

Validating GWAS-Identified Risk Loci for Alzheimer's Disease in Han Chinese Populations

Hui-Zhen Wang · Rui Bi · Qiu-Xiang Hu · Qun Xiang · Chen Zhang ·
Deng-Feng Zhang · Wen Zhang · Xiaohong Ma · Wanjun Guo · Wei Deng ·
Liansheng Zhao · Peiyan Ni · Mingli Li · Yiru Fang · Tao Li · Yong-Gang Yao

Received: 9 September 2014 / Accepted: 18 November 2014 / Published online: 3 December 2014
© Springer Science+Business Media New York 2014

Abstract In recent years, genome-wide association studies (GWASs) have identified many novel susceptible genes/loci for Alzheimer's disease (AD). However, most of these studies were conducted in European and populations of European origin, and limited studies have been performed in Han Chinese. In this study, we genotyped 14 single-nucleotide polymorphisms (SNPs) in eight GWAS-reported AD risk genes in 1509 individuals comprising two independent Han Chinese case-control cohorts. Four SNPs (rs11234495, rs592297, rs676733, and rs3851179) in the *PICALM* gene were significantly associated with late-onset (LO)-AD in populations from Southwest China, whereas SNPs rs744373 (*BINI*), rs9331942 (*CLU*), and rs670139 (*MS4A4E*) were linked to

LO-AD in populations from East China. In the combined Han Chinese population, positive associations were observed between *PICALM*, *CLU*, *MS4A4E* genes, and LO-AD. The association between rs3851179 (*PICALM*), rs744373 (*BINI*), and AD was further confirmed by meta-analysis of Asian populations. Our study verified the association between *PICALM*, *BINI*, *CLU*, and *MS4A4E* variants and AD susceptibility in Han Chinese populations. We also discerned some regional differences concerning AD susceptibility SNPs.

Keywords GWAS · Variants · Alzheimer's disease
Han Chinese

Hui-Zhen Wang and Rui Bi contributed equally to this study.

Electronic supplementary material The online version of this article (doi:10.1007/s12035-014-9015-z) contains supplementary material, which is available to authorized users.

H.-Z. Wang · R. Bi · Q.-X. Hu · Q. Xiang · D.-F. Zhang ·
W. Zhang · Y.-G. Yao

Key Laboratory of Animal Models and Human Disease Mechanisms of the Chinese Academy of Sciences & Yunnan Province, Kunming Institute of Zoology, Kunming, Yunnan 650223, China

H.-Z. Wang
School of Life Sciences, University of Science and Technology of China, Hefei, Anhui 230026, China

R. Bi · Q.-X. Hu · Q. Xiang · D.-F. Zhang · W. Zhang · Y.-G. Yao
Kunming College of Life Science, University of Chinese Academy of Sciences, Kunming, Yunnan 650204, China

C. Zhang · Y. Fang
Division of Mood Disorders, Shanghai Mental Health Center, Shanghai Jiao Tong University School of Medicine, Shanghai 200030, China

X. Ma · W. Guo · W. Deng · L. Zhao · P. Ni · M. Li · T. Li
The Mental Health Center and Psychiatric Laboratory, West China Hospital, Sichuan University, Chengdu, Sichuan 610064, China

X. Ma · W. Guo · W. Deng · L. Zhao · P. Ni · M. Li · T. Li
The State Key Laboratory of Biotherapy, West China Hospital, Sichuan University, Chengdu, Sichuan 610064, China

Y.-G. Yao (✉)
Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming, Yunnan 650223, China
e-mail: yaoyg@mail.kiz.ac.cn

Y.-G. Yao
e-mail: ygyaozh@gmail.com

Y.-G. Yao
CAS Center for Excellence in Brain Science, Chinese Academy of Sciences, Shanghai 200031, China

Introduction

Alzheimer's disease (AD) is the most prevalent neurodegenerative disease, which mainly leads to memory loss in the elderly population over 65 years old. Extracellular senile plaques (SP), which were mainly composed of amyloid β ($A\beta$) peptide and intracellular neurofibrillary tangles (NFT) formed by hyperphosphorylated tau protein, are the hallmarks observed in AD brain [1]. It has been widely accepted that AD can be divided into early-onset (EO-AD, <65 years) and late-onset Alzheimer's disease (LO-AD, \geq 65 years).

Genetic linkage studies have identified rare mutations in the amyloid precursor protein (*APP*), presenilin 1 and presenilin 2 (*PSEN1* and *PSEN2*) genes in early-onset familial Alzheimer's disease (EO-FAD) [2]. Most of these mutations are inherited in an autosomal dominant manner [3, 4], and can lead to an increased ratio of $A\beta_{42}/A\beta_{40}$ or $A\beta$ aggregation [5, 6]. However, the etiology of LO-AD is extremely intricate and influenced by complicated interactions between genetic components and environmental risk factors. The $\epsilon 4$ allele of the apolipoprotein E gene (*APOE*) has been established unequivocally as a susceptible gene for LO-AD, which confers 4–15-fold risk to AD in $\epsilon 4$ allele carriers than in noncarriers [2]. In recent years, several genome-wide association studies (GWAS) were performed to identify risk gene/loci for LO-AD, and ten novel AD risk genes (*BINI*, *CLU*, *ABCA7*, *CRI*, *PICALM*, *MS4A6A*, *CD33*, *MS4A4E*, *CD2AP*, and *EPHA1*) were identified [7–11]. Some of these genes were repeatedly identified as AD risk genes in subsequent independent studies [12–14]. In 2011, two large GWASs of European origin populations identified another five potential AD risk genes (*ABCA7*, *MS4A6A/MS4A4E*, *CD33*, *CD2AP*, and *EPHA1*); meanwhile, these two GWASs successfully replicated the association between those previously reported loci (*CLU*, *CRI*, *PICALM*, and *BINI*) and LO-AD [7, 8]. Collectively, these risk genes can explain around 50 % heritability of LO-AD and are involved in pathways of production, degradation, and clearance of $A\beta$, cholesterol metabolism, immunity, and cellular signaling [2].

It should be noted that these large-scale GWASs about AD were conducted in European and populations of European origin. Several case-control studies had been carried out to validate the potential association of these genes with AD in different populations [12, 13, 15, 16]. Recently, Tan et al. [17] assessed the association of GWAS-linked genes/loci with LO-AD in a Han Chinese population from North China, and they confirmed that the *MS4A6A* and *CD33* genes were associated with LO-AD.

In this study, we aimed to test whether these GWAS-associated AD risk genes confer genetic susceptibility to AD in different Han Chinese populations from Southwest and East China. We screened 14 single-nucleotide polymorphisms

(SNPs) of the GWAS-associated AD risk genes in two independent sample sets and verified the association between several loci of those GWAS-reported genes and AD in Han Chinese populations.

Materials and Methods

Subjects

Two independent samples from Southwest China and East China were recruited and analyzed in this study. The cohort from Southwest China was collected at the Mental Health Center of West China Hospital, which was composed of 333 unrelated sporadic LO-AD patients and 334 cognitively healthy controls. The cohort from East China was composed of 416 unrelated sporadic LO-AD patients and 426 cognitively healthy individuals, which was recruited from the Shanghai Mental Health Center and Tongde Hospital of Zhejiang Province. Around 67 % of the AD patient and control samples had been genotyped for the *LRRK2* genetic polymorphisms in our previous study [18]. All participants were of Han Chinese origin. Patients were independently diagnosed by two psychiatrists according to the criteria of DSM-IV and NINCDS-ADRDA [19]. The healthy controls were confirmed as cognitively intact and neurologically normal. All patients were identified as sporadic LO-AD since none of their first-degree relatives had dementia. Written informed consents following the principles of the Declaration of Helsinki were obtained from each participant or guardian. This study was approved by the institutional review board of the Kunming Institute of Zoology, Chinese Academy of Sciences.

SNP Selection and Genotyping

A total of 14 SNPs, including 8 GWAS-reported variants (*BINI*, rs744373; *CLU*, rs11136000; *ABCA7*, rs3764650; *PICALM*, rs3851179; *MS4A6A*, rs610932; *CD33*, rs3865444; *MS4A4E*, rs670139; *CD2AP*, rs9349407) and 6 tag SNPs (*BINI*, rs1060743; *CLU*, rs9331942; *ABCA7*, rs3752237; *PICALM*, rs11234495, rs592297, and rs676733), were selected according to the linkage disequilibrium (LD) pattern of the respective genes based on data from HapMap (HapMap, <http://hapmap.ncbi.nlm.nih.gov/>, phase 3, CHB) and 1000 Genomes (<http://www.broadinstitute.org/mpg/snap/>). These tag SNPs had a r^2 value >0.8 with the other SNPs and/or were located in different LD blocks with the GWAS hit SNPs. For those genes that span a relatively short genomic region (*MS4A6A*, *CD33*, *MS4A4E*), or tag SNPs that were located in the same LD block with the GWAS hit SNPs (*CD2AP*), we did not analyze tag SNPs but only the GWAS hit SNPs. The detailed information of each SNP is shown in Table 1. We genotyped different alleles of the *APOE* gene,

Table 1 Detailed information of 14 SNPs in eight GWAS-linked genes

Gene	dbSNP rs no. ^a	Allele	MAF ^b	Chr.	Position	Annotation	Source ^c
<i>BINI</i>	rs744373	C/T	0.33	2	127611085	5' near region	GWAS
	rs1060743	C/T	0.39	2	127542003	Synonymous	Tag SNP
<i>CLU</i>	rs11136000	C/T	0.21	8	27520436	Intron 3	GWAS
	rs9331942	C/T	0.34	8	27511031	3' near region	Tag SNP
<i>ABCA7</i>	rs3764650	G/T	0.27	19	997520	Intron 12	GWAS
	rs3752237	A/G	0.16	19	998161	Synonymous	Tag SNP
<i>PICALM</i>	rs3851179	A/G	0.47	11	85546288	5' near region	GWAS
	rs11234495	C/T	0.41	11	85353082	3' near region	Tag SNP
	rs592297	C/T	0.41	11	85403585	Synonymous	Reported
	rs676733	C/T	0.46	11	85411658	Intron 3	Tag SNP
<i>MS4A6A</i>	rs610932	A/C	0.34	11	59695883	3' near region	GWAS
<i>CD33</i>	rs3865444	G/T	0.18	19	56419774	5' near region	GWAS
<i>MS4A4E</i>	rs670139	A/C	0.42	11	59728371	5' near region	GWAS
<i>CD2AP</i>	rs9349407	C/G	0.12	6	47561337	Intron 1	GWAS

^a SNPs in bold were distilled from previous GWAS reports, and other SNPs were selected according to linkage disequilibrium blocks of the respective gene based on HapMap (<http://hapmap.ncbi.nlm.nih.gov/>) CHB data set and 1000 Genomes (<http://www.broadinstitute.org/mpg/snap/>)

^b Minor allele frequency in CHB data set was obtained from 1000 Genomes database (<http://asia.ensembl.org/>)

^c The SNPs marked with “GWAS” were obtained from Alzgene (<http://www.alzgene.org/TopResults.asp>)

which was a well-established risk gene for LO-AD [20, 21], using the same approach described in our previous study [18].

All these 14 SNPs were genotyped by using the SNaPshot assay, which comprising a multiplex PCR and a subsequent single-base extension process, according to the detailed step-by-step procedure in our previous studies [18, 22]. Briefly, multiplex PCR was carried out in a volume of 8- μ L reaction solution containing 20–50 ng genomic DNA, 0.4 mM dNTPs, 0.2–0.5 μ M of each primer (Supplementary Table S1), 2.0 mM MgCl₂, and 1.0 U of FastStart Taq DNA Polymerase (Roche Applied Science). The amplification program was composed of a pre-denaturation cycle at 94 °C for 2 min; 40 amplification cycles at 94 °C for 30 s, 55 °C for 30 s, and 72 °C for 1 min; and ended with an incubation cycle at 4 °C. Multiplex PCR products were cleaned up at 72 °C for 40 min with 1.0 U of shrimp alkaline phosphatase (SAP) and 0.5 U of Exonuclease I (TaKaRa Biotechnology Co. Ltd., Dalian, China), followed by a final incubation at 96 °C for 10 min to inactivate the enzyme. The single-base extension was conducted in a total of 10- μ L reaction solution containing 4- μ L multiplex PCR products, 5- μ L SNaPshot Multiplex Ready Reaction Mix, and 0.4–0.8- μ M pooled SNP-specific oligonucleotide primers (Supplementary Table S1) according to the protocol of the ABI PRISM[®] SNaPshot[®] Multiplex Kit (Applied Biosystems). The single-base extension program contained 25 cycles of 96 °C for 10 s, 50 °C for 5 s, and 60 °C for 30 s. The products were then purified by SAP (1.0 U) at 37 °C for 40 min, followed by a heat inactivation at 75 °C for 20 min. A mixture of 4- μ L purified products and 9 μ L Hi-Di[™] formamide was analyzed by the ABI

PRISM[™] 3730xl DNA analyzer (Applied Biosystems) at the Kunming Biodiversity Large-Apparatus Regional Center, Kunming Institute of Zoology. The GeneMarker software (<http://www.softgenetics.com/GeneMarker.html>) was employed to read the genotyping results..

Statistical Analysis and Meta-Analysis

The power analysis was calculated using the Quanto software [23]. The frequency of allele, genotype, and haplotype was analyzed using the PLINK software [24]. Binary logistic regression under the general model was performed by SPSS 16.0 (SPSS Inc., Chicago, Illinois) to assess the association between these SNPs and LO-AD, with an adjustment of *APOE* $\epsilon 4$ status (*APOE* $\epsilon 4^+$ or *APOE* $\epsilon 4^-$). A *P* value <0.05 was regarded as statistically significant. When considering Bonferroni correction for multiple testing, a cutoff of *P*<0.0035 (0.05/14) was set as statistically significant. The deviation from the Hardy-Weinberg equilibrium of each SNP was calculated for both the control and case populations by using the PLINK software [24]; SNPs with a *P* value less than 0.001 were regarded as departure from the Hardy-Weinberg equilibrium. LD structures of the *BINI*, *CLU*, *PICALM*, and *ABCA7* genes were reconstructed using Haploview software version 4.2 [25]. Haplotypes were reconstructed by using the PHASE 2.0 program [26]. All the above analyses were performed for the two independent cohorts separately. Considering the fact that these individuals are all of Han Chinese origin and a combined sample may increase the power of the statistical test [27], we pooled all the samples together to investigate

the association between these GWAS hit SNPs and tag SNPs and AD risk even though there existed some potential genetic differences between populations from Southwest China and East China. In addition, we investigated whether these SNPs affect messenger RNA (mRNA) expression level of relevant genes in different tissues in the GTEx database (<http://www.gtexportal.org/home/>) [28].

We performed meta-analysis for SNPs rs3851179, rs3764650, rs11136000, rs744373, rs610932, rs3865444, and rs9331942 with AD by using the RevMan 5.2 software (<http://www.cochrane.org/revman>) based on data from this study and reported Asian populations (Supplementary Table S2). Those SNPs with no available data in Asian populations were not considered in the meta-analysis.

Results

Statistical Power and LD Pattern of Analyzed SNPs

As the minor allele frequency (MAF) of all SNPs in this study ranged from 11.9 to 46.4 %, assuming a false positive rate controlled as 0.05, the power to detect the odds ratio (OR) value as 1.5 for risk allele was expected to be from 72.8 to 95.5 % based on the samples from Southwest China. However, when we considered the OR value as 1.2, the power was much reduced, with a value range from 20.0 to 38.2 %.

Total genotyping call rate of all individuals was 99.8 %. The allele and genotype counts for each SNP were shown in Supplementary Table S3. All 14 SNPs analyzed in this study were in Hardy-Weinberg equilibrium in both AD patients and controls (Supplementary Table S3). We validated the genotyping results of SNaP-shot assay by direct sequencing 5 % of total samples and obtained consistent results. The $\epsilon 4$ allele of the *APOE* gene showed a significant association with AD risk in both cohorts and in the combined Han Chinese sample (Supplementary Table S4). The LD patterns of these SNPs in the *BINI* (rs1060743-rs744373), *CLU* (rs9331942-rs11136000), *PICALM* (rs11234495-rs592297-rs676733-rs3851179), and *ABCA7* (rs3764650-rs3752237) genes were similar between AD patients and controls (Fig. 1).

SNPs in the *PICALM* Gene Were Associated with LO-AD in Han Chinese

Genotype frequencies of four SNPs of the *PICALM* gene were significantly different between the case and control populations from Southwest China (rs11234495, $P=0.013$; rs592297, $P=0.035$; rs676733, $P=0.0004$; rs3851179, $P=0.001$), and two of them also showed a significant difference

at the allelic level (rs11234495, $P=0.016$, OR=0.767, 95 % confidence interval (CI) 0.618–0.951; rs676733, $P=0.030$, OR=0.782, 95 % CI 0.626–0.976) (Table 2). We failed to discern any positive association of these SNPs with LO-AD in samples from East China. When we combined these two cohorts together, the results remained to be significant (Table 2). Logistic regression analysis revealed that these SNPs were significantly associated with LO-AD in populations from Southwest China and the combined sample after adjustment for the *APOE4* status (Table 2). However, after Bonferroni correction for multiple testing, only SNPs rs676733 and rs3851179 remained to be significantly associated with AD (Table 2) in the cohort from Southwest China. The inconsistency of association between the two independent cohorts might simply be an issue of low power of the study and/or potential region difference.

Haplotype construction based on the four SNPs of the *PICALM* gene revealed that haplotype TTTG (SNP order: rs11234495-rs592297-rs676733-rs3851179) increased the risk of AD in the Southwest China cohort ($P=0.043$, OR=1.256, 95 % CI 1.013–1.558), haplotype CTCA increased the risk of AD in the East China cohort ($P=0.023$, OR=1.620, 95 % CI 1.070–2.454), and haplotype CCCA decreased the risk of AD in the combined population ($P=0.039$, OR=0.848, 95 % CI 0.726–0.991) (Table 3). However, all haplotype associations did not survive the Bonferroni correction for multiple testing, possibly due to the low power of this study.

Intriguingly, the GWAS hit variant rs3851179 and tag SNP rs592297 of the *PICALM* gene were correlated to its mRNA level in brain cingulate cortex (rs3851179, $P=0.01$; rs592297, $P=0.01$) according to expression data in the GTEx database (<http://www.broadinstitute.org/gtex/>), suggesting that these variants might be functional.

SNP rs3851179 of the *PICALM* Gene Confers Susceptibility to LO-AD in Asian Populations

SNP rs3851179 (*PICALM*) has been reported to be a susceptibility locus for AD in European populations [9–11], whereas recent studies in different Asian populations had controversial results. In three reported Han Chinese studies, rs3851179 was not associated with AD [15, 29, 30], but in one Japanese population, it showed a weak association [31].

To evaluate the association between rs3851179 and AD, we performed a meta-analysis of 7739 individuals from Asian populations, which were composed of the above-mentioned four validation studies and the two cohorts analyzed in this study (Supplementary Table S2). Results of the Cochran's Q test indicated no significant heterogeneity among these populations. The allele test ($Z=2.97$, $P=0.003$, OR=0.90, 95 % CI 0.84–0.96) indicated a significant association between rs3851179 and LO-AD (Fig. 2a).

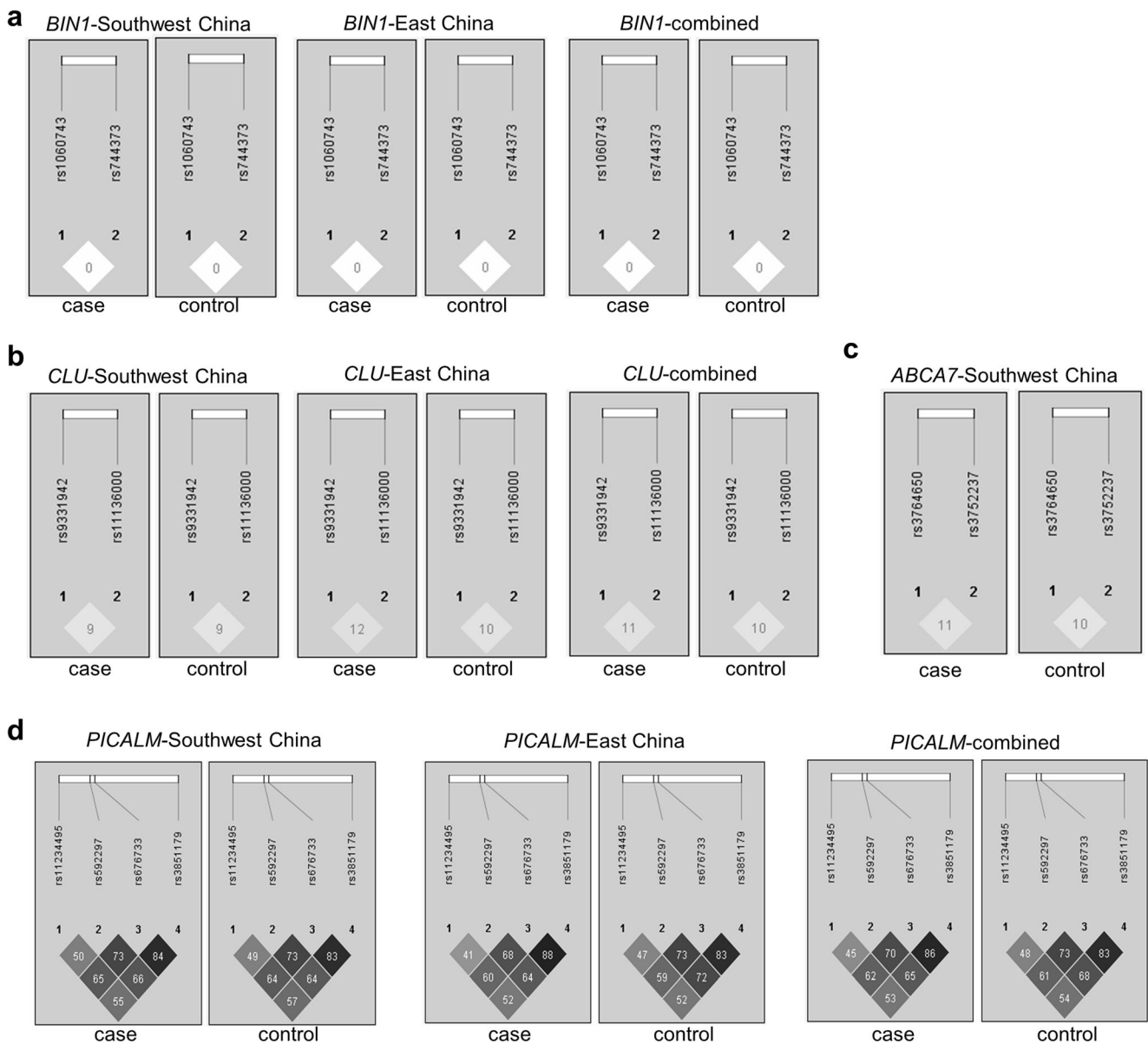


Fig. 1 Linkage disequilibrium (LD) patterns of the *BIN1* (a), *CLU* (b), *ABCA7* (c), and *PICALM* (d) genes in AD cases and controls from Southwest China, East China, and combined population. The value in each square refers to $r^2 \times 100$

SNPs of the *BIN1* (rs744373), *CLU* (rs9331942), *MS4A4E* (rs670139), and *CD2AP* (rs9349407) Genes Were Associated to LO-AD in Han Chinese

In Han Chinese cohort from East China, SNPs rs744373 (*BIN1*, $P=0.038$), rs9331942 (*CLU*, $P=0.013$), and rs670139 (*MS4A4E*, $P=0.022$) were significantly different between cases and controls at the genotypic level. SNP rs744373 (*BIN1*, $P=0.026$, OR=1.256, 95 % CI 1.028–1.535) and rs9349407 (*CD2AP*, $P=0.037$, OR=1.256, 95 % CI 1.013–1.558) showed a significant difference at the allelic level (Table 2) in the East China cohort and the combined

sample, respectively. However, we failed to discern any association between these SNPs (rs744373, rs9331942, rs670139, and rs9349407) and AD in Han Chinese from Southwest China. Note that a positive association of SNPs rs9331942 and rs670139 with LO-AD was observed in the combined Han Chinese populations both before and after adjustment for *APOE4* status. Nonetheless, all these associations were no longer significant after Bonferroni correction for multiple testing.

Haplotype analysis showed that there was no significant difference of haplotype frequency of *BIN1* and *CLU* SNPs between AD cases and controls in both cohorts and the

Table 2 Association between SNPs in GWAS-linked genes and AD in Han Chinese

Gene	SNP ID	Populations	MAF (case/control)	OR (95 % CI)	P value (allelic) ^a	P value (genotypic) ^a	P value trend ^a	Adjusted P value ^b
<i>BIN1</i>	rs1060743	Southwest	0.456/0.464	0.969 (0.781–1.202)	0.777	0.692	0.777	0.557
		East	0.445/0.413	1.138 (0.938–1.380)	0.191	0.238	0.184	0.166
		Combined	0.450/0.436	1.060 (0.918–1.224)	0.428	0.715	0.425	0.667
	rs744373	Southwest	0.366/0.361	1.024 (0.820–1.281)	0.832	0.884	0.838	0.874
		East	0.380/0.328	1.256 (1.028–1.535)	0.026	0.038	0.023	0.038
		Combined	0.374/0.342	1.147 (0.989–1.332)	0.071	0.164	0.073	0.187
<i>CLU</i>	rs11136000	Southwest	0.215/0.196	1.125 (0.862–1.468)	0.385	0.280	0.399	0.209
		East	0.210/0.185	1.170 (0.920–1.487)	0.200	0.442	0.207	0.304
		Combined	0.213/0.190	1.150 (0.962–1.374)	0.124	0.185	0.133	0.107
	rs9331942	Southwest	0.297/0.310	0.942 (0.746–1.190)	0.617	0.331	0.622	0.297
		East	0.325/0.356	0.871 (0.711–1.065)	0.178	0.013	0.177	0.017
		Combined	0.312/0.336	0.900 (0.773–1.048)	0.175	0.006	0.177	0.005
<i>PICALM</i>	rs3851179	Southwest	0.377/0.422	0.828 (0.665–1.031)	0.091	0.001 ^d	0.085	0.002 ^d
		East	0.371/0.385	0.943 (0.774–1.148)	0.557	0.716	0.549	0.771
		Combined	0.374/0.401	0.890 (0.769–1.031)	0.119	0.011	0.112	0.024
	rs11234495	Southwest	0.453/0.520	0.767 (0.618–0.951)	0.016	0.013	0.016	0.019
		East	0.494/0.500	0.976 (0.806–1.182)	0.804	0.139	0.805	0.145
		Combined	0.483/0.509	0.901 (0.781–1.040)	0.153	0.011	0.155	0.019
	rs592297	Southwest	0.298/0.347	0.798 (0.633–1.005)	0.055	0.035	0.055	0.048
		East	0.298/0.331	0.856 (0.696–1.052)	0.140	0.249	0.126	0.268
		Combined	0.298/0.338	0.830 (0.711–0.968)	0.017	0.014	0.015	0.018
	rs676733	Southwest	0.353/0.411	0.782 (0.626–0.976)	0.030	0.0004 ^d	0.028	0.002 ^d
		East	0.383/0.375	1.036 (0.850–1.262)	0.730	0.891	0.725	0.901
		Combined	0.370/0.391	0.914 (0.789–1.060)	0.234	0.027	0.227	0.066
<i>MS4A6A</i>	rs610932	Southwest	0.374/0.377	0.984 (0.788–1.229)	0.888	0.486	0.887	0.695
		East	0.368/0.365	1.012 (0.830–1.234)	0.906	0.960	0.901	0.820
		Combined	0.370/0.370	1.000 (0.862–1.159)	0.997	0.827	0.997	0.973
<i>CD33</i>	rs3865444	Southwest	0.200/0.189	1.073 (0.818–1.408)	0.609	0.831	0.616	0.617
		East	0.208/0.201	1.042 (0.823–1.321)	0.731	0.930	0.732	0.845
		Combined	0.204/0.196	1.055 (0.883–1.262)	0.554	0.837	0.558	0.667
<i>MS4A4E</i>	rs670139	Southwest	0.438/0.423	1.063 (0.856–1.321)	0.580	0.269	0.585	0.318
		East	0.393/0.419	0.897 (0.738–1.090)	0.273	0.022	0.274	0.049
		Combined	0.413/0.421	0.968 (0.838–1.119)	0.622	0.012	0.664	0.019
<i>CD2AP</i>	rs9349407	Southwest	0.146/0.121	1.235 (0.900–1.696)	0.190	0.327	0.191	0.338
		East	0.135/0.109	1.273 (0.950–1.707)	0.106	0.134	0.105	0.091
		Combined	0.140/0.115	1.256 (1.013–1.558)	0.037	0.108	0.037	0.058
<i>ABCA7^c</i>	rs3764650	Southwest	0.332/0.311	1.104 (0.877–1.390)	0.401	0.703	0.404	0.939
	rs3752237	Southwest	0.184/0.182	1.014 (0.768–1.339)	0.923	0.630	0.924	0.639

MAF minor allele frequency

^a P value was calculated by PLINK software [24]

^b Binary logistic regression analysis was performed to assess the association of these GWAS-linked genes with AD, with an adjustment for *APOE4* status

^c SNPs rs3764650 and rs3752237 were not genotyped in samples from East China

^d The association of rs3851179 and rs676733 with AD remained significant after Bonferroni correction for multiple testing, in which $P < 0.0035$ (0.05/14) was set as statistically significant

combined population (Table 3). Notably, SNPs rs744373 (*BIN1*, $P=0.009$) and rs670139 (*MS4A4E*, $P=0.03$) were

found to be significantly associated with their mRNA levels in pituitary, and rs670139 showed a weak association with

Table 3 Haplotype frequencies of SNPs in the *BINI*, *CLU*, *PICALM*, and *ABCA7* genes in 333 AD cases and 334 healthy controls from Southwest China and in 416 AD cases and 426 normal controls from East China

Haplotype ^a	Southwest China		P value	OR (95 % CI)		P value	East China		P value	OR (95 % CI)		P value	Combined	
	Case	Control		Case	Control		Case	Control		Case	Control			
													Case	Control
rs1060743-rs744373 (<i>BINI</i>)														
CC	151	151	1.000	1.004 (0.777–1.297)	198	172	0.078	1.235 (0.980–1.556)	349	323	0.189	1.126 (0.948–1.336)		
TC	93	90	0.812	1.042 (0.763–1.424)	117	107	0.389	1.139 (0.860–1.510)	210	197	0.424	1.095 (0.888–1.349)		
CT	152	159	0.700	0.947 (0.734–1.220)	172	180	0.857	0.973 (0.769–1.231)	324	339	0.660	0.961 (0.809–1.142)		
TT	270	268	0.911	1.018 (0.818–1.267)	345	393	0.056	0.827 (0.682–1.003)	615	661	0.185	0.905 (0.783–1.046)		
rs9331942-rs11136000 (<i>CLU</i>)														
TT	143	130	0.378	1.132 (0.867–1.477)	174	156	0.197	1.180 (0.927–1.501)	317	286	0.111	1.158 (0.969–1.385)		
CC	196	206	0.592	0.935 (0.740–1.182)	269	301	0.198	0.875 (0.715–1.071)	465	507	0.185	0.899 (0.772–1.048)		
TC	325	331	0.785	0.970 (0.783–1.203)	388	393	0.845	1.021 (0.843–1.236)	713	724	1.000	0.999 (0.866–1.152)		
Rare ^b	2	1			1	2			3	3				
rs11234495-rs592297-rs676733-rs3851179 (<i>PICALM</i>)														
CCCA	190	221	0.075	0.807 (0.640–1.019)	240	265	0.338	0.898 (0.729–1.106)	430	486	0.039	0.848 (0.726–0.991)		
CTCA	37	42	0.643	0.877 (0.560–1.383)	60	39	0.023	1.620 (1.070–2.454)	97	81	0.216	1.221 (0.901–1.656)		
TTTA	20	15	0.398	1.348 (0.684–2.656)	10	15	0.422	0.679 (0.303–1.520)	30	30	1.000	1.008 (0.605–1.681)		
CTTG	65	69	0.785	0.939 (0.657–1.342)	103	98	0.599	1.087 (0.810–1.460)	168	167	0.908	1.016 (0.809–1.275)		
TTTG	344	307	0.043	1.256 (1.013–1.558)	404	411	0.922	1.013 (0.837–1.226)	748	718	0.202	1.100 (0.954–1.269)		
Rare ^b	10	14			15	24			25	38				
rs3764650-rs3752237 (<i>ABCA7</i>)														
GG	220	207	0.446	1.099 (0.873–1.383)	–	–	–	–	–	–	–	–		
TA	124	123	0.944	1.014 (0.769–1.336)	–	–	–	–	–	–	–	–		
TG	322	338	0.412	0.914 (0.737–1.133)	–	–	–	–	–	–	–	–		

^aThe haplotype analysis was performed by using PHASE 2.0 program [26]

^bHaplotypes with a frequency less than 3 % were pooled together

mRNA level of the *MS4A4E* gene in brain cingulate cortex ($P=0.07$) based on the GTEx database.

Meta-Analysis of the *ABCA7*, *CLU*, *BIN1*, *MS4A6A*, and *CD33* Genes with LO-AD in Asian Populations

Meta-analysis of SNPs rs3764650 (*ABCA7*), rs11136000 (*CLU*), rs9331942 (*CLU*), rs744373 (*BIN1*), rs610932 (*MS4A6A*), and rs3865444 (*CD33*) by using the reported data from Asian populations and data from this study revealed that rs744373 (*BIN1*) and rs11136000 (*CLU*) were significantly associated with LO-AD risk in Asian populations (rs744373: $Z=3.22$, $P=0.001$, $OR=1.14$, 95 % CI 1.05–1.24; rs11136000: $Z=2.05$, $P=0.04$, $OR=0.93$, 95 % CI 0.87–1.00) (Fig. 2b, c), while no significant association was observed for other SNPs (Fig. 2d–g). It should be mentioned that the association pattern of the two *CLU* SNPs were different in our samples and in the result of meta-analysis: rs9331942 was not associated with AD in the meta-analysis but was positively associated with AD in Han Chinese, whereas rs11136000 had a reverse association pattern with rs9331942.

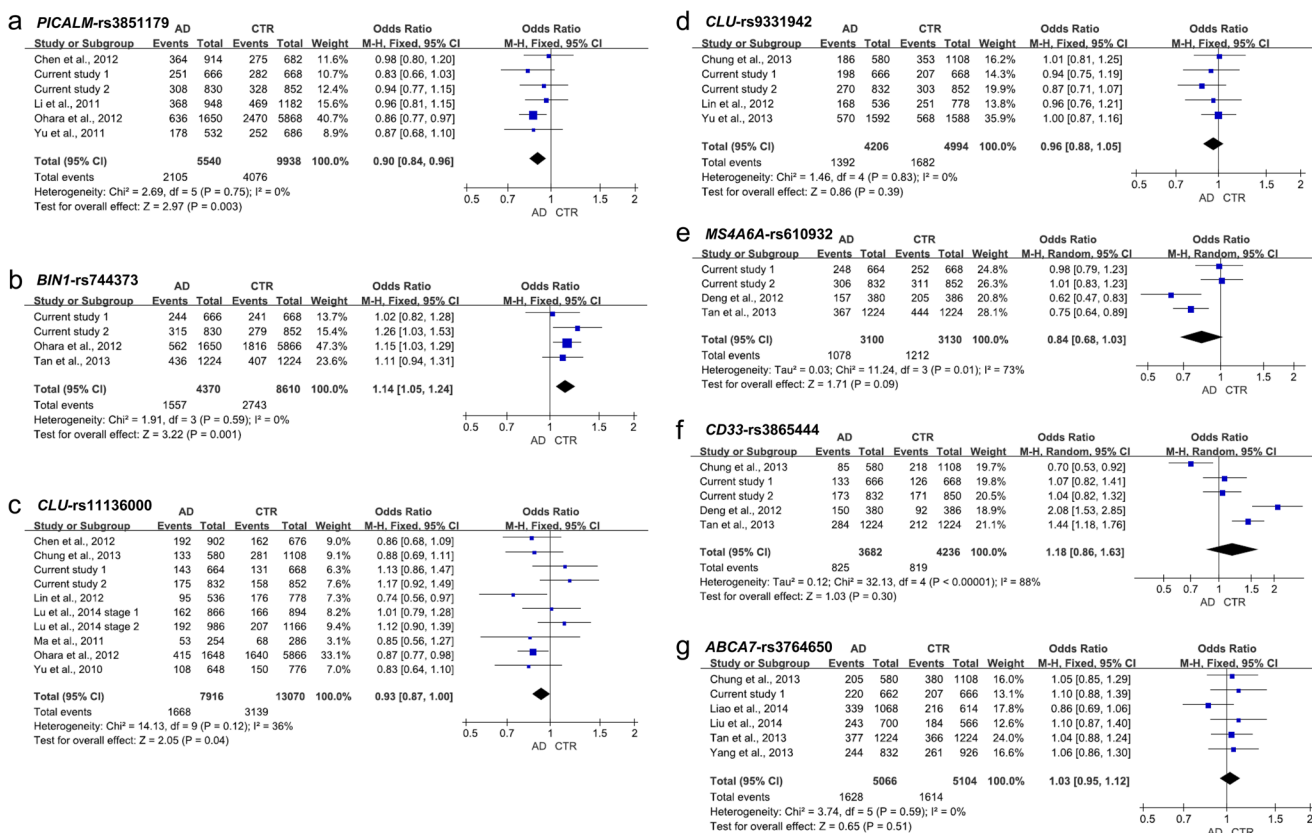


Fig. 2 Meta-analysis of SNPs rs3851179, rs744373, rs11136000, rs9331942, rs610932, rs3865444, and rs3764650 with AD in Asian populations. Allele tests of SNPs rs3851179 (a), rs744373 (b), rs11136000 (c), rs9331942 (d), rs610932 (e), rs3865444 (f), and

Discussion

GWAS is a burgeoning method to seek novel susceptibility genes/loci for AD. Recently, several GWASs had identified many novel AD susceptibility genes/loci in European and populations of European origin [7–11]. Independent replication studies in different populations are important for interpreting the case-and-effect relationship and the mechanism of genetic susceptibility.

In order to investigate whether the reported AD susceptibility loci confer risk to AD in Han Chinese populations, we screened GWAS hit and tag SNPs in eight GWAS-reported AD risk genes in two independent Han Chinese sample sets. Albeit the sample size of our sample was modest, we were able to validate the association of three SNPs (rs744373 of *BIN1*, rs3851179 of *PICALM*, and rs670139 of *MS4A4E*) with LO-AD in our Han Chinese samples, and the effect direction of these AD risk SNPs was as reported in previous GWAS [7, 8, 10, 11]. SNP rs9349407 of *CD2AP* was marginally associated with LO-AD in population from East China, with similar effect direction as reported in previous studies [7, 8]. The GWAS hit SNP rs11136000 in

rs3764650 (g) were performed by using the RevMan 5.2 software (<http://www.cochrane.org/revman>). The detailed information of the reported data from Asian populations is listed in the Supplementary Table S2

the *CLU* gene showed no association with AD in our study, but we observed a positive association of another SNP rs9331942 in this gene with LO-AD.

Of note, the selected *PICALM* tag SNPs in this study (rs11234495, rs592297, and rs676733) were all associated with LO-AD in our Han Chinese population from Southwest China and the combined population, indicating an important role of the *PICALM* gene in AD. Moreover, the association of rs3851179 and rs676733 of the *PICALM* gene with AD risk in the cohort from Southwest China (rs3851179, $P=0.002$; rs676733, $P=0.002$, with an adjustment of the *APOE* $\epsilon 4$ status) remained to be significant even after Bonferroni correction.

The *PICALM* gene encodes phosphatidylinositol-binding clathrin assembly protein (PICALM), which is a key component in clathrin-mediated endocytosis [32]. As APP must be internalized into cells through endocytosis before being cleaved to A β [33], PICALM may play a key role in the A β production and release and influence A β levels. In this study, all four *PICALM* SNPs (rs3851179, rs11234495, rs592297, and rs676733) were significantly associated with LO-AD. Several haplotypes determined by these SNPs also conferred AD risk. Note that this association did not survive the Bonferroni correction, which can be explained by the limited sample size of this study. The GWAS hit SNP rs3851179 (which is located at 88.5-kb upstream of *PICALM*) and tag SNPs rs11234495 and rs676733 (both are within the intron region) may be linked to some functional variants conferring the etiology of AD. SNP rs592297 is a synonymous variant in exon 5 that may potentially influence exon splicing, which was first reported to be associated with LO-AD in a large-scale GWAS, but the association did not reach statistical significance at the genome level (2×10^{-7}) [11]. Furthermore, according to the GTEx database, SNPs rs3851179 and rs592297 were associated with mRNA expression level of the *PICALM* gene in brain cingulate cortex, which was identified to be involved in learning and memory process [34]. Though the association between rs3851179 and LO-AD has been widely identified in European populations [9–13] and was validated in this study, several replication studies produced inconsistent results in different populations [12, 30, 35]. Meta-analysis of rs3851179 based on the data of this study and previously reported studies [15, 29–31] demonstrated that this SNP indeed conferred susceptibility to LO-AD in Asians, which was in accordance with a recent report by Liu and colleagues [36]. Our result and previous reports supported a notion that the *PICALM* gene was a common susceptibility gene for AD in different populations.

Gene expression data in the GTEx database indicated that the GWAS-associated SNPs in the *BINI* (rs744373) and *MS4A4E* (rs670139) genes may affect AD risk through their influence on gene expression. SNP rs744373 of *BINI* was confirmed to be associated with AD risk in different ethnic

backgrounds including East Asian and Caucasian populations according to a meta-analysis based on several validation studies [37]. In this study, we also observed an association between rs744373 and AD risk. Furthermore, meta-analysis for rs744373 in Asian populations supported its role in AD. The *BINI* mRNA expression level was found to be significantly associated with different genotypes of rs744373 in pituitary region by re-analyzing expression data from the GTEx database, which was in line with previous evidence that variant rs59335482 (rs744373 is tightly linked to this SNP) could mediate AD risk by increasing cerebral expression of *BINI* [38]. In addition, we successfully replicated the association between rs670139 and LO-AD in this study. SNP rs670139 lies within an intergenic region between the *MS4A4E* and *MS4A6A* genes [7]. The gene expression data of the GTEx database indicated a potential association between rs670139 and *MS4A4E* mRNA level in both cingulate cortex and pituitary region. One recent study also demonstrated that rs670139 was associated with AD patient's Braak tangle and Braak plaque score [39]. There may be some other functional variants that were tightly linked with rs670139 and affected the expression and function of nearby genes [40]. SNPs rs9331942 in the 3' UTR of *CLU* and rs9349407 in the 5' UTR of *CD2AP* showed an association with AD in the combined Han Chinese sample, which may participate in the transcriptional regulation of the related genes. Further experimental assay should be performed to solidify our speculations.

Failure in replicating GWAS results is a common issue in genetic studies of complex disease, which is influenced by genetic heterogeneity and environmental risk factors [41]. For instance, we recently failed to validate the GWAS hit risk loci for schizophrenia in Han Chinese, albeit these susceptibility loci were initially identified in the same ethnic populations [42]. We encountered similar conditions in this study, in which the association of certain GWAS hit SNPs with AD was inconsistent between the two case-control cohorts from Southwest China and East China (Table 2). The discrepancy may be attributed to several reasons. First, the weak statistical power due to relatively small sample size may account for the inconsistent results in different populations. Second, we only considered the GWAS top hit SNP in each locus and one to four tag SNPs for some of the top hit genes. As the GWAS hit SNPs are most probably not the functional variants, but themselves tag SNPs of the functional variants (in populations from European ancestry). Those top hit SNPs might not correctly tag the functional variants in Han Chinese because the LD pattern differs among populations. Third, the gene-gene and gene-environment interactions may also influence the effect of risk allele between different populations. Genetic heterogeneity of AD may imply more and more risk genes for this disease, as exemplified by the most recent meta-analysis that identified 11 new susceptibility loci [43].

In summary, we verified the association between the *PICALM*, *CLU*, *MS4A4E*, *BINI* genes and LO-AD in Han Chinese populations, whereas the association of the remaining GWAS hit genes with AD was not validated in our populations. Particularly, meta-analysis revealed that the *PICALM*, *BINI*, and *CLU* genes were significantly associated with AD risk in Asian populations. One limitation of this study is that we lacked demographical data, such as age, sex, and education year of subjects for all individuals, which disabled us from retrieving more information in the association analyses. Further independent validating studies and essential functional assays are needed to solidify the current conclusions and to characterize the putative role of these genes concerning the production, transport, release, and clearance of A β from brain into blood and their participation in innate immunity in the central nervous system.

Acknowledgments We thank DNA donors in this study. We are grateful to the two anonymous reviewers for their helpful comments. This study was supported by the Strategic Priority Research Program (B) of the Chinese Academy of Sciences (XDB02020000).

Conflict of Interest The authors declare that they have no conflict of interest

References

- Dickson TC, Vickers JC (2001) The morphological phenotype of beta-amyloid plaques and associated neuritic changes in Alzheimer's disease. *Neuroscience* 105(1):99–107
- Tanzi RE (2012) The genetics of Alzheimer disease. *Cold Spring Harb Perspect Med* 2(10) pii: a006296. doi:10.1101/cshperspect.a006296.
- Ertekin-Taner N (2007) Genetics of Alzheimer's disease: a centennial review. *Neurol Clin* 25(3):611–667, v
- Zetsche T, Rujescu D, Hardy J, Hampel H (2010) Advances and perspectives from genetic research: development of biological markers in Alzheimer's disease. *Expert Rev Mol Diagn* 10(5):667–690
- Tanzi RE, Bertram L (2005) Twenty years of the Alzheimer's disease amyloid hypothesis: a genetic perspective. *Cell* 120(4):545–555
- Scheuner D, Eckman C, Jensen M, Song X, Citron M, Suzuki N, Bird TD, Hardy J, Hutton M, Kukull W, Larson E, Levy-Lahad E, Viitanen M, Peskind E, Poorkaj P, Schellenberg G, Tanzi R, Wasco W, Lannfelt L, Selkoe D, Younkin S (1996) Secreted amyloid beta-protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nat Med* 2(8):864–870
- Hollingworth P, Harold D, Sims R, Gerrish A, Lambert JC, Carrasquillo MM, Abraham R, Hamshere ML, Pahwa JS, Moskvin V, Dowzell K, Jones N, Stretton A, Thomas C, Richards A, Ivanov D, Widdowson C, Chapman J, Lovestone S, Powell J, Proitsis P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Brown KS, Passmore PA, Craig D, McGuinness B, Todd S, Holmes C, Mann D, Smith AD, Beaumont H, Warden D, Wilcock G, Love S, Kehoe PG, Hooper NM, Vardy ER, Hardy J, Mead S, Fox NC, Rossor M, Collinge J, Maier W, Jessen F, Ruther E, Schurmann B, Heun R, Kolsch H, van den Bussche H, Heuser I, Kornhuber J, Wiltfang J, Dichgans M, Frolich L, Hampel H, Gallacher J, Hull M, Rujescu D, Giegling I, Goate AM, Kauwe JS, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K, Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin A, Gwilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tsolaki M, Singleton AB, Guerreiro R, Muhleisen TW, Nothen MM, Moebus S, Jockel KH, Klopp N, Wichmann HE, Pankratz VS, Sando SB, Aasly JO, Barcikowska M, Wszolek ZK, Dickson DW, Graff-Radford NR, Petersen RC, van Duijn CM, Breteler MM, Ikram MA, DeStefano AL, Fitzpatrick AL, Lopez O, Launer LJ, Seshadri S, Berr C, Campion D, Epelbaum J, Dartigues JF, Tzourio C, Alperovitch A, Lathrop M, Feulner TM, Friedrich P, Riehle C, Krawczak M, Schreiber S, Mayhaus M, Nicolhaus S, Wagenpfeil S, Steinberg S, Stefansson H, Stefansson K, Snaedal J, Bjornsson S, Jonsson PV, Chouraki V, Genier-Boley B, Hiltunen M, Soininen H, Combarros O, Zelenika D, Delepine M, Bullido MJ, Pasquier F, Mateo I, Frank-Garcia A, Porcellini E, Hanon O, Coto E, Alvarez V, Bosco P, Siciliano G, Mancuso M, Panza F, Solfrizzi V, Nacmias B, Sorbi S, Bossu P, Piccardi P, Arosio B, Annoni G, Seripa D, Pilotto A, Scarpini E, Galimberti D, Brice A, Hannequin D, Licastro F, Jones L, Holmans PA, Jonsson T, Riemenschneider M, Morgan K, Younkin SG, Owen MJ, O'Donovan M, Amouyel P, Williams J (2011) Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. *Nat Genet* 43(5):429–435
- Naj AC, Jun G, Beecham GW, Wang LS, Vardarajan BN, Buros J, Gallins PJ, Buxbaum JD, Jarvik GP, Crane PK, Larson EB, Bird TD, Boeve BF, Graff-Radford NR, De Jager PL, Evans D, Schmechel JA, Carrasquillo MM, Ertekin-Taner N, Younkin SG, Cruchaga C, Kauwe JS, Nowotny P, Kramer P, Hardy J, Huentelman MJ, Myers AJ, Barmada MM, Demirci FY, Baldwin CT, Green RC, Rogava E, St George-Hyslop P, Arnold SE, Barber R, Beach T, Bigio EH, Bowen JD, Boxer A, Burke JR, Cairns NJ, Carlson CS, Carney RM, Carroll SL, Chui HC, Clark DG, Comeveaux J, Cotman CW, Cummings JL, DeCarli C, DeKosky ST, Diaz-Arrastia R, Dick M, Dickson DW, Ellis WG, Faber KM, Fallon KB, Farlow MR, Ferris S, Frosch MP, Galasko DR, Ganguli M, Gearing M, Geschwind DH, Ghetti B, Gilbert JR, Gilman S, Giordani B, Glass JD, Growdon JH, Hamilton RL, Harrell LE, Head E, Honig LS, Hulette CM, Hyman BT, Jicha GA, Jin LW, Johnson N, Karlawish J, Karydas A, Kaye JA, Kim R, Koo EH, Kowall NW, Lah JJ, Levey AI, Lieberman AP, Lopez OL, Mack WJ, Marson DC, Martiniuk F, Mash DC, Masliah E, McCormick WC, McCurry SM, McDavid AN, McKee AC, Mesulam M, Miller BL, Miller CA, Miller JW, Parisi JE, Perl DP, Peskind E, Petersen RC, Poon WW, Quinn JF, Rajbhandary RA, Raskind M, Reisberg B, Ringman JM, Roberson ED, Rosenberg RN, Sano M, Schneider LS, Seeley W, Shelanski ML, Slifer MA, Smith CD, Sonnen JA, Spina S, Stern RA, Tanzi RE, Trojanowski JQ, Troncoso JC, Van Deerlin VM, Vinters HV, Vonsattel JP, Weintraub S, Welsh-Bohmer KA, Williamson J, Woltjer RL, Cantwell LB, Dombroski BA, Beekly D, Lunetta KL, Martin ER, Kamboh MI, Saykin AJ, Reiman EM, Bennett DA, Morris JC, Montine TJ, Goate AM, Blacker D, Tsuang DW, Hakonarson H, Kukull WA, Foroud TM, Haines JL, Mayeux R, Pericak-Vance MA, Farrer LA, Schellenberg GD (2011) Common variants at MS4A4/MS4A6E, CD2AP, CD33 and EPHA1 are associated with late-onset Alzheimer's disease. *Nat Genet* 43(5): 436–441
- Lambert JC, Heath S, Even G, Campion D, Sleegers K, Hiltunen M, Combarros O, Zelenika D, Bullido MJ, Tavernier B, Letenneur L, Bettens K, Berr C, Pasquier F, Fievet N, Barberger-Gateau P, Engelborghs S, De Deyn P, Mateo I, Franck A, Helisalmi S, Porcellini E, Hanon O, de Pancorbo MM, Lendon C, Dufouil C, Jaillard C, Leveillard T, Alvarez V, Bosco P, Mancuso M, Panza F, Nacmias B, Bossu P, Piccardi P, Annoni G, Seripa D, Galimberti D, Hannequin D, Licastro F, Soininen H, Ritchie K, Blanche H,

- Dartigues JF, Tzourio C, Gut I, Van Broeckhoven C, Alperovitch A, Lathrop M, Amouyel P (2009) Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet* 41(10):1094–1099
10. Seshadri S, Fitzpatrick AL, Ikram MA, DeStefano AL, Gudnason V, Boada M, Bis JC, Smith AV, Carassquillo MM, Lambert JC, Harold D, Schrijvers EM, Ramirez-Lorca R, Debette S, Longstreth WT Jr, Janssens AC, Pankratz VS, Dartigues JF, Hollingworth P, Aspelund T, Hernandez I, Beiser A, Kuller LH, Koudstaal PJ, Dickson DW, Tzourio C, Abraham R, Antunez C, Du Y, Rotter JI, Aulchenko YS, Harris TB, Petersen RC, Berr C, Owen MJ, Lopez-Arrieta J, Varadarajan BN, Becker JT, Rivadeneira F, Nalls MA, Graff-Radford NR, Champion D, Auerbach S, Rice K, Hofman A, Jonsson PV, Schmidt H, Lathrop M, Mosley TH, Au R, Psaty BM, Uitterlinden AG, Farrer LA, Lumley T, Ruiz A, Williams J, Amouyel P, Younkin SG, Wolf PA, Launer LJ, Lopez OL, van Duijn CM, Breteler MM (2010) Genome-wide analysis of genetic loci associated with Alzheimer disease. *JAMA: J Am Med Assoc* 303(18):1832–1840
 11. Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, Pahwa JS, Moskvina V, Dowzell K, Williams A, Jones N, Thomas C, Stretton A, Morgan AR, Lovestone S, Powell J, Proitsi P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Morgan K, Brown KS, Passmore PA, Craig D, McGuinness B, Todd S, Holmes C, Mann D, Smith AD, Love S, Kehoe PG, Hardy J, Mead S, Fox N, Rossor M, Collinge J, Maier W, Jessen F, Schurmann B, van den Bussche H, Heuser I, Komhuber J, Wiltfang J, Dichgans M, Frolich L, Hampel H, Hull M, Rujescu D, Goate AM, Kauwe JS, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K, Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin A, Gwilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tsolaki M, Singleton AB, Guerreiro R, Muhleisen TW, Nothen MM, Moebus S, Jockel KH, Klopp N, Wichmann HE, Carrasquillo MM, Pankratz VS, Younkin SG, Holmans PA, O'Donovan M, Owen MJ, Williams J (2009) Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet* 41(10):1088–1093
 12. Jun G, Naj AC, Beecham GW, Wang LS, Buross J, Gallins PJ, Buxbaum JD, Ertekin-Taner N, Fallin MD, Friedland R, Inzelberg R, Kramer P, Rogava E, St George-Hyslop P, Alzheimer's Disease Genetics Consortium, Cantwell LB, Dombroski BA, Saykin AJ, Reiman EM, Bennett DA, Morris JC, Lunetta KL, Martin ER, Montine TJ, Goate AM, Blacker D, Tsuang DW, Beekly D, Cupples LA, Hakonarson H, Kukull W, Foroud TM, Haines J, Mayeux R, Farrer LA, Pericak-Vance MA, Schellenberg GD (2010) Meta-analysis confirms CR1, CLU, and PICALM as Alzheimer disease risk loci and reveals interactions with APOE genotypes. *Arch Neurol* 67(12):1473–1484
 13. Hu X, Pickering E, Liu YC, Hall S, Fournier H, Katz E, Dechairo B, John S, Van Eerdewegh P, Soares H, Alzheimer's Disease Neuroimaging Initiative (2011) Meta-analysis for genome-wide association study identifies multiple variants at the BIN1 locus associated with late-onset Alzheimer's disease. *PLoS One* 6(2):e16616
 14. Kamboh MI, Demirci FY, Wang X, Minster RL, Carrasquillo MM, Pankratz VS, Younkin SG, Saykin AJ, Jun G, Baldwin C, Logue MW, Buross J, Farrer L, Pericak-Vance MA, Haines JL, Sweet RA, Ganguli M, Feingold E, Dekosky ST, Lopez OL, Bamada MM (2012) Genome-wide association study of Alzheimer's disease. *Transl Psychiatry* 2:e117
 15. Li HL, Shi SS, Guo QH, Ni W, Dong Y, Liu Y, Sun YM, Bei W, Lu SJ, Hong Z, Wu ZY (2011) PICALM and CR1 variants are not associated with sporadic Alzheimer's disease in Chinese patients. *J Alzheim Dis: JAD* 25(1):111–117
 16. Ferrari R, Moreno JH, Minhajuddin AT, O'Bryant SE, Reisch JS, Barber RC (1846) Momeni P (2012) Implication of common and disease specific variants in CLU, CR1, and PICALM. *Neurobiol Aging* 33(8):1846e7–1846e18
 17. Tan L, Yu JT, Zhang W, Wu ZC, Zhang Q, Liu QY, Wang W, Wang HF, Ma XY, Cui WZ (2013) Association of GWAS-linked loci with late-onset Alzheimer's disease in a northern Han Chinese population. *Alzheim Dement: J Alzheim Assoc* 9(5):546–553
 18. Bi R, Zhao L, Zhang C, Lu W, Feng JQ, Wang Y, Ni J, Zhang J, Li GD, Hu QX, Wang D, Yao YG, Li T (2014) No association of the LRRK2 genetic variants with Alzheimer's disease in Han Chinese individuals. *Neurobiol Aging* 35(2):444e5–444e9
 19. McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlan EM (1984) Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology* 34(7):939–944
 20. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, Roses AD, Haines JL, Pericak-Vance MA (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261(5123):921–923
 21. Kurz A, Lautenschlager N, Haupt M, Zimmer R, von Thülen B, Altland K, Lauter H, Müller U (1994) The apolipoprotein E-epsilon 4 allele is a risk factor for Alzheimer disease with early and late onset. *Nervenarzt* 65(11):774–779
 22. Wang D, Feng JQ, Li YY, Zhang DF, Li XA, Li QW, Yao YG (2012) Genetic variants of the MRC1 gene and the IFNG gene are associated with leprosy in Han Chinese from Southwest China. *Hum Genet* 131(7):1251–1260
 23. Gauderman WJ (2002) Sample size requirements for matched case-control studies of gene-environment interaction. *Stat Med* 21(1):35–50
 24. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, Maller J, Sklar P, de Bakker PI, Daly MJ, Sham PC (2007) PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 81(3):559–575
 25. Barrett JC, Fry B, Maller J, Daly MJ (2005) Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 21(2):263–265
 26. Stephens M, Smith NJ, Donnelly P (2001) A new statistical method for haplotype reconstruction from population data. *Am J Hum Genet* 68(4):978–989
 27. Skol AD, Scott LJ, Abecasis GR, Boehnke M (2006) Joint analysis is more efficient than replication-based analysis for two-stage genome-wide association studies. *Nat Genet* 38(2):209–213
 28. Consortium GT (2013) The Genotype-Tissue Expression (GTEx) project. *Nat Genet* 45(6):580–585
 29. Chen LH, Kao PY, Fan YH, Ho DT, Chan CS, Yik PY, Ha JC, Chu LW, Song YQ (2012) Polymorphisms of CR1, CLU and PICALM confer susceptibility of Alzheimer's disease in a southern Chinese population. *Neurobiol Aging* 33(1):210e1–210e7
 30. Yu JT, Song JH, Ma T, Zhang W, Yu NN, Xuan SY, Tan L (2011) Genetic association of PICALM polymorphisms with Alzheimer's disease in Han Chinese. *J Neurol Sci* 300(1–2):78–80
 31. Ohara T, Ninomiya T, Hirakawa Y, Ashikawa K, Monji A, Kiyohara Y, Kanba S, Kubo M (2012) Association study of susceptibility genes for late-onset Alzheimer's disease in the Japanese population. *Psychiatr Genet* 22(6):290–293
 32. Tebar F, Bohlander SK, Sorkin A (1999) Clathrin assembly lymphoid myeloid leukemia (CALM) protein: localization in endocytic-coated pits, interactions with clathrin, and the impact of overexpression on clathrin-mediated traffic. *Mol Biol Cell* 10(8):2687–2702
 33. Koo EH, Squazzo SL (1994) Evidence that production and release of amyloid beta-protein involves the endocytic pathway. *J Biol Chem* 269(26):17386–17389
 34. Teixeira CM, Pomedli SR, Maei HR, Kee N, Frankland PW (2006) Involvement of the anterior cingulate cortex in the expression of

- remote spatial memory. *J Neurosci: Off J Soc Neurosci* 26(29):7555–7564
35. Jiang T, Yu JT, Tan MS, Wang HF, Wang YL, Zhu XC, Zhang W, Tan L (2014) Genetic variation in PICALM and Alzheimer's disease risk in Han Chinese. *Neurobiol Aging* 35(4):934e1–934e3
 36. Liu G, Zhang S, Cai Z, Ma G, Zhang L, Jiang Y, Feng R, Liao M, Chen Z, Zhao B, Li K (2013) PICALM gene rs3851179 polymorphism contributes to Alzheimer's disease in an Asian population. *Neuromol Med* 15(2):384–388
 37. Liu G, Zhang S, Cai Z, Li Y, Cui L, Ma G, Jiang Y, Zhang L, Feng R, Liao M, Chen Z, Zhao B, Li K (2013) BIN1 gene rs744373 polymorphism contributes to Alzheimer's disease in East Asian population. *Neurosci Lett* 544:47–51
 38. Chapuis J, Hansmannel F, Gistelink M, Mounier A, Van Cauwenberghhe C, Kolen KV, Geller F, Sottejeau Y, Harold D, Dourlen P, Grenier-Boley B, Kamatani Y, Delepine B, Demiautte F, Zelenika D, Zommer N, Hamdane M, Bellenguez C, Dartigues JF, Hauw JJ, Letronne F, Ayril AM, Sleegers K, Schellens A, Broeck LV, Engelborghs S, De Deyn PP, Vandenberghe R, O'Donovan M, Owen M, Epelbaum J, Mercken M, Karran E, Bantscheff M, Drewes G, Joberty G, Campion D, Octave JN, Berr C, Lathrop M, Callaerts P, Mann D, Williams J, Buee L, Dewachter I, Van Broeckhoven C, Amouyel P, Moechars D, Dermaut B, Lambert JC, GERAD Consortium (2013) Increased expression of BIN1 mediates Alzheimer genetic risk by modulating tau pathology. *Mol Psychiatry* 18(11):1225–1234
 39. Karch CM, Jeng AT, Nowotny P, Cady J, Cruchaga C, Goate AM (2012) Expression of novel Alzheimer's disease risk genes in control and Alzheimer's disease brains. *PLoS One* 7(11):e50976
 40. Carrasquillo MM, Belbin O, Hunter TA, Ma L, Bisceglia GD, Zou F, Crook JE, Pankratz VS, Sando SB, Aasly JO, Barcikowska M, Wszolek ZK, Dickson DW, Graff-Radford NR, Petersen RC, Passmore P, Morgan K, Alzheimer's Research UKc, Younkin SG (2011) Replication of EPHA1 and CD33 associations with late-onset Alzheimer's disease: a multi-centre case-control study. *Mol Neurodegener* 6(1):54
 41. Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorf LA, Hunter DJ, McCarthy MI, Ramos EM, Cardon LR, Chakravarti A, Cho JH, Guttmacher AE, Kong A, Kruglyak L, Mardis E, Rotimi CN, Slatkin M, Valle D, Whittemore AS, Boehnke M, Clark AG, Eichler EE, Gibson G, Haines JL, Mackay TF, McCarroll SA, Visscher PM (2009) Finding the missing heritability of complex diseases. *Nature* 461(7265):747–753
 42. Ma L, Tang J, Wang D, Zhang W, Liu W, Wang D, Liu XH, Gong W, Yao YG, Chen X (2013) Evaluating risk loci for schizophrenia distilled from genome-wide association studies in Han Chinese from Central China. *Mol Psychiatry* 18(6):638–639
 43. Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, Jun G, Destefano AL, Bis JC, Beecham GW, Grenier-Boley B, Russo G, Thornton-Wells TA, Jones N, Smith AV, Chouraki V, Thomas C, Ikram MA, Zelenika D, Vardarajan BN, Kamatani Y, Lin CF, Gerrish A, Schmidt H, Kunkle B, Dunstan ML, Ruiz A, Bihoreau MT, Choi SH, Reitz C, Pasquier F, Hollingworth P, Ramirez A, Hanon O, Fitzpatrick AL, Buxbaum JD, Campion D, Crane PK, Baldwin C, Becker T, Gudnason V, Cruchaga C, Craig D, Amin N, Berr C, Lopez OL, De Jager PL, Deramecourt V, Johnston JA, Evans D, Lovestone S, Letenneur L, Moron FJ, Rubinsztein DC, Eiriksdottir G, Sleegers K, Goate AM, Fievet N, Huentelman MJ, Gill M, Brown K, Kamboh MI, Keller L, Barberger-Gateau P, McGuinness B, Larson EB, Green R, Myers AJ, Dufouil C, Todd S, Wallon D, Love S, Rogava E, Gallacher J, St George-Hyslop P, Clarimon J, Lleo A, Bayer A, Tsuang DW, Yu L, Tsolaki M, Bossu P, Spalletta G, Proitsi P, Collinge J, Sorbi S, Sanchez-Garcia F, Fox NC, Hardy J, Naranjo MC, Bosco P, Clarke R, Brayne C, Galimberti D, Mancuso M, Matthews F, European Alzheimer's Disease Initiative, Genetic, Environmental Risk in Alzheimer's Disease, Alzheimer's Disease Genetic Consortium, Cohorts for Heart and Aging Research in Genomic Epidemiology, Moebus S, Mecocci P, Del Zompo M, Maier W, Hampel H, Pilotto A, Bullido M, Panza F, Caffarra P, Nacmias B, Gilbert JR, Mayhaus M, Lannfelt L, Hakonarson H, Pichler S, Carrasquillo MM, Ingelsson M, Beekly D, Alvarez V, Zou F, Valladares O, Younkin SG, Coto E, Hamilton-Nelson KL, Gu W, Razquin C, Pastor P, Mateo I, Owen MJ, Faber KM, Jonsson PV, Combarros O, O'Donovan MC, Cantwell LB, Soininen H, Blacker D, Mead S, Mosley TH Jr, Bennett DA, Harris TB, Fratiglioni L, Holmes C, de Bruijn RF, Passmore P, Montine TJ, Bettens K, Rotter JI, Brice A, Morgan K, Foroud TM, Kukull WA, Hannequin D, Powell JF, Nalls MA, Ritchie K, Lunetta KL, Kauwe JS, Boerwinkle E, Riemenschneider M, Boada M, Hiltunen M, Martin ER, Schmidt R, Rujescu D, Wang LS, Dartigues JF, Mayeux R, Tzourio C, Hofman A, Nothen MM, Graff C, Psaty BM, Jones L, Haines JL, Holmans PA, Lathrop M, Pericak-Vance MA, Launer LJ, Farrer LA, van Duijn CM, Van Broeckhoven C, Moskvina V, Seshadri S, Williams J, Schellenberg GD, Amouyel P (2013) Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet* 45(12):1452–1458