

# **The Potential of NLRP3 Infammasome as a Therapeutic Target in Neurological Diseases**

**Wenfang He1 · Zhiping Hu2 · Yanjun Zhong1 · Chenfang Wu1 · Jinxiu Li[1](http://orcid.org/0000-0002-4174-1994)**

Received: 27 April 2022 / Accepted: 10 January 2023 / Published online: 21 January 2023 © The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2023

# **Abstract**

NLRP3 (NLRP3: NOD-, LRR-, and pyrin domain-containing protein 3) infammasome is the best-described infammasome that plays a crucial role in the innate immune system and a wide range of diseases. The intimate association of NLRP3 with neurological disorders, including neurodegenerative diseases and strokes, further emphasizes its prominence as a clinical target for pharmacological intervention. However, after decades of exploration, the mechanism of NLRP3 activation remains indefnite. This review highlights recent advances and gaps in our insights into the regulation of NLRP3 infammasome. Furthermore, we present several emerging pharmacological approaches of clinical translational potential targeting the NLRP3 infammasome in neurological diseases. More importantly, despite small-molecule inhibitors of the NLRP3 infammasome, we have focused explicitly on Chinese herbal medicine and botanical ingredients, which may be splendid therapeutics by inhibiting NLRP3 infammasome for central nervous system disorders. We expect that we can contribute new perspectives to the treatment of neurological diseases.

**Keywords** NLRP3 infammasome · Neurological diseases · Stroke · Alzheimer's disease · Neuroinfammation · Chinese herbal medicine

## **Abbreviations**



 $\boxtimes$  Jinxiu Li jinxiuli2021@csu.edu.cn

<sup>1</sup> Department of Critical Care Medicine, The Second Xiangya Hospital, Central South University, Changsha, China

<sup>2</sup> Department of Neurology, The Second Xiangya Hospital, Central South University, Changsha, China





# **Introduction**

The NLRP3 infammasome is a protein complex composed of a sensor protein (NLRP3), an adaptor apoptosis-associated speck-like protein containing a CARD (ASC), and a downstream efector (pro-caspase-1) (Fig. [1\)](#page-2-0). Following the activation of the infammasome sensor molecule by agonists, ASC self-associates into a helical fbrous assembly [\[1](#page-12-0)] that forms the so-called ASC speck or pyroptosome [\[2](#page-12-1)], which serves as a molecular plateau for caspase-1 activation by proximally induced autocatalysis [\[3\]](#page-12-2). Activated caspase-1 promotes the maturation and secretion of several

pro-infammatory cytokines, including interleukin-1β (IL-1β) and interleukin-18 (IL-18). On the one hand, it also triggers cellular pyroptosis capable of clearing pathogens and damaged cells [[4\]](#page-12-3). Depending on the sensor, the main infammasomes include NLRP3 (NOD-, LRR-, and pyrin domain-containing protein 3), NOD-like receptor family CARD domain-containing protein 4 (NLRC4), interferoninducible protein AIM2, NLRP1, and others. NLRP3 is a NOD-like receptor family (NLRs) member and is the bestdescribed infammasome sensor, which plays as a key component of the innate immune system. NLRP3 contains an amino-terminal PYRIN (PYD) domain, a nucleotide-binding NACHT domain, and a carboxy-terminal leucine-rich repeat (LRR) domain (Fig. [1\)](#page-2-0). The NACHT domain carries ATPase activity, and the ATP binding is essential for the function of NLRP3 [\[5](#page-12-4)]. Gain-of-function mutations of the *Nlrp3* gene result in a dominantly inherited autoinfammatory disease called cryptochrome-associated periodic syndrome (CAPS) [[6–](#page-12-5)[8\]](#page-12-6). Recent research reveals that the inactive endogenous full-length NLRP3 is formed in a double-ring cage that is primarily membrane-localized [\[9](#page-12-7)]. The location is consistent with previous studies of NLRP3 on various membranous organelles [[10\]](#page-12-8). The double-ring cage is combined by LRR-LRR interactions, while the PYD domain is hidden inside to prevent premature activation [\[9\]](#page-12-7). Furthermore, it has been demonstrated that this specifc cage confguration is required for the dispersion of the *trans*-Golgi network (TGN) [[9](#page-12-7)], a common cellular event during NLRP3 activation [\[11\]](#page-12-9) that we will discuss later. Recently, it has been further suggested that NIMA-related protein kinase 7 (NEK7), a serine–threonine kinase previously involved in mitosis, is required to activate NLRP3 infammasome [\[12](#page-12-10), [13](#page-12-11)]. After being triggered by stimuli, NEK7-NLRP3 interacts to form a complex, which is essential for the subsequent recruitment of ASC and the activation of caspase-1. This interaction is thought to occur downstream of reactive oxygen species (ROS) and potassium efflux  $[12]$  $[12]$  and is specific to NLRP3 rather than other infammasome sensors [[12,](#page-12-10) [13](#page-12-11)].

Up to now, NLRP3 infammasome is the most investigated infammasome in the central nervous system (CNS). It is mainly studied in microglia but can also be expressed in astrocytes and oligodendrocytes in patients or animal models with neurologic diseases [\[14](#page-12-12), [15](#page-12-13)]. Regarding CNS, NLRP3 infammasome is particularly sensitive to endosome injury and responds to various aggregated proteins associated with diseases, such as Aβ [\[16\]](#page-13-0) or α-synuclein [[17\]](#page-13-1). Many studies suggested the involvement of NLRP3 infammasomes in neurodegenerative diseases, while the pharmacologic or genetic deletion or inhibition of NLRP3 infammasomes could improve neurologic function [\[18\]](#page-13-2). Previous studies summarized that the use of specifc inhibitors of NLRP3 infammasomes, along with microglial autophagy inducers, could contribute to the elimination of misfolded proteins,



<span id="page-2-0"></span>**Fig. 1** A schematic diagram of mediators and stimulators involved in NLRP3 infammasome activation and the structure of NLRP3. The canonical activation of NLRP3 infammasome consists of two steps: priming (signal 1) and activation (signal 2). Signal 1 (left), triggered by the activation of cytokines (such as IL-β, TNF) or pathogen-associated molecular patterns (PAMPs), is to initiate the transcription of *NLRP3* and pro-infammatory genes and upregulate their expressions in cells. The priming step of NLRP3 infammasome activation provides cells with permission to be activated subsequently. Signal 2 (right) is trigged by various PAMPs and DAMPs. At present, the well-accepted upstream signals which could activate NLRP3 infammasome comprise  $K^+$  efflux, the leakage of cathepsins caused by lysosomal disruption, impaired mitochondrial function,  $Ca^{2+}$  influx and *trans*-Golgi disassembly. Activation of NLRP3 by agonists subsequently attracts ASC and caspase-1 to assemble into NLRP3 infammasome, thereby inducing caspase-1 self-cleavage and activation. Activated caspase-1 then promotes the maturation and secretion of pro-infammatory cytokines including interleukin-1β (IL-1β)

the scavenging of impaired mitochondria and ROS in neurodegenerative diseases [\[19](#page-13-3)]. Recent studies also highlighted that NLRP3 infammasome was also engaged in the pathological processes of other neurological disorders, such as ischemic stroke, which have not traditionally been considered as infammatory in nature [\[20\]](#page-13-4). Moreover, a number of inhibitors targeting NLRP3 infammasomes have demonstrated therapeutic efficacy in animal experiments or *in vivo* models. This paper reviews the latest developments and gaps regarding NLRP3 infammasome activation. Meanwhile, we summarize the recent fndings on inhibiting the NLRP3 infammasome signaling pathway in treating neurological diseases. Besides small-molecule inhibitors and microRNA

and interleukin-18 (IL-18).The non-canonical NLRP3 infammasome activation is triggered by caspase-11 in mice (human homologs caspase-4 and caspase-5). After sensing LPS, caspase-11 is activated via Toll-like receptor 4 (TLR4), leading to the cleavage of gasdermin D (GSDMD). The cleaved GSMD, N-terminal, forms membrane pores, thereby resulting in potassium efflux and pyroptosis. The dotted line indicates the pathway where the mechanism is not yet established. ASC, apoptosis-associated speck-like protein containing a CARD; CARD, caspase recruitment domain; CLIC, chloride intracellular channel protein; GSDMD, gasdermin D; N- terminal, GSDMD amino-terminal cell death domain; IL-1β, interleukin-1β; IL-18, interleukin-18; IL-1R1, IL-1 receptor type 1; LPS, lipopolysaccharide; LRR, leucine-rich repeat; NEK7, NIMA-related kinase 7; NF-κB, nuclear factor-κB; P2X7, purinergic 2X7 receptor; PtdIns4P, phosphatidylinositol-4-phosphate; PYD, pyrin domain; ROS, reactive oxygen species; TLR, Toll-like receptor; TNF, tumor necrosis factor; TNFR, tumor necrosis factor receptor; TWIK2, two-pore domain weak inwardly rectifying K.<sup>+</sup> channel 2

for NLRP3 infammasomes, we specifcally focused on the remarkable studies of herbal medicines, plant-derived ingredients, and drugs used to treat other diseases, which have rarely been systematically reviewed in other similar articles.

# **Activation of NLRP3 Infammasome**

## **Canonical NLRP3 Activation**

It is well established that two-step procedures stimulate the canonical activation of NLRP3 infammasome: priming (signal 1) and activation (signal 2) (Fig. [1](#page-2-0)). Briefy, signal 1 is to initiate the transcription of *Nlrp3* and pro-infammatory genes and upregulate their expressions in cells. This process is involved with pattern recognition receptors (PRRs) such as Toll-like receptors (TLRs), nucleotide-binding oligomerization domain-containing protein 2 (NOD2), and tumor necrosis factor (TNF) receptors induced by various pathogen-associated molecular patterns (PAMPs) or damage-associated molecular patterns (DAMPs) [[21,](#page-13-5) [22](#page-13-6)]. Nuclear factor-κB (NF-κB) is further activated by these receptors, increasing the transcription of the infammasome components [[23\]](#page-13-7). The activation step (signal 2) occurs following the recognition of an NLRP3 infammasome stimulus and induces the recruitment of ASC and caspase-1. Unlike other members of the NLR family, the NLRP3 infammasome responds to various stimuli independent in origin, chemical composition, and structural properties. The stimulants range from intrinsic and extrinsic agents, known as DAMPs and PAMPs. PAMP signals mainly come from invasive pathogens such as bacterial toxins [\[24\]](#page-13-8). Lipopolysaccharide (LPS) is commonly thought to be a prototypical PAMP. DAMP signals perceived by NLRP3 infammasome are usually produced by the damaged body itself, such as triphosphate (ATP), silica crystals [\[25\]](#page-13-9), amyloid β (Aβ) [\[16](#page-13-0)], and α-synuclein [[17](#page-13-1)]. Given the high variability of activators, they are unlikely to interact by directly binding to NLRP3 infammasome. Recent studies suggested that the NLRP3 infammasome may perform as a signal integrator, recognizing any molecules or conditions that could induce alterations in homeostasis or pathological state [[26\]](#page-13-10). However, how NLRP3 infammasome sense and respond to dangerous cellular signals remains indefnite.

Here, we introduced the widely accepted upstream signals and some latest studies.  $K^+$  efflux is a common required upstream event for NLRP3 activation [\[27\]](#page-13-11). After being simulated by ATP, purinergic 2X7 receptor (P2X7R) promotes  $K^+$  efflux with two-pore domain weak inwardly rectifying  $K^+$  channel 2 (TWIK2), as well as  $Ca^{2+}$  and  $Na^+$ influx  $[28, 29]$  $[28, 29]$  $[28, 29]$  $[28, 29]$ . Pore-forming toxins also induce  $K^+$  efflux [\[24\]](#page-13-8). Meanwhile,  $Ca^{2+}$  flux and  $K^+$  efflux are often synergistic during NLRP3 activation  $[28]$ . Ca<sup>2+</sup> release from the endoplasmic reticulum (ER) promoted by  $K^+$  is an upstream signal in NLRP3 infammasome activation [[30](#page-13-14)]. More particularly, ER stress, resulting in  $Ca^{2+}$  release from the ER lumen, magnifes the activation of NLRP3 infammasome. Besides potassium and calcium, chloride is also engaged [\[30](#page-13-14)]. Cl<sup>−</sup> efflux is thought to promote ASC aggregation during NLRP3 infammasome formation, depending on chloride intracellular channel proteins (CLICs) [[31\]](#page-13-15). Lysosomal disruption and the leakage of cathepsins is another common mechanism of NLRP3 activation [\[25](#page-13-9)]. Activation caused by phagocytosis of crystalline material or protein aggregates is mediated via this pathway mainly. Meanwhile, lysosomal damage could trigger  $K^+$  efflux and  $Ca^{2+}$  influx [[27](#page-13-11)]. Early studies revealed ROS production was necessary for NLRP3 infammasome activation [\[32\]](#page-13-16). In addition, mitochondrial dysfunction and the release of mitochondrial reactive oxygen species (mtROS), cardiolipin, and mitochondrial DNA (mtDNA) into the cytoplasm is an equally critical upstream event independent of  $K^+$  efflux and lysosomal disruption involved in NLRP3 activation [\[33](#page-13-17), [34](#page-13-18)]. Despite mitochondria [\[35](#page-13-19)], dysfunction of organelles such as the endoplasmic reticulum [[36](#page-13-20)] and Golgi apparatus [[37\]](#page-13-21) are also engaged in NLRP3 infammasome activation. The Golgi apparatus is defnitely the most unexpected and astonishing among all these organelles. Recent studies implicate that the Golgi apparatus is not only an organelle for intracellular sorting but also plays a crucial role in signal transfer during the inner immune response [\[37](#page-13-21)]. Chen et al. indicated that the trans-Golgi network (TGN), rather than *cis*- or medial-Golgi, was disassembly induced by several NLRP3 activators into dispersed *trans*-Golgi network (dTGN) [\[11](#page-12-9)]. dTGN performed as a scafold that recruited NLRP3 via phosphatidylinositol-4-phosphate (PtdIns4P), promoting NLRP3 infammasome assembly and the downstream caspase-1 activation. Intriguing, it was further suggested that *trans*-Golgi disassembly was the common cellular event in two distinct pathways,  $K^+$  efflux-dependent and mitochondria-dependent NLRP3 activation, which eventually led to NLRP3 aggregation [[11](#page-12-9)]. Recent studies also suggested that the activation of NLRP3 required the ER-to-Golgi translocation of sterol regulatory element binding protein (SREBP) 2 and SREBP cleavageactivating protein (SCAP) [[10](#page-12-8)]. Another study clarifed that NLRP3 stimuli invoked the localization of mitochondriaassociated endoplasmic reticulum membranes (MAMs) close to the Golgi membrane [\[38\]](#page-13-22). This inter-organelle crosstalk relied on the recruitment of protein kinase domain (PKD) at the Golgi DAG site, which promoted the assembly of NLRP3 oligomers into active infammasomes [[38\]](#page-13-22).

Of note, although researchers have made numerous eforts to understand the upstream events during NLRP3 activation, there is still no defnitive conclusion. Many pathways are cross-linked and overlapping, and the results occasionally confict with others. Further investigation is required.

## **Non‑canonical NLRP3** *A***ctivation**

Diferent from the canonical activation, the non-canonical NLRP3 infammasome activation is triggered by caspase-4 and caspase-5 in humans [[39\]](#page-13-23) and caspase-11 in mice [\[40](#page-13-24)], respectively (Fig. [1\)](#page-2-0). Generally speaking, upon sensing LPS produced by Gram-negative bacteria, caspase-4, caspase-5, and caspase-11 are activated via Toll-like receptor 4 (TLR4), leading to the cleavage of gasdermin D (GSDMD). The cleaved N-terminal of GSMD then forms membrane pores, thereby resulting in potassium efflux and pyroptosis  $[41, 42]$  $[41, 42]$  $[41, 42]$  $[41, 42]$ . Nevertheless, the canonical and non-canonical infammasome activation shared similar consequences, namely, the release of pro-infammatory cytokines IL-1β and IL-18. It was reported that the oxidized phospholipid 1-palmitoyl-2-arachidonoyl-sn-glycero-3-phosphorylcholine (oxPAPC) also induced caspase-11-dependent IL-1β release via this non-canonical pathway [\[43](#page-13-27)].

# **Inhibitors of NLRP3 Infammasome**

Many inhibitors have been developed to investigate the regulatory mechanisms of NLRP3 infammasomes and their roles in disease pathogenesis. These inhibitors directly or indirectly inhibit NLRP3 infammasome components or related upstream/downstream signaling events [[44\]](#page-13-28). Among them, the activation step is the favored target for the creation of NLRP3 antagonists, as it should lead to the most direct and specialized suppression. However, the molecular mechanism of inhibition or the precise target has not been fully elucidated for many of these compounds. To date, MCC950 (also known as CP-456773 or CRID3) is the most studied and specifc NLRP3 infammasome inhibitor, as it does not inhibit any other identifed infammasomes such as NLRP1, NLRC4, or AIM2 [[45\]](#page-13-29). Meanwhile, MCC950 could inhibit both the canonical and non-canonical activation of NLRP3 infammasome by all known stimuli at present. Due to its specifcity, it is the most recommended and employed compound to study the relevant events of NLRP3 infammasome *in vitro* and *in vivo*. MCC950 binds to NLRP3 in the NACHT domain and blocks its ability to hydrolyze ATP, thereby preventing it from retaining an active form, thus inhibiting the recruitment to ASC and the cleavage of caspase-1 [\[46\]](#page-13-30). It was also suggested that MCC950 could perform its function by inhibiting the chloride efflux following nigericin stimulation [[47](#page-14-0)]. Shockingly, MCC950 shows therapeutic efficacy in a wide range of disease models *in vivo*, including diabetes [[48](#page-14-1)], neurodegenerative disease [\[49\]](#page-14-2), traumatic brain injury [[50,](#page-14-3) [51](#page-14-4)], atherosclerosis [[52](#page-14-5)], steatohepatitis [[53](#page-14-6)], and colitis [\[32](#page-13-16)]. Other small-molecule inhibitors of NLRP3 infammasome include Bay 11–7082 [[54](#page-14-7)], JC-171 [\[55](#page-14-8)], and β-hydroxybutyrate (BHB) [\[56\]](#page-14-9). Interestingly, many traditional Chinese medicines, botanical ingredients, and drugs used in treating other diseases have also been shown to inhibit the action of NLRP3 infammasome. We summarize the inhibitors in Table [1](#page-5-0) and will discuss these agents later in specifc disease models.

# **Therapeutic Targeting of NLRP3 in Neurological Diseases**

Here we overview the recent studies of NLRP3 infammasome performed in the nervous system and try to assess its potential as a drug target for the treatment of neurological disease, especially in stroke and neurodegenerative diseases such as Alzheimer's disease. Moreover, we have specifcally focused on Chinese herbal medicine, botanical ingredients, and drugs used in treating other conditions, which may be splendid therapeutics by inhibiting NLRP3 infammasome for CNS disorders. Figure [2](#page-6-0) presents a schematic diagram of the possible major drivers of NLRP3 infammasome activation in neurological diseases.

## **Ischemic Stroke**

Ischemic stroke is a severe life-threatening condition that accounts for almost 87% of all strokes [[92\]](#page-15-0). Although intravenous thrombolysis and thrombectomy have been developed recently, ischemic stroke remains the second leading cause of death globally [[93](#page-15-1)]. The primary complications of strokes, including cognitive impairment and depression [[94\]](#page-15-2), consequently carry heavy social and family burdens. However, most neuroprotective agents which proved to be efective in animal studies ended in failure [[95,](#page-15-3) [96\]](#page-15-4). Therefore, fnding new directions for pharmaceutical treatment is urgent and essential.

Previous fndings highlighted the importance of NLRP3 infammasome in mediating the infammatory response in aseptic tissue injury during post-stroke damage [[97](#page-15-5), [98](#page-15-6)]. At the early stage of ischemia, hypoxia and hypoperfusion accompanied by the generation of DAMPs could directly activate NLRP3 infammasome, exacerbating inner immune response and the injuries of stroke. The generation of ROS induced in ischemia–reperfusion could activate the NLRP3 infammasome and cause more neural damage [[99\]](#page-15-7). In addition to ROS, other dangerous signals that can trigger NLRP3 activation include energy depletion, acidosis, cathepsin release, mitochondrial DNA oxidation and dysfunction, intracellular Ca<sup>2+</sup> accumulation, decreased intracellular  $K^+$ concentration, cell swelling, and protein kinase R (PKR) activation [\[100\]](#page-15-8). Pro-infammatory cytokines such as IL-1  $β$  and caspase-1, as well as NLRP3, were highly expressed in cellular models, animal models, and patients with stroke [[101–](#page-16-0)[104\]](#page-16-1). These massive cytokines and chemokines were highly toxic to neurons. It was well established that infammatory cascade response mediated by NLRP3 inflammasome contributed to cerebral edema and hemorrhage, blood–brain barrier damage, and more neuronal death [[105](#page-16-2)].

At present, the cell-specifc expression and distribution of the NLRP3 infammasome in cerebral ischemia–reperfusion injury have not been fully elucidated. In the mouse brain, expression of NLRP3, ASC, and caspase-1 has been observed in microglia after LPS stimulation and was not detected in astrocytes, suggesting that microglia may be the main locus engaged in the generation of NLRP3 infam-masome [\[106\]](#page-16-3). Microglia is the major innate immune cell population in the brain [[107](#page-16-4)]. Impaired microglia immune

<span id="page-5-0"></span>



*AD*, Alzheimer's disease; *EAE*, experimental autoimmune encephalomyelitis; *ICH*, intracerebral hemorrhage; *MS*, multiple sclerosis; *PD*, Parkinson's disease; *SAE*, sepsis-associated encephalopathy; *SAH*, subarachnoid hemorrhage; *TBI*, traumatic brain injury

functions often lead to overproduction of infammatory mediators and exacerbate central nervous system damage [\[108,](#page-16-5) [109](#page-16-6)]. Consistent with this, there were signs that NLRP3 was expressed in microglia and vascular endothelial cells but not neurons [[57](#page-14-10)]. In contrast, data also showed that the levels of NLRP3 infammasome proteins, IL-1β, and IL-18 increased in neurons under ischemic conditions as well as in neurons from stroke patients [\[88](#page-15-9), [103,](#page-16-7) [110\]](#page-16-8). Different ischemia models and interventions and the duration of ischemic injury were possible explanations. It was suggested that the activation pattern of NLRP3 infammasome induced by ischemia–reperfusion injury varied between neurons and glial cells [[88](#page-15-9)].

The NLRP3 infammasome may act through multiple mechanisms to mediate neuronal and glial cell death in ischemic stroke. The mechanisms include enhancing the production and secretion of pro-infammatory cytokines such as IL-1 $\beta$  and IL-18 and through the multiplicity of cleaved caspase-1 in mediating neural apoptosis. Consistently, further studies demonstrated that the interference of NLRP3 activation matched the regulation of neuroprotective function. Blockade of NLRP3 reduced the infarction



<span id="page-6-0"></span>**Fig. 2** A review of mediators and stimulators of NLRP3 infammasome activation presently known to be involved mainly in microglia in neurological dysfunction

volume and neurovascular complications in the middle cerebral artery occlusion mice model, and inhibited neuronal apoptosis *in vivo* and *in vitro* [[111\]](#page-16-9). Compared to the controls, MCC950-treated mice showed signifcant reductions in infarcts, edema, and hemoglobin content, along with improved neurological deficits  $[112]$  $[112]$  $[112]$ . The expression of NLRP3, cleavage products caspase-1, and IL-1β were also decreased in the penumbral region after ischemic. Meanwhile, in parallel, these effects of MCC950 were associated with reduced phosphorylation levels of NFkBp65 and IkBa. Thus the study showed that MCC950 exhibited therapeutic potential in animal models with ischemic stroke. Furthermore, it was indicated that treatment of MCC950 improved cognitive function, neurovascular unit integrity, and neurovascular remodeling in rats after stroke [[113](#page-16-11)]. NLRP3 and IL-1 $\beta$  expressions were also significantly decreased in the hippocampus with MCC950 interference [[113](#page-16-11)]. Recently, it was further suggested that the blood–brain barrier integrity and functional outcome were improved considerably in MCC950-treated mice after ischemic stroke [\[114\]](#page-16-12). Moreover, endothelial cell survival was elevated *in vivo* as well as *in vitro* experiments with consistent results by inhibiting infammatory signaling cascades and pyroptosis [\[114](#page-16-12)]. Astragaloside IV (AST IV), one of the practical components of the traditional Chinese medicine Astragalus membranaceus, exerted protective efects against cerebral ischemia–reperfusion injury through inhibiting NLRP3 infammasome-mediated pyroptosis via activating nuclear factor erythroid-2–related factor 2 (Nrf2) signaling [[75](#page-15-11)]. However, the regulatory mechanism of NLRP3 is not well understood. Fann et al. were the frst to identify that NF-κB and mitogen-activated protein kinase (MAPK) signaling pathways could regulate the activation of NLRP3 infammasome in ischemic primary cortical neurons [\[115](#page-16-13)]. A recent study indicated that NLRP3 infammasome might be directly controlled by JAK2/STAT3 signaling pathway after stroke *in vitro* [\[116](#page-16-14)]*.* NLRP3 infammasome was also involved in the regulation of autophagy. Yue Liu et al. suggested that miR-135a-5p, highly expressed in M2 phenotype microglia-derived extracellular vesicles, could negatively regulate NLRP3 expression via thioredoxin-interacting protein (TXNIP), thereby reducing neuronal autophagy and ischemic brain damage [[83\]](#page-15-19). Another fnding suggested that resveratrol, a Chinese herbal medicine, could alleviate ischemia–reperfusion injury in rats by suppressing NLRP3 infammasome activation via Sirt1-dependent autophagy activity [\[72](#page-14-21)].

Interestingly, some market-approved drugs also present neuroprotection effects via the NLRP3 inflammasome pathway. Lithium  $(L<sup>+</sup>)$  is used in psychiatry to treat acute mania and acute depression. Recently, it has been reported to exert neuroprotection in stroke patients [[117\]](#page-16-15). It was further suggested that the underlying mechanism of Li<sup>+</sup> was related to NLRP3 inflammasomes.  $Li<sup>+</sup>$  inhibited the activation of NLRP3 through two signaling pathways, AKT/GSK3β/βcatenin and AKT/FoxO3a/β-catenin, which suppressed the production of ROS [[88\]](#page-15-9). Idebenone is a well-used antioxidant as a mitochondrial protectant. Peng et al. found that idebenone could efectively alleviate the loss of mitochondrial membrane potential  $(\Delta \psi m)$ , attenuate mitochondrial dysfunction, inhibit the NLRP3-mtDNA interaction and NLRP3 activation, and subsequently reduce neurological defcits in rats with ischemic stroke [[89\]](#page-15-24). Another recent study reported that verapamil acted as a reliable adjuvant to thrombolytic therapy by reversing the tissue plasminogen activator (tPA) induced activation of the TXNIP/NLRP3 infammasome pathway and reducing infarct volume in hyperglycemic stroke [[90\]](#page-15-25). Therefore, targeting upstream and downstream pathways of NLRP3 infammasome signaling may become a promising therapeutic strategy for ischemic stroke.

#### **Hemorrhagic** *S***troke**

Cerebral hemorrhage is a general term for a range of disruptive brain hemorrhagic disorders with high mortality. Mainly two categories are included, intracerebral hemorrhage (ICH) and subarachnoid hemorrhage (SAH).

#### **Intracerebral Hemorrhage**

ICH is the most severe type of stroke. Although the incidence of ICH is less than ischemic stroke [[92](#page-15-0)], it often brings the poorest outcomes and the highest morbidity among stroke subtypes [\[118](#page-16-16)]. More grimly, up to half of the survivors will have a recurrent hemorrhagic or ischemic stroke, sudden cognitive decline, or sudden depression within five years after the brain hemorrhage [[119\]](#page-16-17). Controlling blood pressure is the most efective way to slow or possibly prevent this progression [[120\]](#page-16-18). Therefore, searching for efective early treatment is extremely important for patients with ICH. Rapidly formed hematomas could directly lead to brain damage and neurological defcits after the onset of ICH [\[121](#page-16-19)]. Thus, it was thought that the removal of the hematoma might be benefcial for ICH patients. However, the surgical trial in intracerebral hemorrhage (STICH) failed to show any overall efficacy of early hematoma clearance compared to initial conservative therapies [[122](#page-16-20)]. Eight years later, the STICH II trial yielded the same results: early surgery could not improve the outcomes of ICH patients in terms of the death rate and disability at six months [[123](#page-16-21)]. Since efective treatments for the primary injury are still lacking, the secondary brain injury caused by metabolites or components of hematoma is of increasing interest to researchers. During this phase, the hematoma triggers a series of pathological changes, including cytotoxicity, apoptosis, oxidative stress, and infammations, leading to neurological function deterioration [[124\]](#page-16-22). Of note, controlling the infammatory process may be a potentially beneficial strategy for treating ICH [[125\]](#page-16-23).

Converging evidence shows that NLRP3 infammasome plays a critical role in ICH-induced secondary brain injury and neuroinfammation. Previous studies have demonstrated that NLRP3 infammasome was involved in the infammatory response after ICH [[62](#page-14-12)]. The ATP-gated transmembrane cation channel P2X7R was known as a key mediator in the upstream of NLRP3 infammasome activation and cytokines release [[28\]](#page-13-12). P2X7R and NLRP3 infammasomes were activated sequentially in rats with collagenase-induced cerebral striatum hemorrhage, which promoted the generation of NLRP3 infammasome-dependent pro-infammatory cytokines, such as IL-1 $\beta$  and IL-18, and propelled the onset of brain damage subsequently [\[62\]](#page-14-12). Meanwhile, the neuroinfammation after ICH could be alleviated by blue brilliant G (BBG), a selective P2X7R inhibitor, by inhibiting the P2X7R/NLRP3 infammasome pathway in rats [[62](#page-14-12)]. Researchers further confrmed that activation of NLRP3 infammasome could enhance infammatory response, promote neutrophil infltration, aggravate brain edema, and deteriorate neurological function in mice after ICH [[126](#page-16-24)]. Another research showed that NLRP3 infammasome was essential for complement-induced neuroinfammation after ICH, which played a key role in ICH-induced neurological dysfunction [\[127\]](#page-16-25). As a crucial transcription factor, Nrf2 could suppress the expression of NLRP3 and the subsequent clearance of ROS, thereby mitigating early brain injury [\[128](#page-16-26)]. Upregulation of the Nrf2-mediated signaling pathway leading to NLRP3 inactivation by Silymarin could produce a neuroprotective efect in rats with collagenase-induced ICH [[76\]](#page-15-12). Similar results were observed in other recent studies. Intraperitoneal injection of Ghrelin can protect mice from the secondary brain injury after ICH by inhibiting the activation of NLRP3 infammasome and promoting Nrf2/ARE signaling [[129\]](#page-16-27). Although the relationship between Nrf2 and NLRP3 infammasomes needs further investigation, these studies provide experimental evidence for the involvement of Nrf2 in the regulation of NLRP3 to some extent. The other underlying mechanisms regulating NLRP3-mediated infammation in ICH injury included peroxynitrite [[62](#page-14-12)] and mitophagy [[130\]](#page-16-28). A recent study indicated that NLRP3 infammasome might be regulated via the JAK2-STAT3-A20 pathway after ICH in C57BL/6 mice [[131\]](#page-17-0). Evidence further showed that selective NLRP3 infammasome inhibitor MCC950 can efectively down-regulate the expression of IL-1β and IL-6 by microglia in mouse models with ICH induced by autologous blood and bacterial collagenase [\[58](#page-14-22)]. Moreover, the intervention of MCC950 presented signifcant neuroprotective efects. Intraperitoneally injected MCC950 (10 mg/kg) in mice after ICH reduced cell death in brain tissue, maintained the integrity of the blood–brain barrier, and improved long-term prognosis [\[58\]](#page-14-22). Interestingly, the beneficial effects of MCC950 were eliminated after microglia depletion [[58](#page-14-22)].

In addition to small-molecule inhibitors, post-transcriptional modifcation by specifc RNA interference (RNAi) or microRNA (miRNA) on the expression of NLRP3 showed equivalent neuroprotection in animal models with ICH. NLRP3 RNAi could inhibit the inflammatory response induced by erythrocytelysis, alleviate microglia-mediated neuronal injury, and improve neurological dysfunction after ICH [[132](#page-17-1)]. Previous research has suggested that miR-223 can down-regulate NLRP3 and suppress infammation, thus reducing brain edema and improving neurological function [\[84\]](#page-15-20). Moreover, a recent study showed that miR-152 could inhibit the TXNIP-mediated activation of NLRP3 infammasome, thereby attenuating neuroinfammation, brain injury, and neurological deficits in the collagenase-induced ICH rats [\[85\]](#page-15-21).

Resveratrol, a plant polyphenol complex, has been shown to induce autophagy and inhibit mitochondrial damage in macrophages, thereby inhibiting NLRP3 infammasome activation, NLRP3-mediated IL-1β release, and pyroptosis [\[133](#page-17-2)]. Several studies have shown that resveratrol can reduce neuroinfammation and alleviate secondary brain injury after ICH [[134](#page-17-3)]. As discussed in other parts of our article, resveratrol already showed benefcial efects in Alzheimer's disease [[73\]](#page-15-27), cerebral ischemia [\[72](#page-14-21)], and SAH [[74\]](#page-15-10), which were all closely associated with the regulation of NLRP3. Nevertheless, whether resveratrol also exerts anti-infammatory effects through the inhibition of the NLRP3 inflammasome remains unclear in these studies of ICH and requires further research. In conclusion, early inhibition targeting the NLRP3 may be a potential strategy for the treatment of ICH.

#### **Subarachnoid Hemorrhage**

Subarachnoid hemorrhage (SAH), mainly caused by aneurysms rupture, has a mortality and disability rate of up to 25–50% [[135](#page-17-4)] while only accounts for 3% of all strokes [[92](#page-15-0)]. Early brain injury and delayed cerebral ischemia are two major factors afecting functional outcomes after SAH. Previous and recent research on SAH emphasizes the importance of infammation in aneurysm formation and rupture [[136](#page-17-5), [137](#page-17-6)]. Chronic infammation is also thought to be a promising drug therapy target to stabilize unruptured intracranial aneurysms or prevent the formation of unruptured intracranial aneurysms [\[138\]](#page-17-7). A previous study observed that the P2X7R/cryopyrin inflammatory axis might promote the production of IL-1 β/IL-18 after SAH by activating caspase-1, thus promoting the formation of neuroinfammation [[139](#page-17-8)]. Recent research showed that in the rat model with SAH, melatonin treatment could attenuate early brain injury, which was closely associated with the inhibition of NLRP3 infammasome activation [[91](#page-15-26)]. The pro-infammatory cytokine levels and ROS production were also signifcantly reduced. Evidence also proved that MCC950, a selective NLRP3 inhibitor, could alleviate early brain injury caused by SAH in animal models [[140](#page-17-9), [141](#page-17-10)]. Intraperitoneal injection of MCC950 with a dose of 10 mg/ kg reduced neuroinfammation and improved SAH-induced neurological dysfunction in Sprague–Dawley rats [[141](#page-17-10)]. Resveratrol, as mentioned above, also protected rats and the primary cultured cortical neurons from early brain injury after SAH, at least in part through suppressing the NLRP3 infammasome signaling pathway [[74\]](#page-15-10). It was also suggested that Pterostilbene, a polyphenol mainly derived from blueberries and grapes, could reduce early brain injury possibly through the inhibition of NLRP3 infammasome and Nox2 related oxidative stress [\[77](#page-15-13)]. Moreover, Schisandrin B signifcantly improved the neurological function in rats with SAH, which was related to the inhibition of NLRP3 and neuroinfammation [[142](#page-17-11)]. Inhibition of NLRP3 alleviated cerebral edema, microthrombosis, tight junction disruption, and microglial activation but also prevented middle cerebral artery vasospasm during the delayed period and attenuated sensorimotor disturbances caused by SAH [[140\]](#page-17-9). Furthermore, suppression of the NLRP3 infammasome pathway would promote neurogenesis action following SAH and improve the impaired delayed neurological dysfunction [[143,](#page-17-12) [144](#page-17-13)]. However, the mechanism needs further studies. It has been demonstrated that NLRP3-dependent neuroinfammation could be mediated by the TGR5/cAMP/PKA signaling pathway [[144\]](#page-17-13). In experimental models of SAH induced by endovascular perforation, the NLRP3 infammasome could be activated by triggering receptor expressed on myeloid cells 1 (TREM-1) and mediate microglial pyroptosis [\[145](#page-17-14)].

#### **Alzheimer Disease**

Alzheimer's disease (AD) is the primary dementia disease in the world, which is characterized by progressive cognitive and behavioral disorders. According to the latest report of Alzheimer's Disease International (ADI), up to 6.5 million Americans (>age 65) are sufering from Alzheimer's dementia today. An estimated \$321 billion of total pay-ments were cost [\[146](#page-17-15)]. However, in contrast to the severity of the disease, despite more than a century of exploration, there have been few breakthroughs in drug development for Alzheimer's disease, and many promising drug developments failed [[147\]](#page-17-16). A new direction for drug exploration is required.

The converging pathological features of AD are extracellular senile plaques formed by amyloid ß (Aβ) protein, intracellular neurofbrillary tangles composed of the hyperphosphorylation of tau protein, and the loss of neurons [[148\]](#page-17-17). These pathological changes are closely related to innate immune abnormalities in the brain [[149\]](#page-17-18). Furthermore, it has been identifed that NLRP3 activation contributes to AD without equivocation. The expression of NLRP3 and downstream inflammatory cytokines (IL-1 $\beta$  and IL-18) was elevated in peripheral monocytes in severe AD patients [[150\]](#page-17-19). Similar elevated expression patterns were observed in the temporal cerebral cortex of AD patients by Mahmoudiasl et al. [[151](#page-17-20)]. Almost two decades ago, Aβ, as a kind of DAMPs, had been identifed as the direct activator of the NLRP3 infammasome, leading to the maturation and release of IL-1β for the frst time [[16\]](#page-13-0). The mechanism involved included lysosomal damage and cathepsin B release [\[16](#page-13-0)]. Subsequently, considerable research was carried out. It was further suggested that the activation of NLRP3 infammasome in microglia exacerbated Aβ pathology  $[59]$  $[59]$  by prion-like ASC-speck cross-seeding [[152\]](#page-17-21). Despite fibrillar  $\mathbf{A}\beta$ , it was reported recently that lower molecular weight Aβ oligomers and protofbrils could also trigger the activation of NLRP3, suggesting that innate immune response induced by Aβ species may occur before the initiation of Aβ deposition [[153](#page-17-22)]. NLRP3 inflammasome deficiency would reduce Aβ deposition in the amyloid precursor protein/presenilin 1 (APP/PS1) model of AD [\[49](#page-14-2), [59\]](#page-14-23). Furthermore, neuroinfammation induced by activation of NLRP3 inflammasome impaired  $\text{A}$ β clearance in microglia [[154](#page-17-23)].

Presently, Aβ-activated NLRP3 infammasomes participating in the pathogenesis of AD are convinced, while the relationship between tau and NLRP3 infammasome is more limited studied. Recently, it was confrmed that the activation of NLPR3 in microglia could promote the pathological changes of tau protein, which was conducive to the onset of AD [\[155](#page-17-24)]. Meanwhile, NLRP3 could be activated by tau oligomers and monomers in microglia directly and mediate Aβ-induced tau pathology. Interestingly, tau protein also participated in the priming step for the activation of NLRP3 infammasome [[156,](#page-17-25) [157\]](#page-17-26). NLRP3-defcient mice showed decreased tau pathology along with improved cognitive function. Further research demonstrated that aggregated tau could activate the NLRP3 infammasome in primary microglia [\[158\]](#page-17-27). Tau-induced pathological responses can be alleviated in tau transgenic mice defcient for ASC or by the treatment of NLRP3 inhibitors [[158](#page-17-27)]. The latest study further emphasized the importance of NLRP3 infammasome and suggested that the Drp1-HK1-NLRP3 signal axis may play a vital role in the white matter degeneration in AD [[159\]](#page-17-28).

Dynamin-related protein 1 (Drp1) is a mitochondrial fission guanosine triphosphatase. Knock-down of Drp1 in mature oligodendrocytes-specifc heterozygous could inhibit the activation of NLRP3 infammasome, correct myelin degeneration and axonal loss, and improve cognitive function. White matter loss often presents early in the course of AD and is hypothesized to be the initial affair in AD pathology. The fndings may provide new ideas for the treatment of AD.

Given that the NLRP3 inflammasome signaling axis may play an essential role in the pathogenesis of AD, targeting NLRP3 as a potential treatment for AD attracted more attention and has been discussed extensively. Here, we will primarily discuss the latest research, especially the pharmaceutical research of Chinese traditional herb-derived compounds.

Numerous pieces of research demonstrated that MCC950 could reduce Aβ-induced pathological events and improve cognitive function. MCC950 can inhibit the activation of NLRP3 infammasome in microglia, ameliorate the impairment of synaptic plasticity, and reduce the accumulation of Aβ  $[160]$  $[160]$ . Meanwhile, the inactivation of NLRP3 inflammasome mediated by MCC950 could attenuate the reactivity of microglia induced by Aβ1-42 oligomers and improve memory impairment [\[161](#page-18-0)]. MCC950 could also completely suppress the immune response mediated by the NLRP3 infammasome pathway which was induced by fbrils and low molecular weight  $\text{A}$ β aggregates [[153](#page-17-22)]. In addition, the activation of IL-1 $\beta$  induced by tau aggregates was also prevented by MCC950, as well as the tau-mediated pathological changes [\[158](#page-17-27)]. However, the phase II clinical trial of MCC950 was suspended due to the possible hepatotoxicity in rheumatoid arthritis [\[162](#page-18-1)]. Drug improvement based on MCC950 to reduce the side effect may be the next potential strategy. Bay 11–7082 is another inhibitor of the NF-κB/ NLRP3 pathway. The pre-administration of Bay 11–7082 can block the activation of the NLRP3 infammasome and attenuate neuronal damage and cognitive dysfunction in aged rats [[63\]](#page-14-13). In APP23 mice treated with kainic acid (KA), BAY 11–7082 protected them from KA-induced neuronal degeneration and Aβ deposition, thereby improving their cognitive function [\[64](#page-14-24)]. VX-765 is a reversible caspase-1 inhibitor that has been found to cross the blood–brain barrier and exert neuroprotective efects. In the mouse model of AD, treatment with VX-765 could reduce neuroinfammation and deposition of Aβ plaques, ultimately improving the neurocognitive function [\[67](#page-14-16)]. Unfortunately, the clinical trial of VX-765 was halted due to its hepatotoxicity and immunosuppression [[162](#page-18-1)]. The more inspiring discovery came from fenamate non-steroidal anti-infammatory drugs (NSAIDs), compounds already approved by the FDA for other treatments, which could also act as selective inhibitors of NLRP3 infammasome [\[87](#page-15-23)]. The results showed that fenamate NSAIDs could inhibit NLRP3 activation by blocking

the volume-regulated anion channel (VRAC), independently of cyclooxygenase (COX) enzymes. Importantly, treatment with mefenamic acid protected the mice from Aβ-induced memory deficits and NLRP3-mediated neuroinflammation in two animal models of AD [[87](#page-15-23)]. These encouraging fndings in animal studies may provide a more rapid breakthrough in clinical trials. Other NLRP3 infammasome-specifc inhibitors, such as inzomelid (Infazome/Roche) and NT-0167 (NodThera/Roche), are currently undergoing clinical trials. A recent fnding showed that miR-22 expression levels were lower in peripheral blood of AD patients than in healthy subjects. Moreover, miR-22 attenuated cognitive dysfunction in AD mice by targeting GSDMD to modulate glial cell pyroptosis, inhibit the activation of NLRP3 infammasome, and decrease infammatory cytokine release [\[86\]](#page-15-22).

In addition, many plant extracts and traditional Chinese medicines have also been shown to have neuroprotective efects in AD by modulating the NLRP3 infammasome pathway. Dl-3-n-butylphthalide (Dl-NBP) is the active ingredient extracted from Chinese herbal celery seeds. It is not only a commonly used drug for the treatment of ischemic stroke but also a novel therapeutic approach for neurodegenerative diseases. TXNIP has been demonstrated a key role in the activation of the NLRP3 infammasome [[163\]](#page-18-2). In APP/PS1 mice, Dl-NBP treatment could suppress TXNIP-NLRP3 interaction, inhibit NLRP3 infammasome-mediated infammatory damage by up-regulating Nrf2, ameliorate neuronal apoptosis, and thereby reduce oxidative stress damage [[78\]](#page-15-14). Achyranthes bidentate is a traditional Chinese medicine with anti-infammatory and antioxidant features and has been used for the treatment of dementia for a long time. A recent study showed that Achyranthes bidentate polypeptide fraction κ (ABPPκ) decreased Aβ oligomerinduced IκBα phosphorylation and NLRP3 expression in BV2 microglia. *In vivo*, pre-treatment of ABPPk reduced the expression of NLRP3, cleaved caspase-1, and ASC in the brain, promoted the polarization of M2 phenotype in microglia, and thus attenuated the memory impairment and the loss of hippocampal neurons in mice [[80\]](#page-15-16). Resveratrol is a polyphenol compound derived from natural plants mainly in red grape skins and wine. It was reported that resveratrol could reverse the mitochondrial dysfunction by decreasing the expression of NF-κB/NLRP3/IL-1β, ameliorate learning and memory impairment, and reduce the neural injury in the hippocampus in the  $A\beta1-42$ -induced mouse model of AD [[73](#page-15-27)]. Another recent study highlighted the crucial role of the low-density lipoprotein receptor-related protein 1 (LRP1) in the regulation of the endocytosis of tau and its spread [\[164](#page-18-3)]. Polyphenol (LSP) extracted from lychee seed could inhibit NLRP3 infammasome-mediated neuroinfammation *in vitro* and improve cognitive function in APP/PS1 mice. It was further clarifed that the molecular mechanism of LSP was mediated by up-regulating the expression of the

LRP1/AMPK signaling pathway and enhancing the microglial autophagy [[81\]](#page-15-17). In conclusion, considering their good safety and fewer side effects, patients with AD may benefit from NLRP3-targeting traditional Chinese herbal medicines, but it requires more research.

## **Parkinson's Disease**

It is suggested that the NLRP3 infammasome may play a critical role in the pathogenesis of Parkinson's disease (PD) [\[68](#page-14-17)]. α-Synuclein (αSyn) is the pathologic feature protein of PD whose dysfunction is closely related to PD pathogenesis. Neuronal uptake of αSyn leads to impaired mitochondrial function [[165\]](#page-18-4) and the following release of excess mitochondrial reactive oxygen species (mtROS), which could directly activate the NLRP3 infammasome in macrophages and microglia  $[166]$  $[166]$ . It was further clarified that  $\alpha$ Syn could potently trigger the LPS-independent priming and activation of NLRP3 infammasome assembly in microglia both *in vivo* and *in vitro* models of PD [\[17\]](#page-13-1). Expression of the NLRP3 infammasome component was generally increased in PD animal models [\[18](#page-13-2), [71](#page-14-20)], as well as in brain tissues [[18\]](#page-13-2) and peripheral blood samples [[167](#page-18-6), [168](#page-18-7)] from PD patients. *Nlrp3* knockout mice sufered less infammatory cascade response and hazardous neurotoxicity than *Nlrp3*-expressing wild-type ones [[71](#page-14-20)]. Moreover, NLRP3 deletion markedly reduced motor dysfunction and dopaminergic neurodegeneration in the MPTP-induced mouse model of PD [[71\]](#page-14-20). Similar results were observed in the MCC950-treated mouse. The administration of MCC950 in various animal PD models inhibited the activation of NLRP3 inflammasome and efficiently attenuated motor impairment, nigrostriatal dopaminergic degeneration, and the aggregation of  $\alpha$ -synaptic nucleoproteins [\[18](#page-13-2)]. Inhibition of the downstream of the NLRP3/caspase-1/ IL-1β pathway by *caspase-1* knockout or caspase-1 inhibitor also alleviated dopaminergic neuronal loss and dyskinesia in murine models under PD conditions [[68,](#page-14-17) [69](#page-14-18)]. In addition, the blockade of IL-1 receptors by IL-1 receptor antagonist (IL-1Ra) signifcantly reduced the degeneration of dopaminergic neurons and motor deficits *in vivo* [\[71\]](#page-14-20). DI-NBP has been used in the treatment of acute ischemic stroke in China. Interestingly, both *in vivo* and *in vitro*, Dl-NBP inhibited NLRP3 infammasome activation and α-Syn aggregation, thereby suppressing neuroinfammation, ameliorating mitochondrial damage, and rescuing dopaminergic neurons [[79\]](#page-15-15). Furthermore, Perillyl Alcohol, a monoterpene compound found in the essential oils of plants, could support neuronal survival and improve behavioral activities in the mouse via inhibiting the NLRP3 infammasome activation, thereby elevating levels of various antioxidant enzymes and restoring levels of dopamine and other neurotransmitters in the striatum [\[82](#page-15-18)]. These latest studies highlight the role of NLRP3 infammasome as a potential target for PD treatment.

#### **Multiple Sclerosis**

Previous studies demonstrated that in patients with relapseremitting multiple sclerosis (RRMS), the expression levels of NLRP3 and IL-1 $\beta$  were significantly different in responders and non-responders with the administration of interferon beta (IFN-β) treatment [[169\]](#page-18-8). However, this result is still controversial. Some studies came to similar conclusions [[170](#page-18-9)], and others came to the opposite ones. In contrast, Sunny Malhotra et al.'s fndings were against the involvement of polymorphisms located in the *Nlrp3* gene and the response to IFN-β in multiple sclerosis (MS) patients [[171](#page-18-10)]. Recently, it was shown that the presence of IL-1 $\beta$ signaling in peripheral blood cells of patients with primary progressive MS (PPMS) rather than relapsed clinical types [ $172$ ]. This IL-1 $\beta$  signature was related to the upregulation of an overactive NLRP3 infammasome. More importantly, NLRP3 infammasome was suggested to act as a pathogenic factor in MS. PPMS patients with higher IL-1 $\beta$  levels in blood cells had accelerated disease progression and required walking-help ten years earlier than those with lower IL-1β levels [[172](#page-18-11)].

Experimental autoimmune encephalomyelitis (EAE) is an ideal disease model of human MS. Earlier research indicated that MCC950 treatment could reduce IL-1β generation *in vivo* and attenuate the severity of EAE [[45\]](#page-13-29). The results were reaffirmed by Malhotra et al. Mice treated with MCC950 were associated with less disease severity in EAE and reduced LPS-induced axonal damage [[172](#page-18-11)]. Oral treatment with MCC950 (50 mg/kg/day) halted disease relapses and reversed MS-associated central neuropathic pain in the relapsing–remitting EAE mouse model [\[173](#page-18-12)]. More interestingly, NLRP3 expression was observed in astrocytes, microglia, and neurons, although only microglia and astrocytes displayed IL-1β immunoreactivity [\[172](#page-18-11)]. Another small-molecule NLRP3 infammasome inhibitor, JC-171, has been identifed to reduce NLRP3-mediated IL-1β secretion in a dose-dependent manner by disrupting NLRP3-ASC interactions in both *in vitro* and *in vivo* EAE models [[55](#page-14-8)]. Manoalide is a newly identifed selective NLRP3 inhibitor that acts by blocking the NEK7–NLRP3 interaction [[70\]](#page-14-19). Manoalide treatment could relieve the infammation and attenuate the pathogenesis of EAE in mice [\[70](#page-14-19)].

## **Other Neurological Disorders**

## **Traumatic brain injury (TBI)**

TBI provokes an infammatory cascade including potassium and chloride efflux, ATP transferred to the extracellular space, alteration of calcium signaling, mitochondrial injury, and ROS release, all of which eventually trigger the NLRP3 infammasome activation [[174\]](#page-18-13). Treatment of MCC950 in animals at the acute stages of TBI could reduce microglial activation, maintain the integrity of the blood–brain barrier, attenuate NLRP3-mediated neuroinfammation, and improve neuro-logical outcomes [[50](#page-14-3), [51](#page-14-4)]. Of note, the efficacy of MCC950 was inhibited in microglia-depleted TBI animals [[50\]](#page-14-3). BAY 11–7082, which selectively inhibits the NLRP3 infammasome through interaction with NLRP3 ATPase, significantly attenuated infammatory infltration and damage in the cortex and hippocampus of murine within 24 h after TBI [[65](#page-14-14)]. The synergistic effect of Bay 11–7082 and dexmedetomidine was further found in inhibiting NLRP3/caspase-1 axis activity and improving neurological dysfunction following TBI [[175](#page-18-14)]. The study also indicated that the knockout of NLRP3 presented overlapping outcomes. NLRP3-defcient mice showed preserved cognitive function and less severe brain damage compared to wild-type mice [\[65](#page-14-14)].

#### **Epilepsy**

NLRP3 infammasome activation has been reported to contribute to the pathogenesis of seizures. The elevated expression of NLRP3 infammasome was observed in the hippocampus of diferent epilepsy murine models [\[176](#page-18-15), [177\]](#page-18-16) and in the hippocampus of patients with mesial temporal lobe epilepsy [[178\]](#page-18-17). NLRP3 knockdown in amygdala kindling-induced status epilepticus rat models was associated with reduced expression of IL-1 $\beta$  and caspase-1, along with improved seizure severity [\[177](#page-18-16)]. It was further clarifed that NLRP3 knockout also protected mouse and cellular epilepsy models from neuronal apoptosis [\[60](#page-14-25)]. Similar results were observed in MCC950-treated cellular models of epilepsy [\[60\]](#page-14-25). Unfortunately, as previously described, phase II clinical trials of two caspase-1-inhibiting peptidomimetic prodrugs, VX-740 and VX-765, for the treatment of arthritis, epilepsy, and psoriasis were terminated due to liver toxicity [[162](#page-18-1)].

#### **Others**

In murine cardiac arrest (CA) and cardiopulmonary resuscitation (CPR) models, MCC950-treated mice showed improvements in functional recovery and survival rates [[179\]](#page-18-18). The conclusion was further reconfrmed in another study by intraperitoneal injection of MCC950 (10 mg/ kg) after the return of spontaneous circulation (ROSC) in models of CA. Compared with the control group, cerebral microcirculation, cerebral edema, 24-h survival rate, and neurological deficits were remarkably improved in animals treated with MCC950 [\[180](#page-18-19)]. In addition, NLRP3-mediated neuronal pyroptosis contributed to the pathologic process of sepsis-associated encephalopathy (SAE). Administration of MCC950 could reverse these efects and rescue cognitive deficits in mouse models of SAE  $[61]$  $[61]$ . MCC950 could also attenuate infammation, neuronal apoptosis, and mitochondrial disorders via the Nrf2 pathway in SAE mice [\[181\]](#page-18-20). Nevertheless, the underlying mechanisms of NLRP3 in those diseases require more investigation.

# **Conclusion and Prospect for the Future**

Activation of NLRP3 infammasome is a pathological hallmark common to numerous immune-mediated and degenerative neurological diseases. NLRP3 has been shown to impact various neurological disease models in mice, highlighting the potential application of NLRP3-targeted therapy for these diseases. Several compounds interfering with the NLRP3 infammasome pathway have exhibited therapeutic efects in preclinical trials. Due to the genetic and biological intricacy of neurological disorders such as AD and PD, a single approach may not be efficient for their treatments. Thus infammasome inhibitors may be able to work in conjunction with other drugs and support their therapeutic outcomes. Considering the safety and fewer side effects, traditional Chinese medicines, as well as plant-derived ingredients also offer innovative perspectives and alternative options for intractable conditions. However, despite decades of research, our understanding of NLRP3 infammasome in central nervous system diseases is still lacking. A better appreciation of the physiological mechanisms regulating NLRP3 infammasome will lead to a better assessment of the therapeutic potential of infammasome inhibitors in human neurological disorders. We also expect that targeting NLRP3 infammasome of the neurological diseases will be applied to the clinical treatment soon, so as to beneft more patients.

**Author Contribution** All authors contributed to the study conception and design. Wenfang He had the idea for the article and prepared the draft of the manuscript. Literature search and data analysis were performed by Yanjun Zhong and Chenfang Wu. Jinxiu Li and Zhiping Hu reviewed and critically revised the manuscript. All authors read and approved the fnal manuscript.

**Funding** The study was supported by the Scientifc Research Project of Hunan Provincial Health Commission (202117010086) and Natural Science Foundation of Hunan Province, China (Grant No. 2021JJ40832).

**Data Availability** Data sharing not applicable to this article as no datasets were generated or analyzed during the current study.

# **Declarations**

**Ethics Approval** This is not applicable.

**Consent to Participate** This is not applicable.

**Consent for Publication** This is not applicable.

**Competing Interests** The authors declare no competing interests.

## **References**

- <span id="page-12-0"></span>1. Lu A, Magupalli VG, Ruan J, Yin Q, Atianand MK, Vos MR, Schroder GF, Fitzgerald KA et al (2014) Unifed polymerization mechanism for the assembly of ASC-dependent infammasomes. Cell 156(6):1193–1206. [https://doi.org/10.1016/j.cell.2014.02.](https://doi.org/10.1016/j.cell.2014.02.008) 00<sub>8</sub>
- <span id="page-12-1"></span>2. Fernandes-alnemri T, Wu J, Yu J, Datta P, Miller B, Jankowski W, Rosenberg S, Zhang J et al (2007) The pyroptosome: a supramolecular assembly of ASC dimers mediating infammatory cell death via caspase-1 activation. Cell Death Difer 14(9):1590–1604
- <span id="page-12-2"></span>3. Hoss F, Rodriguez-alcazar JF, Latz E (2017) Assembly and regulation of ASC specks. Cell Mol Life Sci 74(7):1211–1229. <https://doi.org/10.1007/s00018-016-2396-6>
- <span id="page-12-3"></span>4. Broz P, Dixit VM (2016) Infammasomes: mechanism of assembly, regulation and signalling. Nat Rev Immunol 16(7):20–407. <https://doi.org/10.1038/nri.2016.58>
- <span id="page-12-4"></span>5. Duncan JA, Bergstralh DT, Wang Y, Willingham SB, Ye Z, Zimmermann AG, Ting JP (2007) Cryopyrin/NALP3 binds ATP/ dATP, is an ATPase, and requires ATP binding to mediate infammatory signaling. Proc Natl Acad Sci U S A 104(19):6–8041
- <span id="page-12-5"></span>6. Hofman HM, Mueller JL, Broide DH, Wanderer AA, Kolodner RD (2001) Mutation of a new gene encoding a putative pyrinlike protein causes familial cold autoinfammatory syndrome and Muckle-Wells syndrome. Nat Genet 29(3):301–305
- 7. Aksentijevich I, Nowak M, Mallah M, Chae JJ, Watford WT, Hofmann SR, Stein L, Russo R et al (2002) De novo CIAS1 mutations, cytokine activation, and evidence for genetic heterogeneity in patients with neonatal-onset multisystem infammatory disease (NOMID): a new member of the expanding family of pyrin-associated autoinfammatory diseases. Arthritis Rheum 46(12):3340–3348
- <span id="page-12-6"></span>8. Aganna E, Martinon F, Hawkins PN, Ross JB, Swan DC, Booth DR, Lachmann HJ, Bybee A et al (2002) Association of mutations in the NALP3/CIAS1/PYPAF1 gene with a broad phenotype including recurrent fever, cold sensitivity, sensorineural deafness, and AA amyloidosis. Arthritis Rheum 46(9):2445–2452
- <span id="page-12-7"></span>9. Andreeva L, David L, Rawson S, Shen C, Pasricha T, Pelegrin P, Wu H (2021) NLRP3 cages revealed by full-length mouse NLRP3 structure control pathway activation. Cell 184(26):6299- 6312.e22. <https://doi.org/10.1016/j.cell.2021.11.011>
- <span id="page-12-8"></span>10. Guo C, Chi Z, Jiang D, Xu T, Yu W, Wang Z, Chen S, Zhang L et al (2018) Cholesterol homeostatic regulator SCAP-SREBP2 integrates NLRP3 infammasome activation and cholesterol biosynthetic signaling in macrophages. Immunity 49(5):842-856.e7. <https://doi.org/10.1016/j.immuni.2018.08.021>
- <span id="page-12-9"></span>11. Chen J, Chen ZJ (2018) PtdIns4P on dispersed trans-Golgi network mediates NLRP3 inflammasome activation. Nature 564(7734):71–76. <https://doi.org/10.1038/s41586-018-0761-3>
- <span id="page-12-10"></span>12. He Y, Zeng MY, Yang D, Motro B, Nunez G (2016) NEK7 is an essential mediator of NLRP3 activation downstream of potassium efflux. Nature 530(7590):354-357. [https://doi.org/10.1038/](https://doi.org/10.1038/nature16959) [nature16959](https://doi.org/10.1038/nature16959)
- <span id="page-12-11"></span>13. Shi H, Wang Y, Li X, Zhan X, Tang M, Fina M, Su L, Pratt D et al (2016) NLRP3 activation and mitosis are mutually exclusive events coordinated by NEK7, a new infammasome component. Nat Immunol 17(3):250–258. <https://doi.org/10.1038/ni.3333>
- <span id="page-12-12"></span>14. Maturana CJ, Aguirre A, Saez JC (2017) High glucocorticoid levels during gestation activate the infammasome in hippocampal oligodendrocytes of the ofspring. Dev Neurobiol 77(5):625–642. <https://doi.org/10.1002/dneu.22409>
- <span id="page-12-13"></span>15. Johann S, Heitzer M, Kanagaratnam M, Goswami A, Rizo T, Weis J, Troost D, Beyer C (2015) NLRP3 infammasome is

expressed by astrocytes in the SOD1 mouse model of ALS and in human sporadic ALS patients. Glia 63(12):2260–2273. [https://](https://doi.org/10.1002/glia.22891) [doi.org/10.1002/glia.22891](https://doi.org/10.1002/glia.22891)

- <span id="page-13-0"></span>16. Halle A, Hornung V, Petzold GC, Stewart CR, Monks BG, Reinheckel T, Fitzgerald KA, Latz E et al (2008) The NALP3 inflammasome is involved in the innate immune response to amyloidbeta. Nat Immunol 9(8):65–857.<https://doi.org/10.1038/ni.1636>
- <span id="page-13-1"></span>17. Panicker N, Sarkar S, Harischandra DS, Neal M, Kam T, Jin H, Saminathan H, Langley M et al (2019) Fyn kinase regulates misfolded alpha-synuclein uptake and NLRP3 infammasome activation in microglia. J Exp Med 216(6):1411–1430. [https://](https://doi.org/10.1084/jem.20182191) [doi.org/10.1084/jem.20182191](https://doi.org/10.1084/jem.20182191)
- <span id="page-13-2"></span>18. Gordon R, Albornoz EA, Christie DC, Langley MR, Kumar V, Mantovani S, Robertson AAB, Butler MS et al (2018) Infammasome inhibition prevents alpha-synuclein pathology and dopaminergic neurodegeneration in mice. Sci Transl Med 10(465). <https://doi.org/10.1126/scitranslmed.aah4066>
- <span id="page-13-3"></span>19. Wu A, Zhou X, Qiao G, Yu L, Tang Y, Yan L, Qiu W, Pan R et al (2021) Targeting microglial autophagic degradation in NLRP3 infammasome-mediated neurodegenerative diseases. Ageing Res Rev 65:101202.<https://doi.org/10.1016/j.arr.2020.101202>
- <span id="page-13-4"></span>20. Hong P, Gu R, Li F, Xiong X, Liang W, You Z, Zhang H (2019) NLRP3 infammasome as a potential treatment in ischemic stroke concomitant with diabetes. J Neuroinfammation 16(1):121. <https://doi.org/10.1186/s12974-019-1498-0>
- <span id="page-13-5"></span>Franchi L, Eigenbrod T, Nunez G (2009) Cutting edge: TNFalpha mediates sensitization to ATP and silica via the NLRP3 infammasome in the absence of microbial stimulation. J Immunol 183(2):6–792. <https://doi.org/10.4049/jimmunol.0900173>
- <span id="page-13-6"></span>22 Xing Y, Yao X, Li H, Xue G, Guo Q, Yang G, An L, Zhang Y et al (2017) Cutting edge: TRAF6 mediates TLR/IL-1R signaling-induced nontranscriptional priming of the NLRP3 infammasome. J Immunol 199(5):1561–1566. [https://doi.org/10.4049/](https://doi.org/10.4049/jimmunol.1700175) [jimmunol.1700175](https://doi.org/10.4049/jimmunol.1700175)
- <span id="page-13-7"></span>23. Bauernfeind FG, Horvath G, Stutz A, Alnemri ES, Macdonald K, Speert D, Fernandes-alnemri T, Wu J et al (2009) Cutting edge: NF-kappaB activating pattern recognition and cytokine receptors license NLRP3 infammasome activation by regulating NLRP3 expression. J Immunol 183(2):787–91. [https://doi.org/10.4049/](https://doi.org/10.4049/jimmunol.0901363) [jimmunol.0901363](https://doi.org/10.4049/jimmunol.0901363)
- <span id="page-13-8"></span>24. Greaney AJ, Leppla SH, Moayeri M (2015) Bacterial exotoxins and the infammasome. Front Immunol 6:570. [https://doi.org/10.](https://doi.org/10.3389/fimmu.2015.00570) [3389/fmmu.2015.00570](https://doi.org/10.3389/fimmu.2015.00570)
- <span id="page-13-9"></span>25. Hornung V, Bauernfeind F, Halle A, Samstad EO, Kono H, Rock KL, Fitzgerald KA, Latz E (2008) Silica crystals and aluminum salts activate the NALP3 infammasome through phagosomal destabilization. Nat Immunol 9(8):56–847. [https://doi.org/10.](https://doi.org/10.1038/ni.1631) [1038/ni.1631](https://doi.org/10.1038/ni.1631)
- <span id="page-13-10"></span>26. Liston A, Masters SL (2017) Homeostasis-altering molecular processes as mechanisms of infammasome activation. Nat Rev Immunol 17(3):208–214.<https://doi.org/10.1038/nri.2016.151>
- <span id="page-13-11"></span>27. Munoz-planillo R, Kuffa P, Martinez-colon G, Smith BL, Rajendiran TM, Nunez G (2013)  $K(+)$  efflux is the common trigger of NLRP3 infammasome activation by bacterial toxins and particulate matter. Immunity 38(6):1142–1153. [https://doi.](https://doi.org/10.1016/j.immuni.2013.05.016) [org/10.1016/j.immuni.2013.05.016](https://doi.org/10.1016/j.immuni.2013.05.016)
- <span id="page-13-12"></span>28. Di A, Xiong S, Ye Z, Malireddi RKS, Kometani S, Zhong M, Mittal M, Hong Z et al (2018) The TWIK2 potassium efflux channel in macrophages mediates NLRP3 inflammasomeinduced infammation. Immunity 49(1):56-65.e4. [https://doi.](https://doi.org/10.1016/j.immuni.2018.04.032) [org/10.1016/j.immuni.2018.04.032](https://doi.org/10.1016/j.immuni.2018.04.032)
- <span id="page-13-13"></span>29. Katsnelson MA, Lozada-soto KM, Russo HM, Miller BA, Dubyak GR (2016) NLRP3 infammasome signaling is activated by low-level lysosome disruption but inhibited by extensive lysosome disruption: roles for  $K$ + efflux and Ca2+ influx. Am

J Physiol Cell Physiol 311(1):83-C100. [https://doi.org/10.1152/](https://doi.org/10.1152/ajpcell.00298.2015) [ajpcell.00298.2015](https://doi.org/10.1152/ajpcell.00298.2015)

- <span id="page-13-14"></span>30. Murakami T, Ockinger J, Yu J, Byles V, Mccoll A, Hofer AM, Horng T (2012) Critical role for calcium mobilization in activation of the NLRP3 infammasome. Proc Natl Acad Sci U S A 109(28):11282–11287.<https://doi.org/10.1073/pnas.1117765109>
- <span id="page-13-15"></span>31. Green JP, Yu S, Martin-sanchez F, Pelegrin P, Lopez-castejon G, Lawrence CB, Brough D (2018) Chloride regulates dynamic NLRP3-dependent ASC oligomerization and infammasome priming. Proc Natl Acad Sci U S A 115(40):9371-E9380. [https://](https://doi.org/10.1073/pnas.1812744115) [doi.org/10.1073/pnas.1812744115](https://doi.org/10.1073/pnas.1812744115)
- <span id="page-13-16"></span>32 Dostert C, Petrilli V, Bruggen RV, Steele C, Mossman BT, Tschopp J (2008) Innate immune activation through Nalp3 inflammasome sensing of asbestos and silica. Science 320(5876):674–7. <https://doi.org/10.1126/science.1156995>
- <span id="page-13-17"></span>33. Zhou R, Yazdi AS, Menu P, Tschopp J (2011) A role for mitochondria in NLRP3 inflammasome activation. Nature 469(7329):221–225.<https://doi.org/10.1038/nature09663>
- <span id="page-13-18"></span>34. Iyer SS, He Q, Janczy JR, Elliott EI, Zhong Z, Olivier AK, Sadler JJ, Knepper-adrian V et al (2013) Mitochondrial cardiolipin is required for Nlrp3 infammasome activation. Immunity 39(2):311–323.<https://doi.org/10.1016/j.immuni.2013.08.001>
- <span id="page-13-19"></span>35. Liu Q, Zhang D, Hu D, Zhou X, Zhou Y (2018) The role of mitochondria in NLRP3 infammasome activation. Mol Immunol 103:115–124.<https://doi.org/10.1016/j.molimm.2018.09.010>
- <span id="page-13-20"></span>36. Zhou Y, Tong Z, Jiang S, Zheng W, Zhao J, Zhou X (2020) The roles of endoplasmic reticulum in NLRP3 infammasome activation. Cells-Basel 9(5). <https://doi.org/10.3390/cells9051219>
- <span id="page-13-21"></span>37. Tao Y, Yang Y, Zhou R, Gong T (2020) Golgi apparatus: an emerging platform for innate immunity. Trends Cell Biol 30(6):467–477.<https://doi.org/10.1016/j.tcb.2020.02.008>
- <span id="page-13-22"></span>38. Zhang Z, Meszaros G, He W, Xu Y, Magliarelli HDF, Mailly L, Mihlan M, Liu Y et al (2017) Protein kinase D at the Golgi controls NLRP3 infammasome activation. J Exp Med 214(9):2671– 2693.<https://doi.org/10.1084/jem.20162040>
- <span id="page-13-23"></span>39. Shi J, Zhao Y, Wang Y, Gao W, Ding J, Li P, Hu L, Shao F (2014) Infammatory caspases are innate immune receptors for intracellular LPS. Nature 514(7521):187–192. [https://doi.org/10.1038/](https://doi.org/10.1038/nature13683) [nature13683](https://doi.org/10.1038/nature13683)
- <span id="page-13-24"></span>40. Kayagaki N, Warming S, Lamkanf M, Walle LV, Louie S, Dong J, Newton K, Qu Y et al (2011) Non-canonical infammasome activation targets caspase-11. Nature 479(7371):117–121. [https://](https://doi.org/10.1038/nature10558) [doi.org/10.1038/nature10558](https://doi.org/10.1038/nature10558)
- <span id="page-13-25"></span>41. Kayagaki N, Stowe IB, Lee BL, O'rourke K, Anderson K, Warming S, Cuellar T, Haley B et al (2015) Caspase-11 cleaves gasdermin D for non-canonical infammasome signalling. Nature 526(7575):666–671.<https://doi.org/10.1038/nature15541>
- <span id="page-13-26"></span>42. Baker PJ, Boucher D, Bierschenk D, Tebartz C, Whitney PG, D'silva DB, Tanzer MC, Monteleone M et al (2015) NLRP3 inflammasome activation downstream of cytoplasmic LPS recognition by both caspase-4 and caspase-5. Eur J Immunol 45(10):26–2918.<https://doi.org/10.1002/eji.201545655>
- <span id="page-13-27"></span>43. Zanoni I, Tan Y, Gioia MD, Broggi A, Ruan J, Shi J, Donado CA, Shao F et al (2016) An endogenous caspase-11 ligand elicits interleukin-1 release from living dendritic cells. Science 352(6290):1232–6. <https://doi.org/10.1126/science.aaf3036>
- <span id="page-13-28"></span>44. Baldwin AG, Brough D, Freeman S (2016) Inhibiting the infammasome: a chemical perspective. J Med Chem 59(5):1691–1710. <https://doi.org/10.1021/acs.jmedchem.5b01091>
- <span id="page-13-29"></span>45. Coll RC, Robertson AAB, Chae JJ, Higgins SC, Munoz-planillo R, Inserra MC, Vetter I, Dungan LS et al (2015) A small-molecule inhibitor of the NLRP3 infammasome for the treatment of infammatory diseases. Nat Med 21(3):248–255. [https://doi.org/](https://doi.org/10.1038/nm.3806) [10.1038/nm.3806](https://doi.org/10.1038/nm.3806)
- <span id="page-13-30"></span>46. Coll RC, Hill JR, Day CJ, Zamoshnikova A, Boucher D, Massey NL, Chitty JL, Fraser JA et al (2019) MCC950 directly targets

the NLRP3 ATP-hydrolysis motif for infammasome inhibition. Nat Chem Biol 15(6):556–559. [https://doi.org/10.1038/](https://doi.org/10.1038/s41589-019-0277-7) [s41589-019-0277-7](https://doi.org/10.1038/s41589-019-0277-7)

- <span id="page-14-0"></span>47. Jiang H, He H, Chen Y, Huang W, Cheng J, Ye J, Wang A, Tao J et al (2017) Identifcation of a selective and direct NLRP3 inhibitor to treat infammatory disorders. J Exp Med 214(11):3219– 3238. <https://doi.org/10.1084/jem.20171419>
- <span id="page-14-1"></span>48. Zhai Y, Meng X, Ye T, Xie W, Sun G, Sun X (2018) Inhibiting the NLRP3 infammasome activation with MCC950 ameliorates diabetic encephalopathy in db/db mice. Molecules 23(3). [https://](https://doi.org/10.3390/molecules23030522) [doi.org/10.3390/molecules23030522](https://doi.org/10.3390/molecules23030522)
- <span id="page-14-2"></span>49. Dempsey C, Araiz AR, Bryson KJ, Finucane O, Larkin C, Mills EL, Robertson AAB, Cooper MA et al (2017) Inhibiting the NLRP3 infammasome with MCC950 promotes non-phlogistic clearance of amyloid-beta and cognitive function in APP/PS1 mice. Brain Behav Immun 61:306–316. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.bbi.2016.12.014) [bbi.2016.12.014](https://doi.org/10.1016/j.bbi.2016.12.014)
- <span id="page-14-3"></span>50. Xu X, Yin D, Ren H, Gao W, Li F, Sun D, Wu Y, Zhou S et al (2018) Selective NLRP3 infammasome inhibitor reduces neuroinfammation and improves long-term neurological outcomes in a murine model of traumatic brain injury. Neurobiol Dis 117:15– 27.<https://doi.org/10.1016/j.nbd.2018.05.016>
- <span id="page-14-4"></span>51. Ismael S, Nasoohi S, Ishrat T (2018) MCC950, the selective inhibitor of nucleotide oligomerization domain-like receptor protein-3 infammasome, protects mice against traumatic brain injury. J Neurotrauma 35(11):1294–1303. [https://doi.org/10.](https://doi.org/10.1089/neu.2017.5344) [1089/neu.2017.5344](https://doi.org/10.1089/neu.2017.5344)
- <span id="page-14-5"></span>52. Heijden TVD, Kritikou E, Venema W, Duijn JV, Santbrink PJV, Slutter B, Foks AC, Bot I et al (2017) NLRP3 infammasome inhibition by MCC950 reduces atherosclerotic lesion development in apolipoprotein E-defcient mice-brief report. Arterioscler Thromb Vasc Biol 37(8):1457–1461. [https://doi.org/10.1161/](https://doi.org/10.1161/ATVBAHA.117.309575) [ATVBAHA.117.309575](https://doi.org/10.1161/ATVBAHA.117.309575)
- <span id="page-14-6"></span>53. Mridha AR, Wree A, Robertson AAB, Yeh MM, Johnson CD, Rooyen DMV, Haczeyni F, Teoh NC et al (2017) NLRP3 infammasome blockade reduces liver infammation and fbrosis in experimental NASH in mice. J Hepatol 66(5):1037–1046. [https://](https://doi.org/10.1016/j.jhep.2017.01.022) [doi.org/10.1016/j.jhep.2017.01.022](https://doi.org/10.1016/j.jhep.2017.01.022)
- <span id="page-14-7"></span>54. Juliana C, Fernandes-alnemri T, Wu J, Datta P, Solorzano L, Yu J, Meng R, Quong AA et al (2010) Anti-infammatory compounds parthenolide and Bay 11–7082 are direct inhibitors of the infammasome. J Biol Chem 285(13):9792–9802. [https://doi.org/](https://doi.org/10.1074/jbc.M109.082305) [10.1074/jbc.M109.082305](https://doi.org/10.1074/jbc.M109.082305)
- <span id="page-14-8"></span>55. Guo C, Fulp JW, Jiang Y, Li X, Chojnacki JE, Wu J, Wang X, Zhang S (2017) Development and characterization of a hydroxylsulfonamide analogue, 5-chloro-N-[2-(4-hydroxysulfamoylphenyl)-ethyl]-2-methoxy-benzamide, as a novel NLRP3 infammasome inhibitor for potential treatment of multiple sclerosis. ACS Chem Neurosci 8(10):2194–2201. [https://doi.org/10.1021/](https://doi.org/10.1021/acschemneuro.7b00124) [acschemneuro.7b00124](https://doi.org/10.1021/acschemneuro.7b00124)
- <span id="page-14-9"></span>56. Youm Y, Nguyen KY, Grant RW, Goldberg EL, Bodogai M, Kim D, D'agostino D, Planavsky N et al (2015) The ketone metabolite beta-hydroxybutyrate blocks NLRP3 infammasome-mediated infammatory disease. Nat Med 21(3):263–269. [https://doi.org/](https://doi.org/10.1038/nm.3804) [10.1038/nm.3804](https://doi.org/10.1038/nm.3804)
- <span id="page-14-10"></span>57. Yang F, Wang Z, Wei X, Han H, Meng X, Zhang Y, Shi W, Li F et al (2014) NLRP3 defciency ameliorates neurovascular damage in experimental ischemic stroke. J Cerebral Blood Flow Metab 34(4):660–667. <https://doi.org/10.1038/jcbfm.2013.242>
- <span id="page-14-22"></span>58. Ren H, Kong Y, Liu Z, Zang D, Yang X, Wood K, Li M, Liu Q (2018) Selective NLRP3 (pyrin domain-containing protein 3) infammasome inhibitor reduces brain injury after intracerebral hemorrhage. Stroke 49(1):184–192. [https://doi.org/10.](https://doi.org/10.1161/STROKEAHA.117.018904) [1161/STROKEAHA.117.018904](https://doi.org/10.1161/STROKEAHA.117.018904)
- <span id="page-14-23"></span>59. Heneka MT, Kummer MP, Stutz A, Delekate A, Schwartz S, Vieira-saecker A, Griep A, Axt D et al (2013) NLRP3 is

activated in Alzheimer's disease and contributes to pathology in APP/PS1 mice. Nature 493(7434):674–678. [https://doi.org/](https://doi.org/10.1038/nature11729) [10.1038/nature11729](https://doi.org/10.1038/nature11729)

- <span id="page-14-25"></span>60. Shen K, Mao Q, Yin X, Zhang C, Jin Y, Deng A, Gu Z, Chen B (2018) NLRP3 infammasome activation leads to epileptic neuronal apoptosis. Curr Neurovasc Res 15(4):276–281. [https://](https://doi.org/10.2174/1567202616666181122165540) [doi.org/10.2174/1567202616666181122165540](https://doi.org/10.2174/1567202616666181122165540)
- <span id="page-14-11"></span>61. Fu Q, Wu J, Zhou X, Ji M, Mao Q, Li Q, Zong M, Zhou Z et al (2019) NLRP3/caspase-1 pathway-induced pyroptosis mediated cognitive deficits in a mouse model of sepsis-associated encephalopathy. Infammation 42(1):306–318. [https://doi.org/](https://doi.org/10.1007/s10753-018-0894-4) [10.1007/s10753-018-0894-4](https://doi.org/10.1007/s10753-018-0894-4)
- <span id="page-14-12"></span>62. Feng L, Chen Y, Ding R, Fu Z, Yang S, Deng X, Zeng J (2015) P2X7R blockade prevents NLRP3 infammasome activation and brain injury in a rat model of intracerebral hemorrhage: involvement of peroxynitrite. J Neuroinfammation 12:190. <https://doi.org/10.1186/s12974-015-0409-2>
- <span id="page-14-13"></span>63. Liu P, Gao T, Li T, Yang Y, Xu Y, Xu Z, Mi W (2021) Repeated propofol exposure-induced neuronal damage and cognitive impairment in aged rats by activation of NF-kappaB pathway and NLRP3 infammasome. Neurosci Lett 740:135461. [https://](https://doi.org/10.1016/j.neulet.2020.135461) [doi.org/10.1016/j.neulet.2020.135461](https://doi.org/10.1016/j.neulet.2020.135461)
- <span id="page-14-24"></span>64. Ruan Y, Qiu X, Lv Y, Dong D, Wu X, Zhu J, Zheng X (2019) Kainic acid induces production and aggregation of amyloid beta-protein and memory deficits by activating inflammasomes in NLRP3- and NF-kappaB-stimulated pathways. Aging 11(11):3795–3810. <https://doi.org/10.18632/aging.102017>
- <span id="page-14-14"></span>65. Irrera N, Pizzino G, Calo M, Pallio G, Mannino F, Fama F, Arcoraci V, Fodale V et al (2017) Lack of the Nlrp3 infammasome improves mice recovery following traumatic brain injury. Front Pharmacol 8:459. [https://doi.org/10.3389/fphar.](https://doi.org/10.3389/fphar.2017.00459) [2017.00459](https://doi.org/10.3389/fphar.2017.00459)
- <span id="page-14-15"></span>66. Inoue M, Williams KL, Oliver T, Vandenabeele P, Rajan JV, Miao EA, Shinohara ML (2012) Interferon-beta therapy against EAE is efective only when development of the disease depends on the NLRP3 infammasome. Sci Signal 5(225):ra38. [https://](https://doi.org/10.1126/scisignal.2002767) [doi.org/10.1126/scisignal.2002767](https://doi.org/10.1126/scisignal.2002767)
- <span id="page-14-16"></span>67. Flores J, Noel A, Foveau B, Lynham J, Lecrux C, Leblanc AC (2018) Caspase-1 inhibition alleviates cognitive impairment and neuropathology in an Alzheimer's disease mouse model. Nat Commun 9(1):3916. [https://doi.org/10.1038/](https://doi.org/10.1038/s41467-018-06449-x) [s41467-018-06449-x](https://doi.org/10.1038/s41467-018-06449-x)
- <span id="page-14-17"></span>68. Mao Z, Liu C, Ji S, Yang Q, Ye H, Han H, Xue Z (2017) The NLRP3 infammasome is involved in the pathogenesis of Parkinson's disease in rats. Neurochem Res 42(4):1104-1115. [https://](https://doi.org/10.1007/s11064-017-2185-0) [doi.org/10.1007/s11064-017-2185-0](https://doi.org/10.1007/s11064-017-2185-0)
- <span id="page-14-18"></span>69. Qiao C, Zhang L, Sun X, Ding J, Lu M, Hu G (2017) Caspase-1 deficiency alleviates dopaminergic neuronal death via inhibiting caspase-7/AIF pathway in MPTP/p mouse model of Parkinson's disease. Mol Neurobiol 54(6):4292–4302. [https://doi.org/10.](https://doi.org/10.1007/s12035-016-9980-5) [1007/s12035-016-9980-5](https://doi.org/10.1007/s12035-016-9980-5)
- <span id="page-14-19"></span>70. Li C, Lin H, He H, Ma M, Jiang W, Zhou R (2022) Inhibition of the NLRP3 infammasome activation by manoalide ameliorates experimental autoimmune encephalomyelitis pathogenesis. Front Cell Dev Biol 10:822236. [https://doi.org/10.3389/fcell.2022.](https://doi.org/10.3389/fcell.2022.822236) [822236](https://doi.org/10.3389/fcell.2022.822236)
- <span id="page-14-20"></span>71. Lee E, Hwang I, Park S, Hong S, Hwang B, Cho Y, Son J, Yu J (2019) MPTP-driven NLRP3 inflammasome activation in microglia plays a central role in dopaminergic neurodegeneration. Cell Death Difer 26(2):213–228. [https://doi.org/10.1038/](https://doi.org/10.1038/s41418-018-0124-5) [s41418-018-0124-5](https://doi.org/10.1038/s41418-018-0124-5)
- <span id="page-14-21"></span>72. He Q, Li Z, Wang Y, Hou Y, Li L, Zhao J (2017) Resveratrol alleviates cerebral ischemia/reperfusion injury in rats by inhibiting NLRP3 infammasome activation through Sirt1-dependent autophagy induction. Int Immunopharmacol 50:208–215. [https://](https://doi.org/10.1016/j.intimp.2017.06.029) [doi.org/10.1016/j.intimp.2017.06.029](https://doi.org/10.1016/j.intimp.2017.06.029)
- <span id="page-15-27"></span>73. Qi Y, Shang L, Liao Z, Su H, Jing H, Wu B, Bi K, Jia Y (2019) Intracerebroventricular injection of resveratrol ameliorated Abeta-induced learning and cognitive decline in mice. Metab Brain Dis 34(1):257–266. [https://doi.org/10.1007/](https://doi.org/10.1007/s11011-018-0348-6) [s11011-018-0348-6](https://doi.org/10.1007/s11011-018-0348-6)
- <span id="page-15-10"></span>74. Zhang X, Wu Q, Zhang Q, Lu Y, Liu J, Li W, Lv S, Zhou M et al (2017) Resveratrol attenuates early brain injury after experimental subarachnoid hemorrhage via inhibition of NLRP3 infammasome activation. Front Neurosci 11:611. [https://doi.org/10.3389/](https://doi.org/10.3389/fnins.2017.00611) [fnins.2017.00611](https://doi.org/10.3389/fnins.2017.00611)
- <span id="page-15-11"></span>75. Xiao L, Dai Z, Tang W, Liu C, Tang B (2021) Astragaloside IV alleviates cerebral ischemia-reperfusion injury through NLRP3 infammasome-mediated pyroptosis inhibition via activating Nrf2. Oxid Med Cell Longev 2021:9925561. [https://doi.org/10.](https://doi.org/10.1155/2021/9925561) [1155/2021/9925561](https://doi.org/10.1155/2021/9925561)
- <span id="page-15-12"></span>76. Yuan R, Fan H, Cheng S, Gao W, Xu X, Lv S, Ye M, Wu M et al (2017) Silymarin prevents NLRP3 infammasome activation and protects against intracerebral hemorrhage. Biomed Pharmacother 93:308–315. <https://doi.org/10.1016/j.biopha.2017.06.018>
- <span id="page-15-13"></span>77. Liu H, Zhao L, Yue L, Wang B, Li X, Guo H, Ma Y, Yao C et al (2017) Pterostilbene attenuates early brain injury following subarachnoid hemorrhage via inhibition of the NLRP3 infammasome and Nox2-related oxidative stress. Mol Neurobiol 54(8):5928– 5940. <https://doi.org/10.1007/s12035-016-0108-8>
- <span id="page-15-14"></span>78. Wang C, Xu Y, Wang X, Guo C, Wang T, Wang Z (2019) Dl-3-n-Butylphthalide inhibits NLRP3 infammasome and mitigates Alzheimer's-like pathology via Nrf2-TXNIP-TrX axis. Antioxid Redox Signal 30(11):1411–1431. [https://doi.org/10.1089/ars.](https://doi.org/10.1089/ars.2017.7440) [2017.7440](https://doi.org/10.1089/ars.2017.7440)
- <span id="page-15-15"></span>79. Que R, Zheng J, Chang Z, Zhang W, Li H, Xie Z, Huang Z, Wang H et al (2021) Dl-3-n-Butylphthalide rescues dopaminergic neurons in Parkinson's disease models by inhibiting the NLRP3 infammasome and ameliorating mitochondrial impairment. Front Immunol 12:794770. [https://doi.org/10.3389/fmmu.](https://doi.org/10.3389/fimmu.2021.794770) [2021.794770](https://doi.org/10.3389/fimmu.2021.794770)
- <span id="page-15-16"></span>80. Ge X, Wang Y, Yu S, Cao X, Chen Y, Cheng Q, Ding F (2021) Anti-infammatory activity of a polypeptide fraction from Achyranthes bidentate in amyloid beta oligomers induced model of Alzheimer's disease. Front Pharmacol 12:716177. [https://doi.org/](https://doi.org/10.3389/fphar.2021.716177) [10.3389/fphar.2021.716177](https://doi.org/10.3389/fphar.2021.716177)
- <span id="page-15-17"></span>81. Qiu W, Pan R, Tang Y, Zhou X, Wu J, Yu L, Law BY, Ai W et al (2020) Lychee seed polyphenol inhibits Abeta-induced activation of NLRP3 infammasome via the LRP1/AMPK mediated autophagy induction. Biomed Pharmacother 130:110575. [https://](https://doi.org/10.1016/j.biopha.2020.110575) [doi.org/10.1016/j.biopha.2020.110575](https://doi.org/10.1016/j.biopha.2020.110575)
- <span id="page-15-18"></span>82. Ahmed S, Panda SR, Kwatra M, Sahu BD, Naidu V (2022) Perillyl alcohol attenuates NLRP3 infammasome activation and rescues dopaminergic neurons in experimental *in vitro* and *in vivo* models of Parkinson's disease. ACS Chem Neurosci 13(1):53– 68.<https://doi.org/10.1021/acschemneuro.1c00550>
- <span id="page-15-19"></span>83. Liu Y, Li Y, Xiao L, Chen L, Zheng S, Zeng E, Xu C (2021) Extracellular vesicles derived from M2 microglia reduce ischemic brain injury through microRNA-135a-5p/TXNIP/ NLRP3 axis. Lab Investig 101(7):837–850. [https://doi.org/10.](https://doi.org/10.1038/s41374-021-00545-1) [1038/s41374-021-00545-1](https://doi.org/10.1038/s41374-021-00545-1)
- <span id="page-15-20"></span>84. Yang Z, Zhong L, Xian R, Yuan B (2015) MicroRNA-223 regulates infammation and brain injury via feedback to NLRP3 infammasome after intracerebral hemorrhage. Mol Immunol 65(2):267–276. <https://doi.org/10.1016/j.molimm.2014.12.018>
- <span id="page-15-21"></span>85. Hu L, Zhang H, Wang B, Ao Q, He Z (2020) MicroRNA-152 attenuates neuroinfammation in intracerebral hemorrhage by inhibiting thioredoxin interacting protein (TXNIP)-mediated NLRP3 inflammasome activation. Int Immunopharmacol 80:106141.<https://doi.org/10.1016/j.intimp.2019.106141>
- <span id="page-15-22"></span>86. Han C, Guo L, Yang Y, Guan Q, Shen H, Sheng Y, Jiao Q (2020) Mechanism of microRNA-22 in regulating neuroinfammation in

Alzheimer's disease. Brain Behav 10(6):01627. [https://doi.org/](https://doi.org/10.1002/brb3.1627) [10.1002/brb3.1627](https://doi.org/10.1002/brb3.1627)

- <span id="page-15-23"></span>87. Daniels MJD, Rivers-auty J, Schilling T, Spencer NG, Watremez W, Fasolino V, Booth SJ, White CS et al (2016) Fenamate NSAIDs inhibit the NLRP3 infammasome and protect against Alzheimer's disease in rodent models. Nat Commun 7:12504. <https://doi.org/10.1038/ncomms12504>
- <span id="page-15-9"></span>88. Chen B, Zhang M, Ji M, Zhang D, Chen B, Gong W, Li X, Zhou Y et al (2022) The neuroprotective mechanism of lithium after ischaemic stroke. Commun Biol 5(1):105. [https://doi.org/10.](https://doi.org/10.1038/s42003-022-03051-2) [1038/s42003-022-03051-2](https://doi.org/10.1038/s42003-022-03051-2)
- <span id="page-15-24"></span>89. Peng J, Wang H, Gong Z, Li X, He L, Shen Q, Pan J, Peng Y (2020) Idebenone attenuates cerebral infammatory injury in ischemia and reperfusion via dampening NLRP3 infammasome activity. Mol Immunol 123:74–87. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.molimm.2020.04.013) [molimm.2020.04.013](https://doi.org/10.1016/j.molimm.2020.04.013)
- <span id="page-15-25"></span>90. Ismael S, Nasoohi S, Yoo A, Mirzahosseini G, Ahmed HA, Ishrat T (2021) Verapamil as an adjunct therapy to reduce tPA toxicity in hyperglycemic stroke: implication of TXNIP/NLRP3 infammasome. Mol Neurobiol 58(8):3792–3804. [https://doi.org/10.](https://doi.org/10.1007/s12035-021-02384-z) [1007/s12035-021-02384-z](https://doi.org/10.1007/s12035-021-02384-z)
- <span id="page-15-26"></span>91. Cao S, Shrestha S, Li J, Yu X, Chen J, Yan F, Ying G, Gu C et al (2017) Melatonin-mediated mitophagy protects against early brain injury after subarachnoid hemorrhage through inhibition of NLRP3 infammasome activation. Sci Rep 7(1):2417. [https://](https://doi.org/10.1038/s41598-017-02679-z) [doi.org/10.1038/s41598-017-02679-z](https://doi.org/10.1038/s41598-017-02679-z)
- <span id="page-15-0"></span>92. Tsao CW, Aday AW, Almarzooq ZI, Alonso A, Beaton AZ, Bittencourt MS, Boehme AK, Buxton AE et al (2022) Heart Disease and Stroke Statistics-2022 update: a report from the American Heart Association. Circulation 145(8):153-e639. [https://doi.org/](https://doi.org/10.1161/CIR.0000000000001052) [10.1161/CIR.0000000000001052](https://doi.org/10.1161/CIR.0000000000001052)
- <span id="page-15-1"></span>93. Campbell BCV, Silva DAD, Macleod MR, Coutts SB, Schwamm LH, Davis SM, Donnan GA (2019) Ischaemic stroke. Nat Rev Dis Primers 5(1):70.<https://doi.org/10.1038/s41572-019-0118-8>
- <span id="page-15-2"></span>94. Pendlebury ST, Rothwell PM (2009) Prevalence, incidence, and factors associated with pre-stroke and post-stroke dementia: a systematic review and meta-analysis. Lancet Neurol 8(11):1006– 1018. [https://doi.org/10.1016/S1474-4422\(09\)70236-4](https://doi.org/10.1016/S1474-4422(09)70236-4)
- <span id="page-15-3"></span>95. O'collins VE, Macleod MR, Donnan GA, Horky LL, Worp BHVD, Howells DW (2006) 1,026 experimental treatments in acute stroke. Ann Neurol 59(3):467–477. [https://doi.org/10.1002/](https://doi.org/10.1002/ana.20741) [ana.20741](https://doi.org/10.1002/ana.20741)
- <span id="page-15-4"></span>96. Guekht A, Skoog I, Edmundson S, Zakharov V, Korczyn AD  $(2017)$  ARTEMIDA Trial (a randomized trial of efficacy, 12 months international double-blind actovegin): a randomized controlled trial to assess the efficacy of actovegin in poststroke cognitive impairment. Stroke 48(5):1262–1270. [https://doi.org/](https://doi.org/10.1161/STROKEAHA.116.014321) [10.1161/STROKEAHA.116.014321](https://doi.org/10.1161/STROKEAHA.116.014321)
- <span id="page-15-5"></span>97. Fann DY, Lee S, Manzanero S, Chunduri P, Sobey CG, Arumugam TV (2013) Pathogenesis of acute stroke and the role of infammasomes. Ageing Res Rev 12(4):66–941. [https://doi.org/](https://doi.org/10.1016/j.arr.2013.09.004) [10.1016/j.arr.2013.09.004](https://doi.org/10.1016/j.arr.2013.09.004)
- <span id="page-15-6"></span>98. Savage CD, Lopez-castejon G, Denes A, Brough D (2012) NLRP3-infammasome activating DAMPs stimulate an infammatory response in glia in the absence of priming which contributes to brain infammation after injury. Front Immunol 3:288
- <span id="page-15-7"></span>99. Minutoli L, Puzzolo D, Rinaldi M, Irrera N, Marini H, Arcoraci V, Bitto A, Crea G et al (2016) ROS-mediated NLRP3 infammasome activation in brain, heart, kidney, and testis ischemia/ reperfusion injury. Oxid Med Cell Longev 2016:2183026. [https://](https://doi.org/10.1155/2016/2183026) [doi.org/10.1155/2016/2183026](https://doi.org/10.1155/2016/2183026)
- <span id="page-15-8"></span>100. Gong Z, Pan J, Shen Q, Li M, Peng Y (2018) Mitochondrial dysfunction induces NLRP3 inflammasome activation during cerebral ischemia/reperfusion injury. J Neuroinfammation 15(1):242. <https://doi.org/10.1186/s12974-018-1282-6>
- <span id="page-16-0"></span>101. Barrington J, Lemarchand E, Allan SM (2017) A brain in fame; do infammasomes and pyroptosis infuence stroke pathology? Brain Pathol 27(2):205–212.<https://doi.org/10.1111/bpa.12476>
- 102. Denes A, Pinteaux E, Rothwell NJ, Allan SM (2011) Interleukin-1 and stroke: biomarker, harbinger of damage, and therapeutic target. Cerebrovasc Dis 32(6):27–517. [https://doi.org/10.](https://doi.org/10.1159/000332205) [1159/000332205](https://doi.org/10.1159/000332205)
- <span id="page-16-7"></span>103. Fann DY, Lee SY, Manzanero S, Tang SC, Gelderblom M, Chunduri P, Bernreuther C, Glatzel M et al (2013) Intravenous immunoglobulin suppresses NLRP1 and NLRP3 infammasome-mediated neuronal death in ischemic stroke. Cell Death Dis 4:790. <https://doi.org/10.1038/cddis.2013.326>
- <span id="page-16-1"></span>104. Fann DY, Santro T, Manzanero S, Widiapradja A, Cheng Y, Lee S, Chunduri P, Jo D et al (2014) Intermittent fasting attenuates infammasome activity in ischemic stroke. Exp Neurol 257:114– 119. <https://doi.org/10.1016/j.expneurol.2014.04.017>
- <span id="page-16-2"></span>105. Wang Q, Tang XN, Yenari MA (2007) The inflammatory response in stroke. J Neuroimmunol 184(1–2):53–68
- <span id="page-16-3"></span>106. Gustin A, Kirchmeyer M, Koncina E, Felten P, Losciuto S, Heurtaux T, Tardivel A, Heuschling P et al (2015) NLRP3 infammasome is expressed and functional in mouse brain microglia but not in astrocytes. PLoS ONE 10(6):0130624. [https://doi.org/10.](https://doi.org/10.1371/journal.pone.0130624) [1371/journal.pone.0130624](https://doi.org/10.1371/journal.pone.0130624)
- <span id="page-16-4"></span>107. Prinz M, Erny D, Hagemeyer N (2017) Ontogeny and homeostasis of CNS myeloid cells. Nat Immunol 18(4):385–392. [https://](https://doi.org/10.1038/ni.3703) [doi.org/10.1038/ni.3703](https://doi.org/10.1038/ni.3703)
- <span id="page-16-5"></span>108. Salter MW, Stevens B (2017) Microglia emerge as central players in brain disease. Nat Med 23(9):1018–1027. [https://doi.org/10.](https://doi.org/10.1038/nm.4397) [1038/nm.4397](https://doi.org/10.1038/nm.4397)
- <span id="page-16-6"></span>109. Vilalta A, Brown GC (2018) Neurophagy, the phagocytosis of live neurons and synapses by glia, contributes to brain development and disease. FEBS J 285(19):3566–3575. [https://doi.org/](https://doi.org/10.1111/febs.14323) [10.1111/febs.14323](https://doi.org/10.1111/febs.14323)
- <span id="page-16-8"></span>110. Wang C, Zhang L, Li G, Shi Y, Li J, Zhang X, Wang Z, Ding F et al (2014) Mulberroside A protects against ischemic impairment in primary culture of rat cortical neurons after oxygenglucose deprivation followed by reperfusion. J Neurosci Res 92(7):54–944. <https://doi.org/10.1002/jnr.23374>
- <span id="page-16-9"></span>111. Ye X, Shen T, Hu J, Zhang L, Zhang Y, Bao L, Cui C, Jin G et al (2017) Purinergic 2X7 receptor/NLRP3 pathway triggers neuronal apoptosis after ischemic stroke in the mouse. Exp Neurol 292:46–55.<https://doi.org/10.1016/j.expneurol.2017.03.002>
- <span id="page-16-10"></span>112. Ismael S, Zhao L, Nasoohi S, Ishrat T (2018) Inhibition of the NLRP3-infammasome as a potential approach for neuroprotection after stroke. Sci Rep 8(1):5971. [https://doi.org/10.1038/](https://doi.org/10.1038/s41598-018-24350-x) [s41598-018-24350-x](https://doi.org/10.1038/s41598-018-24350-x)
- <span id="page-16-11"></span>113. Ward R, Li W, Abdul Y, Jackson L, Dong G, Jamil S, Filosa J, Fagan SC et al (2019) NLRP3 infammasome inhibition with MCC950 improves diabetes-mediated cognitive impairment and vasoneuronal remodeling after ischemia. Pharmacol Res 142:237–250.<https://doi.org/10.1016/j.phrs.2019.01.035>
- <span id="page-16-12"></span>114. Bellut M, Papp L, Bieber M, Kraft P, Stoll G, Schuhmann MK (2021) NLPR3 inflammasome inhibition alleviates hypoxic endothelial cell death *in vitro* and protects blood-brain barrier integrity in murine stroke. Cell Death Dis 13(1):20. [https://doi.](https://doi.org/10.1038/s41419-021-04379-z) [org/10.1038/s41419-021-04379-z](https://doi.org/10.1038/s41419-021-04379-z)
- <span id="page-16-13"></span>115. Fann DY, Lim Y, Cheng Y, Lok K, Chunduri P, Baik S, Drummond GR, Dheen ST et al (2018) Evidence that NF-kappaB and MAPK signaling promotes NLRP infammasome activation in neurons following ischemic stroke. Mol Neurobiol 55(2):1082– 1096.<https://doi.org/10.1007/s12035-017-0394-9>
- <span id="page-16-14"></span>116. Zhu H, Jian Z, Zhong Y, Ye Y, Zhang Y, Hu X, Pu B, Gu L et al (2021) Janus kinase inhibition ameliorates ischemic stroke injury and neuroinfammation through reducing NLRP3 infammasome

activation via JAK2/STAT3 pathway inhibition. Front Immunol 12:714943. [https://doi.org/10.3389/fmmu.2021.714943](https://doi.org/10.3389/fimmu.2021.714943)

- <span id="page-16-15"></span>117. Mohammadianinejad SE, Majdinasab N, Sajedi SA, Abdollahi F, Moqaddam MM, Sadr F (2014) The effect of lithium in poststroke motor recovery: a double-blind, placebo-controlled, randomized clinical trial. Clin Neuropharmacol 37(3):73–78. [https://](https://doi.org/10.1097/WNF.0000000000000028) [doi.org/10.1097/WNF.0000000000000028](https://doi.org/10.1097/WNF.0000000000000028)
- <span id="page-16-16"></span>118. Giang KW, Mandalenakis Z, Dellborg M, Lappas G, Eriksson P, Hansson P, Rosengren A (2018) Long-term risk of hemorrhagic stroke in young patients with congenital heart disease. Stroke 49(5):1155–1162. [https://doi.org/10.1161/STROKEAHA.117.](https://doi.org/10.1161/STROKEAHA.117.020032) [020032](https://doi.org/10.1161/STROKEAHA.117.020032)
- <span id="page-16-17"></span>119. Rosand J (2021) Preserving brain health after intracerebral haemorrhage. Lancet Neurol 20(11):879–880. [https://doi.org/10.1016/](https://doi.org/10.1016/S1474-4422(21)00339-2) [S1474-4422\(21\)00339-2](https://doi.org/10.1016/S1474-4422(21)00339-2)
- <span id="page-16-18"></span>120. Salman RA, Law ZK, Bath PM, Steiner T, Sprigg N (2018) Haemostatic therapies for acute spontaneous intracerebral haemorrhage. Cochrane Database Syst Rev 4:005951. [https://doi.org/10.](https://doi.org/10.1002/14651858.CD005951.pub4) [1002/14651858.CD005951.pub4](https://doi.org/10.1002/14651858.CD005951.pub4)
- <span id="page-16-19"></span>121. Keep RF, Hua Y, Xi G (2012) Intracerebral haemorrhage: mechanisms of injury and therapeutic targets. Lancet Neurol 11(8):31– 720. [https://doi.org/10.1016/S1474-4422\(12\)70104-7](https://doi.org/10.1016/S1474-4422(12)70104-7)
- <span id="page-16-20"></span>122. Mendelow AD, Gregson BA, Fernandes HM, Murray GD, Teasdale GM, Hope DT, Karimi A, Shaw MDM et al (2005) Early surgery versus initial conservative treatment in patients with spontaneous supratentorial intracerebral haematomas in the International Surgical Trial in Intracerebral Haemorrhage (STICH): a randomised trial. Lancet 365(9457):387–397. [https://](https://doi.org/10.1016/S0140-6736(05)17826-X) [doi.org/10.1016/S0140-6736\(05\)17826-X](https://doi.org/10.1016/S0140-6736(05)17826-X)
- <span id="page-16-21"></span>123 Mendelow AD, Gregson BA, Rowan EN, Murray GD, Gholkar A, Mitchell PM (2013) Early surgery versus initial conservative treatment in patients with spontaneous supratentorial lobar intracerebral haematomas (STICH II): a randomised trial. Lancet 382(9890):397–408. [https://doi.org/10.1016/S0140-6736\(13\)](https://doi.org/10.1016/S0140-6736(13)60986-1) [60986-1](https://doi.org/10.1016/S0140-6736(13)60986-1)
- <span id="page-16-22"></span>124. Xi G, Keep RF, Hoff JT (2006) Mechanisms of brain injury after intracerebral haemorrhage. Lancet Neurol 5(1):53–63
- <span id="page-16-23"></span>125. Shao A, Zhu Z, Li L, Zhang S, Zhang J (2019) Emerging therapeutic targets associated with the immune system in patients with intracerebral haemorrhage (ICH): from mechanisms to translation. EBioMedicine 45:615–623. [https://doi.org/10.1016/j.ebiom.](https://doi.org/10.1016/j.ebiom.2019.06.012) [2019.06.012](https://doi.org/10.1016/j.ebiom.2019.06.012)
- <span id="page-16-24"></span>126. Ma Q, Chen S, Hu Q, Feng H, Zhang JH, Tang J (2014) NLRP3 infammasome contributes to infammation after intracerebral hemorrhage. Ann Neurol 75(2):19–209. [https://doi.org/10.1002/](https://doi.org/10.1002/ana.24070) [ana.24070](https://doi.org/10.1002/ana.24070)
- <span id="page-16-25"></span>127. Yao S, Cao F, Chen J, Chen W, Fan R, Li G, Zeng Y, Jiao S et al (2017) NLRP3 is required for complement-mediated caspase-1 and IL-1beta activation in ICH. J Mol Neurosci 61(3):385–395. <https://doi.org/10.1007/s12031-016-0874-9>
- <span id="page-16-26"></span>128. Luo Y, Reis C, Chen S (2019) NLRP3 infammasome in the pathophysiology of hemorrhagic stroke: a review. Curr Neuropharmacol 17(7):582–589. [https://doi.org/10.2174/1570159X17](https://doi.org/10.2174/1570159X17666181227170053) [666181227170053](https://doi.org/10.2174/1570159X17666181227170053)
- <span id="page-16-27"></span>129. Cheng Y, Chen B, Xie W, Chen Z, Yang G, Cai Y, Shang H, Zhao W (2020) Ghrelin attenuates secondary brain injury following intracerebral hemorrhage by inhibiting NLRP3 infammasome activation and promoting Nrf2/ARE signaling pathway in mice. Int Immunopharmacol 79:106180. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.intimp.2019.106180) [intimp.2019.106180](https://doi.org/10.1016/j.intimp.2019.106180)
- <span id="page-16-28"></span>130. Zheng S, Jian D, Gan H, Wang L, Zhao J, Zhai X (2021) FUNDC1 inhibits NLRP3-mediated infammation after intracerebral hemorrhage by promoting mitophagy in mice. Neurosci Lett 756:135967.<https://doi.org/10.1016/j.neulet.2021.135967>
- <span id="page-17-0"></span>131. Ji N, Wu L, Shi H, Li Q, Yu A, Yang Z (2022) VSIG4 Attenuates NLRP3 and ameliorates neuroinfammation via JAK2-STAT3- A20 pathway after intracerebral hemorrhage in mice. Neurotox Res 40(1):78–88.<https://doi.org/10.1007/s12640-021-00456-5>
- <span id="page-17-1"></span>132. Yuan B, Shen H, Lin L, Su T, Zhong S, Yang Z (2015) Recombinant adenovirus encoding NLRP3 RNAi attenuate infammation and brain injury after intracerebral hemorrhage. J Neuroimmunol 287:5–71.<https://doi.org/10.1016/j.jneuroim.2015.08.002>
- <span id="page-17-2"></span>133. Chang Y, Ka S, Hsu W, Chen A, Chao LK, Lin C, Hsieh C, Chen M et al (2015) Resveratrol inhibits NLRP3 infammasome activation by preserving mitochondrial integrity and augmenting autophagy. J Cell Physiol 230(7):1567–1579. [https://doi.org/10.](https://doi.org/10.1002/jcp.24903) [1002/jcp.24903](https://doi.org/10.1002/jcp.24903)
- <span id="page-17-3"></span>134. Cai J, Liu W, Lu F, Kong W, Zhou X, Miao P, Lei C, Wang Y (2018) Resveratrol attenuates neurological deficit and neuroinfammation following intracerebral hemorrhage. Exp Ther Med 15(5):4131–4138.<https://doi.org/10.3892/etm.2018.5938>
- <span id="page-17-4"></span>135 Kamp MA, Steiger H, Lieshout JHV (2020) Experimental aneurysmal subarachnoid hemorrhage: tiding over. Transl Stroke Res 11(1):1–3. <https://doi.org/10.1007/s12975-019-00726-7>
- <span id="page-17-5"></span>136. Murakami K, Koide M, Dumont TM, Russell SR, Tranmer BI, Wellman GC (2011) Subarachnoid hemorrhage induces gliosis and increased expression of the pro-infammatory cytokine high mobility group box 1 protein. Transl Stroke Res 2(1):72–79. <https://doi.org/10.1007/s12975-010-0052-2>
- <span id="page-17-6"></span>137 Suzuki H (2019) Infammation: a good research target to improve outcomes of poor-grade subarachnoid hemorrhage. Transl Stroke Res 10(6):597–600.<https://doi.org/10.1007/s12975-019-00713-y>
- <span id="page-17-7"></span>138. Frosen J, Cebral J, Robertson AM, Aoki T (2019) Flow-induced, infammation-mediated arterial wall remodeling in the formation and progression of intracranial aneurysms. Neurosurg Focus 47(1):21.<https://doi.org/10.3171/2019.5.FOCUS19234>
- <span id="page-17-8"></span>139. Chen S, Ma Q, Kraft PR, Hu Q, Rolland W, Sherchan P, Zhang J, Tang J et al (2013) P2X7R/cryopyrin infammasome axis inhibition reduces neuroinfammation after SAH. Neurobiol Dis 58:296–307. <https://doi.org/10.1016/j.nbd.2013.06.011>
- <span id="page-17-9"></span>140. Dodd WS, Noda I, Martinez M, Hosaka K, Hoh BL (2021) NLRP3 inhibition attenuates early brain injury and delayed cerebral vasospasm after subarachnoid hemorrhage. J Neuroinfammation 18(1):163. <https://doi.org/10.1186/s12974-021-02207-x>
- <span id="page-17-10"></span>141. Luo Y, Lu J, Ruan W, Guo X, Chen S (2019) MCC950 attenuated early brain injury by suppressing NLRP3 infammasome after experimental SAH in rats. Brain Res Bull 146:320–326. [https://](https://doi.org/10.1016/j.brainresbull.2019.01.027) [doi.org/10.1016/j.brainresbull.2019.01.027](https://doi.org/10.1016/j.brainresbull.2019.01.027)
- <span id="page-17-11"></span>142. Chen S, Ding Y, Shi S, Tu X (2022) Schisandrin B inhibits NLRP3 infammasome pathway and attenuates early brain injury in rats of subarachnoid hemorrhage. Chin J Integr Med. [https://](https://doi.org/10.1007/s11655-021-3348-z) [doi.org/10.1007/s11655-021-3348-z](https://doi.org/10.1007/s11655-021-3348-z)
- <span id="page-17-12"></span>143. Zuo Y, Wang J, Liao F, Yan X, Li J, Huang L, Liu F (2018) Inhibition of heat shock protein 90 by 17-AAG reduces infammation via P2X7 receptor/NLRP3 infammasome pathway and increases neurogenesis after subarachnoid hemorrhage in mice. Front Mol Neurosci 11:401. <https://doi.org/10.3389/fnmol.2018.00401>
- <span id="page-17-13"></span>144. Hu X, Yan J, Huang L, Araujo C, Peng J, Gao L, Liu S, Tang J et al (2021) INT-777 attenuates NLRP3-ASC infammasomemediated neuroinfammation via TGR5/cAMP/PKA signaling pathway after subarachnoid hemorrhage in rats. Brain Behav Immun 91:587–600. <https://doi.org/10.1016/j.bbi.2020.09.016>
- <span id="page-17-14"></span>145. Xu P, Hong Y, Xie Y, Yuan K, Li J, Sun R, Zhang X, Shi X et al (2021) TREM-1 exacerbates neuroinfammatory injury via NLRP3 infammasome-mediated pyroptosis in experimental subarachnoid hemorrhage. Transl Stroke Res 12(4):643–659. <https://doi.org/10.1007/s12975-020-00840-x>
- <span id="page-17-15"></span>146. 2022 Alzheimer's disease facts and fgures. Alzheimer's & dementia: the journal of the Alzheimer's Association, 2022. <https://doi.org/10.1002/alz.12638>
- <span id="page-17-16"></span>147. Long JM, Holtzman DM (2019) Alzheimer disease: an update on pathobiology and treatment strategies. Cell 179(2):312– 339.<https://doi.org/10.1016/j.cell.2019.09.001>
- <span id="page-17-17"></span>148. Hampel H, Nistico R, Seyfried NT, Levey AI, Modeste E, Lemercier P, Baldacci F, Toschi N et al (2021) Omics sciences for systems biology in Alzheimer's disease: state-of-the-art of the evidence. Ageing Res Rev 69:101346. [https://doi.org/10.](https://doi.org/10.1016/j.arr.2021.101346) [1016/j.arr.2021.101346](https://doi.org/10.1016/j.arr.2021.101346)
- <span id="page-17-18"></span>149. Hur J, Frost GR, Wu X, Crump C, Pan SJ, Wong E, Barros M, Li T et al (2020) The innate immunity protein IFITM3 modulates gamma-secretase in Alzheimer's disease. Nature 586(7831):735–740. [https://doi.org/10.1038/](https://doi.org/10.1038/s41586-020-2681-2) [s41586-020-2681-2](https://doi.org/10.1038/s41586-020-2681-2)
- <span id="page-17-19"></span>150. Saresella M, Rosa FL, Piancone F, Zoppis M, Marventano I, Calabrese E, Rainone V, Nemni R et al (2016) The NLRP3 and NLRP1 inflammasomes are activated in Alzheimer's disease. Mol Neurodegener 11:23. [https://doi.org/10.1186/](https://doi.org/10.1186/s13024-016-0088-1) [s13024-016-0088-1](https://doi.org/10.1186/s13024-016-0088-1)
- <span id="page-17-20"></span>151. Ahmed ME, Iyer S, Thangavel R, Kempuraj D, Selvakumar GP, Raikwar SP, Zaheer S, Zaheer A (2017) Co-localization of glia maturation factor with NLRP3 inflammasome and autophagosome markers in human Alzheimer's disease brain. J Alzheimer's Dis 60(3):1143–1160. [https://doi.org/10.3233/](https://doi.org/10.3233/JAD-170634) [JAD-170634](https://doi.org/10.3233/JAD-170634)
- <span id="page-17-21"></span>152. Venegas C, Kumar S, Franklin BS, Dierkes T, Brinkschulte R, Tejera D, Vieira-saecker A, Schwartz S et al (2017) Microgliaderived ASC specks cross-seed amyloid-beta in Alzheimer's disease. Nature 552(7685):355–361. [https://doi.org/10.1038/natur](https://doi.org/10.1038/nature25158) [e25158](https://doi.org/10.1038/nature25158)
- <span id="page-17-22"></span>153. Luciunaite A, Mcmanus RM, Jankunec M, Racz I, Dansokho C, Dalgediene I, Schwartz S, Brosseron F et al (2020) Soluble Abeta oligomers and protofbrils induce NLRP3 infammasome activation in microglia. J Neurochem 155(6):650–661. [https://](https://doi.org/10.1111/jnc.14945) [doi.org/10.1111/jnc.14945](https://doi.org/10.1111/jnc.14945)
- <span id="page-17-23"></span>154. Tejera D, Mercan D, Sanchez-caro JM, Hanan M, Greenberg D, Soreq H, Latz E, Golenbock D et al (2019) Systemic infammation impairs microglial Abeta clearance through NLRP3 infammasome. EMBO J 38(17):101064. [https://doi.org/10.15252/](https://doi.org/10.15252/embj.2018101064) [embj.2018101064](https://doi.org/10.15252/embj.2018101064)
- <span id="page-17-24"></span>155. Ising C, Venegas C, Zhang S, Scheiblich H, Schmidt SV, Vieirasaecker A, Schwartz S, Albasset S et al (2019) NLRP3 infammasome activation drives tau pathology. Nature 575(7784):669– 673. <https://doi.org/10.1038/s41586-019-1769-z>
- <span id="page-17-25"></span>156. Panda C, Voelz C, Habib P, Mevissen C, Pufe T, Beyer C, Gupta S, Slowik A (2021) Aggregated Tau-PHF6 (VQIVYK) potentiates NLRP3 infammasome expression and autophagy in human microglial cells. Cells-Basel 10(7). [https://doi.org/10.3390/cells](https://doi.org/10.3390/cells10071652) [10071652](https://doi.org/10.3390/cells10071652)
- <span id="page-17-26"></span>157. Zhao Y, Tan S, Huang Z, Shan F, Li P, Ning Y, Ye S, Zhao Z et al (2021) NLRP3 infammasome-dependent increases in high mobility group box 1 involved in the cognitive dysfunction caused by Tau-overexpression. Front Aging Neurosci 13:721474. <https://doi.org/10.3389/fnagi.2021.721474>
- <span id="page-17-27"></span>158. Stancu I, Cremers N, Vanrusselt H, Couturier J, Vanoosthuyse A, Kessels S, Lodder C, Brone B et al (2019) Aggregated Tau activates NLRP3-ASC infammasome exacerbating exogenously seeded and non-exogenously seeded Tau pathology *in vivo*. Acta Neuropathol 137(4):599–617. [https://doi.org/10.1007/](https://doi.org/10.1007/s00401-018-01957-y) [s00401-018-01957-y](https://doi.org/10.1007/s00401-018-01957-y)
- <span id="page-17-28"></span>159. Zhang X, Wang R, Hu D, Sun X, Fujioka H, Lundberg K, Chan ER, Wang Q et al (2020) Oligodendroglial glycolytic stress triggers infammasome activation and neuropathology in Alzheimer's disease. Sci Adv 6(49). [https://doi.org/10.1126/sciadv.](https://doi.org/10.1126/sciadv.abb8680) [abb8680](https://doi.org/10.1126/sciadv.abb8680)
- <span id="page-17-29"></span>160. Qi Y, Klyubin I, Cuello AC, Rowan MJ (2018) NLRP3 dependent synaptic plasticity defcit in an Alzheimer's disease

amyloidosis model *in vivo*. Neurobiol Dis 114:24–30. [https://doi.](https://doi.org/10.1016/j.nbd.2018.02.016) [org/10.1016/j.nbd.2018.02.016](https://doi.org/10.1016/j.nbd.2018.02.016)

- <span id="page-18-0"></span>161. Fekete C, Vastagh C, Denes A, Hrabovszky E, Nyiri G, Kallo I, Liposits Z, Sarvari M (2019) Chronic amyloid beta oligomer infusion evokes sustained infammation and microglial changes in the rat hippocampus via NLRP3. Neuroscience 405:35–46. <https://doi.org/10.1016/j.neuroscience.2018.02.046>
- <span id="page-18-1"></span>162. Mackenzie SH, Schipper JL, Clark AC (2010) The potential for caspases in drug discovery. Curr Opin Drug Discov Dev 13(5):568–576
- <span id="page-18-2"></span>163. Yoshihara E (2020) TXNIP/TBP-2: a master regulator for glucose homeostasis. Antioxidants 9(8). [https://doi.org/10.3390/](https://doi.org/10.3390/antiox9080765) [antiox9080765](https://doi.org/10.3390/antiox9080765)
- <span id="page-18-3"></span>164. Rauch JN, Luna G, Guzman E, Audouard M, Challis C, Sibih YE, Leshuk C, Hernandez I et al (2020) LRP1 is a master regulator of tau uptake and spread. Nature 580(7803):381–385. [https://](https://doi.org/10.1038/s41586-020-2156-5) [doi.org/10.1038/s41586-020-2156-5](https://doi.org/10.1038/s41586-020-2156-5)
- <span id="page-18-4"></span>165. Reeve AK, Ludtmann MHR, Angelova PR, Simcox EM, Horrocks MH, Klenerman D, Gandhi S, Turnbull DM et al (2015) Aggregated alpha-synuclein and complex I deficiency: exploration of their relationship in diferentiated neurons. Cell Death Dis 6:1820. <https://doi.org/10.1038/cddis.2015.166>
- <span id="page-18-5"></span>166. Sarkar S, Malovic E, Harishchandra DS, Ghaisas S, Panicker N, Charli A, Palanisamy BN, Rokad D et al (2017) Mitochondrial impairment in microglia amplifes NLRP3 infammasome proinfammatory signaling in cell culture and animal models of Parkinson's disease. NPJ Parkinson's Dis 3:30. [https://doi.org/](https://doi.org/10.1038/s41531-017-0032-2) [10.1038/s41531-017-0032-2](https://doi.org/10.1038/s41531-017-0032-2)
- <span id="page-18-6"></span>167. Fan Z, Pan Y, Zhang Z, Yang H, Yu S, Zheng Y, Ma J, Wang X (2020) Systemic activation of NLRP3 infammasome and plasma alpha-synuclein levels are correlated with motor severity and progression in Parkinson's disease. J Neuroinfammation 17(1):11.<https://doi.org/10.1186/s12974-019-1670-6>
- <span id="page-18-7"></span>168. Wang X, Chi J, Huang D, Ding L, Zhao X, Jiang L, Yu Y, Gao F (2020) alpha-synuclein promotes progression of Parkinson's disease by upregulating autophagy signaling pathway to activate NLRP3 infammasome. Exp Ther Med 19(2):931–938. [https://](https://doi.org/10.3892/etm.2019.8297) [doi.org/10.3892/etm.2019.8297](https://doi.org/10.3892/etm.2019.8297)
- <span id="page-18-8"></span>169. Malhotra S, Rio J, Urcelay E, Nurtdinov R, Bustamante MF, Fernandez O, Oliver B, Zettl U et al (2015) NLRP3 infammasome is associated with the response to IFN-beta in patients with multiple sclerosis. Brain 138(Pt 3):52–644. [https://doi.org/10.1093/brain/](https://doi.org/10.1093/brain/awu388) [awu388](https://doi.org/10.1093/brain/awu388)
- <span id="page-18-9"></span>170. Imani D, Azimi A, Salehi Z, Rezaei N, Emamnejad R, Sadr M, Izad M (2018) Association of nod-like receptor protein-3 single nucleotide gene polymorphisms and expression with the susceptibility to relapsing-remitting multiple sclerosis. Int J Immunogenet 45(6):329–336.<https://doi.org/10.1111/iji.12401>
- <span id="page-18-10"></span>171. Malhotra S, Sorosina M, Rio J, Peroni S, Midaglia L, Villar LM, Alvarez-cermeno JC, Schroeder I et al (2018) NLRP3 polymorphisms and response to interferon-beta in multiple sclerosis patients. Mult Scler 24(11):1507–1510. [https://doi.org/10.1177/](https://doi.org/10.1177/1352458517739137) [1352458517739137](https://doi.org/10.1177/1352458517739137)
- <span id="page-18-11"></span>172. Malhotra S, Costa C, Eixarch H, Keller CW, Amman L, Martinezbanaclocha H, Midaglia L, Sarro E et al (2020) NLRP3 infammasome as prognostic factor and therapeutic target in primary

progressive multiple sclerosis patients. Brain 143(5):1414–1430. <https://doi.org/10.1093/brain/awaa084>

- <span id="page-18-12"></span>173. Khan N, Kuo A, Brockman DA, Cooper MA, Smith MT (2018) Pharmacological inhibition of the NLRP3 infammasome as a potential target for multiple sclerosis induced central neuropathic pain. Infammopharmacology 26(1):77–86. [https://doi.org/10.](https://doi.org/10.1007/s10787-017-0401-9) [1007/s10787-017-0401-9](https://doi.org/10.1007/s10787-017-0401-9)
- <span id="page-18-13"></span>174 O'brien WT, Pham L, Symons GF, Monif M, Shultz SR, Mcdonald SJ (2020) The NLRP3 infammasome in traumatic brain injury: potential as a biomarker and therapeutic target. J Neuroinfammation 17(1):104. [https://doi.org/10.1186/](https://doi.org/10.1186/s12974-020-01778-5) [s12974-020-01778-5](https://doi.org/10.1186/s12974-020-01778-5)
- <span id="page-18-14"></span>175. Zheng B, Zhang S, Ying Y, Guo X, Li H, Xu L, Ruan X (2018) Administration of Dexmedetomidine inhibited NLRP3 infammasome and microglial cell activities in hippocampus of traumatic brain injury rats. Biosci Rep 38(5). 10.1042/BSR20180892
- <span id="page-18-15"></span>176. He Q, Jiang L, Man S, Wu L, Hu Y, Chen W (2018) Curcumin reduces neuronal loss and inhibits the NLRP3 inflammasome activation in an epileptic rat model. Curr Neurovasc Res 15(3):186–192. [https://doi.org/10.2174/15672026156661807311](https://doi.org/10.2174/1567202615666180731100224) [00224](https://doi.org/10.2174/1567202615666180731100224)
- <span id="page-18-16"></span>177. Meng X, Tan L, Tan M, Jiang T, Tan C, Li M, Wang H, Yu J (2014) Inhibition of the NLRP3 infammasome provides neuroprotection in rats following amygdala kindling-induced status epilepticus. J Neuroinfammation 11:212. [https://doi.org/10.](https://doi.org/10.1186/s12974-014-0212-5) [1186/s12974-014-0212-5](https://doi.org/10.1186/s12974-014-0212-5)
- <span id="page-18-17"></span>178. Toscano ECDB, Vieira ELM, Dias BBR, Caliari MV, Goncalves AP, Giannetti AV, Siqueira JM, Suemoto CK et al (2021) NLRP3 and NLRP1 infammasomes are up-regulated in patients with mesial temporal lobe epilepsy and may contribute to overexpression of caspase-1 and IL-beta in sclerotic hippocampi. Brain Res 1752:147230.<https://doi.org/10.1016/j.brainres.2020.147230>
- <span id="page-18-18"></span>179. Jiang M, Li R, Lyu J, Li X, Wang W, Wang Z, Sheng H, Zhang W et al (2020) MCC950, a selective NLPR3 infammasome inhibitor, improves neurologic function and survival after cardiac arrest and resuscitation. J Neuroinfammation 17(1):256. [https://doi.](https://doi.org/10.1186/s12974-020-01933-y) [org/10.1186/s12974-020-01933-y](https://doi.org/10.1186/s12974-020-01933-y)
- <span id="page-18-19"></span>180. Zheng G, Xu J, He F, Hu J, Ge W, Ji X, Wang C, Bradley JL et al (2021) Efects of NLRP3 infammasome blockade on postresuscitation cerebral function in a rat model of cardiopulmonary resuscitation. Biomed Pharmacother 143:112093. [https://doi.org/](https://doi.org/10.1016/j.biopha.2021.112093) [10.1016/j.biopha.2021.112093](https://doi.org/10.1016/j.biopha.2021.112093)
- <span id="page-18-20"></span>181. Xie K, Zhang Y, Wang Y, Meng X, Wang Y, Yu Y, Chen H (2020) Hydrogen attenuates sepsis-associated encephalopathy by NRF2 mediated NLRP3 pathway inactivation. Infamm Res 69(7):697–710.<https://doi.org/10.1007/s00011-020-01347-9>

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.