

Ionic liquid modified carbon paste electrode and investigation of its electrocatalytic activity to hydrogen peroxide

ERHAN CANBAY, HAYATI TÜRKMEN[†] and EROL AKYILMAZ*

Department of Biochemistry, Faculty of Science, Ege University 35100 Bornova-Izmir/Turkey

[†]Department of Chemistry, Faculty of Science, Ege University 35100 Bornova-Izmir/Turkey

MS received 24 December 2012; revised 25 April 2013

Abstract. This paper reports on the preparation and advantages of novel amperometric biosensors in the presence of hydrophobic ionic liquid (IL), 1-methyl-3-butylimidazolium bromide ([MBIB]). Carbon paste biosensor has been constructed by entrapping horseradish peroxidase in graphite and IL mixed with paraffin oil as a binder. The resulting IL/graphite material brings new capabilities for electrochemical devices by combining the advantages of ILs composite electrodes. Amounts of H₂O₂ were amperometrically detected by monitoring current values at reduction potential (−0.15 V) of K₃Fe(CN)₆. Decrease in biosensor responses were linearly related to H₂O₂ concentrations between 10 and 100 μM with 2 s response time. Limit of detection of the biosensor were calculated to be 3.98 μM for H₂O₂. In the optimization studies of the biosensor some parameters such as optimum pH, optimum temperature, enzyme amount, interference effects of some substances on the biosensor response, reproducibility and storage stability were carried out. The promising results are ascribed to the use of an ionic liquid, which forms an excellent charge-transfer bridge and wide electrochemical windows in the bulk of carbon paste electrode.

Keywords. Biosensor; ionic liquid; 1-methyl-3-butylimidazolium bromide; carbon paste electrode; horseradish peroxidase.

1. Introduction

Recently, electrochemical study of the third generation biosensor based on the direct electron transfer between the protein and the electrode has been widely investigated at various electrodes. The main aim is to elucidate the complex mechanisms of biological electron transfer and determine their potential application in biotechnology (Xu *et al* 2006; Gao *et al* 2007). Until now, horseradish peroxidase (HRP) is often used as the molecules for the study of electron transfer reactions because of their commercial availability, moderate cost, and known and documented structure (Chen *et al* 2000; Xiao *et al* 2000). However, direct adsorption of such proteins onto the electrode surface can frequently result in their denaturation and the loss of bioactivity (Liu *et al* 2006). Therefore, it is necessary to develop new immobilization methods to realize direct electrochemistry without loss of the bioactivity of protein.

Carbon paste electrode (CPE), which was made up of carbon particles and organic liquid, has been widely applied in the electroanalytical community due to its low cost, ease of fabrication, high sensitivity for detection and

renewable surface (Wang and Lu 1998; Kulys 1999). However, carbon paste electrodes (CPEs) also exhibit several disadvantages such as relative weaker fabrication reproducibility and mechanical stability comparing with the bare solid electrodes (Svorc *et al* 1997; Mailley *et al* 2003) which greatly limit the practical utility of enzymatic assays and probes. Moreover, the binder is non-conducting, which to some extent weakens the electrochemical response of the sensor.

Ionic liquid modified carbon paste electrode (IL–CPE) is a new kind of working electrode prepared by using ionic liquid (IL) as a binder and a modifier. Due to the specific properties of IL such as high chemical and thermal stabilities, negligible vapour pressure, high ionic conductivity, wide electrochemical windows, low toxicity and the ability to dissolve a wide range of organic and inorganic compounds (Liu *et al* 2005a, b; Pandey 2006), IL–CPEs were demonstrated to exhibit advantages including wide electrochemical windows, inherent electrocatalytic ability and antifouling ability (Wei and Ivaska 2008). Because of their high ionic conductivity and wide electrochemical windows, ILs have been used in electrochemistry and electroanalysis. Compton *et al* (2004) and Endres (2004) have reviewed the recent progress of RTILs in electrochemistry. Recently, Wang *et al* (2007) have shown that it is possible to fabricate an IL-carbon paste (CP) biosensor with low background current via

*Author for correspondence (erol.akyilmaz@ege.edu.tr)

mixing the paste with paraffin oil 'a non-conductive binder' which tends to decrease the background current and hence improve the sensor response. Liu and co-workers (2005a, b) also fabricated an imidazolium-based ionic liquid modified carbon paste electrode, and the presence of ionic liquids caused an increase in the sensitivity of the response toward to nitrite detection (Liu *et al* 2005a, b). Safavi *et al* (2007) investigated the electrochemical oxidation of phenolic compounds on an IL-CPE, which exhibited higher stability than commonly used working electrodes. IL-CPE can also be used as the substrate electrode for further modification with redox proteins or nanoparticles, which exhibited many new characteristics and potential applications (Sun *et al* 2009).

In this paper, an IL, 1-methyl-3-butylimidazolium bromide ([MBIB]), was used as binder for manufacturing the modified carbon paste electrode. IL-CPE electrode was prepared with mixing graphite, paraffin oil, ionic liquid and horseradish peroxidase.

2. Experimental

2.1 Apparatus

In the experiments PalmSens potentiostat (Netherlands), a three-electrodes system from CH Instruments (USA) that contains, a CHI 111 model Ag/AgCl reference electrode and a CHI 115 model platinum wire counter electrode, Gilson P100 and P1000 automatic pipettes (France), Yellow-Line magnetic stirrer (Germany) and Nuve model thermostat (TR) were used.

2.2 Chemicals and reagents

HRP (EC 1.11.1.7), paraffin oil, potassium ferricyanide, hydrogen peroxide and graphite powder and all other chemicals were purchased from Sigma Chemical. The ionic liquid 1-methyl-3-butylimidazolium bromide ([MBIB]) was kindly prepared by the Chemistry Research Laboratory at Ege University according to earlier study (Tasci *et al* 2012). All the chemicals were of high-purity grade and used as purchased without further purification.

2.3 Electrode fabrication

The required amount of the ionic liquid or paraffin oil was mixed using pestle and mortar with the required amount of graphite. This ionic liquid modified carbon paste was tightly packed into a cavity (3.3 mm diameter) of a glass tube and the electrical contact was established via copper wire. Then 2 mg of horseradish peroxidase was added and the paste was mixed for 10 min. Prior to use, the surface of the carbon paste electrode was polished with weighing paper.

2.4 Procedure

Measurements were carried out with potassium ferricyanide in a phosphate buffer (0.05 M, pH 7) containing 5 mM $\text{Fe}(\text{CN})_6^{3-/4-}$ supporting electrolyte medium for amperometric and cyclic voltammetry measurements. Amperometric detection was made under the potential of -0.15 V, whereas cyclic voltammetry experiments was made between the potential of -0.6 and 0.6 V. In the former, the desired working potential was applied and transient currents were allowed to decay to a steady-state value.

3. Results and discussion

3.1 Effect of ionic liquid on modified electrode

In order to detect the effect of ionic liquid on the modified electrode some cyclic voltammetric experiments were done by using the modified CPE electrodes with ionic liquid and without ionic liquid in the presence of $\text{Fe}(\text{CN})_6^{3-/4-}$ redox couple. A pair of redox peaks were observed for $\text{Fe}(\text{CN})_6^{3-/4-}$ on a CPE (figure 1(A), curve a and figure 1(B), curve a). Oxidation peak potential (E_{pa}) is about $+0.305$ V, reduction peak potential (E_{pc}) is about -0.103 V and ΔE_p is 0.408 V. On the other hand, a pair of redox peaks at different potentials were observed by using CPE with ionic liquid (figure 1(A), curves b, c, d and e). Oxidation peak potential (E_{pa}) is about 0.146 V and reduction peak potential (E_{pc}) is about -0.05 V and ΔE_p is 0.196 V for (c). Also, the peak current (I_p) at IL-CPE were much larger than about 15.57 times of that at CPE. It can be observed that the background current increased with an increase in the ionic liquid content. As previously reported in the literature (Maleki *et al* 2006), the addition of ILs to a carbon paste electrode modifies the microstructure of the paste and the charge transfer resistance decreases and charge transfer rate increases, because of the higher conductivity of the electrode containing the ionic liquid. An increase in the ionic liquid content resulted in increasing peak-to-peak separation. The most useful electrochemical results could be obtained by using IL-CPE prepared by 10/20/70 ratio. In addition the best and smooth electrode surface was obtained by using the same ratio. So, the best ratio of ionic liquid was chosen to be 10%.

Figure 2(A) shows CV responses of IL-CPE in 5 mM $\text{Fe}(\text{CN})_6^{3-/4-}$ with scan rates from 10 to 100 mV s^{-1} . There was a good linear relationship between peak current and the square root of scan rates, which showed a typical diffusion-controlled electrochemical behaviour.

3.2 Effect of enzyme amount on biosensor responses

In order to optimize the biosensor, several experimental parameters were investigated for the use with cyclic

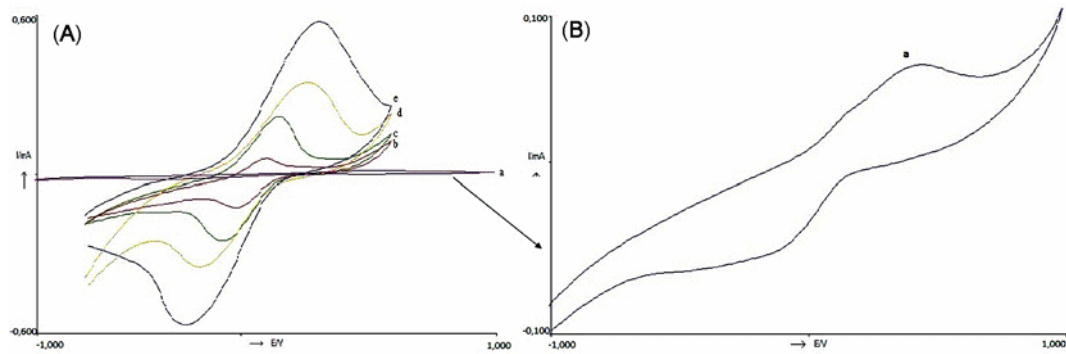


Figure 1. CVs obtained from different ratios of IL/parafin oil/graphite: (a) 0/30/70, (b) 5/25/70, (c) 10/20/70, (d) 15/15/70, (e) 20/10/70 for (A); (a) 0/30/70 for (B), respectively. Conditions: 0.05 M phosphate buffer (pH 7) containing 5 mM potassium ferricyanide, scan rate 50 mV/s and potentials were referred to Ag/AgCl reference electrode. Amount of HRP were kept constant to be 2 mg, respectively.

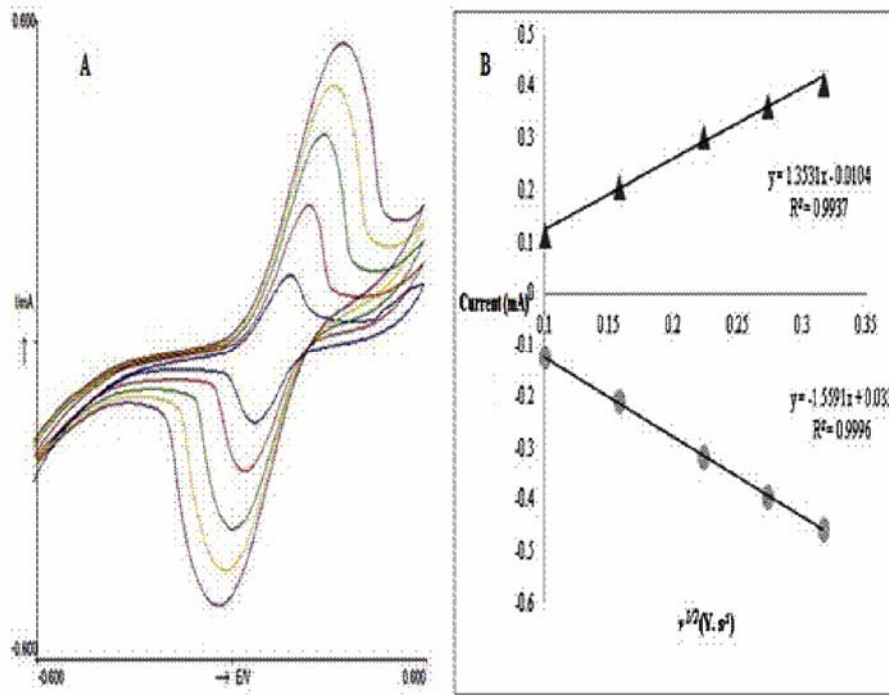


Figure 2. (A) CVs of IL-CPE at different scan rates (from inside to outside): 0.01, 0.025, 0.05, 0.075, 0.1 V s⁻¹, respectively. (B) Plot of cathodic and anodic peak currents vs scan rate ($v^{1/2}$). Conditions: 0.05 M phosphate buffer (pH 7) containing 5 mM potassium ferricyanide and potentials were referred to Ag/AgCl reference electrode. Amount of HRP and ratio of IL/parafin oil/graphite were kept constant to be 2 mg and 10/20/70, respectively.

voltammetry. Initially, to detect the effect of enzyme amount, 1, 2 and 3 mg (113 unit mg⁻¹) of peroxidase were used in electrodes with a composition of 70:20:10 (w/w/w) graphite powder:parafin oil:ionic liquid. From the results, higher biosensor responses and more acceptable calibration curves were achieved when biosensor was used, that contained 2 mg horseradish peroxidase. When enzyme amount was used to be 1 mg, biosensor responses were decreased and when enzyme amount was used to be 3 mg, biosensor responses were increased but

calibration curves were decreased because of diffusion barrier occurrence.

3.3 Amperometric response of developed H₂O₂ biosensor

Amperometric response of IL-CPE to H₂O₂ was investigated at different applied potentials between -0.3 and 0.5 V. The most suitable and highest responses were obtained at -0.15 V. Figure 3(A) shows amperometric

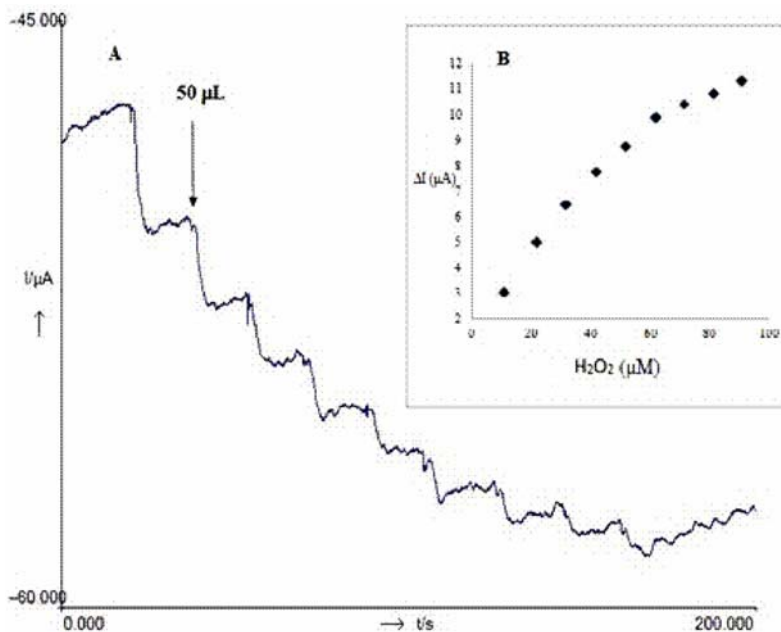


Figure 3. Typical amperometric response of fabricated biosensor to successive addition of H₂O₂ in a stirred 0.05 M PBS (pH 7) containing 5 mM potassium ferricyanide. Applied potential was -0.15 V vs Ag/AgCl. (A) Amperometric response of 50 μ L successive addition of 1 mM H₂O₂ and (B) calibration curve between current and concentration of H₂O₂. Amount of HRP and ratio of IL/parafin oil/graphite were kept constant to be 2 mg and 10/20/70, respectively.

responses of IL-CPE to successive addition of H₂O₂ in 0.05 M PBS (pH 7) containing 5 mM Fe(CN)₆^{3-/4-}. The biosensor responded rapidly when H₂O₂ was added to the stirring PBS and steady-state current could be obtained within 2 s. The inset (figure 3B) displayed calibration curve of the amperometric response of the biosensor to the concentration of H₂O₂. The linear range of the developed biosensor for the determination of H₂O₂ was found to be 1×10^{-5} – 1×10^{-4} M and a correlation coefficient of 0.981 ($n = 9$).

3.4 Optimization of experimental conditions

The effect of the pH, from 6 to 8 on the biosensor response in 0.05 M PBS (pH 7) containing 5 mM Fe(CN)₆^{3-/4-} and 5×10^{-5} M H₂O₂ solution was also investigated. The highest current was obtained at pH 7, in agreement with the results reported for other studies using horseradish peroxidase (Lei *et al* 2004; Garcia *et al* 2007). In addition, the effect of the temperature from 20 to 40 °C was investigated. The best results was obtained at 30 °C. Therefore, a pH value of 7 and temperature 30 °C were used in further experiments.

3.5 Linear range of biosensor

For the determination of a linear range for H₂O₂, cyclic voltammetry and also amperometric methods were used.

According to the results obtained from both methods, responses of the biosensor depend linearly on H₂O₂ concentration between 10 and 100 μ M. The cyclic voltammograms indicated that when concentrations of H₂O₂ were increased, anodic peak current of the biosensor increased correlating at the oxidation potential of K₃Fe(CN)₆ whereas cathodic peak current of the biosensor decreased reduction potential of K₃Fe(CN)₆. From the cyclic voltammograms, it can be said that K₃Fe(CN)₆ is an effective mediator in the regeneration of the enzyme.

Figure 4 shows amperometric responses of separate additions of H₂O₂ at -0.15 V potential. From the figure, it can be said that addition of H₂O₂ into reaction cell decreased linearly with H₂O₂ concentration. Detection limits of the biosensor were found to be 3.98 μ M for amperometric measurements. This result also supported the amperometric results showed in figure 3.

3.6 Reproducibility, stability and interference effects of some substances

The concentration of 5×10^{-4} M H₂O₂ was measured consecutively for 7 times and average value, standard deviation (SD) and coefficients of variation (CV%) were calculated to be 5.4×10^{-4} M, 0.031, 4.2%, respectively. So, it can be said that biosensor showed a good reproducibility for the determination of H₂O₂.

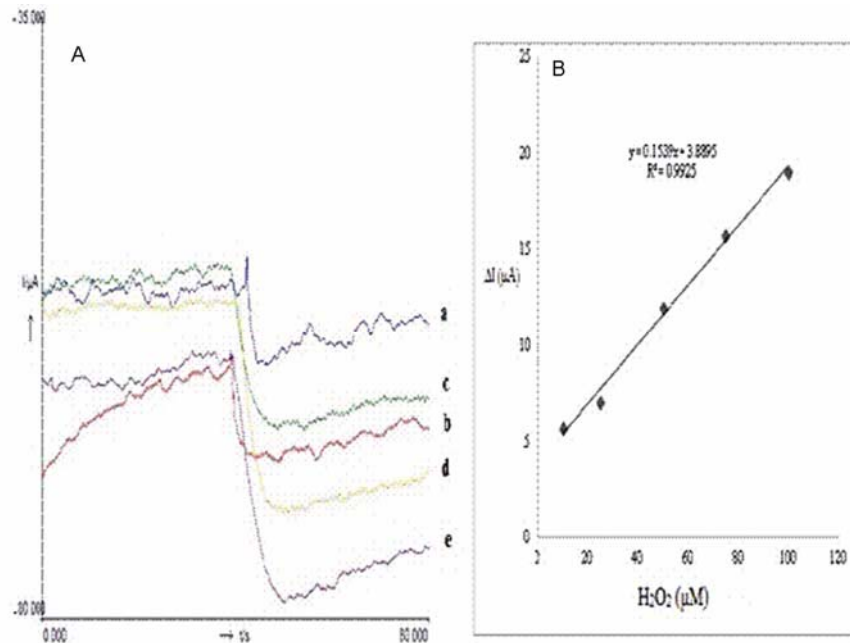


Figure 4. Typical amperometric responses of biosensor in a stirred 0.05 M PBS (pH 7) containing 5 mM potassium ferricyanide. Applied potential was -0.15 V vs Ag/AgCl. (A) Amperometric response of $10 \mu\text{M}$ (a), $25 \mu\text{M}$ (b), $50 \mu\text{M}$ (c), $75 \mu\text{M}$ (d), $100 \mu\text{M}$ H₂O₂ (e) and (B) calibration curve between the current and concentration of H₂O₂. Amount of HRP and ratio of IL/paraffin oil/graphite were kept constant to be 2 mg and 10/20/70, respectively.

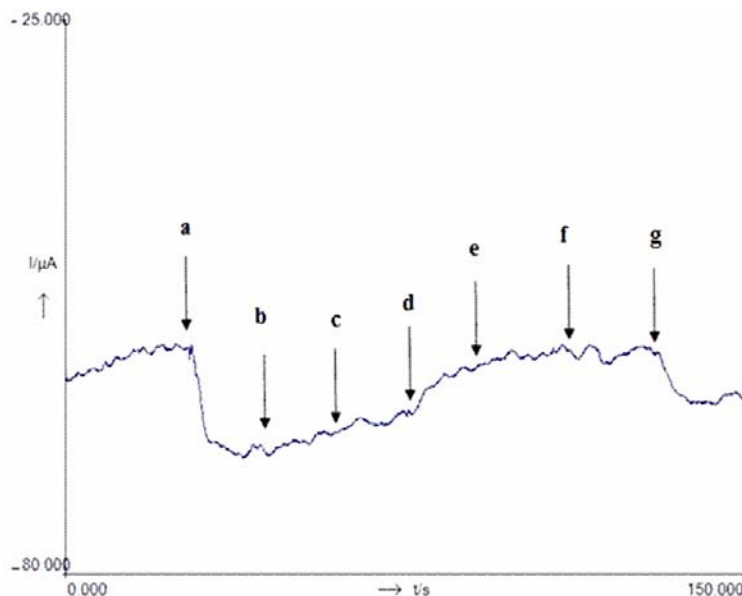


Figure 5. Substrate selectivity of IL-CPE hydrogen peroxide (a), D-glucose (b), D-fructose (c), L-ascorbic acid (d), L-glutamic acid (e), L-aspartic acid (f) and hydrogen peroxide (g) in 0.05 M PBS (pH 7) containing 5 mM potassium ferricyanide. Concentration of all substances is $50 \mu\text{M}$. Amount of HRP and ratio of IL/paraffin oil/graphite were kept constant to be 2 mg and 10/20/70, respectively.

The storage stability of the biosensor has been studied over a period of 2 months. The biosensor was stored at 4°C when not in use. The biosensor lost only 7% of the

initial response after 2 weeks and maintained more than 80% of the initial value after storage for one month. It lost 50% of the initial response after two months.

Table 1. Comparison of H₂O₂ biosensor based on HRP.

Electrode type	Linear range	Response time (s)	Detection limit (μM)	Reference
GCE/FBCS	35–2000 μM	20	15.0	Garcia <i>et al</i> (2007)
GCE/P(GMA-co-VFc)	2–30 mM	4	2.60	Senel <i>et al</i> (2010)
Au/SA–HRP	7–4100 μM	–	1.80	Liu <i>et al</i> (2009)
GCE/DDAB–HRP	1–4.0 mM	5	–	Tang <i>et al</i> (2003)
CCE/nano Au–HRP	12.2–1100 μM	<8	6.10	Lei <i>et al</i> (2004)
IL–CPE–HRP	10–100 μM	2	3.98	This work

GCE, glassy carbon electrode; P(GMA-co-VFc)poly(glycidyl methacrylate-co-vinylferrocene); FBCS, ferrocene branched chitosan; PEGDGE: polyethylene glycol diglycidyl ether; SA, sodium alginate; CCE, carbon ceramic electrode; HRP, horseradish peroxidase; DDAB, didodecyltrimethylammonium bromide.

Anti-interference ability of the biosensor was also studied. For this purpose 5 μM concentration of D-glucose, D-fructose, L-ascorbic acid, L-glutamic acid, L-aspartic acid (figure 5). We can see only the addition of ascorbic acid increased current 20% in comparison with H₂O₂. It was expected because of its universal interference capacity.

From table 1, it can be told that our proposed biosensor shows great performance in terms of response time and detection limit and it shows acceptable linear range for H₂O₂.

4. Conclusions

In our work, we have illustrated the ionic liquid (1-methyl-3-butylimidazolium bromide ([MBIB]) which can be used to prepare attractive composite electrodes for amperometric detection of hydrogen peroxide based on peroxidase enzyme. Comparing with CPE to IL–CPE, when paraffin oil only used as the binder decreases the high background current. However, the addition of the ionic liquids has several advantages such as a higher sensitivity, a wider linear dynamic range, high conductivity and higher stability (pH, thermal and storage). Peroxidase can contain a heme cofactor in its active site, or alternate redox-active cysteine or selenocysteine residues. The nature of the electron donor is dependent on the structure of the enzyme. Horseradish peroxidase can use a variety of organic compounds to be electron donors and acceptors. It has an accessible active site and many compounds can reach the site of the reaction. From the experimental results, it is obvious that the ionic liquid used in the biosensor preparation serves to be an attractive compound for the enzymatic reaction of peroxidase enzyme.

References

Compton R G, Buzzeo M C and Evans R G 2004 *Chem. Phys. Chem.* **5** 1106

- Chen X, Ruan C, Kong J and Deng J 2000 *Anal. Chim. Acta* **412** 89
- Endres F 2004 *Z. Phys. Chem.* **218** 255
- Gao F X, Yuan R, Chai Y Q, Tang M Y, Cao S R and Chen S H 2007 *Coll. Surf. A: Physicochem. Eng. Aspects* **295** 223
- Garcia A, Peniche-Covas C, Chico B, Simpson B K and Villalonga R 2007 *Macromol. Biosci.* **7** 435
- Kulys J 1999 *Biosens. Bioelectron.* **14** 473
- Lei C, Hu S, Gao N, Shen G and Yu R 2004 *Bioelectrochemistry* **65** 33
- Liu C, Guo X and Yuan R 2009 *J. Mol. Catal.* **B60** 151
- Liu H T, He P, Li Z Y, Sun C Y, Shi L H, Liu Y, Zhu G Y and Li J H 2005a *Electrochem. Commun.* **7** 1357
- Liu J F, Jiang G B and Jönsson J Å 2005b *Trends Anal. Chem.* **24** 20
- Liu Y, Yuan R and Chai Y Q 2006 *Sens. Actuators* **B115** 109
- Maleki N, Safavi A and Tajabadi F 2006 *Anal. Chem.* **78** 3820
- Mailley P, Cummings E A, Mailley S C, Eggins B R, McAdams E and Cosnier S 2003 *Anal. Chem.* **75** 5422
- Pandey S 2006 *Anal. Chim. Acta* **556** 38
- Safavi A, Maleki N and Tajabadi F 2007 *Analyst* **132** 54
- Senel M, Cevik E and Abasiyanik F 2010 *Sens. Actuators* **B145** 445
- Sun W, Li X Q, Qin P and Jiao K 2009 *J. Phys. Chem.* **C113** 11294
- Svorc J, Miertu S, Katrlík J and Střed'ansk M 1997 *Anal. Chem.* **69** 2086
- Tang J, Wang B, Wu Z, Han X, Dang S and Wang E 2003 *Biosens. Bioelectron.* **18** 867
- Tasci Z, Kunduracioglu A, Kani I and Çetinkaya B 2012 *Chem. Cat. Chem.* **4** 831
- Wang J and Lu F 1998 *J. Am. Chem. Soc.* **120** 1049
- Wang S, Chen T, Zhang Z and Pang, D 2007 *Electrochem. Commun.* **9** 1337
- Wei D and Ivaska A 2008 *Anal. Chim. Acta* **607** 126
- Xiao Y, Ju H X and Chen H Y 2000 *Anal. Biochem.* **278** 22
- Xu Y X, Wang F, Chen X X and Hu S S 2006 *Talanta* **70** 651