of a potent and long lasting immune response. eHSP72 induces a plethora of immune responses and the list continues to grow. Briefly, as early as 2–4 h post exposure of APC to eHSP72, there is a significant release of cytokines including TNF-*α*, IL-1*β*, IL-6 and IL-12 (Asea *et al* 2000a,b), GM-CSF (Srivastava 2002); nitric oxide, a potent apotogenic mediator (Panjwani *et al* 2002); chemokines including MIP-1, MCP-1 and RANTES (Lehner *et al* 2000; Panjwani *et al* 2002). This part of the immune response does not require peptide, since both peptide-bearing and non peptide-bearing eHSP72 is capable of inducing pro-inflammatory cytokine production by APCs (Asea *et al* 2000a). However, peptide is required for specific CD8⁺ CTL responses (Srivastava et al 1994; Srivastava 2000, 2005). eHSP72 induces the DC maturation by augmenting the surface expression of CD40, CD83, CD86 and MHC class II molecules on DC (Basu *et al* 2000; Singh-Jasuja *et al* 2000; Asea *et al* 2002a; Noessner *et al* 2002) and migration of DC (Binder *et al* 2000) and NK cells (Gastpar *et al* 2005).

2.1 *Dichotomy of HSP70 effects*

A dichotomy now exists between the effects of HSP based on its relative location, intracellular versus extracellular, and the target cell it binds to and activates. The upregulation of intracellular HSP70 (iHSP72) is generally cytoprotective and induces the cell's anti-apoptotic mechanisms (Jaattela *et al* 1998), represses gene expression (Tang *et al* 2001), modulates cell cycle progression (Hut *et al* 2005) and is antiinflammatory (Housby *et al* 1999). On the other hand, the upregulation of extracellular HSP70 (eHSP72) is generally immuno-stimulatory and stimulates pro-inflammatory cytokine synthesis (Asea *et al* 2000c; Asea 2005), augments chemokine synthesis (Lehner *et al* 2000; Panjwani *et al* 2002), upregulates co-stimulatory molecules (Asea *et al* 2002b; Bausero *et al* 2005b) and enhances anti-tumour surveillance (Srivastava *et al* 1994; Srivastava 2000, 2005).

3. Passive release mechanism

Gallucci and colleagues initially demonstrated that dendritic cells (DC) are stimulated by endogenous signals received from stressed, viral-infected or necrosis-induced cells, but not by healthy cells or cells undergoing apoptosis (Gallucci *et al* 1999). In a series of elegantly performed experiments, Basu and co-workers later reported that heat shock proteins including gp96, calreticulin, HSP90 and HSP72 are released from cells by necrotic but not apoptotic cells (Basu *et al* 2000). These authors demonstrated that necrosis induced by freeze thaw, but not apoptosis induced by irradiation, resulted in the release of HSP into the culture supernatant

(Basu *et al* 2000; Basu and Srivastava 2000; Srivastava and Amato 2001; Srivastava 2003). During apoptotic cell death, the contents of the cell are not released into the external milieu but are packaged neatly into apoptotic bodies, which are efficiently scavenged by neighbouring professional phagocytes. However, necrotic cell death results in the discharge of intracellular contents into the extracellular milieu thereby liberating heat shock proteins (for review *see* Srivastava 2003; Calderwood 2005). These results make the necrosis hypothesis an attractive explanation for the mechanism by which heat shock proteins are released into the circulation.

A condition in which necrotic cell death clearly contributes to the release of HSP72 from cells is following severe trauma. Pittet and colleagues (Pittet *et al* 2002) found a significant upregulation in circulating serum HSP72 in severely traumatized patients as early as 30 minutes after injury. Increased circulating serum HSP72 has also been measured in patients after coronary artery bypass grafting (Dybdahl *et al* 2002; Dybdahl *et al* 2004). Importantly, circulating serum HSP72 has been suggested as a marker of myocardial damage, and reported to have a role in the inflammatory response after acute myocardial infarction (AMI) (Dybdahl *et al* 2005). Other conditions in which elevated levels of circulating serum HSP72 has been demonstrated are renal disease (Wright *et al* 2000), hypertension (Pockley *et al* 2002), atherosclerosis (Pockley *et al* 2003), aging (Terry *et al* 2004), and sickle cell disease (Adewoye *et al* 2005). However, in these conditions although necrosis is proposed as the mechanism of release, conclusive experimental data is still lacking.

A more conclusive study was designed in which *in situ* killing of tumour cells using suicide gene transfer to generate death by a non-apoptotic pathway was shown to be associated with high immunogenicity and induction of HSP (Melcher *et al* 1998). The most conclusive demonstration that necrosis accounts for HSP release is seen following infection with lytic viruses. In a study by Moehler and coworkers, it was demonstrated that parvovirus-mediated cell killing enhances tumour immunogenicity by HSP72 release and contributes to the anti-tumour effect of parvoviruses (Moehler *et al* 2005). Although these authors did not directly demonstrate that H1-induced cell killing and its associated HSP72 release promotes the loading and maturation of antigen presenting cells and by extension triggers tumour-specific immune responses, one can speculate that the release of HSP72 could facilitate priming of T cells specific for viral antigens.

Taken together, the passive release clearly is an important mechanism by which HSP72 is released into the circulation. However, is it the only mechanism? An additional mechanism for HSP release is now proposed as being of equal importance in the release of HSP72 into the circulation.

4. Active mechanisms

The active release hypothesis has been proposed as an additional mechanism to the passive release hypothesis. Three lines of evidence strongly support this hypothesis. First, as early as 1998 Pockley and coworkers demonstrated the presence of soluble HSP72 and antibodies against HSP72 in the peripheral circulation of normal individuals (Pockley *et al* 1998). Second, Guzhova and colleagues demonstrated that HSP72 is released by glia cells in the absence of necrotic cell death (Guzhova *et al* 2001). Third, and extremely compelling, psychological stress induced by exposure of a Sprague Dawley rat to a cat results in the release of HSP72 into the circulation (Fleshner *et al* 2004). In this study, a rat was put in a glass cage and a cat placed on top of the cage. This form of psychological stress induced a marked increase in circulating HSP72, as judged by the classical HSP72 sandwich ELISA. The terrified rat did not run around the cage thereby negating the possibility that damage to the muscles played a part in increased HSP72 release. Subsequent studies have demonstrated that HSP72 is released by B cells (Clayton *et al* 2005) and peripheral blood mononuclear cells (Hunter-Lavin *et al* 2004) under non necrotic conditions.

Using conditions that will not induce significant cell death, our group showed that IFN-*γ* and IL-10 induce the active release of constitutively expressed HSP70 as well as designed Hsc70 or HSP73 from tumours (Barreto *et al* 2003). However, these initial studies did not address the mechanism underlying HSP72 release. Recently, our group (Bausero *et al* 2005a; Gastpar *et al* 2005) and others (Lancaster and Febbraio 2005) have begun to elucidate the mechanism of active release of iHSP72 from viable cells. In our study, we demonstrated that certain pro-inflammatory cytokines, normally found in high concentrations within inflammatory foci including IFN-γ, IL-10 but not the antiinflammatory cytokine TGF- β 1, mediate the active release of HSP72. We further showed that whereas some eHSP72 could be found as free HSP72, a proportion of eHSP72 was released within exosomes (Bausero *et al* 2005a). Exosomes are internal vesicles of multi-vesicular bodies (MVB) released into the extracellular milieu upon fusion of MVB with the cell surface (Raposo *et al* 1996; Zitvogel *et al* 1998, 1999). In addition to containing HSP72 (Bausero *et al* 2005a; Gastpar *et al* 2005), exosomes are highly packed with immunostimulatory mediators including MHC class I and II (Raposo *et al* 1996; Zitvogel *et al* 1998, 1999) and costimulatory molecules (Escola *et al* 1998). Additionally, we demonstrated that HSP72 is released by a non classical protein transport pathway and that intact surface membrane lipid rafts are required for efficient stress-induced HSP72 release (Bausero *et al* 2005a; Gastpar *et al* 2005). These studies were recently confirmed in B cells (Clayton *et al*

2005). Recent studies by Lancaster and Febbraio (2005) demonstrate that exosomes provide the major pathway for secretory vesicular release of HSP72. However, using methyl-β-cyclodextrin (the cholesterol depleting agent) to disrupt lipid raft function, these authors were unable to confirm a role for lipid rafts in stress-induced HSP72 release from human peripheral blood mononuclear cells (PBMC) (Lancaster and Febbraio 2005). In order to address the cellular location of HSP72 after stress, a recent study demonstrated that newly synthesized HSP72 protein localizes within the Golgi region of HeLa cells and also concentrates on the surface of the plasma membrane and in the ruffled zone of migrating cells (Schneider *et al* 2002).

Taken together, these studies suggest that the active release hypothesis is an important mechanism by which HSP72 is released into the circulation. However, studies remain to be performed that conclusively demonstrate that T cell responses are primed by active release of HSP72 especially in the case of psychological stress or exercise.

5. Function of HSP70 on NK cells

HSP70 is highly expressed on the surface of certain tumour cells and its surface expression is absent in normal cells (Multhoff *et al* 1997). This unique surface expression of HSP70 may be utilized in the tumour-specific immune response and lysis of tumour cells mediated by non-MHC restricted natural killer cells (Multhoff *et al* 1999; Moser *et al* 2002; Multhoff 2002; Gross *et al* 2003a). Furthermore, a 14-mer peptide isolated from HSP70, termed TKD peptide stimulates the cytolytic and proliferative activity of NK cells similar to that of full length HSP70 (Multhoff *et al* 2001). HSP70-based immunotherapies have been extended to other infectious diseases such as malaria and HIV (Lehner and Anton 2002; Lehner and Shearer 2002; Wang *et al* 2002).

In comparison with immunocompetent cells, malignant tumour cells express high levels of surface bound HSP70 (Botzler *et al* 1996; Multhoff and Hightower 1996; Multhoff *et al* 1997; Botzler *et al* 1998b; Hantschel *et al* 2000). This HSP70 expression on tumours correlates with an increased sensitivity to natural killer (NK)-mediated cytolysis following cytokine stimulation (Botzler *et al* 1998a; Multhoff *et al* 1999, 2001). Recent studies have shown that the cytolytic activity of HSP70 can also be transduced by specific fragments of the HSP70 protein. Both the full-length HSP70 protein and the C-terminal domain of HSP70 stimulates the cytolytic activity of naive NK cells against HSP70-positive tumour target cells (Gross *et al* 2003a). In addition, tumour growth in mice with severe combined immunodeficiency was shown to be inhibited by HSP70-peptide-activated, CD94 positive natural killer cells (Moser *et al* 2002). Recent work from the Multhoff laboratory demonstrates that a 14 amino acid sequence of the HSP70 protein, termed TKD (TKDNNLLGRFELSG, $aa_{450-463}$) is the extracellular recognition site for NK cells (Multhoff *et al* 2001). These authors demonstrate that granzyme B specifically binds to portions of the HSP70 expressed on the plasma surface of tumours but not normal cells (Gross *et al* 2003b). These observations unravel a hitherto unknown mechanism by which cytolytic effector cells eliminate HSP70 expressing tumours in a perforin-independent, granzyme B-dependent manner. These studies are in agreement with recent findings that immunization of the peptide binding C-terminal portion of HSP70 $(aa_{359-610})$ (HSP70₃₅₉₋₆₁₀) is responsible for stimulating Th1-polarizing cytokine (IL-12 and TNF-*α*), C-C chemokine release, and acts as an adjuvant (Wang *et al* 2002). Immunization of nonhuman primates with $HSP70_{359}$ $_{610}$ induced the production of RANTES and IL-12, and acted as an adjuvant when loaded with CC5-peptide (Wang *et al* 2002), suggesting a possible alternative vaccine strategy for HIV infection (Lehner and Anton 2002; Lehner and Shearer 2002). An additional role of extracellular HSP70 as a chemoattractant for NK cells was recently demonstrated (Gastpar *et al* 2005).

6. Biological significance of released HSP72

Irrespective of whether HSP72 enters the circulation via an active or passive release mechanism, what is the role of eHSP72 in circulation? The danger theory postulates that immune activation involves danger/non-danger molecular recognition schemas and suggests that innate immune cells are activated by danger signals that are derived from stressed or damaged self-proteins (Matzinger, 1998; Gallucci and Matzinger 2001). It is now widely accepted that eHSP72 fit this criteria. The hypothesis is further reinforced by studies showing that circulating eHSP72 is increased and upregulated in diseased conditions including renal disease (Wright *et al* 2000), hypertension (Pockley *et al* 2002), atherosclerosis (Pockley *et al* 2003) and sickle cell disease (Adewoye *et al* 2005). However, intriguing questions still remain about the role of increased circulating eHSP72 during psychological stress like that demonstrated when a Sprague Dawley rat is exposed to a cat (Fleshner *et al* 2004) or the release eHSP72 into circulation by human brain in response to exercise (Lancaster *et al* 2004).

Is it possible that in these situations the circulating eHSP72 is priming the immune system to real or perceived danger? These are important questions, the answers to which will provide a keen insight into numerous psychological and pathophysiological conditions. However, extensive studies and concerted efforts that bring together HSP researchers from such disparate fields as immunology, molecular biology, cell biology, neurophysiology and psychology utilizing new breakthrough technologies including the

recently deciphered human genome and proteomics are required before conclusive answers can be given.

Acknowledgements

I thank all the faculty and staff of the Division of Investigative Pathology for helpful discussions. This work was supported in part by the National Institute of Health grant RO1CA91889, Scott & White Clinic, the Texas A&M University System Health Science Center College of Medicine, the Central Texas Veterans Health Administration and an Endowment from the Cain Foundation.

References

- Adewoye A H, Klings E S, Farber H W, Palaima E, Bausero M A, McMahon L, Odhiambo A, Surinder S, Yoder M, Steinberg M H and Asea A 2005 Sickle cell vaso-occlusive crisis induces the release of circulating serum heat shock protein-70; *Am. J. Hematol.* **78** 240–242
- Asea A 2003 Chaperokine-induced signal transduction pathways; *Exerc. Immunol. Rev.* **9** 25–33
- Asea A 2005 Stress proteins and initiation of immune response: chaperokine activity of hsp72; *Exerc. Immunol. Rev.* **11** 34–45
- Asea A, Kabingu E, Stevenson M A and Calderwood S K 2000a HSP70 peptide-bearing and peptide-negative preparations act as chaperokines; *Cell Stress Chaperones* **5** 425–431
- Asea A, Kraeft S K, Kurt-Jones E A, Stevenson M A, Chen L B, Finberg R W, Koo G C and Calderwood S K 2000b HSP70 stimulates cytokine production through a CD14-dependant pathway, demonstrating its dual role as a chaperone and cytokine; *Nat. Med.* **6** 435–442
- Asea A, Rehli M, Kabingu E, Boch J A, Bare O, Auron P E, Stevenson M A and Calderwood S K 2002a Novel signal transduction pathway utilized by extracellular HSP70: role of toll-like receptor (TLR) 2 and TLR4; *J. Biol. Chem.* **277** 15028–15034
- Asea A, Rehli M, Kabingu E, Boch J A, Bare O, Auron P E, Stevenson M A and Calderwood S K 2002b Novel signal transduction pathway utilized by extracellular HSP70: role of toll-like receptor (TLR) 2 and TLR4; *J. Biol. Chem.* **277** 15028–15034
- Barreto A, Gonzalez J M, Kabingu E, Asea A and Fiorentino S 2003 Stress-induced release of HSC70 from human tumours; *Cell. Immunol.* **222** 97–104
- Basu S, Binder R J, Suto R, Anderson K M and Srivastava P K 2000 Necrotic but not apoptotic cell death releases heat shock proteins, which deliver a partial maturation signal to dendritic cells and activate the NF-kappa B pathway; *Int. Immunol.* **12** 1539–1546
- Basu S and Srivastava P K 2000 Heat shock proteins: the fountainhead of innate and adaptive immune responses; *Cell Stress Chaperones* **5** 443–451
- Bausero M A, Gastpar R, Multhoff G and Asea A 2005a Alternative Mechanism by which IFN-{gamma} Enhances Tumour Recognition: Active Release of Heat Shock Protein 72; *J. Immunol.* **175** 2900–2912
- Bausero M A, Gastpar R, Multhoff G and Asea A 2005b Alternative mechanism by which IFN-gamma enhances tumour recognition: active release of heat shock protein 72; *J. Immunol.* **175** 2900–2912
- Binder R J, Anderson K M, Basu S and Srivastava P K 2000 Cutting edge: heat shock protein gp96 induces maturation and migration of CD11c+ cells *in vivo*; *J. Immunol.* **165** 6029–6035
- Botzler C, Issels R and Multhoff G 1996 Heat-shock protein 72 cell-surface expression on human lung carcinoma cells in associated with an increased sensitivity to lysis mediated by adherent natural killer cells; *Cancer Immunol. Immunother.* **43** 226–230
- Botzler C, Li G, Issels R D and Multhoff G 1998a Definition of extracellular localized epitopes of Hsp70 involved in an NK immune response; *Cell Stress Chaperones* **3** 6–11
- Botzler C, Schmidt J, Luz A, Jennen L, Issels R and Multhoff G 1998b Differential Hsp70 plasma-membrane expression on primary human tumours and metastases in mice with severe combined immunodeficiency; *Int. J. Cancer* 77 942–948
- Calderwood S K 2005 Chaperones and slow death a recipe for tumour immunotherapy; *Trends Biotechnol.* **23** 57–59
- Clayton A, Turkes A, Navabi H, Mason M D and Tabi Z 2005 Induction of heat shock proteins in B-cell exosomes; *J. Cell Sci.* **118** 3631–3638
- Dybdahl B, Slordahl S A, Waage A, Kierulf P, Espevik T and Sundan A 2005 Myocardial ischaemia and the inflammatory response: release of heat shock protein 70 after myocardial infarction; *Heart* **91** 299–304
- Dybdahl B, Wahba A, Haaverstad R, Kirkeby-Garstad I, Kierulf P, Espevik T and Sundan A 2004 On-pump versus off-pump coronary artery bypass grafting: more heat-shock protein 70 is released after on-pump surgery; *Eur. J. Cardiothorac Surg.* **25** 985–992
- Dybdahl B, Wahba A, Lien E, Flo T H, Waage A, Qureshi N, Sellevold O F, Espevik T and Sundan A 2002 Inflammatory response after open heart surgery: release of heat-shock protein 70 and signaling through toll-like receptor-4; *Circulation* **105** 685–690
- Escola J M, Kleijmeer M J, Stoorvogel W, Griffith J M, Yoshie O and Geuze H J 1998 Selective enrichment of tetraspan proteins on the internal vesicles of multivesicular endosomes and on exosomes secreted by human B-lymphocytes; *J. Biol. Chem.* **273** 20121–20127
- Fink A L 1999 Chaperone-mediated protein folding; *Physiol. Rev.* **79** 425–449
- Fleshner M, Campisi J, Amiri L and Diamond D M 2004 Cat exposure induces both intra- and extracellular Hsp72: the role of adrenal hormones; *Psychoneuroendocrinology* **29** 1142–1152
- Gallucci S, Lolkema M and Matzinger P 1999 Natural adjuvants: endogenous activators of dendritic cells; *Nat. Med.* **5** 1249–1255
- Gallucci S and Matzinger P 2001 Danger signals: SOS to the immune system; *Curr. Opin. Immunol.* **13** 114–119
- Gastpar R, Gehrmann M, Bausero M A, Asea A, Gross C, Schroeder J A and Multhoff G 2005 Heat shock protein 70 surface-positive tumour exosomes stimulate migratory and cytolytic activity of natural killer cells; *Cancer Res.* **65** 5238–5247
- Gross C, Hansch D, Gastpar R and Multhoff G 2003a Interaction of heat shock protein 70 peptide with NK cells involves the NK receptor CD94; *Biol. Chem.* **384** 267–279
- Gross C, Koelch W, DeMaio A, Arispe N and Multhoff G 2003b Cell surface-bound heat shock protein 70 (Hsp70) mediates perforin-independent apoptosis by specific binding and uptake of granzyme B; *J. Biol. Chem.* **278** 41173–41181
- Guzhova I, Kislyakova K, Moskaliova O, Fridlanskaya I, Tytell M, Cheetham M and Margulis B 2001 In vitro studies show that Hsp70 can be released by glia and that exogenous Hsp70 can enhance neuronal stress tolerance; *Brain Res.* **914** 66–73
- Hantschel M, Pfister K, Jordan A, Scholz R, Andreesen R, Schmitz G, Schmetzer H, Hiddemann W and Multhoff G 2000 Hsp70 plasma membrane expression on primary tumour biopsy material and bone marrow of leukemic patients; *Cell Stress Chaperones* **5** 438–442
- Housby J N, Cahill C M, Chu B, Prevelige R, Bickford K, Stevenson M A and Calderwood S K 1999 Non-steroidal anti-inflammatory drugs inhibit the expression of cytokines and induce HSP70 in human monocytes; *Cytokine* **11** 347–358
- Hunter-Lavin C, Davies E L, Bacelar M M, Marshall M J, Andrew S M and Williams J H 2004 Hsp70 release from peripheral blood mononuclear cells; *Biochem. Biophys. Res. Commun.* **324** 511–517
- Hut H M, Kampinga H H and Sibon O C 2005 Hsp70 protects mitotic cells against heat-induced centrosome damage and division abnormalities; *Mol. Biol. Cell.* **16** 3776–3785
- Jaattela M, Wissing D, Kokholm K, Kallunki T and Egeblad M 1998 Hsp70 exerts its anti-apoptotic function downstream of caspase-3-like proteases; *Embo J.* **17** 6124–6134
- Lancaster G I and Febbraio M A 2005 Exosome-dependent trafficking of HSP70: a novel secretory pathway for cellular stress proteins; *J. Biol. Chem.* **280** 23349–23355
- Lancaster G I, Moller K, Nielsen B, Secher N H, Febbraio M A and Nybo L 2004 Exercise induces the release of heat shock protein 72 from the human brain in vivo; *Cell Stress Chaperones* **9** 276–280
- Lehner T and Anton P A 2002 Mucosal immunity and vaccination against HIV; *Aids (Suppl. 4)* **16** S125–S132
- Lehner T, Bergmeier L A, Wang Y, Tao L, Sing M, Spallek R and van der Zee R 2000 Heat shock proteins generate betachemokines which function as innate adjuvants enhancing adaptive immunity; *Eur. J. Immunol.* **30** 594–603
- Lehner T and Shearer G M 2002 Alternative HIV vaccine strategies; *Science* **297** 1276–1277
- Lindquist S and Craig E A 1988 The heat-shock proteins; *Annu. Rev. Genet.* **22** 631–677
- Matzinger P 1998 An innate sense of danger; *Semin. Immunol.* **10** 399–415
- Melcher A, Todryk S, Hardwick N, Ford M, Jacobson M and Vile R G 1998 Tumour immunogenicity is determined by the mechanism of cell death via induction of heat shock protein expression; *Nat. Med.* **4** 581–587
- Moehler M H, Zeidler M, Wilsberg V, Cornelis J J, Woelfel T, Rommelaere J, Galle P R and Heike M 2005 Parvovirus H-1 induced tumour cell death enhances human immune response *in vitro* via increased phagocytosis, maturation, and crosspresentation by dendritic cells; *Hum. Gene Ther.* **16** 996–1005
- Moser C, Schmidbauer C, Gurtler U, Gross C, Gehrmann M, Thonigs G, Pfister K and Multhoff G 2002 Inhibition of tumour growth in mice with severe combined immunodeficiency is

mediated by heat shock protein 70 (Hsp70)-peptide-activated, CD94 positive natural killer cells; *Cell Stress Chaperones* **7** 365–373

- Multhoff G 2002 Activation of natural killer cells by heat shock protein 70; *Int. J. Hyperthermia* **18** 576–585
- Multhoff G, Botzler C, Jennen L, Schmidt J, Ellwart J and Issels R 1997 Heat shock protein 72 on tumour cells: a recognition structure for natural killer cells; *J. Immunol.* **158** 4341–4350
- Multhoff G and Hightower L E 1996 Cell surface expression of heat shock proteins and the immune response; *Cell Stress Chaperones* **1** 167–176
- Multhoff G, Mizzen L, Winchester C C, Milner C M, Wenk S, Eissner G, Kampinga H H, Laumbacher B and Johnson J 1999 Heat shock protein 70 (Hsp70) stimulates proliferation and cytolytic activity of natural killer cells; *Exp. Hematol.* **27** 1627–1636
- Multhoff G, Pfister K, Gehrmann M, Hantschel M, Gross C, Hafner M and Hiddemann W 2001 A 14-mer Hsp70 peptide stimulates natural killer (NK) cell activity; *Cell Stress Chaperones* **6** 337–344
- Noessner E, Gastpar R, Milani V, Brandl A, Hutzler P J, Kuppner M C, Roos M, Kremmer E, Asea A, Calderwood S K and Issels R D 2002 Tumour-derived heat shock protein 70 peptide complexes are cross- presented by human dendritic cells; *J. Immunol.* **169** 5424–5432
- Panjwani N N, Popova L and Srivastava P K 2002 Heat shock proteins gp96 and hsp70 activate the release of nitric oxide by APCs; *J. Immunol.* **168** 2997–3003
- Pittet J F, Lee H, Morabito D, Howard M B, Welch W J and Mackersie R C 2002 Serum levels of Hsp 72 measured early after trauma correlate with survival; *J. Trauma* **52** 611–617; discussion 617
- Pockley A G, De Faire U, Kiessling R, Lemne C, Thulin T and Frostegard J 2002 Circulating heat shock protein and heat shock protein antibody levels in established hypertension; *J. Hypertens.* **20** 1815–1820
- Pockley A G, Georgiades A, Thulin T, de Faire U and Frostegard J 2003 Serum heat shock protein 70 levels predict the development of atherosclerosis in subjects with established hypertension; *Hypertension* **42** 235–238
- Pockley A G, Shepherd J and Corton J M 1998 Detection of heat shock protein 70 (Hsp70) and anti-Hsp70 antibodies in the serum of normal individuals; *Immunol. Invest.* **27** 367–377
- Raposo G, Nijman H W, Stoorvogel W, Liejendekker R, Harding C V, Melief C J and Geuze H J 1996 B lymphocytes secrete antigen-presenting vesicles; *J. Exp. Med.* **183** 1161–1172
- Schneider E M, Niess A M, Lorenz I, Northoff H and Fehrenbach E 2002 Inducible hsp70 expression analysis after heat and physical exercise: transcriptional, protein expression, and subcellular localization; *Ann. N. Y. Acad. Sci.* **973** 8–12
- Singh-Jasuja H, Scherer H U, Hilf N, Arnold-Schild D, Rammensee H G, Toes R E and Schild H 2000 The heat shock protein gp96 induces maturation of dendritic cells and down-regulation of its receptor; *Eur. J. Immunol.* **30** 2211–2215
- Srivastava P 2002 Interaction of heat shock proteins with peptides and antigen presenting cells: chaperoning of the innate and adaptive immune responses; *Annu. Rev. Immunol.* **20** 395–425
- Srivastava P K 2000 Heat shock protein-based novel immunotherapies; *Drug News Perspect.* **13** 517–522
- Srivastava P K 2003 Hypothesis: controlled necrosis as a tool for immunotherapy of human cancer; *Cancer Immun.* **3** 4
- Srivastava P K 2005 Immunotherapy for human cancer using heat shock protein-Peptide complexes; *Curr. Oncol. Rep.* **7** 104–108
- Srivastava P K and Amato R J 2001 Heat shock proteins: the 'Swiss Army Knife' vaccines against cancers and infectious agents; *Vaccine* **19** 2590–2597
- Srivastava P K, Udono H, Blachere N E and Li Z 1994 Heat shock proteins transfer peptides during antigen processing and CTL priming; *Immunogenetics* **39** 93–98
- Tang D, Xie Y, Zhao M, Stevenson M A and Calderwood S K 2001 Repression of the HSP70B promoter by NFIL6, Ku70, and MAPK involves three complementary mechanisms; *Biochem. Biophys. Res. Commun.* **280** 280–285
- Tavaria M, Gabriele T, Kola I and Anderson R L 1996 A hitchhiker's guide to the human Hsp70 family; *Cell Stress Chaperones* **1** 23–28
- Terry D F, McCormick M, Andersen S, Pennington J, Schoenhofen E, Palaima E, Bausero M, Ogawa K, Perls T T and Asea A 2004 Cardiovascular disease delay in centenarian offspring: role of heat shock proteins; *Ann. N. Y. Acad. Sci.* **1019** 502–505
- Wang Y, Kelly C G, Singh M, McGowan E G, Carrara A S, Bergmeier L A and Lehner T 2002 Stimulation of Th1 polarizing cytokines, C-C chemokines, maturation of dendritic cells, and adjuvant function by the peptide binding fragment of heat shock protein 70; *J. Immunol.* **169** 2422–2429
- Wright B H, Corton J M, El-Nahas A M, Wood R F and Pockley A G 2000 Elevated levels of circulating heat shock protein 70 (Hsp70) in peripheral and renal vascular disease; *Heart Vessels* **15** 18–22
- Zitvogel L, Fernandez N, Lozier A, Wolfers J, Regnault A, Raposo G and Amigorena S 1999 Dendritic cells or their exosomes are effective biotherapies of cancer; *Eur. J. Cancer (Suppl. 3)* **35** S36–S38
- Zitvogel L, Regnault A, Lozier A, Wolfers J, Flament C, Tenza D, Ricciardi-Castagnoli P, Raposo G and Amigorena S 1998 Eradication of established murine tumours using a novel cell-free vaccine: dendritic cell-derived exosomes; *Nat. Med.* **4** 594–600

*e*Publication: 15 March 2007