

POLG1-Related and other “Mitochondrial Parkinsonisms”: an Overview

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Abstract Mitochondrial dysfunction has been implicated in the pathogenesis of sporadic, idiopathic Parkinson disease. In some cases, mitochondrial DNA primary genetic abnormalities, or more commonly, secondary rearrangements due to polymerase gamma (*POLG1*) gene mutation, can directly cause parkinsonism. The case of a Parkinson disease patient with some signs or symptoms suggestive of mitochondrial disease (i.e., ptosis, myopathy, neuropathy) is a relatively common event in the neurological practice. Mitochondrial parkinsonisms do not have distinctive features allowing an immediate diagnosis, and a negative family history does not rule out a possible diagnosis of mitochondrial disorder. In this article, we do not revise the mitochondrial hypothesis of sporadic, idiopathic Parkinson disease, extensively discussed elsewhere, but we review *POLG1*-related parkinsonism and other well-defined forms of “mitochondrial parkinsonisms”, with mtDNA mutations or rearrangements. Lastly, we try to introduce a possible diagnostic approach for patients with parkinsonism and suspected mitochondrial disorder.

Keywords Haplogroups · Mitochondria · Mitochondrial diseases · mtDNA · Parkinson’s disease · *POLG*

Introduction

Parkinson disease (PD) is a common neurodegenerative movement disorder (Dauer and Przedborski 2003). The

cardinal features of PD include resting tremor, rigidity, bradykinesia and postural instability. These symptoms are caused by the progressive loss of the dopamine neurons within the substantia nigra pars compacta and the associated deficiency of the neurotransmitter dopamine in the striatum. Degeneration in other areas (i.e., the locus coeruleus, the dorsal raphe nuclei, and others) may contribute to many of the non-motor symptoms of PD, such as depression, dementia, and autonomic dysfunction (Dauer and Przedborski 2003). In addition, the surviving neurons contain cytoplasmic, eosinophilic inclusions called Lewy bodies (Dauer and Przedborski 2003). PD exists as both familial and idiopathic forms. Linkage studies showed 16 PD-related genetic loci (*PARK1-16*). Mutations in several genes have been linked to the genetic forms of PD (α -synuclein, Parkin, *DJ-1*, *PINK1*, and *LRRK2*; Bueler 2009). Mitochondrial dynamics (fission, fusion, migration) is important for neurotransmission, synaptic maintenance, and neuronal survival. Recent studies have shown that *PINK1* and *Parkin* play crucial roles in the regulation of mitochondrial dynamics and function. Moreover, mutations in *DJ-1* and *Parkin* render animals more susceptible to oxidative stress and mitochondrial toxins implicated in sporadic PD, lending support to the hypothesis that PD may be caused by gene-environmental factor interactions (Bueler 2009). The etiology of sporadic PD remains largely unclear, but accumulating evidence suggests that mitochondrial dysfunction occurs in brain and peripheral tissues of PD patients (Fukae et al. 2007). Since the discovery that the toxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine, an inhibitor of the mitochondrial complex I, was an environmental factor capable of causing a disease indistinguishable from sporadic PD, a number of studies have revealed that mitochondrial dysfunction plays an important role in the neurodegenerative process leading to PD (Fukae et al.

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2007). Especially complex I dysfunction has been implicated in the pathogenesis of PD (Parker et al. 1989).

Accumulation of clonal, somatic mitochondrial DNA (mtDNA) deletions has been observed in the substantia nigra during aging and in PD, suggesting that mtDNA mutations in some instances may predispose to dopamine neuron death by impairing mitochondrial respiration (Bueler 2009). A role of mtDNA has been also suggested by the finding that normal cells transfected with mtDNA from PD patients (cytoplasmic hybrids (cybrids)) showed 25% decreased complex I activity (Gu et al. 1998). In the cybrid technique, culturable cells depleted of endogenous mtDNA are repopulated with mitochondria (with their own mtDNA) from patients (Mancuso et al. 2008a).

The role of mtDNA variants in PD has been extensively studied (Mancuso et al. 2008a). van der Walt and coworkers (van der Walt et al. 2003) genotyped ten single-nucleotide polymorphisms that define the European mtDNA haplogroups in 609 white PD patient and 340 white controls. Mitochondrial haplogroups are specific and unique sets of common mtDNA polymorphisms that have evolved from the same ancestor. These authors observed that haplotype J and K reduced the incidence of PD by 50% (van der Walt et al. 2003). Consistently with this study, it has been subsequently reported that haplotype cluster UKJT was associated with a 22% reduction in population-attributable risk for PD (Pyle et al. 2005). Autere et al. (2004) observed that the supercluster JTIWX increased the risk of both PD and PD with dementia; this cluster was associated with a twofold increase in nonsynonymous substitutions in the mtDNA genes encoding complex I subunits. Another group evaluated the distribution of mtDNA haplogroups in a large cohort of 620 Italian patients with adult-onset idiopathic PD vs. two groups of ethnic-matched controls (Ghezzi et al. 2005). These authors found that haplogroup K was associated with a lower risk; they reported also that the 10398G polymorphism resulted protective against PD (Ghezzi et al. 2005). This last finding (but not the association between haplogroup K and lower risk) was later confirmed in 271 Spanish PD patients vs. 230 healthy controls (Huerta et al. 2005). Overall, despite some negative studies (Latsoudis et al. 2008), mtDNA haplogroups have been associated with a number of different neurodegenerative diseases, but to date, the only disease consistently associated with a different mtDNA haplogroup frequency is PD (Mancuso et al. 2008a). However, recent data failed to demonstrate a bias towards maternal inheritance in familial PD (Simon et al. 2010).

It is now well established that defects in mtDNA replication can lead to mitochondrial dysfunction and disease. DNA polymerase gamma (POLG) is the only DNA polymerase in human mitochondria and is essential for mtDNA replication and repair (Chan and Copeland

2009). In addition to its 5' to 3' polymerase activity, POLG has a 3' to 5' exonuclease activity important in the repair process. Mitochondria have their own small 16.5-kb circular double-stranded DNA that encodes 22 tRNAs, 2 rRNAs, and 13 polypeptides that are essential for electron transport and oxidative phosphorylation. Nuclear genes encode the other mitochondrial proteins, including the proteins involved in mtDNA replication, such as POLG subunits 1 and 2 (codified by *POLG1* and *POLG2* nuclear genes; Chan and Copeland 2009). *POLG1* mutations lead to uncorrect nucleotide incorporation in mtDNA (Batabyal et al. 2010). Homozygous knock-in mice expressing a proof-reading-deficient version of POLG show an “mtDNA mutator” phenotype with increased amounts of point mutations and deleted mtDNA. This increase in somatic mtDNA mutations is associated with reduced lifespan and premature onset of aging-related phenotypes, thus providing a link between mtDNA mutations and aging in mammals (Trifunovic et al. 2004).

POLG1 has a polyglutamine tract (poly-Q) in the N-terminal, encoded by a CAG sequence in exon 2 (Chan and Copeland 2009). Commonly, the poly-Q tract comprises ten repeats (10Q, frequency >80%) or 11Q (frequency 6–12%); however the composition of poly-Q alleles has been reported to vary from 6Q to 14Q. *POLG1* CAG repeat variants have been associated with male infertility, testicular cancer (Chan and Copeland 2009) and, interestingly, with idiopathic sporadic PD (Luoma et al. 2007). However, the role of the CAG trinucleotide repeat in *POLG1* is controversial (Chan and Copeland 2009). Very recently, Eerola et al. (2010) sequenced the poly-Q in 641 North American Caucasian PD patients and 292 controls. Caucasian literature controls were also used. Normal allele was defined either as 10/11Q or as 10Q according to the previous literature (Eerola et al. 2010). Variant alleles defined as non-10Q were significantly increased in the PD patients compared to the North American controls as well as compared to the larger set of 897 controls, strengthening the evidence for involvement of *POLG1* CAG repeats in PD (Eerola et al. 2010). Similar results have been obtained by Anvret et al. (2010) in Swedish PD patients. These authors reported a significant association between the non-10/11Q repeats with PD (Anvret et al. 2010). These findings may be attributable to a beneficial function of the 10/11Q repeat protein or to linkage disequilibrium between the 10Q allele and another variation within *POLG1* (Eerola et al. 2010). However, further large case-control studies are strongly needed, as well as analyses on functional differences of *POLG1* poly-Q variants.

In humans, *POLG1* mutations account for a substantial proportion of patients with mitochondrial myopathy and mtDNA deletions or depletion, and may cause both clinically and genetically heterogeneous disorders (Filosto

et al. 2003). The case of a PD patient with some signs or symptoms suggestive of mitochondrial disease (i.e., ptosis, myopathy, neuropathy) is a relatively common event in the neurological practice. In this article, we do not revise the mitochondrial hypothesis of sporadic, idiopathic PD, extensively discussed elsewhere (Fukae et al. 2007), but we review *POLG1*-related parkinsonism and other well-defined forms of “mitochondrial parkinsonisms”, with mtDNA mutations or rearrangements. At last, we try to introduce a possible diagnostic approach for patients with parkinsonism and suspected mitochondrial disorder.

Mitochondrial Disorders

Mitochondrial disorders (MDs) are a group of disorders caused by impairment of the mitochondrial respiratory chain (DiMauro and Schon 2003). The effects of mutations affecting the respiratory chain may be multisystemic, with involvement of visual and auditory pathways, heart, central nervous system, and skeletal muscle (DiMauro and Schon 2003). The estimated prevalence of MDs is 1–2 in 10,000 (Schaefer et al. 2008). MDs are, therefore, one of the commonest inherited neuromuscular disorders. The genetic classification of MDs distinguishes disorders due to defects in mtDNA from those due to defects in nuclear DNA (nDNA; DiMauro and Schon 2003). The first ones are inherited according to the rules of mitochondrial genetics (maternal inheritance, heteroplasmy and the threshold effect, mitotic segregation; DiMauro and Schon 2003). Each cell contains multiple copies of mtDNA (polyplasm), which in normal individuals are identical (homoplasmy; DiMauro and Schon 2003). Heteroplasmy refers to the coexistence of two populations of mtDNA, normal and mutated. Mutated mtDNA in a given tissue has to reach a minimum critical number before oxidative metabolism is impaired severely enough to cause dysfunction (threshold effect; DiMauro and Schon 2003). Differences in mutational loads surpassing the pathogenic threshold in some tissues but not in others may contribute to the heterogeneity of phenotypes. Because of the mitotic segregation, the mutation load can change from one cell generation to the next and, with time, it can either surpass or fall below the pathogenic threshold (DiMauro and Schon 2003). Further, the pathogenic threshold varies from tissue to tissue, according to the relative dependence of each tissue on oxidative metabolism (DiMauro and Schon 2003). For instance, central nervous system, skeletal muscle, heart, endocrine glands, the retina, the renal tubule, and the auditory sensory cells are highly dependent on oxidative metabolism for energy generation. Moreover, it is important to notice that the occurrence of a single large-scale deletion, common cause of progressive external ophthalmoplegia

(PEO), is almost sporadic (DiMauro and Schon 2003). Therefore, a negative family history does not rule out a possible diagnosis of MD.

MD related to nDNA are caused by mutations in structural components or ancillary proteins of the respiratory chain, by defects of the membrane lipid milieu, of Coenzyme Q10 biosynthetic genes, and by defects in intergenomic signaling (associated with mtDNA depletion or multiple deletions; DiMauro and Schon 2003). Mutations in nuclear genes adenine nucleotide translocator 1 (*ANT1*), twinkle mitochondrial helicase (*PEO1*), and mitochondrial catalytic subunit of DNA polymerase (*POLG1*) result in MD with mtDNA multiple deletions—commonly with a PEO phenotype—or mtDNA depletion with Alpers’ syndrome phenotype (Filosto et al. 2003; Gonzalez-Vioque et al. 2006; Copeland 2010). More recently, *OPA1* gene has been also associated with MD and mtDNA multiple deletions (Stewart et al. 2008). In a family with autosomal dominant PEO and parkinsonism, a heterozygous *PEO1* mutation has been reported to segregate with the disease phenotype (Baloh et al. 2007). No pathogenic mutations were present in *POLG1* or *ANT1* (Baloh et al. 2007). Very rarely, nuclear mutations in structural components of the respiratory chain, such as *NDUFV2* (Nishioka et al. 2010), may also cause familial PD, but further studies are still needed.

“Mitochondrial Parkinsonisms”

As mentioned above, there is increasing evidence that impairment of mitochondrial function and oxidative damage are contributing factors to the pathophysiology of sporadic PD. In some cases, mitochondrial genetic abnormalities can directly cause parkinsonisms (Mancuso et al. 2008a).

mtDNA point mutations causing parkinsonism have been reported only rarely, typically as one feature of a larger syndrome, i.e., with sensorineural deafness and neuropathy (Thyagarajan et al. 2000), MELAS (mitochondrial encephalomyopathy with lactic acidosis and stroke-like episodes; De Coo et al. 1999) or Leber’s hereditary optic neuropathy (Nikoskelainen et al. 1995). A family with maternally inherited, adult-onset multisystem degeneration including prominent parkinsonism was reported in 1999 (Simon et al. 1999). The parkinsonism was levodopa responsive and was associated with the loss of pigmented neurons in the substantia nigra in at least one patient. In this family, a mutation of the mitochondrial *ND4* gene of complex I (previously reported in Leber’s disease) was found (Simon et al. 1999).

Horvath et al. (2007) observed the A8344G “MERRF” (myoclonic epilepsy with ragged red fibers) mutation on the tRNA^{Lys} gene of the mtDNA in a 66-year-old man with

dopa-responsive parkinsonism, reduced muscle strength, and ragged red fibers (pathological hallmark of mitochondrial myopathy). No other clinical signs typical of MERRF were noted. The A8344G mutation was present in a virtually homoplasmic state in the patient's muscle, and 80% mutant was detected in blood DNA (Horvath et al. 2007). In order to evaluate if this mutation could represent a common cause of sporadic PD, our group subsequently analysed 159 Italian patients with PD (Mancuso et al. 2008b). None of the patients carried the A8344 mutation in DNA extracts from peripheral blood lymphocytes. Thus, the screening for this mutation should be restricted to PD patients with other canonical features of mitochondrial encephalomyopathies (i.e., myopathy, ophthalmoplegia, neuropathy, cerebellar ataxia; Mancuso et al. 2008b).

More frequently parkinsonism has been associated with large-scale rearrangements in mtDNA (Siciliano et al. 2001). Further, co-existence of parkinsonism and mutations in the mitochondrial polymerase gamma nDNA gene (*POLG1*) with mtDNA multiple deletions has been reported in several families, suggesting that when defective, this gene can be responsible for some of the mendelian transmission of Parkinsonism (Mancuso et al. 2008a). *POLG1*-related parkinsonisms will be considered in the next paragraph.

mtDNA mutations may also cause movement disorders other from parkinsonism, i.e., our group reported the case of a 36-year-old man who had severe postural and kinetic tremor of the arms since the age of 17 years and a mild myopathy. The levodopa test resulted negative, and DATSCAN showed a normal dopamine transporters concentration in the striatum, suggesting a diagnosis of essential tremor (Mancuso et al. 2008c). Southern Blot analysis revealed the presence of a single deletion of 7.7-kb interesting 58% of mtDNA in the skeletal muscle (Mancuso et al. 2008c).

Interestingly, Simon et al. (2003) reported a mtDNA mutation in the gene encoding the *ND1* subunit of mitochondrial complex I in a patient with adult-onset dystonia, spasticity, and myopathy. The same mutation subsequently was identified in two additional unrelated patients with adult-onset dystonia (Simon et al. 2003). Subsequently, two independent patients presenting with progressive generalized dystonia and bilateral striatal necrosis with a mutation in the mitochondrial *ND6* gene were reported (Solano et al. 2003). More recently, “mitochondrial dystonia” has also been associated with mitochondrial *ND3* gene (Sarzi et al. 2007; Wang et al. 2009).

***POLG1* in Human Disease**

POLG1 mutations can cause dominant or recessive disorders, frequently associated with severe and multisystem involvement. These forms of MD are associated with secondary

accumulation of multiple deletions in mtDNA (Gonzalez-Vioque et al. 2006). Mutations in the *POLG1* gene have emerged as one of the most common causes of inherited MDs in children and adults (Wong et al. 2008). Functional genetic variants of *POLG1* are present in up to 0.5% of the general population, and pathogenic mutations have been described in most exons of the gene (Hudson and Chinnery 2006). The most severe manifestations have been associated with mutations of the “spacer” region of *POLG1* (Luoma et al. 2005). In a large study, the clinical presentation ranged from the neonatal period to late adult life, with an overlapping phenotypic spectrum from severe encephalopathy and liver failure to late-onset PEO, ataxia, myopathy and isolated muscle pain or epilepsy (Horvath et al. 2006). A high incidence of psychiatric disease and primary gonadal failure have also been documented in some families transmitting dominant *POLG1* mutations (Hudson and Chinnery 2006). Very recently, a new nosological entity has been also suggested—“*POLG1*-related multiple sclerosis-like illness” (Echaniz-Laguna et al. 2010; Harris et al. 2010).

The most common *POLG1* disease mutation (467A>T) has been found in all of the major *POLG1*-related diseases, i.e., Alpers syndrome, ataxia-neuropathy syndromes and PEO (Chan and Copeland 2009). 467A>T mutation is generally reported as a recessive mutation; however, it may give rise to a mild dominant phenotype with late-onset ptosis (Chan and Copeland 2009). This mutation was shown to lead to a severe catalytic defect, disrupting physical association between *POLG* catalytic and accessory subunits, with great reduction of both polymerase and exonuclease *POLG* activities (Chan et al. 2005).

Moreover, mutations in *POLG1* cause a recessively inherited syndrome with episodic features and progressive ataxia (Winterthun et al. 2005). Hakonen et al. (2005) reported the high carrier frequency of the 748 W>S *POLG1* substitution in control subjects in Finland, explaining the high prevalence of mitochondrial recessive ataxia syndrome (MIRAS) in Scandinavia. However, although 467A>T and 748 W>S are a common cause of ataxia in Scandinavia, they are rare in other European regions, such as the United Kingdom and Italy (Craig et al. 2007). It is therefore likely that the high prevalence of MIRAS in Finland and Norway is due to a founder effect. Very recently, it has been reported that *POLG1* mutations are rather common in Central European ataxia patients, and cause ataxia with PEO (47%), and, more rarely, with psychiatric comorbidities (20%) and epilepsy (14%; Schicks et al. 2010).

***POLG1*-Associated Parkinsonism**

The first family in which parkinsonism was a prominent manifestation together with PEO and multiple mtDNA

deletions, caused by a novel *POLG1* mutation, was reported in 2004 (Mancuso et al. 2004). A missense mutation in the *POLG1* gene (2492A>G) was found in the proband (a woman with PEO, mild axonal neuropathy and parkinsonism since the age of 47, with slowly progressive resting hand tremor, slowing of her gait, bradykinesia, rigidity, and slurred speech) and in her affected siblings (Mancuso et al. 2004). The patient's brother developed parkinsonism in his early 40s. The proband's muscle biopsy showed numerous ragged-red fibers and cytochrome *c* oxidase-negative fibers, with multiple deletions of mtDNA (Mancuso et al. 2004). This mutation, however, has also been found in Europe in healthy population. Therefore, its pathogenetic role is still unclear.

In the same period, Luoma et al. (2004) studied patients with *POLG1*-related PEO and their healthy relatives in seven families of various ethnic origins, by clinical, biochemical, morphological, and molecular characterisation, and by positron emission tomography (PET) studies. A significant cosegregation of parkinsonism with *POLG1* mutations was observed, and PET findings were consistent with dopaminergic neuron loss (Luoma et al. 2004). Post-mortem examination in two individuals showed loss of pigmented neurons and pigment phagocytosis in the substantia nigra, without Lewy bodies (Luoma et al. 2004). Furthermore, most women with PEO had early menopause (before age 35 years). The *POLG1* gene defect resulted in secondary accumulation of mtDNA deletions in patients' tissues (Luoma et al. 2004).

Tzoulis and co-workers studied 26 patients belonging to 20 families with a disorder caused by mutations in the *POLG1* gene (Tzoulis et al. 2006), homozygous for 1399 G>A or 2243 G>C or compound heterozygotes for these two mutations. Irrespective of genotype, the patients exhibited a progressive neurological disorder usually starting in their teens and characterized by epilepsy, headache, ataxia, neuropathy, myoclonus and late-onset PEO. Compound heterozygotes had a significantly shorter survival time. Epilepsy occurred in 22 of the 26 patients and in the majority of these there was an occipital EEG focus (Tzoulis et al. 2006). Patients with this disorder are at high risk of death from status epilepticus and from liver failure, if exposed to sodium valproate. In this group was included a heterozygous woman who developed epilepsy at the age of 55 years. When examined at age 72, she had axonal peripheral neuropathy, ataxia, mild ptosis/PEO, and features of mild parkinsonism (unilateral bradykinesia, tremor and rigidity; Tzoulis et al. 2006).

The group of Chinnery studied a large family with autosomal dominant PEO and parkinsonism, with multiple mitochondrial DNA deletions (Hudson et al. 2007). These authors identified two novel heterozygous *POLG1* exon

substitutions. Autosomal-dominant progressive PEO segregated with 1532 G>A mutation in exon 8, predicted to alter a highly conserved serine to asparagine in the linker region of POLG. The one patient with parkinsonism had an additional heterozygous substitution in exon 7 (1389 G>T), in trans as regards the other mutation. This mutation, present only in the patient with parkinsonism, was predicted to alter a highly conserved leucine to phenylalanine and, based on this pedigree, it seemed unlikely that it could cause disease on its own (Hudson et al. 2007). None of the substitutions were found in a large group of healthy controls, patients with PEO without parkinsonism, and patients with sporadic PD. These two mutations did not directly affect the polymerase domain of POLG. Like other linker region mutations, they are likely to affect polymerase activity of the enzyme (Hudson et al. 2007). DATSCAN from the index case with PEO and parkinsonism showed reduced uptake in the left caudate nucleus and both putamen, consistent with a nigrostriatal dopaminergic deficiency (Hudson et al. 2007). Although his symptoms improved on a dopamine agonist, he had no tremor and the extrapyramidal rigidity was symmetric.

Pagnamenta et al. (2006) reported a family in which the proband, her mother, and her maternal grandmother presented with PEO and premature ovarian failure. The mother developed parkinsonian-like resting tremor and reduced rapid alternating movements affecting her left arm and leg, and mild bradykinesia, in her sixth decade. The tremor responded to levodopa treatment. Sequence analysis of *POLG1* revealed a dominant 955Y>C mutation segregating with the disease. Southern blot analysis demonstrated mtDNA depletion in fibroblasts (43% of controls), whereas multiple rearrangements of mtDNA were seen in skeletal muscle (Pagnamenta et al. 2006). Mouse and yeast models with this mutation show enhanced amounts of oxidative lesions (Graziewicz et al. 2007). The 955Y>C mutated POLG displays relaxed discrimination when incorporating oxidatively damaged nucleotides, offering a biochemical link between oxidative stress and PD, and suggesting that patients harbouring *POLG1* mutations may undergo enhanced oxidative stress and mtDNA mutagenesis (Graziewicz et al. 2007).

In previous years, other patients with *POLG1* gene mutation and parkinsonism have been reported (Galassi et al. 2008; Betts-Henderson et al. 2009). Two sisters with early-onset parkinsonism (dystonic toe curling, action tremor, masked face, bradykinesia, stooped posture, and rigidity), together with clinical and electrophysiological signs of sensorimotor axonal peripheral neuropathy, were reported to have ragged-red and cytochrome *c* oxidase-negative fibers in muscle biopsy (Davidzon et al. 2006). Multiple mitochondrial DNA deletions were found by long polymerase chain reaction, and sequencing of the *POLG1*

gene showed that the patients were compound heterozygous for two pathogenic mutations (Davidzon et al. 2006). Therefore, *POLG1* mutations can cause early-onset parkinsonism even in the absence of PEO.

Remes et al. (2008) reported the 748 W>S mutation (one of the most common mutations in this gene, found to be a frequent cause of autosomal recessive ataxia in adults and of Alpers syndrome in children) in a 65-year-old man with a late-onset syndrome consisting of ataxia, parkinsonism, PEO, peripheral neuropathy, and sensorineural hearing loss (Remes et al. 2008). Interestingly, Invernizzi et al. (2008) reported a compound heterozygous patient with two novel mutations in *POLG1*, who had autosomal recessive PEO, followed by the development of pseudo-orthostatic tremor evolving into levodopa-responsive parkinsonism.

Conclusion

In some cases, mtDNA primary genetic abnormalities, or more commonly secondary mtDNA rearrangements due to *POLG1* gene mutation, can directly cause parkinsonism. Mitochondrial parkinsonisms do not have distinctive features allowing an immediate diagnosis. Usually, age at onset is about 50 years. Reduced dopamine uptake in the corpus striatum and good response to levodopa or dopamine agonists (Synofzik et al. 2010) have been well documented. Levodopa-induced dyskinesias and motor fluctuations may also occur (Wilcox et al. 2007). *POLG1* mutations should be considered in the differential diagnosis of parkinsonism, especially in families with autosomal dominant transmission. However, a negative family history does not rule out a possible diagnosis of MD.

At present, diagnosis of MD requires a complex approach, including clinical and family data, measurement of serum lactate, exercise testing, electromyography, muscle histology, and enzymology, and genetic analysis (Fig. 1). Ragged red fibers, ragged blue fibers, and cytochrome *c* oxidase-negative muscle fibers are pathological hallmarks. However, reliable biomarkers for these disorders are still needed. “Red flags” for MD are short stature, neurosensory hearing loss, ptosis, ophthalmoplegia, axonal neuropathy, diabetes mellitus, hypertrophic cardiomyopathy, renotubular acidosis, migraine-like headache (Mancuso et al. 2009). Therapy of MD is still inadequate, despite great progress in the molecular understanding of these disorders. Therapies that have been attempted include respiratory chain cofactors, other metabolites secondarily decreased in MD, antioxidants and agents acting on lactic acidosis, but their role in the treatment of the majority of MD remains unclear (Orsucci et al. 2009).

It is important to consider a possible diagnosis of mitochondrial parkinsonism in PD patients with other

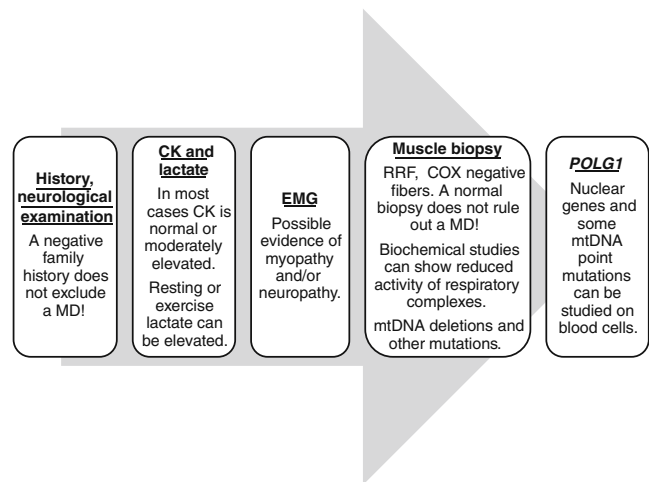


Fig. 1 Diagnostic approach to mitochondrial disorders (MD) in patients with parkinsonism. This figure is conceived as a flow chart, but the real diagnostic process depends on the specific clinical presentation of the patient, and is not a rigid process. *COX* cytochrome *c* oxidase, *EMG* electromyography, *mtDNA* mitochondrial DNA, *RRF* ragged red fibers

features of MD (Fig. 2), and especially of *POLG1*-related mitochondrial encephalomyopathies (i.e., PEO, myopathy, isolated muscle pain, dysphagia, neuropathy, ataxia, psychiatric disorders, hypogonadism, epilepsy). *POLG1* mutations result in extremely heterogenous phenotypes which often have overlapping clinical findings, making it difficult to categorize patients into syndromes. The lack of signs of mitochondrial dysfunction in the muscle biopsy does not exclude a *POLG1*-related disease (Milone and Massie 2010). Analysis of mtDNA of clinically affected tissues is often informative, but not always. The clinical features of the patient are more important to select putative *POLG1*

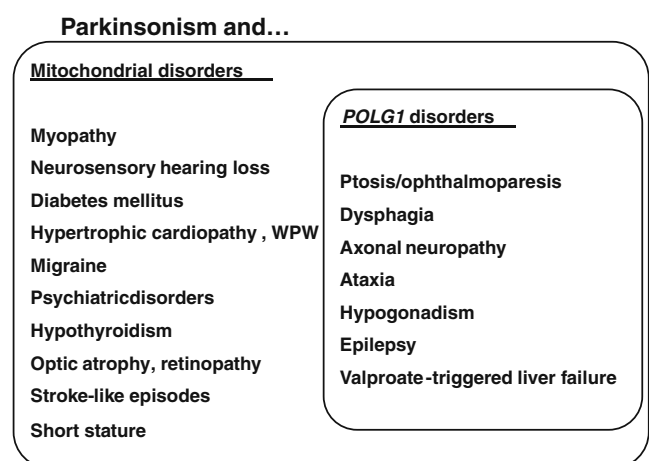


Fig. 2 “Red flags” for mitochondrial disorders and particularly for *POLG1*-related disorders. It is important to consider a possible diagnosis of mitochondrial parkinsonism in patients/families with Parkinson disease and some of these adjunctive features. *WPW* Wolff–Parkinson–White

mutation carriers than the presence of mtDNA deletions or muscle oxidative phosphorylation activity defects (Blok et al. 2009). Molecular analysis of *POLG1* is essential when *POLG1*-related disease is suspected. In these patients, sodium valproate should be avoided because of the risk of liver failure.

The case of a PD patient with some signs or symptoms suggestive of mitochondrial disease is a relatively common event in neurological practice. Conversely, patients with a previous diagnosis of MD may show some features of parkinsonism. Further studies are needed in order to study the prevalence of MD among PD patients, and conversely of movement disorders among MD patients. Furthermore, they should aim to better understand the molecular basis underlying the phenotypic variability among these patients. Large, multicenter studies are strongly needed to better characterize the phenotype and natural history of MD, in order to identify some countermeasures (i.e., pharmacological, physical, or others) capable of benefit patients with these chronic, still incurable disorders.

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