

Down-regulation of Dicer in hepatocellular carcinoma

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Abstract Dicer, the key enzyme in the RNAi pathway, is misregulated in tumor tissues. The altered expression of Dicer is associated with clinical characteristics in patients with cancer. Liver carcinoma and adjacent non-neoplastic tissues were obtained from 36 patients with hepatocellular carcinoma (HCC) undergoing surgery. Expressions of Dicer mRNA were evaluated using the Real-time reverse transcription-PCR in 36 liver carcinoma tissues and 36 adjacent histologically non-cancerous liver tissues. Dicer mRNA levels were evaluated in relation to age, sex, tumor number, tumor size, tumor stage, and distant metastasis. Dicer mRNA level was significantly lower in malignant tissues than in the corresponding non-neoplastic tissues in 34 of the

36 patients with HCC (94.4%). The Dicer expression level was not associated with clinical characteristics, including age, sex, tumor number, tumor size, tumor stage, or distant metastasis in HCC cases. These results demonstrate that Dicer is significantly down-regulated in HCC, suggesting that reduced expression of Dicer may play an important role during the process of hepatocarcinogenesis.

Keywords HCC · Dicer · Real-time PCR

Introduction

Hepatocellular carcinoma (HCC) is one of the leading causes of cancer-related death [1, 2]. A total of 80–90% of the HCC cases in China, India, Korea, Singapore, and Vietnam are associated with Hepatitis B virus (HBV) infection [3]. Mutation, amplification, and epigenetic changes in various genes have been identified in liver cancers, suggesting that these genes are involved in hepatocarcinogenesis [4, 5]. In addition, emerging evidence suggests that components of the RNA interference (RNAi) pathway and miRNAs may play important roles in the carcinogenesis of HCC [6–16].

Dicer, the key enzyme in the RNAi pathway, is upregulated in prostate adenocarcinoma [17], ovarian serous carcinomas [18], pleomorphic adenomas of the salivary gland [19], and acute myeloid leukemia (AML) [20]. Reduced expression of Dicer is associated with poor prognosis in ovarian carcinomas and patients with lung cancer [21, 22]. However, the Dicer level in patients with AML is not associated with clinical outcomes [20]. Disruption of Dicer promotes hepatocarcinogenesis [16]. In this study, we compared the Dicer mRNA between tumor tissues and corresponding adjacent non-neoplastic tissues from patients

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with HCC and investigated whether the level of Dicer mRNA is associated with clinical parameters.

Materials and methods

Patients and tissues

Liver carcinoma and adjacent non-neoplastic tissues were obtained from 36 patients with HCC undergoing surgery in

Southwest Hospital (Chongqing, China) between April 2008 and January 2009. None of the patients had received chemotherapy before operation. Tissue samples were immediately frozen in liquid nitrogen and stored at -80° until use. All patients gave their informed consent, and the study was approved by the ethics and scientific committees of Southwest Hospital. The tumor types and stages were determined according to the WHO classification. All HCC cases were confirmed to be HCC by clinical pathology. The clinicopathological characteristics are summarized in Table 1.

Table 1 Clinicopathological features and Dicer/GAPDH of patients included in the study

Patient no.	Sex	Age (years)	Relative Dicer mRNA (mean \pm SD)		Tumor		Size (cm)	Metastasis
			Tumor	Non-cancerous	Stage	Number		
1	Male	53	0.014 \pm 0.005	0.017 \pm 0.006	Ib	1	4.0	No
2	Female	40	0.006 \pm 0.003	0.043 \pm 0.007	Ib	1	3.5	No
3	Male	56	0.015 \pm 0.009	0.053 \pm 0.013	IIa	1	6.4	No
4	Male	52	0.025 \pm 0.005	0.055 \pm 0.013	IIb	>3	>10	No
5	Female	65	0.06 \pm 0.012	0.029 \pm 0.007	IIIa	1	8.0	Yes
6	Male	36	0.012 \pm 0.005	0.022 \pm 0.002	IIIa	1	7.8	Yes
7	Female	43	0.032 \pm 0.006	0.025 \pm 0.005	IIIa	1	11.0	Yes
8	Male	56	0.055 \pm 0.012	0.178 \pm 0.039	IIb	1	8.0	No
9	Male	57	0.399 \pm 0.067	0.208 \pm 0.048	IIa	1	6.1	No
10	Male	35	0.067 \pm 0.011	0.120 \pm 0.027	Ib	1	4.7	No
11	Male	37	0.117 \pm 0.034	0.198 \pm 0.021	Ib	1	4.6	No
12	Female	73	0.04 \pm 0.008	0.128 \pm 0.016	IIIa	1	7.0	No
13	Male	43	0.02 \pm 0.005	0.012 \pm 0.001	IIa	2	7.0	No
14	Male	63	0.000 \pm 0.000	0.001 \pm 0.000	IIIa	2	14.8	Yes
15	Female	31	0.017 \pm 0.005	0.018 \pm 0.003	IIIa	>3	>10	Yes
16	Male	51	0.004 \pm 0.001	0.008 \pm 0.001	Ib	1	4.6	No
17	Male	44	0.139 \pm 0.047	0.168 \pm 0.018	Ib	2	4.8	No
18	Male	52	0.679 \pm 0.086	0.742 \pm 0.093	IIb	1	10.2	No
19	Male	46	0.061 \pm 0.012	0.084 \pm 0.022	IIa	1	9.7	No
20	Male	62	0.071 \pm 0.020	0.398 \pm 0.070	IIIa	1	12.0	Yes
21	Male	53	0.0340 \pm 0.012	0.061 \pm 0.016	IIa	1	7.5	No
22	Female	40	0.0116 \pm 0.005	0.032 \pm 0.006	IIa	1	5.4	No
23	Male	56	0.022 \pm 0.008	0.045 \pm 0.005	IIb	1	12.4	No
24	Male	52	0.050 \pm 0.015	0.5678 \pm 0.013	IIa	2	9.2	No
25	Male	56	0.2000 \pm 0.04	0.408 \pm 0.043	IIa	1	6.4	No
26	Male	52	0.096 \pm 0.019	0.208 \pm 0.043	IIb	2	10.0	No
27	Female	65	0.136 \pm 0.044	0.357 \pm 0.057	IIIa	1	8.0	Yes
28	Male	36	0.080 \pm 0.023	0.104 \pm 0.018	IIIa	1	6.8	Yes
29	Female	43	0.026 \pm 0.005	0.056 \pm 0.009	IIIa	1	12.0	Yes
30	Male	49	0.019 \pm 0.009	0.042 \pm 0.010	IIb	1	11.0	No
31	Male	58	0.134 \pm 0.031	0.327 \pm 0.036	IIb	1	4.0	No
32	Female	54	0.007 \pm 0.001	0.009 \pm 0.001	IIIa	1	8.5	Yes
33	Male	51	0.132 \pm 0.038	0.207 \pm 0.023	IIb	>3	12.5	No
34	Male	60	0.325 \pm 0.066	0.648 \pm 0.065	IIa	1	7.0	No
35	Male	65	0.072 \pm 0.025	0.150 \pm 0.029	IIa	1	8.6	No
36	Male	45	0.099 \pm 0.020	0.203 \pm 0.026	IIa	1	6.0	No

Table 2 Sequence of primers used in real-time RT-PCR assay

Target gene	Function	Sequence (5'-3')
GAPDH	Forward primer	CTCTCTGCTCCTCCTGTTCGAC
	Reverse primer	TGAGCGATGTGGCTCGGCT
Dicer	Forward primer	TCCACGAGTCACAATCAACACGG
	Reverse primer	GGGTTCTGCATTTAGGAGCTAGATGAG

RNA isolation and quantitative real-time RT-PCR

Total RNA was prepared using TRIzol (Invitrogen) and reverse-transcribed using MML-V reverse transcriptase (Promega) according to the manufacturer's protocol. Samples prepared without reverse transcription served as negative control templates. SYBR Green(ABI) PCR was performed in triplicate using the ABI PRISM 7300 Sequence Detection System. All samples were normalized to the signal generated from GAPDH. Primer sequences are presented in Table 2.

Western blot analysis

Total protein content was measured using the Bradford protein assay. Samples were denatured at 100°C for 5 min. Equal amounts of total protein were loaded to each well for electrophoresis in SDS–polyacrylamide gels and then transferred to polyvinylidene fluoride microporous membranes (Millipore). Membranes were then incubated with primary antibody followed by incubation with horseradish peroxidase–linked secondary antibodies. The primary antibodies included anti-Dicer (ab14601; Abcam) and anti-GAPDH (KC-5G4; Kangchen Biotech).

Statistical analysis

Statistical analysis was performed with SPSS 11.5. Statistical comparisons for significance were made with

Wilcoxon signed-rank test for paired samples and with Mann–Whitney *U* Kruskal–Wallis test for unpaired samples. The correlation between continuous variables was examined by means of Spearman's rank-order coefficients. A level of $P < 0.05$ was considered significant.

Results

Expression of Dicer mRNA in cancerous liver tissues and adjacent non-neoplastic liver tissues of HCC cases

The Dicer mRNA was quantified by real-time RT-PCR in paired samples of tumor tissues and adjacent non-tumor tissues from 36 patients with HCC. The Dicer mRNA level was normalized to the level of GAPDH mRNA. In contrast to prostate adenocarcinoma [17], lung adenocarcinoma [23], ovarian serous carcinomas [18], and AML [20], where Dicer is upregulated, the Dicer mRNA level was significantly lower in malignant tissues than in the corresponding non-neoplastic tissues in 34 of the 36 patients with HCC (94.4%) (Table 1; Fig. 1). To determine whether the decreased Dicer mRNA level was mirrored by reduced protein expression, western blot was performed. As expected, our data indicated that Dicer protein was down-regulated in most of the malignant tissues. The expressions of Dicer protein from a typical case are indicated in Fig. 2.

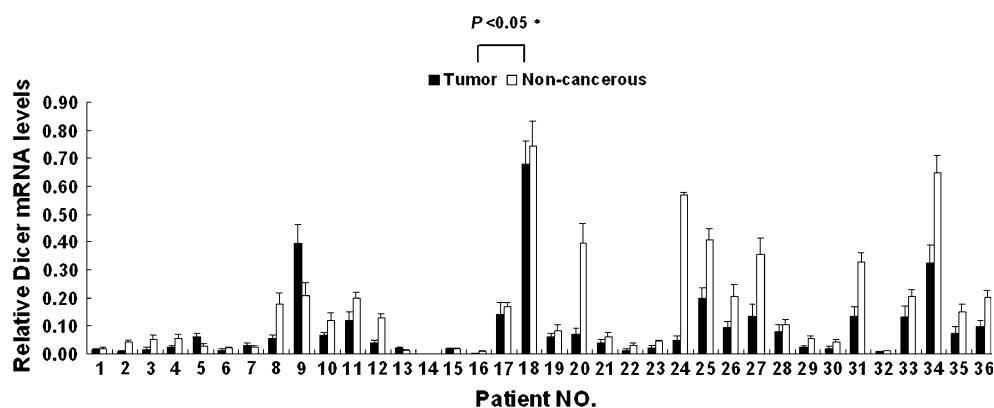


Fig. 1 Relative Dicer mRNA in paired HCC tumor tissues and the adjacent non-cancerous liver tissues ($n = 36$). The relative Dicer mRNA levels (normalized to the corresponding GAPDH mRNAs)

were significantly lower in tumor tissues compared to adjacent non-cancerous liver tissues.* Wilcoxon signed rank Test

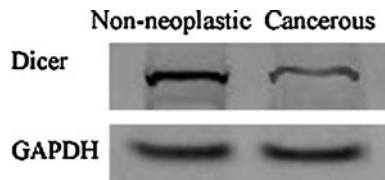


Fig. 2 Dicer protein expression from a typical case. GAPDH was used as an internal control

Relationship between Dicer mRNA level and the clinical parameters in patients with HCC

Dicer expression level is associated with clinicopathological features, including clinical outcomes, in ovarian carcinoma and patients with lung cancer [12–23]. Therefore, we investigated whether the expression level of Dicer is associated with any clinicopathologic characteristics of HCC. Statistical analysis indicated that there was no significant association between *Dicer* expression level and clinical characteristics, including age, sex, tumor number, tumor size, tumor stage, and distant metastasis (Table 3; Figs. 3, 4).

Table 3 Correlation of clinical and pathological features with relative Dicer mRNA levels in 36 patients with HCC

Clinical features	No.	P value	Significance
Sex			
Male	9	0.079 ^a	No
Female	27		
Stage			
I	10		
II	15	0.172 ^b	No
III	11		
Tumor number			
Single	25	0.612 ^a	No
Multiple	11		
Tumor size(cm)			
≥10	26	0.374 ^a	No
<10	10		
Metastasis			
Yes	12	0.280 ^a	No
No	24		

^a Mann–Whitney *U* test

^b Kruskal–Wallis test

Fig. 3 Boxplot graphs of correlation between clinical features and relative Dicer mRNA levels in patients with HCC. The relative Dicer mRNA levels of patients with HCC have no significant correlation with sex (a), tumor numbers (b), metastasis (c), stage (d), and tumor size (cutpoint: 10 cm) (e). *Mann–Whitney *U* Test, [§]Kruskal–Wallis Test

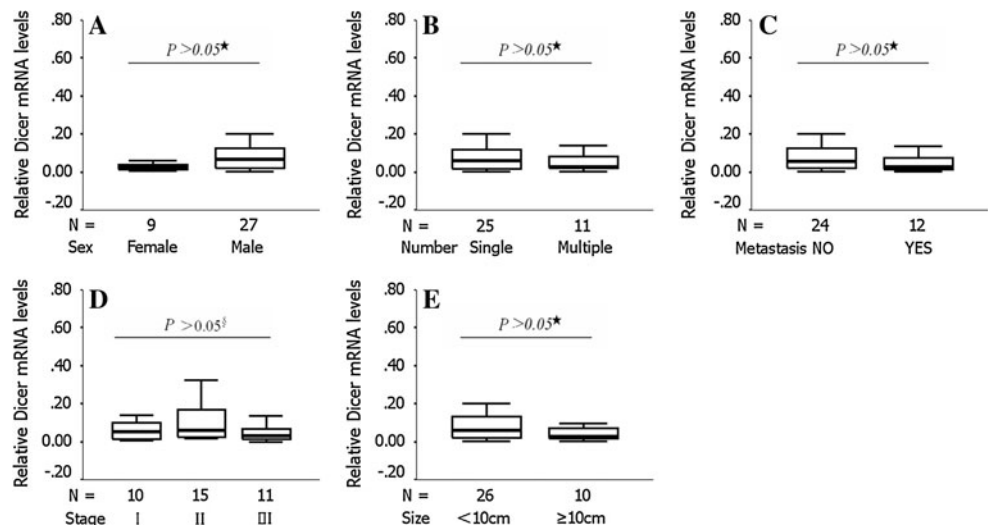
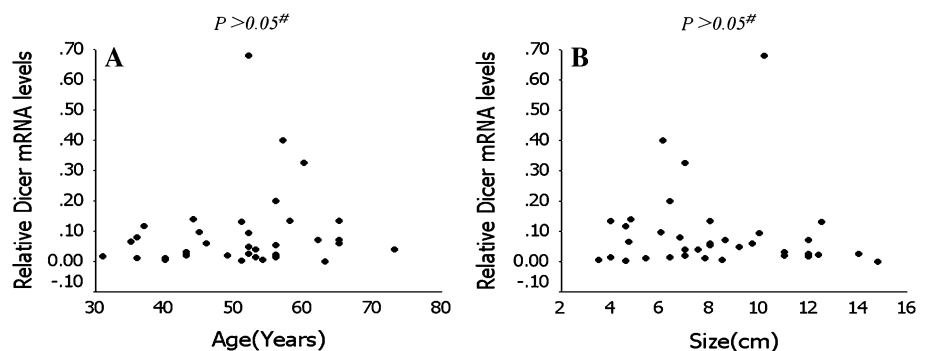


Fig. 4 Scatterplot graphs of correlation between clinical features and relative Dicer mRNA levels in patients with HCC. The relative Dicer mRNA levels of patients with HCC have no significant correlation with age (a) or tumor size (b). [#]Spearman’s rank-order coefficients test



Discussion

Dicer is upregulated in prostate adenocarcinoma [17], ovarian serous carcinomas [18], pleomorphic adenomas of the salivary gland [19], and acute myeloid leukemia (AML) [20]. Here, we found that Dicer is down-regulated in hepatocellular carcinoma. Reduced expression of Dicer is associated with poor prognosis in ovarian carcinomas and patients with lung cancer [21, 22]. Prognosis of patients with HCC is poor [24], suggesting that reduced Dicer expression is associated with the poor outcomes of patients with HCC. However, Martin et al. [20] found that Dicer level is not associated with clinical outcomes in patients with AML, suggesting that reduced Dicer expression may not be related to the poor prognosis of patients with HCC.

Altered Dicer expression is associated with the progress of lung adenocarcinoma. In the proposed multistep carcinogenesis model, peripheral adenocarcinoma of the lung develops from a precursor lesion referred to as atypical adenomatous hyperplasia (AAH). AAH transforms into non-mucinous bronchioalveolar carcinoma (BAC), which progresses into invasive adenocarcinoma. Dicer is upregulated in AAH and BAC and down-regulated in invasive lung adenocarcinoma [23]. We found that the Dicer mRNA level is not associated with the stage of HCC. However, we cannot rule out the possibility that Dicer expression is a dynamic progress during hepatocarcinogenesis. It is possible that Dicer is upregulated in HCC at the very early stage during hepatocarcinogenesis.

Approximately 50% of all annotated human miRNAs are located in areas of the genome associated with cancer or “fragile sites” [25], and thus, miRNAs might have a crucial function in cancer progression. In fact, increasing evidence shows that expression of miRNAs is down-regulated in most of the human cancer [26, 27]. Dicer is essential for miRNA biogenesis; loss of Dicer in mice disrupts embryonic stem-cell differentiation and is lethal during early development. Low levels of Dicer mRNA also affect normal cellular development and differentiation [25–28]. Disruption of Dicer1 promotes hepatocarcinogenesis [16]. Furthermore, abnormalities in the copy number of the Dicer gene have been found in human melanoma, breast, and ovarian cancers [29]. It is therefore possible that the abnormal miRNA expression observed in HCC [6–15] is secondary to defective RNA silencing machinery and that decreased Dicer expression is a cause rather than a consequence of HCC.

Conclusions

Our data demonstrate that Dicer is significantly down-regulated in HCC, suggesting that reduced expression of

Dicer may play an important role during the process of hepatocarcinogenesis.

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Conflict of interest statement Authors declare that no conflicting or competing interests, of any nature, exist between the Authors of this work and their Academic activity.

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