



Association of *CILP*, *COL9A2* and *MMP3* Gene Polymorphisms with Lumbar Disc Degeneration in an Indian Population

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Abstract

Lumbar disc degeneration (LDD) is a multifactorial disorder caused by genetic and environmental factors. Polymorphisms in several structural and inflammatory genes like collagens, aggrecan, matrix metalloproteinases are associated with the risk of disc degeneration. In this study, we analyzed the role of a few important single nucleotide polymorphisms in cartilage intermediate layer protein (*CILP*), collagen 9A2 (*COL9A2*) and matrix metalloproteinase 3 (*MMP3*) genes in LDD from an Indian population. Two hundred patients with LDD and 200 healthy controls were recruited for the study. Genotyping was performed by allelic discrimination assay. The rs2073711 polymorphism (*CILP* gene - GG genotype) was associated with reduced risk of LDD in the Indian population (OR = 0.43, $p = 0.016$). The rs591058 polymorphism (*MMP3* gene - TT genotype) is found to be associated with lower risk among women (OR = 0.34, $p = 0.041$). No significant association was found between *COL9A2* polymorphism rs7533552 and the risk of LDD. We conclude that the *CILP* gene polymorphism (rs2073711) is associated with a lower risk of LDD, the *MMP3* (rs591058) gene polymorphism is associated with LDD among women, and the TT genotype confers a lower risk of LDD.

Keywords Lumbar disc degeneration (LDD) · *CILP* · *MMP3* · *COL9A2* · Polymorphism · Indian population

Introduction

Lumbar disc degeneration (LDD) is a common musculoskeletal disorder involving degeneration of lumbar intervertebral discs (Andersson 1999). It is a complex disease with genetic and environmental factors playing important roles (Matsui et al. 1998; Battie et al. 1995; Sambrook et al. 1999). Several genes such as collagen, aggrecan, matrix metalloproteinases (MMPs), vitamin D receptor (VITDR) play a role in the pathology of LDD. Other risk factors such as physical loading, obesity and smoking are also thought to contribute to the disease significantly (Liuke et al. 2005; Battie et al. 1991, Zitting et al. 1998).

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The intervertebral disc is a specialized form of fibrocartilage divided into an outer layer, the annulus fibrosus and inner layer, and the nucleus pulposus. The annulus fibrosus has a characteristic ring-like structure consisting mainly of densely packed collagen I fibers. The nucleus pulposus is rich in collagen II (about 20% of its dry weight) and contains minor amounts of two other cartilage collagens, types IX and XI. Both layers contain small amounts of collagen VI and minor amounts of collagens III and V are present throughout the disc. Proteoglycans form almost 50% of the dry weight of the nucleus pulposus. Collagens provide tensile support for the disc and the extensively hydrated proteoglycans give the tissue its resistance to compressive forces (Ala-Kokko 2002). Collagen IX is a structural protein with an important role in connecting various types of collagens together, particularly collagen II with proteoglycans in the matrix. Previous studies have shown that a few gene polymorphisms in the collagen gene are associated with LDD risk (Pluijm et al. 2004).

Cartilage Intermediate Layer Protein (*CILP*) gene is abundantly expressed in intervertebral discs and its expression increases when degeneration occurs (Seki et al. 2005). So the SNPs in this gene may play a role in the risk of LDD. Since matrix degradation is one of the hallmarks of LDD, single

nucleotide polymorphisms (SNPs) in matrix-degrading enzymes like MMPs and a disintegrin and metalloproteinase with thrombospondin motif (ADAMTS) may play an important role in LDD. MMP3 is one of the important enzymes present in the disc. MMP3 contributes to disc degeneration by degrading matrix proteoglycans and collagens. Polymorphisms in *MMP3* gene (rs679620, rs591058 and rs650108) have been associated with tendon/ligament injury prevalence (e.g., Achilles tendinopathy/rupture and anterior cruciate ligament rupture) (Bell et al. 2012; Laguetta et al. 2011; Collins and Raleigh 2009).

In this study, we explore the role of single nucleotide polymorphisms (rs2073711 of *CILP* gene, rs591058 of *MMP3* gene and rs7533552 of *COL9A2* gene) associated with lumbar disc degeneration.

Materials and Methods

Study Population

A total of 200 patients with LDD were recruited from the Department of Neurosurgery, National Institute of Mental Health and Neurosciences, Bangalore, India, for this study during January 2016 to April 2018. Patients with low back ache with sciatica and whose MRI showed lumbar disc degeneration were considered as LDD patients and recruited to the study. Patients with physical damage to the disc and other genetic disorders were excluded from the study. The control subjects ($n = 200$) were randomly selected from general population who were healthy adults without a history of back pain in the recent past. This study was approved by the institute's ethics committee for human studies, NIMHANS, Bangalore, India (Item No.XII, Sl.No.12.04, Neurosciences). Informed consent was obtained from all participants in this study. Five-milliliter blood samples were collected from patients and control subjects for molecular analysis.

Genotyping

Genomic DNA from blood was isolated using Macherey-Nagel (MN) kits according to the manufacturer's protocol. Nanodrop ND2000c spectrophotometer was used for quantitation and purity checking. DNA with A260/280 1.75–1.85 was used for genotyping. SNP genotyping was performed using Taqman® allelic discrimination assay (Applied Biosystems, Foster City, CA, USA) with a commercially available primer probe set (C_16027328_10 for rs2073711, C_785960_1_ for rs591058, AN7DRM6 for rs7533552). Genotyping was performed in an Applied Biosystems 7500 Fast machine.

Statistical Analysis

Statistical software R.3.3.1 was used for the analysis of data. Continuous variables were expressed as mean \pm SD and categorical variables are expressed as percent values or absolute numbers, as indicated. Differences in demographic characteristics between patients and controls were compared by using the χ^2 test for all categorical variables. To estimate the deviation of frequency of gene alleles in the tested population, we performed the Hardy-Weinberg equilibrium using χ^2 tests. Genotypes and allele frequencies were compared by χ^2 analysis. Association between risk and variables were expressed as odds ratio (OR) with 95% confidence interval (CI). The results were considered statistically significant at $p \leq 0.05$.

Results

Clinical and Demographical Details of Study Subjects

Table 1 provides the details of 200 LDD patients and 200 control subjects enrolled in the study. Median age of patients was 42 years (range 18–73 years) and controls was 40.5 years (18–80). There was no significance difference between distribution of age and sex among cases and controls.

CILP Gene (rs2073711)

The genotypic and allelic distribution of rs2073711 is given in supplementary table 1. The minor allele frequency was 0.333 for patients and 0.393 for controls. This SNP did not follow Hardy Weinberg equilibrium (HWE) in patients ($p = 0.023$) but followed HWE in controls ($p = 0.814$). Significant association was found between GG genotype and risk of LDD. GG genotype conferred significantly lower risk ($p = 0.016$, OR = 0.43, 95% CI = 0.21–0.85). In recessive model (GGvsAG + AA), significant association was found ($p = 0.009$). The genotypes were compared between different parameters like sex, smoking and alcohol consumption history. Among men, the GG genotype was found to be significantly associated (OR = 0.28, 95% CI = 0.09–0.81, $p = 0.019$). Also, among subjects with no history of smoking or alcohol consumption, the GG genotype was significantly associated with reduced risk of LDD. The data is given in supplementary table 2.

COL9A2 Gene (rs7533552)

The distribution of genotypes of rs7533552 was in HWE ($p = 0.339$). The allelic frequency for the T allele was 71.5% and for the C allele was 28.5% in patients and 72.2% for T and 27.8% for C in controls. The genotypic and allelic distribution of rs7533552 is given in supplementary table 3. Chi-square test was performed to see if there was any difference between

Table 1 Characteristics of patients and controls

Characteristics	Healthy controls (<i>n</i> = 200)	Patients (<i>n</i> = 200)	<i>p</i> value
Mean age (years ± SD)	42.92 ± 18.23	42.94 ± 12.47	0.543
Males, <i>n</i> (%)	117 (58.5)	128 (64.0)	0.259
Females, <i>n</i> (%)	83 (41.5)	72 (36.0)	
Smoking history, <i>n</i> (%)			
Yes	–	48(24.0)	
No	–	152(76)	
Alcohol consumption, <i>n</i> (%)			
Yes	–	45(22.5)	
No	–	155(77.5)	

genotypes of patients and controls. No significant association was found between genotypes and risk of LDD. The genotypes were compared between different parameters such as sex, smoking and drinking history, and no association was found. The data is given in supplementary table 4.

MMP3 Gene (Rs591058)

The distribution of genotypes of rs591058 was in HWE ($p = 0.377$). The allelic frequency for the C allele was 70.5% and for the T allele was 29.5% in patients and 64.7% for C and 35.3% for T in controls. The genotypic and allelic distribution of rs591058 is given in supplementary table 5. Chi-square test was performed to see if there was any difference between genotypes of patients and controls. No significant association was found between genotypes and risk of LDD. The genotypes were compared between different parameters such as sex, smoking and alcohol consumption history. There was no association between genotypes and smoking history. In women, the TT genotype was found to be significantly associated (OR = 0.34, 95% CI = 0.12–0.97, $p = 0.041$). In subjects with no history of alcohol consumption, the T allele was almost associated (OR = 0.73, 95% CI = 0.53–1.00, $p = 0.052$) with LDD. The data is given in supplementary table 6.

Discussion

LDD is a disease that involves complex molecular pathological mechanisms, and many genes and other environmental factors contribute to its risk. Disc degeneration is a process that begins early in life and is a result of a variety of intrinsic and extrinsic factors, including normal aging (Min et al. 2010). Despite extensive research in this field, the etiology of the disease is still not defined clearly. Polymorphisms in collagen, matrix degradation and inflammatory genes have been studied previously as these are the key players thought to be involved in disc degradation. In this study we analyzed the association of three SNPs

namely, rs2073711 (*CILP* gene), rs7533552 (*COL9A2* gene) and rs591058 (*MMP9* gene) with LDD.

rs2073711 (1184 T → C) is a polymorphism present in exon 8 of the *CILP* gene. T is the ancestral allele and C is the disease-associated risk allele. The change in allele leads to a substitution of amino acid Ile395Thr. *CILP* directly interacts with transforming growth factor- β 1 (TGF- β 1) and inhibits the TGF- β 1 mediated induction of extracellular matrix proteins such as collagen II and aggrecan (Seki et al. 2005). Previous studies have shown that the C allele (coding for Thr 395) of 1184 T/C increases binding and inhibition of TGF- β 1, suggesting that regulation of TGF- β 1 signaling by *CILP* plays a crucial role in the origin and pathogenesis of lumbar disc disease (Seki et al. 2005).

rs2073711 SNP has been associated with LDD in other populations. It was significantly associated with lumbar disc disease in Japanese subjects (Seki et al. 2005; Min et al. 2009, 2010). But this SNP was not associated with lumbar disc disease in Finnish or Chinese populations (Virtanen et al. 2007). However, in another Finnish population study, SNP rs2073711 was associated with disc degeneration among women (Kelempisioti et al. 2011). In an Indian study by Rajasekaran et al. (2013), this SNP was not associated with LDD. In our study, the GG genotype had a protective effect for LDD. Although the G allele is considered a risk allele, in our population it had a protective effect. Study in a larger cohort will give a clear idea about the genotypic distribution of this SNP in Indian population. Also, in our study the GG genotype showed a protective effect in men (OR = 0.28, 95% CI = 0.09–0.81, $p = 0.019$) and in subjects with no history of smoking (OR = 0.46, 95% CI = 0.22–0.97, $p = 0.042$) and no alcohol consumption (OR = 0.39, 95% CI = 0.18–0.85, $p = 0.017$).

We studied the polymorphism present in the *COL9A2* gene (rs7533552). In normal tissues collagen IX serves as a bridge between collagenous and non-collagenous proteins (Bagheri et al. 2016). The *COL9A2* gene is located in the chromosome 1p34. Collagen IX is made up of three unique polypeptides, namely α 1, α 2, and α 3, which are encoded by genes, collagen type 9 alpha 1 (*COL9A1*), collagen type 9 alpha 2 (*COL9A2*),

and collagen type 9 alpha 3 (*COL9A3*), respectively (Toktas et al. 2015). As collagen has an important role in maintaining the structural integrity of the intervertebral discs, genetic polymorphisms affecting the function or abundance of collagen can predispose a patient to disc degeneration. Collagen IX is an attractive candidate for LDD because it serves as a minor component in both the main structures of the intervertebral disk, the annulus fibrosus and the nucleus pulposus, in addition to its presence in cartilage and the vitreous body of the eye (Buckwalter 1995).

Several groups have studied the association of the *COL9A2* gene polymorphisms with the risk of LDD. A Finnish study reported that the *COL9A2* gene polymorphism (rs12077871) was associated with LDD (Annunen et al. 1999). Another Finnish study contradicted the previous study stating that rs12077871 was not relevant to LDD, but rs61734651 in the *COL9A3* gene was relevant to LDD (Paassilta et al. 2001).

rs7533552 is a missense variant. This is the first study involving this variant in an Indian population. rs7533552 was associated with hip osteoarthritis (Näkki et al. 2010). It was not associated with lumbar disc disease in a Japanese population (Seki et al. 2006). It was not associated with degenerative disc disease in a Han Chinese population (Song et al. 2010). No association was found between this SNP and lumbar spinal stenosis patients (Hyun et al. 2011). SNP rs7533552 was associated with disc bulging in a Finnish male twin cohort study (Videman et al. 2009). In our study this SNP was not associated with the risk of LDD.

The polymorphism rs591058 in the *MMP3* gene was also studied. MMPs are members of the zinc-endopeptidases, which can be divided into five groups: collagenases (MMP-1, MMP-8 and MMP-13), gelatinases (MMP-2 and MMP-9), stromelysins (MMP-3, MMP-7 and MMP-10), matrilysins and membrane-type MMPs (MT-MMPs) (Nagase et al. 2006; Cawston and Billington 1996). MMP-3 is involved in several functions such as extracellular matrix remodeling and degradation, cell proliferation and angiogenesis (Chakraborti et al. 2003). MMP-3 is directly involved in degradation of components of the extracellular matrix including proteoglycans, laminin, fibronectin, gelatins and collagens (Matrisian 1990). It is also indirectly involved in degradation of the extracellular matrix by activating other latent MMPs (Colombini et al. 2014; Takahashi et al. 2001). Hence, MMP-3 can mediate disc degeneration by the degradation of collagens and proteoglycans.

Promoter region polymorphism in the *MMP3* gene, which produces five or six repeats of adenine (5A and 6A, respectively) was associated with degenerative disc disease (Takahashi et al. 2001). The 5A allele has twice the promoter activity compared to the 6A allele, which leads to more protein production and a potential mechanism for increased degeneration (Ye et al. 1996). This polymorphism was associated with disc degeneration in Japanese population, Chinese

population and English women (Takahashi et al. 2001; Valdes et al. 2005; Yuan et al. 2010). In another study, two polymorphisms in the *MMP3* gene (rs645419 and rs646910) were studied and no association was found with the disease (Videman et al. 2009). One study found that rs591058 was associated with an increased risk of Achilles tendinopathy (Raleigh et al. 2009). The T allele is the minor allele for rs591058. In our study, the T allele showed a protective effect in women (OR = 0.34, 95% CI = 0.12–0.97, $p = 0.041$). Even though the p value is only borderline significant, we can consider it since our sample size is moderately large. Also, the 95% CI is 0.12–0.97 and it does not include one. So, from the p value and CI, 95% significance can be considered. Another Indian study by Rajasekaran et al. (2013) studied rs591058 SNP and they did not find any association with lumbar disc degeneration. In that study, overall association was studied, and female subgroup analysis was not provided.

In conclusion, we studied the role of three SNPs in LDD. rs7533552 (*COL9A2*) was not associated with LDD. rs591058 (*MMP3*) showed that the TT genotype had a reduced risk of LDD in women. rs2073711 (*CILP*) results showed that the GG genotype had a reduced risk of LDD in an Indian population. Our study gives a basic understanding about the distribution of these genotypes in Indian population. Further studies in larger cohorts are needed help us understand the role of these SNPs better. Functional analysis of these genes can be a useful tool in understanding their role in LDD.

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Compliance with Ethical Standards

Conflict of Interest The authors have declared that there is no conflict of interest financially or findings specified in this paper.

References

- Ala-Kokko L (2002) Genetic risk factors for lumbar disc disease. *Ann Med* 34:42–47
- Andersson GB (1999) Epidemiological features of chronic low-back pain. *Lancet* 354:581–585
- Annunen S, Paassilta P, Lohiniva J, Perala M, Pihlajamaa T, Karppinen J, Tervonen O, Kroger H, Lahde S, Vanharanta H, Ryhanen L, Goring HH, Ott J, Prockop DJ, Ala-Kokko L (1999) An allele of *COL9A2* associated with intervertebral disc disease. *Science* 285:409–412
- Bagheri MH, Honarpisheh AP, Yavarian M, Alavi Z, Siegelman J, Valtchinov VI (2016) MRI phenotyping of *COL9A2/Trp2* and *COL9A3/Trp3* alleles in lumbar disc disease: a case-control study in south-western Iranian population reveals a significant *Trp3*-disease association in males. *Spine* 41:1661–1667
- Battie MC, Videman T, Gil K et al (1991) Volvo award in clinical sciences. Smoking and lumbar intervertebral disc degeneration: an MRI study of identical twins. *Spine* 16:1015–1021

- Battie MC, Haynor DR, Fisher LD, Gill K, Gibbons LE, Videman T (1995) Similarities in degenerative findings on magnetic resonance images of the lumbar spines of identical twins. *JBJS* 77:1662–1670
- Bell RD, Shultz SJ, Wideman L, Henrich VC (2012) Collagen gene variants previously associated with anterior cruciate ligament injury risk are also associated with joint laxity. *Sports Health* 4(4):312–318
- Buckwalter JA (1995) Aging and degeneration of the human intervertebral disc. *Spine* 20:1307–1314
- Cawston TE, Billington C (1996) Metalloproteinases in the rheumatic diseases. *J Pathol* 180(2):115–117
- Chakraborti S, Mandal M, Das S, Mandal A, Chakraborti T (2003) Regulation of matrix metalloproteinases: an overview. *Mol Cell Biochem* 253(1–2):269–285
- Collins M, Raleigh SM (2009) Genetic risk factors for musculoskeletal soft tissue injuries. *Med Sport Sci* 54:136–149
- Colombini A, Brayda-Bruno M, Lombardi G, Croiset SJ, Vrech V, Maione V, Banfi G, Cauci S (2014) FokI polymorphism in the vitamin D receptor gene (VDR) and its association with lumbar spine pathologies in the Italian population: a case-control study. *PLoS One* 9(5):e97027
- Hyun SJ, Park BG, Rhim SC, Bae CW, Lee JK, Roh SW, Jeon SR (2011) A haplotype at the COL9A2 gene locus contributes to the genetic risk for lumbar spinal stenosis in the Korean population. *Spine* 36(16):1273–1278
- Kelampisioti A, Eskola PJ, Okuloff A, Karjalainen U, Takatalo J, Daavittila I, Niinimäki J, Sequeiros RB, Tervonen O, Solovieva S, Kao PY (2011) Genetic susceptibility of intervertebral disc degeneration among young Finnish adults. *BMC Med Genet* 12(1):153
- Laguet M-J, Abrahams Y, Prince S, Collins M (2011) Sequence variants within the 3'-UTR of the COL5A1 gene alters mRNA stability: implications for musculoskeletal soft tissue injuries. *Matrix Biol* 30(5):338–345
- Liuke M, Solovieva S, Lamminen A, Luoma K, Leino-Arjas P, Luukkonen R, Riihimäki H (2005) Disc degeneration of the lumbar spine in relation to overweight. *Int J Obes* 29(8):903
- Matrisian LM (1990) Metalloproteinases and their inhibitors in matrix remodeling. *Trends Genet* 6:121–125
- Matsui H, Kanamori M, Ishihara H, Yudoh K, Naruse Y, Tsuji H (1998) Familial predisposition for lumbar degenerative disc disease. A case-control study. *Spine* 23:1029–1034
- Min SK, Nakazato K, Okada T, Ochi E, Hiranuma K (2009) The cartilage intermediate layer protein gene is associated with lumbar disc degeneration in collegiate judokas. *Int J Sports Med* 30(9):691–694
- Min SK, Nakazato K, Yamamoto Y, Gushiken K, Fujimoto H, Fujishiro H, Kobayakawa Y, Hiranuma K (2010) Cartilage intermediate layer protein gene is associated with lumbar disc degeneration in male, but not female, collegiate athletes. *Am J Sports Med* 38(12):2552–2557
- Nagase H, Visse R, Murphy G (2006) Structure and function of matrix metalloproteinases and TIMPs. *Cardiovasc Res* 69(3):562–573
- Näkki A, Videman T, Kujala UM, Suhonen M, Männikkö M, Peltonen L, Battié MC, Kaprio J, Saarela J (2010) Candidate gene association study of magnetic resonance imaging-based hip osteoarthritis (OA): evidence for COL9A2 gene as a common predisposing factor for hip OA and lumbar disc degeneration. *J Rheumatol*. <https://doi.org/10.3899/jrheum.100080>
- Paasilta P, Lohiniva J, Goring HH, Perala M, Raina SS, Karppinen J, Hakala M, Palm T, Kroger H, Kaitila I, Vanharanta H, Ott J, Ala-Kokko L (2001) Identification of a novel common genetic risk factor for lumbar disk disease. *JAMA* 285:1843–1849
- Pluijm SM, Van Essen HW, Bravenboer N, Uitterlinden AG, Smit JH, Pols HA, Lips P (2004) Collagen type I $\alpha 1$ Sp1 polymorphism, osteoporosis, and intervertebral disc degeneration in older men and women. *Ann Rheum Dis* 63(1):71–77
- Rajasekaran S, Kanna RM, Senthil N, Raveendran M, Cheung KM, Chan D, Subramaniam S, Shetty AP (2013) Phenotype variations affect genetic association studies of degenerative disc disease: conclusions of analysis of genetic association of 58 single nucleotide polymorphisms with highly specific phenotypes for disc degeneration in 332 subjects. *Spine J* 13(10):1309–1320
- Raleigh SM, van der Merwe L, Ribbans WJ, Smith RKW, Schwellnus MP, Collins M (2009) Variants within the MMP3 gene are associated with Achilles tendinopathy: possible interaction with the COL5A1 gene. *Br J Sports Med* 43(7):514–520
- Sambrook PN, MacGregor AJ, Spector TD (1999) Genetic influences on cervical and lumbar disc degeneration: a magnetic resonance imaging study in twins. *Arthritis Rheum* 42:366–372
- Seki S, Kawaguchi Y, Chiba K, Mikami Y, Kizawa H, Oya T, Mio F, Mori M, Miyamoto Y, Masuda I, Tsunoda T (2005) A functional SNP in CILP, encoding cartilage intermediate layer protein, is associated with susceptibility to lumbar disc disease. *Nat Genet* 37(6):607
- Seki S, Kawaguchi Y, Mori M, Mio F, Chiba K, Mikami Y, Tsunoda T, Kubo T, Toyama Y, Kimura T, Ikegawa S (2006) Association study of COL9A2 with lumbar disc disease in the Japanese population. *J Hum Genet* 51(12):1063
- Song HF, Wu ZH, Fei Q, Yan JZ, Liu Z, Zhang JG, Li SG, Qiu GX (2010) Association study of Trp2 allele polymorphism with degenerative disc disease in a Chinese Han nationality. *Zhonghua Yi Xue Za Zhi* 90(3):148–152
- Takahashi M, Haro H, Wakabayashi Y, Kawauchi T, Komori H, Shinomiya K (2001) The association of degeneration of the intervertebral disc with 5a/6a polymorphism in the promoter of the human matrix metalloproteinase-3 gene. *J Bone Joint Surg* 83:491–495
- Toktaş ZO, Ekşi MŞ, Yılmaz B, Demir MK, Özgen S, Kılıç T, Konya D (2015) Association of collagen I, IX and vitamin D receptor gene polymorphisms with radiological severity of intervertebral disc degeneration in southern European ancestor. *Eur Spine J* 24(11):2432–2441
- Valdes AM, Hassett G, Hart DJ, Spector TD (2005) Radiographic progression of lumbar spine disc degeneration is influenced by variation at inflammatory genes: a candidate SNP association study in the Chingford cohort. *Spine* 30(21):2445–2451
- Videman T, Saarela J, Kaprio J, Näkki A, Levälähti E, Gill K, Peltonen L, Battié MC (2009) Associations of 25 structural, degradative, and inflammatory candidate genes with lumbar disc desiccation, bulging, and height narrowing. *Arthritis Rheum* 60(2):470–481
- Virtanen IM, Song YQ, Cheung KM, Ala-Kokko L, Karppinen J, Ho DW, Luk KD, Yip SP, Leong JC, Cheah KS, Sham P (2007) Phenotypic and population differences in the association between CILP and lumbar disc disease. *J Med Genet* 44:285–288
- Ye S, Eriksson P, Hamsten A, Kurkinen M, Humphries SE, Henney AM (1996) Progression of coronary atherosclerosis is associated with a common genetic variant of the human stromelysin-1 promoter which results in reduced gene expression. *J Biol Chem* 271(22):13055–13060
- Yuan HY, Tang Y, Liang YX, Lei L, Xiao GB, Wang S, Xia ZL (2010) Matrix metalloproteinase-3 and vitamin d receptor genetic polymorphisms, and their interactions with occupational exposure in lumbar disc degeneration. *J Occup Health* 52(1):23–30
- Zitting P, Rantakallio P, Vanharanta H (1998) Cumulative incidence of lumbar disc diseases leading to hospitalization up to the age of 28 years. *Spine* 23:2337–2343