#### **REVIEW ARTICLE**



# Small cell carcinoma of the bladder: the characteristics of molecular alterations, treatment, and follow-up

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## Abstract

Small cell carcinoma of the bladder (SCCB) is a rare disease associated with high invasiveness and mortality. Histologically, SCCB is difficult to distinguish from small cell lung cancer (SCLC); however, it shares more similar molecular alterations with urothelial carcinoma (UC). As a result, now, the widely accepted theory about the cells of origin is that SCCB and UC probably have a common clone origin. Even the former probably comes from a preexisting UC. At present, given its rarity, early diagnoses, treatments, and follow-ups are not well established, which are vital to patients with SCCB. Inspirationally, in recent years, with the development of molecular diagnostic methods, molecular alterations of SCCB have been understood partially, which are propitious to excavate new potential therapeutic strategies and establish sound follow-ups. Therefore, the future will be light for patients with SCCB.

**Keywords** Small cell carcinoma of the bladder  $\cdot$  Urothelial carcinoma  $\cdot$  Small cell lung cancer  $\cdot$  Molecular alterations  $\cdot$  Treatment  $\cdot$  Follow-up

# Introduction

Bladder cancer is the 10th most common form of malignancy worldwide. In 2018, 549,000 new cases were diagnosed and 200,000 cases died of bladder cancer [1]. Neuroendocrine cancer of bladder (NECB) represents approximately less than 1% of all urinary bladder malignancies. SCCB is the major subtype of NECB and often mixes with urothelial

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components [2, 3]. Similar to UC, SCCB is more common in men (ratio of male to female = 3:1) and usually appears in the seventh to eighth decades of life. Painless gross hematuria and irritative voiding symptoms are most common initial signs [4, 5]. However, SCCB is usually advanced when it is diagnosed, with poorer prognosis compared to UC in the same stage [2].

Histologically, SCCB is similar to SCLC, but clinical manifestations are consistent with UC. Therefore, researchers devote to seeking for features of molecular alterations in SCCB.

Some studies found that SCCB had a high somatic mutational rate, which was mainly driven by APOBEC-mediated mutational process (accounted for  $60 \pm 23.7\%$  of all somatic mutations) [6, 7]. The APOBEC-mediated mutational process was mainly C>G or C>T mutation. The similar mutational process was also observed in UC (accounted for 65.5% of all somatic mutations) [6–9]. SCLC harbored also a high mutational rate (7.4 protein-changing mutations per million base pairs) [10]. However, the mutational process was predominantly C: G>A: T transversions [11]. Hence, here, we summarize features of molecular alterations in SCCB by comparing with SCLC and UC, including cell cycle-related genes (TP53, RB1, CKDN2A, and MDM2), Telomerase reverse transcriptase (TERT) promoters, and chromosome modifiers. Meantime, we discuss potential clinical significance of these molecular alterations and current status of treatments and follow-ups in SCCB.

# **Molecular alterations in SCCB**

## The relevance of TP53, MDM2, RB1, CDKN2A

TP53 gene is located on chromosome 17p13 and codes the tumor suppressor protein, p53 protein. p53 protein participates in numerous cellular processes, containing cell cycle arrest, DNA repair, apoptosis, metabolism, autophagy, and so on [12, 13]. More than half of malignant tumors are associated with alterations of TP53 gene, and missense mutation of is mainly in the form of mutation [14–16]. TP53 gene mutations not only deprive p53 protein of its antitumor effect, namely loss-of-function, but may make some mutant p53 protein gain new ability to promote tumor formation, that is gain-of-function [17, 18].

Mutant p53 protein plays an important role in promoting tumorigenesis and development, but it needs the assistance of other molecules, such as MDM2, RB1, and CDKN2A.

It is recognized that MDM2 is a negative regulatory factor of p53 protein. The MDM2 gene is a pseudogene on chromosome 12, and encodes a MDM2 protein with distinctive E3 ubiquitin ligase effect, degrading p53 protein. In addition, MDM2 protein binds to the transactivated domain of p53 protein and inhibits its activity [19, 20]. Interestingly, p53 protein induces the transcription of MDM2 gene and prevents it from overexpressing in healthy conditions [21]. Therefore, the relationship of p53 protein and MDM2 protein is negative feedback. The balance is broken in tumors, leading to tumorigenesis [22].

Other than MDM2 gene, RB1 gene also implicates in tumor formation. RB1 gene is one of the most important tumor suppressor genes and encodes the pRb protein. pRb protein is also a negative regulator of cell cycle and limits the cell proliferation via inhibiting E2F transcription factors [23]. Meantime, pRb protein and p16 protein have negative feedback relationship. The p16 protein is encoded by tumor suppressor gene, CDKN2A gene. And it is a negative feedback role in cyclinD-CDK4/6-pRb-E2F cell cycle regulation pathway. The p16 protein prevents the phosphorylation of pRb by inhibiting CDK4/6, while hyperphosphorylated pRb can induce the expression of p16 protein and feedback inhibits the phosphorylation of pRb, thereby inhibiting the proliferation of the cells [24–26]. Mutations of CDKN2A gene, including homozygous deletion, loss of heterozygosity, and promoter abnormal methylation, can lead to cell proliferation out of control and promote tumor formation [27].

## Molecular alterations and clinical significance of the TP53, MDM2, RB1, and CDKN2A genes in SCCB

Molecular alterations of the TP53, MDM2, RB1, and CDKN2A Genes in SCCB are also common. In a genomic analysis of 110 cases of SCLC, inactivating events of TP53 and RB1 were approximately 65% and 90%, respectively, and bi-allelic losses involving TP53 and RB1 were nearly 100% [28]. Consistent with SCLC, alterative events in TP53 and RB1 each affected up to 90% of patients and also had a high frequency of bi-allelic mutation of these two genes in SCCB [6, 29, 30]. UC also harbored the alteration of these two genes, however, the rate of which was lower than SCCB. Hence, mutations of TP53 and RB1 genes in small cell carcinoma were related to tissue [6].

However, in a review, researchers showed that the loss of TP53 and RB1 proteins may be the feature of invasive tumors rather than the basic characteristics of neuroendocrine tumors [31]. Similar to the above point of view, although dual loss of TP53 and RB1 was universal in SCCB, these genes were not sufficient for small cell phenotype [6, 8]. Therefore, other factors, other than the loss of TP53 and RB1, could contribute to the small cell phenotype. Furthermore, alterations of these genes allowed tumor cells getting lineage plasticity and made cells transit to alternative lineages, which building up a tolerance to drugs, escaping immune surveillance and so on [31].

Taken together, TP53 and RB1 are probably therapeutic targets for drug resistance. And further studies need to be conducted aiming to elucidate other driver gene mutations in SCCB.

In addition loss of TP53 and RB1, in SCCB, the inactivation of CDKN2A gene was mainly due to loss of heterozygous (frequency of LOH: 35-47%), as had been shown in UC (frequency of LOH: 30-47%) [32, 33]. By contrast, the deletion frequency of CDKN2A gene was 5% in SCLC [28]. CDKN2A gene deletion mutation was a predictor of increased aggressiveness and worse prognosis in UC [34, 35]. SCCB shares similar frequency of CDKN2A gene deletion mutation with UC, but it is undefined whether effects of CDKN2A gene deletion mutation are similar to that in UC. Recently, Chang et al. demonstrated that SCCB lacked CDKN2A gene deletion mutation (P = 0.02) [6]. Thus, CDKN2A gene deletion mutation in SCCB is controversial. Further confirmation of the existence of CDKN2A gene deletion mutation and its role in SCCB studies will be useful in future.

Meantime, several studies have also found that SCCB has high-level amplifications of 1p22-32, 3q26.3, 8q24, 12q14-21, and gains of 5q, 6p, 8q, and 20q. What is more, endogens are located at some of these sites, like MDM2 [32, 36]. On the basis of the negative relationship between

p53 protein and MDM2 protein, Gupta et al. suggested that the inhibition of MDM2 ubiquitin ligase activity and the interaction of MDM2-p53 could rescue the p53 protein and achieve the purpose of inhibiting tumor [37]. Deliver the goods, there were several ongoing clinic trials of the MDM2 inhibitors in other cancers, such as acute myelocytic leukemia and myelodysplastic syndrome [38]. More interestingly, MDM2 gene amplification and TP53 gene mutation were opposed to each other in human cancer [39]. The preclinical data support this point, which was drug resistance of MDM2 inhibitors in TP53 mutant cells [40]. The mechanism of drug resistance in TP53 mutant cell is not very clear and is worth to be studied deeply.

## Special-bladder molecular alterations

SCCB also harbors special-bladder molecular alterations, including TERT promoter mutation and epigenetic modifier alteration.

#### **TERT promoter mutation**

*TERT* promoter mutations have been found to be related to many human tumors, such as UC (55.6% and 82.8%) [41, 42]. In a retrospective study with a sample size of 11 cases, TERT promoter mutations were also high frequency in SCCB (nearly 100%), but it did not exist in small cell tumor of other organs, including lung and prostate [43]. Subsequently, by sequencing of 341 key cancer-associated genes, Chang et al. also discovered that 95% of patients with SCCB had high frequency of TERT promoter mutations, which were not found in SCLC [6]. Thus, SCCB can distinguish from small cell tumors of other organs via TERT promoter mutations.

Several studies highlighted that TERT promoter mutations were no relativity with tumor stage and grade, but were meaningful urine biomarkers for patients with papillary and flat noninvasive UC in the follow-up [44, 45]. TERT promoter mutations probably also are meaningful urine biomarker in SCCB, but hypothesis requires to be verified.

#### **Epigenetic modifier alteration**

The majority of patients with SCCB harbored a high rate of bladder-specific mutations in diverse epigenetic modifiers including ARID1A, KDM6A, CREBBP, EP300, and KMT2A/C/D [6, 7]. SCCB shared a similar frequency of these chromatin-modifying genes with UC, but not with SCLC ( $P < 10^{-6}$ ). It suggested that SCCB was distinct from SCLC and likely came from a UC precursor [6, 8, 31]. Furthermore, these genes may be potential differential diagnosis markers in future.

## The cellular origin of the SCCB

So far, the cellular origin of the SCCB is still unclear. According to histology analysis, SCCB usually mixed with other components, the most common of which is urothelium component [7]. As early as 2005, Cheng et al. presented that these two components of SCCB had common clone origin [32]. One study showed that urothelial cells could differentiate into various cell types, including glandular cells, squamous cells, and neuroendocrine cells via modulating microRNA-145 [46]. The molecular changes of SCCB are more similar to UC (discussed in 'Molecular alterations in SCCB'), which provided stronger evidence that SCCB came from a preexisting urothelial cancer. The study in preclinical model has discovered that different lesions arised from distinct precursors of UC and displayed diverse invasiveness and prognosis [47]. It is worth to discuss what kind of lesion SCCB originates from UC, because it can influence clinic treatment and management.

#### **Treatment and follow-up**

At present, there are two widely used staging systems for SCCB: The Veterans Administration Lung Study Group staging system (limited disease and extensive disease) [48] and The American Joint Committee on Cancer staging system (limited disease:  $T_{1-4}$ ,  $N_{0-1}$ ,  $M_0$ , and extensive disease:  $T_X$ ,  $N_X$ ,  $M_1$  or  $T_X$ ,  $N_{2-3}$ ,  $M_0$ ) [49].

#### Limited disease

Given the rarity of SCCB, therapeutic strategies are not well established. Over 3 decades, researchers have attempted to improve survival rate via various therapeutic strategies, including cystectomy, chemotherapy, radiotherapy, and any combination of these three treatments [50]. However, survival data are still stagnant and treatments are still based on stages and the state of patient.

Patients with limited disease (LD) can be divided into two groups: operable and bladder sparing. In 2004, a retrospective study showed that there was no survival difference between patients with cystectomy and those without cystectomy (P = 0.65, N = 64) [51]. The conclusion, which patients received initial cystectomy alone with poor survival, was consistent with some other retrospective studies [52, 53]. Therefore, researchers proposed cystectomy plus multimodal treatment (chemotherapy, radiotherapy). Some retrospective reviews suggested that patients got potential benefits of improved survival from preoperative chemotherapy [52, 54, 55]. In 2009, the results of the small phase II clinical trial are no difference with previous retrospective studies, which provided stronger evidence for the benefit of neoadjuvant chemotherapy [56]. In a retrospective analysis of 175 cohort, Lynch et al. demonstrated that besides improving long-term survival, neoadjuvant chemotherapy also could make pathological downstaging to  $\leq pT1N0$  compared with initial cystectomy [62% vs. 9%; odds ratio: 44.55; 95% CI (10.39–191)] [57]. Thus, in 2016, The National Comprehensive Cancer Network guidelines for Oncology recommend that patients with SCCB undergo initial chemotherapy followed by radiotherapy or cystectomy without systemic disease [36].

However, the effect of adjuvant chemotherapy on survival is still not very clear in contrast with neoadjuvant chemotherapy. In some early studies, the impact of adjuvant chemotherapy on survival was not superior to cystectomy alone [52, 58]. Conversely, Kaushik et al. suggested that the 5-year overall survival was improved in patients receiving adjuvant chemotherapy, compared with those who did not (43% vs. 20%; P = 0.03) [59]. The reasons for the difference in the effect of adjuvant chemotherapy on survival may include the following: First, these results mainly are on basis of some retrospective and single-institution studies, which have some limitations, such as small sample size, heterogeneity of chemotherapy regimen, incomparability of baseline data, and so on. Then, so far, the diagnostic criterion has not been established well and SCCB often mixes with other components. Therefore, a subset of patients only is diagnosed after cystectomy and some patients are overlooked or misdiagnosed with other bladder tumors. So, Lynch et al. promoted the effect of adjuvant chemotherapy may have been underestimated. And they suggested neoadjuvant chemotherapy was provided for SCCB diagnosed before cystectomy; however, adjuvant chemotherapy was provided when diagnosed by cystectomy [57].

In conclusion, neoadjuvant chemotherapy is necessary for patients with SCCB who can be operated on. Although

Table 1 Treatments, survival, and recurrence of bladder preservation

the effect of adjuvant chemotherapy on survival is uncertain and need to be verified by prospective study, whether or not to use adjuvant chemotherapy depends on the situation of patient.

Patients, who have poor basic condition or are unwilling to operate, are treated with bladder-preservation therapy. In histology, SCCB was consistent with SCLC, so local radiotherapy and systemic chemotherapy was still advocated. Therefore, the bladder-preservation therapeutic strategies included TURBT plus chemoradiation or chemoradiation (sequential or concurrent chemoradiation) [50, 60, 61]. Some retrospective studies demonstrated that chemoradiation improved survival in patients with bladder sparing [50, 60-65], which is shown in Table 1. And cystectomy alone could be substituted by sequential chemoradiation, when patients aimed to preserve the bladder [50, 61, 64]. However, it is still unclear whether the curative effect of chemoradiation can be equated with that of chemotherapy plus surgery, which needs to be studied deeply.

Patients with bladder preservation have the following two problems: The optimal dose of radiation is still uncertain. In a retrospective study based on the National Cancer Database, they took 79 years old as the critical point and demonstrated that the optimal total dose between 54 and 60 Gy was recommended for patients aged 79 or younger. Conversely, among patients over the age of 79, the overall survival time was not affected by the total dose. Taking into account toxicity of radiotherapy, they also suggested that this group of patients might be better to get total dose less than 54 Gy [66]. However, this is, after all, a retrospective study, which also has many limitations as has been shown in other retrospective studies. In addition, since early retrospective studies have reported median/mean dose of radiotherapy between 59 and 64.5 Gy (Table 1), it is difficult to decide the rationality of

Author	Bex et al.	Bex et al.	Meijer et al.	Mattes et al.	Bryant et al.	Kamp et al.
Reference	[62]	[63]	[61]	[64]	[50]	[67]
Sample size	25	42	66	19	11	110
Stage of LD	$T_X N_0 - N_1 M_0$	$pT_{X}cN_{0-1}cM_{0}$	$T_{X-4}N_{0-1}M_0$	$T_{1-4}N_{0-2}M_0$	$T_{1-4}N_{0-2}M_0$	$cT_{1-4}N_{0-1}M_0$
LD (%)	17 (68%)	17 (40.5%)	27 (40.9%)	19 (100%)	11 (100%)	89 (80.9%)
Neoadjuvant chemotherapy	58.8%	100%	100%	89.5%	100%	73%
Local therapy TURBT	62.5%	100%	100%	100%	100%	-
Sequential radiotherapy	80%	100%	100%	100%	100%	73%
Median/mean dose of radio- therapy	60 Gy	60 Gy	60 Gy	64.8 Gy	59 Gy	60 Gy
OS	-	56% (2-year)	22% (5-year)	78% (2-year)	24% (3-year)	-
		47% (3-year)				
		36% (5-year)				
Local recurrence	12.5% (24 month)	23.5%	25.9% (29 month)	25% (24 month)	27%	22% (24 month)
Distant recurrence	-	47.1% (6 month)	44.4% (10 mo)	40% (24 month)	73% (9 month)	35% (9 month)

the choice of the critical point of age and total dose. The optimal total dose still needs further prospective studies and clinical trials to identify.

Another problem for patients receiving a bladder-conservation treatment is very little known about frequency and treatment of recurrence. The recurrence included local recurrence and distant recurrence. In some previous retrospective studies, local recurrence occurred mainly at 2 years of bladder-preservation therapy, with a recurrence rate of about 12-30% [61, 62, 64]. At the same time, the tissue components of the second primary tumor were mainly UC, carcinoma in situ (CIS), or the mixture of UC and CIS, and the small cell carcinoma (SCC) was rare [61, 63]. And the rate of response of salvage therapies, such as TURB, cystectomy, neoadjuvant chemotherapy, and BCG, was 64% [67]. Compared with local recurrence, distant recurrence first occurred 6-10 months after treatment with 30-80% recurrence frequency [50, 61, 63, 67]. The main organs involved in distant recurrence were live, lung, and bone [50]. For patients with distant recurrence, salvage therapies are still not well established. Hence, researchers promoted a reasonable follow-up schedule, which was cystoscopy and chest, abdominal/pelvic CT scanning every 3 months for the first 2 years and then 6 monthly till year 5 and then annually [67].

## **Extensive disease**

The treatment for patients with extensive disease was similar to SCLC, which was cisplatin-based palliative systemic chemotherapy [4, 36].

## Immunotherapy

Nowadays, immunotherapy is very promising therapeutic strategy for tumors, especially immune checkpoint inhibitors (ICBs). At present, the most representative ICBs are programmed cell death protein 1 (PD-1)/programmed cell death-ligand 1 (PD-L1) inhibitor and cytotoxic T-lymphocyte protein 4 (CTLA4) inhibitor. In 2017, the US Food and Drug Administration (FDA) approved two PD-1 inhibitor (Nivolumab, Pembrolizumab) and three PD-L1 inhibitor (Atezolizumab, Durvalumab, Avelumab) for the treatment of advanced or metastatic UC [68]. SCCB shares similar molecular alterations with UC, so can ICB be used in advanced or metastatic SCCB?

In 2015, a study demonstrated that PD-L1 was absent in small cell neuroendocrine tumor cells. However, PD-L1 and PD-1 expressed in tumor-infiltrating cells (TIC), which are macrophages and lymphocytes, respectively [69]. Subsequently, in a transcriptomic and protein analysis of SCCB, Koshkin and colleagues discovered that the expression of PD-L1 was seen on TIC via immunohistochemical (IHC), but not on tumor cells [70]. Recently, the results of the study conducted by Mandelkow were concordant with Koshkin. And they promoted that SCCB exhibited an immune-excluded phenotype [71, 72]. The mechanism of the immune-excluded phenotype is not clear in SCCB. We put forward the following hypotheses: (1) The loss of RB1 and TP53 allows tumor cells getting lineage plasticity (discussed in 'Molecular alterations in SCCB'), which lets tumor camouflage to prevent TIC infiltration. (2) Although SCCB do not express PD-L1, it may release other inhibitors to keep from TIC infiltrating. To date, immunotherapy in small cell carcinoma of the bladder is not very clear; however, it also brings dawn to SCCB's novel treatment.

## **Conclusion and outlook**

SCCB is a low incidence and high invasiveness disease. Although SCCB has small cell component, clinical manifestations and molecular alterations are similar to UC. As we all known, SCCB is difficult to distinguish from other small cell carcinomas. As we discuss in special-bladder molecular alterations, TERT promoter mutations and epigenetic modifiers highly express in SCCB, but lack in small cell carcinoma of other organs. These special mutations may be potential markers for differential diagnosis, but they need to be translated into clinically measurable indicators.

Because of the rarity of SCCB, standard treatment strategies are not well established. The treatment of SCCB mainly consists of systemic chemotherapy, surgery, and radiation, but survival data are still stagnant. Hence, we need to seek for novel therapeutic targets. Cell cycle-related therapeutic strategies may be also promising treatment options. Although immunotherapy is in full swing in treatment of tumors, in SCCB it is not optimistic. PD-L1 expresses on TIC, so does the immune checkpoint inhibitor have any effect? Indeed, tumor microenvironment and immuneexcluded phenotype may be potential treatment targets.

SCCB still faces many challenges and opportunities: how to assess whether the patient with sparing bladder has a recurrence and the remedial measures after recurrence. During follow-up, are there any more sensitive serological or urine biomarkers to monitor tumor progression?

Although it is long way to improve prognosis of patients with SCCB, we believe advances in medicine will bring light.

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#### Compliance with ethical standards

**Conflict of interests** The authors have declared that no competing interest exists.

# References

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2018. CA Cancer J Clin. 2018;68(1):7–30. https://doi.org/10.3322/caac.21442.
- Niu Q, Lu Y, Xu S, Shi Q, Guo B, Guo Z, et al. Clinicopathological characteristics and survival outcomes of bladder neuroendocrine carcinomas: a population-based study. Cancer Manag Res. 2018;10:4479–89. https://doi.org/10.2147/CMAR.S175286.
- 3. Humphrey PA, Moch H, Cubilla AL, Ulbright TM, Reuter VE. The 2016 WHO classification of tumours of the urinary system and male genital organs-part B: prostate and bladder tumours. Eur Urol. 2016;70(1):106–19. https://doi.org/10.1016/j.eurur o.2016.02.028.
- Erdem GU, Ozdemir NY, Demirci NS, Sahin S, Bozkaya Y, Zengin N. Small cell carcinoma of the urinary bladder: changing trends in the current literature. Curr Med Res Opin. 2016;32(6):1013–21. https://doi.org/10.1185/03007995.2016.1155982.
- Schreiber D, Rineer J, Weiss J, Leaf A, Karanikolas N, Rotman M, et al. Characterization and outcomes of small cell carcinoma of the bladder using the surveillance, epidemiology, and end results database. Am J Clin Oncol Cancer. 2013;36(2):126–31. https:// doi.org/10.1097/COC.0b013e3182438c71.
- Chang MT, Penson A, Desai NB, Socci ND, Shen R, Seshan VE, et al. Small-cell carcinomas of the bladder and lung are characterized by a convergent but distinct pathogenesis. Clin Cancer Res. 2018;24(8):1965–73. https://doi.org/10.1158/1078-0432. CCR-17-2655.
- Shen PY, Jing Y, Zhang RY, Cai MC, Ma P, Chen H, et al. Comprehensive genomic profiling of neuroendocrine bladder cancer pinpoints molecular origin and potential therapeutics. Oncogene. 2018;37(22):3039–44. https://doi.org/10.1038/s4138 8-018-0192-5.
- Oser MG, Janne PA. Small-cell neuroendocrine tumors: cell state trumps the oncogenic driver. Clin Cancer Res. 2018;24(8):1775– 6. https://doi.org/10.1158/1078-0432.CCR-17-3646.
- Robertson AG, Kim J, Al-Ahmadie H, Bellmunt J, Guo G, Cherniack AD, et al. Comprehensive molecular characterization of muscle-invasive bladder cancer. Cell. 2017;171(3):540.e25–556. e25. https://doi.org/10.1016/j.cell.2017.09.007.
- Peifer M, Fernandez-Cuesta L, Sos ML, George J, Seidel D, Kasper LH, et al. Integrative genome analyses identify key somatic driver mutations of small-cell lung cancer. Nat Genet. 2012;44(10):1104–10. https://doi.org/10.1038/ng.2396.
- Augert A, Zhang Q, Bates B, Cui M, Wang X, Wildey G, et al. Small cell lung cancer exhibits frequent inactivating mutations in the histone methyltransferase KMT2D/MLL2: CALGB 151111 (alliance). J Thorac Oncol. 2017;12(4):704–13. https://doi. org/10.1016/j.jtho.2016.12.011.
- Fischer M. Census and evaluation of p53 target genes. Oncogene. 2017;36(28):3943–56. https://doi.org/10.1038/onc.2016.502.
- Freed-Pastor WA, Prives C. Mutant p53: one name, many proteins. Genes Dev. 2012;26(12):1268–86. https://doi.org/10.1101/ gad.190678.112.
- Lawrence MS, Stojanov P, Mermel CH, Robinson JT, Garraway LA, Golub TR, et al. Discovery and saturation analysis of cancer genes across 21 tumour types. Nature. 2014;505(7484):495–501. https://doi.org/10.1038/nature12912.

- Berman-Booty LD, Knudsen KE. Models of neuroendocrine prostate cancer. Endocr Relat Cancer. 2015;22(1):R33–49. https ://doi.org/10.1530/ERC-14-0393.
- Di Agostino S, Fontemaggi G, Strano S, Blandino G, D'Orazi G. Targeting mutant p53 in cancer: the latest insights. J Exp Clin Cancer Res. 2019;38(1):290. https://doi.org/10.1186/s1304 6-019-1302-0.
- 17. Mantovani F, Collavin L, Del Sal G. Mutant p53 as a guardian of the cancer cell. Cell Death Differ. 2019;26(2):199–212. https ://doi.org/10.1038/s41418-018-0246-9.
- Kim MP, Lozano G. Mutant p53 partners in crime. Cell Death Differ. 2018;25(1):161–8. https://doi.org/10.1038/cdd.2017.185.
- Nag S, Zhang X, Srivenugopal KS, Wang MH, Wang W, Zhang R. Targeting MDM2-p53 interaction for cancer therapy: are we there yet? Curr Med Chem. 2014;21(5):553–74. https://doi. org/10.2174/09298673113206660325.
- Frezza AM, Assi T, Lo Vullo S, Ben-Ami E, Dufresne A, Yonemori K, et al. Systemic treatments in MDM2 positive intimal sarcoma: a multicentre experience with anthracycline, gemcitabine, and pazopanib within the World Sarcoma Network. Cancer. 2019. https://doi.org/10.1002/cncr.32508.
- Zhang Q, Zeng SX, Lu H. Targeting p53-MDM2-MDMX loop for cancer therapy. Subcell Biochem. 2014;85:281–319. https ://doi.org/10.1007/978-94-017-9211-0\_16.
- Yang L, Song T, Cheng Q, Chen L, Chen J. Mutant p53 sequestration of the MDM2 acidic domain inhibits E3 ligase activity. Mol Cell Biol. 2019. https://doi.org/10.1128/mcb.00375-18.
- Dyson NJ. RB1: a prototype tumor suppressor and an enigma. Genes Dev. 2016;30(13):1492–502. https://doi.org/10.1101/ gad.282145.116.
- D'Arcangelo D, Tinaburri L, Dellambra E. The role of p16(INK4a) pathway in human epidermal stem cell selfrenewal, aging and cancer. Int J Mol Sci. 2017. https://doi. org/10.3390/ijms18071591.
- 25. Fu HC, Chuang IC, Yang YC, Chuang PC, Lin H, Ou YC, et al. Low P16(INK4A) expression associated with high expression of cancer stem cell markers predicts poor prognosis in cervical cancer after radiotherapy. Int J Mol Sci. 2018. https://doi. org/10.3390/ijms19092541.
- He SH, Sharpless NE. Senescence in health and disease. Cell. 2017;169(6):1000-11. https://doi.org/10.1016/j. cell.2017.05.015.
- Noe M, Hackeng WM, de Leng WWJ, Vergeer M, Vleggaar FP, Morsink FHM, et al. Well-differentiated pancreatic neuroendocrine tumor in a patient with familial atypical multiple mole melanoma syndrome (FAMMM). Am J Surg Pathol. 2019. https://doi. org/10.1097/PAS.00000000001314.
- George J, Lim JS, Jang SJ, Cun Y, Ozretic L, Kong G, et al. Comprehensive genomic profiles of small cell lung cancer. Nature. 2015;524(7563):47–53. https://doi.org/10.1038/nature14664.
- Meder L, Konig K, Ozretic L, Schultheis AM, Ueckeroth F, Ade CP, et al. NOTCH, ASCL1, p53 and RB alterations define an alternative pathway driving neuroendocrine and small cell lung carcinomas. Int J Cancer. 2016;138(4):927–38. https://doi. org/10.1002/ijc.29835.
- Al-Ahmadie H, Iyer G, Hohl M, Asthana S, Inagaki A, Schultz N, et al. Synthetic lethality in ATM-deficient RAD50-mutant tumors underlies outlier response to cancer therapy. Cancer Discov. 2014;4(9):1014–21. https://doi.org/10.1158/2159-8290. CD-14-0380.
- Rickman DS, Beltran H, Demichelis F, Rubin MA. Biology and evolution of poorly differentiated neuroendocrine tumors. Nat Med. 2017;23(6):1–10. https://doi.org/10.1038/nm.4341.
- 32. Cheng L, Jones TD, McCarthy RP, Eble JN, Wang MS, MacLennan GT, et al. Molecular genetic evidence for a common clonal origin of urinary bladder small cell carcinoma and coexisting

urothelial carcinoma. Am J Pathol. 2005;166(5):1533–9. https:// doi.org/10.1016/S0002-9440(10)62369-3.

- Pan CX, Zhang H, Lara PN Jr, Cheng L. Small-cell carcinoma of the urinary bladder: diagnosis and management. Expert Rev Anticancer Ther. 2006;6(12):1707–13. https://doi.org/10.1586/14737 140.6.12.1707.
- 34. Abat D, Demirhan O, Inandiklioglu N, Tunc E, Erdogan S, Tastemir D, et al. Genetic alterations of chromosomes, p53 and p16 genes in low- and high-grade bladder cancer. Oncol Lett. 2014;8(1):25–32. https://doi.org/10.3892/ol.2014.2108.
- 35. Worst TS, Weis CA, Stohr R, Bertz S, Eckstein M, Otto W, et al. CDKN2A as transcriptomic marker for muscle-invasive bladder cancer risk stratification and therapy decision-making. Sci Rep. 2018. https://doi.org/10.1038/s41598-018-32569-x.
- Kouba EJ, Cheng L. Understanding the genetic landscape of small cell carcinoma of the urinary bladder and implications for diagnosis, prognosis, and treatment: a review. JAMA Oncol. 2017;3(11):1570-8. https://doi.org/10.1001/jamao ncol.2016.7013.
- Gupta A, Shah K, Oza MJ, Behl T. Reactivation of p53 gene by MDM2 inhibitors: a novel therapy for cancer treatment. Biomed Pharmacother. 2019;109:484–92. https://doi.org/10.1016/j.bioph a.2018.10.155.
- Khurana A, Shafer DA. MDM2 antagonists as a novel treatment option for acute myeloid leukemia: perspectives on the therapeutic potential of idasanutlin (RG7388). Oncotargets Ther. 2019;12:2903–10. https://doi.org/10.2147/Ott.S172315.
- Oliner JD, Saiki AY, Caenepeel S. The role of MDM2 amplification and overexpression in tumorigenesis. Cold Spring Harb Perspect Med. 2016. https://doi.org/10.1101/cshperspect.a0263 36.
- Weisberg E, Halilovic E, Cooke VG, Nonami A, Ren T, Sanda T, et al. Inhibition of wild-type p53-expressing AML by the novel small molecule HDM2 inhibitor CGM097. Mol Cancer Ther. 2015;14(10):2249–59. https://doi.org/10.1158/1535-7163. Mct-15-0429.
- Giedl J, Rogler A, Wild A, Riener MO, Filbeck T, Burger M, et al. TERT core promotor mutations in early-onset bladder cancer. J Cancer. 2016;7(8):915–20. https://doi.org/10.7150/jca.15006.
- Avogbe PH, Manel A, Vian E, Durand G, Forey N, Voegele C, et al. Urinary TERT promoter mutations as non-invasive biomarkers for the comprehensive detection of urothelial cancer. EBioMedicine. 2019;44:431–8. https://doi.org/10.1016/j.ebiom .2019.05.004.
- 43. Zheng XY, Jian ZG, Bezerra SM, Faraj SF, Munari E, Fallon JT, et al. High frequency of TERT promoter mutation in small cell carcinoma of bladder, but not in small cell carcinoma of other origins. J Hematol Oncol. 2014. https://doi.org/10.1186/s1304 5-014-0047-7.
- 44. Rane JK, Simms MS, Maitland NJ. Re: Yves Allorya, Willemien Beukers, Ana Sagrera, et al. Telomerase reverse transcriptase promoter mutations in bladder cancer: high frequency across stages, detection in urine, and lack of association with outcome. Eur Urol 2014;65:360-6: telomerase expression and stem cells: urologic epithelial perspective. Eur Urol. 2014;65(6):E85–6. https://doi. org/10.1016/j.eururo.2014.02.030.
- 45. Kinde I, Munari E, Faraj SF, Hruban RH, Schoenberg M, Bivalacqua T, et al. TERT promoter mutations occur early in urothelial neoplasia and are biomarkers of early disease and disease recurrence in urine. Can Res. 2013;73(24):7162–7. https://doi. org/10.1158/0008-5472.Can-13-2498.
- 46. Fujii T, Shimada K, Tatsumi Y, Hatakeyama K, Obayashi C, Fujimoto K, et al. microRNA-145 promotes differentiation in human urothelial carcinoma through down-regulation of syndecan-1. BMC Cancer. 2015;15:818. https://doi.org/10.1186/s1288 5-015-1846-0.

- Van Batavia J, Yamany T, Molotkov A, Dan H, Mansukhani M, Batourina E, et al. Bladder cancers arise from distinct urothelial sub-populations. Nat Cell Biol. 2014;16(10):982–91. https://doi. org/10.1038/ncb3038.
- Micke P, Faldum A, Metz T, Beeh KM, Bittinger F, Hengstler JG, et al. Staging small cell lung cancer: Veterans Administration Lung Study Group versus International Association for the Study of Lung Cancer: what limits limited disease? Lung Cancer. 2002;37(3):271–6.
- 49. Nicholson AG, Chansky K, Crowley J, Beyruti R, Kubota K, Turrisi A, et al. The International Association for the Study of Lung Cancer Lung Cancer Staging Project: proposals for the revision of the clinical and pathologic staging of small cell lung cancer in the forthcoming eighth edition of the TNM classification for lung cancer. J Thorac Oncol. 2016;11(3):300–11. https ://doi.org/10.1016/j.jtho.2015.10.008.
- Bryant CM, Dang LH, Stechmiller BK, Gilbert SM, Morris CG, Zlotecki RA. Treatment of small cell carcinoma of the bladder with chemotherapy and radiation after transurethral resection of a bladder tumor. Am J Clin Oncol. 2016;39(1):69–75. https ://doi.org/10.1097/COC.00000000000027.
- Montie JE. Small cell carcinoma of the urinary bladder. A clinicopathologic analysis of 64 patients. J Urol. 2005;173(6):1920– 1. https://doi.org/10.1097/01.ju.0000161245.95285.b2.
- 52. Siefker-Radtke AO, Dinney CP, Abrahams NA, Moran C, Shen Y, Pisters LL, et al. Evidence supporting preoperative chemotherapy for small cell carcinoma of the bladder: a retrospective review of the M.D. Anderson cancer experience. J Urol. 2004;172(2):481–4.
- Quek ML, Nichols PW, Yamzon J, Daneshmand S, Miranda G, Cai J, et al. Radical cystectomy for primary neuroendocrine tumors of the bladder: the university of southern california experience. J Urol. 2005;174(1):93–6. https://doi.org/10.1097/01. ju.0000162085.20043.1f.
- Walther PJ. Adjuvant/neo-adjuvant etoposide/cisplatin and cystectomy for management of invasive small cell carcinoma of the bladder. J Urol. 2002;167(4):285.
- 55. Pasquier D, Barney B, Sundar S, Poortmans P, Villa S, Nasrallah H, et al. Small cell carcinoma of the urinary bladder: a retro-spective, multicenter rare cancer network study of 107 patients. Int J Radiat Oncol Biol Phys. 2015;92(4):904–10. https://doi.org/10.1016/j.ijrobp.2015.03.019.
- Siefker-Radtke AO, Kamat AM, Grossman HB, Williams DL, Qiao W, Thall PF, et al. Phase II clinical trial of neoadjuvant alternating doublet chemotherapy with ifosfamide/doxorubicin and etoposide/cisplatin in small-cell urothelial cancer. J Clin Oncol. 2009;27(16):2592–7. https://doi.org/10.1200/Jco.2008.19.0256.
- 57. Lynch SP, Shen Y, Kamat A, Grossman HB, Shah JB, Millikan RE, et al. Neoadjuvant chemotherapy in small cell urothelial cancer improves pathologic downstaging and long-term outcomes: results from a retrospective study at the MD Anderson Cancer Center. Eur Urol. 2013;64(2):307–13. https://doi.org/10.1016/j.eururo.2012.04.020.
- Bhatt VR, Loberiza FR Jr, Tandra P, Krishnamurthy J, Shrestha R, Wang J. Risk factors, therapy and survival outcomes of small cell and large cell neuroendocrine carcinoma of urinary bladder. Rare Tumors. 2014;6(1):5043. https://doi.org/10.4081/rt.2014.5043.
- Kaushik D, Frank I, Boorjian SA, Cheville JC, Eisenberg MS, Thapa P, et al. Long-term results of radical cystectomy and role of adjuvant chemotherapy for small cell carcinoma of the bladder. Int J Urol. 2015;22(6):549–54. https://doi.org/10.1111/iju.12729.
- Lohrisch C, Murray N, Pickles T, Sullivan L. Small cell carcinoma of the bladder: long term outcome with integrated chemoradiation. Cancer. 1999;86(11):2346–52.
- 61. Meijer RP, Meinhardt W, van der Poel HG, van Rhijn BW, Kerst JM, Pos FJ, et al. Local control rate and prognosis after sequential

chemoradiation for small cell carcinoma of the bladder. Int J Urol. 2013;20(8):778–84. https://doi.org/10.1111/iju.12038.

- Bex A, Nieuwenhuijzen JA, Kerst M, Pos F, van Boven H, Meinhardt W, et al. Small cell carcinoma of bladder: a single-center prospective study of 25 cases treated in analogy to small cell lung cancer. Urology. 2005;65(2):295–9. https://doi.org/10.1016/j.urology.2004.09.049.
- 63. Bex A, de Vries R, Pos F, Kerst M, Horenblas S. Long-term survival after sequential chemoradiation for limited disease small cell carcinoma of the bladder. World J Urol. 2009;27(1):101–6. https://doi.org/10.1007/s00345-008-0304-x.
- Mattes MD, Kan CC, Dalbagni G, Zelefsky MJ, Kollmeier MA. External beam radiation therapy for small cell carcinoma of the urinary bladder. Pract Radiat Oncol. 2015;5(1):e17–22. https:// doi.org/10.1016/j.prro.2014.03.013.
- 65. Akamatsu H, Nakamura K, Ebara T, Inaba K, Itasaka S, Jingu K, et al. Organ-preserving approach via radiotherapy for small cell carcinoma of the bladder: an analysis based on the Japanese Radiation Oncology Study Group (JROSG) survey. J Radiat Res. 2019. https://doi.org/10.1093/jrr/rrz018.
- 66. Germino E, Fischer-Valuck BW, Rudra S, Rao YJ, Contreras J, Abraham C, et al. Radiation therapy as definitive local treatment in patients with limited-stage small cell carcinoma of the bladder: does total dose matter? Bladder Cancer. 2018;4(3):311–7. https:// doi.org/10.3233/BLC-180165.
- 67. van de Kamp M, Meijer R, Pos F, Kerst M, van Werkhoven E, van Rhijn B, et al. Intravesical recurrence after bladder sparing treatment of small cell carcinoma of the bladder: characteristics, treatment, and outcome. Urol Oncol. 2018;36(6):307.e1–8. https://doi.org/10.1016/j.urolonc.2018.02.015.

- Hsu FS, Su CH, Huang KH. A comprehensive review of US FDA-approved immune checkpoint inhibitors in urothelial carcinoma. J Immunol Res. 2017;2017:6940546. https://doi. org/10.1155/2017/6940546.
- Schultheis AM, Scheel AH, Ozretic L, George J, Thomas RK, Hagemann T, et al. PD-L1 expression in small cell neuroendocrine carcinomas. Eur J Cancer. 2015;51(3):421–6. https://doi. org/10.1016/j.ejca.2014.12.006.
- Koshkin VS, Garcia JA, Reynolds J, Elson P, Magi-Galluzzi C, McKenney JK, et al. Transcriptomic and protein analysis of small-cell bladder cancer (SCBC) identifies prognostic biomarkers and DLL3 as a relevant therapeutic target. Clin Cancer Res. 2019;25(1):210–21. https://doi.org/10.1158/1078-0432. Ccr-18-1278.
- Mandelkow T, Blessin NC, Lueerss E, Pott L, Simon R, Li WC, et al. Immune exclusion is frequent in small-cell carcinoma of the bladder. Dis Markers. 2019. https://doi.org/10.1155/2019/25325 18.
- Chen DS, Mellman I. Elements of cancer immunity and the cancer-immune set point. Nature. 2017;541(7637):321–30. https://doi.org/10.1038/nature21349.

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